Trait-space disparity in fish communities spanning 380 million years from the Late Devonian to present

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Abstract

Aim: The diversity and distribution of traits in an ecological community determine how it functions. Modern fish communities conserve trait space across similar habitats. However, little is known about trait-space variation through deep time (or across vastly different habitats). In this paper, we sought to identify how trait diversity varies through space and time.

Location: Global Devonian and modern fish communities.

Methods: We examined two Devonian fish communities — a tropical reef (Gogo, Australia) and a tropical estuary (Miguasha, Canada) — and compared their trait space with that of six modern communities from diverse habitats.

Results: Trait-space metrics reflecting the amount of diversity found in each community (functional richness: FRic) and the similarity among species in each community (functional nearest-neighbour distance: FNND) indicated Devonian communities had similar scores to modern communities, but were less functionally rich than their closest modern analogues. Metrics describing location in trait space (centroid distances and hypervolume overlap) also showed distinctions between Devonian and modern communities. Indeed, location in trait space of Devonian and modern communities differed on several axes associated with a range of traits. Functional richness and nearest-neighbour distance also varied with habitat type and climate zone: estuarine communities had the highest nearest-neighbour distances followed by reef communities; reefs were the most functionally rich communities followed by estuaries; and functional richness was highest in tropical communities.

Main Conclusions: Fish community trait space varies with time, habitat and climate, suggesting (*i*) lability in fish trait space and (*ii*) evolutionary history, environmental filtering, and stochasticity influence community assembly. Furthermore, the large changes in community trait space that have occurred over deep time indicate changes in community structure and function.

Key words: community assembly, evolutionary constraints, fish communities, functional diversity, palaeoecology, trait diversity.

Introduction

The combination of a species' traits mediate its interactions with its environment [1,2], so the range and distribution of traits among interacting species is important in determining function and resilience of ecological communities [3–5]. Further, the diversity and distribution of traits in a community — also known as community 'trait space' — can provide insights into the rules of community assembly, including what roles evolution, species filtering, and niche differentiation play in controlling how communities form [6–8]. Indeed, recent research on the trait space of contemporary communities has addressed how communities develop and adjust to their ever-shifting environments [9,10]. However, it is still unclear whether patterns observed within and among modern communities also apply to ecological communities over deep time. Consistent spatial and temporal patterns in community trait space would suggest that community assembly rules are constant, whereas variation could indicate phylogenetic constraints [11], stochasticity in evolutionary pathways [12,13], and/or shifts in the environmental parameters that determine community composition [14].

A range of metrics are commonly used to describe and investigate community trait space [15]. Such trait-diversity metrics often indicate variation among communities in different habitats and climates [16,17]. For example, trait diversity in fish (non-tetrapod vertebrate) communities differs depending on substratum (e.g., reef *versus* seagrass *versus* sand) [18–20] and habitat complexity [21,22]. Despite this variation, there is also evidence for the conservation of trait space across similar habitats, even when there are substantial phylogenetic and taxonomic differences among the resident communities [23]. That evolutionarily distinct communities occupying similar habitats have similar trait spaces suggests convergence (through evolution and/or niche filtering) and highlights the

importance of deterministic processes in shaping community trait space (i.e., environment
determining trait space) [24,25].

Although trait space varies among habitats and is conserved within habitat types across
locations in contemporary communities, it remains unclear how it varies through deep time.
Few studies have examined changes in functional diversity and trait space over millions of
years (but see [26] for impacts of extinction events). By testing for patterns across time as
well as across contemporaneous habitats, it is possible to test whether similar habitats select
for similar community trait space irrespective of phylogeny and evolutionary history.

The Devonian Period (419.2 to 358.9 million years ago [Ma]) is popularly termed the
"Age of Fishes". During that time, diverse fish communities established and radiated across
the planet, giving rise to the first tetrapods. Four mass extinctions have since occurred (not
including the current one) [27], which along with the passing of time and millions of
'background' extinction and speciation events, have caused massive phylogenetic turnover
[28,29]. In addition, Devonian and modern fish communities differ in terms of the non-fish
species and nutrient inputs that make up the wider ecological community [30,31]. For
example, Devonian reefs were primarily built of stromatoporoid sponges, calcareous algae,
and microbial communities rather than hexacorallian corals (but also included some rugose
and tabulate corals) [32,33], and ancient estuaries generally had low invertebrate and algal
diversity compared to modern estuaries [34,35]. Thus, although the environments Devonian
fish communities experienced were physically similar to their modern counterparts, their
vertebrate and non-vertebrate biota differed greatly. By comparing these ancient assemblages
to modern communities, it is therefore possible to test if physical environment is the primary
determinant of fish community trait diversity across space and time, or whether biological,
phylogenetic, and evolutionary differences associated with the passing of hundreds of
millions of years have a dominant effect on trait space. Such knowledge will improve our
understanding of how communities have responded to change in the past, and how they could
change in response to anthropogenic and non-anthropogenic disturbances in the future.

We compared the trait space of two of the best-described Devonian fish communities — a
tropical reef (Gogo Formation, northern Australia) and a tropical estuary (Miguasha, eastern
Canada) — to that of six modern fish communities from different habitats and climate zones,
to identify how trait diversity varies through space and time and thereby shedding light on the
processes of community assembly. We compared these communities in terms of two trait-
space metrics that reflect diversity and distribution of traits within each community
(functional richness [FRic] and functional nearest-neighbour distance [FNND]) and two that

quantify each community's position in trait space relative to other communities (distance between hypervolume centroids and overlap of hypervolumes) [36]. We hypothesise that (*i*) if community trait space is primarily determined by habitat, there will be larger differences between communities from different habitats (i.e., reef *versus* estuary *versus* freshwater) than between Devonian and modern communities from the same habitat (e.g., Devonian reef community similar to modern reef community) or between tropical and temperate/subtropical communities from the same habitat (e.g., tropical reef similar to temperate reef). (*ii*) If community trait space is primarily determined by climate, there will be larger differences between tropical *versus* temperate/subtropical communities than between communities from different periods of time or different habitat types within the same climate zone. (*iii*) If community trait space is primarily determined by evolutionary history and the phylogenetic makeup of its constituent species, there will be larger differences between Devonian and modern communities compared to between communities from different habitat types or different climate zones within the same period. (*iv*) If time, habitat, and climate all substantially affect community trait space, there will be consistent groupings/patterns across all these variables (e.g., community metric scores will group together based on time, habitat, and climate); and (*v*) If community trait space is independent of time, habitat, and climate, trait space metrics will not be grouped by period (Devonian *versus* modern), habitat or climate zone. By identifying which of these five patterns are observed in trait space metrics, we provide insight into the factors shaping community trait space (i.e., community assembly) in the past, present and future.

