Homoploid Hybrid Speciation in a Marine Pelagic Fish *Megalaspis cordyla* (Carangidae)

NOZOMU MUTO[1,*], YONG-CHAO SU[2], HARUTAKA HATA[3], NGUYEN VAN QUAN[4], VEERA VILASRI[5], MAZLAN ABD. GHAFFAR[6] AND RICARDO P. BABARAN[7]

[1]Department of Marine Biosciences, Kitasato University, Sagamihara, Kanagawa 252-0373, Japan;

[2]Department of Biomedical Science and Environmental Biology, Kaohsiung Medical University, Kaohsiung City, 80708, Taiwan;

[3]National Museum of Natural History, Smithsonian Institution, Washington, DC 20560, USA;

[4]Institute of Marine Environment and Resources, Vietnam Academy of Science and Technology, Hai Phong City, Vietnam;

[5]Natural History Museum, National Science Museum, Pathum Thani 12120, Thailand;

[6]Institute of Climate Adaption and Marine Biotechnology, Universiti Malaysia Terengganu, Kuala Nerus 21030, Malaysia;

[7]College of Fisheries and Ocean Sciences, University of the Philippines Visayas, Miagao Iloilo 5023, Philippines

*Correspondence to be sent to: Department of Marine Biosciences, Kitasato University, Sagamihara, Kanagawa 252-0373, Japan; E-mail: muto.nozomu@kitasato-u.ac.jp / nzmutou@gmail.com.

Abstract.—Homoploid hybrid speciation (HHS) is an enigmatic evolutionary

process where new species arise through hybridization of divergent lineages

without changes in chromosome number. Although increasingly documented in

various taxa and ecosystems, convincing cases of HHS in marine fishes have

been lacking. This study presents evidence of HHS in Torpedo scad *Megalaspis*

cordyla based on comprehensive genomic, morphological, and ecological

analyses. A Principal Component Analysis using thousands of SNPs identified

three sympatric clusters in the western Pacific. Genome-wide differentiation

between the clusters and the admixed nature of a cluster between the others

were evident from population genomic analyses, species tree estimation,

mitochondrial DNA divergence, and tests of introgression. Multiple statistical

methods for hybrid detection also supported the admixed ancestry of this

cluster. Moreover, model-based demographic inference favored a hybrid

speciation scenario over introgression. Examination of occurrence data and

ecologically relevant morphological characters suggested ecological differences

between the clusters, potentially contributing to reproductive isolation and niche

partitioning in sympatry. The clusters are morphologically distinguishable and

thus can be taxonomically recognized as separate species. The hybrid cluster is

restricted to the coasts of Taiwan and Japan, where all three clusters coexist.

The parental clusters are additionally found in lower latitudes such as the coasts

of the Philippines, Vietnam, Thailand, and Malaysia, where they display non-

overlapping distributions. Given the geographical distributions, estimated times

of the species formation, and patterns of historical demographic changes, we

propose that the Pleistocene glacial cycles were the primary driver of HHS in

this system. Based on this argument, we develop an ecogeographic model of

HHS in marine coastal ecosystems, including a novel hypothesis to explain the

initial stages of HHS.

Key words: climate change; ecological divergence; hybridization; latitudinal

gradient; morphology; Pleistocene; phylogeography; reproductive isolation]

Running head: HYBRID SPECIATION IN A MARINE PELAGIC FISH

Hybridization can drive adaptation and species diversification. One possible

outcome of hybridization is hybrid speciation, where interbreeding between

diverged species results in a new population evolving independently from the

parental species (Coyne and Orr 2004; Mallet 2007; Peñalba et al. 2024). In

allopolyploid hybrid speciation, a hybrid population arises with whole-genome

duplication, reported frequently in plants (Wood et al. 2009). In contrast,

homoploid hybrid speciation (HHS) occurs without changes in ploidy, with the

nascent hybrid population possessing a recombinant genome of parental species

(Mallet 2007). There is much debate about the criteria for a case to be regarded

as HHS. An emerging view is that at least the first and second of the following

criteria must be satisfied, whereas a more stringent definition requires the third

as well: 1) the species arose via hybridization with detectable genomic signatures,

2) intrinsic or extrinsic reproductive isolation from parental species has been

established, and 3) the reproductive barriers directly resulting from hybridization

(Abbott et al. 2013; Schumer et al. 2014; Nieto Feliner et al. 2017; Long and

Rieseberg 2024). Cases of HHS fulfilling the first two criteria are increasingly

documented. However, it is still rare compared to other hybridization-related

phenomena, such as the formation of hybrid zones and introgression. Moreover,

reported cases are strongly biased towards terrestrial and freshwater organisms

(Abbott et al. 2013; Yakimowski and Rieseberg 2014; Taylor and Larson 2019

and references therein).

A major challenge in HHS is the persistence of the nascent hybrid population.

Immediately after formation, it faces ecological competition with parental species

and the swamping effects of gene flow (Buerkle et al. 2000). The hybrid

population can overcome both and persist on an evolutionary timescale by

acquiring a novel niche not occupied by the parental species. The acquisition of

a novel niche drives HHS in several ways. First, it may directly cause extrinsic

pre- or postzygotic reproductive isolation between the hybrid and parental

species (Rieseberg et al. 2003; Schwarz et al. 2005). Second, it enables the

hybrid and parental species to coexist in sympatry by alleviating competition

through niche partitioning (Lamichhaney et al. 2018; Olave et al. 2022). Third, it

may allow the hybrid species to occupy a spatially separated habitat from the

parental species, facilitating allopatric or parapatric divergence (Donovan et al.

2010; Brennan et al. 2012; Lopes et al. 2023). Supporting the importance of the

third role, the independence of the hybrid species in most compelling cases of

homoploid hybrid speciation seems to rely on geographical separation from at

least one parental species (Rieseberg et al. 2003; Mavárez and Linares 2008;

Larsen et al. 2010; Brennan et al. 2012; Kang et al. 2013; Lamichhaney et al.

2018; Maier et al. 2019; Noguerales and Ortego 2022; but see Olave et al. 2022

and Rosser et al. 2024 for counterexamples). Theory suggests that the

recombinant hybrid genome can express transgressive phenotypes via additive

and/or epistatic effects of parental genetic variants, leading to the exploitation of

novel niches (Rieseberg et al. 2003; Gross and Rieseberg 2005; Mallet 2007;

Abbott et al. 2013). However, in what ecogeographic settings the niche

divergence leads to reproductive or geographical isolation remains largely

unknown.

The Pleistocene glacial cycles significantly impacted the evolutionary histories

of organisms at and below the species level across various ecosystems and taxa.

In terrestrial ecosystems, ice sheets formed in high altitudes and latitudes during

glacial periods disrupted existing habitats, while glacial retreat during interglacial

periods created new, unoccupied habitats. Coupled with temperature changes,

these habitat dynamics resulted in allopatric divergence, secondary contact,

altitudinal and latitudinal range shifts, population size changes, and local

adaptation (Hewitt 2004). Mountain systems are among the most well-studied

(Hewitt 2000, 2004), including a few possible cases of homoploid hybrid

speciation (Gompert et al. 2006; Maier et al. 2019). Marine coastal areas are

another ecosystem strongly influenced by historical climate changes. Eustatic

sea level changes cyclically disrupted and opened habitats for marine organisms

in shallow waters, creating barriers or corridors to gene flow (Bowen et al. 2016).

Changes in water temperature and associated environmental factors caused

latitudinal range shifts, population size changes, and adaptive responses, similar

to those ongoing under contemporary climate change (Kiessling et al. 2012;

Neiva et al. 2014; Donelson et al. 2019). These, in turn, resulted in various

hybridization-related phenomena in coastal organisms via secondary contact:

introgression (Duranton et al. 2020), formation of stable hybrid zones (Hirase et

al. 2021), and divergence with gene flow (Muto and Kai 2023). Nonetheless, HHS

in this environment has not been documented to date.

Here, we provide genomic and phenotypic evidence for HHS in the torpedo

scad *Megalaspis cordyla* (Linnaeus, 1758) in the western Pacific, representing

the first compelling case of this phenomenon in a marine pelagic fish. The

torpedo scad is a pelagic marine predator in the family Carangidae (Teleostei),

usually reaching 40 cm in total length. Widely distributed in tropical to temperate

coastal waters throughout the Indian Ocean and the western Pacific, it is an

important fisheries resource in the surrounding countries (Smith-Vaniz 1999).

Previous phylogeographic studies based on mitochondrial DNA (mtDNA)

sequences suggested historical vicariance followed by secondary contact in the

western Pacific region. Specimens from the southwestern part of the region (the

Pacific coast of the Asian continent, from Vietnam to Malaysia) and the

southeastern part (the islands of the Philippines and Indonesia) represent

reciprocally monophyletic lineages, whereas specimens from the northern part

(Taiwan) represent both lineages (Su et al. 2020; Muto et al. 2021). In the

present study, we provided evidence for genome-wide divergence between the

two lineages. We also discovered a third genetic cluster restricted to locations

where the two lineages are sympatric. We tested the hypotheses that this third

cluster originated from the admixture of the other two and that they represent

separate species, using detailed genomic, ecological, and morphological

analyses. Based on the results, we suggest that the Pleistocene glacial cycles

primarily drove HHS in the present system. Furthermore, we develop an

ecogeographic model of HHS in coastal marine environments, where

demographic dynamics under climate change and adaptation across latitudinal

environmental gradients play critical roles. This model bridges a major gap in

the understanding of how the initial stages of HHS proceed.

151   **MATERIALS AND METHODS**

152   A full description of methods is provided in Online Appendix 1.

Specimens

We analyzed a total of 160 specimens of *M. cordyla* collected from various

localities in the western Pacific (Fig. 1a), where two distinct mtDNA lineages

have been discovered (Su et al. 2020; Muto et al. 2021). For genotyping, we

used tissues stored in natural history collections in respective countries (Table

S1). Total genomic DNA was extracted from the fin or muscle tissue of each

specimen using the Favorgen Tissue Genomic DNA Extraction Mini Kit

(Favorgen). The specimens from Vietnam, Thailand, Malaysia, and the

Philippines were analyzed in the previous study (Muto et al. 2021).

