Phylogenomics, Classification, and Lifestyle Evolution in Raft- and Nursery Web-Spiders (Araneae: Dolomedidae and Pisauridae)

Kuang-Ping Yu[1,2,*,+], Ren-Chung Cheng[3,4,*], Charles R. Haddad[5], Akio Tanikawa[6], Brogan L. Pett[7,8], Luis N. Piacentini[9], Peter Jäger[10], Ho Yin Yip[11], Yuya Suzuki[12], Arnaud Henrard[13], Christina J. Painting[14,15], Cor J. Vink[16], Eileen A. Hebets[17], Mark S. Harvey[18,19], Matjaž Kuntner[1,2,20,21,22,+]

[1]Department of Organisms and Ecosystems Research, National Institute of Biology, Večna pot 121, 1000 Ljubljana, Slovenia.

[2]Department of Biology, Biotechnical Faculty, University of Ljubljana, Večna pot 111, 1000 Ljubljana, Slovenia.

[3]Department of Life Sciences, National Chung Hsing University, No. 145, Xingda Rd., South Dist., 40227 Taichung, Taiwan.

[4]Research Center for Global Change Biology, National Chung Hsing University, No. 145, Xingda Rd., South Dist., 40227 Taichung, Taiwan.

[5]Department of Zoology and Entomology, University of the Free State, P.O. Box 339, 9300 Bloemfontein, South Africa.

[6]Laboratory of Biodiversity Science, School of Agriculture and Life Sciences, The University of Tokyo, Japan.

[7]SpiDiverse, Biodiversity Inventory for Conservation (BINCO), 3380 Walmersumstraat,

Glabbeek, Belgium.

[8]Centre for Ecology and Conservation, University of Exeter, Penryn Campus, Penryn,

Cornwall, UK.

[9]Arachnology Division, Argentine Museum of Natural Science "Bernardino Rivadavia"

CONICET, Av. Angel Gallardo 470, C1405DJR, Buenos Aries, Argentina.

[10]Senckenberg Research Institute, Arachnology, Mertonstraße 17-21, 60325 Frankfurt am

Main, Germany

[11] Discovery & Education Department, Ocean Park Hong Kong, 180 Wong Chuk Hang Road,

Aberdeen, Hong Kong.

[12]Tokushima Prefectural Museum, Mukoterayama, Tokushima City, Tokushima Prefecture

(post code 770-8070), Japan.

[13]Royal Museum of Central Africa, Leuvensesteenweg 13, B-3080, Tervuren, Belgium.

[14]Te Aka Mātuatua School of Science, University of Waikato, New Zealand.

[15]Te Pūnaha Matatini, Centre of Research Excellence, New Zealand.

[16]Department of Pest-management and Conservation, Lincoln University, Lincoln, New

Zealand.

[17]School of Biological Sciences, University of Nebraska-Lincoln, Lincoln, NE 68588, USA.

[18]Collections & Research, Western Australian Museum, 49 Kew Street, Welshpool, Western

Australia 6106, Australia.

[19]School of Biological Sciences, University of Western Australia, Crawley, Western Australia

6009, Australia.

[20]Jovan Hadži Institute of Biology, ZRC SAZU, Novi trg 2, 1000 Ljubljana, Slovenia.

[21]Department of Entomology, National Museum of Natural History, Smithsonian Institution,

10th and Constitution, NW, Washington, DC 20560-0105, USA.

[22]State Key Laboratory of Biocatalysis and Enzyme Engineering, and Centre for Behavioural

Ecology and Evolution, School of Life Sciences, Hubei University, Hubei, China.

* = shared first authorship.

+ = corresponding authors: Department of Organisms and Ecosystems Research, National

Institute of Biology, Večna pot 121, 1000 Ljubljana, Slovenia; Kuang-Ping.Yu@nib.si; +386

(0)59 232 701.

ABSTRACT

Pisauridae Simon, 1890 or "nursery web spiders" are a global and heterogenous

assemblage of spider genera with diverse lifestyles, containing web builders and webless

species, as well as terrestrial and semi-aquatic species, notably "fishing spiders", genus

Dolomedes Latreille, 1804. Incomplete, unresolved, or conflicting phylogenies have so far

hampered testing for *Dolomedes* and pisaurid monophyly and evolution. Here, we broadly

address these questions within a phylogenomic and comparative framework. Our goals are i)

reconstruction of a robust phylogeny to test the monophyly of *Dolomedes* and Pisauridae and

to amend *Dolomedes* classification; ii) estimation of evolutionary shifts and trends in

lifestyles and capture webs; and iii) evaluation of hypotheses of morphological trait

association with a semi-aquatic lifestyle. To this end we generate subgenomic data

(ultraconserved elements or UCE) for 53 *Dolomedes* species and 28 pisaurid genera. We

analyze these data using maximum likelihood, Bayesian, and multi-species coalescence

approaches, as well as using two different phylogenetic time calibration methods, RelTime

and MCMCtree. Consistent across analytical approaches, our phylogenies reject the

monophyly of both Pisauridae and *Dolomedes*. "Pisaurid" genera fall into three clades: 1)

Focal Clade I groups the majority, including *Pisaura* Simon, 1886, hence representing true

pisaurids; 2) Focal Clade II = *Blandinia* Tonini et al., 2016 is sister to Trechaleidae Simon,

1890 and Lycosidae Sundevall, 1833; 3) Focal Clade III with fishing and raft spiders groups

Dolomedes, *Megadolomedes* Davies and Raven, 1980, and *Ornodolomedes* Raven and

Hebron, 2018 and is sister to Focal Clade II, Trechaleidae, and Lycosidae. Our taxonomy,

based on complementary taxa and morphological evidence, resurrects Dolomedidae Simon,

1876 to include *Dolomedes* and the Oceanic genera *Bradystichus* Simon, 1884,

Megadolomedes, *Caledomedes* Raven and Hebron, 2018, *Mangromedes* Raven and Hebron,

2018, *Ornodolomedes*, and *Tasmomedes* Raven and Hebron, 2018. Both RelTime and

MCMCtree analyses yield comparable divergence estimations: Pisauridae origin is estimated

between 29 and 40 Ma; *Blandinia* between 21 and 34 Ma; Dolomedidae between 10 and 17

Ma; and *Dolomedes* between 9 and 16 Ma. In order to avoid misleading significant

correlations and/or over-resolved ancestral states, we performed taxon sampling bias

correction in all evolutionary analyses. Evolutionary analyses reconstruct semi-aquatic

lifestyle as ancestral to a large clade containing pisaurids, lycosids, trechaleids, *Blandinia*,

and dolomedids, with several reversals to terrestrial lifestyle. Capture webs evolved at least

three times, with reversals. Counter to expectation, the evolution of lifestyles and capture

webs are independent. Although leg and tarsus lengths do not indicate lifestyles, semi-aquatic

taxa are significantly larger than terrestrial ones. We explain this pattern with a biomechanical

threshold over which surface tension can be broken while spiders forage under water. Our

time-calibrated analyses indicate that the evolution of terrestrial and web-building lifestyles

from semi-aquatic ancestors in Pisauridae coincided with cooling and drying climates in the

mid-Miocene. We therefore hypothesize that climatic changes have acted as strong selection

pressures toward lifestyle diversification.

Keywords: Fishing spiders, semi-aquatic spiders, ultraconserved elements, ancestral state

reconstruction, comparative analysis, taxon sampling bias

INTRODUCTION

Known as raft or fishing spiders, *Dolomedes* Latreille, 1804 (Figs. 1a–b) is a globally

distributed genus with 105 species of large wandering spiders, currently under the family

Pisauridae Simon, 1890 (World Spider Catalog 2024). *Dolomedes* species are well known for

their semi-aquatic lifestyle as most of them inhabit a variety of waterbodies. Their ecology is

accompanied by remarkable behaviors such as rafting on the water surface, diving under the

surface (McAlister 1960; Deshefy 1981; Suter 1999), and, quite unusual for spiders, preying

on aquatic vertebrates (Williams 1979; McCormick and Polis 1982; Bleckmann and Lotz

1987; Zimmermann and Spence 1989; Nyffeler and Pusey 2014; Baba et al. 2019; Nyffeler

and Gibbons 2022). *Dolomedes* species are therefore popular models for studying semi-

aquatic adaptations in arthropods, focusing on neurosensing of water ripples (Roland and

Rovner 1983; Bleckmann and Barth 1984; Bleckmann et al. 1994; Suter 2003), locomotion

on water (Suter and Wildman 1999; Suter and Gruenwald 2000a, 2000b; Stratton et al. 2004;

Suter 2013), as well as the associated morphologies (e.g., appendage proportions, setae

morphologies and densities, cuticle structures, and respiratory organs; see Suter et al. 2004;

Stratton and Suter 2009; Stratton et al. 2004; see also Lapinski et al. 2015).

Although most *Dolomedes* species strongly depend on freshwater-associated habitats

(Carico 1973a; Zhang et al. 2004; Tanikawa and Miyashita 2008; Vink and Dupérré 2010;

Raven and Hebron 2018; Yu and Kuntner 2024), not all species are semi-aquatic. For

example, *D. sulfureus* L. Koch, 1878 (Fig. 1b), *D. silvicola* Tanikawa and Miyashita, 2008,

D. tenebrosus Hentz, 1844, *D. albineus* Hentz, 1845, *D. minor* L. Koch, 1876, and *D.*

schauinslandi Simon, 1899, are reported to be terrestrial, and largely independent of water

bodies (Carico 1973a; Tanikawa and Miyashita 2008; Ono 2009; Vink and Dupérré 2010).

Their distinct morphologies and disparate distributions in Asia, North America, and New

Zealand suggest that these terrestrial species are not each other's closest relatives. The

currently understood lifestyle diversity within the genus raises questions of how, when,

where, and why terrestrial versus semi-aquatic lifestyles evolved in *Dolomedes*. The lack of a

comprehensive *Dolomedes* species phylogeny currently precludes answering these questions.

Similar questions related to the evolution of lifestyles can be asked more broadly about

phylogenetic proximity of *Dolomedes*, i.e. to the family Pisauridae, whose constituent taxa

display an even more complex array of lifestyle and trait diversity (Sierwald 1997; Zhang et

al. 2004; Santos 2007a; Ono 2009; Jäger 2011; Raven and Hebron 2018; Dippenaar-

Schoeman et al. 2020). With 52 genera and 365 species (World Spider Catalog 2024), this

diverse global family is thought to be defined by the construction of nursery webs to protect

their spiderlings (Fig. 1c). As adults, most pisaurid genera are terrestrial wandering spiders

(Figs. 1b–f), others construct sheet or three-dimensional capture webs of numerous shapes

and sizes in terrestrial habitats (Figs. 1g–l), and some are semi-aquatic (Figs. 1a, m–p) (Table

1). This phenotypic and ecological diversity makes pisaurids an ideal model group for

studying the evolution of lifestyles and capture webs, yet their evolution has not been

adequately analyzed due to unknown or conflicting phylogenies (Griswold 1993; Santos

2007b; Bayer and Schönhofer 2013; Moradmand et al. 2014; Polotow et al. 2015; Albo et al.

2017; Wheeler et al. 2017; Cheng and Piel 2018; Fernández et al. 2018; Piacentini and

Ramírez 2019; Kallal et al. 2021; Hazzi and Hormiga 2023; Kulkarni et al. 2023) (Table 2;

see also Taxonomy). Here, we address these issues within an original phylogenomic-

evolutionary framework.

Our first goal is to test the monophyly of *Dolomedes* and Pisauridae family, as well as

the phylogenetic position of *Dolomedes*, using phylogenomic data, and to use these results to

update the classification of *Dolomedes*. Although the monophyly of Pisauridae and the

affiliation of *Dolomedes* have been tested multiple times under different frameworks (see

citations above), none of the recent studies covered more than a quarter of the 52 pisaurid

genera (Table 2). Limited taxon coverage has also led to an unclear topology within

Dolomedes, the most speciose "pisaurid" genus (World Spider Catalog 2024). So far,

Dolomedes species phylogenies have been regional and based on a few Sanger loci (Vink and

Dupérré 2010; Tanikawa 2012; Yu and Kuntner, 2024). For example, a species level

phylogeny of *Dolomedes* used a single gene fragment for 13 Japanese species (Tanikawa

2012) and failed to resolve all the nodes. Considering that Raven and Hebron's (2018)

taxonomic review of Australian pisaurids increased the number of *Dolomedes*' close relatives

from two to six genera with different degrees of intra- and intergeneric morphological

variation, a better sampled *Dolomedes* phylogeny is also needed to test the boundaries of this

diverse genus. We therefore set about to reconstruct a robust and well-sampled global

phylogeny based on ultraconserved elements (UCE; Faircloth et al. 2012, 2013, 2015;

Faircloth 2017; Miles Zhang et al. 2019) data and including more than half of the *Dolomedes*

species as well as other genera currently in Pisauridae. To test the robustness of the

phylogenetic results, we utilize maximum likelihood, Bayesian, and multi-species

coalescence approaches. In order to add a historic time component to the evolution of the

target clades, we also subject the phylogenomic data to two different time calibration

methods, *a posteriori* (RelTime; Tamura et al. 2012; 2018) and *a priori* (MCMCtree; Yang

2007) (see Beavan et al. 2020).

Our second goal is to utilize the newly obtained phylogeny to reconstruct the evolution

of lifestyles in *Dolomedes* and Pisauridae. To this end, we first populate the data for the

terminals in the phylogeny on semi-aquatic vs. terrestrial lifestyles as well as the

presence/absence of capture webs. Next, we test: i) whether a semi-aquatic lifestyle is

ancestral in *Dolomedes* but not in Pisauridae, as suggested by a prior phylogenetic study of

Lycosoidea (Piacentini and Ramírez 2019); ii) whether the absence of a capture web is

ancestral in *Dolomedes* and Pisauridae (Piacentini and Ramírez 2019); and iii) if lifestyles

and capture webs are correlated traits. This third, original hypothesis, predicts that evolution

of a semi-aquatic lifestyle excludes the presence of a capture web.

Our third goal is to test the hypotheses that body size, leg length, and tarsus length all

relate to a semi-aquatic lifestyle. Although researchers have analyzed how spider

morphologies differ across foraging guilds on the spider tree of life (Wolff et al. 2022), semi-

aquatic lifestyle was not considered as a separate guild and was therefore not discussed. To

fill this knowledge gap, we test the correlation between a semi-aquatic lifestyle and

morphological traits on the phylogeny. Although there are no rigorous empirical comparisons,

the literature suggests that several semi-aquatic spiders exhibit iv) larger body sizes

(*Ancylometes* Bertkau 1880, see Lapinski et al. 2015); v) relatively shorter legs (Tanikawa

and Miyashita 2008); and vi) relatively longer tarsi (Lapinski et al. 2015) compared to

terrestrial species (non-pisaurid species). We test these hypotheses on the phylogeny of

Dolomedes and Pisauridae.

MATERIALS & METHODS

Phylogenetic taxon sampling

Phylogenetic taxon sampling comprised 96 terminals, including 53 *Dolomedes*

morphospecies and 31 pisaurid species of 28 genera (of which one is without a certain genus

affiliation, Fig. 1d) as ingroups (Table 3). Each pisaurid genus was represented by only one

species, except for the diverse genera occupying more than one biogeographic realm:

Dendrolycosa Doleschall, 1859*, Nilus* O. Pickard-Cambridge, 1876*,* and *Hygropoda* Thorell,

1895 were thus each represented by two species, one per biogeographic realm (Table 3). As

outgroups to Pisauridae and *Dolomedes*, we used representatives of Trechaleidae Simon,

1890, Lycosidae Sundevall, 1833, Zoropsidae Bertkau, 1882, Psechridae Simon, 1890,

Oxyopidae Thorell, 1869, Thomisidae Sundevall, 1833, and Viridasiidae Lehtinen, 1967

(Table 3).

Lifestyle, capture web, and morphological data

We scored lifestyle and capture web as binary characters: semi-aquatic versus terrestrial

lifestyle, and capture web presence versus absence (Table 3). A semi-aquatic lifestyle in our

study refers to the adult spiders being documented to inhabit any water bodies in direct

contact with the water. In the absence of such documentation, species are taken to be

terrestrial. The presence of a capture web was only scored such in those species for which

literature documents a prey capturing web (not merely webbing, nursery web, or nest; see

Eberhard 2020) in adults. In the absence of such information for adults, species were scored

as webless. We obtained data on carapace width (CW), leg I length (L1), and tarsus I length

(T1), either by microscopic measuring or from the literature. We used CW to represent the

size of a spider while relative leg and tarsus lengths were calculated by dividing L1 by CW

and T1 by L1, respectively. We emphasized the measurement of adult females, but in the four

cases where females were not available, we measured males instead (Table 3). This

introduces little bias, as most pisaurids are not extremely sexually size dimorphic (Kuntner

and Coddington 2020). Measurements were taken using a Leica M205C stereomicroscope or

a Keyence VHX7000 digital microscope. For the two genera whose phylogenetic terminals

were represented by juveniles and therefore lacked precise species identification, we

averaged female measurements from the literature (Table 3).

DNA extraction and phylogenomic pipeline

We used one to four legs from each specimen for DNA extraction. For small spiders, we

also used the whole prosoma. We extracted genomic DNA using QIAamp DNA Micro Kits

(Qiagen, Hilden, Germany), with the DNA quality checked using electrophoresis and a

Qubit™ 4 Fluorometer (Invitrogen, Carlsbad, CA, USA) with the dsDNA HS Assay Kit.

DNA samples containing more than 320 ng of DNA were then diluted to equal volumes and

sent to Rapid Genomics LLC (Gainesville, FL, USA) for library preparation to capture and

sequence UCE data. We used the RTA spider probe set (Zhang et al. 2023) for capturing UCE

loci.

To assemble and filter the UCE matrix we employed the Phyluce v1.7.3 pipeline

(Faircloth 2016) following the online tutorial

(https://phyluce.readthedocs.io/en/latest/tutorials/index.html). After trimming the adaptors,

we assembled the data using the package's command lines under the SPAdes de novo

assembly algorithm (Prjibelski et al. 2020). UCE loci were then found and extracted by

matching the assembled contigs to the RTA spider probe set (Zhang et al. 2023). We ran both

edges and internal trimming on each captured locus before aligning them. Finally, we aligned

the captured loci with 75% and 95% taxon completeness into a sparse and a dense matrix,

respectively, for subsequent phylogeny reconstruction. Both data matrices were uploaded to

DRYAD.

Phylogeny reconstruction

We reconstructed the phylogenies on both sparse and dense matrices using maximum

likelihood (ML), Bayesian inference (BI), and multi-species coalescence (MSC) approaches.

We chose the topology that received overall the highest nodal supports as the best tree and

mapped the statistics from all these analyses onto it.

To partition both the sparse and the dense UCE matrices, we first split each locus into

three fragments, a core region, a right, and a left flank, using entropy-based Sliding-Window

Site Characteristics (SWSC-EN; Tagliacollo and Lanfear 2018). We then used PartionFinder

2 (Lanfear et al. 2017) to reclassify all fragments into the best-fitted partition subsets using

AICc algorithm with the substitution models, suggested by SWSC-EN, set as GTR+G.

