Is variation in female aggressiveness across *Drosophila* species associated with

reproductive potential?

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Data and code accessibility

Data and code for this paper are available in the Dryad repository:

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Abstract

Aggression is a key determinant of fitness in many species, mediating access to mates, food, and

breeding sites. Variation in intrasexual aggression across species is likely driven by variation in resource

availability and distribution. To date, most studies have focused on male aggression, but female

aggression is common and has fitness consequences. While males primarily compete over access to

mates, females are likely to compete over resources to maximize offspring quantity and/or quality, such

as food or breeding sites. To investigate why reproduction and aggression are often linked, we tested

predictions about resource valuation, life-history, and physiological correlates of reproduction. We used

machine learning classifiers developed for *D. melanogaster* to identify and quantify aggressive behaviors

displayed in the presence of yeast for mated and unmated females of eight *Drosophila* species. We found

that female aggression was linked to ovariole number across species, suggesting that females that lay

more eggs are more aggressive. A need for resources for egg production or oviposition sites or similar

physiological mechanisms regulating aggression and reproduction may therefore be drivers of female

aggression.

Introduction

Aggression towards members of the same species is almost ubiquitous throughout the animal kingdom. In many species, aggression is a key determinant of fitness, establishing an individual's social dominance, access to resources (mates, food, shelter, and/or territory), and vulnerability to predation (Clutton-Brock and Huchard 2013b). Intrasexual aggression has been extensively studied in males, but, until recently, relatively neglected in females (Ah-King 2022). It is now commonly accepted that females compete aggressively in a wide range of taxa, with consequences for survival and reproduction (Clutton-Brock and Huchard 2013a; Rosvall 2013; Stockley and Campbell 2013).

Female-female aggression (hereafter 'female aggression') is diverse. Among species, female aggression varies in the frequency, intensity, and types of aggression, as well as the diversity of resources females compete over (e.g. food, nesting sites, territory,Clutton-Brock and Huchard 2013b; Rosvall 2011; Stockley and Campbell 2013). Within species, females commonly shift their levels of aggression in response to their current reproductive stage. For example, oestrous baboons fight over mates (Huchard and Cowlishaw 2011), mosquitofish increase fighting in the late stages of pregnancy (Seebacher et al. 2013), *Drosophila melanogaster* females increase fighting after mating (Bath et al. 2017; Nilsen et al. 2004). Although females fight predominantly over resources for reproduction, we have few testable predictions about the relationship between reproduction and patterns of female aggression across species because the majority of studies on female aggression focus on the benefits and costs within a species (Clutton-Brock and Huchard 2013a; Rosvall 2011; Stockley and Campbell 2013). The only systematic study (to our knowledge) investigating female aggression and resource valuation across taxa, found increased territorial aggression in cavity-nesting bird species relative to sister species that nested in open nests, suggesting selection for females to fight to protect limited nesting sites (Lipshutz and Rosvall 2021).

We propose that reproduction and female aggression may be related across species because of perceived resource value for maximizing reproduction, because life-history is closely tied to reproductive strategies, and because the physiology of reproduction is tightly linked to the physiology of aggression (all discussed below). We set out to test these ideas in a set of *Drosophila* species chosen for their differences in reproductive traits.

***Drosophila* species**

The genus *Drosophila* encompasses over 2000 species (Markow and O'Grady 2008), which vary

in their ecology, physiology, mating behavior and life-history, all of which are likely to influence levels of

aggression (Markow and O'Grady 2005a; Markow and O'Grady 2005b; Markow and O'Grady 2008).

Aggression is understudied in non-melanogaster *Drosophila* species, with a few exceptions (Amino and

Takashi 2020; Belenioti and Chaniotakis 2020; Boake et al. 1998; Gupta et al. 2019; Ringo et al. 1983).

Most studies have focused on male aggression, though unmated females have been examined in a total

of 10 species other than *D. melanogaster* (Belenioti and Chaniotakis 2020; Shultzaberger et al. 2019).

Female aggression in *D. melanogaster* consists of a range of behaviors, from high-intensity headbutts to

low-intensity wing threats (Nilsen et al. 2004). Female aggression is linked to reproduction in *Drosophila*

melanogaster: mated females fight more than unmated females over food (Nilsen et al. 2004, Bath et al.

2017), while unmated females attack mating pairs but mated females do not (Gaspar et al. 2022). The

change in aggression with mating status over food is stimulated by the transfer of sperm and seminal fluid

proteins during mating (Bath et al. 2017) and may be driven by increased need for protein-rich food to fuel

egg production in mated females (Lee et al. 2013). The clear relationship between reproduction and

aggression in *D. melanogaster* sets the groundwork to test if this relationship holds when comparing

species with different reproductive strategies (e.g. how many eggs they lay, their mating system, and

lifespan).

In *Drosophila*, female fecundity is determined by the number of ovarioles (egg-producing

compartments in the ovaries), each of which has the capacity to produce two eggs per day (David 1970);

species with more ovarioles produce more offspring than females of species with few ovarioles (R'kha et

al. 1997; Sarikaya et al. 2019). Ovariole number varies across *Drosophila* species and is strongly

correlated with ecology (Sarikaya et al. 2019). Thus, the reproductive potential of females of different

species is limited by physiology. Because females need protein to produce eggs after mating (Walker et

al. 2015), competition for protein-rich food may lead to the possibility that variation in reproductive

potential (i.e. ovariole number) across species may drive aggressive responses to mating due to

increased need or valuation of the contested resource (see Predictions below).

Mating systems also vary across the genus: in some species, females mate multiply each day

whereas in other species females have a refractory period or may never remate (Markow and O'Grady

2005b). Characteristics of the male ejaculate influence female remating rate. In *Drosophila melanogaster*,

proteins transferred with sperm modify female physiology and behavior, inducing a refractory period (Avila

et al. 2011). In addition, the quantity of sperm transferred affects the female's propensity to remate, with

females receiving few sperm needing to remate often to maximize their reproductive potential (Markow

2002). Across species, the amount of sperm transferred is inversely related to the size of sperm (Markow

1996; Pitnick 1996; Syed et al. 2021) and sperm investment, either in size or number of sperm, varies

across the genus (Lüpold et al. 2016; Markow 1996; Pitnick et al. 1995; Schärer et al. 2008; Syed et al.

2021). Variation in remating rate and sperm provisioning across species is likely to result in differences

among species in the relationship between a female's mating status and her investment in aggression.

In this study, we tested if reproductive potential is related to female aggression in eight species of

Drosophila. We used a comparative approach among *Drosophila* species to test the predicted drivers of

female aggression across species: remating rate, sperm length, ovariole number, and lifespan. We used

machine learning to automatically score common aggressive behaviors in eight *Drosophila* species,

measuring aggression in unmated and mated females and investigating cross-species relationships with

reproductive drivers of aggression. We tested the specific predictions below.

Resource valuation

Aggression theory predicts that an individual will be most motivated to fight (and continue fighting) when it

perceives the resource it is competing over to be valuable (Arnott and Elwood 2008). The reproductive

stage of a female modifies how valuable resources are at any given time, influencing her likelihood of

engaging, persisting in, and escalating aggressive contests (Huchard and Cowlishaw 2011; Palanza et al.

1996; Seebacher et al. 2013). By framing resources in terms of their value to females, we can predict

when females should compete over resources within and across species and when females should

display the most aggression. For some species, resources required for offspring production and

provisioning (e.g. food, nesting sites) may be the most limiting resource, causing aggression to peak at

times when fecundity is at a maximum (Rosvall 2008; Seebacher et al. 2013; Viblanc et al. 2016). For

other species, mates or the resources they provide (e.g. sperm, nuptial gifts, paternal care) may be a

limited resource, resulting in increased female competition before mating for access to mates (e.g. Bro-

Jørgensen 2002; Gwynne 1991; Papadopoulos et al. 2009). We, therefore, predict that that the mating

system and reproductive potential of a species may explain patterns of female aggression.

We focused on aggression over access to food as this is a form of competition common across

taxa (Stockley and Campbell 2013). Reproductive costs include the energy demands for production of

offspring, gestation, lactation, parental care, and locating appropriate breeding sites (Stockley and

Campbell 2013). The more females invest in reproduction, the greater their energetic demands, leading to

increased consumption of food in general, as well as increased intake of specific nutrients (e.g. protein,

specific amino acids (Carvalho et al. 2006; Lorenz and Gäde 2009). Because females need food for

investment in reproduction, females should be motivated to fight over access to food when it is limited,

leading to a positive correlation between investment in reproduction and aggression over food. Thus

investment in reproduction leads to higher resource need and/or valuation. When females compete for

food, females should be most aggressive when energetic demands are highest, for example during

gestation and lactation for mammals, during incubation for birds, and during egg production after mating

for most invertebrates (Gittleman and Thompson 1988; Nord and Williams 2015; Shingleton 2015). We

can also use this theory to predict patterns of aggression across species: species that have high

reproductive energy demands (i.e. producing offspring at high rates) will be more aggressive over food

than those with low demands.

With respect to *Drosophila*, resource valuation leads to several predictions. 1) Within species,

mated females will show more aggression than unmated females as they have increased egg production

and the need to find oviposition sites to lay eggs. Producing more eggs requires increased consumption

of food, driving increased aggression after mating in all species. 2). Among species, species with more

ovarioles will display more aggression than those with fewer (fig. 1A), as they produce more eggs so have

higher energy and resource requirements. 3) Among species, we also predict that there will be a greater

change in aggression after mating in species with more ovarioles as they should exhibit the largest

change in resource demands. 4) Across species, we expect a negative correlation between mated female

aggression and sperm length (fig. 1B). Species with longer sperm transfer fewer sperm in each mating,

meaning that fewer eggs are fertilized than in species with shorter sperm. We would therefore expect

females receiving few, long sperm to have low energetic demands from egg laying and therefore show

lower rates of aggression than females of species who receive many, short sperm.

**Figure 1: Predictions for the relationship between aggression and potential drivers of aggression
across species** A) All theories predict a positive relationship between reproduction (measured as
ovariole number) and female aggression. The exact nature of this relationship is unclear, so we have
plotted a solid line to indicate a linear relationship and a dashed line to indicate a situation where a
maximum level of aggression is reached and cannot increase any further. B) Predicted negative
correlation between sperm length and female aggression. C) Predicted negative correlation between
lifespan and female aggression. D) The underlying assumption for life-history theory is that there is a
negative relationship between reproduction and lifespan.