Methods

Sites and species lists

We compared two Late Devonian (Frasnian) and six modern fish communities. The Late Devonian communities of Miguasha (Escuminac Formation) in north-eastern Canada and the Gogo Formation in north-western Australia are two of only six recognised fish Lagerstätten from this period [37]. We selected these communities based on (1) their fossil records capturing most of the *in situ* fish diversity, and (2) the availability of trait data for a range of traits and species that make up these communities (Table S1.1, see Appendix S1 in Supporting Information). Miguasha (modern: 48° 48′ N, 66° 16′ W) was a tropical estuary in Laurussia 385 to 374 million years ago [38–40]. Thousands of fish specimens have been preserved and recovered from Miguasha, representing at least 19 species [39]. The Gogo

Formation (modern: 18° 26′ S, 125° 55′ E) represents the basinal and channel facies adjacent

to a tropical, stromatoporoid sponge-dominated reef that was on the edge of the Gondwanan

landmass 384 to 382 million years ago; the fish preserved in the Gogo Formation were

deposited under anoxic marine conditions and came from the adjacent reef [33]. Fifty-three

fish species have been identified from Gogo (including 47 described and 6 undescribed

species; see Appendix S1, Tables S1.1 and S1.2) [33,41]. Discovery curves suggest all

(fossilised) fish species from Miguasha have been described [34] and that most (> 95%) fish

species from Gogo have been identified (see Appendix S1, Fig. S1.1 and Table S1.2).

We selected six modern fish communities from a diversity of environments (different

habitat types and climate zones) for comparison to the Devonian communities. Our selection

of modern communities was further guided by the availability of species lists for the

assemblage and the presence of information on trophic interactions (allowing for later

development of trophic-network models of those communities). We included reef, estuarine,

and freshwater communities, with tropical and temperate or subtropical representatives of

each (i.e., we included each habitat-type/climate-zone combination). The modern

communities were: Caribbean reefs (tropical; 18° 3′ N, 65° 28′ W) [42]; Chilean reefs

(temperate/subtropical; 31° 30′ S, 71° 35′ W) [43]; Santa Cruz Channel, north-eastern Brazil

(tropical estuary; 7° 46′ S, 34° 53′ W) [44,45]; Ythan Estuary, Scotland (temperate; 57° 19′

N, 1° 59′ W) [46–48]; Braço Morto Acima and Abaixo, central Brazil (tropical freshwater

oxbow lakes; 19° 41′ S, 56° 59′ W) [49–58]; and the Nepean River, New South Wales,

Australia (temperate/subtropical freshwater river section; 33° 47′ S, 150° 38′ E; ala.org.au;

see Appendix S1, Table S1.3) [59].

Traits

We collected information on 11 traits from the 488 species in our study (416 extant species

and 72 Devonian species; see Appendix S1, Table S1.4). We chose traits based on their

ecological relevance and their availability for both living and extinct species. The traits were:

sagittal body shape, transverse body shape, maximum total body length (tip of snout to tip of

caudal fin), head length, eye diameter, pre-orbital length, body depth, position of mouth,

position of eyes, presence of spiracle, and caudal fin shape (see Appendix S1, Fig. S1.2 and

Table S1.4). All morphometric variables (except maximum total body length) were expressed

as a proportion of maximum total body length as a size standardisation and to reduce

correlations among these traits.

We collected trait data for Devonian species from the literature, specimens, and

photographs of specimens. However, it was not possible to collect all traits for the species

from the Gogo Formation because some of them are only known from incomplete or

disarticulated body parts (due to the preservational environment) [60]. We addressed these

gaps by estimating traits using: (1) expert opinion in cases where traits could be confidently

inferred (inferences made by J. Long, A. Clement, B. Choo, and K. Trinajstic) or (2) multiple

imputation using species' traits, coarse taxonomy, and the 'missForest' R package [61]

(GitHub folder). Evaluating multiple imputation performance indicated out-of-bag error

estimates of 0.30 (normalised root-mean-square error) and 0.11 (proportion of falsely

classified) for continuous and categorical variables, respectively. For modern species, we

extracted trait data from *Fishbase* (using the 'rFishbase' package) [62], photographs,

and scientific literature; GitHub folder). We visually inspected the distribution of continuous

traits, identifying two as right-skewed (total length and pre-orbital length); we $\log_e$-

transformed these traits to normalise them.

Analyses

We built a Gower dissimilarity matrix [63] to quantify trait differences among species using

the 'gawdis' package in R [64]. Gower dissimilarity can handle mixed data types (our trait

data included continuous and categorical variables) and can normalise the contribution of

variables [6,63]. Including all traits resulted in an unbalanced distribution, so we also

calculated Gower distances with the maximum number of traits that did not result in uneven

distributions (there were three such combinations of nine traits). Observed patterns in

functional diversity metrics were consistent across these trait sets (see Appendix S1, Fig.

S1.3 and S1.4). We therefore only present results from the 'all traits' Gower distance trait set

in the main text.

We applied principal coordinate analysis (PCoA) to the Gower distances to ordinate

species within a lower-dimensional space and calculate functional diversity metrics using the

'mFD' R package [65]. The first eight principal coordinate axes explained 73% of the total

variation (GitHub folder); we identified that eight was the optimal number of axes based on

scaled mean absolute deviation and scaled root of mean square error (minimising the

deviation between trait-based distances and functional space distances; GitHub folder). We

therefore used these eight axes to calculate each community's functional richness (FRic) [66]

and functional nearest neighbour (FNND) [67] scores. Functional richness, defined as the

proportion of functional space filled by a community, quantifies the functional diversity of a community. In contrast, functional nearest neighbour is the average distance in trait space to each species' nearest neighbour within the community, quantifying functional similarity (redundancy) among species in a community. These metrics, calculated from PCoA coordinates, are unitless 'scores' that can be used to compare communities. We also calculated functional evenness, functional divergence, and functional specialisation (indicating how regularly species are distributed in trait space, how distinct species are from the centroid of the community's trait space, and how distinct species in a community are from the global centroid, respectively) (see Appendix S1, Fig. S1.5).

Species diversity varied substantially among the included communities, ranging from 17 species in Ythan Estuary to 208 species in Caribbean reefs (see Appendix S1, Tables S1.1 and S1.3). Functional diversity metrics can be correlated with species diversity [15,68]. We therefore also calculated standardized effect sizes — a method that controls for species richness — for both metrics (functional richness and nearest-neighbour scores) [69]. To do this, we generated 1000 null models by randomly shuffling species among the communities while maintaining the species richness of each community. We then calculated the mean and standard deviation of the functional diversity metrics for each community in the null models. We subtracted null mean values from observed functional diversity metrics to calculate the effect sizes, which we divided by the null standard deviations to standardize across scales. Controlling for species diversity in this way allowed us to (*i*) separate patterns due to species diversity from those due to functional diversity, (*ii*) detect patterns in functional diversity that were masked by species diversity, and (*iii*) limit the effects of species with extreme traits. We plotted both the 'observed' and standardized effect size (species-diversity controlled) metrics, allowing us to evaluate visually whether period, habitat type, or climate zone grouped communities in either of these metrics.

We also used the principal coordinates to build trait-space hypervolumes for each community using Gaussian kernel density estimation and the 'hypervolume' package in R [70]. We set the requested quantile to 0.95 and estimated the bandwidth for each axis in each community separately. We then used the mean estimated bandwidth for each axis across the eight communities as fixed bandwidths to recalculate hypervolumes (i.e., ensuring the same bandwidths for each community). The resulting hypervolumes were used to quantify distances between community centroids (geometric centre) and overlap in multidimensional trait space. We measured overlap using the Jaccard index, and we further decomposed non-overlap into its turnover and nestedness components [71]. We made these comparisons

between all pairwise community combinations to test if communities were grouped/clustered

according to their period, habitat, or climate zone (see hypotheses listed in the introduction).

