EFFECTS OF TWO DIFFERENT COMPOUNDS ON SEIZURE SUPPRESSION USING THE ZEBRAFISH PTZ-SEIZURE MODEL

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Abstract

Epilepsies are a common and severe neurological condition characterized by spontaneous and recurrent seizures. Although anti-seizure medications are effective for most patients, about 30% remain pharmacoresistant. Moreover, uncontrolled seizures are associated with risk factors and shortened life expectancy for individuals with refractory epilepsy. Preclinical studies are an essential step for drug discovery and the zebrafish (*Danio rerio*) has been successfully employed for this purpose. In this study, we applied the zebrafish PTZ-seizure model to investigate the effect of two compounds on seizure suppression, Tripeptide (p-BTX-I) and the Cx43 peptide CX2. Zebrafish larvae at 6 days post-fertilization (dpf) were exposed to both compounds, according to their group, 24h prior to PTZ-seizure induction. We quantified the compounds' effect on seizure latency, number of seizures and transcript levels of genes related to inflammation, oxidative stress, and apoptosis (*il1b, tnfa, cox1, cox2a, il6, casp3a, casp9, baxa, bcl2a, nox1, sod1* and *cat*).

Our results showed that CX2 at a concentration of 0.1 μM/mL yielded the best outcome for seizure suppression as it reduced the number of seizures and increased the seizure latency. Additionally, CX2 treatment before PTZ-induced seizures decreased the transcript of *il1b, il6, tnfa* and *cox1* genes, all related to inflammation. A bio-distribution study showed that the CX2 reached the zebrafish brain at both times investigated, 1h and 6h. Similarly, the tripeptide exhibited anti-inflammatory and anti-apoptotic action, reducing mRNA expression of the *il1b* and *casp9* genes. Our findings suggest that both Tripeptide and CX2 hold translational potential for seizure suppression.

Keywords: zebrafish$_1$, drug screening$_2$, seizure$_3$, compounds$_4$, pentylenetetrazole$_5$.

Introduction

Epilepsies are a common neurological disease characterized by recurrent unprovoked seizures, affecting approximately 50 million people worldwide [1,2]. Currently, anti-seizure medications are effective for 66% of people with epilepsy in developed countries [3,4]. However, more than 30% of people with epilepsy do not respond well to conventional therapies, making them pharmacoresistant [5]. Uncontrolled seizures can lead to various risks for people with epilepsy, such as risk of injury, neuropsychological impairment, and shortened lifespan [6]. Therefore, the search for new drugs or substances that could improve the treatment of people with epilepsy is urgent.

Pre-clinical trials are crucial in testing the therapeutic or toxicological effects of new substances, using *in vivo*, *in vitro*, or *ex-vivo* strategies. The scalability and rapid access to the results significantly contribute to the value of such studies. In this context, the zebrafish emerges as a pivotal model, offering distinct advantages over other animal models for pre-clinical trials [7]. These include its small size, low maintenance cost, high fecundity, and optical transparency during embryogenesis [8]. Furthermore, the zebrafish genome exhibits approximately 70% homology with the human genome, with 84% of known genes associated with human diseases, including epilepsy [9,10]. In recent years, several platforms for automatic data acquisition and analysis in the zebrafish model have been developed, enabling a variety of multiplexed phenotypic assays with minimal human intervention [11,12]. Moreover, the zebrafish aligns with the 3Rs (reduction, refinement, and replacement) philosophy [13]. Beyond its investigation of molecular pathways and behaviors relevant to various diseases, the zebrafish offers a time and cost-effective model due to its small size, among other attributes, which is advantageous for scalable studies involving simultaneous analysis of animals and drugs [14].

In the context of epilepsy, the zebrafish is a well-characterized seizure model in adults and larvae, as it is used for modeling epilepsy disorders by genetic manipulation [15–17]. In 2005, Baraban and

71 colleagues used the pro-convulsant agent pentylenetetrazol (PTZ) to induce seizures in zebrafish larvae
72 [16]. They observed a specific seizure behavior, up-regulation of the *c-fos* gene in the brain, and
73 electrographic discharges in the optic tectum after exposing the larvae to 15 mM PTZ at seven days
74 post-fertilization. Moreover, these responses were attenuated by common anti-seizure medications.
75 Overall, these findings suggest that zebrafish exhibit many similarities to traditional models such as
76 rodents.