Fig. 1. a) Sampling sites (circles) and distributions of three genetic clusters (polygons) of
Megalaspis cordyla in the western Pacific. The grey-shaded area represents regions
currently shallower than 130 m, repeatedly exposed during Pleistocene glacial cycles. The
dotted arrow indicates the approximate route of the Kuroshio Current. b) Scatter plots for
the principal component analysis (PCA) based on the genome-wide SNPs, with polygons
and arrows indicating the three genetic clusters and their putative early-generation hybrids,
respectively. The clusters are named according to their geographical distributions. c) Result
of the STRUCTURE analysis assuming the number of clusters (*K*) = 2, which was the
optimal according to the delta *K* values. Horizontal bars and arrows indicate individual
assignments to the three clusters and putative early-generation hybrids, respectively, based
on the PCA. The color code for the horizontal bars matches that for the PCA plot polygons.
Note that individuals in the North cluster and putative early-generation hybrids show

admixed ancestry. d) Statistically parsimonious network of haplotypes of the concatenated
control region and cytochrome b gene sequences of the mitochondrial genome. Circle/pie-
chart size is proportional to haplotype frequency. Black dots and a hatch mark along
branches represent unobserved hypothetical haplotypes and one mutational step,
respectively.
180

181   *Mitochondrial DNA Genotyping*

We obtained partial sequences of the mitochondrial cytochrome b gene

(CYTB: 873 bp) from all specimens. Additionally, we obtained partial sequences

of the following seven loci from a subset of the specimens: cytochrome c

oxidase subunits 1 (COI: 612 bp) and 2 (COII: 657 bp), NADH dehydrogenase

subunits 1 (ND1: 828 bp), 2 (ND2: 531 bp), 4 (ND4: 828 bp), and 5 (ND5: 801

bp), and the control region (CR: 760 bp) (Table S1). We amplified the COI and

CYTB sequences following Ivanova et al. (2007) and Cárdenas et al. (2005),

respectively. The remaining loci were amplified using newly developed primers

(Table S2). We edited and aligned the sequences using BioEdit v7.0.5.3 (Hall

1999) and CLUSTAL X (Larkin et al. 2007).

193   *Genome-Wide SNPs Genotyping*

We genotyped 115 specimens using MIG-seq (Suyama and Matsuki 2015;

Suyama et al. 2022), a reduced representation sequencing method targeting

SNPs in the inter-simple sequence repeat region (ISSR). We performed the first

PCR using 12 multiplex primers to amplify various ISSR across the genome

(Suyama and Matsuki 2015; Suyama et al. 2022). The first PCR used 12

multiplex primers to amplify various ISSRs across the genome. The second

PCR added Illumina sequencing adapters and a unique dual index. We pooled

the second PCR products, size-selected for 400–800 bp, purified, quality-

checked, quantified, and sequenced on an Illumina Novaseq 6000 in 150 bp

paired-end mode at Novogen Co. Ltd., China, yielding 148M reads.

We processed the raw MIG-seq reads by the single-end mode of fastp

v0.20.1 (Chen et al. 2018), merging the processed reads 1 and 2 into a single

FASTQ file, and mapped them to the *Caranx ignobilis* reference genome

(GenBank JAFHLA000000000.1) (Pickett et al. 2022) using BWA-mem v0.7.17

(Li 2013) with default settings. Unmapped or low-quality reads were removed

using Samtools v1.12 (Danecek et al. 2021). The resulting BAM files were used

to prepare 5 datasets for downstream analyses (see the next paragraph). We

also downloaded the whole-genome shotgun reads of *C. ignobilis* (a different

individual from the reference) and *Caranx melampygus* from the NCBI Short

Read Archive (accession nos.SRR1002875 and SRR1002876) (Santos 2014)

as outgroups. We processed these reads using the paired-end mode of fastp,

mapped them to the reference genome using BWA-mem with default settings,

and removed unmapped, low-quality, or duplicate reads using Samtools. The

resulting BAM files were used for the analyses requiring outgroups (datasets 2–

5: see below).

The following 5 datasets were prepared for downstream analyses: 1) 2,926

unlinked biallelic SNPs in all available (105) specimens of *M. cordyla* without an

outgroup for population genetic analyses using various tools and ancestry

estimation using HIest v2.0 (Fitzpatrick 2012); 2) 6,789 unlinked biallelic SNPs

in 24 specimens (8 per genetic cluster) with *C. ignobilis* as an outgroup for

admixture detection using HyDe v1.0.0 (Blischak et al. 2018) and

ADMIXTOOLS v7.0.2 (Patterson et al. 2012); 3) 3,179 unlinked biallelic SNPs in

9 specimens (3 per genetic cluster) with *C. ignobilis* as an outgroup for

divergence time estimation using SNAPP v1.5.0 (Bryant et al. 2012); 4) 6,341

biallelic SNPs in the same 24 specimens, polarized using the *C. ignobilis* and *C.*

melampygus genomes as outgroups, for demographic model selection using

fastsimcoal v2.7.0.9 (Excoffier et al. 2013, 2021); and 5) 22,326 biallelic SNPs

(linked and unlinked) and 920,184 monomorphic sites in the same 24

specimens, polarized using the *C. ignobilis* and *C. melampygus* genomes as

outgroups, for demographic parameter estimation using fastsimcoal 2 and

historical population size estimation using Stairway Plot v2.1.2 (Liu and Fu

2020). Note that monomorphic sites and linked SNPs in the 5th dataset are

needed for direct parameter estimations using fastsimcoal 2, whereas only

unlinked SNPs are used for the model selection (Excoffier et al. 2013). For each

dataset, we called SNPs using Stacks v2.5.4 (Catchen et al. 2013) or *mpileup*

implemented in bcftools v1.8 (Danecek et al. 2021). We then filtered the SNPs

and created final vcf files using VCFtools v0.1.16 (Danecek et al. 2011) and R

package 'SNPfiltR' v1.0.0 (DeRaad 2022) on R v4.2.2 (R Core Team 2022).

We used *easySFS.py* (https://github.com/isaacovercast/easySFS) to

calculate the unfolded site frequency spectrum (SFS) from the 4th and 5th

datasets, assuming the reference alleles fixed in both outgroups represent the

ancestral state. The resulting one-dimensional SFS from the 5th dataset was

used for the historical population size estimation by Stairway Plot 2, whereas

multi-dimensional SFS was used for the demographic inference by fastsimcoal

2. For input file preparations for the other analyses, we used PGDSpider

v2.1.1.2 (Lischer and Excoffier 2012) to convert the vcf files to appropriate

formats.

252   *Population Genetic Analyses*

We constructed minimum-spanning haplotype networks of mtDNA sequences

using PopART v1.7 (Leigh and Bryant 2015).

The following population genetic analyses were conducted using 1st dataset

of the genome-wide SNPs. We performed a principal component analysis

(PCA) using the 'adegenet' package v2.1.4 in R (Jombart 2008). We used

STRUCTURE v2.3.4 (Pritchard et al. 2000) to identify admixed individuals with

10 replicate runs for each predefined number of genetic clusters (K) = 1–8,

each with 100,000 Markov Chain Monte Carlo (MCMC) generations after

100,000 generations of burn-in. These runs were parallelized using StrAuto v1.0

(Chhatre and Emerson 2017). Statistical support for K values was evaluated by

calculating the ΔK value (Evanno et al. 2005) using Structure Harvester v0.6.94

(Earl and vonHoldt 2012). The results were summarized and visualized with

'pophelper' v2.3.1 in R (Francis 2017). We used SplitsTree v5.3.0 (Huson and

Bryant 2006) to build an unrooted phylogenetic network based on the Neighbor-

Net algorithm.

The population genetic analyses showed one genetic cluster was genetically

intermediate to the other two clusters (Fig. 1b, c). This intermediate cluster is

hereafter called the "North" cluster, while the others are the "East" and "West"

clusters, according to their geographical distributions (Fig. 1a). Putative early-

generation hybrids (F1, F2, or backcross) between these clusters were also

detected. Excluding these putative hybrids, we calculated locus-specific

pairwise F_{ST} between the clusters using *populations* implemented in Stacks,

and pairwise F_{ST} across all loci and associated *p*-values using Arlequin v3.5.2.2

(Excoffier and Lischer 2010).

We estimated historical population size changes in each cluster using

Stairway Plot v2.1.2 (Liu and Fu 2020) based on the 5th dataset. We assumed

a mutation rate of $1E^{-8}$ per site per generation (Tine et al. 2014) and a

generation time of 1.2 years (Froese and Pauly 2023). Other settings were

default or those recommended by the author of the software.

283   *Divergence Time Estimation*

We estimated the divergence times of the three genetic clusters using

genome-wide SNP dataset 3 and mtDNA sequence data. The former was done

under the multispecies coalescent model using the SNAPP v1.5.0 package

(Bryant et al. 2012) of BEAST v2.7.1 (Bouckaert et al. 2014), following Stange

et al. (2018). Time calibration was based on the estimated node age of 6.349

(95% CI: 4.434-8.698) million years ago (Ma) for the most recent common

ancestor (MRCA) of *C. ignobilis* and *M. cordyla* (Glass et al. 2023),

approximated by a lognormal distribution with a mean of 6.349 Ma, a standard

deviation of 0.205, and an offset of -0.01. We performed two independent runs

of 50,000,000 MCMC generations, sampling every 10,000 steps and discarding

the first 10% of the trees as burn-in, then combined the posterior tree

distributions from the two runs. We estimated a maximum clade credibility tree

using TreeAnnotator v2.7.1 (Bouckaert et al. 2014) and visualized the tree using

FigTree v1.4.4 (http://tree.bio.ed.ac.uk/software/figtree/). The estimation based

on the concatenated sequences of the 8 mtDNA loci was done using BEAST

v2.7.1. We partitioned the sequences by locus and estimated the substitution

model for each partition using the bModelTest v1.3.3 (Bouckaert and

Drummond 2017), which averages over different substitution models in BEAST.

We assumed the optimized relaxed clock model and the Yule model for the

clock model and the tree prior, respectively. We used the mitogenome sequence

of *C. ignobilis* (accession No.: NC_022932) as an outgroup. Time calibration,

MCMC simulations, convergence check, tree estimation, and visualization were

the same as described above.

Ancestry Estimation and Admixture Detection

We estimated the ancestries of the individuals in the North cluster and the

putative early-generation hybrids using the 'HIest' package v2.0 (Fitzpatrick

2012) in R. This program estimates the ancestries of target individuals relative

to predefined parents via log-likelihood maximization, using target genotypes

and parental allele frequencies. First, we assumed the East and West clusters

as potential parents and used 108 unlinked SNPs with allele frequencies

differing by >0.5 between them. Second, we estimated the ancestries of

putative hybrids only, assuming the three clusters as potential parents, using

130 SNPs with allele frequencies differing by >0.5 between one or more pairs of

the clusters. In the first analysis, we estimated the ancestry index (S) and the

inter-class heterozygosity (H) using the *HIest* function, employing the SANN

method for searching the likelihood surface, 500 MCMC generations, and the

option surf = TRUE. S represents the proportion of an individual's alleles

derived from one predefined parental population (here, the East cluster),

equivalent to the hybrid index. H indicates the proportion of loci with one allele

from each parental population. Expected H values vary between classes of

early-generation hybrids (F1: 1.0; F2 and backcross: 0.5), decreasing if

succeeding reproduction occurs exclusively among hybrids and alleles are lost

by drift. We also estimated the likelihoods for six possible genotype classes

resulting from 2 generations of inter- or intraspecific crosses (parental, F1, F2,

and backcrosses) using the *HIclass* function. We compared the maximum

likelihood of the six classes with that of the maximum likelihood estimation of S

and H using the *HItest* function. For the second analysis, we estimated the

ancestry indices S_{East}, S_{West}, and S_{North} of the putative early-generation hybrids

using the *threeway* function. These indices are equivalent to S, representing the

proportion of alleles derived from the three predefined parental populations. We

also calculated the likelihoods for 15 possible genotype classes in the first two

generations of hybridization between pairs of parental populations using the

thirdclass function and compared the maximum likelihood with that obtained

from the *threeway* analysis using the *HItest3* function.