Life-history

Life-history theory provides additional insight into the link between reproduction and female

aggression across species. Species lie along a continuum of life-history strategies: on one end lie species

with 'fast' life-histories (short lifespans and fast rates of reproduction) while at the other end are species

with 'slow' life-histories (long lifespans and slow rates of reproduction, fig 1D, Stearns 1989). Behaviors

that increase access to resources or mates, such as aggression or boldness, come with costs in the form

of increased mortality, and are likely to coevolve negatively with lifespan (fig. 1C) for two different reasons

(Eckerström-Liedholm et al. 2016): 1) Species with different life-history strategies differ in their relative

energetic demands. Species with fast life-histories have high productivity and short lifespans so need

energetic resources to grow fast and start reproducing quickly (Dammhahn et al. 2018). Fast species

should therefore exhibit high risk-taking behaviors to acquire resources due to the high relative value of

the resources. 2) Species will vary in their valuation of current vs future reproduction. Traits that prioritize

current reproduction, even at the cost of future reproduction, are favored in short-lived species that

receive the benefits of such behaviors but tend to die before facing the costs (Eckerström-Liedholm et al.

2016). Aggression increases current reproduction through increasing access to vital resources but with an

elevated risk of injury or other costs (Smith and Blumstein 2008).

We can test the trade-off between reproduction and lifespan in our dataset with ovariole number

and lifespan with the following predictions: 1) Within species, mated females will fight more than unmated

females as there is no point taking risky behavior if an individual is not yet reproducing. 2) Among

species, faster-reproducing species (those with more ovarioles) will display more aggression. We

therefore expect to see a positive correlation between ovariole number and aggression (fig. 1A). 3)

Among species, we also predict that there will be a greater change in aggression after mating in species

with more ovarioles as they should need to increase their risk-taking more due to increased reproduction.

4) Across species, shorter-lived species will display more aggression, leading to a negative correlation

between lifespan and aggression (fig. 1C).

Physiology

Changes in aggression associated with reproduction may be a by-product of changes in

physiology involved in reproduction. Many of the hormones that regulate reproductive traits, including egg

production, ovulation, mating, gestation and lactation, are also implicated in regulating aggression. For

example, the monoamine octopamine, the invertebrate counterpart to vertebrate norepinephrine,

functions as a neurotransmitter, neuromodulator, and neurohormone (White et al. 2021). Octopamine

signaling regulates a range of reproductive behaviors, including ovulation, sperm storage, and

physiological changes in mated females in a range of insect species (White et al. 2021). Octopamine also

regulates aggression in both males and females, suggesting a possible link between reproductive and

aggressive regulators (Cunningham et al. 2014; Jia et al. 2021; Zhou et al. 2008). In vertebrates,

testosterone regulates aggression in both sexes and mediates ovulation or fertility (reviewed in Rosvall et

al. 2020). Female testosterone levels fluctuate seasonally in bird species, peaking during territory

establishment and diminishing during periods of parental care (reviewed in Rosvall et al. 2020),

suggesting adaptive expression of hormones that are involved in both aggression and reproduction.

Changes in aggression regulated by hormones may therefore track changes in reproductive state, and

induce changes in aggression, but this may be a side-effect of reproduction rather than an adaptive trait

(Sibbald and Plowright 2015). Conversely, the use of the same mechanisms in reproduction and

aggression may be an adaptive way of linking the two to ensure aggression occurs at the optimal time.

If shared physiological mechanisms regulate reproduction and aggression, we predict: 1) Within

species, mated females will be more aggressive than unmated females due to the changes in physiology

stimulated by mating regulating both reproduction and aggression. 2) Across species, aggression and

ovariole number will be positively correlated. If the rate of aggression increases in response to the rate of

reproduction due to similar hormones regulating the behaviors, as reproductive rate increases, so should

aggression. 3) Because females that have more ovarioles should exhibit larger changes in physiology

due to reproduction, females in species with more ovarioles will have a greater increase in aggression

after mating than those with fewer. 4) Because females can only respond to sperm after mating, any

relationship between sperm production and female aggression should only be seen in mated, and not

unmated females. 5) If the number of sperm produced by a male is related to change in aggression, then

a larger response to mating will occur in species that produce many, small sperm.

To test these predictions and measure female aggression in a range of species, we used machine

learning to automatically score headbutting and fencing in mated and unmated females in eight

Drosophila species. The development of software to track animal locomotion and behavior has improved

our ability to study broad-scale and individual-level behaviors (Dell et al. 2014), but the use of such

analyses identifying and classifying behaviors is usually focused on one species at a time (though see

DeLellis et al. 2014; Hernández et al. 2021). Here we examined eight *Drosophila* species with a range of

phylogenetic relatedness, and differences in reproductive and life-history traits, to investigate cross-

species relationships with reproductive drivers of aggression.

Materials and Methods

Strain maintenance and unmated fly collection

All strains (table S1) were maintained on standard cornmeal-molasses food in multiple 75 ml

bottles at 25˚C on a 12:12 light:dark cycle. Unmated males and females were collected on ice within a

day of eclosion except for species that are sexually mature in a day or less (*D. willistoni* and *D. nebulosa*),

which were collected within 7 hours of eclosion. Males were housed in 12.5 ml vials in groups of 10 with

ad libitum access to food. Females were housed individually.

Aggression assay

The aggression assay followed the method of Bath et al. (2017). We collected flies before the age

at which they become sexually mature (table S1). When sexual maturity for one sex was unknown, we

collected flies at the same age as the sex for which we had age at maturity information on the basis that if

one sex was not mature, the other sex could not mate even if they were mature (e.g. *D. ananassae* males

are reported to be sexually mature at 4 days old, so we collected both males and females before they

were 4 days-old). Once both males and females were sexually mature, a subset of the unmated females

were paired with one or two males of their species at lights on for up to two hours or until the first mating

finished. Females that mated were separated from the male and transferred to fresh vials to be kept

overnight. Females that did not mate were discarded. Unmated females that were never exposed to

males were moved to fresh vials at the same time as mated females to be kept in identical conditions

overnight.

At 24-hrs after mating, single mated and unmated females of a single species were separately

placed in individual empty vials with a moist cotton ball. Females remained in these vials for two hours to

increase their motivation to compete for food (Edwards et al. 2006). Each female was then paired with a

female from the same treatment (mated or unmated) in a contest arena (20 mm in diameter), with apple

juice agar and a small patch of yeast paste (Supplementary Video). Up to 20 dyads were video recorded

simultaneously with a Basler ac A2440-75µm camera. Where possible, equal numbers of mated and

unmated dyads were included in each video to control for time-of-day effects. Pairs were given 5 minutes

to acclimatize to the arena. We recorded videos for 15 minutes using Kinovea software (v 0.9.5;

https://www.kinovea.org) at 30 frames-per-second. Flies were subsequently discarded.

Automated tracking and machine learning methods

Following recording, we used automated tracking and machine learning to score flies' aggressive

and movement behavior (Bath et al. 2020). Briefly, the Caltech fly tracker (Eyjolfsdottir et al. 2014)

determined the location and per-frame characteristics (e.g. velocity, distance to other flies, orientation) of

each fly within an arena. Each species was tracked separately, with species-specific tracking parameters

(e.g. background and foreground thresholds) to account for differences in body size. Videos were

inspected manually after tracking to check the accuracy of tracking. Videos that did not track correctly

were retracked with video-specific parameters to ensure the highest quality video tracking.

The tracking data were transferred to JAABA (Kabra et al. 2013), where we used supervised

machine learning to train classifiers for female aggression. The two most overt contact behaviors used to

score female aggression are headbutts (attacks using the head) and fencing (striking the opponent with

legs; see Supplementary Video). Headbutting is the most commonly used measure of female aggression

in *D. melanogaster* studies (Bath et al. 2017; Chiu et al. 2021; Schretter et al. 2020), so we focus our

results section on this behavior, with fencing results reported in the Supplementary Information. Both

headbutting and fencing have been observed in females of multiple species (Belenioti and Chaniotakis

2020; Shultzaberger et al. 2019), suggesting consistent behavioral repertoires across species. The

existence of a common behavioral repertoire allows us to use the same machine learning classifiers for

all species in our experiments.

For all species, we used an existing lab classifier for headbutts trained on *Drosophila*

melanogaster (Bath et al. 2020). We trained one new classifier for fencing and one new locomotion

classifier, defined as any time a female moved around the arena. A new species-specific classifier was

trained for the *D. nebulosa* wing lifting display, a behavior unique to this species (Supplementary Video).

All classifiers were ground-truthed by manually scoring a subset of videos with the scorer blind to

the automated classifier score. Ground-truthing was conducted on each species to test for differences in

accuracy among species. We used 30 randomly chosen intervals for each behavior in each species,

using the 'balanced random' option in JAABA, which ensures that some of the intervals contain the

behavior. We selected bout lengths (the number of frames an instance of that behavior was scored) the

same length as the average bout length for each species, because bout lenghts differed among species

(table S2). As is often the case when scoring behaviors, some instances of behaviors were clear

('Certain'), while others were more difficult to classify ('Difficult'). When manually scoring during ground-

truthing, we therefore scored the frames as either containing the behavior ('Behavior') or not ('None'), and

within each of those two groups, whether the behavior was easy to distinguish ('Certain') or difficult to

score ('Difficult'). This resulted in four possible outcomes for a frame: 'Behavior Certain', 'Behavior

Difficult', 'None Certain', 'None Difficult'. We then separately analyzed the frames where the behavior was

clear from those where it was difficult, because we expected differences in the performance of the

classifier between 'Certain' and 'Difficult' cases.

To quantify behaviors, we obtained the number of frames in which JAABA scored that a focal

individual in a dyad displayed that behavior. We divided this number of frames by the total number of

frames in which the individual was tracked to produce the proportion of frames an individual engaged in

the behavior. Using the proportion of frames allowed us to account for slight differences in frame numbers

among videos or individuals, due to recording or tracking discrepancies. We calculated values for each

individual in a dyad, but for some analyses (see below), we calculated a combined score for the dyad

based on both individual's tracking data.