77 Historically, natural substances from plants and animals are a source of numerous medicinal
78 preparations, widely employed for preventive and therapeutic care [18,19]. This fact is corroborated
79 by the Food and Drug Administration (FDA), which reports that between 1981 and 2019, 34% of drugs
80 were based on substances from natural products [20].

81 Considering the advantages of zebrafish for drug screening based on phenotype and the need to find
82 new treatments for controlling seizures, we aimed to evaluate the effect of different natural compounds
83 on seizure suppression using the PTZ-zebrafish model.

84 Materials and Methods

85 Zebrafish maintenance and embryo acquisition

86 Wild-type adult zebrafish were obtained from the Laboratory of Zebrafish and Husbandry at the School
87 of Medical Sciences, Unicamp. The animals were housed in 30 – 50 liter tanks, accommodating two
88 animals per liter of water. The tanks were maintained under controlled physicochemical conditions of
89 temperature (26±2°C), pH (7-7.5), levels of ammonia (< 0.1 ppm), nitrite (< 0.2 ppm) and dissolved
90 oxygen (4-11 ppm). A photoperiod cycle of 14 h of light and 10 h of darkness was maintained. Adult
91 fish were fed three times a day with commercial flake food (Tetramin, Tetra, Blacksburg, VA, USA)
92 and once a day with brine shrimp and paramecium. Embryos were collected following natural

93 spawning and nurtured in Petri dishes containing water from the aquariums maintaining consistent
94 temperature and photoperiod with the adults. From the 5th day onwards, the larvae were fed with
95 paramecium. Ethical approval for all experimental protocols was obtained from the Ethics Committee
96 for Animal Research of the State University of Campinas (CEUA 4895-1/2019 and CEUA 5757-
97 1/2021).

98 ## Compounds

99 **Tripeptide (p-BTX-I)**

100 The compound sequence (Glu-Val-Trp) was obtained from AminoTech – Research and Development
101 (São Paulo, Brazil). Subsequently, it was diluted in Milli-Q water (Merck KGaA, Darmstadt, Germany)
102 and then aliquoted. These aliquots were stored at -20°C. The employed concentrations (as listed in
103 Table 1) were determined based on research conducted in cell models [21,22].

104 **Table 1.** Compounds and their concentrations were analyzed to determine their effect on the
105 suppression of seizures induced by the proconvulsant agent pentylenetetrazole.

106

Compound	Concentration
Tripeptide (p-BTX-I)	25, 10 and 5 µg/mL
CX2	0.5, 0.1 and 0.05 µM

107
108
109

110 **CX2**

111 The CX2 (ARG-Cx43p) sequence corresponds to the C-terminal domain of the zebrafish Cx43 gene
112 (PCT/EP2020/071242). CX2 was assembled following standard Fmoc/tBu solid-phase MW-assisted

113 peptide synthesis protocols [23,24], purified by reverse-phase HPLC, and their identity confirmed by
114 HPLC-MS(ESI). The concentrations used (as listed in Table 1) were determined through studies
115 involving zebrafish models for regeneration and senescence. Like Tripeptide, CX2 was diluted in Milli-
116 Q water, aliquoted, and stored at -20°C.

117 Compounds pretreatment

118 For all compounds, zebrafish larvae at 6 days post-fertilization (dpf) were randomly placed into Petri
119 dishes, each one containing 25 larvae. The larvae were incubated in the respective solutions and
120 concentrations for 24h (as specified in Table 1) prior to seizure induction. The temperature and
121 photoperiod conditions were consistent with those described for the adult zebrafish. The study groups
122 consisted of the Control Group (CG), PTZ Group (PTZ), Treatment 1, Treatment 2, and Treatment 3.
123 Treatments 1 to 3 employed the previously determined concentrations mentioned above.

124 Seizure induction by pentylenetetrazol

125 Seizure induction followed the pretreatment of the compounds. Larvae with 7 dpf were carefully
126 transferred to a 96-well plate filled with 100 µL of aquarium water, with one larva per well.
127 Subsequently, 100 µL of 30 mM of the proconvulsant agent pentylenetetrazol (PTZ) (Sigma-Aldrich,
128 St. Louis, MO, USA) was added to each well, resulting in a final concentration of 15 mM of PTZ. The
129 larvae were exposed to PTZ for 20 minutes. The control group underwent the same procedure but in
130 PTZ-free water.