We preformed ML analyses in IQ-TREE v2.3.2 (Minh et al. 2020) using the partitioned

matrices. Nodal supports were derived from ultrafast bootstrap approximation (UFBoot;

Minh et al. 2013; Hoang et al. 2017) and SH-like approximate likelihood ratio test (SH-aLRT;

Guindon et al. 2010), both with 1,000 replicates.

We carried out the BI analyses using the unpartitioned matrices in Exabayes 1.5.1

(Aberer et al. 2014) with default settings. On each matrix, we ran the Markov chain Monte

Carlo (MCMC) searches in two independent and two coupled chains. Each chain had one

million generations sampled every 50 generations with a 25% burn-in. We then generated the

best consensus trees after evaluating the results by their effective sample sizes (ESS). The

posterior probabilities of the MCMC searches were used as nodal supports.

For MSC, we first generated a gene tree per partition using IQ-TREE v2.3.2 with 1,000

UFBoot replicates. We then imported all gene trees into Accurate Species Tree Algorithm

(ASTRAL-IV; Zhang et al. 2018; Zhang and Mirarab 2022) with default settings. Each

analysis had 16 rounds of search and 16 rounds of subsampling and the posterior probabilities

were used as nodal supports.

Divergence time estimation

Due to the lack of synapomorphies, the classification of all fossils previously assigned to

Pisauridae has recently been questioned (Wunderlich 2008; Magalhaes et al. 2020), so these

fossils could not be used for time calibration. We calibrated the phylogeny using three

relatively reliable fossils with a relevant family affiliation, following Magalhaes et al. (2020),

as follows. First, we treated an unidentified juvenile lycosid (Penney 2001) from Dominican

amber (15–20 million years ago (Ma); Iturralde-Vinent and MacPhee 1996) as a crown

lycosid. Secondly, we treated *Oxyopes succini* Petrunkevitch, 1958 from Baltic amber (43–

48.5 Ma; Ritzkowski 1997) as a stem oxyopid. Finally, we treated *Syphax* cf. *megacephalus*

L. Koch and Berendt, 1854 (see Wunderlich 2004) from Baltic amber as a stem thomisid (see

also Piacentini and Ramírez 2019; Magalhaes et al. 2020).

A posteriori calibration was performed in RelTime (Tamura et al. 2012; 2018) using a

relaxed local clock model under MEGA 11 (Tamura et al. 2021) with *Viridasius* sp. set as the

outgroup to all Lycosoidea. We used the unpartitioned sparse UCE matrix and its best ML

topology, generated from IQ-TREE v2.3.2, for time calibration analyses. The probability

distributions of time at the calibration points were set as log-normal with their 95%

confidence intervals specified close to their proposed fossil ages. This reconstructed

ultrametric tree was then used in subsequent evolutionary analyses.

To test whether RelTime yielded reliable absolute divergence times (see Lozano-

Fernandez et al. 2017; Beavan et al. 2020), we also ran *a priori* time calibration under

MCMCtree (Yang 2007) on a matrix of 300 loci with 99% taxon completeness, randomly

selected using Phyluce v1.7.3, in PAML 4.10 (Yang 2007). We selected the ML topology

generated from the dense matrix as the starting tree after removing the branch lengths.

Calibration points were selected and set as in the RelTime analysis using MCMCtreeR

(Puttick 2019) in R 4.3.3 (R Core Team 2024). Following the program tutorial (dos Reis et al.

2017), we first estimated the average substitution rate per time unit as well as the length,

gradient, and Hessian of each branch. With the above estimated parameters, we ran

divergence time dating in two independent runs. Each run had 500,000 generations sampled

every 50 generations and with a 10% burn-in. We compared the results of the two runs in

MCMCtreeR, using the script from Gutiérrez-Trejo et al. (2024).

Evolutionary analyses

We performed all evolutionary analyses in R 4.3.3 using the packages "ape" (Paradis et

al. 2004), "phytools" (Revell 2024), and "MCMCglmm" (Hadfield 2010). We first pruned the

distantly related outgroups (Zoropsidae, Psechridae, Oxyopidae, Thomisidae, Viridasiidae)

from the ultrametric tree (using the command code "drop.tip" and "extract.clade" in "ape") as

they were irrelevant for the evolutionary questions posed, but retained Lycosidae and

Trechaleidae because they nested with our ingroups (see Results). We also removed the

pisaurid *Tallonia picta* Simon, 1889 due to insufficient trait data. Given the strong emphasis

on *Dolomedes* species in our study, our taxon sample, if left unaltered, could lead to biased

evolutionary outcomes (Ackerly 2000; Li et al. 2008; Sidlauskas 2008; Jorgensen et al.

2023). For all evolutionary analyses, we therefore used both the full taxon tree (putatively

biased), as well as the average of randomized pruned taxon trees (putatively less biased), as

follows. First, we assigned *Dolomedes* terminals to their biogeographic realms modified from

Udvardy (1975), then randomly selected one terminal per realm to generate a reduced subset

of *Dolomedes* terminals and pruned the other *Dolomedes* terminals from the tree. We

repeated this randomization 30 times to generate 30 different reduced taxon trees, each

presumably with a relatively balanced taxon sampling.

To reconstruct the evolution of semi-aquatic vs. terrestrial lifestyles and the

presence/absence of capture webs, we first sought the best evolutionary model by fitting, on

i) the full taxon tree and ii) the reduced taxon trees, four different assumptions of trait

transition rate matrices: equal rates (ER), all-rates-different (ARD), state A to B only and B to

A only, into k-state Markov models (Mk model; Pagel 1994; Lewis 2001). For the

evolutionary models estimated from the full taxon tree, we selected the one with the lowest

AIC as the best model (Fig. 2a). For the evolutionary models estimated from the reduced

taxon trees, we first averaged the AIC and the estimated trait transition probability generated

from each model with the same assumptions on all reduced taxon trees (Fig. 2b). Then, we

selected the evolutionary model with the lowest average AIC as the most suitable across all

reduced taxon trees. To reconstruct the ancestral states of lifestyles and capture webs in

Dolomedes and pisaurids, we simulated stochastic character maps 100 times on the full taxon

tree, once using the best model from the full taxon tree, and again using the best-averaged

model generated from the reduced taxon trees (Fig. 2b).

We used Pagel's (1994) binary character correlation test to examine the evolutionary

correlation between lifestyles and capture web on i) the full taxon tree and ii) reduced taxon

trees. We first fit the two traits into a two-trait independent, and a dependent ARD-Mk model.

For the models estimated from the full taxon tree, we applied model selection by analysis of

variance (ANOVA) to test whether the dependent model significantly better fit our data (Fig.

2c). For the models estimated from the reduced taxon trees, we further averaged the values in

the statistic tables generated from the ANOVA model selections and the estimated trait

transition probabilities of all the reduced taxon trees. We then used the averaged p-value and

the number of reduced taxon trees that showed significance in each ANOVA model selection

to seek the consensus on whether semi-aquatic versus terrestrial lifestyles and the presence or

absence of capture webs were correlated across all reduced taxon trees (Fig. 2d).

We tested the correlation between lifestyles and three morphological traits (CW; L1/CW;

T1/L1) using Markov chain Monte Carlo Sampler for Multivariate Generalized Linear Mixed

Models (MCMCglmm; Hadfield 2010) on i) the full taxon tree (Fig. 2e) and ii) the reduced

taxon trees (Fig. 2f) with 10 million MCMC iterations. The thinning interval and burn-in

were set to 5,000 and 2.5 million, respectively. For those morphological trait(s) that were

significantly (pMCMC < 0.05) correlated with lifestyles on the full taxon tree, we further

examined their relationship with lifestyles by generating density curves of estimated moment-

matching values (Hadfield 2010). To test for correlations between lifestyles and the three

morphological traits on the reduced taxon trees, we averaged all values in the MCMCglmm

tables from all the reduced taxon trees, and also counted the number of the reduced taxon

trees that showed significance (Fig. 2f). Then, as above, we further examined the relationship

between morphological traits with lifestyles by generating density curves of estimated

moment-matching values (Hadfield 2010). As phylogenetic correlations among traits can be

detected at some, but not all, hierarchical levels of a given phylogeny (see Wolff et al. 2022),

we performed an additional set of MCMCglmm analyses on a full set of *Dolomedes* terminals

only following the above algorithm for the full taxon tree (Fig. 2g).

RESULTS

The RTA spider probe set, on average, captured 2,790 UCE loci consisting of 2,815,984

base pairs (Table S1) with *Peucetia* sp. (Oxyopidae) yielding only 1,125 loci. After trimming

and aligning, the sparse matrix with 75% taxon completeness contained 2,676 UCE loci with

1,221,957 base pairs, and the dense matrix with 95% taxon completeness contained 1,175

UCE loci with 875,429 base pairs. Only 94 loci were shared by all taxa.

Phylogenies reconstructed from the sparse and the dense matrix show identical

topologies in all analyses except within *Dolomedes* (Fig. S1), where MSC recovered slightly

different topologies (Figs. S1e–f), interpreted in the following paragraphs. With very few

exceptions, all nodes receive full or high statistical support from UFboot, SH-aLRT, and BI

posterior probabilities. Despite yielding a few additional poorly supported nodes within

Dolomedes, most nodes are still strongly supported by MSC. Given the best overall supports,

we choose the ML topology reconstructed from the sparse matrix as the preferred tree (Figs.

3 and 4; see also Supplementary Material).

Systematics of raft- and nursery web- spiders

All analyses reject the monophyly of Pisauridae as currently classified. This family-level

name refers to a paraphyletic assemblage of genera that fall into three clades, labeled as focal

clades in this paper (Fig. 3; see Taxonomy): 1) **Focal Clade I** groups the majority of the

genera classified in Pisauridae, and because it contains *Pisaura* Simon, 1886, this clade

represents the "true pisaurids". Focal Clade I (Pisauridae) is sister to a taxon-rich clade

containing all the taxa listed below; 2) **Focal Clade II** contains one genus that is currently

still cataloged as a pisaurid, i.e. the monotypic *Blandinia* Tonini et al., 2016 from Madagascar

(Fig. 1K). Because it is recovered as sister to Trechaleidae and Lycosidae, *Blandinia* cannot

be considered a pisaurid, and is instead *incertae sedis*, pending better taxonomic resolution;

3) **Focal Clade III** contains *Dolomedes*, *Megadolomedes* Davies and Raven, 1980, and

Ornodolomedes Raven and Hebron, 2018. This clade is sister to Focal Clade II, Trechaleidae,

and Lycosidae (Fig. 3). Considering the phylogenetic results, the topological stability, the

strong clade supports, and our understanding of the morphology of *Dolomedes* and its

relatives, detailed in the Taxonomy section below, we will from here on formally refer to the

Focal Clade III as Dolomedidae Simon, 1876, colloquially named *raft spiders*. We suggest

the use of *fishing spiders* for *Dolomedes*.

Within Dolomedidae, we tested for the monophyly of *Dolomedes*. Strictly speaking,

Dolomedes is not monophyletic. The majority of *Dolomedes* species, including the type

species *D. fimbriatus* (Clerck, 1757), are in a clade that represents true *Dolomedes*, but one

species, *D. karijini* Raven and Hebron, 2018, is recovered in the sister clade that also contains

Ornodolomedes and *Megadolomedes* (Figs. 3 and 4). While the Afrotropical *Dolomedes*

species are monophyletic within their biogeographic realm, other biogeographic realms do

not show strict species monophyly (Fig. 4). For example, in addition to the distal Palearctic

Dolomedes clade, other Palearctic species are found in the "raptor" clade, which is sister to

all other true *Dolomedes* (Fig. 4). The Australasian species are not monophyletic, but the

New Zealand species are (Fig. 4). An Indomalayan clade groups some, but not all, species

from this region, as many Indomalayan terminals also form the "raptor" clade (Fig. 4). Most

Nearctic species are monophyletic, but *D. striatus* Giebel, 1869 is sister to the Palearctic

clade (Fig. 4).

Multi-species coalescence recovers three topological differences within *Dolomedes*

(Figs. S1e–f) from the preferred tree (Fig. 4), two among the distal "raptor" clade and the

other among the Indomalayan, Afrotropic, and Nearctic plus Palearctic (Holarctic) clades.

The MSC phylogenies reconstructed from both matrices suggest the two morphospecies, *D.*

cf. *raptoroides* HK and *D*. cf. *raptoroides* VN, are not sister as they are in the preferred tree

(Fig. 4). Additionally, MSC has *D. yawatai* Ono, 2002 as sister to a clade with four

Dolomedes morphospecies from Taiwan (Figs. S1e–f) instead of nesting within that clade

(Fig. 4). MSC on the dense matrix suggests the Indomalayan clade as sister to a large clade

including the Afrotropic and Holarctic *Dolomedes* (Figs. S1e–f). However, such topologies

are poorly supported with their posterior probabilities (PP) between 0.6 and 0.7. In the

preferred tree, Afrotropic *Dolomedes* are sister to a clade uniting the Indomalayan and the

Holarctic clades, with better nodal supports (> 0.9; see Fig. 4).

Divergence times of raft- and nursery web- spiders

RelTime and MCMCtree (results in parentheses) yield comparable divergence times of

the three focal clades. Both analyses reconstruct relatively recent origins for the major clades

in question. The crown that gave rise to all three focal clades plus Trechaleidae and Lycosidae

is estimated to have originated between 32 and 37 Ma (34–43 Ma), in the late Eocene to early

Oligocene (Fig. 5). The origin of Pisauridae (Focal Clade I), estimated between 29 and 35 Ma

(31–40 Ma) (Fig. 5), is the earliest of the three focal clades. The Oligocene-Miocene split of

Blandinia (Focal Clade II) from Trechaleidae plus Lycosidae is estimated between 21 and 31

Ma (24–34 Ma) (Fig. 5). Compared to the above two focal clades, the origin of Dolomedidae

(Focal Clade III) is estimated to be more recent, between 10 and 15 Ma (12–17 Ma) in the

mid-Miocene (Fig. 5). The origin of *Dolomedes* is estimated to be in the Miocene, between 9

to15 Ma (11–16 Ma) (Fig. 5), and since then, the clade has undergone a rapid diversification,

with the 95% confidence intervals and highest posterior densities for final splits between

species doublets lying between 0.2 Ma (0.3 Ma) (*D. hydatostella* Yu and Kuntner, 2024 and

D. rotundus Yu and Kuntner, 2024 from Madagascar) and 9.2 Ma (8.9 Ma) (*D. lesserti*

Roewer, 1955 from Benin and an unnamed *Dolomedes* species from D.R. Congo) (Fig. 5).

On the other hand, some species that group with larger clades in our phylogeny could

plausibly be much older, e.g. between 7.1 and 14.7 Ma (10.2–15 Ma) (*D. facetus* Koch, 1876

from Australia).

Evolution of lifestyles in raft- and nursery web- spiders

With the lowest average AIC, the model allowing only transitions from a semi-aquatic to

a terrestrial lifestyle ("semi-aquatic to terrestrial" model) is the best evolutionary model

(Table 4) on the reduced taxon trees (Fig. S2). Partially rejecting our hypothesis, the semi-

aquatic lifestyle is the ancestral state not only of Dolomedidae (Focal Clade III), but also of

the entire clade that unites Pisauridae (Focal Clade I), Focal Clade II, Lycosidae, and

Trechaleidae (Fig. 6A, see also Fig. S3). On the full taxon tree, the model detects no fewer

than 15 reversals to a terrestrial lifestyle, including three in Pisauridae and eight in

Dolomedidae. The terrestrial lifestyle in the focal clades of our tree, thus evolved from semi-

aquatic ancestors. In Focal Clade I, all semi-aquatic terminals or genera, *Hygropoda*,

Thaumasia Perty, 1833, *Qianlingula* Zhang et al., 2004, and *Nilus*, have retained the

ancestral, semi-aquatic lifestyle (Fig. 6a). The seven terrestrial *Dolomedes* species are not

closely related to each other (Fig. 6a), suggesting independent terrestrialization in each

species. This result is not strongly affected by taxon sampling bias. Model selection on the

full taxon tree namely also suggests the "semi-aquatic to terrestrial" model as the best (Table

4), and thus the reconstructed ancestral lifestyle is identical to that from the reduced taxon

trees, albeit with different rates of trait transition (Fig. 6b, see also Fig. S4).

After averaging on the reduced taxon trees (Fig. S2), the ARD is the best model to

explain the evolution of the capture web (Table 4); the reconstruction does not recover a clear

ancestral state (Fig. 6c, see also Fig. S5). Due to this ambiguity, the hypothesis of an ancestral

absence of a capture web for the clade containing Pisauridae, *Blandinia*, and Dolomedidae is

not supported. Reconstructions on the full taxon tree suggest at least three independent

origins of the capture web: one each in Lycosidae, *Blandinia*, and distally within the Focal

Clade I, the latter containing numerous reversals to no web (Fig. 6c). Within *Dolomedes*,

however, the absence of a capture web is ancestral, as partially predicted by the hypothesis

(Fig. 6c). Furthermore, the absence of a web is ancestral for the entire Dolomedidae, with no

reversals (Fig. 6c). This analysis is strongly affected by the taxon sampling bias and its

correction. Model selection on the full taxon tree namely suggests that the model allowing

only transitions from web presence to absence, rather than the ARD, is the best model. This

model reconstructs the presence of a capture web as the ancestral state of all three focal

clades, Trechaleidae, and Lycosidae with numerous reversals to no web (Fig. 6d, see also Fig.

S7). Such results would outright reject our hypothesis of an ancestral absence of a web in

Pisauridae.

Our analyses also reject the hypothesis of an evolutionary correlation between capture

webs and lifestyles. Instead, the evolution of the web presence or absence and semi-aquatic

versus terrestrial lifestyles are independent (Fig. 7a). None of the reduced taxon trees suggest

that the two-trait dependent model fits our data significantly better than the independent

model (average p-value of the ANOVA model selection = 0.1, see also Supplementary

Material). However, this result is strongly affected by the likely bias in the taxon sample on

the tree. On the full taxon tree, the two-trait dependent ARD model fits the data significantly

better than the two-trait independent ARD model (see also Supplementary Material),

suggesting that a semi-aquatic lifestyle excludes the presence of a capture web (Fig. 7b). This

result would corroborate the predicted evolutionary correlation between these traits.

Our final set of hypotheses predicted that semi-aquatic taxa would exhibit wider

carapaces, relatively shorter legs I, and relatively longer tarsi I compared to the terrestrial

taxa in the phylogeny. The results support only the first hypothesis (Table 5). The significance

of a wider carapace in semi-aquatic taxa is recovered in 29 out of 30 reduced taxon trees (see

Supplementary Material). On average, the carapace of semi-aquatic spiders is 2.4 mm wider

than that of the terrestrial spiders in the phylogeny (Fig. 8a). On the other hand, no

significance is detected in the relative length of leg I and tarsus I between semi-aquatic and

terrestrial taxa (Table 5). These results appear to be less susceptible to taxon bias, as analyses

using the full taxon tree yield identical results (Table 5, Fig. 8b). However, when only

Dolomedes terminals are considered, all three of the above hypotheses are rejected (Table 5).