Because distances between centroids suggested the fish community trait spaces were

segregated, we identified the specific traits responsible by determining along which

hypervolume axes the centroids differed, and which traits were correlated with these axes.

We then investigated how these traits were distributed among the fish communities.

Results

Functional richness of the Devonian communities fell within the range observed in modern

communities, irrespective of whether we controlled for species richness (Fig. 1a,b; not

controlling for species richness: Gogo $= 2.4 \times 10^{-3}$, Miguasha $= 4.7 \times 10^{-4}$, modern

assemblages $= 2.6 \times 10^{-5}$ to 0.29; controlling for species richness: Gogo $= -2.4$, Miguasha $= -$

1.0, modern assemblages $= -0.8$ to -2.9). However, the Devonian communities had lower

functional richness than their closest modern counterparts when not controlling for species

richness (Fig. 1a; Gogo *versus* Caribbean: 2.4×10^{-3} *vs.* 0.29; Miguasha *versus* Santa Cruz

Estuary 4.7×10^{-4} *vs.* 4.5×10^{-2}). Standardized effect sizes controlling for species diversity

indicated that Devonian communities had similar functional richness to their modern

counterparts (Fig. 1b; Gogo *versus* Caribbean: -2.4 *vs.* -2.0; Miguasha *versus* Santa Cruz

Estuary -1.0 *vs.* -1.8). There was habitat-linked variation in functional richness, but the pattern

differed depending on whether we controlled for species diversity (Fig. 1a,b). Reef

communities had the highest score followed by estuarine communities in observed/raw

functional richness (Fig. 1a), whereas estuarine communities had the highest scores followed

by reefs after controlling for species diversity (Fig. 1b). This habitat-functional richness

pattern, including the switching between reefs and estuaries as the most functionally rich

communities when controlling for species diversity, emerged in both modern and Devonian

communities (Fig. 1a,b). Among the modern assemblages, tropical communities had higher

observed functional richness than their temperate/subtropical counterparts (Fig. 1a; mean

richness tropical *versus* temperate/subtropical: 0.11 vs. 0.0006), whereas temperate and

subtropical communities had higher functional richness when controlling for species diversity

(Fig. 1b; mean richness tropical *versus* temperate/subtropical: -2.2 vs. -1.0).

Figure 1. Functional diversity metrics for two Devonian and six modern fish communities. Two functional diversity metrics are shown: functional richness (a. and b.) and nearest neighbour (c. and d.). Panels in the left column show the observed metrics (not controlling for species diversity), whereas panels in the right column show standardized effect sizes (controlling for species diversity). Pink background indicates the five tropical communities (ancient and modern), yellow background indicates the three temperate/subtropical communities (modern only).

Nearest-neighbour scores, reflecting the distance in functional space of each species to the nearest species in its community, were similar for Devonian and modern communities (Fig. 1c,d). Miguasha had a slightly higher score than the other communities when not controlling for species diversity (Fig. 1c; Miguasha = 0.421, Gogo = 0.207; modern assemblages = 0.181 to 0.411). After accounting for species diversity, nearest-neighbour scores for Gogo and

Miguasha fell within the range observed in the modern communities (Fig. 1d; Gogo = -5.2,

Miguasha = -1.7; modern assemblages = -1.6 to -5.8).

There were differences among habitats in nearest-neighbour scores within periods and

climate zones. Freshwater communities consistently had the lowest nearest-neighbour scores

followed by reef communities (Fig. 1c,d). Further, there was a consistent latitudinal pattern

among modern communities — species in temperate and subtropical communities were

farther from their nearest neighbour than were species in tropical communities (Fig. 1c,d;

mean nearest neighbour tropical *versus* temperate/subtropical: 0.21 vs. 0.36 [not controlling

for species diversity] and -4 vs. -2.2 [controlling for species diversity]).

The distances between hypervolume centroids of Devonian *versus* modern communities

were greater than those between centroids of communities from the same period (Fig. 2a,b;

distance between Devonian and modern: 0.16 ± 0.01, distance between communities from the

same period: 0.07 ± 0.02; reported values are means ± SD unless otherwise stated), indicating

that community trait spaces were centred in different locations depending on their time

period. Centroids of communities from the same habitat type or the same climate zone were

not closer together than centroids from different habitats or climate zones (Fig. 2c,d).

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301 **Figure 2.** Distances between centroids of functional trait space hypervolumes for fish communities. Two
302 Devonian (Gogo and Miguasha) and six modern fish communities from different habitats and climates are
303 compared. On the left, plot a. is a heatmap showing distances between the centroids of each community. On the
304 right are three violin plots showing distances between communities from either the same (within) or different

(between): b. periods, c. habitat types, and d. climate zones. Plot points in the violin plots represent individual comparisons between communities, jittered on the *x* axis for clarity.

The hypervolume Jaccard index measuring trait-space overlap also indicated less similarity between Devonian and modern communities compared to between communities from the same period (Fig. 3a,b; overlap between Devonian and modern: 0.05 ± 0.02, overlap between communities from the same period: 0.18 ± 0.07). The overlap index between Devonian communities was within the range observed among modern communities (Devonian overlap = 0.16, modern community overlap = 0.09-0.37; Fig. 3a). There was no evidence of greater overlap between hypervolumes of communities from the same *versus* different habitat types or climate zones (Fig. 3c,d). Partitioning the Jaccard index into components of dissimilarity due to turnover *versus* nestedness indicated that turnover largely explained differences among trait spaces i.e., communities occupied different trait spaces rather than subsets of each other's trait spaces (dissimilarity due to turnover: 0.52 to 0.96; dissimilarity due to nestedness: 0 to 0.3; see Appendix S1, Fig. S1.6 and S1.7).

Figure 3. Jaccard Index comparing overlap in functional trait space hypervolumes for fish communities. Two Devonian (Gogo and Miguasha) and six modern fish communities from different habitats and climates are compared. On the left, plot a. is a heatmap showing Jaccard indices for all community comparisons. On the right is a series of violin plots showing Jaccard indices between communities from either the same (within) or different (between): b. periods, c. habitat types, and d. climate zones. Plot points in the violin plots represent individual comparisons between communities, jittered on the *x* axis for clarity.

The centroids for both Devonian communities were outside the range observed in modern communities on axis 1 (Devonian ≤ -0.089, modern ≥ -0.045), axis 2 (Devonian ≥ 0.061, modern ≤ 0.009), and axis 7 (Devonian ≥ 0.026, modern ≤ -0.003; see Appendix S1, Table S1.5 and Fig. S1.8).