131 Latency and number of seizures

132 The latency and number of seizures were monitored through visual observation, following the method
133 described by Barbalho et al., 2016a [25]. Specifically, latency was defined as the period between the
134 initiation of PTZ exposure and the larva reaching stage 3 of seizure-like activity, according to the

135 criteria established by Baraban et al., 2005 [16]. Larvae were evaluated for 10 minutes of PTZ
136 exposure, and a complete seizure was considered when they reached stage 3.

137 RNA extraction

138 Immediately after PTZ exposure, fish were cryo-anesthetized and their heads were cut, collected, and
139 then incubated in TRIzol® (Invitrogen, Carlsbad, CA, USA). The biological material was lysed using
140 the TissueLyzer equipment (QIAGEN, GmbH, Germany) at 25 beats per second (BPS) for 2 minutes.
141 Subsequently, the heads were stored at -80 ºC until further processing. Each group consisted of five
142 samples (n = 5); however, each sample was composed by pooling five larval heads, to obtain sufficient
143 biological material for RNA extraction. Total RNA extraction was performed using TRIzol® according
144 to the manufacturer's protocol. The concentration and quality were determined using the Epoch™
145 spectrophotometer (BioTek, Winooski, VT, USA) and gel electrophoresis.

146 RT-qPCR

147 The synthesis of cDNA was carried out using the High-Capacity cDNA Reverse Transcription kit
148 (Applied Biosystems™, Califórnia, USA) following the manufacturer's instructions. Quantitative PCR
149 (qPCR) was performed using the SYBR ® Green Master Mix reagent (Bio-Rad) using the ABI 7500
150 system (Applied Biosystems, Foster City, CA, USA). Target genes (as listed in Table 2) were designed
151 using the Primer-BLAST online tool (https://www.ncbi.nlm.nih.gov/tools /primer-blast/) from NCBI
152 (National Center for Biotechnology Information) zebrafish database. Runs were carried out in triplicate
153 using *eef1a1l1* as a housekeeping gene [26] to normalize the genes from the inflammation pathway
154 (*il1b, tnfa, cox1, cox2a* and *il6*), the apoptosis pathway (*casp3a, casp9, baxa* and *bcl2a*), and the
155 oxidative stress pathway (*nox1, sod* and *cat*) were analyzed. Data were evaluated using 7500 software
156 v2.03 (Applied Biosystems). Relative gene expression analysis was calculated using the Livak and
157 Schmittgen equation RQ = 2-ΔΔCT [27].

158 The three pathways (inflammation, apoptosis, and oxidative stress) are all closely linked to the
159 underlying mechanisms of seizure generation and progression, which is why we selected representative
160 genes from these pathways for analysis.

161 **Table 2:** Primer sequences designed using the Primer-Blast online tool.

Gene	Seq.Ref.NCBI	Forward	Reverse	Size (pb)
eef1a1l1	NM_131263.1	AGCAGCAGCTGAGGAGTGAT	CCGCATTTGTAGATCAGATGG	140
il1b	NM_212844.2	GCTGGAGATCCAAACGGATA	ATTTGACGGACTCGAAGGTG	85
tnfa	NM_212859.2	TCGGGTGTATGGAGGGTGTT	TTGATTGCCCTGGGTCTTATGG	96
cox1	NM_153656.2	CTGGGAGGCTTATTCCAACA	CCAGAAGTTTAGGGTCTGGAAG	119
cox2a	NM_153657.1	ACCAGGGCGTGTGTTTATCC	GTGAGAAGCTCAGGGGTAGTG	100
il6	NM_001261449.1	GGCATTTGAAGGGGTCAGGA	GCGTTAGACATCTTTCCGTGC	92
casp3a	NM_131877.3	CAGCTTGAACTACCCCAACA	AAGCTTTCGAAACACGTTCA	133
casp9	NM_001007404.2	GCGACAAGCTGGAGAAAAGA	GATGACCACACAGCAGTCGTA	140
baxa	NM_131562.2	GAGCTGCACTTCTCAACAACTTT	GAAGATCTCACGGGCCACTC	245
bcl2a	NM_001030253.2	ACTACCTGAACGGGCCACT	AAAACGGGTGGAACACAGAG	105

nox1	XM_009291119.3	TCGTCAAGAAAACCCATCGTCT	TGTTGATCTTGCAGCACAGTCT	213
sod1	NM_131294.1	GGCCTTACTCCAGGAAAACA	TCTCCGACGTGTCTCACACT	140
cat	NM_130912.2	CAACGCTGAAGGGAAAAAGAACA	GCATTTGCTAAGATCACTTGTGTTG	250