Discussion

The lack of comprehensive phylogenies has so far precluded a thorough systematic

account of *Dolomedes* and Pisauridae *sensu lato*, as well as generalizations of the lifestyle

evolution of this global assemblage of spider genera. We pursue these broad goals in the

phylogenomic and comparative study reported here. Our phylogenetic results reject the

monophyly of both Pisauridae and *Dolomedes*. The genera previously placed within

Pisauridae fall into three focal clades: the true Pisauridae, *Blandinia*, and Dolomedidae,

whose family rank we formally resurrect (see Taxonomy). The origin of each of the focal

clades is estimated to be relatively recent, i.e. 40 Ma or less. *Dolomedes* is paraphyletic, as *D.*

karijini groups with *Ornodolomedes* and *Megadolomedes*. Our reconstructions partially reject

our first hypothesis that the semi-aquatic lifestyle is ancestral to *Dolomedes* but not to

Pisauridae. A semi-aquatic lifestyle is reconstructed as ancestral for all three focal clades,

followed by independent reversals to terrestrial lifestyle. Our second hypothesis of the

ancestral absence of a capture web in *Dolomedes* and Pisauridae is also partially rejected. The

evolution of a capture web is ancestrally ambiguous, with a web origin in both Pisauridae and

Blandinia. In Dolomedidae, on the other hand, the unequivocal ancestral absence of a capture

web is retained through their evolutionary history. Our third hypothesis, which predicted that

a semi-aquatic lifestyle would exclude the presence of a capture web, is also rejected, because

our evolutionary tests reveal an independent evolution of lifestyle and capture web. Of the

hypotheses that semi-aquatic pisaurids having larger body sizes, relatively shorter legs, and

relatively longer tarsi, our comparative analyses support only the larger body size hypothesis,

but only at higher hierarchical levels and not within *Dolomedes* species. Contrary to our

prediction, the relative lengths of legs and tarsi do not differ between lifestyles. Our study is

the first step towards stability in systematics of raft and nursery web spiders and at the same

time provides a phylogenetic basis for further comparative research.

Dolomedidae: A step towards the redefinition of Pisauridae

We provide a well-sampled and robust phylogenomic backbone to support the

resurrection of Dolomedidae, Lehtinen's (1967) taxonomic hypothesis that has regained

recent attention (Albo et al. 2017; Hazzi and Hormiga 2023; Kulkarni et al. 2023). Our

phylogeny is consistent with the topologies found by Hazzi and Hormiga (2023) and

Kulkarni et al. (2023) that *Dolomedes* is sister to the clade including Lycosidae, Trechaleidae,

and additionally *Blandinia*. Although several putatively closely related genera of *Dolomedes*,

including *Bradystichus* Simon, 1884, *Caledomedes* Raven and Hebron, 2018, *Mangromedes*

Raven and Hebron, 2018, and *Tasmomedes* Raven and Hebron, 2018, were not available for

inclusion with UCE data, additional evidence from morphological (Platnick and Forster 1993;

Raven and Hebron 2018) and molecular (Piacentini and Ramírez 2019; Kulkarni et al. 2023)

studies both suggest that these genera fit our definition of Dolomedidae rather than

Pisauridae. This updated classification of Dolomedidae is formalized in the Taxonomy.

The *Dolomedes* paraphyly hinges on one species, *D. karijini*, and could be resolved

through an establishment of a new genus for this species group. Although we envision this

solution being valid in the future, we refrain from doing so here due to our limited access to

Australian species. Based on details of the genitalia, Raven and Hebron (2018) treated *D.*

karijini in the *D. albicomus* group with *Dolomedes* species from Australia and New

Caledonia. Considering the incomplete taxon sampling and the current ambiguity of species

boundaries within *Dolomedes* (Tanikawa and Miyashita 2008; Vink and Dupérré 2010; Yu

and Kuntner 2024), it is unclear whether the entire species group, or only *D. karijini* should

be elevated to a new genus. This decision will await future integrative taxonomic work with

more Australasian species.

Our taxonomy is only the first step towards a better definition of Pisauridae and its

subfamilies. Although our phylogenies reject the classification of *Blandinia* as a pisaurid, it is

unclear whether this monotypic genus can be assigned to another existing family. Its

placement in Trechaleidae would be unfounded, given conflicting phylogenies (see Piacentini

and Ramírez 2019; Hazzi and Hormiga 2023; Kulkarni et al. 2023). In the Taxonomy below,

we therefore avoid creating a new family and hence treat *Blandinia* as *incertae sedis* until

further evidence is available. Our phylogeny also partially rejects the currently used pisaurid

subfamilial classifications, Thaumasinae, Thalassinae, and Pisaurinae (Simon 1890; Sierwald

1990), a problem that awaits more focused systematics efforts.

Independent evolution of lifestyles and capture web

Most spiders are terrestrial, so the evolution of aquatic or semi-aquatic lifestyles in this

group is an evolutionary oddity worthy of in-depth research. Species and genera within at

least 22 spider families are known to have aquatic or semi-aquatic lifestyles (Crews et al.

2020). A comparative study across 42 spider families by Stratton et al. (2004) recovered

aquatic adaptive locomotion and exoskeleton morphologies as ancestral to Pisauridae,

Lycosidae, and Trechaleidae. These results agree with ours that a semi-aquatic lifestyle is

ancestral to those three families as well as to Dolomedidae. However, research focusing on

Dictynidae O. Pickard-Cambridge 1871 (Crews et al. 2020) and Lycosoidea (Piacentini and

Ramírez 2019), which includes Pisauridae and *Dolomedes*, both recovered a terrestrial

lifestyle as ancestral to these clades, followed by multiple origins of aquatic or semi-aquatic

lifestyles. Our results, which suggest that a semi-aquatic lifestyle is ancestral to both

Pisauridae and Dolomedidae, do not align with those above (Piacentini and Ramírez 2019).

This discrepancy may stem from the less taxon sampling focus on terrestrial lycosids and a

more complete taxon sample of pisaurids and dolomedids in this present study compared to

that of Piacentini and Ramírez (2019). Character states of long and basal branches are more

likely to be reconstructed as ancestral (see Li et al. 2008), and our study indeed detected

several newly sequenced semi-aquatic pisaurids at the base of the family, and, additionally,

improved the phylogenetic resolution of *Dolomedes*.

Aside from the widely documented freshwater bound, semi-aquatic lifestyle,

Dolomedidae also comprises the marine intertidal *Mangromedes* (Raven and Hebron 2018)

and the mud-dwelling *Bradystichus* (Platnick and Forster 1993; Raven and Hebron 2018;

Harvey M.S., personal observation). The marine intertidal lifestyle is very rare in the spider

tree of life (Raven 1988; Crews et al. 2020; Yu et al. 2023). Mud-dwelling, likewise, is rare

and may represent an intermediate state between semi-aquatic and terrestrial lifestyles. These

extraordinary cases could imply a more complex and diverse evolutionary history of water-

associated lifestyles in Dolomedidae compared to Pisauridae. However, detailed evolutionary

analyses of these additional lifestyles will require a phylogeny including all dolomedid

genera.

The evolution of capture webs in spiders has been the focus of numerous recent studies

(Blackledge et al. 2009; Bond et al. 2014; Garrison et al. 2016; Cheng and Piel 2018;

Fernández et al. 2018; Kallal et al. 2021). By focusing on all spiders, these studies were

unable to explore the lifestyle diversity within large and speciose clades, such as Pisauridae.

The above studies have not reached a consensus on the ancestral presence or absence of a

capture web in Pisauridae, Lycosoidea, or the RTA clade by either reconstructing no web as

ancestral (Garrison et al. 2016; Fernández et al. 2018; Piacentini and Ramírez 2019), the

presence of a ground sheet web as ancestral (Blackledge et al. 2009), or producing an

ambiguous/unresolved result (Bond et al. 2014; Cheng and Piel 2018; Kallal et al. 2021).

These differing results are not surprising, considering that only three of the above studies

(Cheng and Piel 2018; Piacentini and Ramírez 2019; Kallal et al. 2021) included any web-

building pisaurids, in addition to non-web-building representatives. Our study covers most of

the pisaurids known to build capture webs, but does not unambiguously resolve the ancestral

state. This strongly suggests that the results of the above works, which were based on very

few selected pisaurid taxa were over-resolved. Among the known web-building pisaurids

unavailable in our study is the South American *Architis* Simon, 1898, which, based on

published morphological (Santos 2007a) and phylogenetic studies (Wheeler et al. 2017;

Piacentini and Ramírez 2019; Hazzi and Hormiga 2023), might belong to the clade

containing *Thaumasia*, *Hala* Jocqué, 1994, and *Tolma* Jocqué, 1994—all webless genera. If

this is the case, we expect this clade to represent another case of independent origin of a

capture web in pisaurids.

As no pisaurid or dolomedid taxa are known to be both semi-aquatic *and* build a capture

web (Table 1), we hypothesized that these traits are evolutionarily exclusive, at least in

wandering spiders. Although some studies may suggest that in Araneoidea, these two traits

may co-occur in Tetragnathidae Menge, 1866, Linyphiidae Blackwall, 1859, Theridiidae

Sundevall, 1833, and Theridiosomatidae Simon, 1881 (see Crews et al. 2020), the most

convincing cases are rare (Eberhard 2020) and seemingly aberrant. One such case is

Conculus Kishida, 1940 (Anapidae Simon, 1895) that is known for its diving behaviors and

for building a part of its capture web underwater (Komatsu 1940). The other case is

Wendilgarda Keyserling, 1886 (Theridiosomatidae) that attaches its web anchors to the water

surface (Coddington and Valerio 1980). Curiously, this is the first study to test for an

evolutionary correlation between these two traits. Even more curious is that, contrary to our

expectations, the result suggests their independence. The above cases of semi-aquatic *and*

web-building araneoids may not be so aberrant after all and the evolution of semi-aquatic

lifestyle and the presence of a capture web may not be mutually exclusive across Araneae.

Aside from capture and nursery webs, literature also documents other webbing types

constructed by pisaurids and dolomedids, both terrestrial and semi-aquatic, throughout their

ontogeny. Such examples are juvenile webbing in *Thaumasia* (Eberhard 2020), *Qianlingula*

(Yu K.-P.; personal observations), and *Dolomedes fimbriatus* (Jäger P.; personal

observations). Although these types of non-capture webs were not considered in our

comparative analyses, which only considered adult phenotypes, the abilities of semi-aquatic

spiders to build large-scale webbing may also agree with the independent evolution of

lifestyles and capture webs. Future research might profitably look into similarities and

differences in web structure, composition, and web building behaviors among these different

types of webs at different ontogenetic stages.

Morphological adaptations to a semi-aquatic lifestyle

Our comparative analysis at a higher phylogenetic hierarchy revealed that semi-aquatic

representatives of all three focal clades have wider carapaces compared to terrestrial ones.

This finding is consistent with the general understanding that semi-aquatic and aquatic

animals are larger or heavier than terrestrial ones, whether in mammals (Gearty et al. 2018),

reptiles (Meiri 2008), birds (Gaston and Blackburn 1973), or invertebrates (Sabo et al. 2002;

Braddy et al. 2008). Biological explanations for this phenomenon usually invoke buoyancy

and gravity (Reynolds 1977; Reynolds and Karlotski 1977), trophic structure (Shurin et al.

2006; Tucker and Rogers 2014), thermoregulation (Downhower and Blumer 1988; Meiri

2008), aquatic locomotion (Schmidt-Nielsen 1971; Anderson et al. 1979), energy trade-off

(Gearty et al. 2018), and predator avoidance (Wolff and Guthrie 1985). However, these

explanations are more relevant to vertebrates, whereas gravity and thermoregulation may be

less applicable to much smaller invertebrates, such as spiders.

We hypothesize that larger sizes of semi-aquatic pisaurids and dolomedids may be

related to a biomechanical threshold above which surface tension can be broken when spiders

forage under water. For generalist predators, aquatic habitats offer additional food resources

compared to the terrestrial habitats, e.g. crustaceans, fishes, and amphibians, but only if the

predator can reach underwater and subdue this relatively large prey. Semi-aquatic spiders that

are large enough to break through the surface tension to access these additional food sources,

while avoiding being eaten, may therefore have a competitive advantage over terrestrial

species. Our field observations on *Dolomedes plantarius* (Clerck, 1757) that smaller juveniles

usually do not dive when disturbed but larger adults and juveniles usually do (Yu K.-P.,

Klemen Č, and Kuntner M., personal observation) may add credibility to this hypothesis.

Additionally, larger size in animals can facilitate aquatic locomotion through increased area

in touch with the water surface and consequentially increased buoyancy (Soncini and Klein

2023), and by holding larger and denser hydrophobic structures. However, semi-aquatic

species do not simply evolve to become increasingly larger as their maximum size cannot

break the physical equilibrium of rafting on water surfaces (Soncini and Klein 2023).

We repeated the test of correlation between body size and lifestyle on *Dolomedes*

terminals only and found that semi-aquatic species of *Dolomedes* were not significantly

larger than the exclusively terrestrial ones. It is not unusual for comparative work to find

conflicting results at different phylogenetic hierarchies, including examples in a study about

correlation between morphological traits and foraging guilds in spiders (Wolff et al. 2022). In

many such cases, explanations are not readily available, but our case does not seem

complicated. Why is that? The vast majority of known *Dolomedes* are semi-aquatic, and this

lifestyle is ancestral and nearly universal in this clade, with only a few derived species having

recently evolved terrestrial lifestyles. In these cases, the time on land, and presumably a

relaxed selection for larger sizes, has simply not been long enough for these species to evolve

smaller sizes that are otherwise more typical of terrestrial inhabitants. This analysis of

Dolomedes terminals only may therefore be a less powerful test of terrestrial traits compared

than an analysis of all focal clades at the level of genera.

Reducing taxon sampling bias in trait evolution analyses

Our research provides a case study of how taxon sampling bias can affect the outcome

of commonly used evolutionary analyses and how to avoid this bias. Trait transition

probability matrices (Q), estimated by fitting the extended M*k* model to the phylogeny, are

sensitive to the number of terminals of each character state (Lewis 2001; see also Jorgensen

et al. 2023). Therefore, if the trees or clades do not contain all extant taxa, a biased taxon

coverage in each character state could result in misleading or less robust results (Ackerly

2000; Sidlauskas 2008; Jorgensen et al. 2023). Two solutions have been proposed to reduce

such bias: i) calibration of Q by additional parameters derived from the taxon sampling

completeness (Jorgensen et al. 2023); and ii) data selection (Ackerly 2000; Sidlauskas 2008).

In our study, taxon sampling was heavily weighted towards *Dolomedes*, a genus, in which

most species are semi-aquatic and none have a capture web. The relative over-sampling of a

homogenous clade could lead to inflation or reduction of certain transition probabilities. Due

to the unknown lifestyles in many pisaurid genera, calibration parameters by taxon sampling

completeness were not feasible. We therefore approached the reduction of taxonomic bias in

our analyses through data selection by randomly pruning *Dolomedes* terminals from the full

taxon tree, corrected for their representation of biogeographic realms. We subjected all our

evolutionary and comparative tests on trees with and without bias correction, i.e. the full

taxon tree, and 30 reduced taxon trees.

Comparing the outcomes of the analyses with and without bias correction reveals that

the ancestral state reconstruction of lifestyles and the correlation test between lifestyles and

morphological traits are relatively robust to taxon sampling bias. However, the ancestral state

reconstruction of a capture web and the correlation between lifestyle and capture web are

sensitive to taxon sampling bias. Without reducing the taxon sampling bias, both analyses

yielded over-resolved ancestral states, as well as misleading correlation outcomes between

pairs of traits.

RelTime and MCMCtree yield congruent results

We subjected the data to two approaches to divergence time estimation, the *a posteriori*

and the *a priori* methods implemented in RelTime (Tamura et al. 2012; 2018) and MCMCtree

(Yang 2007), respectively. Both types of analyses yielded largely overlapping results with

interpretable confidence intervals and posterior densities, which may give considerable

credibility to the estimated absolute and relative ages of the focal clades in our study.

Credibility of the estimated absolute ages in our study is also suggested by comparing the

hypothesized ages of the two island endemic species *Dolomedes schauinslandi* (0.9–4.2 Ma)

and *D. orion* Tanikawa, 2003 (2.3–6.4 Ma) with the proposed time of the major geological

events having shaped the Chatham Islands (2–3 & 4–6 Ma; Heenan et al. 2010) and Okinawa

Islands (6–10 Ma; Wang et al. 2014).

The estimated origin of Pisauridae (29–40 Ma) is consistent with the literature (25–75

Ma; see Piacentini and Ramírez 2019; Magalhaes et al. 2020). However, our estimation

yields a markedly younger origin of the crown Dolomedidae (compare 10–17 Ma in our study

with 25–50 Ma in Piacentini and Ramírez 2019; Magalhaes et al. 2020). These differences

could be due to variation in taxon sampling, amount of molecular data, and different

topologies. Despite the relatively recent origin of the crown Dolomedidae, it sits on a long

stem, as the split between Dolomedidae and its sister clade is estimated to be between 30 and

40 Ma. While the shape of this chronogram could plausibly be an artifact of missing some

critical (extinct) taxa, we find the interpretation more likely that this shape reflects a recent

diversification of Dolomedidae with relatively rapid species radiations, similar to the pattern

in the extant segmented spiders (Xu et al. 2015). The incomplete reproductive isolation

implied by the introgression between the sister species *Dolomedes minor* and *D. aquaticus*

Goyen, 1888 (Lattimore et al. 2011) may reinforce the interpretation that *Dolomedes*

undergoes rapid species radiations.

Did Miocene climate change drive lifestyle diversification?

Cenozoic climate oscillations may have shaped the evolution of the terrestrial lifestyles

and the presence of capture webs in Pisauridae. Combining the results of time calibration and

ancestral state reconstructions, the three reversals from semi-aquatic to terrestrial lifestyles

and the origin of capture webs in Pisauridae are all estimated between 10 and 20 Ma. This

time interval matches the era when the Earth's temperature and humidity reached the optima

and started dropping (Zachos et al. 2001; Sun et al. 2020). Considering i) the extent to which

semi-aquatic pisaurids are all distributed in the tropics, and ii) the distant phylogenetic

proximity among Palearctic and Nearctic pisaurids, one can hypothesize that the cooling and

drying climate in the mid-Miocene may have driven the evolution from semi-aquatic to

terrestrial lifestyles (see also Ye et al. 2018), which may have spurred the emergence of web-

based foraging behaviors.