We tested if the North cluster originated from an admixture of the East and

West clusters by three methods based on the 3rd dataset of the genome-wide

SNPs. First, we used HyDe v1.0.0 (Blischak et al. 2018), which assesses the

evidence for hybridization under a model of hybrid speciation and estimates the

associated ancestry proportions in a putative hybrid population from two

parental populations (Γ and 1−Γ). We tested for hybridization at the population

level for each cluster, leaving the remaining clusters as putative parental

populations, using the *run_hyde.py* script. We then performed 500 bootstrap

resampling using the *bootstrap_hyde.py* script to detect individual variation in r

within a hybrid population. Second, we calculated the f_3 statistic using 'admixr'

v0.9.1 (Petr et al. 2019). The f_3 statistic is a three-population statistic based on

branch lengths between populations. A significantly negative f_3 statistic indicates

that a predefined outgroup population is a mixture of two ingroup populations

(Patterson et al. 2012). We calculated the f_3 statistics designating each cluster

as an outgroup. Third, we calculated the *D*-statistic using 'admixr' to test for

introgression between the clusters. The *D*-statistic detects introgression

between non-sister taxa under the user-defined topology by asking if the allele-

sharing pattern between the taxa deviates from what is expected under

incomplete lineage sorting without introgression (Patterson et al. 2012). Given

the possible hybrid origin of the North cluster (see Results), two strictly

bifurcating topologies were possible for the user-defined topology, i.e., one with

a sister relationship between the East and North clusters, i.e.,

(East,North),West), and another between the West and North clusters,

(West,North),East). These were referred to as topologies 1 and 2, respectively.

We calculated the *D*-statistics for both.

365   *Demographic Model Selection and Parameter Estimation*

We inferred the demographic history of *M. cordyla* by testing alternative

scenarios using fastsimcoal v2.7.0.9 (Excoffier et al. 2021). Given the admixed

nature of the North cluster, we aimed to determine if it originated from an

admixture of the East and West clusters ("hybrid speciation" scenario) or an

instantaneous introgression event following divergence ("pulse introgression").

We also considered a null scenario of strict bifurcation between the clusters

without a pulse introgression event ("bifurcation"), following Noguerales and

Ortego (2022). Alternative topologies 1 and 2, i.e., (East,North),West) and

(West,North),East), were assumed for the pulse introgression and bifurcation

scenarios, resulting in five scenarios. These scenarios were simulated with or

without gene flow between the clusters before (ancestral gene flow) and after

(contemporary gene flow) the formation of the North cluster, resulting in 20

models (Fig. S1).

We performed 100 independent fastsimcoal runs for each demographic

model using the 4th dataset for model selection. Each run comprised 100

expectation-conditional maximization (ECM) cycles with 100,000 coalescent

simulations. The run with the smallest difference between the maximum

expected likelihood and the maximum observed likelihood was considered the

best fit. For model selection, we compared 1) Akaike's information criterion

(AIC) based on the best-fit run of each model, and 2) the likelihood distributions

from 100 expected SFS, generated by 1,000,000 coalescent simulations under

the parameters estimated from the best-fit run (Meier et al. 2017). Under the

selected model, we then estimated demographic parameters using the 5th

dataset. To obtain 95% confidence intervals (CIs) for each parameter, we

generated 100 parametric bootstrap replicates of SFS under the maximum

likelihood parameters of the best-fit run following the program manual. The

parameters were re-estimated using 100 independent runs for each

bootstrapped SFS with the same settings, except that the initial values were set

to the maximum likelihood parameters of the best-fit run, and the number of

ECM cycles was set to 50. We rescaled the estimated parameters assuming a

mutation rate of $1E^{-8}$ per site per generation and a generation time of 1.2 years

(see above).

399   *Morphology*

We examined 37 linear measurements and 17 countable characters in 37

specimens from Japan, Taiwan, Vietnam, and Thailand (Tables S1, S3). This

analysis aimed to determine if the genetic clusters 1) differ in ecologically

relevant morphological characters, suggesting ecological divergence, and 2)

can be recognized as separate species in a taxonomic sense. Linear

measurements were made to the nearest 0.01 mm using a digital vernier

caliper. Methods for measuring and counting generally followed Marr and

Schaefer (1949), Gibbs and Collette (1967), and Muto et al. (2016). Linear

measurements are expressed as % of the Standard length (SL). To illustrate the

variation in overall morphology, we performed a PCA of 24 characters with no

missing data using the R package 'FactoMineR' v2.11 (Lê et al. 2008). All linear

measurements were corrected for allometry using the R package 'GroupStruct'

v0.1.0 (Chan and Grismer 2022) before performing the PCA.

414   *Ecology*

To investigate potential ecological divergence between the genetic clusters,

we re-analyzed the ecological data of *M. cordyla* in Taiwan reported by Su et al.

(2020). The data include sampling season and location, individual body length

and weight, gonad weight, gonad somatic index (GSI: [gonad weight/body

weight] × 100), and sex (Table S4), along with the mtCR sequences of the

specimens. Su et al. (2020) compared the ecological factors between two major

lineages of mtCR sequences. In the present study, we show that one of these

two major lineages corresponded to the West cluster, while the other

corresponded to the East and North clusters (see Results). Additionally, we

demonstrate that the East and North clusters can be distinguished by a minor

mtCR divergence within the major lineage. These observations allowed us to

reclassify the specimens examined by Su et al. (2020) into the three clusters

based solely on the mtCR sequences reported by the authors, albeit with some

uncertainty. First, we tested the association between the ecological factors and

genetic clusters under the Random Forest model using the R package 'ranger'

v0.16.0 (Wright and Ziegler 2017). We assessed the importance of each

ecological factor using the mean decrease in accuracy and estimated the

significance of this importance metric with a permutation test using the R

package 'rfPermute' v2.5.2 (Archer 2021). Second, we performed a hierarchical

cluster analysis to create a dendrogram of the specimens based on the

ecological data. We calculated the pairwise Gower's distance between each

specimen and constructed Ward's hierarchical agglomerative clustering using

the R package 'stats' v4.2.2 (R Core Team 2022). Third, we performed a Factor

Analysis of Mixed Data (FAMD) using the R package 'FactomineR'.

RESULTS

The first two PCA axes separated the specimens into three distinct clusters:

1) individuals from Japan to Malaysia along the east coast of the Eurasian

continent, 2) individuals from Japan to the Philippines along the Kuroshio

Current, and 3) individuals from Japan and Taiwan (Fig. 1a, b). These clusters

are referred to as the West, East, and North clusters, respectively, based on

their geographical distributions. The East and West clusters occupied opposite

positions along PC 1, with the North cluster in an intermediate position.

Principal component 2 further separated the North cluster from the others. Two

individuals from Japan were intermediate between the North and West clusters

and are hereinafter referred to as putative early-generation hybrids. One

individual from Thailand was plotted near the East cluster, while all other

individuals from Thailand were included in the West cluster. This individual from

Thailand is also regarded as a putative early-generation hybrid. In the

STRUCTURE analysis, the most likely number of K was 2 (Fig. S2). Assuming

$K = 2$, individuals of the North cluster showed signatures of admixture between

the East and West clusters, with a greater ancestry contribution from the former

(Fig. 1c). The putative early-generation hybrids from Japan showed

intermediate ancestry between the North and West clusters, while that from

Thailand was intermediate between the North and East clusters. Assuming $K =$

3–8, individuals in the North cluster showed admixed ancestry from three

clusters: two corresponding to the East and West clusters, and an additional

minor cluster (Fig. S2). The Neighbor-Net network showed similar patterns

regarding the separation of the East, West, and North clusters, the intermediate

nature of the North cluster, and the positions of the putative early-generation

hybrids (Fig. S3). Pairwise F_{ST} across all SNPs was greatest in the East-West

comparison, followed by the West-North and East-North comparisons (0.2989,

0.2167, and 0.0861, respectively); all comparisons were significant (Table S5).

Significant locus-by-locus pairwise F_{ST} was most frequently observed in the

East-West comparison, followed by the West-North and East-North

comparisons (Fig. S4). Several SNPs showed significant differentiation in both

East-North and West-North comparisons, highlighting the unique evolutionary

history of the North cluster at these loci. In terms of allele frequency, a total of

130 SNPs were highly differentiated (i.e., differences >0.5) between one or

more cluster pairs (Fig. S5; Table S6). The majority of these SNPs (95 loci)

were highly differentiated in the East-West comparison, with the North cluster

showing intermediate allele frequencies. For 56 of these 95 SNPs, the North

cluster's allele frequency was closer to the East cluster, while for others, it was

closer to the West cluster, suggesting that alleles from the East and West

clusters were alternatively sorted in the North cluster following admixture. In

contrast, the North cluster was highly differentiated from both the East and West

clusters at 4 SNPs, due to the alleles reaching high frequency in the North

cluster while being almost absent in the East and West (0.56–0.67 in the North

vs. 0–0.05 in East and West).

The population sizes of the East and West clusters were inferred to have

declined approximately 70–100 thousand years ago (Ka), while the North

cluster remained stable (Fig. S6). Subsequently, all three clusters expanded in

population size towards the present time.

490   *mtDNA Differentiation*

The minimum spanning network based on concatenated sequences of CYTB

and CR (873 + 760 bp) consisted of three major haplogroups, each

corresponding to one of the three genetic clusters (Fig. 1d). We refer to these

haplogroups as the East, West, and North haplogroups, corresponding to their

respective clusters. The East-West and East-North divergences of the

haplogroups were equivalent to 58 and 4 mutations, respectively. However, the

East and North clusters were not reciprocally exclusive: one individual of the

North cluster was included in the East haplogroup. Moreover, the East-North

divergence disappeared when only the CYTB sequences or concatenated

sequences of the seven protein-coding genes were analyzed (Fig. S7),

indicating the divergence is due to variation in CR sequences. The putative

early-generation hybrids from Japan were included in the West haplogroup (Fig.

1d). We were unable to obtain the CR sequence for the putative hybrid from

Thailand; this individual was included in the "East + North" haplogroup in the

CYTB-only network (Fig. S7).