162

163 **Statistical analysis**

164 Statistical analysis was performed using GraphPad Prism version 5.0 (GraphPad Software, San Diego,
165 CA, USA). One-way ANOVA was performed, followed by the Bonferroni method for multiple
166 comparisons. Differences were considered significant if $p<0.05$. Results are presented as mean ± SEM.

167 **Bio-distribution**

168 Bio-distribution analysis was conducted on Tripeptide and CX2, labeled with tetramethylrhodamine-
169 TMR and carboxytetramethylrhodamine – TAMRA, respectively.

170 Larvae at 6 dpf were transferred to a 96-well plate, with one larva per well, containing 100 µL of
171 aquarium water. Subsequently, 100 µL of the Tripeptide compound at 20 µg/mL or CX2 at 1 µM was
172 added according to the respective group, totalizing 200 ml of solution at final concentration of 10 µg/ml
173 for Tripeptide and 0.5 µM for CX2. Exposure times for both compound groups were 1, 6, 18, and 24
174 hours (n=3 each time). The control group underwent similar manipulation but in water only. The
175 incubation temperature during the experiments was maintained at 26±2°C. upon completion of
176 exposure, larvae were rapidly and carefully transferred to a Becker containing 0.02% tricaine (Tricaine,
177 Sigma) for approximately 2 minutes. Subsequently, the larvae were transferred to a Petri dish
178 containing 1% agarose as a base, along with a drop of water. Larvae were examined and photographed

179 using the Multizoon AZ100 microscope (Nikon, Tokyo, Japan) at 2X magnification. Two photographs
180 of each larva were taken, one in black and white, and the other capturing only the fluorescence
181 emission. Imaging merging was performed using the NIS-Elements software (Nikon, Tokyo, Japan).
182 The experiments and protocols were approved by the animal care and use committee of the Universidad
183 de Santiago de Compostela and the standard protocols of Spain (CEEA-LU-003 and Directive 2012-
184 63-EU).

185 # Results

186 ## Behavioral assay

187 We analyzed the effects of tripeptide and CX2 on seizure behavior. We found an increase in latency
188 only for animals pretreated with the CX2 at 0.1 µM, ($p \leq 0.05$) as well as a significant decrease in the
189 number of seizures ($p \leq 0.001$) when compared to the PTZ group (Fig 1B). No statistical significance
190 was found for the tripeptide (Fig 1A).

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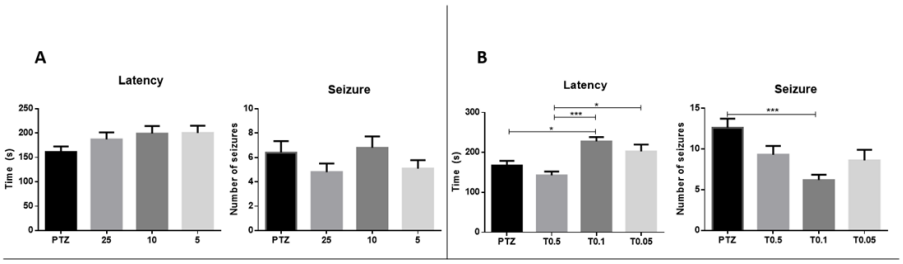
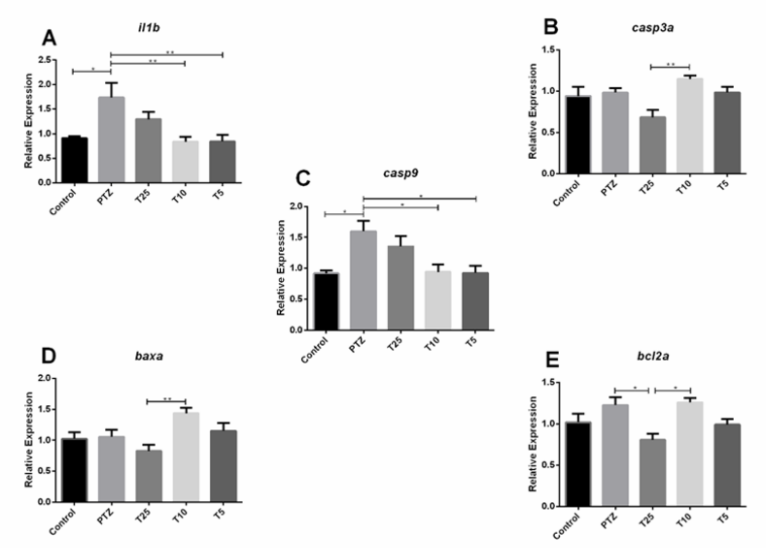