The terrestrial lifestyle in *Dolomedes* may have undergone a different evolutionary

scheme compared to pisaurids, judging from the fact that the reversals to terrestrial lifestyle

in *Dolomedes* are phylogenetically derived and independent. Not unlike the semi-aquatic

pisaurids, extant *Dolomedes* species of early branching lineages are distributed in subtropical

to tropical Asia and Australasia, corresponding to regions that harbored humid refugia during

the Miocene (Milne 2006; Steinthorsdottir et al. 2021). However, the more northern

distribution ranges of some terminals from the same lineages strongly suggest that

Dolomedes species can cope more easily with cold climates than the semi-aquatic pisaurids.

During Miocene climate oscillations, *Dolomedes* may therefore have taken over niches and

habitats that became less hospitable to other large semi-aquatic spiders. This could perhaps

explain their colonization of most continents, and the current biogeographic patterns of

Dolomedes as the only large semi-aquatic spiders inhabiting temperate or even boreal regions

of the Northern Hemisphere. This scenario, on the one hand, and the observed cohabitation of

certain terrestrial and semi-aquatic sister species of *Dolomedes*, –on the other (Ono 2009;

Vink and Dupérré 2010), suggest that the evolution of the terrestrial lifestyle in *Dolomedes*

stems from episodes of rapid radiation and niche partitioning.

TAXONOMY

Taxonomic and phylogenetic history of Dolomedidae and Pisauridae

Among the first organisms to be named using Linnaeus' binomial nomenclature,

Araneus fimbriatus Clerck, 1757 and *Araneus plantarius* Clerck, 1757 from Sweden,

represent the first descriptions of the species now known as *Dolomedes*. Latreille (1804)

introduced the genus name *Dolomedes* and classified it under "wolf spiders" to include these

two species, along with *Araneus mirabilis* Clerck, 1757 [today *Pisaura mirabilis* (Clerck,

1757), the type species of *Pisaura* and thus Pisauridae]. Simon (1864) placed *Dolomedes* in

Lycosidae and later in the lycosid subfamily "Dolomedinae" (Simon 1876). This was the

original designation of Dolomedinae as a family-level taxon, and thus we attribute

Dolomedidae to this author, eventhough Simon (1898b) himself later transferred *Dolomedes*

to Pisauridae. In 1898, Simon then classified Pisauridae into "Pisaurieae", "Thalassieae", and

"Dolomedeae" based on the arrangement of their eyes. This classification was followed by

Petrunkevitch (1928), but renamed as Pisaurinae, Thalassiinae (=Thalassinae), and

Thaumasiinae (=Thaumasinae), respectively. Lehtinen (1967) was the first to propose the

family Dolomedidae which included the elevation of *Dolomedes* and Thaumasiinae. This

proposed taxonomic act was subsequently considered unjustified (Sierwald 1990; Griswold

1993). Although the subfamily placement of *Dolomedes* in Simon (1898b) and Petrunkevitch

(1928) has been followed by some authors (Roewer 1955; Raven and Hebron 2018), the

proximity of *Dolomedes* and *Thaumasia* has never been confirmed phylogenetically. Detailed

studies and analyses of pisaurid genitalia seemed to support the monophyly of Pisauridae

(Sierwald 1990; Griswold 1993; Zhang et al. 2004; Santos 2007b), with *Dolomedes* deemed a

close relative of *Thalassius* Simon, 1885 (=*Nilus*) (Sierwald 1990; Zhang et al. 2004) or

Pisaurina Simon, 1898 (Santos 2007b). Sierwald (1989, 1990) outlined some unique genital

features in *Dolomedes*.

While some phylogenetic studies recovered results similar to the above morphological

hypotheses (Bayer and Schönhofer 2013; Moradmand et al. 2014; Fernández et al. 2018), the

majority suggested otherwise. Some studies still recovered a monophyletic Pisauridae, albeit

with *Dolomedes* and the New Caledonian endemic *Bradystichus* Simon, 1884, if included,

forming a clade sister to all other pisaurids (Wheeler et al. 2017; Cheng and Piel 2018;

Piacentini and Ramírez 2019; Kallal et al. 2021). Other studies suggest that Pisauridae is

paraphyletic (Polotow et al. 2015; Albo et al. 2017; Hazzi and Hormiga 2023; Kulkarni et al.

2023). They recover *Dolomedes* either as sister to a large clade with Pisauridae, Trechaleidae,

and Lycosidae (Albo et al. 2017) or to Trechaleidae and Lycosidae (Hazzi and Hormiga 2023;

Kulkarni et al. 2023). Kulkarni and co-workers (2023) recovered *Dolomedes* and

Bradystichus as sister to Trechaleidae and Lycosidae.

Family Dolomedidae Simon, 1876, family rank resurrected

Bradystichidae Simon 1884, new synonymy

(*Raft spiders*)

Composition:

Dolomedes Latreille, 1804 (*Fishing spiders*), *Bradystichus* Simon, 1884, *Megadolomedes*

Davies and Raven 1980, *Caledomedes* Raven and Hebron, 2018, *Mangromedes* Raven and

Hebron, 2018, *Ornodolomedes* Raven and Hebron, 2018, and *Tasmomedes* Raven and

Hebron, 2018.

Diagnosis:

Dolomedidae resemble Pisauridae, Trechaleidae, and Lycosidae. According to Simon

(1876), these four families share the combination of the following habitus characteristics: i)

carapace pear-shaped and longer than wide, dorsally elevated without tubercles or spines; ii)

eight eyes in two rows: the posterior row strongly recurved with the lateral eyes distantly

separated from the anterior eyes, all posterior eyes larger than the anterior eyes; iii) legs

prograde with three claws, the fourth legs are the longest while the third legs are the shortest;

iv) abdomen longer than wide; and v) ecribellate, spinnerets regular without elongated

sections.

Dolomedidae genera can be distinguished from Trechaleidae, Lycosidae, and Pisauridae

by the following characters: i) a tiny stem and head of spermathecae, which together form an

accessory bulb, connected to the base of the spermathecae (Figs. S7–S9, after Sierwald

1989); ii) an elongated, coiled, or spirally arranged base of the spermatheca and spermathecal

lumen (Figs. S7–S9; see also Remarks); iii) the presence of all three, the retrolateral tibial

apophysis, the ventral tibial apophysis, and the basal cymbium apophysis on the male palp

(Figs. S10–S11, see also Raven and Hebron 2018); v) the presence of a round sclerotized

"saddle" instead of a distal tegular apophysis (Figs. S10–S11, see Sierwald 1990; Raven and

Hebron 2018); and vi) the presence of a lateral subterminal apophysis, which is connected to

the distal sclerotized tube of the apical division together with the embolus and fulcrum (Figs.

S12).

Dolomedidae genera differ from Lycosidae and Trechaleidae by i) having both the

copulatory duct and fertilization duct connected to the base of the spermatheca (Figs. S7–S9,

see also Sierwald 1989); ii) the presence of a distal tegular projection, which forms a U-

shaped tegular ring with the tegulum and conductor (Figs. S10–S11); and iii) the stereotypical

behavior whereby females carry their egg sac on the chelicerae (as opposed to attached to the

spinnerets in Lycosidae and in Trechaleidae).

Dolomedidae genera can also be diagnosed from the subfamily Pisaurinae by i) a nearly

straight or strongly recurved, but never strongly procurved, anterior eye row (see Raven and

Hebron 2018); ii) the carapace not protruding anteriorly at the anterior eye row in the lateral

view (see Blandin 1979; Raven and Hebron 2018); iii) the absence of a "carina" in the female

epigyne (Figs. S7–S8; see Sierwald 1997); iv) the copulatory and fertilization ducts both

connected to the base of spermatheca (Figs. S7–S9, see also Sierwald 1989); v) a U-shaped

tegular ring with a triangular conductor (Figs. S10–S11).

Remarks:

Female vulvae in Dolomedidae are usually elongated tubes whose structures are difficult

to differentiate. Sierwald (1989) considered female *Dolomedes* to have a rather small base of

spermatheca connected to a thick tubular fertilization duct (see Fig. 4 in Sierwald 1989); such

nomenclature was followed subsequently (e.g., Yu and Kuntner 2024). However, further

dissection into those tubular "fertilization ducts" detects matter that can be recognized as a

sperm mass (Fig. S13). This newly emerged detail suggests that those tubular structures are

responsible for sperm storage and should therefore be classified as the elongation of a base of

spermatheca rather than a tubular fertilization duct (see also Jäger 2008). As in Sierwald

(1989), we did not find any glandular pores on those tubular structures. In *Dolomedes* and in

the investigated pisaurids, those pores are found exclusively on the tip of accessory bulbs or

the head of spermathecae but not on its base (Sierwald 1989). The actual function of different

parts of the female vulva in Dolomedidae awaits further detailed studies.

Genus *Blandinia* Tonini et al., 2016, *incertae sedis*

Remarks:

Blandin (1979) first established the genus as *Ransonia* Blandin, 1979, based on female

specimens of a single species from Madagascar. The genus was renamed to *Blandinia* by

Tonini et al. (2016) because *Ransonia* had been preoccupied by a jellyfish genus described by

Kramp (1947). Blandin (1979) classified this genus in Pisauridae and Pisaurinae, owing to its

strongly procurved anterior eye row. Blandin also pointed out that the genus had a distinct

number of cheliceral teeth and a unique epigyne among Pisaurinae genera (Blandin 1979).

Our phylogenomic analyses suggest that *Blandinia* is not a pisaurid but is sister to

Trechaleidae and Lycosidae. Although such a topology might suggest the establishment of a

new family, we refrain from doing so due to unknown and conflicting topologies in

Trechaleidae (Piacentini and Ramírez 2019; Hazzi and Hormiga 2023; Kulkarni et al. 2023),

as well as the unknown male morphology of *Blandinia*. We therefore suggest to treat

Blandinia as *incertae sedis* for the time being.

Family Pisauridae Simon, 1890

Remarks:

Based on our phylogeny (Fig. 3) Pisauridae should be delimited to *exclude* the genera in

Dolomedidae (above) as well as *Blandinia*. This is the first step towards pisaurid

delimitation. Although we refrain from providing a diagnosis for this family here, the current

phylogenetic evidence (Fig. 3) suggests these 24 genera could be classified as pisaurids:

Afropisaura Blandin, 1979, *Caripetella* Strand, 1928, *Charminus* Thorell, 1899, *Cispius*

Simon, 1898, *Cladycnis* Simon, 1898, *Dendrolycosa* Doleschall, 1859, *Euprosthenops*

Pocock, 1897, *Euprosthenopsis* Blandin, 1974, *Hala* Jocqué, 1994, *Hygropoda* Thorell, 1895,

Maypacius Simon, 1898, *Nilus* O. Pickard-Cambridge, 1876, *Paracladycnis* Blandin, 1979,

Perenethis L. Koch, 1878, *Pisaura* Simon, 1886, *Pisaurina* Simon, 1898, *Polyboea* Thorell,

1895, *Qianlingula* Zhang et al., 2004, *Rothus* Simon, 1898, *Sphedanus* Thorell, 1877,

Tallonia Simon, 1889, *Tetragonophthalma* Karsch, 1878, *Thaumasia* Perty, 1833, and *Tolma*

Jocqué, 1994.

The following 20 genera were not included in our phylogenetic analyses but may or may

not be true pisaurids: *Archipirata* Simon, 1898, *Architis* Simon, 1898, *Chiasmopes* Pavesi,

1883, *Cispinilus* Roewer, 1955, *Conakrya* Schmidt, 1956, *Eucamptopus* Pocock, 1900,

Ilipula Simon, 1903, *Inola* Davies, 1982, *Papakula* Strand, 1911, *Phalaeops* Roewer, 1955,

Stoliczka O. Pickard-Cambridge, 1885, *Tapinothele* Simon, 1898, *Tapinothelella* Strand,

1909, *Tapinothelops* Roewer, 1955, *Thalassiopsis* Roewer, 1955, *Tinus* F. O. Pickard-

Cambridge, 1901, *Voraptipus* Roewer, 1955, *Voraptus* Simon, 1898, *Vuattouxia* Blandin,

1979, and *Walrencea* Blandin, 1979.

SUPPLEMENTARY MATERIAL

All supplementary material including the UCE matrices is available at *doi* and DRYAD,

https://doi.org/10.5061/dryad.12jm63z6x.

ACKNOWLEDGMENTS

We thank Robert J. Raven for useful suggestions and kind help with materials and

images; as well as Peter Michalik for the help in spider genial nomenclature. We thank Collin

Chu, Volker Framenau, Nuria Macías Hernández, Kyle Knysh, Joseph Koh, Yuri Marusik,

Sean McCann, Daniel Schoenberg, Michael Skvarla, and Lok Ming Tang for kindly

providing materials, as well as Martín Ramírez, Lauren Esposito, Denise Montelongo, Petra

Sierwald, Maureen Turcatel, Jessica Wadleigh, Dmitri Logunov, Michael Rix, Joseph

Schubert, Owen Seeman, Julianne Waldock, Christophe Allard, and Julia Altmann for

facilitating museum loans. We also thank Matjaž Bedjanič, Han-Po Chang, Matjaž Gregorič,

Daiqin Li, Mu-Ming Lin, Ying-Yuan Lo, Annie Rasoanoeliarimanana, Rhina Harin' Hala

Rasolondalao, Jeremia Ravelojaona, Tiana Vololona, and Yi-Hua Yo for their support in the

field. We would also like to express our thanks to Klemen Čandek, Yi-Yan Li, David

Stanković, Eva Turk and an anonymous peer reader for bioinformatics suggestions and

supports.

FUNDING

Matjaž Kuntner and Kuang-Ping Yu were supported by the Slovenian Research and

Innovation Agency (grants P1-0255, J1-50015). Ren-Chung Cheng was supported by the

National Science and Technology Council, Taiwan (112-2621-B-005-002-MY3). Charles

Haddad was supported by the National Research Foundation of South Africa (IFRR

#132687).

REFERENCES

Aberer A.J., Kobert K., Stamatakis A. 2014. ExaBayes: Massively parallel Bayesian tree

inference for the whole-genome era. Molecular biology and evolution. 31:2553–2556.

Ackerly D.D. 2000. Taxon sampling, correlated evolution, and independent contrasts.

Evolution. 54:1480–1492.

Albo M.J., Bidegaray-Batista L., Bechsgaard J., Silva E.L.C. Da, Bilde T., Pérez-Miles F.

2017. Molecular phylogenetic analyses show that Trechaleidae and Lycosidae are sister

groups. Arachnology. 17:169–176.

Anderson J.F., Hermann R., Prange H.D. 1979. Scaling of supportive tissue mass. The

Quarterly Review of Biology. 54:139–148.

Baba Y.G., Watari Y., Nishi M., Sasaki T. 2019. Notes on the feeding habits of the Okinawan

fishing spider, *Dolomedes orion* (Araneae: Pisauridae), in the southwestern islands of

Japan. Journal of Arachnology. 47:154–158.

Bayer S., Schönhofer A.L. 2013. Phylogenetic relationships of the spider family Psechridae

inferred from molecular data, with comments on the Lycosoidea (Arachnida: Araneae).

Invertebrate Systematics. 27:53–80.

Beavan A.J., Donoghue P.C., Beaumont M.A., Pisani D. 2020. Performance of a priori and a

posteriori calibration strategies in divergence time estimation. Genome biology and

evolution 12:1087–1098.

Bertkau P. 1880. Verzeichniss der von Prof. Ed. van Beneden auf seiner im Auftrage der

Belgischen Regierung unternommen wissenschaftlichen Reise nach Brasilien und La

Plata im Jahren 1872-73 gensammelten Arachniden. Mémoires Couronnés et Mémoires

des Savants Étrangers de l'Académie Royale des Sciences, des Lettres et des Beaux-Arts

de Belgique. 43:1–120.

Bertkau P. 1882. Über das Cribellum und Calamistrum. Ein Beitrag zur Histologie, Biologie

und Systematik der Spinnen. Archiv für Naturgeschichte. 48:316–362.

Bishop S.C. 1924. A revision of the Pisauridae of the United States. New York State Museum

Bulletin. 252:1–140.

Blackledge T.A., Scharff N., Coddington J.A., Szü T., Wenzel J.W., Hayashi C.Y., Agnarsson

I. 2009. Reconstructing web evolution and spider diversification in the molecular era.

PNAS. 106:5229–5234.

Blackwall J. 1859. Descriptions of newly discovered spiders captured by James Yate Johnson

Esq., in the island of Madeira. Annals and Magazine of Natural History. 4:255–267.

Blandin P. 1974. Etudes sur les Pisauridae africaines II. Définition du genre *Euprosthenops*

Pocock, 1897 et description du genre *Euprosthenopsis* n. gen. (Araneae – Pisauridae –

Pisaurinae). Revue Suisse de Zoologie. 81:933–947.

Blandin P. 1979. Etudes sur les Pisauridae afrícaines XI. Genres peu connus ou nouveaux des

Iles Canaries, du Continent africaín et de Madagascar. Revue Zoologique Africaine.

93:347–375.

Bleckmann H., Barth F.G. 1984. Sensory ecology of a semi-aquatic spider (*Dolomedes*

triton): II. The release of predatory behavior by water surface waves. Sociobiology.

14:303–312.

Bleckmann H., Borchardt M., Horn P., Görner P. 1994. Stimulus discrimination and wave

source localization in fishing spiders (*Dolomedes triton* and *D. okefinokensis*). Journal

of Comparative Physiology A. 174:305–316.

Bleckmann H., Lotz T. 1987. The vertebrate-catching behaviour of the fishing spider

Dolomedes triton (Araneae, Pisauridae). Animal Behaviour. 35:641–651.

Bond J.E., Garrison N.L., Hamilton C.A., Godwin R.L., Hedin M., Agnarsson I. 2014.

Phylogenomics resolves a spider backbone phylogeny and rejects a prevailing paradigm

for orb web evolution. Current Biology. 24:1765–1771.

Bösenberg W., Strand E. 1906. Japanische Spinnen. Abhandlungen der Senckenbergischen

Naturforschenden Gesellschaft. 30:93–422.

Braddy S.J., Poschmann M., Tetlie O.E. 2008. Giant claw reveals the largest ever arthropod.

Biol Lett. 4:106–109.

Brady A.R. 2012. Nearctic species of the new genus *Tigrosa* (Araneae: Lycosidae). Journal of

Arachnology. 40:182–208.

Carico J.E. 1973a. The Nearctic Species of the Genus *Dolomedes* (Araneae: Pisauridae).

Bulletin of the Museum of Comparative Zoology. 144:435–488.

Carico J.E. 1973b. The Nearctic spider genus *Pisaurina* (Pisauridae). Psyche, Cambridge.

79:295–310.

Carico J.E. 1976. The spider genus *Tinus* (Pisauridae). Psyche, Cambridge. 83:63–78.

Chae J.H., Lee J.G., Lee J.H., Kim S.K. 2023. Taxonomic notes on *Dolomedes*

nigrimaculatus (Araneae: Pisauridae) from Korea. Acta Arachnol. 72:29–37.

Cheng D.Q., Piel W.H. 2018. The origins of the Psechridae: Web-building lycosoid spiders.

Molecular Phylogenetics and Evolution. 125:213–219.