For all behavioral classifiers, we used the proportion of frames scored for a behavior as our

response variable (e.g. proportion of frames scored as headbutting). The proportion of frames in which

one individual displayed aggression (headbutting and fencing) was positively correlated with the

proportion of frames her opponent displayed aggression (fig. S1 & S2), thus counting all frames with

aggression leads to pseudoreplication. The exception was *D. sturtevanti,* which did not have a positive

correlation in the behavior of both females. Nevertheless, to avoid pseudoreplication in all species, we

summed the number of frames for both individuals in a dyad to calculate the total number of frames in

which the behavior was observed in the arena to produce one value per dyad. We then divided this

number by the number of frames in the video to produce the total proportion of frames in which the

behavior happened for the pair. We then used this composite value for each pair, resulting in only one

value per dyad. For locomotion behavior, how much one individual moved was correlated with how much

its opponent moved (fig. S3). However, both individuals could be moving at the same time so combining

the number of frames in which each individual moved would have produced an overestimate of

locomotion within a pair. We therefore randomly chose one individual from each dyad and ran the

analysis on those individuals to avoid pseudoreplication.

Phylogenetic tree and species data

We built a phylogeny for our eight *Drosophila* species using the Open Tree of Life data store

(McTavish et al. 2015) and custom synthesis tools (Redelings and Holder 2017), based on phylogenies

from multiple sources (Finet et al. 2021; Oliveira et al. 2013; Sarikaya et al. 2019; Suvorov et al. 2022).

Branch lengths were estimated from date using Chronosynth (McTavish et al. 2023,

https://github.com/OpenTreeOfLife/chronosynth). Once a tree containing 200 species had been built for

another research project, we trimmed this tree to contain only our eight species (fig. S4). The eight

species were chosen as they represented a range of mating system characteristics (lifespan, remating

rate, sperm length, ovariole number; table S1). We attempted to account for phylogeny by pairing

species, when possible, in the same species group (i.e. *D. ananassae, D. bipectinata* in the melanogaster

group, *D. willistoni, D. nebulosa* in the willistoni group, *D. saltans, D. sturtevanti* in the saltans group).

In addition to the phylogeny, we collected values on ovariole number, sperm length, and remating

rate from the literature (table S1). For several of our species, we did not have data for ovariole number or

remating rate. We collected remating data for *D. saltans, D. sturtevanti,* and *D. willistoni*. We attempted to

collect these data for *D. nebulosa* but could not get a large enough sample size. To measure remating

rate, we mated females as described above. Once those pairs had finished mating, we removed the male

and left the female in the vial alone overnight. Twenty-four hours later, we placed two new unmated males

(which had not been used the previous day) in the vial with the female. Flies were observed for two hours

to determine if remating occurred (following Denis et al. 2017; Holman et al. 2008). Our sample sizes

were 29 *D. saltans* vials (4 remated)*,* 37 *D. sturtevanti* (37 remated)*,* and 48 *D. willistoni* vials (1 remated).

We then calculated the percentage of females that remated for use in our analysis of remating rate.

We counted the number of ovarioles in species for which we could not find ovariole number in the

literature. We followed the procedures outlined in Lobell et al. (2017). Briefly, unmated females were

collected and housed individually. When sexually mature, a female was placed on food with yeast paste

overnight before freezing at -20˚C. Within a month, females were dissected using fine needles in a drop

of PBS and stained in a solution of PBS saturated with crystal violet (Thermo Scientific). Ovarioles were

counted from both ovaries. Strains measured are given in table S1. At least 20 females were dissected

from each species.

Data analysis

All analyses were run in R v4.2.1 (R Core Team 2017). To analyze our behavioral data, we used

generalized linear models (GLMs) from the *lme4* package (Bates et al. 2015). Model fit and assumptions

were checked by visual examination of diagnostic plots. We acquired p-values using the 'Anova' function

from the *car* package (Fox and Weisberg 2018) To test if classifier accuracy was correlated with the

frequency of the behavior, we conducted a linear model with classifier accuracy (% of instances correct)

as the response variable and the proportion of frames in which the behavior was observed as the

explanatory variable. To analyze the effects of species and mating status, we used the total proportion of

frames in which flies performed the behavior as the response variable, with species, mating status, and

their interaction as explanatory variables. All variables and their interaction were included in all models as

these were key to the experimental design and answering the question at hand. We fitted these models

with a quasibinomial error distribution to account for underdispersion in our proportion data. We

performed this analysis for all species for headbutting, fencing, and locomotion (tables S3, S4).

To assess whether locomotion and aggression were correlated, we ran the following GLM with a

quasibinomial distribution:

*Proportion of frames aggressive ~ Proportion of frames moving * Species * Mating status*

As differences in aggression may be related to the proximity of flies within the arena, we also tested if the

average distance between females varied across species, as has been found in previous studies

(Shultzaberger et al. 2019). We used the minimum distance from the focal fly's head to the closest side of

the other fly that lies in front of the fly (-30°- 30° in front of the fly). We chose this measure as the distance

to an opponent in front of the fly is more relevant to aggression than absolute difference between flies as

flies can only headbutt flies in front of them (Nilsen et al. 2004); the measure is generated automatically

by the JAABA per-frame feature: 'dnose2ell_angle_min30to30'.

To test relationships between aggression and the reproductive characteristics, we performed

phylogenetic analyses using the mean values for each behavioral trait (i.e. headbutting and fencing) for

each species and mating treatment as the response variable and the mean value for the reproductive trait

(i.e. ovariole number, sperm length, remating rate) as the explanatory variable. We ran a phylogenetic

generalized least squares models with female median lifespan, average ovariole number, sperm length

and remating rate per species as explanatory variables in separate models (ape package: Paradis and

Schliep 2019; nlme package: Pinheiro et al. 2022). For the analyses with ovariole number and lifespan,

we included average body size as a covariate because ovariole number and lifespan scale with body size

across *Drosophila* species (Sarikaya et al. 2019; Syed et al. 2021). We log-transformed ovariole number,

body size, and sperm length before including them as explanatory variables in phylogenetic generalized

least squares models. As we ran four models for each of our response variables, we corrected for multiple

testing by applying a Bonferroni correction and reducing the threshold for significance (dividing our

significance threshold by the number of models: 0.05/4 = 0.0125). We then judged whether terms were

significant based on this new threshold.

***Drosophila nebulosa* specific analysis**

We observed female *D. nebulosa* performing a wing lifting movement that was first described with

courtship (Spieth 1947). In this movement, wings are lifted, spread, and dropped. Both *D. saltans* and *D.*

sturtevanti also performed a wing spreading behavior, but the movement was subtle and difficult to score.

The *D. nebulosa* movement is speculated to be a signaling behavior (Spieth 1947), thus we tested if its

performance was consistent with being an aggressive signal by describing when the behavior is

performed. We tested if the proportion of frames an individual spent performing the wing movement

varied with mating status. Because the proportion of frames one individual spent doing the wing behavior

was correlated with how much time their opponent spent performing the behavior, we randomly selected

one individual from each pair for the analysis to test if the wing movement varied with mating status. To

investigate if the wing behavior was a social behavior, we also tested if the distance between individuals,

and the orientation of the individuals, differed between frames in which flies were performing the wing

behavior and the frames in which they did not.

385   **Results**

Accuracy of machine learning across species

We used machine learning classifiers (Bath et al. 2020) to automatically score headbutts, fencing,

and locomotion in eight species. We scored classifier accuracy by ground-truthing a subset of frames

within each species (fig. 2 and table S2). We evaluated frames in which the behavior was unequivocally

happening or clearly not happening as 'Certain' instances and frames where it was less clear if the

behavior was happening or not as 'Difficult' frames. We used these two categories to allow us to separate

clear instances from less clear instances where manual observers often struggle to define behaviors. For

the headbutting classifier, accuracy ranged from 67-100% in the 'Certain' frames and from 63-100% in the

'Difficult' frames. Fencing was more variable with 51-100% for 'Certain' frames and 44-100% accuracy in

the 'Difficult' frames. We included locomotion to investigate differences induced by mating, as increased

movement is a post-mating response in *D. melanogaster* (Isaac et al. 2010). Locomotion is a less

complicated behavior for the classifier to score because it focuses only on the behavior of one fly,

providing us with a non-aggressive behavior for assessing classifier accuracy. Locomotion was scored

with similar accuracy to headbutts and fencing with a range of 73-100% for the 'Certain' frames and 60-

100% for the 'Difficult' frames (Table S2). Overall, headbutting and locomotion were more accurately

classified than fencing, though this did vary between species (e.g. for *D. nebulosa*, all behaviors had

similarly high levels of classifier accuracy, but for *D. hydei* headbutting was more accurately classified

than fencing).

Figure 2: Machine learning classifier accuracy across eight species and three behaviors. Black
circles represent the percentage of frames in the ground-truthing dataset where both the machine
learning classifier and the manual observer scored the frame as containing that behavior. Grey triangles
represent frames where both the machine learning classifier and the manual observer scored the frame
as not containing that behavior. The lines connect points within a species for ease of visualization across
species. These represent the 'Certain' instances of the classifier—those that were judged to be
unequivocally containing the behavior by the manual observer. Results for frames that were less clear are
included in table S2. For Drosophila ananassae, no frames were scored as containing the 'Certain'
behavior by either the classifier or the manual observer, so no circle is included. Points higher on the y-
axis mean the classifier was more accurate than those lower. When the black circle is higher than the

gray triangle, false positives exceeded false negatives in that species. If the points are the other way
around, false negatives exceeded false positives.

We tested if the frequency of behaviors influenced classifier accuracy, on the assumption that

species that had more instances of behaviors may be more likely to have higher accuracy. We found that

classifier accuracy was not related to how frequently the behavior was observed for headbutting ($F_{1,5}$ =

0.3, P = 0.61) or locomotion ($F_{1,5}$ = 1.03, P = 0.35). For fencing, accuracy and frequency were slightly

negatively associated ($F_{1,5}$ = 6.76, P = 0.041): species that had more instances of fencing were less

accurate, driven primarily by low accuracy in *D. hydei*. Therefore, the differences between species and

behaviors in accuracy were not due to how often the behaviors were displayed.