507   *Ancestry Estimation and Admixture Detection*

In the *HIest* analysis, assuming the East and West clusters as potential

parents, S and H for individuals in the North cluster were 0.51–0.76 and 0.10–

0.35, respectively (Fig. 2a). The values of S suggest a greater ancestry

contribution from the East cluster, while those of H suggest that the individuals

are later-generation hybrids (>F_2), having exclusively reproduced within the

same cluster. *HIclass* assigned each individual in the North cluster to either

backcross to the East cluster or F_2, but the likelihood ratio test by *HItest*

rejected these simple assignments in favor of the more nuanced results of *HIest*

(Table S7). Compared to the North cluster individuals, the putative early-

generation hybrids from Japan showed lower S (i.e., more closely related to the

West cluster) and higher H, whereas the hybrid from Thailand showed greater S

and lower H. *HIclass* assigned them to F_2 and backcross to the East cluster,

respectively, but *HItest* again rejected these assignments (Fig. 2a). In the

threeway analysis, where the three clusters were assumed as potential parents,

the putative early hybrids from Japan and Thailand showed mixed ancestries

predominantly between the North and West clusters and between the North and

East clusters, respectively (Table S8). *Thirdclass* assigned them to F_2 between

the North and West clusters and F_2 between the North and East clusters,

respectively; *HItest3* did not reject these assignments.

Fig. 2. a) Estimates of ancestry index (*S*), equivalent to hybrid index, and inter-class heterozygosity (*H*) by HIest, assuming the East and West clusters as potential parents. Individual assignments to the three clusters and putative early-generation hybrids are based on the PCA (Fig. 1b). Double circles indicate specimens for which simple classification as one of 6 genotype classes within 2 generations of inter- or intraspecific crosses (parentals, F1, F2, and backcrosses) was rejected by *HItest*. See Table S8 for results assuming the three clusters as potential parents. b) Density plot of estimated gamma (Γ) values for the North cluster across 500 individual bootstraps in HyDe. Γ and 1-Γ represent estimated ancestry contributions from the East and West clusters, respectively. c) A schematic diagram of the best-fit demographic model, i.e., hybrid speciation with ancestral and contemporary gene flow between clusters, tested using Fastsimcoal 2. Parameters include; N: ancestral ($_{Anc}$) or contemporary ($_{Con}$) effective population size of the common ancestor ($_A$) or the three genetic clusters ($_E$ / $_W$ / $_N$); T: time of divergence ($_{Div}$) or fusion ($_{Fus}$) of the East and West clusters; Γ: ancestry contribution from the East (Γ$_{Fus}$) or West (1−Γ$_{Fus}$) cluster upon fusion; M: ancestral ($_{Anc}$) or contemporary ($_{Con}$) gene flow between clusters, with the letters following the underscore indicating the direction of the gene flow (e.g., M$_{Con\_EW}$ represents contemporary gene flow from the East to West cluster forward in time). See Fig. S1 for all 20 models tested and Fig. S9 and Table S10 for model evaluation by likelihood and Akaike's Information Criterion values. d)–g) Demographic parameters estimated using Fastsimcoal 2 under the best-fit model. Dots and vertical bars represent maximum likelihood point estimates and 95% confidence intervals based on 100 bootstrap replicates. Timings of demographic events are in units of years. Effective population sizes are in units of the number of alleles. Migration rates are in units of proportions of alleles moving per generation. See Table S11 for tabulated values.

Admixture detection using HyDe was significant only for the North cluster (Z-

score = 2.584, p = 0.0049; Table S9), supporting its hybrid origin. Bootstrap

resampling of the individuals in the North cluster showed an unimodal

distribution of ɼ values, indicating uniform admixture across individuals (Fig. 2b).

This uniform admixture is consistent with hybrid speciation rather than ongoing

introgression (Blischak et al. 2018). The departure of the ɼ values from 0.5

(mean = 0.636; 95% CI = 0.633–0.638) suggested a greater ancestry

contribution from the East cluster. The f_3 statistic was negative when the North

cluster was designated as an outgroup, supporting its admixed ancestry (f_3 = -

0.023, Z-score = -4.338; Table S9). Finally, the D-statistics detected gene flow

between both West-North and East-North combinations (West-North: D = 0.185,

Z-score = 7.079; East-North: D = 0.296, Z-score = 11.389; Table S9).

565   *Divergence Time Estimation*

The East and North clusters formed a single clade, leaving the West cluster

in another clade, in both the species tree based on genome-wide SNPs and the

mtDNA phylogeny (Fig. S8). The MRCA age of the three clusters was estimated

to be 0.269 Ma (95% CI: 0.168–0.387) based on genome-wide SNPs and 1.333

Myr (0.244–3.208) based on mtDNA phylogeny. The divergence between the

North and East clusters was estimated at 0.126 Ma (0.075–0.184) and 0.196

Ma (0.015–0.574), respectively. All estimated divergence ages fell within the

Pleistocene. Note that the East and North clusters were not reciprocally

monophyletic in the mtDNA phylogeny; the divergence time mentioned above is

the node age for two clades approximately corresponding to the East and North

clusters.

578   *Demographic Model Selection and Parameter Estimation*

The hybrid speciation model with gene flow between clusters both before and

after the formation of the North cluster (i.e., ancestral and contemporary gene

flow) was the best fit among the 20 models tested, based on the highest log-

likelihood and lowest AIC (Fig. 2c; Fig. S9; Table S10). The other three hybrid

speciation models, differing in ancestral and contemporary gene flow, were not

better supported than the pulse-introgression models assuming a sister

relationship between the West and North clusters (topology 2) (Fig. S9; Table

S10). The models assuming topology 2 were favored over the alternative

topology 1 if accompanied by a pulse introgression event or contemporary gene

flow.

Under the best-fit model, the divergence and fusion of the East and West

clusters were estimated to have occurred 479.9 Ka (95% CI: 256.1–549.9) and

96.5 Ka (94.9–106.4), respectively (Fig. 2d; Table S11), both within the

Pleistocene. The ancestry contribution from the East cluster (Γ_{Fus}) upon the

fusion was less than that from the West cluster ($1-\Gamma_{Fus}$) (Fig. 2e). Effective

population size was greater in the West than in the East cluster, both before and

after the fusion, with both clusters expanding over time (Fig. 2f). Contemporary

population size was smallest in the North cluster, consistent with its restricted

geographical range. Gene flow between the clusters was highly asymmetric;

contemporary gene flow from the East to North cluster was orders of magnitude

greater than that in the opposite direction and other cluster combinations (Fig.

2g; Table S11). Similarly, ancestral gene flow from the West to East cluster was

significantly greater than that in the opposite direction.

603   *Ecology*

A NJ tree based on the 592 bp alignment of the mtCR sequences, including

both the present and Su et al's. (2020) sequences, showed that 20, 55, and 15

haplotypes reported by Su et al. (2020) are most likely identified as the East,

West, and North clusters, respectively (Fig.S10; Table S4). Based on this

identification, we tested possible ecological divergence between the three

clusters. The Random Forest model showed that sampling location and season

were significant predictors of the cluster ($p < 0.05$) (Fig. 3a), suggesting that the

clusters tend to be found in different locations and seasons along the coast of

Taiwan. This implies habitat segregation or differences in migration patterns

contribute to reproductive isolation between the clusters. The Random Forest

proximity scores of the three clusters, reduced to two dimensions through

multidimensional scaling, largely overlapped (Fig. 3b), whereas the individual

assignments to the clusters predicted from the Random Forest model were

more accurate for the North cluster than for the other two clusters (Fig. 3c). This

suggests that the ecological divergence is more pronounced between the North

and the other clusters. In contrast, the dendrogram constructed through the

hierarchical cluster analysis could not fully explain the split of expected East,

West, and North clusters (Fig. S11a). Similarly, the three genetic clusters were

 not clearly separated on the first four dimensions (explaining 61.5% of the total

 variation) derived from the FAMD (Fig. S11b–g).

Fig. 3. Results of the ecological (a–c) and morphological (d–h) analyses. a) Importance of ecological factors in the Random Forest model, shown by the mean decrease in accuracy. Red bars indicate significant results (permutation test, $p < 0.05$). GSI: gonad somatic index. b) Confusion matrix of the Random Forest model comparing true (genetic) individual assignments to the clusters with those predicted by the model. c) Proximity scores reduced to two multidimensional scaling dimensions, showing ecological similarity between samples as determined by the Random Forest model. Inner points represent true (genetic) assignments to the clusters, and outer circles indicate those predicted by the model. d) Principal component analysis of 24 morphological characters. The proportion of variance explained by each principal component is in parentheses. e)–h) Plots of selected morphological characters. e) The distance between the tip of the snout and the origin of the 1st dorsal fin. f) The distance between the tip of the snout and the origin of the 2nd dorsal fin g) The distance between the tip of the snout and the origin of the 1st detached anal-fin spine. h) The number of gill rakers.

Morphology

Principal components (PCs) 1–4 explained 26.0%, 14.4%, 12.6%, and 6.7%

of the total variation in overall morphology, respectively (Fig. S12a). The North

cluster was separated from the East and West clusters on PC 1, suggesting its

distinct morphology (Fig. 3d). This reflects the North cluster's relatively

elongated anterior body: it is distinguishable from the East and West clusters by

the distances from the tip of the snout to the origins of the first-dorsal, second-

dorsal, and anal fins (Fig. 3e–g), with the former two characters strongly

contributing to PC 1 (Fig. S12b–d). The East and West clusters were separated

from each other on PCs 2 and 3, to which gill-raker number and head width

strongly contributed, respectively (Figs. 3d, S12b–e). The West cluster had

fewer gill-rakers (24–29) compared to the East and North clusters (31–35 and

31–34, respectively) (Fig. 3h), implying a distinct trophic niche, as gill-raker

number generally correlates with diet size and shape in fishes (e.g., Gerking

1994). Other characters did not differ considerably between the three clusters

(Fig. S13). The putative hybrids occupied intermediate positions between the

East and North clusters on PC 1. The discovery of distinct morphological

characteristics, coupled with genetic and ecological differences, would allow us

to describe the three clusters as taxonomically distinct species. See Appendix 2

for taxonomic implications.

660   **DISCUSSION**

661   *Reproductive Isolation and Gene Flow*

We demonstrated that the North cluster is an evolutionarily independent

lineage of hybrid origin. The genome-wide differentiation (Figs. 1b, S3–S5),

coupled with the presence of loci where the North cluster showed significant

differentiation from both the East and West clusters (Figs. 1d, S4, S5), provides

strong evidence that the North cluster is evolving separately from the others in

sympatry. Eco-morphological differences further support its ecological and

evolutionary independence (Figs. 3, S11–13). These results highlight that the

North cluster can be recognized as a distinct species under multiple species

concepts (reviewed in Coyne and Orr, 2004), such as the biological, genotypic,

evolutionary, and cohesion concepts. Moreover, the genetic analyses

consistently indicated the intermediate and admixed nature of the North cluster

(Figs. 1c, 2a–b, S3; Tables S7–S9), and demographic model selection favored

the hybrid speciation scenario over the introgression scenario (Figs. 2c, S9;

Table S10). To our knowledge, this is the most compelling example of

homoploid hybrid speciation in marine fishes.