Clerck C. 1757. Svenska spindlar: uti sina hufvud-slägter indelte samt under några och sextio

särskildte arter beskrefne: och med illuminerade figurer uplyste. Stockholmiae: Literis

Laur. Salvii, MDCCLVII.

Coddington J., Valerio C.E. 1980. Observations on the web and behavior of *Wendilgarda*

spiders (Araneae: Theridiosomatidae). Psyche: A Journal of Entomology. 87:93–105.

Crews S.C., Garcia E.L., Spagna J.C., Van Dam M.H., Esposito L.A. 2020. The life aquatic

with spiders (Araneae): repeated evolution of aquatic habitat association in Dictynidae

and allied taxa. Zoological Journal of the Linnean Society. 189:862–920.

Dankittipakul P., Singtripop T., Zhang Z.-S. 2008. A review of the spider genus *Hygropoda* in

Thailand (Araneae, Pisauridae). Revue Suisse de Zoologie. 115:311–323.

Davies V.T. 1982. *Inola* nov. gen., a web-building pisaurid (Araneae: Pisauridae) from

northern Australia with descriptions of three species. Memoirs of the Queensland

Museum. 20:479–487.

Davies V.T., Raven R.J. 1980. *Megadolomedes* nov. gen. (Araneae: Pisauridae) with a

description of the male of the type-species, *Dolomedes australianus* Koch 1865.

Memoirs of the Queensland Museum. 20:135–141.

Deshefy G.S. 1981. 'Sailing' behaviour in the fishing spider, *Dolomedes triton* (Walckenaer).

Animal Behaviour. 29:965–966.

Dippenaar-Schoeman A.S., Haddad C.R., Foord S.H., Lotz L.N. 2020. The Pisauridae of

South Africa. South African National Survey of Arachnida Photo Identification Guide. p.

1–61.

Doleschall L. 1859. Tweede Bijdrage tot de kennis der Arachniden van den Indischen

Archipel. Acta Societatis Scientiarum Indica-Neerlandica. 5:1–60.

dos Reis M., Álvarez-Carretero S., Yang Z. 2017. MCMCTree tutorials. Online at:

https://gensoft.pasteur.fr/docs/paml/4.9j/MCMCtree.Tutorials.pdf.

Downhower J.F., Blumer L.S. 1988. Calculating just how small a whale can be. Nature.

335:675.

Eberhard W. 2020. Spider webs: behavior, function, and evolution. Chicago, USA: University

of Chicago Press.

Faircloth B.C. 2016. PHYLUCE is a software package for the analysis of conserved genomic

loci. Bioinformatics. 32:786–788.

Faircloth B.C. 2017. Identifying conserved genomic elements and designing universal bait

sets to enrich them. Methods in Ecology and Evolution. 8:1103–1112.

Faircloth B.C., Branstetter M.G., White N.D., Brady S.G. 2015. Target enrichment of

ultraconserved elements from arthropods provides a genomic perspective on

relationships among hymenoptera. Molecular Ecology Resources. 15:489–501.

Faircloth B.C., McCormack J.E., Crawford N.G., Harvey M.G., Brumfield R.T., Glenn T.C.

2012. Ultraconserved elements anchor thousands of genetic markers spanning multiple

evolutionary timescales. Systematic Biology. 61:717–726.

Faircloth B.C., Sorenson L., Santini F., Alfaro M.E. 2013. A phylogenomic perspective on the

radiation of ray-finned fishes based upon targeted sequencing of ultraconserved

elements (UCEs). PLoS One. 8:e65923.

Fernández R., Kallal R.J., Dimitrov D., Ballesteros J.A., Arnedo M.A., Giribet G., Hormiga

G. 2018. Phylogenomics, diversification dynamics, and comparative transcriptomics

across the spider tree of life. Current Biology. 28:1489-1497.e5.

Garrison N.L., Rodriguez J., Agnarsson I., Coddington J.A., Griswold C.E., Hamilton C.A.,

Hedin M., Kocot K.M., Ledford J.M., Bond J.E. 2016. Spider phylogenomics:

Untangling the spider tree of life. PeerJ. 4:e1719.

Gaston K.J., Blackburn T.M. 1973. The frequency distribution of bird body weights: aquatic

and terrestrial species. Ibis. 137:237–240.

Gearty W., McClain C.R., Payne J.L. 2018. Energetic tradeoffs control the size distribution of

aquatic mammals. Proceedings of the National Academy of Sciences of the United

States of America. 115:4194–4199.

Giebel C.G. 1869. Über einige Spinnen aus Illinois. Zeitschrift für die Gesammten

Naturwissenschaften. 33:248–253.

Goyen P. 1888. Descriptions of new species of New Zealand Araneae. Transactions and

Proceedings of the New Zealand Institute. 20:133–139.

Griswold C.E. 1993. Investigations into the Phylogeny of the Lycosoid Spiders and Their Kin

(Arachnida: Araneae: Lycosoidea). Smithsonian Contributions to Zoology. 539:1–39.

Guindon S., Dufayard J.F., Lefort V., Anisimova M., Hordijk W., Gascuel O. 2010. New

algorithms and methods to estimate maximum-likelihood phylogenies: Assessing the

performance of PhyML 3.0. Syst Biol. 59:307–321.

Gutiérrez-Trejo N., Van Dam M.H., Lam A.W., Martínez-Herrera G., Noguera F.A.,

Weissling T., Ware J.L., Toledo-Hernández V.H., Skillman F.W. Jr., Farrell B.D., Pérez-

Flores O., Prendini L., Carpenter J.M. 2024. Phylogenomics of *Tetraopes* longhorn

beetles unravels their evolutionary history and biogeographic origins. Scientific Reports.

14:7285.

Hadfield J.D. 2010. MCMC Methods for Multi-Response Generalized Linear Mixed Models:

The MCMCglmm R Package. Journal of Statistical Software. 33:1–22.

Hazzi N.A., Hormiga G. 2023. Molecular phylogeny of the tropical wandering spiders

(Araneae, Ctenidae) and the evolution of eye conformation in the RTA clade. Cladistics.

39:18–42.

Heenan P.B., Mitchell A.D., De Lange P.J., Keeling J., Paterson A.M. 2010. Late-Cenozoic

origin and diversification of Chatham Islands endemic plant species revealed by

analyses of DNA sequence data. New Zealand Journal of Botany. 48:83–136.

Hentz N.M. 1844. Descriptions and figures of the araneides of the United States. Boston

Journal of Natural History. 4:386–396.

Hentz N.M. 1845. Descriptions and figures of the araneides of the United States. Boston

Journal of Natural History. 5:189–202.

Hoang D.T., Chernomor O., von Haeseler A., Quang Minh B., Sy Vinh L., Rosenberg M.S.

2017. UFBoot2: Improving the Ultrafast Bootstrap Approximation. Molecular Biology

and Evolution. 35:518–522.

Iturralde-Vinent M.A., MacPhee R.D.E. 1996. Age and Paleogeographical Origin of

Dominican Amber. Science. 273:1850–1852.

Jäger P. 2008. Revision of the huntsman spider genus *Heteropoda* Latreille 1804: species

with exceptional male palpal conformations from Southeast Asia and Australia

(Arachnida, Araneae: Sparassidae: Heteropodinae). Senckenbergiana biologica. 88:239–

310.

Jäger P. 2011. Revision of the spider genera *Nilus* O. Pickard-Cambridge 1876, *Sphedanus*

Thorell 1877 and *Dendrolycosa* Doleschall 1859 (Araneae: Pisauridae). Zootaxa.

3046:1–38.

Jocqué R. 1994. Halidae, a new spider family from Madagascar (Araneae). Bulletin of the

British Arachnological Society. 9:281–289.

Jorgensen D., Pons-Salort M., Grassly N.C. 2023. A simple correction to adjust for sampling

biases in phylogeographic discrete trait analysis. bioRxiv:1–41.

Kallal R.J., Kulkarni S.S., Dimitrov D., Benavides L.R., Arnedo M.A., Giribet G., Hormiga

G. 2021. Converging on the orb: denser taxon sampling elucidates spider phylogeny and

new analytical methods support repeated evolution of the orb web. Cladistics. 37:298–

316.

Karsch F. 1878. Exotisch-araneologisches. Zeitschrift für die Gesammten

Naturwissenschaften. 51:322|771–333|826.

Kishida K. 1936. A synopsis of the Japanese spiders of the genus *Dolomedes*. Acta

Arachnologica. 1:114–127.

Kishida K. 1940. In: Komatsu T. 1940. On five species of spiders found in the Ryûgadô

Cave, Tosa province. Acta Arachnol. 5:186–195.

Koch C.L., Berendt G.C. 1854. Die im Bernstein befindlichen Myriapoden, Arachniden und

Apteren der Vorwelt. In: Berendt G.C., editor. Die in Bernstein befindlichen organischen

Reste der Vorwelt gesammelt in Verbindung mit mehreren bearbeitetet und

herausgegeben. Berlin, Germany: Nicolaische Buchhandlung. p. 1–124.

Koch L. 1876. Die Arachniden Australiens, nach der Natur beschrieben und abgebildet.

Nürnberg: Bauer & Raspe.

Koch L. 1878a. Japanesische Arachniden und Myriapoden. Verhandlungen der Kaiserlich-

Königlichen Zoologisch-Botanischen Gesellschaft in Wien. 27:735–798.

Koch L. 1878b. Die Arachniden Australiens, nach der Natur beschrieben und abgebildet

[Erster Theil, Lieferung 22–23]. Nürnberg: Bauer & Raspe.

Komatsu T. 1940. On five species of spiders found in the Ryûgadô Cave, Tosa province. Acta

Arachnol. 5:186–195.

Kramp P.L. 1947. Medusae. Part III. Trachylina and Scyphozoa, with zoogeographical

remarks on all the medusae of the northern Atlantic. Danish Ingolf Expedition. 5:1–66.

Kulkarni S., Wood H.M., Hormiga G. 2023. Advances in the reconstruction of the spider tree

of life: A roadmap for spider systematics and comparative studies. Cladistics. 39:479–

532.

Kuntner M., Coddington J.A. 2020. Sexual size dimorphism: Evolution and perils of extreme

phenotypes in spiders. Annual Review of Entomology. 65:57–80.

Lanfear R., Frandsen P.B., Wright A.M., Senfeld T., Calcott B. 2017. PartitionFinder 2: New

methods for selecting partitioned models of evolution for molecular and morphological

phylogenetic analyses. Molecular Biology and Evolution. 34:772–773.

Lapinski W., Walther P., Tschapka M. 2015. Morphology reflects microhabitat preferences in

an assemblage of neotropical wandering spiders. Zoomorphology. 134:219–236.

Latreille P.A. 1804. Tableau methodique des Insectes. Nouveau Dictionnaire d'Histoire

Naturelle Paris. 24:129–295.

Lehtinen P.T. 1967. Classification of the cribellate spiders and some allied families, with

notes on the evolution of the suborder Araneomorpha. Annales Zoologici Fennici.

4:199–468.

Lenz H. 1886. Beiträge zur Kenntniss der Spinnenfauna Madagascars. Zoologische

Jahrbücher, Zeitschrift für Systematik, Geographie und Biologie der Thiere. 1:379–408.

Lessert R. 1928. Araignées du Congo recueillies au cours de l'expedition par l'American

Museum (1909-1915). Deuxieme partie. Revue Suisse de Zoologie. 35:303–352.

Lewis P.O. 2001. A likelihood approach to estimating phylogeny from discrete morphological

character data. Systematic Biology. 50:913–925.

Li G., Steel M., Zhang L. 2008. More taxa are not necessarily better for the reconstruction of

ancestral character states. Systematic Biology. 57:647–653.

Lozano-Fernandez J., dos Reis M., Donoghue, P.C., Pisani D. 2017. RelTime rates collapse to

a strict clock when estimating the timeline of animal diversification. Genome Biology

and Evolution. 9:1320–1328.

Magalhaes I.L.F., Azevedo G.H.F., Michalik P., Ramírez M.J. 2020. The fossil record of

spiders revisited: implications for calibrating trees and evidence for a major faunal

turnover since the Mesozoic. Biological Reviews. 95:184–217.

Marusik Y.M. 1988. New species of spiders (Aranei) from the Upper Kolyma. Zool Zhurnal.

67:1469–1482.

McAlister W.H. 1960. The diving and surface-walking behavior of *Dolomedes triton*

sexpunctatus (Araneida: Pisauridae). Animal Behaviour. 8:109–111.

McCormick S., Polis G.A. 1982. Arthropods that prey on vertebrates. Biological reviews.

57:29–58.

Meiri S. 2008. Evolution and ecology of lizard body sizes. Global Ecology and
Biogeography. 17:724–734.

Menge A. 1866. Preussische Spinnen. Schriften der Naturforschenden Gesellschaft in Danzig
(N.F.).

Milne R.I. 2006. Northern hemisphere plant disjunctions: A window on tertiary land bridges
and climate change? Annals of Botany. 98:465–472.

Minh B.Q., Nguyen M.A.T., Von Haeseler A. 2013. Ultrafast approximation for phylogenetic
bootstrap. Molecular Biology and Evolution. 30:1188–1195.

Minh B.Q., Schmidt H.A., Chernomor O., Schrempf D., Woodhams M.D., Von Haeseler A.,
Lanfear R., Teeling E. 2020. IQ-TREE 2: New models and efficient methods for
phylogenetic inference in the genomic era. Molecular Biology and Evolution. 37:1530–
1534.

Moradmand M., Schönhofer A.L., Jäger P. 2014. Molecular phylogeny of the spider family
Sparassidae with focus on the genus *Eusparassus* and notes on the RTA-clade and
"Laterigradae." Molecular Phylogenetics and Evolution. 74:48–65.

Nadolny A.A., Kovblyuk M.M. 2012. Members of *Pardosa amentata* and *P. lugubris* species
groups in Crimea and Caucasus with notes on *P. abagensis* (Aranei: Lycosidae).
Arthropoda Selecta. 21:67–80.

Nadolny A.A., Ponomarev A. V, Kovblyuk M.M., Dvadnenko K. V. 2012. New data on

Pisaura novicia (Aranei: Pisauridae) from eastern Europe. Arthropoda Selecta. 21:255–

267.

Nyffeler M., Gibbons J.W. 2022. Spiders feeding on vertebrates is more common and

widespread than previously thought, geographically and taxonomically. Journal of

Arachnology. 50:121–134.

Nyffeler M., Pusey B.J. 2014. Fish predation by semi-aquatic spiders: A global pattern. PLoS

One. 9:1–21.

Ono H. 2002. New and remarkable spiders of the families Liphistiidae, Argyronetidae,

Pisauridae, Theridiidae and Araneidae (Arachnida) from Japan. Bulletin of the National

Museum of Nature and Science Tokyo. 28:51–60.

Ono H. 2009. The spiders of Japan with keys to the families and genera and illustrations of

the species. Kanagawa: Tokai University Press.

Pagel M. 1994. Detecting correlated evolution on phylogenies: A general method for the

comparative analysis of discrete characters. Proceedings: Biological Sciences. 255:37–

45.

Paradis E., Claude J., Strimmer K. 2004. APE: Analyses of phylogenetics and evolution in R

language. Bioinformatics. 20:289–290.

Pavesi P. 1883. Spedizione italiana nell'Africa equatoriale. Risultati zoologici. Aracnidi del

regno di Scioa. Annali del Museo Civico di Storia Naturale di Genova. 20:5–105.

Penney D. 2001. Advances in the taxonomy of spiders in miocene amber from the Dominican

Republic (Arthropoda: Araneae). Palaeontology. 44:987–1009.

Perty M. 1833. Arachnides Brasilienses. In: de Spix J.B., Martius F.P., editors. Delectus

animalium articulatorum quae in itinere per Braziliam ann. 1817 et 1820 colligerunt.

Monachii: p. 191–209.

Petrunkevitch A. 1928. System Aranearum. Transactions of The Connecticut Academy of

Arts and Sciences. 29:1–271.

Petrunkevitch A.I. 1958. Amber spiders in European collections. Transactions of the

Connecticut Academy of Arts and Sciences. 41:97–400.

Piacentini L.N., Ramírez M.J. 2019. Hunting the wolf: A molecular phylogeny of the wolf

spiders (Araneae, Lycosidae). Molecular Phylogenetics and Evolution. 136:227–240.

Pickard-Cambridge F.O. 1897. On the cteniform spiders of Africa, Arabia and Syria.

Proceedings of the Zoological Society of London. 66:13–32.

Pickard-Cambridge F.O. 1901. Arachnida - Araneida and Opiliones. Biologia Centrali-

Americana, Zoology. London: p. 193–312.

Pickard-Cambridge O. 1871. Arachnida. The Zoological Record. 7:207–224.

Pickard-Cambridge O. 1876. Catalogue of a collection of spiders made in Egypt, with

descriptions of new species and characters of a new genus. Proceedings of the

Zoological Society of London. 44:541–630.

Pickard-Cambridge O. 1885. Araneidea. Scientific results of the second Yarkand mission;

based upon the collections and notes of the late Ferdinand Stoliczka, Ph. D. Calcutta:

Government of India. p. 1–115.

Platnick N.I., Forster R.R. 1993. A revision of the New Caledonian spider genus *Bradystichus*

(Araneae, Lycosoidea). American Museum Novitates. 3075:1–14.

Pocock R.I. 1897. Descriptions of two new species of scorpions from East Africa. Annals and

Magazine of Natural History. 19:116–119.

Pocock R.I. 1900. The fauna of British India, including Ceylon and Burma. Arachnida.

London: Taylor and Francis.

Pocock R.I. 1900. On the scorpions, pedipalps and spiders from tropical West-Africa,

represented in the collection of the British Museum. Proceedings of the Zoological

Society of London. 67:833–885.

Polotow D., Carmichael A., Griswold C.E. 2015. Total evidence analysis of the phylogenetic

relationships of Lycosoidea spiders (Araneae, Entelegynae). Invertebrate Systematics.

29:124–163.

Prjibelski A., Antipov D., Meleshko D., Lapidus A., Korobeynikov A. 2020. Using SPAdes de

novo assembler. Current Protocols in Bioinformatics. 70:e102.

Puttick, M.N. 2019. MCMCtreeR: Functions to prepare MCMCtree analyses and visualize

posterior ages on trees. Bioinformatics. 35:5321–5322.

R Core Team. 2024. _R: A Language and Environment for Statistical Computing_. R

Foundation for Statistical Computing, Vienna, Austria.

Raven R. 1988. A revision of the mygalomorph spider genus *Idioctis* (Araneae,

Barychelidae). American Museum Novitates. 2929:1–14.

Raven R., Hebron W. 2018. A review of the water spider family Pisauridae in Australia and

New Caledonia with descriptions of four new genera and 23 new species. Memoirs of

the Queensland Museum. Nature 60:233–381.

Revell L.J. 2024. phytools 2.0: an updated R ecosystem for phylogenetic comparative

methods (and other things). PeerJ. 12:e16505.

Reynolds W.W. 1977. Skeleton weight allometry in aquatic and terrestrial vertebrates.