Effects of species and mating status on aggression

Headbutting is a commonly measured in the analysis of female aggression (e.g. Bath et al. 2017;

Nilsen et al. 2004; Schretter et al. 2020). Results for fencing were broadly the same as for headbutting,

thus we present the higher intensity headbutting (Nilsen et al. 2004). Results for fencing are presented in

the online supporting information. We found a significant effect of species on the proportion of frames in

which pairs were headbutting ($\chi^2_{7,538}$ = 49.76, P < 0.001; fig. 3). Mating status did not have a significant

effect ($\chi^2_{1,538}$ = 0.35, P = 0.55), nor was there an interaction between species and mating status ($\chi^2_{7,538}$ =

6.82, P = 0.45). *Drosophila pseudoobscura* was the most aggressive species with respect to headbutting

(proportion of frames: 0.018 ± 0.0017) and was significantly more aggressive than all species except *D.*

saltans (post-hoc Tukey tests, table S3).

Figure 3: Total proportion of frames labeled with headbutting by species and mating status. Each small point represents one dyad of females, with black circles representing unmated pairs (U) and grey triangles representing mated pairs (M). Large points represent model means with error bars representing 95% confidence interval (asymmetric due to proportion data). The phylogenetic relationships among the species are shown underneath the figure (see fig. S4 for branch lengths).

Although we found no evidence for an interaction between species and mating status, visual inspection o the data revealed trends in how species responded to mating. We performed pairwise comparisons within each species between mated and unmated females. Unmated females were significantly more aggressive than mated females in *D. bipectinata* (Odds Ratio Unmated/Mated = 2.46, z = 2.409, *P* = 0.016; fig. 3) and *D. pseudoobscura* (OR = 1.84, z = 3.065, *P* = 0.002). Trending in the same direction were *D. hydei* (OR = 1.55, z = 1.672, *P* = 0.0945), *D. nebulosa* (OR = 1.96, z = 1.943, *p* = 0.052), and *D. saltans* (OR = 1.68, z = 1.898, *P* = 0.058).

To investigate the effect of phylogeny on headbutting, we ran a phylogenetic generalized least squares model with a phylogeny containing our eight species for mated and unmated headbutts separately. We found that both mated and unmated headbutting had a significant phylogenetic signal with λ = 1.23 and λ = 1.07 respectively. Closely related species were similar in their levels of headbutting for

both mated and unmated females. However, the relative change in aggression after mating did not have a

significant phylogenetic signal (λ = - 0.24).

Locomotion and distance between individuals

Species and mating status had a significant interaction with respect to the proportion of frames in

which individual flies were moving ($\chi^2_{7, 538}$ = 34.569, P < 0.0001; fig. S5). Unmated females moved

significantly more than mated females in *D. bipectinata* (OR V/M = 5.64, z = 3.804, P = 0.0001), but

mated females moved more than unmated female *D. sturtevanti* (OR = 0.42, z = -3.071, P = 0.0021). No

other species exhibited a significant difference in locomotion with respect to mating status (fig. S5). We

found no significant effect of locomotion on headbutts, nor were there significant interactions between

locomotion, mating status or species (fig. S6). Observed differences in aggression, therefore, are not

attributable to differences in how much flies move.

We tested if the proximity of flies within the arena differed across species. We found a significant

effect of species in how close an opponent was in the area in front of the focal fly ($\chi^2_{1, 22}$ = 10.83, P =

0.0005, fig. S7). Female *D. hydei* were significantly further apart than *D. saltans* and *D. sturtevanti* when

not headbutting (Tukey post-hoc comparisons: *D. hydei vs. D. saltans*: z = 8.44, P < 0.001; *D. hydei vs.*

D. sturtevanti: z = 7.58, P < 0.001), but no other pairs of species were significantly different.

Ovariole number and aggression

Ovariole number and body size were not correlated with each other in our sample of species ($F_{1,6}$

= 3.24, p = 0.12). The relationship between ovariole number and the proportion of frames with

headbutting was positive– i.e. the more ovarioles a species had, the more time they spent headbutting

(Unmated females: $\chi^2_{1, 5}$ = 40.2, P < 0.0001, fig. 4A; Mated females: $\chi^2_{1, 5}$ = 14.43, P = 0.0001; λ= 1.26,

fig. 4B). Unmated females had a negative relationship between body size and headbutting—larger

species spent less time headbutting ($\chi^2_{1, 5}$ = 8.51, P = 0.004; λ= 0.82). This relationship was not seen in

mated females ($\chi^2_{1, 5}$ = 0.37, P = 0.54). Ovariole number did not predict the difference in aggression

between unmated and mated flies ($\chi^2_{1, 5}$ = 2.6, P = 0.11; λ= -0.68), whereas mating-induced changes in

aggression were linked to body size: species with small body sizes experienced a larger difference in

headbutting between mated and unmated females ($\chi^2_{1,5}$ = 5.88, *P* = 0.015).

Figure 4: Relationship between headbutts and fecundity traits. Relationship between headbutts and ovariole number in A) unmated females B) mated females and relationship between headbutts and sperm length in C) unmated females D) mated females. Dots (triangles in A and C and circles in B and D) represent the species mean proportion of frames in which headbutting was recorded. Confidence intervals represent 95% confidence intervals calculated in the phylogenetic analysis including body size as a covariate. The black line represents the model predictions. Species in the order from least ovariole number to highest are *Drosophila willistoni*, *D. ananassae*, *D. bipectinata*, *D. nebulosa*, *D. sturtevanti*, *D. saltans*, *D. pseudoobscura*, *D. hydei*. For sperm length, the order is: *Drosophila pseudoobscura*, *D. bipectinata*, *D. ananassae*, *D. nebulosa*, *D. willistoni*, *D. saltans*, *D. sturtevanti*, *D. hydei*.

Sperm length and aggression

Sperm length was not correlated with unmated female headbutting ($\chi^2_{1,6}$ = 3.07, *P* = 0.08; λ= 1.17; Fig. 4C), whereas species with larger sperm tended to have less headbutting in mated females ($\chi^2_{1,6}$ = 20.86, *P* < 0.0001; λ= 1.26; fig. 4D). The relative change in headbutting behavior between mated and unmated females was not associated with sperm length across species (sperm length: $\chi^2_{1,6}$ = 0.47, *P* = 0.49; λ= -0.69).

Lifespan

Across species, body size and lifespan were not correlated ($\chi^2_{1,5}$ = 0.12, *P* = 0.73; λ= -0.57), but ovariole number and lifespan had a significant positive correlation ($\chi^2_{1,5}$ = 6.88, *P* = 0.009; λ= -0.57),

which was the opposite to the life-history prediction that there should be a negative relationship between

lifespan and ovariole number (fig. 1D). There was no association between lifespan and mated or unmated

headbutting (mated: $\chi^2_{1,5} = 0.12$, $P = 0.73$; $\lambda = 1.26$; unmated: $\chi^2_{1,5} = 3.46$, $P = 0.06$; $\lambda = 1.26$). Lifespan

and change in headbutting with mating were not associated ($\chi^2_{1,5} = 1.66$, $P = 0.2$; $\lambda = -0.33$).

Species-specific aggressive behavior: *Drosophila nebulosa*

The only species in which we observed a potential alternative aggressive behavior was *D.*

nebulosa. Females periodically performed wing flicks in which they lifted their wings (Supplementary

Video), which might be a signal of size or aggressive intent. To test if wing movements happened in a

social context, we identified which per-frame features were significantly different between frames in which

females were performing the movement and those where they were not. We found no significant

difference in distance to the opponent between frames in which a female was performing and not

performing the wing movement ($t_7 = 2.08$, $P = 0.076$). However, females were more likely to be facing

their opponent when performing the movement than when they were not ($t_7 = 3.2$, $P = 0.015$).

Mating status and the number of wing flicks that the opponent performed had an interaction effect

on the number of wing flicks the focal individual performed ($\chi^2_{1, 62} = 4.4$, $P = 0.036$). For unmated females,

the number of wing flicks was positively correlated within a pair ($t_{1,62} = 5.72$, $P < 0.0001$), but not in mated

females ($t_{1,62} = 0.84$, $P = 0.41$; fig. S8). As a result of this correlation, we chose one individual randomly

from each pair to test if mating influenced how often females performed the wing behavior. Unmated

females performed the wing behavior more than mated females ($\chi^2_{1, 62} = 4.76$, $P = 0.03$; fig. S8).

Discussion

When evaluating our results with respect to the predictions from resource valuation, life-history,

and hormonal correlation (Table 1), life-history is the least supported hypothesis. Aggression in the

species studied is not correlated with lifespan, though theory predicts a negative relationship. In addition,

with the life-history hypothesis, a negative relationship is predicted for the relationship between ovariole

number and lifespan, but we found a positive relationship in our species. Distinguishing between the

resource evaluation hypothesis and the hormonal correlation hypothesis is not possible because they

have the same predictions (table 1) but, for the most part, our results support these hypotheses.

Table 1: Predictions from each hypothesis and the results from this study

Relationship	Hypothesis			Results from this study	
	Resource valuation	Life-history	Hormonal correlation		
Within species predictions					
Aggression and mating	Mated > Unmated	Mated > Unmated	Mated > Unmated	No overall effect; in no species were Mated > Unmated *D. bipectinata* and *D. pseudoobscura*: Unmated > Mated	
Across species predictions				Unmated	Mated
Aggression and ovariole number	**Positive**	**Positive**	**Positive**	**Positive**	**Positive**
Change in aggression after mating and ovariole number	Positive (species with many ovarioles show large response to mating)	Positive	Positive	-	None
Aggression and lifespan	NA	Negative	NA	None	None
Aggression and sperm length	**Negative (mated females)**	NA	**Negative (mated females)**	None	**Negative**
Change in aggression after mating and sperm length	Negative (species with long sperm show small responses to mating)	NA	Negative	-	None
Ovariole number and lifespan	NA	Negative	NA	Positive	Positive

Notes: For the results, only statistically significant relationships are given, or "none" is indicated. Bold text indicates that the hypothesis is supported by the results. NA= not applicable (no prediction).