The present system represents a relatively rare case of HHS where parental

and hybrid species coexist in sympatry. This pattern could most readily be

realized by ecological divergence between the species, which facilitates

extrinsic pre- or postzygotic reproductive isolation while reducing competition

(Buerkle et al. 2000). Indeed, our Random Forest analysis suggested spatio-

temporal segregation of the North cluster from the others, i.e., the seasonal and

locational factors strongly associated with the predictions of clusters, and the

distinct number of gill-rakers in the West cluster implied its unique trophic niche

(Fig. 3). We suggest that these ecological differences maintain the eco-

evolutionary independence of the sympatric species. However, the ecological

divergence is likely a by-product of allopatry under historical climate changes

(rather than a direct outcome of hybridization), with this allopatry being the

primary driver of HHS, given the estimated demographic histories and

ecogeographical setting of the present system (see the next section

"*Ecogeographic Scenario of Hybrid Speciation in M. cordyla*").

It remains uncertain whether the reproductive isolation of the North cluster

from the sympatric parental clusters is complete: the results of our genomic

analyses were equivocal. We observed three putative F_2s between the North

and parental clusters (Table S8), indicating incomplete pre- and postzygotic

isolation before the F_2 generation. Nevertheless, postzygotic isolation due to

hybrid breakdown can mainly manifest in F_2 or later generations, effectively

preventing gene flow (Orr 1995; Stelkens et al. 2015; Lollar et al. 2023). The

demographic parameter estimation suggested the presence of "contemporary

gene flow" (Fig. 2g); however, gene flow may be no longer ongoing, as the

estimation represents an average over generations from the formation of the

North cluster until the present (Fig. 2c). Another result, the uniform ancestry

composition across the individuals in the North cluster (Fig. 2a–b), rather

supports exclusive reproduction among the members of the same cluster, i.e.,

the lack of gene flow.

We observed unequal parental genomic contributions in the North cluster

(Figs. 1b–c, 2a–b, S3–S5), which is typical in HHS (e.g., Elgvin et al. 2017;

Barrera-Guzmán et al. 2018; Noguerales and Ortego 2022; Lopes et al. 2023;

Rosser et al. 2024). Relative parental contributions in hybrid species are

determined by initial contributions upon fusion, subsequent gene flow, and

differential fixation of ancestry blocks via drift and selection (genomic

stabilization) (Baack and Rieseberg 2007; Runemark et al. 2019). In the present

system, the predominance of East cluster ancestry seems to largely reflect

asymmetric gene flow following the fusion; the demographic parameter

estimation suggested that the West cluster initially made a greater contribution

$(1−\Gamma_{Fus} > 0.5$: Fig. 2e), and subsequent gene flow was predominantly from the

East to the North cluster (Fig. 2f). Additional support for the initial closer

relatedness of the West and North clusters and the importance of subsequent

gene flow comes from comparing suboptimal demographic models, where the

models assuming both a sister relationship between West and North (topology

2) and subsequent gene flow or pulse introgression were generally better

supported than those without (Fig. S9, Table S10). The observed asymmetric

gene flow may have resulted from asymmetry in the strength of reproductive

isolation (Rosser et al. 2024). Alternatively, it may reflect unequal frequencies of

repeated secondary contacts under the Pleistocene climatic oscillations (see

the next section).

The limited number of SNPs available constrained our analytical power. This

limitation led us to compromise the complexity of the demographic models,

excluding events like population size changes, multiple pulses of introgression,

or temporal shifts in the magnitude of gene flow, even though such events are

plausible (e.g., De Jode et al. 2023; Muto and Kai 2023). Additionally, the

preference for the hybrid speciation scenario over the pulse introgression was

not robust, depending on the presence of both ancestral and contemporary

gene flow in the model (Fig. S9). This lack of robustness might also be

attributed to the limited number of SNPs. We plan a future study to address

these limitations using whole genome sequencing.

738   *Ecogeographic Scenario of Hybrid Speciation in M. cordyla*

The Pleistocene glacial cycles led to changes in sea levels and seawater

temperatures, causing allopatric divergence, admixture, local adaptation, and

population size changes in coastal marine organisms (Bowen et al. 2016).

Based on our results and the geographic setting, we propose that multiple

cycles of these processes primarily drove HHS in *M. cordyla*. Below, we outline

a possible scenario in detail. Moreover, based on this scenario, we propose a

model for HHS in marine coastal ecosystems, including a novel hypothesis on

the initial persistence of nascent hybrid populations (see the next section

"*Hybrid Speciation in Coastal Marine Environments*").

In the Indo-West Pacific, a vast shallow continental shelf, the Sunda Shelf

(Fig. 1a), strongly limited marine connectivity between the Indian and Pacific

Oceans during the Pleistocene low sea-level (glacial) periods. This "Indo-Pacific

barrier" drove allopatric divergence in various marine organisms, evidenced by

concordant distribution patterns of sister species and phylogeographic breaks

around the Sunda Shelf (Carpenter et al. 2011; Gaither and Rocha 2013). The

distributions of the East and West clusters of *M. cordyla* in the tropical West

Pacific align with this pattern (Fig. 1a), and their estimated divergence time fell

within the Pleistocene, suggesting that they originated from the allopatric

divergence across the Indo-Pacific Barrier (Fig. 4a).

Fig. 4. a)–d) The proposed ecogeographic scenario of HHS in the present system.
Pleistocene glacial-interglacial cycles drive a) allopatric divergence of the East and West
clusters, b) their northward range expansions and secondary contact, leading to the
formation of the North cluster via hybridization, c) range contractions of the East and West
clusters resulting in the geographical isolation of the three clusters, during which ecological
and other isolating barriers develops, and d) range overlap and coexistence in sympatry.
e)–h) The ecogeographic model of hybrid speciation in coastal environments driven by the
Pleistocene glacial cycles. Two coastlines stretching from low to high latitudes in parallel
along a landmass, joining at high-latitude extreme, are shown. Gene flow can only occur
along the coastline, not across the landmass. This process can also occur where
topographic features are latitudinally reversed (i.e., two coastlines join at low-latitude
extreme) or the barrier to gene flow is any other uninhabitable zone, such as an open
ocean as in the present system. e) During a glacial period, high-latitude coastlines become
uninhabitable due to extreme conditions (e.g., low temperature), resulting in allopatric
divergence of parental species. Adaptation over latitudinal gradients in temperature and
other factors occurs independently, based on different loci in each species. f)
Environmental change from glacial to interglacial makes the high-latitude area habitable,
leading to range expansions and secondary contact of parentals, forming a hybrid
population. Hybridization occurs among individuals best adapted to high latitudes since the
direction of the environmental change is aligned with the latitudinal gradient. g) In the next
glacial period, the hybrids have greater local fitness in high-latitude areas due to
additive/epistatic effects of adaptive variants from the parentals. Additionally, heterosis in
hybrids via the combination of recessive deleterious alleles, having been accumulated in
the expansion front of the parentals, can also contribute the greater fitness (not depicted).
The greater fitness allows hybrids to persist in high latitudes while parentals contract to low-
latitude refugia, resulting in geographical isolation. During this period of allopatry, isolating
barriers such as pre- and post-mating barriers based on ecological divergence may

develop as by-products. h) With isolating barriers in place, the hybrid population maintains
independence from parentals upon contact in subsequent interglacial periods.
788

Following the initial divergence, the ranges of the two clusters likely

expanded and contracted in response to the glacial cycles, as demonstrated in

many organisms (reviewed in Hewitt 2004; Hofreiter and Stewart 2009;

Seersholm et al. 2020). Specifically, during warm interglacial periods, poleward

range expansions likely occurred along mainland Asia for the West cluster and

along the Kuroshio current for the East cluster (Kuriiwa et al. 2014; Motomura

and Matsunuma 2022). One such poleward expansion may have resulted in

their range overlap in the mid-latitude temperate area, similar to the present

distribution pattern, triggering hybridization and the formation of the North

cluster (Fig. 4b). In the subsequent glacial period, the North cluster survived in

the temperate area with a stable population while the parentals contracted

towards tropical refugia, as the Stairway Plot analysis implies (Fig. S6). This, in

turn, resulted in their geographical isolation, securing the persistence of the

nascent North cluster without being swamped by gene flow from the parentals

(Fig. 4c). This allopatric period facilitated the development of inferred ecological

difference and other unobserved isolating barriers, allowing the coexistence in

sympatry during the next contact (Fig. 4d). The ancestral and contemporary

gene flow inferred from the demographic modeling likely reflects relatively minor

secondary contacts between the clusters.

We hypothesize that the North cluster's survival in higher latitudes during the

glacial period (i.e., Fig. 4c) was driven by clinal adaptations in the parental

clusters. Prior to hybridization, parallel adaptation along latitudinal gradients in

temperature and associated environmental factors likely occurred within each

parental cluster, resulting in clinal patterns of adaptive genetic variants (Fig. 4e)

(Jeffery et al. 2017; Teng et al. 2023). Subsequent hybridization probably took

place at the high-latitude edge of the parental ranges, where these adaptive

variants were most prevalent (Fig. 4f). In the North cluster, these variants

recombined, enhancing fitness through additive and/or epistatic effects (Mallet

2007; Abbott et al. 2013), which enabled its survival in high latitudes during the

subsequent glacial period (Fig. 4g). Conversely, deleterious alleles may have

also accumulated at the expansion front of the parental clusters before

hybridization, via allele surfing (Excoffier et al. 2009). However, if these

deleterious alleles were recessive, they might have contributed to heterosis in

hybrids, aiding their long-term persistence (MacPherson et al. 2022).

Supporting our hypothesis, a recent simulation study demonstrated that

admixture between two allopatric parental populations adapting to identical

environments can result in hybrids that adapt more rapidly than the parentals to

a novel environment (Kulmuni et al. 2024).

828   *Hybrid Speciation in Coastal Marine Environments*

Hybrid populations can express transgressive phenotypes via combinations

of parental genetic variants, exploiting novel niches and thereby becoming

geographically isolated from the parental populations (Rieseberg et al. 2003;

Gross and Rieseberg 2005; Mallet 2007; Abbott et al. 2013; MacPherson et al.

2022; Kulmuni et al. 2024). Although this process may underlie HHS in various

organisms (Mavárez and Linares 2008; Larsen et al. 2010; Brennan et al. 2012;

Kang et al. 2013; Lamichhaney et al. 2018; Maier et al. 2019; Noguerales and

Ortego 2022), the ecogeographic settings enabling the process are largely

unknown. The scenario described in the preceding section illustrates how this

process can occur in the ocean, including the necessary ecogeographic setting.

We propose this scenario as a general model of hybrid speciation in coastal

marine environments driven by the Pleistocene glacial cycles (Fig. 4e–h). This

model provides realistic, testable explanations for how parental adaptive genetic

variants are created and how hybrids' distinct niches lead to geographical

isolation, filling the gaps in the current understanding.

