Nanopore-based analysis unravels the genetic landscape and phylogenetic placement of human-infecting *Trichuris* species in Côte d'Ivoire, Tanzania, Uganda, and Laos

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Abstract

Soil-transmitted helminthiases (STH), including trichuriasis, pose a significant global burden, affecting numerous mammalian hosts. Traditional diagnostic methods have struggled to differentiate species within the *Trichuris* genus, prompting the use of molecular techniques to understand the diversity and zoonotic transmission of *Trichuris* spp. among humans and non-human primates (NHP), dogs and pigs. In this study, we used nanopore-based full-length ITS2 rDNA sequencing to genetically characterize and confirm the phylogenetic placement of the newly identified human-infecting *Trichuris hominis* and the well-known *Trichuris trichiura*. Using 670 samples from clinical studies in Côte d'Ivoire, Laos, Tanzania, and Uganda, along with publicly available ITS2 sequences, we confirmed two distinct clusters of human-infecting

Trichuris species. We found a varying prevalence of *T. hominis* and *T. trichiura* across the four countries and that these were also present in NHP. Additionally, we demonstrated that ITS2 fragment length accurately differentiates both human-infecting *Trichuris* species and thus makes it a good diagnostic marker for future point-of-care applications in resource-constrained settings.

Introduction

Trichuriasis, an infection caused by the whipworm *Trichuris sp*. is of considerable public health significance and is considered a Neglected Tropical Disease (1). This particular NTD has been estimated to disproportionately infect over 465 million people worldwide, mainly prevalent among children living in impoverished communities with limited hygiene or sanitation facilities, leading to a significant burden of 0.64 million disability-adjusted life years (DALYs) lost annually (2). However, several other mammals such as non-human primates (NHP), rodents, ruminants, and marsupials to name a few encompass the more than 20 specific taxonomic groups of hosts associated specifically with the genus *Trichuris,* meaning they are multi-host parasites (3,4).

Previous research approaches using traditional diagnostic methods like Kato-Katz have always regarded *Trichuris sp*. found in humans and NHP as *Trichiura trichiura* except in host-specific cases like the NHP Colobus monkeys (*Trichuris colobae*) (5) and Chacma baboons (*Trichuris ursinus*) (6) where there is a partial understanding of the *Trichuris* multi-host complexity. Morphologically, both eggs and adult worms of *T. trichiura* from humans and NHP are indistinguishable making molecular methods a promising approach (7–10). Recent efforts employed mostly sequences from the internal transcribed spacer (ITS) regions 1 and 2 (ribosomal DNA), mitochondrial markers and β tubulin gene employed to delineate this cryptic difference, hence resolving the *Trichuris* complex or multi-host specificity (5,9–15). These sequences were used to prove the existence of two separate *Trichuris* species in humans with the consensus being that the two separate *Trichuris* species were *T. trichiura* (6,9–15). Furthermore, a recent study in Côte d'Ivoire revealed a novel *Trichuris* species, called *T.*

hominis, that was observed in a monophyletic cluster with *T.suis* and *T. colobae* (16). However, ITS sequences corresponding to this species have been observed in single human patients in Uganda, and Cameroon, as well as several NHP (14). This sequence has also been shown to be a reliable marker to distinguish between *T. suis* isolated from pigs, *T. hominis* isolated from humans, and *T. trichiura* isolated from humans and non-human primates (16,17).

In this study, we analyzed the full ITS2 region (~500-600bp) from whipworms in stool samples collected from a total of 670 patients from clinical studies in Côte d'Ivoire, Tanzania, Uganda, and Laos. These samples were collected as part of randomized controlled trials (18,19) aiming at testing the efficacy of anthelmintic treatments. We next integrated the dataset generated from these studies with all publicly available and complete ITS2 sequences for the *Trichuris* genus, including sequences derived from humans, non-human primates, pigs, livestock, and rodents from multiple locations. Subsequently, we investigated the genetic makeup and phylogenetic relationships within our study populations and in a broader context. Finally, we identified a potential PCR-based diagnostic marker using the ITS2 fragment length that effectively differentiates between between the two known human-infecting species, *Trichuris trichiura* and *Trichuris hominis*. This method demonstrates a simplified, cost-effective, and robust alternative to expensive sequencing approaches and is especially suitable for resource-limited settings.

Results

Description of the study population

The population studied consisted of patients from the Pujini and Dodo shehias on Pemba Island, Tanzania; the Kisoro and Kabale districts in Uganda; the Dabou and Jacqueville regions around the Lagunes district in Côte d'Ivoire; and the Nambak district in Laos (**Figure 1**). This study included frozen faecal samples from participants aged 6 to 60 years in Côte

d'Ivoire, Tanzania and Laos, as well as ethanol-preserved faecal samples from school-aged children (6-12 years) in Uganda.