Although directly linking the principal coordinate axes to traits is not possible because the axes are based on Gower distances between species, correlations between species coordinates along these axes and traits suggest they mostly reflect eye diameter (correlation: $\eta^2 = 0.623$, p < 0.000), body depth ($\eta^2 = 0.537$, p < 0.000), and caudal fin shape ($\eta^2 = 0.339$, p < 0.000) on axis 1; position of mouth ($\eta^2 = 0.415$, p < 0.000) and eye position ($\eta^2 = 0.371$, p < 0.000) on axis 2; and transverse body shape ($\eta^2 = 0.378$, p < 0.000), and presence of spiracles ($\eta^2 = 0.221$, p < 0.000) on axis 7 (see Appendix S1, Table S1.6). Accordingly, Devonian communities were distinct from modern communities in these traits. Fish in the Devonian communities tended to have smaller eye diameters, their bodies were not as deep, and they displayed less variation in these traits compared to fish in the modern communities (eye diameter relative to total body length: 0.039 ± 0.015 *versus* 0.052 ± 0.023, body depth relative to total body length: 0.190 ± 0.048 *versus* 0.257 ± 0.113, for Devonian and modern communities, respectively; see Appendix S1, Fig. S1.9c,e). None of the fish in these Devonian communities had superior mouth position whereas all the modern communities had some fish with this mouth position (see Appendix S1, Fig. S1.10). None of the Devonian fish had eyes migrated to one side of the body (as in flatfish; see Appendix S1, Fig. S1.10), none of the modern fish had hypocercal tails (see Appendix S1, Fig. S1.10), spiracles and heterocercal tails were more prevalent in the Devonian than modern communities (large spiracles: 56% *versus* < 2%; heterocercal tails: 83% *versus* < 3% for Devonian *versus* modern communities, respectively; see Appendix S1, Fig. S1.10), and the proportion of fish with a compressed transverse body shape is lower in the Devonian fishes (25% *versus* > 55% in Devonian *versus* modern communities, respectively; see Appendix S1, Fig. S1.10c).

Gogo's centroid was also extreme on axes 5 and 6 (with values of 0.056 and 0.019, respectively, compared to values ≤ 0.002 and ≤ 0.017 for the other communities on the same axes; see Appendix S1, Table S1.5), while Miguasha's centroid was extreme on axis 4 (Miguasha = -0.049, other communities ≥ -0.026; see Appendix S1, Table S1.5). Sagittal (not transverse) body shape was the most strongly correlated trait with these axes, showing a

slightly stronger correlation with axis 5 ($\eta^2 = 0.397$, p < 0.000) than with axes 4 and 6 ($\eta^2 =$

0.311, p < 0.000 and $\eta^2 = 0.252$, p < 0.000 for axes 4 and 6, respectively; see Appendix S1,

Table S1.6). These correlations suggest that the extreme positions of Gogo and Miguasha

could be attributed to Gogo's lack of eel-shaped and elongate (e.g., anguilliform) fish and to

Miguasha's higher occurrence of fish with unconventional ('other') body shapes (see

Appendix S1, Fig. S1.10a), which included the tadpole-shaped *Escuminaspis laticeps* and

Levesquaspis patteni.

Discussion

Our analysis of modern and ancient fish communities revealed patterns in community trait

space associated with period, habitat type, and climate zone. The importance of these

variables differed among trait-space metrics, indicating that different factors are important for

different trait-space attributes. The two metrics that reflect diversity and distribution of traits

within a community (functional richness and nearest neighbour) suggest: (1) broad similarity

between Devonian and modern assemblages, but with Devonian communities having lower

functional richness than their closest modern counterparts when not accounting for species

diversity, (2) an association with habitat type, and (3) an association with climate zone (Fig.

1; consistent with hypothesis *iv*). In contrast, period was the only variable that clearly

differentiated fish assemblages in the two metrics that reflect a community's position in trait

space relative to other communities (distance between hypervolume centroids and Jaccard

index); we observed no associations between these metrics and habitat type or climate zone

(Fig. 2, 3; consistent with hypothesis *iii*). The trait-space positions of Devonian communities

were distinct from modern communities on several axes associated with seven of the eleven

traits we included in our analyses (see Appendix S1, Table S1.5 and S1.6); such trait

differences could indicate disparity between Devonian and modern communities in terms of

their structure and function. Further, that communities from the same period have similar trait

spaces, while those from different periods are dissimilar (consistent with hypothesis *iii*),

suggests evolutionary constraints and stochasticity in evolutionary pathways are potentially

important in determining community trait space and community assembly.

Period

If fish community trait space has changed through time, we would expect to see communities

from different periods segregated in terms of their trait-space metrics (i.e., modern

communities similar to each other and different to Devonian communities). This pattern is

clear in three of the four metrics we calculated: Devonian communities differed from modern

communities in terms of functional richness, centroid location, and hypervolume overlap

(Fig. 1a, 2, 3).

Functional richness of Devonian communities was within the range spanned by all modern

communities, but the scores of Devonian communities in this metric were substantially lower

than those observed in modern communities from the same habitat type and climate zone

when not controlling for species diversity (i.e., Gogo Reef *versus* Caribbean Reef, Miguasha

versus Santa Cruz Estuary; Fig. 1a). Despite the Devonian period being popularly named the

'Age of Fishes', Devonian fish communities tended to have lower species diversity compared

to modern fish communities; our results suggest they also had lower trait diversity compared

to their modern counterparts (Fig. 1a,b). Discovery curves confirm that most species in the

Devonian communities we examined have been described (see Appendix S1, Table S1.2, Fig.

S1.1 and [34]), and, therefore, the relatively low functional and species richness in Devonian

communities compared to their modern analogues are probably real features instead of

preservational artefacts. When we controlled for the effect of species diversity on functional

richness using standardized effect sizes, Devonian communities were comparable to their

modern counterparts (Fig 1b). This result emphasizes that changes in trait diversity between

time periods are associated with changes in species diversity, although it remains unclear

whether higher trait diversity is a cause or effect (or both) of higher species diversity.

Lower functional richness in Devonian fish assemblages does not necessarily mean that

more trait space was vacant in these communities. It is possible that non-fish taxa filled some

of the space occupied by fish in modern communities. For example, conodonts (early, non-

fish, but fish-like, jawless vertebrates) might have occupied the trait space of modern

lampreys, hagfish, and small eels [72,73], while invertebrates such as Eurypterids (sea

scorpions) might have filled trait and trophic spaced occupied by predatory fishes in modern

communities [74]. Similarly, non-fish taxa, such as sea snakes, turtles, and marine mammals,

make unique contributions to the trait space in modern aquatic communities [75].

Centroid distances and the Jaccard index (trait-space overlap) are metrics that reflect the

location of a community's trait space relative to other communities. The fact that Devonian

and modern communities are separated by these metrics indicates variation in trait values

(continuous traits), trait status (categorical traits), and/or trait combinations. By examining

which hypervolume axes separated modern and Devonian communities, and which traits

were correlated with these axes, we were able to identify the traits involved in differentiating

the trait spaces of these communities. Seven of the eleven traits we included in our analysis appear to contribute to this differentiation, including: eye diameter, body depth, presence of large spiracles, transverse body shape, caudal fin shape, mouth position, and eye position (see Appendix S1, Fig. S1.6 and S1.7). Of these traits, eye diameter, body depth, mouth position and eye position had lower diversity in Devonian compared to modern communities (see Appendix S1, Fig. S1.6 and S1.7). Thus, differences between modern and Devonian communities as indicated by centroid distances and the Jaccard index reflect, at least in part, trait space that is absent in the Devonian fish, consistent with the observed differences in functional richness between communities from these periods (Fig. 1a,b). However, decomposition of the Jaccard index dissimilarity suggested differences between Devonian and modern fish community trait spaces were primarily due to turnover rather than nestedness (see Appendix S1, Fig. S1.6 and S1.7). In other words, Devonian communities occupied different trait space to modern communities rather than a subset of that trait space. This result is counterintuitive because the modern communities largely encompass the trait variation found in the Devonian communities when traits are considered individually (see Appendix S1, Fig. S1.9 and S1.10). Thus, the dominance of turnover in explaining trait space dissimilarity is likely due to unique combinations of traits rather than unique trait values among Devonian fish.