Under the model, hybrid speciation can occur where coastal marine habitats

in two parallel latitudinal arrays (e.g., coastlines or archipelagos) in a relevant

geographical scale, separated by an uninhabitable zone (e.g., land or open

ocean) but joined at latitudinal extremes (Fig. 4e). The uninhabitable zone

prevents gene flow between parental species, and hybridization occurs at the

latitudinal extreme under glacial maxima/minima (Fig. 4f). The following regions

not only fit these topographic conditions, but also are characterized by

secondary contact driven by the Pleistocene glacial cycles: temperate to tropical

Indo-west Pacific (Gaither and Rocha 2013); the Japanese Archipelago (Hirase

2022); the Aleutian Archipelago (Liu et al. 2012); southern Africa (Gaither et al.

2015); and the Florida peninsula (Soltis et al. 2006). We predict that these

regions harbor undiscovered cases of marine hybrid speciation.

Another key component of the model is the adaptation of the nascent hybrid

population to environmental conditions at the latitudinal extreme, leveraging the

preexisting adaptative genetic variants of the parents established through

parallel adaptation over a common latitudinal gradient (Fig. 4f–g). The parent's

parallel adaptation may be based on similar phenotypes, but their genetic

architectures are most likely *not* identical (e.g., Elmer and Meyer 2011; Jones et

al. 2012), due to the lack of gene flow. Therefore, the recombinant hybrid

genome can express transgressive phenotypes, conferring greater fitness than

parents (Mallet 2007; Abbott et al. 2013). This greater fitness leads to local

persistence under subsequent global cooling/warming and eventual allopatry

from the parents, during which isolating barriers develop as by-products (Fig.

4h). These can be tested by investigating the eco-evolutionary forces underlying

inter- and intraspecific genomic and phenotypic variations in parental and hybrid

populations. For instance, Wang et al. (2021) employed a genome-wide

association study, genomic cline analysis, and examinations of correlations

between climate and genotype to investigate persistent hybrid populations of

the *Setophaga* warbler complex. They demonstrated that these hybrids are

differentiated from parental populations in genomic regions related to climatic

adaptation, and that ancestry in these regions covaries with climatic variation.

ACKNOWLEDGMENTS

We thank the following persons who facilitated the sampling and data

collection: H. Motomura and members of The Kagoshima University Museum

(KAUM) Fish Team; H-C Ho and J-F Huang (National Museum of Marine

Biology and Aquarium, Taiwan); W-J Chen and M-Y Lee (National Taiwan

University); M. Nakae and K. Matsuura (National Museum of Nature and

Science, Japan); T. Nakabo, Y. Kai, and M. Matsunuma (Kyoto University

Museum); S. Kimura (Mie University); K. Koeda (University of the Ryukyus); M.

I. Mahyam, R. H. Raja Bidin, O. Muda, W. M. Arshaad, M. Katoh, and O. Abe

(Marine Fishery Resources Development and Management Department of

Southeast Asian Fisheries Development Centre: SEAFDEC); Y. G. Seah

(Universiti Malaysia Terengganu: UMT); A. Arshad (Universiti Putra Malaysia:

UPM); S. Arnupapboon and K. Phuttharaksa (Eastern Marine Fisheries

Research and Development Centre: EMDEC); N. D. The, P. V. Chien, D. V.

Nhan, V. V. Chien, and P. T. Thu (Institute of Marine Environment and

Resources, Vietnam: IMER); R. F. M. Traifalar, S. S. Garibay, late U. B. Alama,

A. C. Gaje, and R. Cruz (University of the Philippines Visayas: UPV); Satoshi

Ishikawa and members of the 'Coastal Area Capability Enhancement in

Southeast Asia' project (Research Institute for Humanity and Nature: RIHN). We

also thank Y. Kumazawa, F. Hibatullah, K. N. A. Kholil (Nagoya City University),

W. Kusuma (Universitas Brawijaya), and J. R. Glass (University of Alaska

Fairbanks) for their insightful discussions. The Philippines samples were

collected under a Memorandum of Agreement for joint research made by and

among the Department of Agriculture of the Republic of the Philippines (DA),

UPV, KAUM, RIHN, and Tokai University, facilitated by S. L. Sanchez [Bureau of

Fisheries and Aquatic Resources (BFAR), DA]. P. J. Alcala (DA) provided a

Prior Informed Consent Certificate, and I. P. Cabacaba and S. M. S. Nolasco

(BFAR, DA) provided a Fish Specimen Export Certificate (no. 2016-39812). The

Malaysian samples were collected during the Japan Society for the Promotion

of Science (JSPS) Asian Core Program "Establishment of Research and

Education Network on Coastal Marine Science in Southeast Asia" and the JSPS

Core-to-Core Program B "Asia-Africa Science Platforms" supported by UPM,

UMT, and the Ministry of Higher Education of the Government of Malaysia. The

Thai samples were collected with support from the EMDEC, the Training

Department of SEAFDEC, and the Office of Natural Science, National Science

Museum, Pathum Thani. The Vietnamese samples were collected with the

support of the IMER and the Ha Long Bay Management Department and with

permission to use the samples from the Biodiversity Conservation Agency,

Ministry of Natural Resources and Environment, Hanoi. Computations were

performed, in part, on the NIG supercomputer at ROIS National Institute of

Genetics, Japan.

918   **FUNDING**

This work was supported by the Japan Society for the Promotion of Science

(JSPS) Grants-in-Aid for Scientific Research (No. JP26840131, 19K23691,

19KK0190, 23H02242, and 24K02087), JSPS Overseas Research Fellowships

(No. 202160519), Sumitomo Fund (No.173246), Interdisciplinary Collaborative

Research Program of Atmosphere and Ocean Research Institute, The

University of Tokyo (No. JURCAOSIRG24-08), fund for integrated research

activities (I.8b, 47 sub-project) from the Ministry of Agriculture and Rural

Development, Vietnam, and Vietnam Academy of Science and Technology's

Grant in aid (VAST 06.04/18-19, NVCC 23.04/20-20).

929   **CONFLICT OF INTEREST**

930   The authors declare no competing interests.

AUTHORS' CONTRIBUTIONS

933   NM conceived and designed the study; NM, HH, NVQ, VV, MAG, and RPB

934   conducted sampling and sample preparation; NM conducted genetic analysis;

935   HH and NM conducted morphological analysis; SYC conducted ecological

936   analysis; NM wrote the manuscript with input from HH and SYC; all co-authors

937   contributed to improving the manuscript.

DATA AVAILABILITY

940   Data will be available on an online data repository upon publication of the

941   peer-reviewed version of this manuscript.

REFERENCES

Abbott R., Albach D., Ansell S., Arntzen J.W., Baird S.J.E., Bierne N., Boughman J.,
Brelsford A., Buerkle C.A., Buggs R., Butlin R.K., Dieckmann U., Eroukhmanoff F.,
Grill A., Cahan S.H., Hermansen J.S., Hewitt G., Hudson A.G., Jiggins C., Jones J.,
Keller B., Marczewski T., Mallet J., Martinez-Rodriguez P., Möst M., Mullen S.,
Nichols R., Nolte A.W., Parisod C., Pfennig K., Rice A.M., Ritchie M.G., Seifert B.,
Smadja C.M., Stelkens R., Szymura J.M., Väinölä R., Wolf J.B.W., Zinner D. 2013.
Hybridization and speciation. J. Evol. Biol. 26:229–246.
Archer E. 2021. rfPermute: Estimate Permutation p-Values for Random Forest
Importance Metrics. https://CRAN.R-project.org/package=rfPermute.
Baack E.J., Rieseberg L.H. 2007. A genomic view of introgression and hybrid speciation.
Curr. Op. Genet. Dev. 17:513–518.
Barrera-Guzmán A.O., Aleixo A., Shawkey M.D., Weir J.T. 2018. Hybrid speciation leads
to novel male secondary sexual ornamentation of an Amazonian bird. Proc. Natl.
Acad. Sci. USA. 115:E218–E225.
Blischak P.D., Chifman J., Wolfe A.D., Kubatko L.S. 2018. HyDe: A Python package for
genome-scale hybridization detection. Syst. Biol. 67:821–829.

Bouckaert R., Heled J., Kühnert D., Vaughan T., Wu C.H., Xie D., Suchard M.A.,
Rambaut A., Drummond A.J. 2014. BEAST 2: A software platform for Bayesian
evolutionary analysis. PLOS Comp. Biol. 10:1–6.

Bouckaert R.R., Drummond A.J. 2017. bModelTest: Bayesian phylogenetic site model
averaging and model comparison. BMC Evol. Biol. 17:1–11.

Bowen B.W., Gaither M.R., DiBattista J.D., Iacchei M., Andrews K.R., Grant W.S.,
Toonen R.J., Briggs J.C. 2016. Comparative phylogeography of the ocean planet.
Proc. Natl. Acad. Sci. USA. 113:7962–7969.

Brennan A.C., Barker D., Hiscock S.J., Abbott R.J. 2012. Molecular genetic and
quantitative trait divergence associated with recent homoploid hybrid speciation: a
study of *Senecio squalidus* (Asteraceae). Heredity. 108:87–95.

Bryant D., Bouckaert R., Felsenstein J., Rosenberg N.A., RoyChoudhury A. 2012.
Inferring species trees directly from biallelic genetic markers: bypassing gene trees
in a full coalescent analysis. Mol. Biol. Evol. 29:1917–1932.

Buerkle C.A., Morris R.J., Asmussen M.A., Rieseberg L.H. 2000. The likelihood of
homoploid hybrid speciation. Heredity. 84:441–451.

Cárdenas L., Hernández C.E., Poulin E., Magoulas A., Kornfield I., Ojeda F.P. 2005.
Origin, diversification, and historical biogeography of the genus *Trachurus*
(Perciformes: Carangidae). Mol. Phylogenet. Evol. 35:496–507.

Carpenter K.E., Barber P.H., Crandall E.D., Ablan-Lagman M.C.A., Ambariyanto,
Mahardika G.N., Manjaji-Matsumoto B.M., Juinio-Meñez M.A., Santos M.D., Starger
C.J., Toha A.H.A. 2011. Comparative phylogeography of the coral triangle and
implications for marine management. J. Mar. Biol. 2011:1–14.

Catchen J., Hohenlohe P.A., Bassham S., Amores A., Cresko W.A. 2013. Stacks: an
analysis tool set for population genomics. Mol. Ecol. 22:3124–3140.

Chan K.O., Grismer L.L. 2022. GroupStruct: An R Package for Allometric Size Correction.
Zootaxa. 5124:471–482.

Chen S., Zhou Y., Chen Y., Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor.
Bioinformatics. 34:i884–i890.

Chhatre V.E., Emerson K.J. 2017. StrAuto: automation and parallelization of
STRUCTURE analysis. BMC Bioinformatics. 18:192.

Coyne J.A., Orr H.A. 2004. Speciation. Sunderland, Massachusetts: Sinauer Associates.

Danecek P., Auton A., Abecasis G., Albers C.A., Banks E., DePristo M.A., Handsaker
R.E., Lunter G., Marth G.T., Sherry S.T., McVean G., Durbin R., 1000 Genomes

Project Analysis Group. 2011. The variant call format and VCFtools. Bioinformatics.
27:2156–2158.