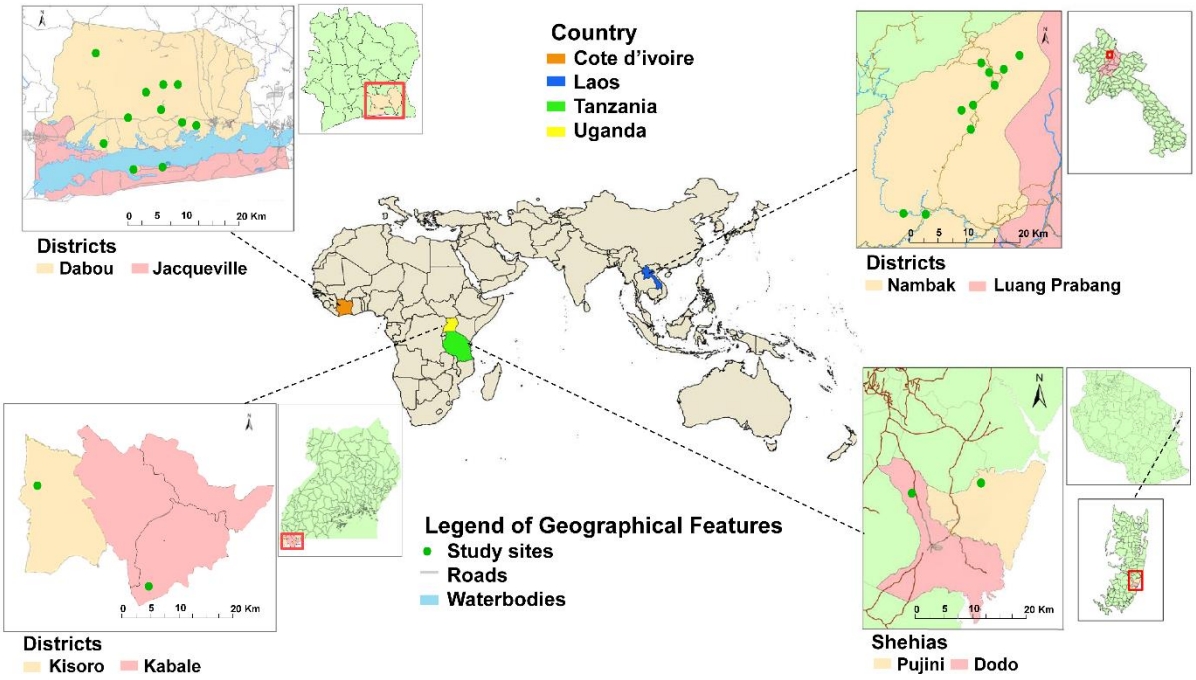

Figure 1: Sampling locations of the study population. The map of the world highlights the four different study locations: the Lagunes region in Côte d'Ivoire, Pujini and Dodo Shehias on Pemba Island in Tanzania, Nambak districts in Laos PDR (adapted from Keller et al. (20)), and the Kabale and Kisoro districts in Uganda each depicted in varying colours on the map.

Stool samples used in this study were collected in the framework of a previous double-blind, placebo-controlled randomized trial conducted between September 2018 and June 2020 in Côte d'Ivoire, Tanzania, and Laos (18,21,22) as well as a parallel open-label randomized controlled superiority trial conducted between October to November 2023 in Uganda (registered as NCT06037876; clinicaltrials.gov/). The metadata for the demographics and helminth infection characteristics of the 670 study participants are shown in **Table 1**. The infection intensities as diagnosed with Kato-Katz (KK) and sex distribution of the study participants depict a good balance between treatment groups. Of the 670 participants, 54.6% (336/670) were female and 45.4% (304/670) were male across the four countries. Overall, the Laos participants were older (mean age 23·1 years [SD 15·2]) than the other three cohorts (mean age 16·5 years [13·0] in Côte d'Ivoire, 14·1 years [9.8] in Tanzania, and 15.8 years [12.6] in Uganda). Samples of all 670 participants were included in PCR-based investigations of the internal transcribed spacer 2 (ITS2) region.

Table 1 Description of the study population

	Côte d'Ivoire (n=189)	Laos (n=156)	Tanzania (n=190)	Uganda (n=135)
Age in Years				
6 -12	116 (61.4%)	71 (45.5%)	119 (62.6%)	135 (100%)
13 – 24	32 (16.9%)	14 (9.0%)	49 (25.8%)	0
≥25	41 (21.7%)	71 (45.5%)	22 (11.6%)	0
Sex				
Female	96 (50.8%)	84 (53.8%)	107 (56.4%)	88 (65.2%)
Male	93 (49.2%)	72 (46.2%)	83 (43.6%)	47 (34.8%)
Infection intensity[+]				
Light	144 (76.2%)	136 (87.2%)	144 (75.8%)	118 (87.4%)
Moderate	42 (22.2%)	20 (12.8)	43 (22.6%)	17 (12.6%)
Heavy	3 (1.6%)	0	3 (1.6%)	0

[+] Infection intensity was classified according to mean EPG into light (1–999 EPG), moderate (1000–9999 EPG), and heavy (≥10 000 EPG).

Molecular Analysis of ITS2 Nuclear Marker

Following DNA extraction of the faecal samples from the patients partaking in the clinical trials across the four countries, we successfully produced amplicons from 188, 154, 174 and 113 participants in Tanzania, Laos, Côte d'Ivoire, and Uganda, respectively. The resulting ITS2 amplicons were sequenced using a MinION platform to achieve 5 to 9896 unprocessed reads per sample. Using a stringent DADA2-based pipeline, we retained a total of 198,792 reads (ranging between 5 to 9420 high-quality reads per sample) that were processed into 474 unique amplicon sequence variants (ASVs) (**Supplementary Figure 1**).

Population Differentiation and Network Analysis

Analyses of haplotypes showed the presence of 233 haplotypes of the *Trichuris* population across the four different countries (**Supplementary Table 4**). Ten haplotypes were shared

among the ASVs from the different countries, with four haplotypes shared within the four countries, while 223 haplotypes were revealed across all settings (51, 63, 74 and 35 from Côte d'Ivoire, Laos, Tanzania and Uganda, respectively). A few of the haplotypes analyzed showed high frequencies, such as Hap 1 in Laos (19%), Hap 2 in Côte d'Ivoire (31%), Hap 3 in Tanzania (14%) and Hap4 in Laos and Tanzania (18% and 24%), although these haplotypes were also present in varying proportions across the other countries (**Supplementary Table 4**). Comparison of these observed haplotypes across the four countries through NCBI BLAST showed that five haplotypes showing 100% identity with already described haplotypes: Hap1, Hap3 and Hap 4 were identical to *T. trichiura* isolated from samples analyzed from Ugandan patients (Accession number KJ588133 and KJ588134 respectively, (9)); Hap2 and Hap9 identified as *T. trichiura* isolated in worms expelled from patients in Cameroon (Accession number GQ301555, (14)). Also, the samples analyzed in Hap2 and Hap9 had a 96% to 100% identity match to ITS2 sequences from the newly described *T. hominis* species (17) following NCBI BLAST to its sequence. The comprehensive list encompassing the BLAST results is available as supplementary material (**Supplementary Table 5**).

Table 2: Genetic diversity estimations of *Trichuris* population per region

Country	N	N_H	Gene diversity	π	H_e
Côte d'Ivoire	189	58	0.9889±0.005	0.236±0.113	0.20
Laos	156	70	0.9951±0.003	0.153±0.073	0.13
Tanzania	190	83	0.9951±0.016	0.143±0.069	0.13
Uganda	135	42	0.9929±0.050	0.173±0.083	0.16

N = Number of samples sequenced; N_H = Number of haplotypes; π = Nucleotide diversity ; H_e = Expected heterozygosity

The analysis of genetic diversity indexes revealed that gene diversity within the populations ranged from 0.9889 to 0.9951, with the highest levels in Laos and Tanzania (both 0.9951), and the lowest in Côte d'Ivoire (0.9889). Conversely, nucleotide diversity (π) and expected

heterozygosity (H_e) showed a different pattern, with the sample population from Côte d'Ivoire and Uganda exhibiting higher diversity. 58 haplotypes from Côte d'Ivoire had the highest nucleotide diversity (π = 0.236±0.113) and expected heterozygosity (H_e = 0.20), followed by Uganda (π = 0.173±0.083, H_e = 0.16). In contrast, 70 haplotypes from Laos and Tanzania had lower nucleotide diversity (π = 0.153±0.0730 and 0.143±0.069 respectively) and expected heterozygosity (H_e = 0.13 for both), indicating fewer nucleotide differences between sequences in these populations. This suggests that while 153 haplotypes from Laos and Tanzania have extensive allelic variation, the alleles are more similar, whereas 100 haplotypes from Côte d'Ivoire and Uganda have more distinct variations at the nucleotide level (**Table 2**).