Community trait space is linked to community structure and function [76,77]. Our trait space results therefore suggest substantial ecological differences between Devonian and modern fish communities. For example, the dearth of Devonian fish with a superior mouth position — an adaptation to feeding from above [78,79] — indicates differences in trophic interactions. While superior mouth position has been documented in Devonian species [80,81], the rarity of this trait in the Devonian contrasts with its widespread occurrence in modern communities (see Appendix S1, Fig. S1.7e). Greater diversity of mouth position does not only indicate a greater variety of feeding strategies used today, but also suggests that species in modern communities experience different predation pressures to those present in the Devonian (e.g., in terms of predator archetypes) [82], which could lead to the evolution of distinct antipredator strategies. Similarly, the lack of fish with deep, laterally compressed bodies in the Devonian communities (i.e., fish from these communities had body depths relative to their total length < 33%, and only 25% were laterally compressed; see Appendix S1, Fig. S1.6 and S1.7) likely indicates ecological differences associated with microhabitat use and how a species interacts with conspecifics and other species [83,84]. Deep-bodied, laterally compressed jawless fish have been documented in older communities (Late Silurian

and Early Devonian) [85], but they were less widespread compared to deep-bodied fish in modern communities, and they appear to have gone extinct before the Late Devonian (i.e., the epoch we examined). Thus, the relatively low variation of traits in Late Devonian communities suggests they were less ecologically/functionally diverse overall than modern fish communities.

The greater trait variation we observed in modern *versus* Devonian fish communities could result from (*i*) greater species diversity (which could be a cause or effect of trait diversity), (*ii*) increased habitat/environmental complexity through time [86], (*iii*) the longer time modern fish assemblages have had to evolve and diversify [87], and/or (*iv*) the morphological flexibility of teleost fish, a group that did not evolve until the Triassic Period (> 100 million years after the Devonian; although greater variation in actinopterygian traits started to emerge immediately following the Hangenberg extinction event at the end of the Devonian) [88]. In modern fish communities, there is a positive correlation between habitat complexity and functional richness [22,89]. Thus, given that Devonian habitats appear to have been less complex than modern habitats (e.g., even the most complex Devonian reefs tended to be less structurally complex than modern coral reefs [90] and diversity of benthic/habitat-forming organisms was lower [91]), this factor might contribute to the lower trait diversity in Devonian fish. Conversely, teleosts have several traits that foster morphological flexibility. For example, the prehensile mouth parts of teleosts give great flexibility in mouth-linked traits, leading to the evolution of different mouth positions and head shapes [92]. Other attributes that promote morphological flexibility in teleosts and other osteichthyans — fish that are more prevalent in modern communities compared to Devonian ones — include the presence of a swim bladder for buoyancy control and bony endoskeletons, facilitating diversification in movement patterns and body shape [93,94]. While our results suggest higher trait diversity in modern fish communities, we acknowledge that the choice of traits and the groupings used for categorical variables could influence conclusions regarding which taxa or communities are most diverse [6]. For example, if we had focused on bony armour plates and lobe-paired fin structure, Devonian communities would be classified as more trait-diverse than modern communities [95]. Regardless, our main conclusion — that trait space of modern and Devonian communities differ — should hold true for any broad sample of traits across the entire phenotype.

Habitat

Functional nearest-neighbour scores did not show consistent differences between periods, but

did display variation with habitat type: freshwater communities consistently had the lowest

scores (their constituent species were the most similar to each other), while estuarine

communities had the highest nearest-neighbour scores (Fig. 1c,d). The low functional

richness and low nearest-neighbour scores in the freshwater communities we examined (i.e.,

indicating low trait diversity and high similarity among species; Fig. 1a,b,c,d) is likely the

result of strong niche filtering, corroborating previous conclusions that habitat filters

determine species (and therefore trait) composition in freshwater communities [96,97]. Reef

communities in both periods have high functional richness when species diversity was not

controlled (Fig. 1a), and low-to-intermediate nearest-neighbour scores (Fig. 1c,d). This

combination suggests that many, densely concentrated and diverse niches are facilitated by

complex and productive reef habitats (corroborating research on modern communities) [98].

Estuarine communities have the highest nearest-neighbour scores in their climate zone and

time period (Fig. 1c,d), indicating that their species are distinct from each other in trait space.

The trait variation among fish in estuarine communities might reflect that these assemblages

are an admixture of species from different origins — they include marine, freshwater, and

diadromous fish, as well as species that complete their life cycle in estuaries — and therefore

include species that have experienced different niche/environmental filters [99,100]. The

association between nearest-neighbour scores and habitat type, and the lack of a consistent

difference between communities from different periods, suggests that environmental factors

are the primary determinants of how similar species within a community can be to each other.

We also found that functional richness varied with habitat type, but the functional

richness-habitat pattern depended on whether we controlled for species diversity (Fig. 1a,b).

Across climate zones, reef communities had the highest observed functional richness

followed by estuarine communities, whereas estuarine communities had the highest richness

followed by reef communities when controlling for species diversity (Fig. 1a,b). We observed

this switching between which habitat type had the highest functional richness in modern and

Devonian communities (Fig. 1a,b). High habitat complexity could explain the elevated

species and functional richness in reef communities by allowing greater niche differentiation

among resident fish [98]. Indeed, that species richness was higher for reef *versus* estuarine

communities (e.g., 208 *versus* 86 species for tropical communities, and 27 *versus* 17 species

for temperate/subtropical communities) also potentially explains why controlling for species

diversity switches which habitat type had the highest functional richness. The difference

might result from saturation of trait space in species-rich reef communities (i.e., the

expansion of a community's trait space associated with the addition of a species diminishes
as more species are added to the community) [6,101–104], a hypothesis supported by the low
nearest-neighbour scores for the reef compared to estuarine communities (Fig. 1c,d).
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Climate
Functional richness of modern tropical communities was higher than that of modern
temperate and subtropical communities when not controlling for species diversity, consistent
with the previously reported pattern of increasing functional richness with decreasing latitude
in fish, other vertebrates, and invertebrates [104–109]. This pattern was reversed when we
controlled for species diversity (Fig. 1b), indicating that high trait diversity in tropical fish
communities is associated with high species richness.
Our results also revealed a latitudinal/climate pattern among modern communities in terms
of their nearest-neighbour scores, with tropical communities having lower scores than their
temperate/subtropical counterparts (Fig. 1c,d). Together, functional richness and nearest-
neighbour scores indicate that fish communities in the tropics have greater variation in traits
and their constituent species are packed more closely together in trait space than observed in
temperate and subtropical communities (where species richness also tends to be lower)
[105,110]. This suggests stronger niche differentiation and reduced functional redundancy at
higher latitudes, potentially indicating that competition is more limiting in temperate than
tropical zones [111]. We were unable to assess whether latitudinal patterns occurred among
the Devonian fish communities because both Devonian communities in our study were
tropical at deposition. Thus, future research could test whether latitude (climate) has been
important in determining community trait space through time, or whether this pattern
developed recently.
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Conclusion
The differences we detected between modern and Devonian fish communities indicate that
community trait space has changed through time. Although the Devonian period is known for
its diverse and abundant fish fauna, our results suggest that modern fish communities are in
fact more trait-diverse (and species-diverse) than their Devonian counterparts, and that their
trait spaces are centred in different locations in trait space. These differences could indicate
several phenomena, including the longer time modern communities have had to develop and
diversify [87], greater variation in habitats today [91, 112], phylogenetic constraints of
different fish lineages [113], and/or stochasticity in evolutionary pathways [114]. Irrespective