Danecek P., Bonfield J.K., Liddle J., Marshall J., Ohan V., Pollard M.O., Whitwham A.,
Keane T., McCarthy S.A., Davies R.M., Li H. 2021. Twelve years of SAMtools and
BCFtools. GigaScience. 10:giab008.

De Jode A., Le Moan A., Johannesson K., Faria R., Stankowski S., Westram A., Butlin
R., Rafajlović M., Fraïsse C. 2023. Ten years of demographic modelling of
divergence and speciation in the sea. Evol. App. 16:542–559.

DeRaad D.A. 2022. snpfiltr: An R package for interactive and reproducible SNP filtering.
Mol. Ecol. Resour. 22:2443–2453.

Donelson J.M., Sunday J.M., Figueira W.F., Gaitán-Espitia J.D., Hobday A.J., Johnson
C.R., Leis J.M., Ling S.D., Marshall D., Pandolfi J.M., Pecl G., Rodgers G.G., Booth
D.J., Munday P.L. 2019. Understanding interactions between plasticity, adaptation
and range shifts in response to marine environmental change. Phil. Trans. R. Soc.
B. 374:20180186.

Donovan L.A., Rosenthal D.R., Sanchez-Velenosi M., Rieseberg L.H., Ludwig F. 2010.
Are hybrid species more fit than ancestral parent species in the current hybrid
species habitats? J. Evol. Biol. 23:805–816.

Duranton M., Allal F., Valière S., Bouchez O., Bonhomme F., Gagnaire P.-A. 2020. The
contribution of ancient admixture to reproductive isolation between European sea
bass lineages. Evol. Lett. 4:226–242.

Earl D.A., vonHoldt B.M. 2012. STRUCTURE HARVESTER: a website and program for
visualizing STRUCTURE output and implementing the Evanno method. Conserv.
Genet. Resour. 4:359–361.

Elgvin T.O., Trier C.N., Tørresen O.K., Hagen I.J., Lien S., Nederbragt A.J., Ravinet M.,
Jensen H., Sætre G.-P. 2017. The genomic mosaicism of hybrid speciation. Sci. Adv.
3:e1602996.

Elmer K.R., Meyer A. 2011. Adaptation in the age of ecological genomics: insights from
parallelism and convergence. Trends Ecol. Evol. 26:298–306.

Evanno G., Regnaut S., Goudet J. 2005. Detecting the number of clusters of individuals
using the software STRUCTURE: a simulation study. Mol. Ecol. 14:2611–2620.

Excoffier L., Dupanloup I., Huerta-Sánchez E., Sousa V.C., Foll M. 2013. Robust
demographic inference from genomic and SNP data. PLOS Genet. 9:e1003905.

Excoffier L., Foll M., Petit R.J. 2009. Genetic consequences of range expansions. Annu.
Rev. Ecol. Evol. Syst. 40:481–501.

Excoffier L., Lischer H.E.L. 2010. Arlequin suite ver 3.5: a new series of programs to
perform population genetics analyses under Linux and Windows. Mol. Ecol. Resour.
10:564–567.

Excoffier L., Marchi N., Marques D.A., Matthey-Doret R., Gouy A., Sousa V.C. 2021.
fastsimcoal2: demographic inference under complex evolutionary scenarios.
Bioinformatics. 37:4882–4885.

Fitzpatrick B.M. 2012. Estimating ancestry and heterozygosity of hybrids using molecular
markers. BMC Evol. Biol. 12:131.

Francis R.M. 2017. pophelper: an R package and web app to analyse and visualize
population structure. Mol. Ecol. Resour. 17:27–32.

Froese   R.,   Pauly   D.   2023.   Fishbase.   Available   from
https://www.fishbase.se/popdyn/KeyfactsSummary_1.php?ID=384&GenusName=
Megalaspis&SpeciesName=cordyla&vStockCode=398&fc=314.

Gaither M.R., Bernal M.A., Fernandez-Silva I., Mwale M., Jones S.A., Rocha C., Rocha
L.A. 2015. Two deep evolutionary lineages in the circumtropical glasseye
Heteropriacanthus cruentatus (Teleostei, Priacanthidae) with admixture in the south-
western Indian Ocean. J. Fish Biol. 87:715–727.

Gaither M.R., Rocha L.A. 2013. Origins of species richness in the Indo-Malay-Philippine
biodiversity hotspot: evidence for the centre of overlap hypothesis. J. Biogeogr.
40:1638–1648.

Gerking S. D. 1994. Feeding Ecology of Fish. San Diego: Academic Press.

Gibbs R.H., Collette B.B. 1967. Comparative anatomy and systematics of the tunas,
genus *Thunnus*. Fish. Bull. 66:65–130.

Glass J.R., Harrington R.C., Cowman P.F., Faircloth B.C., Near T.J. 2023. Widespread
sympatry in a species-rich clade of marine fishes (Carangoidei). Proc. R. Soc. B.
290:20230657.

Gompert Z., Fordyce J.A., Forister M.L., Shapiro A.M., Nice C.C. 2006. Homoploid hybrid
speciation in an extreme habitat. Science. 314:1923–1925.

Gross B.L., Rieseberg L.H. 2005. The ecological genetics of homoploid hybrid speciation.
J. Hered. 96:241–252.

Hall T.A. 1999. BioEdit : a user-friendly biological sequence alignment editor and analysis
program for Windows 95/98/NT. Nucleic Acids. Symp. Ser. 41:95–98.

Hewitt G.M. 2000. The genetic legacy of the Quaternary ice ages. Nature. 405:907–913.

Hewitt G.M. 2004. Genetic consequences of climatic oscillations in the Quaternary. Phil.
Trans. R. Soc. B. 359:183–195.

Hirase S. 2022. Comparative phylogeography of coastal gobies in the Japanese
Archipelago: future perspectives for the study of adaptive divergence and speciation.
Ichthyol. Res. 69:1–16.

Hirase S., Tezuka A., Nagano A.J., Sato M., Hosoya S., Kikuchi K., Iwasaki W. 2021.
Integrative genomic phylogeography reveals signs of mitonuclear incompatibility in
a natural hybrid goby population. Evolution. 75:176–194.

Hofreiter M., Stewart J. 2009. Ecological change, range fluctuations and population
dynamics during the Pleistocene. Curr. Biol. 19:R584–R594.

Huson D.H., Bryant D. 2006. Application of phylogenetic networks in evolutionary studies.
Mol. Biol. Evol. 23:254–267.

Ivanova N.V., Zemlak T.S., Hanner R.H., Hebert P.D.N. 2007. Universal primer cocktails
for fish DNA barcoding. Mol. Ecol. Notes. 7:544–548.

Jeffery N.W., Stanley R.R.E., Wringe B.F., Guijarro-Sabaniel J., Bourret V., Bernatchez
L., Bentzen P., Beiko R.G., Gilbey J., Clément M., Bradbury I.R. 2017. Range-wide
parallel climate-associated genomic clines in Atlantic salmon. Roy. Soc. Open Sci.
4:171394.

Jombart T. 2008. Adegenet: A R package for the multivariate analysis of genetic markers.
Bioinformatics. 24:1403–1405.

Jones F.C., Grabherr M.G., Chan Y.F., Russell P., Mauceli E., Johnson J., Swofford R.,
Pirun M., Zody M.C., White S., Birney E., Searle S., Schmutz J., Grimwood J.,
Dickson M.C., Myers R.M., Miller C.T., Summers B.R., Knecht A.K., Brady S.D.,
Zhang H., Pollen A.A., Howes T., Amemiya C., Lander E.S., Di Palma F., Lindblad-
Toh K., Kingsley D.M. 2012. The genomic basis of adaptive evolution in threespine
sticklebacks. Nature. 484:55–61.

Kang J.H., Schartl M., Walter R.B., Meyer A. 2013. Comprehensive phylogenetic
analysis of all species of swordtails and platies (Pisces: Genus *Xiphophorus*)
uncovers a hybrid origin of a swordtail fish, *Xiphophorus monticolus*, and
demonstrates that the sexually selected sword originated in the ancestral lineage of
the genus, but was lost again secondarily. BMC. Evol. Biol. 13:25.

Kiessling W., Simpson C., Beck B., Mewis H., Pandolfi J.M. 2012. Equatorial decline of
reef corals during the last Pleistocene interglacial. Proc. Natl. Acad. Sci. USA.
109:21378–21383.

Kulmuni J., Wiley B., Otto S.P. 2024. On the fast track: hybrids adapt more rapidly than
parental populations in a novel environment. Evol. Lett. 8:128–136.

Kuriiwa K., Chiba S.N., Motomura H., Matsuura K. 2014. Phylogeography of Blacktip
Grouper, *Epinephelus fasciatus* (Perciformes: Serranidae), and influence of the
Kuroshio Current on cryptic lineages and genetic population structure. Ichthyol. Res.
61:361–374.

Lamichhaney S., Han F., Webster M.T., Andersson L., Grant B.R., Grant P.R. 2018. Rapid
hybrid speciation in Darwin's finches. Science. 359:224–228.

Larkin M.A., Blackshields G., Brown N.P., Chenna R., Mcgettigan P.A., McWilliam H.,
Valentin F., Wallace I.M., Wilm A., Lopez R., Thompson J.D., Gibson T.J., Higgins
D.G. 2007. Clustal W and Clustal X version 2.0. Bioinformatics. 23:2947–2948.

Larsen P.A., Marchán-Rivadeneira M.R., Baker R.J. 2010. Natural hybridization
generates mammalian lineage with species characteristics. Proc. Natl. Acad. Sci.
USA. 107:11447–11452.

Lê S., Josse J., Husson F. 2008. FactoMineR: An R Package for Multivariate Analysis. J.
Stat. Soft. 25:1–18.

Leigh J.W., Bryant D. 2015. POPART: Full-feature software for haplotype network
construction. Methods Ecol. Evol. 6:1110–1116.

Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-
MEM. arXiv. 1303.3997v2:1–3.

Lischer H.E.L., Excoffier L. 2012. PGDSpider: an automated data conversion tool for
connecting population genetics and genomics programs. Bioinformatics. 28:298–
299.

Liu M., Lin L., Gao T., Yanagimoto T., Sakurai Y., Grant W.S. 2012. What maintains the
central North Pacific genetic discontinuity in Pacific Herring ? PLOS ONE. 7:e50340.

Liu X., Fu Y.-X. 2020. Stairway Plot 2: demographic history inference with folded SNP
frequency spectra. Genome Biol. 21:280.

Lollar M.J., Biewer-Heisler T.J., Danen C.E., Pool J.E. 2023. Hybrid breakdown in male
reproduction between recently diverged *Drosophila melanogaster* populations has a
complex and variable genetic architecture. Evolution. 77:1550–1563.

Long Z., Rieseberg L.H. 2024. Documenting homoploid hybrid speciation. Mol. Ecol.
n/a:e17412.