192 **Fig 1: The effect of compound treatment prior to pentylenetetrazol (PTZ)-induced seizures on**
193 **the number of seizure-like behaviors and latency was investigated.** The number of seizures and
194 latency were determined through visual inspection during a 10-minute exposure to PTZ (15 mM). (A)
195 Animals exposed to the tripeptide for 24 hours: Pentylenetetrazol (PTZ) group; treatment group 25
196 µg/mL (T25); 10 µg/ml treatment group (T10); treatment group 5 µg/ml (T5). (B) Animals exposed to
197 CX2 for 24 hours: Pentylenetetrazol (PTZ) group; treatment group 0.5 µM (T0.5); treatment group 0.1

198 μM (T0.1); treatment group 0.05 μM (T0.01). Data are presented as mean ± SEM. Statistical analyses
199 were performed using one-way ANOVA, followed by the Bonferroni method for multiple
200 comparisons. Differences were considered significant if $p < 0.05$. An asterisk (*) indicates that $p \leq 0.05$;
201 three asterisks (***), $p \leq 0.001$.

202 Molecular assay

203 Regarding the tripeptide, our results indicated downregulation of the *il1b* and *casp9* genes (Fig 2A and
204 2B), with significant effects at low concentrations (10 and 5 μg/mL). Additionally, we observed that
205 the expression of the *casp3a*, *baxa,* and *bcl2a* genes was most significantly reduced at a concentration
206 of 25 μg/mL ($p<0.05$), compared to the other concentrations. Similary, at a concentration of 10 μg/mL,
207 the *casp3a*, *baxa,* and *bcl2a* genes exhibited notably high expression levels, roughly equivalent to or
208 slightly higher than those of the PTZ group. However, no statistical difference was detected.



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210 **Fig 2: Expression of *il1b*, *casp3a*, *casp9*, *baxa* and *bcl2a* genes in zebrafish brain after**
211 **pentylenetetrazol (PTZ)-induced seizures.** Each treatment group was initially exposed to the
212 tripeptide for 24 h and subsequently to 15 mM PTZ for 20 min, and the control and PTZ groups were
213 handled identically, but with exposure to water (n = 5 per group). Data are presented as mean ± SEM.

Statistical analyses were performed with the one-way ANOVA, followed by the Bonferroni method for multiple comparisons. Differences were considered significant if $p<0.05$. An asterisk (*) indicates that $p\leq0.05$; two asterisks (**), $p\leq0.01$. Control group (Control); pentylenetetrazol (PTZ) group; treatment group 25 µg/mL (T25); 10 µg/ml treatment group (T10); treatment group 5 µg/mL (T5).

In the case of the CX2 compounds, we found down-regulating of the *il1b, cox1, il16, and tnfa* genes for most of the analyzed concentrations (Fig 3) compared to the PTZ group. The most significant down-regulation was observed at a concentration of 0.1 µM. Conversely, the *cox2a* gene displayed increased expression across all the three concentrations tested, with the most significant increase occurring at a concentration of 0.05 µM ($p\leq0.001$).