Within species predictions

All three hypotheses predicted that mated females are more aggressive than unmated females within species. Unlike *D. melanogaster*, in many of these species unmated and mated females had similar levels of headbutting. The lack of change after mating was surprising because females of all species are expected to experience an increased demand for protein and other nutrients essential for egg production after mating. However, we do not know if females of other species experience the same post-mating shifts in food preference and feeding behavior as *Drosophila melanogaster* (Barnes et al. 2008; Ribeiro and Dickson 2010). Aggression may not change if nutrient requirements do not change substantially. We know little about nutritional needs across species, or if investment in eggs changes after mating. For instance, *D. melanogaster* females can produce mature eggs on a sucrose-only diet, while *D. hydei* females require protein to synthesize essential proteins and develop eggs to maturity (Lamnissou 2000). Some species may therefore require yeast as a high value resource, increasing motivation to compete over yeast, whereas other species may not. Further research investigating the nutritional requirements and patterns of egg production across species are needed to understand changes in female aggression, as well as the patterns of female post-mating responses.

The higher level of unmated females aggression relative to that of mated females in two species

(*D. bipectinata* and *D. pseudoobscura*) is contrary to previous work on *D. melanogaster* (Bath et al.

2017)*.* Females in these species possibly acquire food and/or oviposition sites before mating, resulting in

a peak in aggression before they mate. Unmated females in these species may therefore value food

before mating to supply future egg production, rather than having a shift stimulated by mating.

Alternatively, aggression may be suppressed by mating through seminal fluid proteins or changes in

hormones regulating behaviors, such as juvenile hormone (Scott 2006; Tibbetts et al. 2013). Regardless,

these results illustrate that the behavior and physiology of *Drosophila* species is not always the same as

D. melanogaster. These results raise questions about why *D. melanogaster* shows an increase in

aggression after mating when none of the other species studied do.

Across species predictions – ovariole number

Ovariole number determines the number of eggs that can be produced in a day and,

consequently, lifetime fecundity (Sarikaya et al. 2019). All three of our hypotheses predicted a positive

correlation between ovariole number and aggression (table 1), but with different underlying reasons. The

resource valuation hypothesis is based on egg production requiring a large quantity of protein (Wheeler

1996) suggesting that females who have more ovarioles (and therefore produce more eggs) are likely to

require more protein than females producing fewer eggs over the same time. The increased need for

protein may drive increased aggression, helping to explain the pattern we see across species (the

resource valuation hypothesis). However, this pattern may also be explained by differential changes in

hormones and physiology across species that are associated with differences in ovariole number

(hormonal correlation). Alternatively, life-history theory suggests that faster-reproducing species (those

with more ovarioles) prioritize current reproduction over future reproduction, thus they are aggressive to

increase their current brood's chance of success. We found a positive correlation between ovariole

number and aggression, but are currently not able to distinguish the underlying cause of this relationship

in this experiment.

We also predicted that ovariole number is positively correlated with changes in aggression after

mating—i.e. that species with more ovarioles show a greater change in aggression after mating than

those with fewer ovarioles. However, we found no evidence of this in these species, which is likely driven

by the few species in which aggression changed in response to mating. Because these species differ in

ovariole number but do not change aggression in relation to mating may indicate that aggression is not

driven by energetic requirements of egg production. Alternatively, females across species may not differ

substantially in their valuation of this particular food resource despite laying very different numbers of

eggs.

Drosophila species vary in their ecology, which is likely to generate different selection pressures

on female aggression with respect to different resources (Clutton-Brock and Huchard 2013b). However,

we know little about the behavior and selective pressures in nature for any of the species we studied. All

the species in this study, with the possible exception of *D. saltans* (Valadão et al. 2019), are generalists

that eat and lay eggs on rotting fruit (Nunney 1996; Valadão et al. 2019). Generalist species may

experience less competition for food or oviposition sites than specialists because of their ability to use

multiple hosts. The Drosophilidae family consists of cosmopolitan generalists (e.g. *D. melanogaster*), as

well as specialized species (e.g. *Scaptomyza chauliodon* and *Scaptomyza magnipalpa*; Sarikaya et al.

2019), making the group an excellent study system to explore how ecology affects female aggression

over food and oviposition sites.

Across species predictions – sperm length

Mated female aggression and sperm length were negatively correlated, which was predicted by

both the resource valuation and hormonal correlation hypotheses. The resource valuation hypothesis

argues that some species may compete over access to mates, particularly if mates are potentially limiting,

while the hormonal correlation hypothesis suggests that the strength of female post-mating responses will

be related to the amount of sperm female receive in each mating. Female *D. pseudoobscura* receive up

to 25,000 sperm per mating and can continue to lay fertilized eggs up to 2 weeks after a single mating

(Markow et al. 2012; Snook et al. 1994). In contrast, female *D. hydei* receive around 120 sperm per

mating, remate frequently, and appear to become sperm depleted after 1-2 days in the absence of males

(Pitnick and Markow 1994). Wild caught female *D. hydei* are less likely to be fertilized than are *D.*

melanogaster and *D. pseudoobscura* (Markow et al. 2012). Thus, for female *D. hydei*, sperm is potentially

a limiting resource. All the species in this study, except *D. pseudoobscura,* transfer few sperm during

mating, suggesting sperm may be a limiting factor for these species. Nonetheless, females of some

species fought over food, so females may compete over multiple resources. Future research using a

range of contested resources in experiments is likely to yield different results. For example, unmated *D.*

melanogaster females are more aggressive than mated females towards mating pairs (Gaspar et al.

2022) implying access to mates is driving aggression in that context.

Species with long sperm generally transfer few sperm to females in each mating (Markow 1996;

Syed et al. 2021). Receiving few sperm may explain the negative relationship between sperm length and

mated female headbutting. In *D. melanogaster*, the transfer of sperm induces more fighting than does

mating alone, and the strength of this response may depend on the quantity and/or quality of sperm

received (Bath et al. 2021a; Bath et al. 2021b). If sperm induction of a female's response is dose

dependent, then females receiving few sperm will have less altered aggressive behavior (either up- or

down-regulated) than those who receive more sperm. In contrast to our headbutting results, we found a

significant effect of sperm length on changes in fencing after mating (Supplementary Information):

females of species with longer sperm fenced more after mating than before. Headbutting and fencing may

be regulated separately and serve different functions in aggressive interactions. If fencing is less

energetically costly than headbutting, then fencing may be a low-cost form of aggression favored by

mated females in species that need to expend energy in finding new mates to maximize reproductive

output (i.e. females receiving large sperm).

Male ejaculate components transferred during mating can trigger changes in female behavior and

physiology. Seminal fluid proteins (Sfps) induce strong post-mating responses in *D. melanogaster*

females, altering their receptivity to mating, egg production, sleeping and locomotion patterns, feeding

preferences and behavior, and lifespan (reviewed in Avila et al. 2011; Kubli and Bopp 2012). One Sfp, sex

peptide, is involved in the mating-induced increase in aggression in *D. melanogaster* (Bath et al. 2017).

Differences in aggressive responses to mating among species may indicate different effects of Sfps on

female aggression, and potentially female behavior more broadly. We found contrasting effects of mating

on locomotion in our species: *D. bipectinata* increased movement after mating, while *D. sturtevanti*

reduced movement after mating, and other species showed no change in movement due to mating. While

the functional categories of Sfps are well-conserved across *Drosophila* taxa, the genes encoding Sfps

turnover and diverge rapidly (Hurtado et al. 2022; Sirot 2019; Wigby et al. 2020). Putative Sfps have been

identified in several species in our dataset (e.g. *D. ananassae*, Findlay et al. 2014; *D. pseudoobscura*,

Karr et al. 2019 ; *D. bipectinata*, Santhosh and Krishna 2013), but few have been functionally validated or

empirically tested for variation in female post-mating responses due to a lack of tools in non-model

organisms. However, recent developments in gene editing technologies may allow the targeting of

specific seminal fluid proteins to determine function and effects on female post-mating responses (Denis

et al. 2017).

Across species predictions - lifespan

Species appear along a continuum of fast to slow life-history traits, where 'fast' species have high

growth and reproductive rates, accompanied by shorter lifespans than 'slow' species (Stearns 1989). In

our species, we predicted a negative correlation between lifespan and ovariole number such that species

with high ovariole numbers invest more in reproduction and have short lives. However, we found a

positive correlation between ovariole number and lifespan, suggesting species with more ovarioles live

longer. Why this is the case is unclear. In *Drosophila* species, sperm length is positively correlated with

the number of days post-eclosion for males to reach sexual maturity, which is likely due to the amount of

time it takes for larger sperm to develop (Pitnick et al. 1995; Syed et al. 2021). However, for the majority

of species, sperm length was not related to male lifespan (for an exception, see Syed et al. 2021). Adult

lifespan and maturation time are unlikely to be determined by the development of ovarioles, as ovaries

develop during the larval and pupal stages and ovariole number is set upon eclosion (Büning 1994).

Within species, females with larger bodies tend to have more ovarioles (Honek 1993), but the pattern

does not hold across insect species (Church et al. 2021; Sarikaya et al. 2019). Large species might have

both long lifespans and many ovarioles, but we found no evidence for this in our dataset. Why species

with more ovarioles live longer than species with fewer ovarioles is not clear, but testing more species will

enable us to see if this is a broad pattern among *Drosophila*.

Based on the assumption of a trade-off between reproduction and lifespan, theory predicts a

negative relationship between lifespan and aggression, as short-lived species can receive the benefits of

aggression but die before facing the costs (Eckerström-Liedholm et al. 2016). However, we did not find a

relationship between aggression and lifespan. Either the relationship may not exist without a pre-existing

trade-off between reproduction and lifespan or all our species are reasonably short-lived, though there

654 was a significant amount of variation within our species (female median lifespan varies from 28-59 day;

655 table S1). The lack of a trade-off between reproduction and lifespan in our species, as well as the lack of

656 relationship between lifespan and aggression, suggests that differences in life-histories do not explain the

657 observed relationship between reproduction and aggression.

658 **Evolutionary History**

659 Both headbutting and fencing (Supporting Information) had a high phylogenetic signal, suggesting

660 aggression is conserved among closely related species. Behavioral traits are more evolutionarily labile

661 than morphological or physiological traits (Blomberg et al. 2003), although numerous behavioral traits

662 have significant phylogenetic signal (reviewed in Cicero et al. 2020). Our study consisted of only eight

663 species, which reduces our ability to detect phylogenetic signal, but some of our species were paired

664 within species groups (i.e. *D. ananassae* and *D. bipectinata*, *D. willistoni* and *D. nebulosa*, *D. saltans* and

665 *D. sturtevanti*), which may artificially inflate our measure of phylogenetic signal.