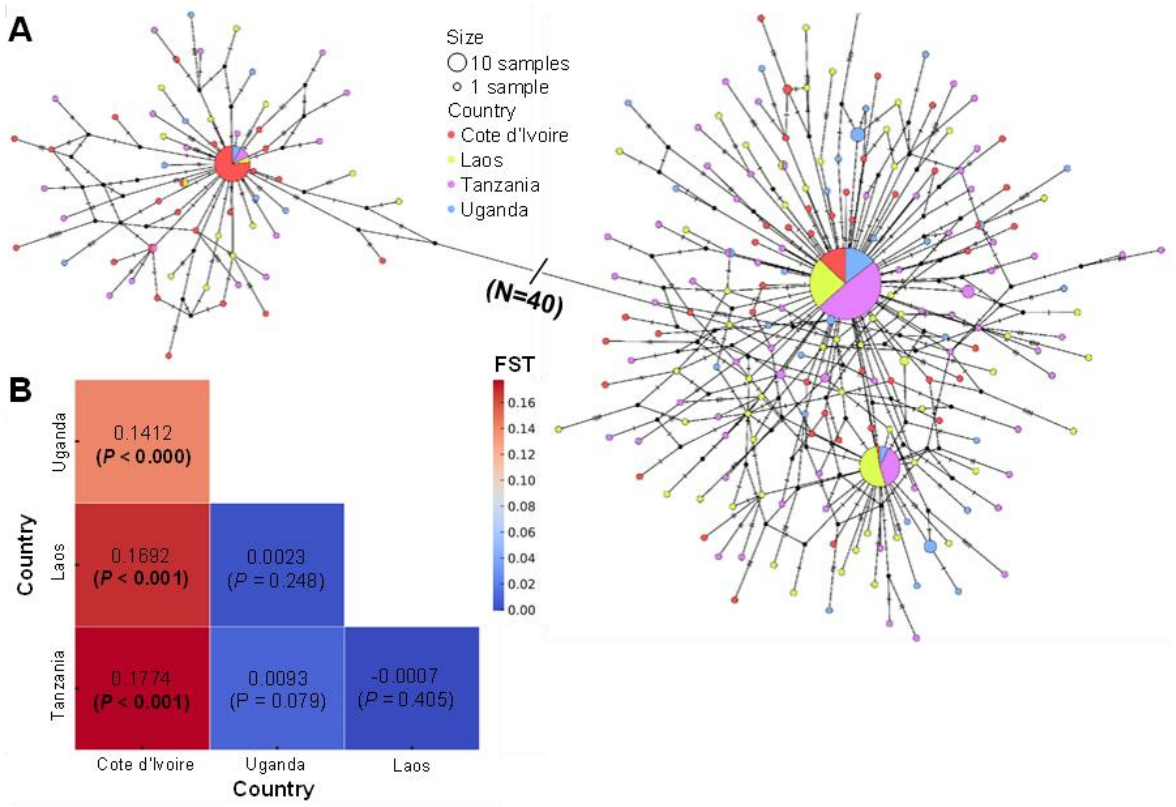

Figure 2: Underlying population genetic structure and network analysis among the four countries A. Pairwise comparison (FST) among the *Trichuris* population in different countries. **B.** Haplotype network inferred from ASVs of *Trichuris* species haplotype sequences from Côte d'Ivoire, Laos, Tanzania and Uganda. Each circle represents a unique haplotype and the size of the circle is proportional to the corresponding haplotype frequency. The colours of the pie charts represent the country of origin (red, Côte d'Ivoire; yellow, Laos; purple, Tanzania; blue, Uganda). Hatch marks along the network branches indicate the number of nucleotide differences between adjacent haplotypes and N = Number of point mutations or nucleotide difference.

Analysis of the haplotype network analyses revealed a reticulate relationship among the ASVs in the four countries scattered in separate clusters (**Figure 2A**). Specifically, the ASVs present in *Trichuris* populations of Côte d'Ivoire, Laos, Tanzania and Uganda were observed in the two clusters separated by 40 point mutations or nucleotide differences. By comparison, a further internal subdivision among haplotypes with a higher proportion was recorded in Tanzania than in the other three countries while the other sub-cluster revealed a higher proportion of haplotypes from Laos study participants than in the other three countries. The remaining haplotypes were observed in a different cluster, which represented a greater proportion of the *Trichuris* population in Côte d'Ivoire than those found in the Tanzania, Laos and Uganda populations.

Comparison of the pairwise FST values between the sampled populations revealed a range from -0.0007 to 0.1774 (**Figure 2B**). ASVs of the sampled population from Côte d'Ivoire were highly differentiated from those in Tanzania, Laos and Uganda with FSTs of 0.1774, 0.1692 and 0.14112 respectively ($P < 0.001$ for all FST values). Consequently, ASVs between Laos and Uganda, Laos and Pemba, as well as Uganda and Tanzania represented an undifferentiated population ($FST < 0.005$, $P > 0.001$).

Phylogenetic and comparative sequence analysis

Phylogenetic analysis of the ITS2 ASVs from the four countries together with publicly available ITS2 sequences of other related species was inferred using NJ, and Maximum Likelihood algorithms and their corresponding tree topologies provided a consistent, robust phylogenetic discrimination (**Figure 3A**). We identified five clades, each corresponding to different but taxonomically related host species and broadly matching published whole genome sequences (WGS) of *Trichuris* species. All the pairwise nucleotide identities between sequences generated in this study and publicly available sequences are summarized in **Figure 3B**. Clade A consisted of published ITS2 sequences of *Trichuris* species of Indian bison, camel, deer and sheep but did not contain any well-characterized WGS of a specific *Trichuris* species.