Lopes F., Oliveira L.R., Beux Y., Kessler A., Cárdenas-Alayza S., Majluf P., Páez-Rosas
D., Chaves J., Crespo E., Brownell R.L., Baylis A.M.M., Sepúlveda M., Franco-Trecu
V., Loch C., Robertson B.C., Peart C.R., Wolf J.B.W., Bonatto S.L. 2023. Genomic
evidence for homoploid hybrid speciation in a marine mammal apex predator. Sci.
Adv. 9:eadf6601.

MacPherson A., Wang S., Yamaguchi R., Rieseberg L.H., Otto S.P. 2022. Parental
population range expansion before secondary contact promotes heterosis. Am. Nat.
200:E1–E15.

Maier P.A., Vandergast A.G., Ostoja S.M., Aguilar A., Bohonak A.J. 2019. Pleistocene
glacial cycles drove lineage diversification and fusion in the Yosemite toad (*Anaxyrus*
canorus). Evolution. 73:2476–2496.

Mallet J. 2007. Hybrid speciation. Nature. 446:279–283.

Marr J.C., Schaefer F.B. 1949. Definitions of body dimensions used in describing tunas.
Fish. Bull. Fish. Wild. Serv. 51:241–244.

Mavárez J., Linares M. 2008. Homoploid hybrid speciation in animals. Mol Ecol.
17:4181–4185.

Meier J.I., Sousa V.C., Marques D.A., Selz O.M., Wagner C.E., Excoffier L., Seehausen
O. 2017. Demographic modelling with whole-genome data reveals parallel origin of
similar *Pundamilia* cichlid species after hybridization. Mol. Ecol. 26:123–141.

Motomura H., Matsunuma M. 2022. Fish diversity along the Kuroshio Current. In:
Yoshiaki K., Motomura H., Matsuura K., editors. Fish Diversity of Japan. Singapore:
Springer Nature Singapore. p. 63–78.

Muto N., Alama U.B., Hata H., Guzman A.M.T., Cruz R., Gaje A., Traifalgar R.F.M.,
Kakioka R., Takeshima H., Motomura H., Muto F., Babaran R.P., Ishikawa S. 2016.
Genetic and morphological differences among the three species of the genus
Rastrelliger (Perciformes: Scombridae). Ichthyol. Res. 63:275–287.

Muto N., Kai Y. 2023. Allopatric origin, secondary contact and subsequent isolation of
sympatric rockfishes (Sebastidae: *Sebastes*) in the north-western Pacific. Biol. J.
Linn. Soc. 138:37–50.

Muto N., Kakioka R., Ghaffar M.A., Quan N.V., Vilasri V., Traifalgar R.F.M., Babaran R.P.,
Muto F., Takeshima H., Motomura H., Ishikawa S. 2021. Fine-scale phylogeography

of coastal fishes in the South China Sea: Possible roles of biological traits and
geography. J. Biogeogr. 48:2785–2800.

Neiva J., Assis J., Fernandes F., Pearson G.A., Serrão E.A. 2014. Species distribution
models and mitochondrial DNA phylogeography suggest an extensive
biogeographical shift in the high-intertidal seaweed *Pelvetia canaliculata*. J.
Biogeogr. 41:1137–1148.

Nieto Feliner G., Álvarez I., Fuertes-Aguilar J., Heuertz M., Marques I., Moharrek F.,
Piñeiro R., Riina R., Rosselló J.A., Soltis P.S., Villa-Machío I. 2017. Is homoploid
hybrid speciation that rare? An empiricist's view. Heredity. 118:513–516.

Noguerales V., Ortego J. 2022. Genomic evidence of speciation by fusion in a recent
radiation of grasshoppers. Evolution. 76:2618–2633.

Olave M., Nater A., Kautt A.F., Meyer A. 2022. Early stages of sympatric homoploid
hybrid speciation in crater lake cichlid fishes. Nat. Commun. 13:5893.

Orr H.A. 1995. The population genetics of speciation: the evolution of hybrid
incompatibilities. Genetics. 139:1805–1813.

Patterson N., Moorjani P., Luo Y., Mallick S., Rohland N., Zhan Y., Genschoreck T.,
Webster T., Reich D. 2012. Ancient admixture in human history. Genetics. 192:1065–
1093.

Peñalba J.V., Runemark A., Meier J.I., Singh P., Wogan G.O.U., Sánchez-Guillén R.,
Mallet J., Rometsch S.J., Menon M., Seehausen O., Kulmuni J., Pereira R.J. 2024.
The role of hybridization in species formation and persistence. Cold Spring Harb.
Perspect. Biol. doi: 10.1101/cshperspect.a041445

Petr M., Vernot B., Kelso J. 2019. admixr—R package for reproducible analyses using
ADMIXTOOLS. Bioinformatics. 35:3194–3195.

Pickett B.D., Glass J.R., Johnson T.P., Ridge P.G., Kauwe J.S.K. 2022. The genome of
a giant (trevally): *Caranx ignobilis*. Gigabyte. 2022:1–16.

Pritchard J.K., Stephens M., Donnelly P. 2000. Inference of population structure using
multilocus genotype data. Genetics. 155:945–959.

R Core Team. 2022. R: A language and environment for statistical computing.
https://www.r-project.org/.

Rieseberg L.H., Raymond O., Rosenthal D.M., Lai Z., Livingstone K., Nakazato T.,
Durphy J.L., Schwarzbach A.E., Donovan L.A., Lexer C. 2003. Major ecological
transitions in wild sunflowers facilitated by hybridization. Science. 301:1211–1216.

Rosser N., Seixas F., Queste L.M., Cama B., Mori-Pezo R., Kryvokhyzha D., Nelson M.,
Waite-Hudson R., Goringe M., Costa M., Elias M., Mendes Eleres de Figueiredo C.,
Freitas A.V.L., Joron M., Kozak K., Lamas G., Martins A.R.P., McMillan W.O., Ready
J., Rueda-Muñoz N., Salazar C., Salazar P., Schulz S., Shirai L.T., Silva-Brandão
K.L., Mallet J., Dasmahapatra K.K. 2024. Hybrid speciation driven by multilocus
introgression of ecological traits. Nature. 628:811–817.

Runemark A., Vallejo-Marin M., Meier J.I. 2019. Eukaryote hybrid genomes. PLOS Genet.
15:e1008404.

Schumer M., Rosenthal G.G., Andolfatto P. 2014. How common is homoploid hybrid
speciation? Evolution. 68:1553–60.

Schwarz D., Matta B.M., Shakir-Botteri N.L., McPheron B.A. 2005. Host shift to an
invasive plant triggers rapid animal hybrid speciation. Nature. 436:546–549.

Seersholm F.V., Werndly D.J., Grealy A., Johnson T., Keenan Early E.M., Lundelius E.L.,
Winsborough B., Farr G.E., Toomey R., Hansen A.J., Shapiro B., Waters M.R.,
McDonald G., Linderholm A., Stafford T.W., Bunce M. 2020. Rapid range shifts and
megafaunal extinctions associated with late Pleistocene climate change. Nat.
Commun. 11:2770.

Smith-Vaniz W.F. 1999. Carangidae. In: Carpenter K.E., Niem V.H., editors. THE LIVING
MARINE RESOURCES OF THE WESTERN CENTRAL PACIFIC Volume 4: Bony
fishes part 2 (Mugilidae to Carangidae). Rome: FAO. p. 2659–2756.

Soltis D.E., Morris A.B., McLACHLAN J.S., Manos P.S., Soltis P.S. 2006. Comparative
phylogeography of unglaciated eastern North America. Mol. Ecol. 15:4261–4293.

Stange M., Sánchez-Villagra M.R., Salzburger W., Matschiner M. 2018. Bayesian
divergence-time estimation with genome-wide single-nucleotide polymorphism data
of sea catfishes (Ariidae) supports Miocene closure of the Panamanian Isthmus.
Syst. Biol. 67:681–699.

Stelkens R.B., Schmid C., Seehausen O. 2015. Hybrid breakdown in cichlid fish. PLOS
ONE. 10:e0127207.

Su Y.-C., Su S.-H., Li H.-Y., Wang H.-Y., Lee S.-C. 2020. Implication of single year
seasonal sampling to genetic diversity fluctuation that coordinates with
oceanographic dynamics in torpedo scads near Taiwan. Sci. Rep. 10:16829.

Suyama Y., Hirota S.K., Matsuo A., Tsunamoto Y., Mitsuyuki C., Shimura A., Okano K.
2022. Complementary combination of multiplex high-throughput DNA sequencing
for molecular phylogeny. Ecol. Res. 37:171–181.

Suyama Y., Matsuki Y. 2015. MIG-seq: an effective PCR-based method for genome-wide
single-nucleotide polymorphism genotyping using the next-generation sequencing
platform. Sci. Rep. 5:16963.

Taylor S.A., Larson E.L. 2019. Insights from genomes into the evolutionary importance
and prevalence of hybridization in nature. Nat. Ecol. Evol. 3:170–177.

Teng W., Fu H., Li Z., Zhang Q., Xu C., Yu H., Kong L., Liu S., Li Q. 2023. Parallel
evolution in *Crassostrea* oysters along the latitudinal gradient is associated with
variation in multiple genes involved in adipogenesis. Mol. Ecol. 32:5276–5287.

Tine M., Kuhl H., Gagnaire P.-A., Louro B., Desmarais E., Martins R.S.T., Hecht J.,
Knaust F., Belkhir K., Klages S., Dieterich R., Stueber K., Piferrer F., Guinand B.,
Bierne N., Volckaert F.A.M., Bargelloni L., Power D.M., Bonhomme F., Canario
A.V.M., Reinhardt R. 2014. European sea bass genome and its variation provide
insights into adaptation to euryhalinity and speciation. Nat. Commun. 5:1–10.

Wang S., Ore M.J., Mikkelsen E.K., Lee-Yaw J., Toews D.P.L., Rohwer S., Irwin D. 2021.
Signatures of mitonuclear coevolution in a warbler species complex. Nat. Commun.
12:1–11.

Wood T.E., Takebayashi N., Barker M.S., Mayrose I., Greenspoon P.B., Rieseberg L.H.
2009. The frequency of polyploid speciation in vascular plants. Proc. Natl. Acad. Sci.
USA. 106:13875–13879.

Wright M.N., Ziegler A. 2017. ranger: a fast implementation of Random Forests for high
dimensional data in C++ and R. J. Stat. Soft. 77:1–17.

Yakimowski S.B., Rieseberg L.H. 2014. The role of homoploid hybridization in evolution:
A century of studies synthesizing genetics and ecology. Am. J. Bot. 101:1247–1258.

[dataset]* Santos S.R. 2014. Data from: "Complete mitochondrial and partial nuclear
genomes for the jack species Caranx ignobilis (Forsskål, 1775) and C. melampygus
(Cuvier, 1833) (Perciformes: Carangidae) from the High Hawaiian Islands" in
Genomic Resources Notes accepted 1 October 2013 – 30 November 2013. Dryad.
https://doi.org/10.5061/dryad.6gr7t.