666 Aggression is a very plastic trait, responding to the social and asocial environment, internal

667 condition, and resource availability and distribution (Clutton-Brock and Huchard 2013b; Stockley and

668 Campbell 2013). In *D. melanogaster*, for instance, females double the amount of time they spend fighting

669 after mating (Bath et al. 2017), while unmated *D. bipectinata* and *D. pseudoobscura* in the current

670 experiment were around twice as aggressive as mated females (fig. 3). Given how plastic aggression is

671 within species in response to mating and environmental factors, phylogenetic conservation of aggression

672 in both mated and unmated females seems surprising. Closely related species may inhabit similar niches

673 or have similar mating systems and morphologies, leading to similar selection pressures shaping

674 aggression. For example, *D. ananassae* and *D. bipectinata,* which belong to the same subgroup, have

675 relatively small sperm and low remating rates, which may shape their response to mating in a similar way

676 (Syed et al. 2021). Further studies expanding the range of species in which we study both male and

677 female aggression will give us more insight into the selection pressures and constraints shaping

678 aggression.

Machine learning for comparative behavior

We used machine learning classifiers originally developed for *D. melanogaster* across eight different species. To our knowledge, this is the first time machine learning has been used for a comparative analysis of social behavior in arthropods (for an example of non-social behavior, see Hernández et al. 2021). The classifier trained to identify headbutts in female *D. melanogaster* scored headbutts for all eight species, with varying levels of accuracy. Our range of accuracies was similar to other classifiers used only for *Drosophila melanogaster* (Bath et al. 2020; Chiu et al. 2023; Robie et al. 2017), with some of our accuracies exceeding that of same species (*D. melanogaster*) classifiers. For our species, classifiers were less accurate in some species, suggesting potential caution and ground-truthing must be used in interpreting data from the same classifier on multiple species.

For most species, in clear instances of headbutting, the classifier was more likely to miss instances of aggression than falsely categorize frames with no headbutting (table S2). Our scores of aggression are, therefore, likely underestimates rather than overestimates of female aggression. For frames in which the behavior was difficult to categorize, the classifier had an even distribution of false negatives and false positives. Classifier accuracy was dependent on tracking quality; however, we ensured high-quality tracking for all species (see Materials and Methods), thus differences in tracking accuracy among species are unlikely to be responsible for differences in classifier accuracy. Differences in classifier accuracy may be due to species-specific differences in headbutting and fencing behaviors. Slight differences in speed, direction, and duration of flies' movements during aggression may alter the classifier's ability to score behaviors (Bath et al. 2020). Detailed studies are needed for an in-depth analysis of qualitative differences among species in these shared behaviors.

Behaviors differed in the accuracy of their relevant classifier, which is consistent with other uses of other classifiers (Robie et al. 2017). The headbutting classifier was the most accurate, while the classifiers for fencing and locomotion were similar to each other in accuracy. We expected the headbutting classifier to have higher accuracy than the fencing classifier because headbutting is easier to score manually. We expected the locomotion classifier to be more accurate than either of the aggressive behaviors because locomotion is easiest behavior to score manually (i.e. is the fly moving or not). The

similar level of accuracy between the locomotion and fencing classifiers implies that species may have

qualitative (different movement styles) and quantitative (duration of movement) differences in locomotion.

Using the same classifiers on all species allowed us to quickly calculate aggression scores and

directly compare species in shared experimental conditions. The ability to use one algorithm for multiple

species (with caveats) greatly increases the number of individuals and species that can be scored for

complex and subtle behaviors. We therefore demonstrate the potential use of machine learning for

comparative behavioral analysis, though caution that a human still needs to do ground truthing as well as

determine if additional behaviors exist that are not shared across species. Comparative behavioral

analysis using machine learning can help us to understand the evolution of complex social behaviors,

including elucidating ecological and genetic factors that shape behaviors across species (Hernández et

al. 2021; Moulin et al. 2021).

Wing-flicking in *D. nebulosa*

Although our study focused on aggressive behaviors shared by species, we observed a

distinctive species-specific behavior that may function in aggressive encounters. A female *D. nebulosa*

was more likely to be facing her opponent when performing the wing flicking behavior than when she was

not performing the behavior, suggesting communication. A female often wing-flicked before approaching

the other fly and engaging in headbutting or fencing. The wing behavior may provide a signaling function,

perhaps indicating size or motivation to engage in aggression. If large females are more likely to win

encounters, wing size may be a signal of resource holding potential and a female's likelihood of winning a

subsequent aggressive encounter. Selection favors processes that allow individuals to evaluate the costs

and benefits of aggression, leading to the evolution of signals that demonstrate resource holding potential

to reduce the costs of engaging in aggression (Parker 1974). Large flies are more likely to win aggressive

encounters for both male and female *D. melanogaster* (Bath et al. 2018; Hoffmann 1987), but how

aggressive encounters are decided is unstudied in *D. nebulosa* and most other Drosophilids. However,

both male and female *D. nebulosa* perform this behavior in mixed-sex groups, as well as in male-female

pairs (Gleason et al. 2012; Spieth 1947) so it is possible it serves an alternative signaling function, or no

function at all. Further work on the sequence of aggressive behaviors, assessment rules, and what

determines winners in a variety of species, will help to elucidate the behaviors involved in aggression

across species.

Conclusion

Species vary in female aggression level in the presence of yeast. The effect of mating in

changing female behavior is not universal among *Drosophila* species and therefore additional study is

needed in species other than *D. melanogaster*. The results indicate that resource valuation and hormonal

correlation are the most likely drivers of female aggression, but more work needs to be done to determine

which has a stronger effect. Studies are needed to understand how female behavior varies and if variation

is dependent on ecology, mating system, phylogenetic history, or other aspects of female biology.

742   **References**

Ah-King, M. 2022. The history of sexual selection research provides insights as to why females are still
understudied. Nature Communications 13:6976.
Amino, K., and M. Takashi. 2020. Intra-versus inter-sexual selection on sexually dimorphic triaits in
Drosophila prolongata. Zoological Science 37:210-216.
Arnott, G., and R. W. Elwood. 2008. Information gathering and decision making about resource value in
animal contests. Animal Behaviour 76:529-542.
Avila, F. W., L. K. Sirot, B. A. Laflamme, C. D. Rubinstein, and M. F. Wolfner. 2011. Insect seminal fluid
proteins: identification and function. Annual Review of Entomology 56:21-40.
Barnes, A. I., S. Wigby, J. M. Boone, L. Partridge, and T. Chapman. 2008. Feeding, fecundity and lifespan
in female *Drosophila melanogaster*. Proceedings of the Royal Society B: Biological Science
275:1675-1683.
Bates, D., M. Mächler, B. M. Bolker, and S. C. Walker. 2015. Fitting linear mixed-effects models using
lme4. Journal of Statistical Software 67:1-48.
Bath, E., E. R. Biscocho, A. Easton-Calabria, and S. Wigby. 2020. Temporal and genetic variation in
female aggression after mating. PLoS ONE 15:e0229633.
Bath, E., S. Bowden, C. Peters, A. Reddy, J. A. Tobias, E. Easton-Calabra, N. Seddon et al. 2017. Sperm
and sex peptide stimulate aggression in female *Drosophila*. Nature Ecology and Evolution
1:0154.
Bath, E., D. Buzzoni, T. Ralph, S. Wigby, and I. Sepil. 2021a. Male condition influences female post
mating aggression and feeding in *Drosophila*. Functional Ecology 35:1288-1298.
Bath, E., D. Edmunds, J. Norman, C. Atkins, L. Harper, W. G. Rostant, T. Chapman et al. 2021b. Sex ratio
and the evolution of aggression in fruit flies. Proceedings of the Royal Society B: Biological
Science 288:20203053.
Bath, E., J. Morimoto, and S. Wigby. 2018. The developmental environment modulates mating-induced
aggressoin and fighting success in adult female *Drosophila*. Functional Ecology 32:2542-2552.
Belenioti, M., and N. Chaniotakis. 2020. Aggressive behavior of *Drosophila suzukii* in relation to
environmental and social factors. Scientific Reports 10:7898.
Blomberg, S. P., J. Garland, T., and A. R. Ives. 2003. Testing for phylogenetic signal in comparative data:
behavioral traits are more labile. Evolution 57:717-745.
Boake, C. R. B., D. K. Price, and D. K. Andreadis. 1998. Inheritance of behavioral differences between
two interfertile, sympatric species, *Drosophila silvestri*s and *D. heteroneura*. Heredity 80:642-650.
Bro-Jørgensen, J. 2002. Overt female mate competition and preference for central males in a lekking
antelope. Proceedings of the National Academy of Sciences, USA 99:9290-9293.

Büning, J. 1994, The Insect Ovary: Ultrastructure, Previtellogenic Growth and Evolution, Springer.

Carvalho, G. B., P. Kapahi, D. J. Anderson, and S. Benzer. 2006. Allocrine modulation of feeding behavior
by the sex peptide of *Drosophila*. Current Biology 16:692-696.

Chen, J., and O. T. Lewis. 2023. Climate and competition combine to set elevational distributions of
tropical rainforest *Drosophila*, BioRxiv.

Chiu, H., E. D. Hoopfer, M. L. Coughlan, and D. J. Anderson. 2021. A circuit logic for sexually shared and
dimorphic aggressive behaviors in *Drosophila*. Cell 184:507-520.

Chiu, H., A. A. Robie, K. M. Branson, T. Vippa, S. Epstein, G. M. Rubin, D. J. Anderson, and C. E.
Schretter. 2023. Cell type-specific contributions to a persistent aggressive internal state in female
Drosophila. eLife.

Church, S. H., B. A. S. de Medeiros, S. Donoughe, N. L. Márquez Reyes, and C. G. Extavour. 2021.
Repeated loss of variation in insect ovary morphology highlights the role of development in life-
history evolution. Proceedings of the Royal Society B: Biological Science 288:20210150.