of the root cause(s), the observed differences in trait space suggests Devonian communities

were structured and functioned differently to modern communities. Further research on how

community structure and function has changed through time, combining information from

ancient and modern communities [115], could provide important insights into community-

assembly rules and the ecological and evolutionary responses of communities to

environmental disturbances in the past and the future.

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Data availability statement

All data and code is available on Github (github.com/JohnLlewelyn/Trait-space-disparity). De-

identified data and code will be made available to reviewers in a zip file.

References

1. McGill BJ, Enquist BJ, Weiher E, Westoby M. 2006 Rebuilding community ecology
from functional traits. *Trends in Ecology & Evolution* **21**, 178–185.
(doi:10.1016/j.tree.2006.02.002)
2. Kiørboe T, Visser A, Andersen KH. 2018 A trait-based approach to ocean ecology. *ICES*
Journal of Marine Science **75**, 1849–1863. (doi:10.1093/icesjms/fsy090)
3. Cadotte MW. 2017 Functional traits explain ecosystem function through opposing
mechanisms. *Ecology Letters* **20**, 989–996. (doi:10.1111/ele.12796)
4. McLean M, Auber A, Graham NAJ, Houk P, Villéger S, Violle C, Thuiller W, Wilson
SK, Mouillot D. 2019 Trait structure and redundancy determine sensitivity to disturbance
in marine fish communities. *Global Change Biology* **25**, 3424–3437.
(doi:10.1111/gcb.14662)
5. Micheli F *et al.* 2014 High vulnerability of ecosystem function and services to diversity
loss in Caribbean coral reefs. *Biological Conservation* **171**, 186–194.
(doi:10.1016/j.biocon.2013.12.029)
6. Mouillot D *et al.* 2021 The dimensionality and structure of species trait spaces. *Ecology*
Letters **24**, 1988–2009. (doi:10.1111/ele.13778)
7. Schöb C, Butterfield BJ, Pugnaire FI. 2012 Foundation species influence trait-based
community assembly. *New Phytologist* **196**, 824–834. (doi:10.1111/j.1469-
8137.2012.04306.x)
8. Weiher E, Clarke GDP, Keddy PA. 1998 Community assembly rules, morphological
dispersion, and the coexistence of plant species. *Oikos* **81**, 309–322.

(doi:10.2307/3547051)

9. Frimpong EA, Angermeier PL. 2010 Trait-based approaches in the analysis of stream fish communities. *American Fisheries Society Symposium* **73**, 109–136.

10. Vogel A *et al.* 2019 Chapter Three - Lost in trait space: Species-poor communities are inflexible in properties that drive ecosystem functioning. In *Advances in Ecological Research* (eds N Eisenhauer, DA Bohan, AJ Dumbrell), pp. 91–131. Academic Press. (doi:10.1016/bs.aecr.2019.06.002)

11. Kohli BA, Rowe RJ. 2019 Beyond guilds: The promise of continuous traits for mammalian functional diversity. *Journal of Mammalogy* **100**, 285–298. (doi:10.1093/jmammal/gyz054)

12. Chase JM, Myers JA. 2011 Disentangling the importance of ecological niches from stochastic processes across scales. *Philosophical Transactions of the Royal Society B: Biological Sciences* **366**, 2351–2363. (doi:10.1098/rstb.2011.0063)

13. Zhou J, Ning D. 2017 Stochastic community assembly: Does it matter in microbial ecology? *Microbiology and Molecular Biology Reviews* **81**, 10.1128/mmbr.00002-17. (doi:10.1128/mmbr.00002-17)

14. Powell JR, Karunaratne S, Campbell CD, Yao H, Robinson L, Singh BK. 2015 Deterministic processes vary during community assembly for ecologically dissimilar taxa. *Nature Communications* **6**, 8444. (doi:10.1038/ncomms9444)

15. Mammola S, Carmona CP, Guillerme T, Cardoso P. 2021 Concepts and applications in functional diversity. *Functional Ecology* **35**, 1869–1885. (doi:10.1111/1365-2435.13882)

16. Pease AA, González-Díaz AA, Rodiles-Hernández R, Winemiller KO. 2012 Functional diversity and trait–environment relationships of stream fish assemblages in a large tropical catchment. *Freshwater Biology* **57**, 1060–1075. (doi:10.1111/j.1365-2427.2012.02768.x)

17. Ingram T, Shurin JB. 2009 Trait-based assembly and phylogenetic structure in northeast Pacific rockfish assemblages. *Ecology* **90**, 2444–2453. (doi:10.1890/08-1841.1)

18. Rincón-Díaz MP, Pittman SJ, Arismendi I, Heppell SS. 2018 Functional diversity metrics detect spatio-temporal changes in the fish communities of a Caribbean marine protected area. *Ecosphere* **9**, e02433. (doi:10.1002/ecs2.2433)

19. Pecuchet L, Törnroos A, Lindegren M. 2016 Patterns and drivers of fish community assembly in a large marine ecosystem. *Marine Ecology Progress Series* **546**, 239–248. (doi:10.3354/meps11613)

20. Henseler C, Nordström MC, Törnroos A, Snickars M, Pecuchet L, Lindegren M, Bonsdorff E. 2019 Coastal habitats and their importance for the diversity of benthic communities: A species- and trait-based approach. *Estuarine, Coastal and Shelf Science* **226**, 106272. (doi:10.1016/j.ecss.2019.106272)

21. Sgarlatta MP, Ramírez-Valdez A, Ladah LB, Calderón-Aguilera LE. 2023 Fish functional diversity is modulated by small-scale habitat complexity in a temperate ecosystem. *Hydrobiologia* **850**, 747–759. (doi:10.1007/s10750-022-05061-x)

22. Quirino BA, Lansac-Tôha FM, Thomaz SM, Heino J, Fugi R. 2021 Macrophyte stand complexity explains the functional α and β diversity of fish in a tropical river-floodplain. *Aquatic Sciences* **83**, 12. (doi:10.1007/s00027-020-00768-2)

23. McLean M, Stuart-Smith RD, Villéger S, Auber A, Edgar GJ, MacNeil MA, Loiseau N, Leprieur F, Mouillot D. 2021 Trait similarity in reef fish faunas across the world's oceans. *Proceedings of the National Academy of Sciences* **118**, e2012318118. (doi:10.1073/pnas.2012318118)

24. Vellend M. 2010 Conceptual synthesis in community ecology. *The Quarterly Review of Biology* **85**, 183–206. (doi:10.1086/652373)