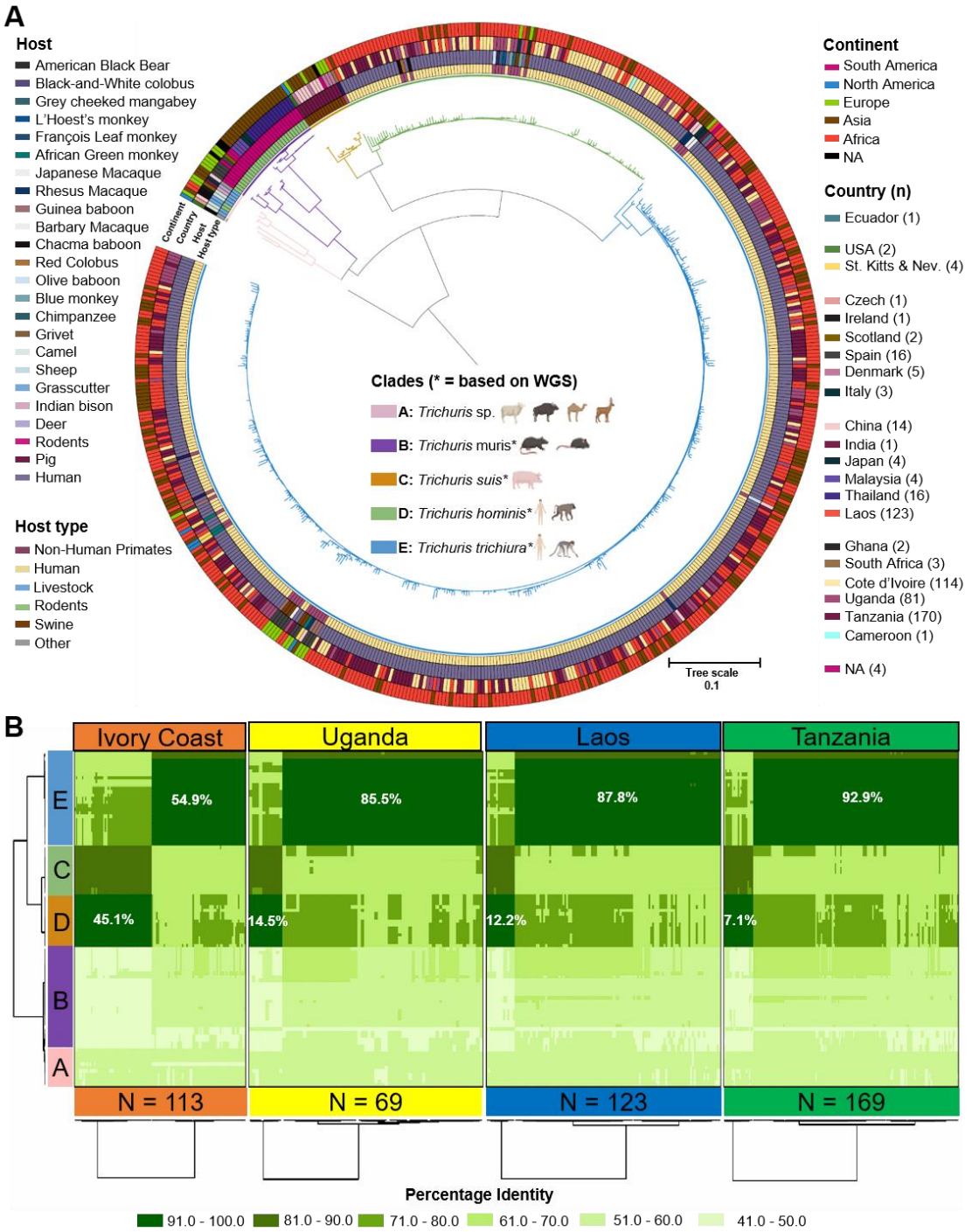

Figure 3: Phylogenetic and sequence characterization of *Trichuris* population A. Maximum likelihood tree based on the ITS2 rDNA using Tamura-Nei with gamma distribution as the substitution model and *Trichinella spiralis* (GenBank accession: KC006432) as an outgroup. Coloured branches correspond to major clades A to E. External rings (from outer to inner one) represent metadata from the samples (as well as other *Trichuris* reference sequences available from rodents, livestocks, pigs, humans, and non-human primates), namely continent, country of origin, host species and host-type respectively. **B.** Pairwise comparison of sequence similarity between samples from the four countries and reference sequences incorporated in the phylogenetic tree using ward clustering method and Euclidean correlation on OriginPro 2021 graphing software.

Clade B consisted of published sequences of *Trichuris* species of arvicolids and murid rodents – and included the sequence of an ITS2 region from a previously published WGS of *T. muris* (23,24). Clade C consisted of published sequences of *Trichuris* species of pigs which correctly matched the WGS of *T. suis* (25). Clade D was distinctly differentiated from *T. suis* sequences with nucleotide similarities ranging from 82 to 88% and contains ITS2 sequences from 20.5% (N = 97 samples) included in this study, as well as 14 publicly available sequences of human and NHP species. Sequences from clade D shared approximately 98% - 100% nucleotide sequence identity with the WGS sequence of a novel species of human-infecting *Trichuris*, named *T. hominis*, observed in Côte d'Ivoire (17), as well as other published partial sequences from humans in Cameroon (accession number GQ301555) (14), non-human primates (NHP) from Italy and Uganda, and more distantly with Chacma baboons in South Africa (accession number GQ301554) (14) with 90% to 94% nucleotide sequence identity. It is noteworthy that over 52.6% of samples contained in this clade were from Côte d'Ivoire (N = 51) but it also contained samples from other countries, Laos (N = 15 samples), Tanzania (N = 21 samples), and Uganda (N = 10 samples). Clade E contained 377 *Trichuris* sequences derived from our study as well as 36 ITS2 reference sequences of human and NHP species and 1 WGS sequence corresponding to the most common human-infecting species, *T. trichiura*. Clade E shared 96% - 100% nucleotide sequence identity to published sequences from humans in Ecuador and Uganda (9,23), as well as sequences from captive and non-captive NHP from Uganda, China, South Africa, Italy, Spain, and St. Kitts & Nevis (10,11,14,26,27). It showed a more distant nucleotide sequence identity (79% to 86%) with sequences obtained from Francois Leaf monkeys in China (26). The majority of sequences clustering in this clade were from Tanzania (N = 148), Laos (N =108), and Uganda (N = 59), but also contained approximately 16.4% (N = 62) of sequences generated from samples collected in Côte d'Ivoire. Furthermore, clade E was also more distant from clades C and D with nucleotide identity values ranging from 64% to 69%.

In summary, the phylogenetic analysis of the ITS2 fragment revealed that the *Trichuris* sequences – both, publicly available sequences and those generated during this study - form distinct clusters which are found to be strongly associated with groups of taxonomically-related host species. In addition, we observed the presence of a country-specific ratio of the human-infecting *Trichuris* species, *T. hominis* and *T. trichiura*, and that both human-infecting species were also found in other non-human primates across the globe.