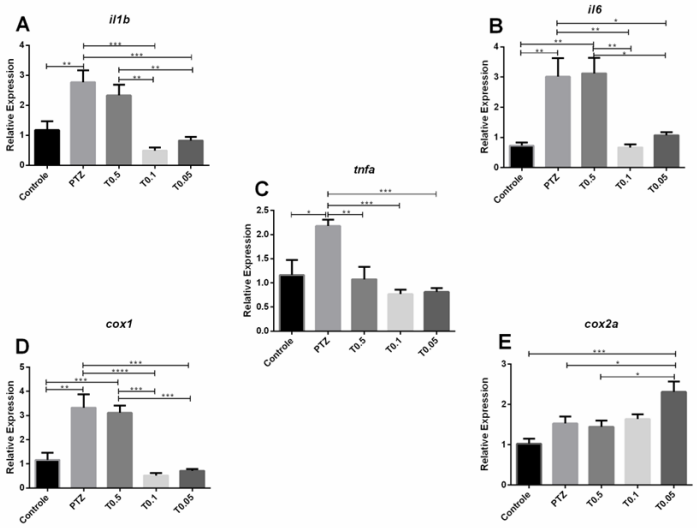
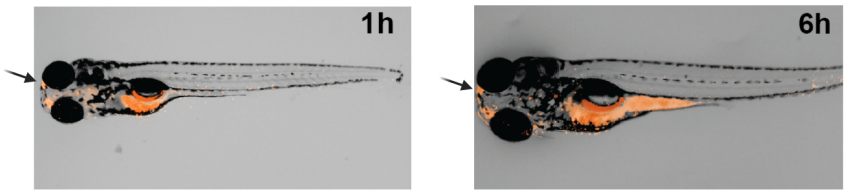


Fig 3: Expression of *il1b*, *il6*, *tnfa*, *cox1* and *cox2a* genes in zebrafish brain after pentylenetetrazol (PTZ)-induced seizures. Each treatment group was initially exposed to the CX2 for 24 h and subsequently to 15 mM PTZ for 20 min, and the control and PTZ groups were treated identically but with water exposure (n = 5 per group). Data are presented as mean ± SEM. Statistical analyses were performed with the one-way ANOVA, followed by the Bonferroni method for multiple comparisons. Differences were considered significant if $p<0.05$. An asterisk (*) indicates that $p\leq0.05$; two asterisks

230 (**), p≤0.01; three asterisks (***), p≤0.001; four asterisks (****), p≤0.0001. Control group (Control);
231 pentylenetetrazol (PTZ) group; treatment group 0.5 μM (T0.5); treatment group 0.1 μM (T0.1);
232 treatment group 0.05 μM (T0.01).

233 **Bio-distribution**

234 By tagging both the Tripeptide and CX2 with fluorescence, we tracked their biodistribution at four
235 time points, 1h, 6h, 18h and 24h. The CX2 compound exhibited robust detection in the brain after 1h
236 and persisted at 6 hours (Fig 4). Additionally, the compound was detected throughout the body,
237 indicating successful absorption by the zebrafish. In contrast, the tripeptide compound did not reach
238 the brain, and it was predominantly retained in the zebrafish intestine (S1 Fig).



239

240 **Figure 4: Time course of CX2 (8ARG-Cx43p, corresponding to the C-terminal domain sequence**
241 **of Cx43) efficiently crossing the blood-brain barrier (BBB) in a 7-day-old zebrafish larva.** The
242 carboxytetramethylrhodamine-TAMRA labeled peptide exhibited fluorescence microscopy detection
243 in the brain and throughout the body (indicated by red color) at 1 hour and 6 hours post-treatment.

244 # Discussion

245 Despite the availability of pharmacological treatments, uncontrolled seizures remain a concern,
246 requiring further investigations for new seizure-suppression approaches [28]. Seizures occur due to a
247 complex process involving multiple factors and affecting various cellular pathways [29]. This study
248 focused on targeting inflammation, oxidative stress, and cell death pathways for seizure modulation

249 [30–33]. To achieve our goal, we used tripeptide (p-BTX-I) to address inflammation and cell death,
250 and the CX2 for its anti-inflammatory action[21,22].

251 Zebrafish offer many advantages in drug screening and phenotype assessment. The zebrafish PTZ-
252 seizure model mimics human seizure behavior and electrographic patterns and is responsive to anti-
253 seizure drugs. The scalability and rapid results of the zebrafish make them an advantageous model for
254 discovering new anti-seizure compounds [8,9,16,34,35].

255 The process of inflammation involves the recruitment of many inflammatory mediators such as
256 interleukins (ILs), interferons (IFNs), tumor necrosis factors (TNFs), and growth factors. *IL1B*, *TNF*-
257 α, and *IL6* genes are among the most studied inflammatory cytokines in the CNS [32]. After a seizure,
258 genes such as *IL1B*, *IL2*, *IL6*, *TNF*-α, and *VEGF,* which are normally present in low concentrations,
259 increase rapidly, leading to damaging changes in synapses and heightened neuronal excitability
260 [33,36].