25. Triantis KA *et al.* 2022 Deterministic assembly and anthropogenic extinctions drive

convergence of island bird communities. *Global Ecology and Biogeography* **31**, 1741–1755. (doi:10.1111/geb.13556)

26. Reeves JC, Moon BC, Benton MJ, Stubbs TL. 2021 Evolution of ecospace occupancy by Mesozoic marine tetrapods. *Palaeontology* **64**, 31–49. (doi:10.1111/pala.12508)

27. Ceballos G, Ehrlich PR, Barnosky AD, García A, Pringle RM, Palmer TM. 2015 Accelerated modern human–induced species losses: Entering the sixth mass extinction. *Science Advances* **1**, e1400253. (doi:10.1126/sciadv.1400253)

28. Sallan LC, Coates MI. 2010 End-Devonian extinction and a bottleneck in the early evolution of modern jawed vertebrates. *Proceedings of the National Academy of Sciences* **107**, 10131–10135. (doi:10.1073/pnas.0914000107)

29. McGhee GR, Clapham ME, Sheehan PM, Bottjer DJ, Droser ML. 2013 A new ecological-severity ranking of major Phanerozoic biodiversity crises. *Palaeogeography, Palaeoclimatology, Palaeoecology* **370**, 260–270. (doi:10.1016/j.palaeo.2012.12.019)

30. Beerling DJ, Chaloner WG, Woodward FI, Algeo TJ, Scheckler SE. 1998 Terrestrial-marine teleconnections in the Devonian: Links between the evolution of land plants, weathering processes, and marine anoxic events. *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences* **353**, 113–130. (doi:10.1098/rstb.1998.0195)

31. Brett CE, Walker SE. 2002 Predators and predation in Paleozoic marine environments. *Paleontological Society Papers.* **8**, 93–118. (doi:10.1017/S1089332600001078)

32. Copper P. 2011 100 million years of reef prosperity and collapse: Ordovician to Devonian interval. *The Paleontological Society Papers* **17**, 15–32. (doi:10.1017/S1089332600002424)

33. Trinajstic K, Briggs DEG, Long JA. 2021 The Gogo Formation Lagerstätte: A view of Australia's first great barrier reef. *Journal of the Geological Society* **179**, jgs2021-105. (doi:10.1144/jgs2021-105)

34. Cloutier R. 2013 Great Canadian Lagerstätten 4. The Devonian Miguasha biota (Québec): UNESCO world heritage site and a time capsule in the early history of vertebrates. *Geoscience Canada* **40**, 149–163.

35. Gess RW, Whitfield AK. 2020 Estuarine fish and tetrapod evolution: Insights from a Late Devonian (Famennian) Gondwanan estuarine lake and a southern African Holocene equivalent. *Biological Reviews* **95**, 865–888. (doi:10.1111/brv.12590)

36. Mouchet MA, Villéger S, Mason NWH, Mouillot D. 2010 Functional diversity measures: An overview of their redundancy and their ability to discriminate community assembly rules. *Functional Ecology* **24**, 867–876. (doi:10.1111/j.1365-2435.2010.01695.x)

37. Anderson PSL, Friedman M, Brazeau MD, Rayfield EJ. 2011 Initial radiation of jaws demonstrated stability despite faunal and environmental change. *Nature* **476**, 206–209. (doi:10.1038/nature10207)

38. Cloutier R, Loboziak S, Candilier A-M, Blieck A. 1996 Biostratigraphy of the Upper Devonian Escuminac Formation, eastern Québec, Canada: A comparative study based on miospores and fishes. *Review of Palaeobotany and Palynology* **93**, 191–215. (doi:10.1016/0034-6667(95)00126-3)

39. Cloutier R, Proust J-N, Tessier B. 2011 The Miguasha Fossil-Fish-Lagerstätte: a consequence of the Devonian land–sea interactions. *Palaeobio Palaeoenv* **91**, 293–323. (doi:10.1007/s12549-011-0058-0)

40. Chevrinais M, Jacquet C, Cloutier R. 2017 Early establishment of vertebrate trophic interactions: Food web structure in Middle to Late Devonian fish assemblages with exceptional fossilization. *Bulletin of Geosciences* 491–510. (doi:10.3140/bull.geosci.1651)

41. Mory AJ, Hocking RM. 2011 *Permian, carboniferous and upper devonian geology of the*

42. Bascompte J, Melián CJ, Sala E. 2005 Interaction strength combinations and the overfishing of a marine food web. *Proceedings of the National Academy of Sciences* **102**, 5443–5447. (doi:10.1073/pnas.0501562102)

43. Pérez-Matus A *et al.* 2017 Temperate rocky subtidal reef community reveals human impacts across the entire food web. *Marine Ecology Progress Series* **567**, 1–16. (doi:10.3354/meps12057)

44. Lira AS, Lucena-Frédou F, Lacerda CHF, Eduardo LN, Ferreira V, Frédou T, Ménard F, Angelini R, Le Loc'h F. 2022 Effect of fishing effort on the trophic functioning of tropical estuaries in Brazil. *Estuarine, Coastal and Shelf Science* **277**, 108040. (doi:10.1016/j.ecss.2022.108040)

45. Ferreira V, Le Loc'h F, Ménard F, Frédou T, Frédou FL. 2019 Composition of the fish fauna in a tropical estuary: The ecological guild approach. *Scientia Marina* **83**, 133. (doi:10.3989/scimar.04855.25A)

46. Cohen JE, Schittler DN, Raffaelli DG, Reuman DC. 2009 Food webs are more than the sum of their tritrophic parts. *Proceedings of the National Academy of Sciences* **106**, 22335–22340. (doi:10.1073/pnas.0910582106)

47. Hall SJ, Raffaelli D. 1991 Food-web patterns: Lessons from a species-rich web. *The Journal of Animal Ecology* **60**, 823. (doi:10.2307/5416)

48. Huxham M, Beaney S, Raffaelli D. 1996 Do Parasites Reduce the Chances of Triangulation in a Real Food Web? *Oikos* **76**, 284. (doi:10.2307/3546201)

49. Angelini R, De Morais RJ, Catella AC, Resende EK, Libralato S. 2013 Aquatic food webs of the oxbow lakes in the Pantanal: A new site for fisheries guaranteed by alternated control? *Ecological Modelling* **253**, 82–96. (doi:10.1016/j.ecolmodel.2013.01.001)

50. Resende EKD. 2000 Trophic structure of fish assemblages in the Lower Miranda River, Pantanal, Mato Grosso do Sul State, Brazil. *Revista brasileira de Biologia* **60**, 389–403. (doi:10.1590/S0034-71082000000300004)

51. Pereira RAC, de Resende EK. 1998 Peixes detritívoros da planície inundável do Rio Miranda, Pantanal, Mato Grosso do Sul, Brasil. *EMBRAPA-CPAP. Research Bulletin* **12**, 1–50.

52. de Resende EK, Pereira RAC. 1998 Peixes herbívoros da planície inundável do rio Miranda, Pantanal, Mato Grosso do Sul, Brasil. *EMBRAPA-CPAP. Research Bulletin* **10**, 1–24.