Hydrobiologia. 56:35–37.

Reynolds W.W., Karlotski W.J. 1977. The allometric relationship of skeleton weight to body

weight in teleost fishes: A preliminary comparison with birds and mammals. Copeia.

1977:160–163.

Ritzkowski S. 1997. K-Ar-Altersbestimmungen der bernsteinführenden Sedimente des

Samlandes (Paläogen, Bezirk Kaliningrad). Metalla (Sonderheft). 66:19–23.

Roewer C.F. 1955. Araneae Lycosaeformia I. (Agelenidae, Hahniidae, Pisauridae) mit

Berücksichtigung aller Arten der äthiopischen Region. Exploration du Parc National de

l'Upemba, Mission G. F. de Witte. 30:1–420.

Roland C., Rovner J.S. 1983. Chemical and vibratory communication in the aquatic pisaurid

spider *Dolomedes triton*. Journal of Arachnology. 11:77–85.

Sabo J.L., Bastow J.L., Power M.E. 2002. Length-mass relationships for adult aquatic and

terrestrial invertebrates in a California watershed. Journal of The North American

Benthological Society. 21:336–343.

Santos A.J. 2007a. A revision of the Neotropical nursery-web spider genus *Architis* (Araneae:

Pisauridae). Zootaxa. 1578:1–40.

Santos A.J. 2007b. A phylogenetic analysis of the nursery-web spider family Pisauridae, with

emphasis on the genera *Architis* and *Staberius* (Araneae: Lycosoidea). Zoologica

Scripta. 36:489–507.

Santos A.J., Álvares S.S., Brescovit A.D. 2003. On the third valid species of the genus

Aglaoctenus Tullgren (Araneae, Lycosidae). Revista Ibérica de Aracnología. 8:89–92.

Schmidt G. 1956. Liste der in den Jahren 1953 and 1954 mit Bananen nach Hamburg

eingeschleppten Spinnen aus Franz.-Guinea. Zoologischer Anzeiger. 157:239–241.

Schmidt-Nielsen K. 1971. Locomotion: Energy cost of swimming, flying, and running.

Science (1979). 177:222–228.

Shurin J.B., Gruner D.S., Hillebrand H. 2006. Review All wet or dried up? Real differences

between aquatic and terrestrial food webs. Proceedings of the Royal Society B:

Biological Sciences. 273:1–9.

Sidlauskas B. 2008. Continuous and arrested morphological diversification in sister clades of

characiform fishes: A phylomorphospace approach. Evolution. 62:3135–3156.

Sierwald P. 1989. Morphology and ontogeny of female copulatory organs in American

Pisauridae, with special reference to homologous features (Arachnida: Araneae).

Smithsonian Contributions to Zoology. 484:1–24.

Sierwald P. 1990. Morphology and homologous features in the male palpal organ in

Pisauridae and other spider families, with notes on the taxonomy of Pisauridae

(Arachnida: Araneae). Nemouria, Occasional Papers of the Delaware Museum of

Natural History. 35:1–59.

Sierwald P. 1997. Phylogenetic analysis of Pisaurine nursery web spiders, with revisions of

Tetragonophthalma and *Perenethis* (Araneae, Lycosoidea, Pisauridae). Journal of

Arachnology. 25:361–407.

Silva E.L.C. da, Carico J.E. 2012. Revision of the Neotropical nursery-web spider genus

Thaumasia Perty, 1833 (Araneae: Lycosoidea: Pisauridae: Thaumasiinae). Zootaxa.

3567:1–64.

Silva E.L.C. Da, Griswold C.E. 2013. The first record and the description of a new species of

Dendrolycosa Doleschall, 1859 (Araneae: Pisauridae: Pisaurinae) from Madagascar.

Zootaxa. 3682:572–578.

1255 Silva E.L.C. Da, Sierwald P. 2013. On three monotypic nursery web spider genera from

1256  Madagascar with first description of the male of *Tallonia picta* Simon, 1889 and

1257  redescription of the type-species of *Paracladycnis* Blandin, 1979 and *Thalassiopsis*

1258  Roewer, 1955 (Araneae: Lycosoidea: Pisauridae). Zootaxa. 3750:277–288.

1259 Silva E.L.C. Da, Sierwald P. 2015. First description of the male of *Rothus vittatus* Simon,

1260  1898 (Araneae: Pisauridae), with a review of the African nursery web spider genus

1261  *Rothus* Simon, 1898. Afr Invertebr. 56:325–341.

1262 Simon E. 1864. Histoire naturelle des araignées (aranéides). Paris, France: Roret.

1263 Simon E. 1876. Les arachnides de France. Tome troisième. Paris, France: Roret.

1264 Simon E. 1881. Les arachnides de France. Tome cinquième, première partie. Paris, France:

1265  Roret.

1266 Simon E. 1884. Description d'une nouvelle famille de l'ordre des Araneae (Bradystichidae).

1267  Annales de la Société Entomologique de Belgique. 28:297–301.

1268 Simon E. 1885. Matériaux pour servir à la faune arachnologiques de l'Asie méridionale. I.

1269  Arachnides recueillis à Wagra-Karoor près Gundacul, district de Bellary par M. M.

1270  Chaper. II. Arachnides recueillis à Ramnad, district de Madura par M. l'abbé Fabre.

1271  Bulletin de la Société Zoologique de France. 10:1–39.

1272 Simon E. 1886. Etudes arachnologiques. 18e Mémoire. XXVI. Matériaux pour servir à la

1273  faune des Arachnides du Sénégal. (Suivi d'une appendice intitulé: Descriptions de

plusieurs espèces africaines nouvelles). Annales de la Société Entomologique de France.

5:345–396.

Simon E. 1889. Etudes arachnologiques. 21e Mémoire. XXXI. Descriptions d'espèces et the

genres nouveaux de Madagascar et de Mayotte. Annales de la Société Entomologique de

France. 8:223–236.

Simon E. 1890. Etudes arachnologiques. 22e Mémoire. XXXIV. Etude sur les arachnides de

l'Yemen. Annales de la Société Entomologique de France. 10:77–124.

Simon E. 1895. Histoire naturelle des araignées. Paris: Roret.

Simon E. 1898a. Descriptions d'arachnides nouveaux des familles des Agelenidae,

Pisauridae, Lycosidae et Oxyopidae. Annales de la Société Entomologique de Belgique.

42:5–34.

Simon E. 1898b. Histoire naturelle des araignées. Deuxième édition, tome second. Paris,

France: Roret.

Simon E. 1899. Ergebnisse einer Reise nach dem Pacific (Schauinsland 1896-1897).

Arachnoideen. Zoologische JahrbücherAbtheilung für Systematik, Geographie und

Biologie der Thiere. 12:411–437.

Simon E. 1903. Descriptions d'arachnides nouveaux. Annales de la Société Entomologique

de Belgique. 47:21–39.

Soncini R., Klein W. 2023. Surface tension in biological systems - a common problem with a

variety of solutions. Comparative Biochemistry and Physiology -Part A : Molecular and

Integrative Physiology. 284:111475.

Steinthorsdottir M., Coxall H.K., de Boer A.M., Huber M., Barbolini N., Bradshaw C.D.,

Burls N.J., Feakins S.J., Gasson E., Henderiks J., Holbourn A.E., Kiel S., Kohn M.J.,

Knorr G., Kürschner W.M., Lear C.H., Liebrand D., Lunt D.J., Mörs T., Pearson P.N.,

Pound M.J., Stoll H., Strömberg C.A.E. 2021. The Miocene: The future of the past.

Paleoceanogr Paleoclimatol. 36:e2020PA004037.

Strand E. 1908. Verzeichnis der von Oscar Neumann in Süd-Aethiopien gesammelten

Spinnen. Archiv für Naturgeschichte. 74:13–66.

Strand E. 1909. Spinnentiere von Südafrika und einigen Inseln gesammelt bei der deutschen

Südpolar-Expedition. Deutsche Südpolar-Expedition 1901-1905. Berlin: p. 541–596.

Strand E. 1928. Miscellanea nomenclatorica zoologica et palaeontologica, I-II. Archiv für

Naturgeschichte. 92:30–75.

Stratton G.E., Suter R.B., Miller P.R. 2004. Evolution of water surface locomotion by spiders:

a comparative approach. Biological Journal of the Linnean Society. 81:63–78.

Stratton G.E., Suter R.B. 2009. Water repellent properties of spiders: Topographical

variations and functional correlates. In: Grob S.N., editor. Functional Surfaces in

Biology. Dordrecht, Netherlands: Springer Netherlands. p. 77–95.

Sun Y., Liu J., Liang Y., Ji J., Liu W., Aitchison J.C., Sun J., Lu J., Song B., Xu Y., Zhang K.,

Liu Z. 2020. Cenozoic moisture fluctuations on the northeastern Tibetan Plateau and

association with global climatic conditions. J Asian Earth Sci. 200.

Sundevall C.J. 1833. Conspectus Arachnidum. Londini Gothorum, Sweden: C. F. Berling.

Suter R.B. 1999. Cheap transport for fishing spiders (Araneae, Pisauridae): The physics of

sailing on the water surface. Journal of Asian Earth Sciences. 27:489–496.

Suter R.B. 2003. Trichobothrial mediation of an aquatic escape response: Directional jumps

by the fishing spider, *Dolomedes triton*, foil frog attacks. Journal of Insect Science. 3:19.

Suter R.B. 2013. Spider locomotion on the water surface: Biomechanics and diversity.

Journal of Arachnology. 41:93–101.

Suter R.B., Gruenwald J. 2000a. Spider size and locomotion on the water surface (Araneae,

Pisauridae). Journal of Arachnology. 28:300–308.

Suter R.B., Gruenwald J. 2000b. Predator avoidance on the water surface? Kinematics and

efficacy of vertical jumping by *Dolomedes* (Araneae, Pisauridae). Journal of

Arachnology. 28:201–210.

Suter R.B., Stratton G.E., Miller P.R. 2004. Taxonomic variation among spiders in the ability

to repel water: Surface adhesion and hair density. Journal of Arachnology. 32:11–21.

Suter R.B., Wildman H. 1999. Locomotion on the water surface: Hydrodynamic constraints

on rowing velocity require a gait change. Journal of Experimental Biology. 202:2771–

2785.

Tagliacollo V.A., Lanfear R. 2018. Estimating Improved Partitioning Schemes for

Ultraconserved Elements. Molecular Biology and Evolution. 35, 2018:1798–1811.

Tamura K., Battistuzzi F.U., Billing-Ross P., Murillo O., Filipski A., Kumar S. 2012.

Estimating divergence times in large molecular phylogenies. Proceedings of the National

Academy of Sciences of the United States of America. 109:19333–19338.

Tamura K., Stecher G., Kumar S. 2021. MEGA11: Molecular Evolutionary Genetics Analysis

Version 11. Molecular Biology and Evolution. 38:3022–3027.

Tamura K., Tao Q., Kumar S. 2018. Theoretical foundation of the RelTime method for

estimating divergence times from variable evolutionary rates. Molecular biology and

evolution. 35:1770–1782.

Tanikawa A. 2003. Two new species and two newly recorded species of the spider family

Pisauridae (Arachnida: Araneae) from Japan. Acta Arachnologica. 52:35–42.

Tanikawa A. 2012. Further notes on the spiders of the genus *Dolomedes* (Araneae:

Pisauridae) from Japan. Acta Arachnologica. 61:11–17.

Tanikawa A., Miyashita T. 2008. A revision of Japanese spiders of the genus *Dolomedes*

(Araneae: Pisauridae) with its phylogeny based on mt-DNA. Acta Arachnologica.

57:19–35.

Thorell T. 1869. On European spiders. Review of the European genera of spiders, preceded

by some observations on zoological nomenclature. Nova Acta Regiae Societatis

Scientiarum Upsaliensis (3). 7:1–108.

Thorell T. 1877. Studi sui Ragni Malesi e Papuani. I. Ragni di Selebes raccolti nel 1874 dal

Dott. O. Beccari. Annali del Museo Civico di Storia Naturale di Genova. 10:341–637.

Thorell T. 1895. Decas aranearum in ins. Singapore a Cel. Th. Workman inventarum.

Bullettino della Società Entomologica Italiana. 26:321–355.

Thorell T. 1897. Araneae paucae Asiae australis. Bihang till Kongliga Svenska Vetenskaps-

Akademiens Handlingar. 22:1–36.

Thorell T. 1899. Araneae Camerunenses (Africae occidentalis) quas anno 1891 collegerunt

Cel. Dr Y. Sjöstedt aliique. Bihang till Kungliga Svenska Vetenskaps-Akademiens

Handlingar. 25:1–105.

Tonini L., Da Silva J.P., Filho A.S., Freitas J. 2016. Replacement names for two preoccupied

generic names in Arthropoda. Zootaxa. 4097:125–126.

Tucker M.A., Rogers T.L. 2014. Examining predator-prey body size, Trophic level and body

mass across marine and terrestrial mammals. Proceedings of the Royal Society B:

Biological Sciences. 281:20142103.

Udvardy M.D.F. 1975. A classification of the biogeographical provinces of the world. IUCN

Occasional Paper no. 18. Morges: IUCN.

Vink C.J., Dupérré N. 2010. Pisauridae (Arachnida: Araneae). Fauna of New Zealand. 64:1–

60.

Walckenaer C.A. 1837. Histoire naturelle des insectes. Aptères. Paris: Roret.

Wang J.F. 1993. Four new species of the spiders of Pisauridae from China (Arachnida:

Araneae). Acta Zootaxonomica Sinica. 18:156–161.

Wang P., Li Q., Li C.-F. 2014. Geology of the China Seas. Oxford: Elsevier.

Wheeler W.C., Coddington J.A., Crowley L.M., Dimitrov D., Goloboff P.A., Griswold C.E.,

Hormiga G., Prendini L., Ramírez M.J., Sierwald P., Almeida-Silva L., Alvarez-Padilla

F., Arnedo M.A., Benavides Silva L.R., Benjamin S.P., Bond J.E., Grismado C.J., Hasan

E., Hedin M., Izquierdo M.A., Labarque F.M., Ledford J., Lopardo L., Maddison W.P.,

Miller J.A., Piacentini L.N., Platnick N.I., Polotow D., Silva-Dávila D., Scharff N.,

Szűts T., Ubick D., Vink C.J., Wood H.M., Zhang J. 2017. The spider tree of life:

phylogeny of Araneae based on target-gene analyses from an extensive taxon sampling.

Cladistics. 33:574–616.

Williams D.S. 1979. The feeding behaviour of New Zealand *Dolomedes* species (Araneae:

Pisauridae). New Zealand Journal of Zoology. 6:95–105.

Wolff J.O., Guthrie R.D. 1985. Why are aquatic small mammals so large? Oikos. 45:365–

373.

Wolff J.O., Wierucka K., Paterno G.B., Coddington J.A., Hormiga G., Kelly M.B.J.,

Herberstein M.E., Ramírez M.J. 2022. Stabilized morphological evolution of spiders

despite mosaic changes in foraging ecology. Systematic Biology. 71:1487–1503.

World Spider Catalog. 2024. World Spider Catalog. Version 25. Natural History Museum

Bern. Available from http://wsc.nmbe.ch.

Wunderlich J. 1987. Die Spinnen der Kanarischen Inseln und Madeiras: Adaptive Radiation,

Biogeographie, Revisionen und Neubeschreibungen. Langen, Germany: Triops.

Wunderlich J. 2004. Fossil crab spiders (Araneae: Thomisidae) in Baltic and Dominican

amber. Beiträge zur Araneologie. 3:1747–1760.

Wunderlich J. 2008. Fossil and extant spiders (Araneae). Beiträge zur Araneologie. 5:1–870.

Xu X., Liu F., Cheng R.C., Chen J., Xu X., Zhang Z., Ono H., Pham D.S., Norma-Rashid Y.,

Arnedo M.A., Kuntner M., Li D. 2015. Extant primitively segmented spiders have

recently diversified from an ancient lineage. Proceedings of the Royal Society B:

Biological Sciences. 282:20142486.

Yang Z. 2007. PAML 4: Phylogenetic analysis by maximum likelihood. Molecular Biology

and Evolution. 24:1586–1591

Ye Z., Zhen Y., Damgaard J., Chen P., Zhu L., Zheng C., Bu W. 2018. Biogeography and

diversification of Holarctic water striders: Cenozoic temperature variation, habitat

shifting and multiple intercontinental dispersals. Systematic Entomology. 43:19–30.

Yin C.M., Peng X.J., Yan H.M., Bao Y.H., Xu X., Tang G., Zhou Q.S., Liu P. 2012. Fauna

Hunan: Araneae in Hunan, China. Changsha: Hunan Science and Technology Press.

Yu K.-P., Lo Y.-Y., Cheng R.-C., Raven R. J., Kuntner M. 2023. Discovery of a new intertidal

trapdoor spider of the genus *Idioctis* (Araneae: Barychelidae), with a generic range

extension to Taiwan. Journal of Arachnology. 51:238–248.

Yu K.-P., Kuntner M. 2024. Discovering unknown Madagascar biodiversity: integrative

taxonomy of raft spiders (Pisauridae: *Dolomedes*). PeerJ. 12:e16781.

Zachos J., Pagani M., Sloan L., Thomas E., Billups K. 2001. Trends, Rhythms, and

Aberrations in Global Climate 65 Ma to Present. Science. 292:686–693.

Zhang C., Rabiee M., Sayyari E., Mirarab S. 2018. ASTRAL-III: Polynomial Time Species

Tree Reconstruction from Partially Resolved Gene Trees. BMC Bioinformatics. 19:153.

Zhang C., Mirarab S. 2022. Weighting by gene tree uncertainty improves accuracy of quartet-

based species trees, Molecular Biology and Evolution. 39:msac215.

Zhang J., Li Z., Lai J., Zhang Z., Zhang F. 2023. A novel probe set for the phylogenomics and

evolution of RTA spiders. Cladistics. 39:116–128.

Zhang J.X. 2000. Taxonomy studies on Chinese spiders of the genus *Pisaura* (Araneae:

Pisauridae) I. Acta Arachnologica Sinica. 9:1–9.

Zhang J.X., Zhu M.S., Song D.X. 2004. A review of the Chinese nursery-web spiders

(Araneae: Pisauridae). Journal of Arachnology. 32:353–417.

Zhang Y.M., Williams J.L., Lucky A. 2019. Understanding UCEs: A comprehensive primer

on using ultraconserved elements for Arthropod phylogenomics. Insect Systematics and

Diversity. 3:1–12.

Zimmermann M., Spence J.R. 1989. Prey use of the fishing spider *Dolomedes triton*

(Pisauridae, Araneae): an important predator of the neuston community. Oecologia.

80:187–194.