Fragment length as a robust diagnostic marker to differentiate human-infecting *Trichuris* species

The fragment lengths - in basepair (bp) - of the ITS2 regions corresponding to the clade structure (clades A to E) are summarized in **Figure 4**. The length of the ITS2 regions ranged from 424.9±47.6bp for *T. muris*-related sequences, 443.7±7.9bp for *Trichuris* species found in livestock, 579.9±2.4bp for *T. suis*-related sequences, 603.7±13.5bp for *T. hominis*-related sequences, and 539.1±30.4bp for *T. trichiura* related sequences. We observed significant

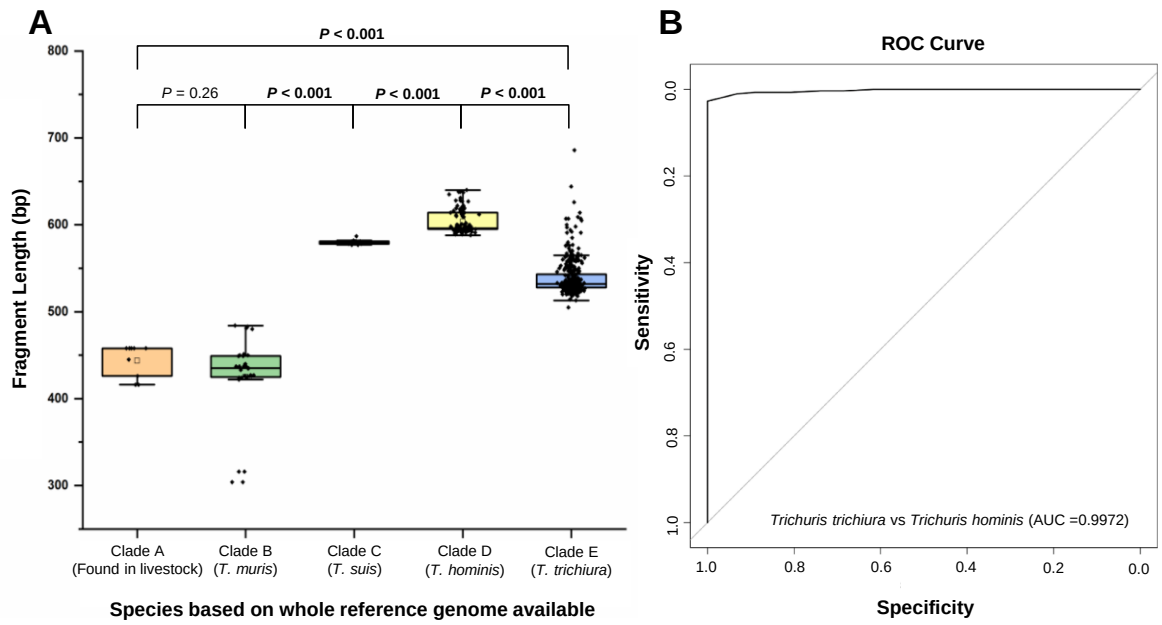

Figure 4: Fragment length could distinguish between different human-infecting *Trichuris* species. **A** Comparison of the fragment length between the five clades. Box plots include the median line, p-value estimated by the one-tailed Mann–Whitney U test. **B** ROC curve showing the performance of random-forest models built by combining the 2 signatures features of fragment length using the lose-1-keep1- m. bp: base pair.

differences in fragment length between all clades, except between *T. muris* and the livestock-related clade A.

We then evaluated the predictive power of the ITS2 region length to discriminate between the two human-infecting species, *T. hominis* and *T. trichiura*. For this purpose, we used a random forest model on our dataset as shown in **Figure 4B**. The model exhibited comparable classification performance with an area under the receiver operating curve of this model of 99.7% (95% CI: 99.4-100). The fragment length-based random forest model accurately classified 95.4% (Kappa = 85.9%) of samples into their respective *Trichuris* species. Taken together, our results showed that the fragment length is an accurate diagnostic marker to differentiate between the two human-infecting *Trichuris* species.

Human-infecting *Trichuris* species and metadata associations

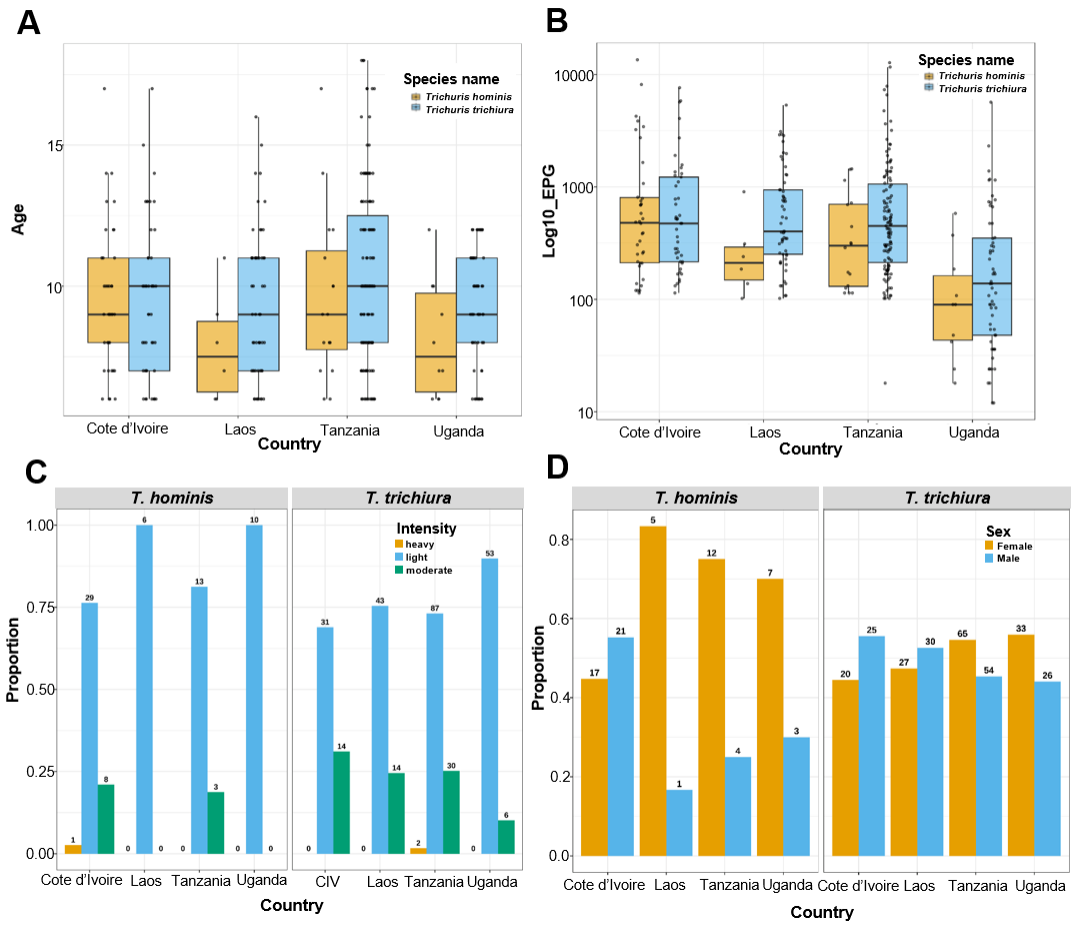

Figure 5: Distribution of the epidemiological metadata observed across the four countries in relation to human-infecting *Trichuris* species. **A** = age; **B** = log10EPG; **C** = infection intensity; and **D** = sex.