Cicero, C., N. Mason, L. Benedict, and J. D. Rising. 2020. Behavioral, morphological, and ecological trait
evolution in two clades of New World Sparrows (*Aimophila* and *Peucaea*, Passerellidae). Peer J
8:e9249.

Clutton-Brock, T., and E. Huchard. 2013a. Social competition and its consequences in female mammals.
Journal of Zoology 289:151-171.

Clutton-Brock, T. H., and E. Huchard. 2013b. Social competition and selection in males and females.
Philosophical Transactions of the Royal Society of London B 368:20130074.

Cunningham, C. B., M. K. Douthit, and A. J. Moore. 2014. Octopaminergic gene expression and flexible
social behaviour in the subsocial burying beetle *Nicrophorus vespilloides*. Insect Molecular
Biology 23:391-404.

Dammhahn, M., N. J. Dingemanse, P. T. Niemelä, and D. Réale. 2018. Pace-of-life syndromes: a
framework for the adaptive integration of behaviour, physiology and life history. Behavioral
Ecology and Sociobiology 72:62.

David, J. R. 1970. Le nombre d'ovarioles chez la Drosophile : relation avec la fécondité et valeur
adaptive. Archives de Zoologie Expérimentale et Générale 111:357-370.

DeLellis, P., G. Polverino, G. Ustuner, N. Abaid, S. Macrì, E. M. Bollt, and M. Porfiri. 2014. Collective
behaviour across animal species. Scientific Reports 4:3723.

Dell, A. I., J. A. Bender, K. Branson, I. D. Couzin, G. G. de Polavieja, L. P. J. J. Noldus, A. Pérez-Escudero
et al. 2014. Automated image-based tracking and its application in ecology. Trends in Ecology &
Evolution 29:417-428.

Denis, B., G. Claisse, A. Le Rouzic, C. Wicker-Thomas, G. Lepennitier, and D. Joly. 2017. Male accessory
gland proteins affect differentially female sexual receptivity and remating in closely related
Drosophila species. Journal of Insect Physiology 99:67-77.

Eckerström-Liedholm, S., W. Sowersby, S. Morozov, W. vander Bijl, P. K. Rowinski, A. Gonzalez-Voyer,
and B. Rogell. 2016. Macroevolutionary evidence suggests trait-dependent coevolution between
behavior and life-history. Evolution 73:2312-2323.

Edwards, A. C., S. M. Rollman, T. J. Morgan, and T. F. C. Mackay. 2006. Quantitative genomics of
aggressive behavior in *Drosophila melanogaster*. PLoS Genetics 2:e154.

Eyjolfsdottir, E., S. Branson, X. P. Burgos-Artizzu, E. D. Hoopfer, J. Schor, D. J. Anderson, and P. Perona.
2014. Detecting social actions of fruit flies, Pages 772-787 *in* D. Fleet, T. Pajdia, B. Schiele, and
T. Tuytelaars, eds. Computer Vision - EECV 2014. Lecture Notes in Computer Science, Springer,
Cham.

Findlay, G. D., J. L. Sitnik, W. Wang, C. F. Aquadro, N. L. Clark, and M. F. Wolfner. 2014. Evolutionary
rate covariation identifies new members of a protein network required for *Drosophila*
melanogaster female post-mating response. PloS ONE 10:e1004106.

Finet, C., V. A. Kassner, A. B. Carvalho, H. Chung, J. P. Day, S. Day, E. K. Delaney et al. 2021.
DrosoPhyla: Resources for Drosophilid phylogeny and systematics. Genome Biology and
Evolution 13:evab179.

Fox, J. H., and S. Weisberg. 2018, An R Companion to Applied REgression. Thousand Oaks, CA, Sage
Publications.

Gaspar, M., S. Dias, and M. L. Vasconcelos. 2022. Mating pair drives aggressive behavior in female
Drosophila. Current Biology 32:4734-4742.

Gittleman, J. L., and S. D. Thompson. 1988. Energy allocation in mammalian reproduction. American
Zoologist 28:863-875.
Gleason, J. M., E. C. Griffith, and J. R. Powell. 1998. A molecular phylogeny of the *Drosophila willistoni*
group: conflicts between species concepts? Evolution 52:1093-1103.
Gleason, J. M., A. A. Pierce, A. L. Vezeau, and S. F. Goodman. 2012. Different sensory modalities are
required for successful courtship in two species of the *Drosophila willistoni* group. Animal
Behaviour 83:217-227.
Gupta, T., S. E. Howe, M. L. Zorman, and B. L. Lockwood. 2019. Aggression and discrimination among
closely versus distantly related species of *Drosophila*. Royal Society Open Science 6:190069.
Gwynne, D. T. 1991. Sexual competition among females: What causes courtship-role reversal? Trends in
Ecology & Evolution 6:118-121.
Hernández, D. G., C. Rivera, J. Cande, B. Zhou, D. L. Stern, and G. J. Berman. 2021. A framework for
studying behavioral evolution by reconstructing ancestral repertoires. eLife 10:e61806.
Hoffmann, A. A. 1987. Territorial encounters beetween *Drosophila* males of different sizes. Animal
Behaviour 35:1899-1901.
Holman, L., R. P. Freckleton, and R. R. Snook. 2008. What use is an infertile sperm? A comparative
study of sperm-heteromorphic *Drosophila*. Evolution 62:374-385.
Honek, A. 1993. Intraspecific variation in body size and fecundity in insects: a general relationship. Oikos
66:483-492.
Huchard, E., and G. Cowlishaw. 2011. Female-female aggression around mating: an extra cost of
sociality in a multimale primate society. Behavioral Ecology 22.
Hurtado, J., F. C. Almeida, S. A. Belliard, S. Revale, and E. Hasson. 2022. Research gaps and new
insights in the evolution of Drosophila seminal fluid proteins. Insect Molecular Biology 31:139-
158.
Isaac, R. E., C. Li, A. E. Leedale, and A. D. Shirras. 2010. *Drosophila* male sex peptide inhibits siesta
sleep and promotes locomotor activity in the post-mated female. Proceedings of the Royal
Society B: Biological Science 277:65-70.
Jia, Y., S. Jin, K. Hu, L. Geng, C. Han, R. Kang, Y. Pang et al. 2021. Gut microbiome modulates
Drosophila aggression through octopamine signaling. Nature Communications 12:2698.
Kabra, M., A. A. Robie, M. Rivera-Alba, S. Branson, and K. Branson. 2013. JAABA: interactive machine
learning for automatic annotation of animal behavior. Nature Methods 10:64-67.
Karr, T. L., H. Southern, M. A. Rosenow, T. I. Gossmann, and R. R. Snook. 2019. The old and the new:
discovery proteomics identifies putative novel seminal fluid proteins in Drosophila. Molecular &
Cellular Proteomics 18:S23-S33.
Krishna, M. S., and S. N. Hegde. 1997. Reproductive success of large and small flies in *Drosophila*
bipectinata complex. Current Science 72:747-750.
Kubli, E., and D. Bopp. 2012. Sexual Behavior: How sex peptide flips the postmating switch of female
flies Current Biology 22:R520-R522.
Lamnissou, K. 2000. Nutritional effects on vitellogenesis in species of *Drosophila*. Journal of
Entomological Science 35:452+464.
Lee, K. P., J.-S. Kim, and K.-J. Min. 2013. Sexual dimorphism in nutrient intake and life span is mediated
by mating in *Drosophila melanogaster*. Animal Behaviour 86:987-992.
Lipshutz, S. E., and K. A. Rosvall. 2021. Nesting strategy shapes territorial aggression but not
testosterone: a comparative approach in female and male birds. Hormones and Behavior
133:104995.
Lobell, A. S., R. R. Kaspari, Y. L. S. Negron, and S. T. Harbison. 2017. The genetic architecture of ovariole
number in *Drosophila melanogaster*: genes with major, quantitative, and pleiotropic effects.
Genes, Genomes, Genetics 7:2391.
Lorenz, M. W., and G. Gäde. 2009. Hormonal regulation of energy metabolism in insects as a driving
force for performance. Integrative and Comparative Biology 49:380-392.
Lüpold, S., M. K. Manier, N. Puniamoorthy, C. Schoff, W. T. Starmer, S. H. B. Luepold, J. M. Belote, and
S. Pitnick. 2016. How sexual selection can drive the evolution of costly sperm ornamentation.
Nature 533:535-541.
Macartney, E. L., and R. Bonduriansky. 2021. Does female resistance to mating select for live-fast-die-
young strategies in males? A comparative analysis in the genus *Drosophila*. Journal of
Evolutionary Biology 35:192-200.

Markow, T. A. 1996. Evolution of *Drosophila* mating systems. Evolutionary Biology 29:73-106.

—. 2002. Perspective: Female remating, operational sex ratio, and the arena of sexual selection in
Drosophila species. Evolution 56:1725-1734.

Markow, T. A., S. Beall, and S. J. Castrezana. 2012. The wild side of life Drosophila reproduction in
nature. Fly 6:98-101.

Markow, T. A., and P. M. O'Grady. 2005a, Drosophila: A Guide to Species Identification and Use.
Burlington, MA, Academic Press.

—. 2005b. Evolutionary genetics of reproductive behavior in *Drosophila*: Connecting the dots. Annual
Review of Genetics 39:263-291.

—. 2008. Reproductive ecology of *Drosophila*. Functional Ecology 22:747-759.

McTavish, E. J., C. E. Hinchliff, J. F. Allman, J. W. Brown, K. A. Cranston, M. T. Holder, J. A. Rees, and S.
A. Smith. 2015. Phylesystem: a git-based data store for community-curated phylogenetic
estimates. Bioinformatics 31:2794-2800.

Moreteau, B., P. Gibert, G. Pétavy, J.-C. Moreteau, R. B. Huey, and J. R. David. 2003. Morphometrical
evolution in a Drosophila clade: the *Drosophila obscura* group. Journal of Zoological Systematics
and Evolutionary Research 41:64-71.

Moulin, T. C., L. E. Covill, P. M. Itskov, M. J. Williams, and H. B. Schiöth. 2021. Rodent and fly models in
behavioral neuroscience: An evaluation of methodological advances, comparative research, and
future perspectives. Neuroscience and Biobehavioral Reviews 120:1-12.