1429

1431

FIGURE 1. A glimpse into lifestyle diversity of the genera currently in Pisauridae Simon,

1890: (a) Female of semi-aquatic *Dolomedes mizhoanus* Kishida, 1936 preying on a fish, (b)

Female of terrestrial *Dolomedes sulfureus* L. Koch, 1878 standing on vegetation; c–f,

terrestrial lifestyle without capture web: (c) A female *Pisaura mirabilis* (Clerck, 1757)

guarding her egg sac and nursery web, (d) Male of an unknown Pisauridae genus from

Madagascar hanging under a leaf, (e) Male of *Perenethis fascigera* Bösenberg and Strand,

1906 roving on grassland litter layer (credit: Han-Po Chang), (f) Female of *Hala* sp. standing

on the forest understory; g–l, terrestrial lifestyle with capture web: (g) Female of *Sphedanus*

quadrimaculatus Thorell, 1897 hanging under her 3-dimensional web with retreat made of

debris, (h) Female of *Caripetella madagascariensis* Lenz, 1886 hanging under her sheet web,

(i) Juvenile of *Euprosthenopsis* sp. on its sheet web (credit: Matjaž Bedjanič), (j) Female of

Dendrolycosa cf. *songi* (Zhang, 2000) standing on her sheet web (credit: Lok Ming Tang), (k)

Female of *Blandinia mahasoana* (Blandin, 1979) standing on her sheet web, (l) Female of

Polyboea zonaformis (Wang, 1993) standing in the center of her 3-dimentional web (credit:

Lok Ming Tang); m–p, semi-aquatic lifestyle, (m) Subadult male of *Thaumasia velox* Simon,

1898 floating on water, (n) Female of *Qianlingula* cf. *turbinata* Zhang et al., 2004 in hunting

posture, (o) Subadult female of *Nilus phipsoni* (F. O. Pickard-Cambridge, 1897) in hunting

posture, (p) Female of *Hygropoda higenaga* (Kishida, 1936) in hunting posture.

1451

FIGURE 2. Analytical design for investigating how biased taxon sampling can influence the

three evolutionary analyses; and how different phylogenetic hierarchies change the

correlation between lifestyles and morphological traits. a–b, analytical design for ancestral

state reconstruction using (a) the full taxon tree, and (b) 30 reduced taxon trees; c–d,

analytical design for comparative analyses between lifestyles and capture webs using (c) the

full taxon tree, and (d) 30 reduced taxon trees; e–g, analytical design for comparative

analyses between lifestyles and morphological traits using (e) the full taxon tree, (f) 30

reduced taxon trees, and (g) the *Dolomedes*-only tree.

1460

1461

FIGURE 3. Maximum likelihood phylogeny of the three focal clades reconstructed from the

sparse UCE matrix; nodal supports are labeled according to the legend at the bottom left,

nodes with full supports are represented by "*"; each terminal is color-coded by its

biogeographic region according to the map (modified from Udvardy 1975) at the top right

with the outgroups in light gray. UFboot: ultrafast bootstrap (Minh et al. 2013; Hoang et al.

2017), SH-aLRT: SH-like approximate likelihood ratio test (Guindon et al. 2010), BI:

1468   posterior probability (%) of Bayesian inference, MSC: posterior probability (%) of multi-

1469   species coalescence. See supplementary materials for the full branch lengths of the outgroups.

1470

1471

FIGURE 4. Maximum likelihood phylogeny of *Dolomedes* Latreille, 1804 and its relatives

reconstructed from the sparse UCE matrix; nodal supports are labeled according to the legend

at the bottom left, nodes with full supports are represented by "*"; each terminal is color-

coded by its biogeographic region according to the map at the bottom left; images in panels

a–g show habitus morphologies of the species labeled correspondingly in the phylogeny: (a)

an unknown morphospecies of the "raptor" clade hiding between leaves, (b) a female *D.*

minor L. Koch, 1876 guarding her nursery web, (c) a female *D. transfuga* Pocock, 1900 in

hunting posture, (d) a male *D. mizhoanus* Kishida, 1936 in hunting posture, (e) a female *D.*

okefinokensis Bishop, 1924 placed on a white background, (f) a *D. fimbriatus* (Clerck, 1757)

carrying her egg sac, and (g) a female *D. plantarius* (Clerck, 1757) standing on macrophytes.

UFboot: ultrafast bootstrap (Minh et al. 2013; Hoang et al. 2017), SH-aLRT: SH-like

approximate likelihood ratio test (Guindon et al. 2010), BI: posterior probability (%) of

Bayesian inference, MSC: posterior probability (%) of multi-species coalescence.

1485

FIGURE 5. Phylogenies of all three focal clades (a–c) calibrated by RelTime (black lines and blue bars; Tamura et al. 2012; 2018) and MCMCtree (gray lines and red bars; Yang 2007) with 95% confidence intervals and highest posterior densities of diversification time; red dots represent the fossil calibration points. A red curve of paleotemperature, modified from Zachos et al. (2001), shows the overlap between diversification and paleoclimate changes. Ma: million years ago.

1493

1494

FIGURE 6. Ancestral state reconstructions rendered from 100 stochastic character mapping on

1496   the full taxon tree and trait transition probabilities of: a–b, semi-aquatic vs terrestrial lifestyle

1497   with their best *k*-state Markov models (M*k* model, Pagel 1994; Lewis 2001) estimated from

1498   the (a) 30 reduced taxon trees, and (b) full taxon tree; c–d, capture web absence vs presence

1499   with their best M*k* model estimated from the (c) 30 reduced taxon trees, and (d) full taxon

1500   tree. PP: posterior probability.

1501

1502

FIGURE 7. Pagel's (1994) binary character correlation test between semi-aquatic vs terrestrial

lifestyle and capture web absence vs presence based on the: a–b, 30 reduced taxon trees with

their estimated and averaged trait transition probabilities in two-trait (a) independent all-

rates-different (ARD) models, and (b) dependent ARD models; c–d, full taxon tree and their

estimated trait transition probabilities in two-trait (c) independent ARD models, and (d) two-

trait dependent ARD models. The model selected by model selection highlighted by red

dotted line.

1510

FIGURE 8. Posterior density curves of moment-matching carapace width estimated via Monte

Carlo Sampler for Multivariate Generalized Linear Mixed Models (MCMCglmm; Hadfield

2010) from the (a) 30 reduced taxon trees and (b) full taxon tree; estimated mean carapace

width of each lifestyle pinpointed by the vertical dotted lines. The box plots below show the

medians (bold lines), first quartiles (Q1, left margins of the boxes), third quartiles (Q3, right

margins of the boxes), Q1 − 1.5 × interquartile range (IQR) (left whisker), and Q3 + 1.5 ×

IQR (right whisker) of all estimated values in both lifestyles.

1519

1520 TABLES

1521 TABLE 1. The 52 genera currently listed in Pisauridae Simon 1890 and their known

1522 distribution ranges and lifestyles.

Genus and author	Biogeographic realm	Lifestyle	Capture web	Data source	Include in this study
Afropisaura Blandin, 1979	Afrotropic	terrestrial	absent	Dippenaar-Schoeman et al. 2020	yes
Archipirata Simon, 1898	Indomalayan	unknown	unknown		no
Architis Simon, 1898	Neotropic	terrestrial	present	Santos 2007	no
Blandinia Tonini et al., 2016	Afrotropic	terrestrial	present	Yu original observation	yes
Bradystichus Simon, 1884	Australasian	terrestrial	absent	Platnick and Forster 1993	no
Caledomedes Raven and Hebron, 2018	Australasian	terrestrial	absent	Raven and Hebron 2018	no
Caripetella Strand, 1928	Afrotropic	terrestrial	present	Yu original observation	yes
Charminus Thorell, 1899	Afrotropic	terrestrial	absent	Dippenaar-Schoeman et al. 2020	yes
Chiasmopes Pavesi, 1883	Afrotropic	terrestrial	absent	Dippenaar-Schoeman et al. 2020	no
Cispinilus Roewer, 1955	Afrotropic	unknown	unknown		no
Cispius Simon, 1898	Afrotropic	terrestrial	absent	Dippenaar-Schoeman et al. 2020	yes
Cladycnis Simon, 1898	Palearctic	terrestrial	absent	Wunderlich 1987	yes
Conakrya Schmidt, 1956	Afrotropic	unknown	unknown		no

Dendrolycosa Doleschall, 1859	Afrotropic & Australasian & Indomalayan	terrestrial	present	Jäger 2011; Raven and Hebron 2018	yes
Dolomedes Latreille, 1804	Global	semi-aquatic	absent	see main text	yes
Eucamptopus Pocock, 1900	Indomalayan	unknown	unknown		no
Euprosthenops Pocock, 1897	Afrotropic	terrestrial	present	Dippenaar-Schoeman et al. 2020	yes
Euprosthenopsis Blandin, 1974	Afrotropic	terrestrial	present	Dippenaar-Schoeman et al. 2020	yes
Hala Jocqué, 1994	Afrotropic	terrestrial	absent	Jocqué 1994; Yu original observation	yes
Hygropoda Thorell, 1895	Afrotropic & Indomalayan	semi-aquatic	absent	Zhang et al. 2004; Dankittipakul et al. 2008	yes
Ilipula Simon, 1903	Indomalayan	unknown	unknown		no
Inola Davies, 1982	Indomalayan	terrestrial	present	Davies 1982	no
Mangromedes Raven and Hebron, 2018	Australasian	semi-aquatic	absent	Raven and Hebron 2018	no
Maypacius Simon, 1898	Afrotropic	terrestrial	absent	Dippenaar-Schoeman et al. 2020	yes
Megadolomedes Davies and Raven, 1980	Australasian	semi-aquatic	absent	Raven and Hebron 2018	yes
Nilus O. Pickard-Cambridge, 1876	Afrotropic & Indomalayan	semi-aquatic	absent	Dippenaar-Schoeman et al. 2020	yes
Ornodolomedes Raven and Hebron, 2018	Australasian	terrestrial	absent	Raven and Hebron 2018	yes
Papakula Strand, 1911	Indomalayan	unknown	unknown		no

Paracladycnis Blandin, 1979	Afrotropic	terrestrial	absent	Yu original observation	yes
Perenethis L. Koch, 1878	Afrotropic & Australasian & Indomalayan	terrestrial	absent	Sierwald 1997; Dippenaar-Schoeman et al. 2020	yes
Phalaeops Roewer, 1955	Afrotropic	unknown	unknown		no
Pisaura Simon, 1886	Palearctic	terrestrial	absent	Zhang et al. 2004	yes
Pisaurina Simon, 1898	Nearctic	terrestrial	absent	Carico 1973b	yes
Polyboea Thorell, 1895	Indomalayan	terrestrial	present	Sierwald 1997	yes
Qianlingula Zhang et al., 2004	Indomalayan	semi-aquatic	absent	Yu original observation	yes
Rothus Simon, 1898	Afrotropic	terrestrial	absent	Dippenaar-Schoeman et al. 2020	yes
Sphedanus Thorell, 1877	Indomalayan	terrestrial	present	Jäger 2011	yes
Stoliczka O. Pickard-Cambridge, 1885	Palearctic	unknown	unknown		no
Tallonia Simon, 1889	Afrotropic	unknown	unknown		yes
Tapinothele Simon, 1898	Afrotropic	unknown	unknown		no
Tapinothelella Strand, 1909	Afrotropic	unknown	unknown		no
Tapinothelops Roewer, 1955	Afrotropic	unknown	unknown		no
Tetragonophthalma Karsch, 1878	Afrotropic	terrestrial	present	Pett B. L. original observation; but see Sierwald 1997	yes
Thalassiopsis Roewer, 1955	Afrotropic	unknown	unknown		no
Thaumasia Perty, 1833	Neotropic	semi-aquatic	absent	Silva and Carico 2012	yes
Tinus F. O. Pickard-Cambridge, 1901	Neotropic	semi-aquatic	absent	Carico 1976	no

Tolma Jocqué, 1994	Afrotropic	terrestrial	absent	Jocqué 1994; Yu original observation	yes
Voraptipus Roewer, 1955	Afrotropic	unknown	unknown		no
Voraptus Simon, 1898	Afrotropic & Indomalayan	unknown	unknown		no
Vuattouxia Blandin, 1979	Afrotropic	unknown	unknown		no
Walrencea Blandin, 1979	Afrotropic	terrestrial	absent	Dippenaar-Schoeman et al. 2020	no

1523

1524　TABLE 2. Monophyly of Pisauridae Simon, 1890 and the affiliation of *Dolomedes* Latreille,

1525　1804 discussed and analyzed in the previous literature.

Literature	Evidence	Monophyly of Pisauridae	Affiliation of *Dolomedes*	Number of genera discussed
Simon 1898b	eye arrangement	Yes	Dolomedeae	33
Petrunkevitch 1928	eye arrangement	Yes	Thaumasiinae	after Simon 1898b
Lehtinen 1967	morphological comparisons	No	Dolomedidae	16
Griswold 1993	morphological cladogram	Yes	sister to *Pisaura*	2
Sierwald 1990	genital structure homologous	Yes	closely related to *Thalassius* (=*Nilus*)	55
Zhang et al. 2004	morphological cladogram	Yes	sister to *Thalassius* (=*Nilus*)	9
Santos 2007b	morphological cladogram	Yes	sister to *Pisaurina*	8
Bayer and Schönhofer 2013	multi-genes phylogeny	No	sister to *Thalassius* (=*Nilus*)	3

Moradmand et al. 2014	multi-genes phylogeny	No	sister to Thalassinae	2
Polotow et al. 2015	mutli-evidence phylogeny	No	NA	3
Albo et al. 2017	multi-genes phylogeny	No	separate clade, basal to Lycosioidea	5
Wheeler et al. 2017	multi-genes phylogeny	Yes	sister to *Bradystichus*, basal pisaurid	9
Fernández et al. 2018	transcriptomic phylogeny	Yes	sister to *Pisaurina*	2
Cheng and Piel 2018	transcriptomic phylogeny	Yes	separate clade, basal pisaurid	4
Piacentini and Ramírez 2019	multi-genes phylogeny	Yes	sister to *Bradystichus*, basal pisaurid	9
Kallal et al. 2021	transcriptomic phylogeny	Yes	separate clade, basal pisaurid	5
Kulkarni et al. 2023	UCE + multi-genes phylogeny	No	separate clade, sister to Lycosidae + Trechaleidae	13
Hazzi and Hormiga 2023	multi-genes phylogeny	No	separate clade, sister to Lycosidae + Trechaleidae	7

1526

TABLE 3. Raw data matrix of all the specimens used in the study, showing the region where they were collected and their traits used in the

analyses. All measurements are averages from female specimens. If no female data or specimen was available, we then used data from males

(italic numbers). If the terminals were represented by juveniles lacking species-level identification, the averages of the female of their genus

were used (numbers underlined). Measurements or data from a single specimen are in bold. All measurements are in millimeters and the

biogeographic realms were modified from Udvardy (1975). CW: carapace width; L1: leg I length; T1: tarsus I length; "-": outgroups that are not

in question. Voucher code and deposition: ABARA, YKP: National Chung Hsing University, Taiwan; ARA, KPARA: National Institute of

Biology, Slovenia; BINCO: Biodiversity Inventory for Conservation, Belgium; CASENT: California Academy of Sciences, USA; FMNH: Field

Museum, USA; MACN: Bernardino Rivadavia Natural Sciences Argentine Museum, Argentina; QM: Queensland Museum, Australia; RMCA:

Royal Museum of Central Africa, Belgium; SMF: Senckenberg Natural History Museum, Germany; WAM: Western Australian Museum,

Australia; ZRCENT: Lee Kong Chian Natural History Museum, Singapore.

Family	Species	Voucher code	Biogeographic realm	Country/Region	Admin	Life style	Capture web	CW	L1	T1	Data source

Family	Species	Voucher	Realm	Country	Location	Habitat	Web				Source
Lycosidae	*Aglaoctenus oblongus*	MACN-Ar41748	Neotropic	Argentina	Buenos Aires	terrestrial	web present	6.30	20.9	2.70	Santos et al. 2003
Lycosidae	*Hogna radiata*	KPARA_00623	Palearctic	Slovenia	Ljubljana	terrestrial	no web	5.74	19.7	2.60	Brady 2012; original data
Lycosidae	*Pardosa lugubris*	KPARA_00017	Palearctic	Slovenia	Ljubljana	terrestrial	no web	2.00	8.20	1.10	Nadolny and Kovblyuk 2012
Lycosidae	*Pirata piscatorius*	KPARA_00638	Palearctic	Slovenia	Jesenice	semi-aquatic	no web	3.77	12.25	1.33	original data
Lycosidae	*Trochosa* aff. *robusta*	KPARA_00639	Palearctic	Slovenia	Ljubljana	terrestrial	no web	3.16	9.48	1.50	original data
Trechaleidae	*Neoctenus* sp.	MACN-Ar43641	Neotropic	Argentina	Chaco	terrestrial	no web	3.65	14.84	1.92	original data
Pisauridae	*Afropisaura valida*	YKP-GB01	Afrotropic	Gabon	Anzem	terrestrial	no web	5.00	*21.68*	*2.81*	Sierwald 1997; original data
Pisauridae	*Blandinia mahasoana*	KPARA_00205	Afrotropic	Madagascar	Antsiranana	terrestrial	web present	2.40	19.61	2.70	original data
Pisauridae	*Caripetella madagascariensis*	KPARA_00178	Afrotropic	Madagascar	Toamasina	terrestrial	web present	4.70	37.23	3.54	original data
Pisauridae	*Charminus atomarius*	KPARA_00581	Afrotropic	South Africa	KwaZulu-Natal	terrestrial	no web	**2.47**	**15.71**	**2.07**	original data

Family	Species	Code	Realm	Country	Locality	Habitat	Web				Source
Pisauridae	*Cispius* sp.	KPARA_0 0587	Afrotropic	South Africa	Free State	terrestrial	no web	2.93	13.18	1.56	original data
Pisauridae	*Cladycnis insignis*	KPARA_0 0617	Palearctic	Spain	Santa Cruz de Tenerife	terrestrial	no web	4.24	21.8	2.82	original data
Pisauridae	*Dendrolycosa* cf. *songi*	YKP-Ar0935	Indomalayan	China	Hong Kong	terrestrial	web present	3.70	17.5	1.80	Zhang 2000
Pisauridae	*Dendrolycosa rossi*	KPARA_0 0180	Afrotropic	Madagascar	Antsiranana	terrestrial	web present	2.09	10.23	1.23	Silva and Griswold 2013; original data
Pisauridae	*Dolomedes* aff. *horishanus* JP	KPARA_ DH_2-1	Indomalayan	Japan	Okinawa	semi-aquatic	no web	7.26	25.55	2.88	original data
Pisauridae	*Dolomedes* aff. *mizhoanus* SING	KPARA_0 0590	Indomalayan	Singapore	Chestnut Ave	semi-aquatic	no web	5.61	16.58	2.50	original data
Pisauridae	*Dolomedes* aff. *raptor* HK	KPARA_0 0614	Indomalayan	China	Hong Kong	semi-aquatic	no web	8.69	32.54	3.85	original data
Pisauridae	*Dolomedes* aff. *raptor* LoTW	YKP-Ar1262	Indomalayan	Taiwan	Hualian	semi-aquatic	no web	**10.62**	**40.72**	**4.94**	original data
Pisauridae	*Dolomedes* aff. *raptor* NETW	YKP-Ar1251	Indomalayan	Taiwan	Taipei	semi-aquatic	no web	11.61	47.67	5.55	original data
Pisauridae	*Dolomedes* aff. *raptor*	KPARA_0	Indomalayan	Taiwan	Taitung	semi-	no web	9.53	39.05	4.71	original data