261 Prostaglandins/*COX-2* are also produced, potentially disrupting the blood-brain barrier and
262 significantly contributing to seizure onset and recurrence [37].

263 Among the compounds analyzed, CX2 at a concentration of 0.1 μM showed the most significant results
264 in seizure suppression, reducing seizure frequency and increasing latency (Fig 1B). Treatment with
265 CX2 prior to PTZ-induced seizures down-regulated the *il1b, cox1, tnfa, and il16* genes, indicating its
266 role in regulating inflammation during seizures [33]. In contrast, the *cox2a* gene showed up-regulation
267 compared to the PTZ group (Fig 3E). *COX2,* which is responsible for prostaglandin production [38],
268 increases during seizures [39]. While *COX2* inhibitors have been explored as therapeutic agents;
269 controversies exist due to types of inhibitors and timing of administration [39,40]

270 Studies in rats and zebrafish suggest that *cox1* may play a more crucial role in inhibiting seizure [41–
271 43]. Barbalho et al. found that inhibiting *cox-1* had a positive impact on seizure in zebrafish larvae,
272 while *cox-2* inhibition did not affect seizures [25].

273 Our results align with these findings. CX2 decreased the levels of *cox1* transcript (Fig 3D) and reduced
274 the occurrence of seizures by increasing latency and decreasing their frequency (Fig 1B). The
275 compound was quickly absorbed and distributed, as it was detected in the brain at the earliest time
276 point examined (Fig 4), and also in the optic tectum, a region associated with significant neuronal
277 activity during seizures [44].

278 As for the Tripeptide compound (Fig 2), we observed various gene responses, particularly the down-
279 regulation of genes related to inflammation. Neuroinflammation, triggered by factors such as tissue
280 damage, infection, stress, and seizures, has been associated with epilepsy [28,45]. Pathways
281 connecting epilepsy to neuroinflammation have been identified, with animal models of seizure
282 induction showing increased mRNA levels of genes involved in inflammatory cascades [46–48]. Thus,
283 efforts have focused on low molecular weight compounds acting on these pathways. Tripeptide (10
284 µg/mL) downregulated *il1b* gene and *casp9* gene indicating its positive effect on these pathways.

285 Regarding the bio-distribution of the Tripeptide compound, it was primarily detected in the zebrafish
286 larvae intestine (S1 Fig). This could be attributed to the fluorophore, tetramethylrhodamine-TMR,
287 which along with the compound, may interfere with absorption or crossing the blood-brain barrier for
288 CNS delivery.

289 Conclusion

290 Our study provides evidence that Tripeptide and CX2 – Cx43 peptide have the potential to modulate
291 gene expression and reduce epileptic seizures in zebrafish. Tripeptide increased the expression of genes
292 associated with antioxidant functions while decreasing the expression of inflammatory genes, with no

293 observed behavioral changes. The compound CX2 peptide demonstrated high effectiveness in reducing
294 the expression of inflammatory genes, lowering the number of seizures, and increasing latency time.
295 CX2 was widely distributed in the zebrafish brain and the body. These compounds could represent a
296 promising avenue for further research in the development of novel anti-seizure medications.
297 Nevertheless, further studies are needed to assess their effects in other animal models and determine
298 their efficacy in humans.
299

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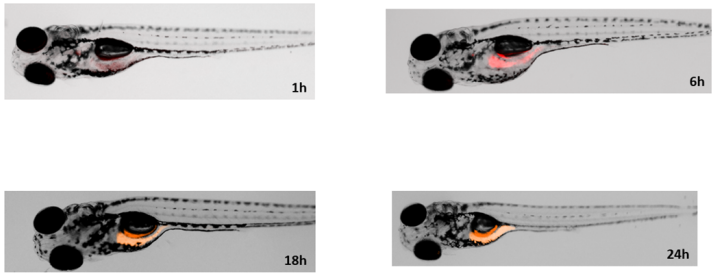
Supporting information



S1 Fig. Time course of the tripeptide in a 7-days post fertilization zebrafish larva. The tetramethylrhodamine-TMR labeled peptide was not detected in the brain through fluorescence microscopy.

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Competing interests

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.