Nilsen, S. P., Y.-B. Chan, R. Huber, and E. A. Kravitz. 2004. Gender-selective patterns of aggressive
behavior in *Drosophila melanogater*. Proceedings of the National Academy of Sciences, USA
101:12343-12347.

Nord, A., and J. B. Williams. 2015. The energetic costs of incubation *in* D. C. Deeming, and S. J.
Reynolds, eds. Nests, Eggs, & Incubation. Oxford, Oxford University Press.

Nunney, L. 1996. The colonization of oranges by the cosmopolitan *Drosophila*. Oecologia 108:552-561.

Oliveira, C. C., M. H. Manfrin, F. M. Sene, and W. J. Etges. 2013. Evolution of male courtship songs in the
Drosophila buzattii species cluster, Pages 137-163 *in* P. Michalak, ed. Speciation: Natural
Processes, Genetics and Biodiversity. Hauppauge, NY, Nova Science Publishers.

Palanza, P., L. Re, D. Mainardi, P. F. Brain, and S. Parmigiani. 1996. Male and female competitive
strategies of wild house mice pairs (*Mus musculus domesticus*) confronted with intruders of
different sex and age in artificial territories. Behaviour 133:863-882.

Papadopoulos, N. T., J. R. Carey, P. Liedo, H.-G. Müller, and D. Sentürk. 2009. Virgin females compete
for mates in the male lekking species *Ceratitis capitata*. Physiological Entomology 34:238-245.

Paradis, E., and K. Schliep. 2019. ape 5.0: an environment for modern phylogenetics and evolutionary
analyses in R. Bioinformatics 35:526-528.

Parker, G. A. 1974. Assessment strategy and the evolution of fighting behaviour. Journal of Theoretical
Biology 47:223-243.

Pinheiro, J., D. Bates, and R. C. Team. 2022. _nlme: Linear and Nonlinear Mixed Effects Models_. R
package version 3.1-160, <https://CRAN.R-project.org/package=nlme>.

Pitnick, S. 1996. Investment in testes and the cost of making long sperm in Drosophila. American
Naturalist 148:57-80.

Pitnick, S., and T. A. Markow. 1994. Large male advantages associated with costs of sperm production in
Drosophila hydei, a species with giant sperm. Proceedings of the National Academy of Sciences,
USA 91:9277-9281.

Pitnick, S., T. A. Markow, and G. S. Spicer. 1995. Delayed male maturity is a cost of producing large
sperm in *Drosophila*. Proceedings of the National Academy of Sciences, USA 92:10614-10618.

—. 1999. Evolution of multiple kinds of female sperm-storage organs in *Drosophila*. Evolution 53:1804-
1822.

Promislow, D. E. L., and T. S. Haselkorn. 2002. Age-specific metabolic rates and mortality rates in the
genus *Drosophila*. Aging Cell 1:66-74.

R Core Team. 2017.R: A language and environment for statistical computing.R Foundation for Statistical
Computing, Viena, Austria.

R'kha, S., B. Moreteau, J. A. Coyne, and J. R. David. 1997. Evolution of a lesser fitness trait: egg
production in the specialist *Drosophila sechellia*. Genetics Research 69:17-23.

Redelings, B. D., and M. T. Holder. 2017. A supertree pipeline for summarizing phylogenetic and
taxonomic information for millions of species. Peer J 5:e3058.

Ribeiro, C., and B. J. Dickson. 2010. Sex peptide receptor and neuronal TOR/S6K signaling modulate
944   nutrient balancing in *Drosophila*. Current Biology 20:1000-1005.

945   Ringo, J., M. K. Kananen, and D. Wood. 1983. Aggression and mating success in three species of
946   *Drosophila*. Zeitschrift für Tierpsychologie 61:341-350.

947   Robie, A. A., J. Hirokawa, A. W. Edwards, L. A. Umayam, A. Lee, M. L. Phillips, G. M. Card et al. 2017.
948   Mapping the neural substrates of behavior. Cell 170:393-406.

949   Rosvall, K. A. 2008. Sexual selection on aggressiveness in females: Evidence from an experimental test
950   with tree swallows. Animal Behaviour 75:1603-1610.

951   —. 2011. Intrasexual competition in females: evidence for sexual selection. Behavioral Ecology 22: 1131–
952   1140.

953   —. 2013. Proximate perspectives of female aggression: good for the gander, good for the goose?
954   Philosophical Transactions of the Royal Society of London B 368:2030083.

955   Rosvall, K. A., A. B. Bentz, and E. M. George. 2020. How research on female vertebrates contributes to
956   an expanded challenge hypothesis. Hormones and Behavior 123:104565.

957   Santhosh, H. T., and M. S. Krishna. 2013. Relationship between male age, accessory gland, sperm
958   transferred, and fitness traits in *Drosophila bipectinata*. Journal of Insect Science 13:159.

959   Sarikaya, D. P., S. H. Church, L. P. Lagomarsino, K. N. Magnacca, S. L. Montgomery, D. K. Price, K. Y.
960   Kaneshiro, and C. G. Extavour. 2019. Reproductive capacity evolves in response to ecology
961   through common changes in cell number in Hawaiian *Drosophila*. Current Biology 29:1877–1884.

962   Schärer, L., J.-L. Da Lage, and D. Joly. 2008. Evolution of testicular archtecture in the Drosophilidae: a
963   role for sperm length. BMC Ecology and Evolution 8:143.

964   Schretter, C. E., Y. Aso, A. A. Robie, M. Dreher, M.-J. Dolan, N. Chen, M. ito et al. 2020. Cell types and
965   neuronal circuitry underlying female aggression in *Drosophila*. eLife 9:e58942.

966   Scott, M. P. 2006. Resource defense and juvenile hormone: The "challenge hypothesis" extended to
967   insects. Hormones and Behavior 49:276-281.

968   Seebacher, F., A. J. W. Ward, and R. S. Wilson. 2013. Increased aggression during pregnancy comes at a
969   higher metabolic cost. Journal of Experimental Biology 216:771-776.

970   Shingleton, A. W. 2015. Physiology: female flies have the guts for reproduction. Current Biology 25:R716-
971   R718.

972   Shultzaberger, R. K., S. J. Johnson, J. Wagner, K. Ha, T. A. Markow, and R. J. Greenspan. 2019.
973   ShConservation of the behavioral and transcriptional response to social experience among
974   *Drosophilids*. Genes, Brain and Behavior 18:e12487.

975   Sibbald, E. D., and C. M. S. Plowright. 2015. Reproductive potential and its behavioural consequences in
976   orphaned bumblebee workers (*Bombus impatiens*). Apidologie 46:618-627.

977   Singh, S. R., and B. N. Singh. 2003. Behavioral genetics of *Drosophila ananassae*. Genetics and
978   Molecular Research 2:394-409.

979   Sirot, L. K. 2019. On the evolutionary origins of insect seminal fluid proteins. General and Comparative
980   Endocrinology 278:104-111.

981   Smith, B. R., and D. T. Blumstein. 2008. Fitness consequences of personality: a meta-analysis.
982   Behavioral Ecology 19:448-455.

983   Snook, R. R., and T. A. Markow. 2001. Mating system evolution in sperm-heteromorphic *Drosophila*.
984   Journal of Insect Physiology 47:957-964.

985   Snook, R. R., T. A. Markow, and T. L. Karr. 1994. Functional nonequivalence of sperm in *Drosophila
986   pseudoobscura*. Proceedings of the National Academy of Sciences, USA 91:11222-11226.

987   Spieth, H. T. 1947. Sexual behavior and isolation in Drosophila I. The mating behavior of species of the
988   *willistoni* group. Evolution 1:17-31.

989   Stearns, S. C. 1989. Trade-offs in life-history evolution. Functional Ecology 3:259-268.

990   Stockley, P., and A. P. Campbell. 2013. Female competition and aggression: interdisciplinary
991   perspectives. Philosophical Transactions of the Royal Society of London B 368:20130073.

992   Suvorov, A., B. Y. Kim, J. Wang, E. E. Armstrong, D. Peede, E. R. R. D'Agostino, D. K. Price et al. 2022.
993   Widespread introgression across a phylogeny of 155 Drosophila genomes. Current Biology
994   32:111-123.

995   Syed, Z. A., R. Dallai, N. Nasirzadeh, J. A. Brill, P. M. O'Grady, S. Cong, E. M. Leef et al. 2021. Sperm
996   cyst "looping": A developmental novelty enabling extreme male ornament evolution. Cells
997   10:2762.

Tibbetts, E. A., C. Vernier, and J. Jinn. 2013. Juvenile hormone influences precontest assessment
behavior in *Polistes dominulus* paper wasps. Animal Behaviour 85:1177-1181.
Valadão, H., C. E. B. Proença, M. P. Kuhlmann, S. A. Harrison, and R. Tidon. 2019. Fruit-breeding
drosophilids (Diptera) in the Neotropics: playing the field and specialising in generalism?
Ecological Entomology 44:721-737.
Viblanc, V. A., C. Pasquaretta, C. Sueur, R. Boonstra, and F. S. Dobson. 2016. Aggression in Columbian
ground squirrels: Relationships with age, kinship, energy allocation, and fitness. Behavioral
Ecology 27:1716-1725.
Walker, S. J., V. M. Corrales-Carvajal, and C. Ribeiro. 2015. Postmating circuitry modulates salt taste
processing to increase reproductive output in *Drosophila*. Current Biology 25:2621-2630.
Wheeler, D. 1996. The role of nourishment in oogenesis. Annual Review of Entomology 41:407-431.
White, M. A., D. S. Chen, and M. F. Wolfner. 2021. She's got nerve: roles of octopamine in insect female
reproduction. Journal of Neurogenetics 35:132-153.
Wigby, S., N. C. Brown, S. E. Allen, S. Misra, J. L. Sitnik, I. Sepil, A. G. Clark, and M. F. Wolfner. 2020.
The *Drosophila* seminal proteome and its role in postcopulatory sexual selection. Philosophical
Transactions of the Royal Society of London B 375:20200072.
Yoon, J. S., K. P. Gagen, and D. L. Zhu. 1990. Longevity of 68 species of *Drosophila*. Ohio Journal of
Science 90:16-32.
Zhou, C., Y. Rao, and Y. Rao. 2008. A subset of octopaminergic neurons are important for *Drosophila*
aggression. Nature Neuroscience 11:1059-1067.