By stratifying our epidemiological data of study participants between the ages of 6-18 years, we sought to understand whether there were differences in the metadata associated with both, *T. hominis* and *T. trichiura*. We did not find any association between age and *Trichuris* species in any of the settings, revealing a homogeneous distribution of both species in the population traditionally most at risk, school-aged children (**Figure 5A**). Similarly, we did not find any differences in infection intensity (as a categorical variable defined by WHO) between both species (**Figure 5B/C**). Finally, we found that in 3 out of 4 settings, namely, Laos, Tanzania, and Uganda, females were more likely to present *T. hominis* than males indicating a potential sex-specific transmission route (**Figure 5D**).

Discussion

The etiological agent of human trichuriasis has traditionally been identified as *T. trichiura* based on morphological characteristics. However, morphological traits alone are often insufficient to distinguish closely related species, especially when adaptive changes or when species divergence has occurred relatively recently. Molecular epidemiological investigations using coding and non-coding regions of nuclear ribosomal DNA have revealed a second *Trichuris* species infecting humans, closely resembling those found in non-human primates (NHP) and pigs (14,16,17). Here, we utilized the nuclear ITS2 region to molecularly identify *Trichuris* species infecting humans from samples from randomized control trials across diverse geographical regions.

Our phylogenetic analysis of ASVs revealed that *Trichuris* populations from humans were grouped into two distinct *Trichuris* genotypes across these study sites, which we distinguish as Clade D and Clade E (**Figure 3A**). Haplotype networks, pairwise FST, and pairwise distance analysis (**Figure 2A-B**; **Figure 3B**) corroborated these results, indicating well-differentiated and genetically divergent clades across the study sites. These findings align with previous studies using ITS2 rDNA, demonstrating the multi-host affiliation of *Trichuris* species among humans and NHP (9–11,14–16,28,29).

Clade D contained majorly of the ITS2 ASVs from Côte d'Ivoire supporting results from recent studies (16,17). However, our study not only incorporates the enhanced resolution of full-length ITS2 sequencing but also significantly broadens the geographical scope including samples from Central Côte d'Ivoire and an entirely new country, Uganda, thereby enriching the diversity and depth of our data. To the best of our knowledge, this is the first time that *T. hominis* has been described from Tanzania, Laos, and Uganda—albeit in a smaller proportion—suggesting that this species has been infecting humans and bypassing standard microscopy-based diagnostic techniques across at least two continents with endemic trichuriasis for a significant amount of time.

Furthermore, Clade D shares a monophyletic lineage with the *T. suis* clade, with an estimated 82 to 88% nucleotide sequence similarity. Although both share similar physiological features, including overall morphology, egg production, and range of hosts, it suggests that *T. hominis* and *T. trichiura* evolved in parallel from a distant common ancestor.

In addition, we showed that Clade D also contains sequences from samples collected previously from 13 primate species spanning multiple studies and countries, as well as one human patient in Cameroon. With approximately 98-100% nucleotide identity of the ITS2 region among these sequences, it shows that *T. hominis* consists of a multi-host lineage suggesting that it is able – just like *T. trichiura* – to infect multiple hosts thus suggesting its potential zoonotic origin.

In contrast to Clade D, Clade E contained over 70% of the ASVs generated from our study. These sequences clustered closely with those derived from 36 human and NHP species, as well as one whole-genome sequence (WGS). Clade E sequences conformed to the most common human-infecting species, sharing 96% - 100% nucleotide similarity with sequences from humans in Ecuador and Uganda (9,23). Additionally, they matched sequences from both captive and non-captive NHP in Uganda, China, South Africa, Italy, Spain, and St. Kitts & Nevis (10,11,14,26,27). This result is in agreement with a recently published study using ITS2 rDNA (16) and further supports that whipworms in primates comprise a *Trichuris* species

complex. Additionally, within this clade, results based on ITS2 revealed an interpopulation similarity of approximately 79% to 86% between *T. trichiura* populations and *Trichuris sp.* from Francois Leaf monkeys, hence, agreeing with Liu et al. (26) that *Trichuris* populations from Francois Leaf monkeys potentially represent a new *Trichuris* species (**Figure 3A-B**). Although both *T. hominis* and *T. trichiura* share similar physiological features, including overall morphology, egg production, and range of hosts, there is every possibility that they both evolved in parallel from a distant common ancestor.

Despite earlier theories suggesting zoonotic transmission between pigs, humans, and NHP from an African origin to Asia and subsequently to South America (30,31), recent evidence suggests limited transmission influenced by natural host dispersion and anthropogenic factors in recent evolutionary history (30,32). Furthermore, comparative analyses of gene duplication, gain, and loss patterns in *T. hominis* compared to *T. suis* and *T. trichiura* indicate a more ancient and divergent lineage (17). All these observations collectively indicate a complex evolutionary history, resulting in a rich diversity of co-evolving species within the *Trichuris* genus. With reports of drug resistance emerging in the veterinary world (33–37) and expected resistance in human trichuriasis within the next decade (38), it is critical to understand the distribution and transmission of *Trichuris* etiologies. This understanding is essential to sustain the development of effective control strategies and achieve the WHO's 2030 goal of disease elimination as a public health problem (39).

To that end, while next-generation sequencing poses challenges for adopting species-level diagnostic tools in resource-constrained environments, we have identified a potential PCR-based diagnostic marker that can accurately differentiate between the two human-infecting *Trichuris* species, *T. hominis* and *T. trichiura*. This marker simplifies the detection process, eliminating the need for expensive sequencing platforms, and establishes a cost-effective, robust method for distinguishing these morphologically similar species. By using the length of ITS2 fragments, a random forest model validated its diagnostic utility, achieving a 95.4% classification accuracy (Kappa = 85.9%). This demonstrates the reliability of fragment length

as a marker in epidemiological studies and control programs, complementing previous findings on ITS2 rDNA's effectiveness in distinguishing closely related nematode species (11,13,16,28,40–42). The potential use of ITS2 length as a diagnostic marker is particularly suitable for resource-constrained environments and could be combined with PCR or further developed into LAMP assays. While MinION sequencing is relatively affordable and allows high multiplexing with our approach, a length-based diagnostic tool is simpler and more accessible for widespread application (43–46).