Family	Species	Code	Realm	Country	Region	Habitat	Web				Source
	SETW	0483				aquatic					
Pisauridae	*Dolomedes* aff. *raptor* WTW	ABARA_ 01442	Indomalayan	Taiwan	Taichung	semi-aquatic	no web	9.32	36.1	4.32	original data
Pisauridae	*Dolomedes* aff. *raptoroides* CN	SMF_DS5 7	Indomalayan	China	Sichuan	semi-aquatic	no web	9.72	40.47	4.86	original data
Pisauridae	*Dolomedes albineus*	KPARA_0 0097	Nearctic	USA	Kentucky	terrestrial	no web	5.75	27.35	3.15	original data
Pisauridae	*Dolomedes angustivirgatus*	KPARA_J P002	Palearctic	Japan	Chiba	semi-aquatic	no web	6.46	28.42	3.81	Tanikawa and Miyashita 2008; original data
Pisauridae	*Dolomedes aquaticus*	KPARA_ NZ_N38	Australasian	New Zealand	Blenheim	semi-aquatic	no web	8.41	28.13	3.86	Vink and Dupérré 2010; original data
Pisauridae	*Dolomedes bedjanic*	KPARA_0 0227	Afrotropic	Madagascar	Toamasina	semi-aquatic	no web	9.34	45.28	5.54	Yu and Kuntner 2024
Pisauridae	*Dolomedes bukhkaloi*	KPARA_ YURI-1	Palearctic	Russia	Omolon	semi-aquatic	no web	6.55	*23.43*	*3.26*	Marusik 1988; original data
Pisauridae	*Dolomedes* cf. *petalinus* HK	KPARA_0 0604	Indomalayan	China	Hong Kong	semi-aquatic	no web	**7.29**	**24.28**	**2.64**	original data
Pisauridae	*Dolomedes* cf.	KPARA_0	Indomalayan	China	Hong Kong	semi-	no web	9.66	38.24	4.51	original data

	raptoroides HK	0606				aquatic					
Pisauridae	*Dolomedes* cf. *raptoroides* VN	SMF_562 15	Indomalayan	Vietnam	Quang Binh	semi-aquatic	no web	10.10	42.55	5.17	original data
Pisauridae	*Dolomedes* CONsp1	BINCO_ DRC_22_ 0309	Afrotropic	D. R. Congo	Tshuapa	semi-aquatic	no web	*4.73*	*20.03*	*2.82*	original data
Pisauridae	*Dolomedes* CONsp2	BINCO_ DRC_21_ 0031_2	Afrotropic	D. R. Congo	Mai-Ndombe	semi-aquatic	no web	5.82	29.04	3.79	original data
Pisauridae	*Dolomedes crosbyi*	BINCO_ DRC_22_ 0285	Afrotropic	D. R. Congo	Tshuapa	semi-aquatic	no web	5.45	17.9	2.24	original data
Pisauridae	*Dolomedes dondalei*	KPARA_0 0597	Australasian	New Zealand	Canterbury	semi-aquatic	no web	8.79	37.7	5.07	Vink and Dupérré 2010; original data
Pisauridae	*Dolomedes facetus*	QM_S116 445	Australasian	Australia	Queensland	semi-aquatic	no web	5.71	23.73	2.59	Raven and Hebron 2018; original data
Pisauridae	*Dolomedes fimbriatus*	KPARA_0 0014	Palearctic	Slovenia	Malo Mraševo	semi-aquatic	no web	5.74	19.17	2.49	original data

Family	Species	Voucher	Realm	Country	Region	Habitat	Web	A	B	C	Reference
Pisauridae	*Dolomedes gracilipes*	BINCO_DRC_22_0286	Afrotropic	D. R. Congo	Tshuapa	semi-aquatic	no web	6.64	43.54	5.90	Lessert 1928, original data
Pisauridae	*Dolomedes gregoric*	KPARA_00253	Afrotropic	Madagascar	Toamasina	semi-aquatic	no web	7.68	38.86	4.81	Yu and Kuntner 2024
Pisauridae	Dolomedes horishanus	ABARA_01583	Indomalayan	Taiwan	Nantou	semi-aquatic	no web	7.26	25.55	2.88	original data
Pisauridae	Dolomedes hydatostella	KPARA_00258	Afrotropic	Madagascar	Antsiranana	semi-aquatic	no web	6.43	23.69	2.36	Yu and Kuntner 2024
Pisauridae	Dolomedes japonicus	KPARA_JP009	Palearctic	Japan	Tokyo	semi-aquatic	no web	10.49	43.65	5.95	Tanikawa and Miyashita 2008; original data
Pisauridae	Dolomedes kalanoro	KPARA_00185	Afrotropic	Madagascar	Toamasina	semi-aquatic	no web	9.15	44.03	5.38	Yu and Kuntner 2024
Pisauridae	*Dolomedes karijini*	WAM_T153133	Australasian	Australia	Western Australia	semi-aquatic	no web	10.17	39.39	4.93	Raven and Hebron 2018; original data
Pisauridae	Dolomedes lesserti	RMCA_214998	Afrotropic	Benin	Ouémé	semi-aquatic	no web	5.63	20.97	2.73	original data
Pisauridae	*Dolomedes minor*	KPARA_	Australasian	New Zealand	Southland	terrestrial	no web	6.65	22.39	3.07	Vink and

		NZ_R75								Dupérré 2010; original data
Pisauridae	*Dolomedes mizhoanus*	ABARA_ 01630	Indomalayan	Taiwan	Taichung	semi-aquatic	no web	8.53	36.49	4.57	Zhang et al. 2004; Yin et al. 2012; original data
Pisauridae	*Dolomedes nigrimaculatus*	SMF_DS8 2	Palearctic	China	Sichuan	terrestrial	no web	8.18	33.68	3.96	Chae et al. 2023
Pisauridae	*Dolomedes okefinokensis*	KPARA_0 0620	Nearctic	USA	Florida	semi-aquatic	no web	9.18	40.45	4.74	original data
Pisauridae	*Dolomedes orion*	KPARA_J P012-1	Indomalayan	Japan	Okinawa	semi-aquatic	no web	16.17	59.78	6.67	Tanikawa 2003; original data
Pisauridae	*Dolomedes pegasus*	KPARA_J P016	Palearctic	Japan	Aomori	semi-aquatic	no web	8.33	27.07	3.41	Tanikawa 2012; original data
Pisauridae	*Dolomedes plantarius*	KPARA_0 0021	Palearctic	Slovenia	Jesenice	semi-aquatic	no web	7.24	25.38	3.31	original data
Pisauridae	*Dolomedes raptor*	KPARA_J P019-2	Palearctic	Japan	Kanagawa	semi-aquatic	no web	8.00	33.78	4.15	Ono 2009; original data
Pisauridae	*Dolomedes reuniascar*	KPARA_0 0408	Afrotropic	Madagascar	Antsiranana	semi-aquatic	no web	6.19	23.39	2.76	original data

Pisauridae	*Dolomedes rotundus*	KPARA_0 0240	Afrotropic	Madagascar	Toamasina	semi-aquatic	no web	6.20	22.3	2.31	Yu and Kuntner 2024
Pisauridae	*Dolomedes saganus*	YKP-Ar0684	Palearctic	China	Fujian	semi-aquatic	no web	7.00	27.69	3.79	Tanikawa and Miyashita 2008
Pisauridae	*Dolomedes schauinslandi*	KPARA_ NZ_DS-M1	Australasian	New Zealand	Rangatira Island	terrestrial	no web	9.73	30.3	3.76	Vink and Dupérré 2010; original data
Pisauridae	*Dolomedes scriptus*	KPARA_0 0118	Nearctic	USA	Kentucky	semi-aquatic	no web	7.25	31.67	4.46	original data
Pisauridae	*Dolomedes senilis*	KPARA_ DS-2-2	Palearctic	Japan	Hokkaido	semi-aquatic	no web	7.00	22.08	2.92	Tanikawa 2012
Pisauridae	*Dolomedes silvicola*	KPARA_J P024-1	Palearctic	Japan	Kanagawa	terrestrial	no web	5.28	28.14	4.14	Tanikawa and Miyashita 2008
Pisauridae	*Dolomedes spathularis*	ZRCENT _61059	Indomalayan	Malaysia	Petaling	semi-aquatic	no web	7.65	26.87	2.79	original data
Pisauridae	*Dolomedes striatus*	FMNH_2 857312	Nearctic	USA	Illinois	semi-aquatic	no web	*4.20*	*18.83*	*2.93*	original data
Pisauridae	*Dolomedes sulfureus*	KPARA_J P027	Palearctic	Japan	Kanagawa	terrestrial	no web	8.19	38.64	5.54	Tanikawa and Miyashita 2008; original data

Family	Species	Code	Realm	Country	Region	Habitat	Web				Source
Pisauridae	*Dolomedes tenebrosus*	KPARA_0 0100	Nearctic	USA	Kentucky	terrestrial	no web	8.68	37.62	4.33	original data
Pisauridae	*Dolomedes transfuga*	RMCA_2 45953	Afrotropic	Guinea	Selinngbala	semi-aquatic	no web	10.7	55.08	7.25	original data
Pisauridae	*Dolomedes triton*	KPARA_0 0115	Nearctic	Canada	Prince Edward Island	semi-aquatic	no web	6.10	22.67	2.64	original data
Pisauridae	*Dolomedes vittatus*	KPARA_0 0088	Nearctic	USA	Kentucky	semi-aquatic	no web	9.75	40.63	5.14	original data
Pisauridae	*Dolomedes yawatai*	KPARA_J P029-1	Indomalayan	Japan	Okinawa	semi-aquatic	no web	10.72	46.1	5.12	Ono 2002, original data
Pisauridae	*Euprosthenops australis*	KPARA_0 0577	Afrotropic	South Africa	Free State	terrestrial	web present	**5.89**	**40.72**	**4.48**	original data
Pisauridae	*Euprosthenopsis* sp.	KPARA_0 0624	Afrotropic	Madagascar	Toliara	terrestrial	web present	3.90	23.26	2.61	original data
Pisauridae	*Hala* sp.	KPARA_0 0149	Afrotropic	Madagascar	Antsiranana	terrestrial	no web	<u>1.89</u>	<u>9.48</u>	<u>1.04</u>	Jocqué 1994; original data
Pisauridae	*Hygropoda higenaga*	YKP-Ar1199	Indomalayan	Taiwan	Taitung	semi-aquatic	no web	3.55	38.92	8.42	Wang 1993
Pisauridae	*Hygropoda* MADsp.	KPARA_0	Afrotropic	Madagascar	Antsiranana	semi-	no web	3.10	31.38	5.19	original data

		0198				aquatic					
Pisauridae	*Maypacius bilineatus*	KPARA_0 0578	Afrotropic	South Africa	KwaZulu-Natal	terrestrial	no web	2.40	19.8	2.50	Strand 1908
Pisauridae	*Megadolomedes johndouglasi*	QM_S109 171	Australasian	Australia	Tasmania	semi-aquatic	no web	8.67	46.83	8.17	Raven and Hebron 2018
Pisauridae	*Nilus majungensis*	KPARA_0 0223	Afrotropic	Madagascar	Toamasina	semi-aquatic	no web	7.12	32.94	3.33	original data
Pisauridae	*Nilus phipsoni*	ABARA_ 01717	Afrotropic	Taiwan	Pingtung	semi-aquatic	no web	8.29	33.3	3.11	original data
Pisauridae	*Ornodolomedes* sp.	KPARA_ QMgift	Australasia	Australia	Queensland	terrestrial	no web	3.75	16.26	1.84	Raven and Hebron 2018
Pisauridae	*Paracladycnis vis*	KPARA_0 0176	Afrotropic	Madagascar	Antananarivo	terrestrial	no web	5.71	35.76	4.12	Silva and Sierwald 2013
Pisauridae	*Perenethis fascigera*	ABARA_ 01868	Indomalayan	Taiwan	Pingtung	terrestrial	no web	3.12	20.31	2.47	original data
Pisauridae	*Pisaura mirabilis*	KPARA_0 0040	Palearctic	Slovenia	Spodnje Prapreče	terrestrial	no web	3.39	18.75	2.29	Nadolny et al. 2012
Pisauridae	Pisauridae gen. MADsp.	KPARA_0 0138	Afrotropic	Madagascar	Antsiranana	terrestrial	no web	2.06	18.36	3.63	original data
Pisauridae	*Pisaurina mira*	FMNH_3	Nearctic	USA	Indiana	terrestrial	no web	5.26	25.11	2.71	original data

195381

		KPARA_ HK439	Indomalayan	China	Hong Kong	terrestrial	web present	2.60	20.4	2.5	Wang 1993
Pisauridae	*Polyboea zonaformis*	KPARA_ HK439	Indomalayan	China	Hong Kong	terrestrial	web present	2.60	20.4	2.5	Wang 1993
Pisauridae	*Qianlingula* cf. *turbinata*	YKP-Ar1265	Indomalayan	Taiwan	Hualian	semi-aquatic	no web	7.19	46.15	3.77	Zhang et al. 2004; original data
Pisauridae	*Rothus vittatus*	KPARA_0 0576	Afrotropic	South Africa	Western Cape	terrestrial	no web	3.26	21.84	3.19	Silva and Sierwald 2015; original data
Pisauridae	*Sphedanus quadrimaculatus*	YKP-Ar0320	Indomalayan	Malaysia	Sabah	terrestrial	web present	3.08	15.12	1.84	original data
Pisauridae	*Tallonia picta*	CASENT _9017992	Afrotropic	Madagascar	Antsiranana	unknown	unknown	4.45	25.47	2.59	Silva and Sierwald 2013
Pisauridae	*Tetragonophthalma vulpina*	BINCO_ DRC_22_ 0258_12	Afrotropic	D. R. Congo	Tshuapa	terrestrial	web present	6.68	42.82	4.17	original data
Pisauridae	*Thaumasia velox*	MACN-Ar34778	Neotropic	Argentina	Corrientes	semi-aquatic	no web	6.31	26.72	2.94	Silva and Carico 2012; original data
Pisauridae	*Tolma* sp.	KPARA_0	Afrotropic	Madagascar	Antsiranana	terrestrial	no web	2.22	10.49	1.12	original data

		0141									
Oxyopidae	*Peucetia* MADsp.	ARA1147	Afrotropic	Madagascar	Toamasina	-	-	-	-	-	-
Psechridae	*Fecenia cylindrata*	ARA4196	Indomalayan	China	Yunnan	-	-	-	-	-	-
Psechridae	*Psechrus* sp.	KPARA_0 0641	Indomalayan	China	Yunnan	-	-	-	-	-	-
Thomisidae	*Xysticus* sp.	MACN-Ar43641	Palearctic	Slovenia	Ljubljana	-	-	-	-	-	-
Viridasiidae	*Viridasius* sp.	KPARA_0 0152	Afrotropic	Madagascar	Antsiranana	-	-	-	-	-	-
Zoropsidae	*Zoropsis spinimana*	KPARA_0 0635	Palearctic	Slovenia	Ljubljana	-	-	-	-	-	-

1537

1538   TABLE 4. AIC comparisons among the *k*-state Markov models (Mk model, Pagel

1539   1994; Lewis 2001) with different trait transition rates assumptions conducted on the

1540   30 reduced taxon trees and the full taxon tree. ER: equal rates, ARD: all-rates-

1541   difference.

Data set	Trait	model	logL	AIC	ΔAIC
		ER	-21.115	44.230	3.932
		ARD	-18.971	41.941	1.643
	lifestyle	semi-aquatic to terrestrial	-19.149	40.298	0.000
30 reduced		terrestrial to semi-aquatic	-25.661	53.321	13.023
taxon trees		ER	-21.892	45.784	1.466
	capture	ARD	-20.159	44.317	0.000
	web	no web to web present	-22.362	46.724	2.407
		web present to no web	-21.239	44.479	0.161

	Trait	model	Average logL	Average AIC	ΔAIC
		ER	-43.226	88.452	2.403
		ARD	-41.793	87.587	1.537
	lifestyle	semi-aquatic to terrestrial	-42.025	86.049	0.000
Full taxon		terrestrial to semi-aquatic	-63.244	128.488	42.438
tree		ER	-23.621	49.242	4.753
	capture	ARD	-21.244	46.489	2.000
	web	no web to web present	-24.159	50.318	5.830
		web present to no web	-21.244	44.489	0.000

1542

TABLE 5. Statistic tables of Monte Carlo Sampler for Multivariate Generalized Linear

Mixed Models (MCMCglmm; Hadfield 2010) between lifestyles and carapace width,

relative length of teg I, and relative length of tarsus I, showing the correlation between

morphological traits and semi-aquatic versus terrestrial lifestyles. CW: carapace

width, L1: leg I length, T1: tarsus I length, 95HPD: 95% highest posterior density,

pMCMC, twice the probabilities of the posterior values that are above or below zero.

Data set	Trait	Factor	Average post mean	Average 95HPD lower bound	Average 95HPD upper bound	Average effective samples	Average pMCMC
	CW	intercept	6.149	4.063	8.182	1510	**< 0.001**
	CW	life style: terrestrial	-2.408	-3.911	-0.858	1549	**0.014**
30 reduced taxon trees	L1/C W	intercept	5.792	4.064	7.483	1545	**< 0.001**
	L1/C W	life style: terrestrial	-0.049	-1.222	1.112	1535	0.812
	T1/L1	intercept	0.131	0.090	0.172	1518	**< 0.001**
	T1/L1	life style: terrestrial	0.005	-0.031	0.041	1498	0.789

	Trait	Factor	Post mean	95HPD lower bound	95HPD upper bound	Effective samples	pMCMC
	CW	intercept	5.451	3.131	7.769	1500	**< 0.001**
	CW	life style: terrestrial	-1.194	-2.210	-0.074	1500	**0.025**
full taxon tree	L1/C W	intercept	5.595	4.071	7.007	1636	**< 0.001**
	L1/C W	life style: terrestrial	0.263	-0.148	0.702	1500	0.231
	T1/L1	intercept	0.132	0.103	0.160	1500	**< 0.001**
	T1/L1	life style: terrestrial	0.002	-0.015	0.019	1500	0.803
	CW	intercept	7.821	5.196	9.878	1665	**< 0.001**

	CW	life style: terrestrial	-0.571	-2.007	0.546	1500	0.361
Dolomedes -only tree	L1/CW	intercept	4.037	3.200	4.736	1500	**< 0.001**
	L1/CW	life style: terrestrial	0.315	-0.106	0.759	1500	0.157
	T1/L1	intercept	0.124	0.092	0.156	1377	**< 0.001**
	T1/L1	life style: terrestrial	-0.001	-0.027	0.032	1500	0.944

1549