Our study has a few drawbacks. First, our observations are based on samples collected from clinical studies in Côte d'Ivoire, Laos, Tanzania, and Uganda, which may not fully capture the global diversity of the newly identified human-infecting *Trichuris* species due to the limited geographical scope. Second, our dependence on nuclear ribosomal DNA for species identification, without comprehensive morphological analysis or whole-genome sequencing, might miss other genetic variations and phenotypic traits. Furthermore, we did not evaluate temporal aspects of infections, such as seasonal variations or long-term changes, and our study did not include an extensive analysis of environmental factors that could influence the distribution and genetic diversity of *Trichuris* species. Importantly, the samples were collected within the context of clinical trials using the drug of choice, but we did not explore how genetic diversity might relate to decreased drug efficacy. While we believe our findings are robust, further studies with broader geographical coverage, and integrated genomic, morphological, and environmental approaches are needed to expand upon these findings.

In conclusion, our study utilized an enhanced resolution of full-length ITS2 rDNA sequencing to identify two distinct human-infecting *Trichuris* species, *T. hominis* and *T. trichiura*, across diverse regions. Our research expands the geographical scope of previous studies, including new regions and countries such as Uganda. We also identified *T. hominis* in Tanzania, and Laos, for the first time, reinvigorating the delimitations of traditional diagnostics in distinguishing between different *Trichuris* species. Additionally, we identified a PCR-based marker using ITS2 fragment length as a powerful diagnostic marker for distinguishing human-

infecting Trichuris species. This tool promises improved epidemiological studies and control programs, hence, aiding the WHO 2030 trichuriasis elimination goal. Future research should refine this diagnostic method and explore other genetic markers in conjunction with ITS2 fragment length to enhance diagnostic precision and provide deeper insights into the evolutionary relationships within the *Trichuris* genus.

Methods

Samples and ethics statement

The data and samples used in this study were derived from two studies. The first study was a double-blind, placebo-controlled randomized trial conducted between September 2018 and June 2020 in Côte d'Ivoire, Laos, and Tanzania. The study examined the efficacy and safety of co-administered albendazole and ivermectin versus albendazole monotherapy against whipworm infections in children and adults with the rationale, procedures, and results of the study published previously (18,21,22).

Briefly, a faecal sample was collected from community members aged between 6 and 60 years with *T. trichiura.* Participants were considered eligible who at baseline had at least 100 eggs per gram of stool. Participants were then randomly selected in a ratio of 1:1 to be administered either a single dose of ivermectin (200 µg/kg) plus albendazole (400 mg) or albendazole (400 mg) plus placebo, stratified by baseline *T trichiura* infection intensity. The egg reduction rate (ERR), and cure rate against *T trichiura*, were assessed 14–21 days post-treatment using Kato-Katz (KK) thick smears. From every sample, about 1g of faeces was transferred into a 2ml screw cap cryotube using a UV sterilized plastic spatula and immediately frozen at -20°C which were then shipped to Swiss TPH on dry ice at the end of the trial.

Additionally, data was also derived from a parallel open-label randomized controlled superiority trial conducted between October to November 2023 in two primary schools in Kabale district, southwestern Uganda among individuals aged 6-12 years to investigate the superiority of co-administered ivermectin plus albendazole to albendazole monotherapy in

terms of cure rates against *T. trichiura* infections (registered as NCT06037876; clinicaltrials.gov/). Briefly, two stool samples were collected at baseline and children found to be infected with *T. trichiura* based on quadruplicate Kato-Katz thick smear readings were enrolled into the study with efficacy of treatment determined at 14-21 days post-treatment by collecting another two stool samples. Following this, a portion of stool (1.5-2 g) from 135 participants found to be positive at both baseline and follow-up (14-21 days post-treatment) identified with *T. trichiura* infections were preserved in 95% ethanol, shipped to Swiss TPH in Basel and subjected to amplicon deep sequencing for characterization of *Trichuris* species. Faecal egg count and epidemiological data are available in **Supplementary Table 1**.

The trial received approval from independent ethics committees in Côte d'Ivoire (reference numbers 088–18/MSHP/CNESVS-km and ECCI00918), Laos (reference number 093/NECHR), Tanzania (Zanzibar, reference number ZAMREC/0003/Feb/2018), Uganda (reference number HS3160ES/UNCST; Division of Vector-borne and neglected tropical diseases VCDR-2023-29) and the institutional research commission of the Swiss TPH and the ethics committee of Switzerland (EKNZ: Ethics Committee of North-Western and Central Switzerland; O _2023-00066; reference number BASEC Nr Req-2018-00494). The trial protocols have also been registered as NCT03527732 and NCT06037876 on ClinicalTrials.gov. Participants gave informed consent to study participation and no compensation was offered in this regard.

DNA isolation and sample processing

Two separate extraction approaches were used in the isolation of DNA from stool received from the multi-country study and the study from Uganda. For the multi-country study, a total of 658 (226, 191, and 221 respectively from Côte d'Ivoire, Laos, and Tanzania) frozen 100 mg of stool samples from the baseline were extracted using a QiAmp PowerFecal DNA extraction kit (Qiagen, Hilden, Germany) according to the manufacturer's protocol with slight modification. For the faecal samples collected in Southern Uganda, 150 - 250mg of samples preserved in Ethanol was used in the extraction of the DNA. Before extraction with the QiAmp

PowerFecal DNA extraction kit, a few modification steps were taken to mechanically rupture the *Trichuris* egg shells which include the removal of ethanol from the stool, washing with phosphate buffer saline (PBS), as well as a freeze-thaw-boiling step to enhance egg shell rupture and minimize inhibition (16,47) (**see** Supplementary appendix 1 for more details of the modified ethanol-preserved sample extraction protocol).

PCR Amplification, Quantification and Purification

Primers used in the ITS2 rDNA reactions were Trichuris_ITS2_F 5'-ATGTCGACGCTACGCCTGTC-3' and Trichuris_ITS2_R 5'-TAGCCTCGTCTGATCTGAGG-3' (16) with a reverse primer containing an added custom barcodes that were adapted from the Illumina 12bp barcode sequences (Caporaso et al., 2012; Dommann et al., 2024) (See **Supplementary Table 2** for full primer sequences). For each run, 96-well plates were multiplexed with the Trichuris_ITS2_F primers and eight custom barcoded reverse primers to amplify the ITS2- regions yielding unique barcodes of 592 bp ITS2 regions for each sample per plate column. LongAmp® Hot Start Taq 2X Master Mix (cat no. M0533S, New England BioLabs, USA) and 2µL of source DNA were used in each amplification. Thermocycling conditions included the following: 94°C for 30 sec initially, 40 cycles of 94°C for 30 sec, 65°C for 15 sec, 65°C for 50 sec, followed by a final extension at 65°C for 10 min carried out on an Eppendorf Mastercycler Nexus Gradient Thermal Cycler (Eppendorf AG, Hamburg, Germany). Barcoded amplicons were quantified directly using a dsDNA HS Assay kit on a Qubit 4 Fluorometer (Invitrogen, USA) according to the manufacturer's protocol to ensure even throughput in the DNA pool using 2 µL of PCR reaction as input. 1% agarose and capillary gel electrophoresis (100V, 20min) were used to verify the amplification of the fragment at the expected band size. Afterwards, sample plates were stored overnight for a short period at 4°C overnight or at -20°C for long-term storage.

Library preparation and sequencing

Nanopore sequencing libraries were prepared using the Native Barcoding Kit 24 v14 (SQK-NBD114.24, Oxford Nanopore Technologies, UK) according to the Native Barcoding Genomic DNA protocol with slight modifications aiming to normalize sample input, allowing for multiplexing as described in more details elsewhere (48). Briefly, samples were pooled column-wise from a 96-well plate before the end-repair step, requiring 12 different outer barcodes for every 8 samples. Each column, representing 8 samples, was processed together with approximately 80 ng input corresponding to 200 fmol of the expected 0.59 kb fragments for the end-repair steps. Following end repair, samples were assessed using the 1X dsDNA HS Assay kit on a Qubit 4 fluorometer. Equimolar masses of the pooled reactions were used for native barcode ligation to ensure equal read numbers per barcode. Adapter ligation and clean-up followed the protocol, utilizing a short-fragment buffer to preserve the ITS2 rDNA fragments. Afterwards, 1 µl of the prepared library was quantified on the Qubit 4.0 (Invitrogen, USA) using the Qubit dsDNA HS Assay Kit (Invitrogen, USA). The protocol from ONT recommends loading 10-20 fmol of amplicon product onto the Flongle Flow Cell (Flongle Flow Cell, R10 Version, FLO-FLG114, Oxford Nanopore Technologies, UK), so we diluted to a concentration of 2-3 ng/µL in Elution Buffer to load approximately 15 fmol of the library onto the Flongle Flow Cell. The Flongle Flow Cell was primed for loading by flushing the flow cell with 120 µL priming mix (3 µl of Flush Tether in 117 µL of Flow Cell Flush), taking care to avoid the introduction of air bubbles. The library was prepared for loading by mixing 15 µl Sequencing Buffer, 10 µl Loading Beads and 5 µl diluted DNA library, after which the sample was added to a Flongle Flow Cell on a MinION Mk1C (Oxford Nanopore Technologies, UK) and sequenced at depth for 24 h

Read processing and sequencing analysis

All POD5 data resulting from the sequencing of the amplicons were already basecalled and demultiplexed based on the native ONT barcode in real-time on the MinKNow v3.6.5 and the built-in Guppy (v6.5.7) using the high accuracy basecalling model. The base-called POD5 files

were then re-run through Dorado (v. 0.5.1; base-caller model dna_r10.4.1_e8.2_400bps_sup@v4.3.0) generating high-quality simplex reads) which was fed through our custom demultiplexing scripts that combine seqkit (v.2.6.1) (49) and custom Python scripts available at https://github.com/STPHxBioinformatics/ITS2_Trichuriasis_nanopore_processing. The script demultiplexes, quality filters, and length filters, as well as utilizing a minimum and maximum, trimmed-read cutoff of 450 bp and 750 bp, respectively with zero mismatches. Consequently, it trims the reads based on the inner PCR barcode (the reverse barcode in this case) ensuring the barcodes are found in the correct orientation. NanoPlot v1.30.1 (50) was then used for descriptive statistics on the runs. Demultiplexed ITS2 reads were then subjected to the R package 'dada2' version 1.3.2 (51) pipeline to include further filtering to remove reads containing unresolved nucleotides (maxN = 0) as well as reads exceeding the expected error number (maxEE = 15) and size range (425–700 bp). The dataset generated was then used as input to define the error rates and perform the removal of identical reads (derepFastq) while inferring the composition of the sequenced pool using the dereplicated sequence dataset as input (dadaFs), and removal of chimeric sequences. Following this, Amplicon sequence Variants (ASVs) generated through the DADA2 pipeline with ASVs below 5 reads across all samples were removed.

Genetic Variation and Phylogenetic Analysis

A curated database of reference ITS2 sequences was created by downloading all publicly available full ITS1-5.8S-ITS2 reference sequences of closely related species on NCBI using the search term "Trichuris"[porgn:__txid36086] as well as from the WGS data of *Trichuris muris* (23,24), *Trichuris trichiura* (23), *Trichuris suis* (25) and an adult worm from Côte d'Ivoire (17). Afterwards, seqkit (49) was used to extract only ITS2 sequences within the forward and reverse primers used in this study with the command seqkit -j 4 amplicon –m 2 -p primers.tab -r 1:-1 20240201_Trichuris.txid_36086.fasta --bed > 20240201_Trichuris_txid_36086.bed. The ASVs were then subjected to multiple sequence alignments with the newly curated

reference sequences (**Supplementary Table 3**), as well as an outgroup species *Trichinella spiralis* (Accession No: KC006432), using the MAFFT tool (Katoh et al., 2019). Phylogenetic relatedness was inferred using Neighbour- Joining, NJ in MEGA v11.0 (52) and the Maximum Likelihood (ML) method using RaxML v8.0 (53) employing the jModelTest v0.1.1 (54) to choose the best-to-fit model.

Population Genetic Structure

Alignment from good quality ITS2 ASVs was used for population genetic data and haplotype polymorphic analysis such as numbers of variable sites, number of haplotypes, nucleotide diversity (π), expected heterozygosity, and Pairwise FST calculations between the *Trichuris* populations in the four countries were calculated using DnaSP v6.0 (55) and Arlequin v3.5.2 (56) using 1000 permutations. Statistical Parsimonious networks were inferred and visualized using Population Analysis with Reticulate Trees, POPART v1.7 (57) with the connection limit set to 95% and gaps being treated as missing. Furthermore, the number of nucleotide differences per sequence in each country and other *Trichuris* species incorporated in the phylogenetic analysis was analyzed using the Compute Pairwise Distance prompt based on the number of differences method of MEGA v11 (52) to assess the sequence similarity.

Data Accessibility

The custom demultiplexing script and DADA2 analysis code used for this study are available at: https://github.com/STPHxBioinformatics/HITS2. The raw ONT sequence data (ITS2 rDNA amplicon sequencing) generated in this study is available in the NCBI Short Read Archive under accession code PRJNA1141773.

Acknowledgements

We want to acknowledge the participants and investigators of the IVB-ALB and FACEITeffic trial. The authors also thank all the Keiser's lab members for their valuable discussions. Computational analyses were performed using the sciCORE (https://scicore.unibas.ch)

scientific computing center at the University of Basel. We are grateful to the European Research Council (No. 101019223) for financial support.

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