53. de Resende EK, Pereira RAC. 2000 Peixes onívoros da planície inundável do rio Miranda, Pantanal, Mato Grosso do Sul, Brasil. *EMBRAPA-CPAP. Research Bulletin* **16**, 1–60.

54. de Resende EK, Pereira RAC. 2000 Peixes insetívoros e zooplanctófagos da planície inundável do rio Miranda, Mato Grosso do Sul, Brasil. *EMBRAPA-CPAP. Research Bulletin* **17**, 1–42.

55. Ferreira FS, Duarte GS do V, Severo-Neto F, Froehlich O, Súarez YR. 2017 Survey of fish species from plateau streams of the Miranda River Basin in the Upper Paraguay River Region, Brazil. *Biota Neotropica* **17**, e20170344. (doi:10.1590/1676-0611-BN-2017-0344)

56. Severo-Neto F, Tencatt LFC, Costa-Pereira R, Tavares LER. 2015 Fishes from Baía da Medalha, southern Pantanal, Brazil: A 20 years review. *Biota Neotropica* **15**. (doi:10.1590/1676-06032015011614)

57. Costa-Pereira R, Severo-Neto F, Yule TS, Pereira APT. 2011 Fruit-eating fishes of Banara arguta (Salicaceae) in the Miranda River floodplain, Pantanal wetland. *Biota*

Northern Canning Basin, Western Australia: A field guide. Perth: Government of Western Australia.

Neotropica **11**, 373–376. (doi:10.1590/S1676-06032011000400033)

58. Ferreira FS, Duarte GS do V, Severo-Neto F, Froehlich O, Súarez YR. 2017 Survey of fish species from plateau streams of the Miranda River Basin in the Upper Paraguay River Region, Brazil. *Biota Neotropica* **17**, e20170344. (doi:10.1590/1676-0611-BN-2017-0344)

59. Growns I, Gehrke PC, Astles KL, Pollard DA. 2003 A comparison of fish assemblages associated with different riparian vegetation types in the Hawkesbury–Nepean River system. *Fisheries Management and Ecology* **10**, 209–220. (doi:10.1046/j.1365-2400.2003.00337.x)

60. Long JA, Trinajstic K. 2010 The Late Devonian Gogo Formation lägerstatte of Western Australia: Exceptional early vertebrate preservation and diversity. *Annual Review of Earth and Planetary Sciences* **38**, 255–279. (doi:10.1146/annurev-earth-040809-152416)

61. Stekhoven DJ. 2022 missForest: Nonparametric missing value imputation using Random Forest. **R package version 1.5**.

62. Boettiger C, Lang DT, Wainwright PC. 2012 rfishbase: Exploring, manipulating and visualizing FishBase data from R. *Journal of Fish Biology* **81**, 2030–2039. (doi:10.1111/j.1095-8649.2012.03464.x)

63. Gower JC, Legendre P. 1986 Metric and Euclidean properties of dissimilarity coefficients. *Journal of Classification* **3**, 5–48. (doi:10.1007/BF01896809)

64. de Bello F, Botta-Dukát Z, Lepš J, Fibich P. 2021 Towards a more balanced combination of multiple traits when computing functional differences between species. *Methods in Ecology and Evolution* **12**, 443–448. (doi:10.1111/2041-210X.13537)

65. Magneville C *et al.* 2022 mFD: An R package to compute and illustrate the multiple facets of functional diversity. *Ecography* **2022**. (doi:10.1111/ecog.05904)

66. Mouillot D, Graham NAJ, Villéger S, Mason NWH, Bellwood DR. 2013 A functional approach reveals community responses to disturbances. *Trends in Ecology & Evolution* **28**. (doi: 10.1016/j.tree.2012.10.004)

67. Magneville C, Loiseau N, Albouy C, Casajus, N. Claverie T. 2022 mFD: An R package to compute and illustrate the multiple facets of functional diversity. *Ecography* **2022**. (doi: 10.1111/ecog.05904)

68. Ricklefs RE, Miles DB. 1994 Ecological and evolutionary inferences from morphology: An ecological perspective. In *Ecological morphology: Integrative organismal biology*, pp. 13–41. The University of Chicago Press.

69. Hurtado-Materon MA, Murillo-García OE. 2023 An integrative approach to understanding diversity patterns and assemblage rules in Neotropical bats. *Scientific Reports* **13**. (doi: 10.1038/s41598-023-35100-z)

70. Blonder B, Morrow CB, Brown S, Butruille G, Chen D, Laini A, Harris DJ. 2023 hypervolume: High dimensional geometry, set operations, projection, and inference using kernel density estimation, support vector machines, and convex hulls. *R package version 3.1.3 ,* <https://CRAN.R-project.org/package=hypervolume>.

71. Baselga. 2012 The relationship between species replacement, dissimilarity derived from nestedness, and nestedness. *Global ecology and biogeography* **21**, 463–468. (doi:10.1016/0169-5347(96)10048-3)

72. Aldridge RJ, Purnell MA. 1996 The conodont controversies. *Trends in Ecology & Evolution* **11**, 463–468. (doi:10.1016/0169-5347(96)10048-3)

73. Aldridge RJ, Donoghue PCJ. 1998 Conodonts: A sister group to hagfishes? In *The Biology of Hagfishes* (eds JM Jørgensen, JP Lomholt, RE Weber, H Malte), pp. 15–31. Dordrecht: Springer Netherlands. (doi:10.1007/978-94-011-5834-3_2)

74. McCoy VE, Lamsdell JC, Poschmann M, Anderson RP, Briggs DEG. 2015 All the better to see you with: Eyes and claws reveal the evolution of divergent ecological roles in giant

pterygotid eurypterids. *Biology Letters* **11**, 20150564. (doi:10.1098/rsbl.2015.0564)

75. Pimiento C, Leprieur F, Silvestro D, Lefcheck JS, Albouy C, Rasher DB, Davis M,
Svenning J-C, Griffin JN. 2020 Functional diversity of marine megafauna in the
Anthropocene. *Science Advances* **6**, eaay7650. (doi:10.1126/sciadv.aay7650)

76. Le Bagousse-Pinguet Y *et al.* 2021 Functional rarity and evenness are key facets of
biodiversity to boost multifunctionality. *Proceedings of the National Academy of*
Sciences **118**, e2019355118. (doi:10.1073/pnas.2019355118)

77. Schleuning M *et al.* 2020 Trait-based assessments of climate-change impacts on
interacting species. *Trends in Ecology & Evolution* **35**, 319-328.
(doi:10.1016/j.tree.2019.12.010)

78. Brind'Amour A, Boisclair D, Dray S, Legendre P. 2011 Relationships between species
feeding traits and environmental conditions in fish communities: A three-matrix
approach. *Ecological Applications* **21**, 363–377. (doi:10.1890/09-2178.1)

79. Moyle PB, Cech JJ. 2004 *Fishes: An introduction to ichthyology*. 5th edition. New
Jersey, USA: Pearson Prentice Hall.

80. Jobbins M, Rücklin M, Sánchez Villagra MR, Lelièvre H, Grogan E, Szrek P, Klug C.
2024 Extreme lower jaw elongation in a placoderm reflects high disparity and modularity
in early vertebrate evolution. *Royal Society Open Science* **11**, 231747.
(doi:10.1098/rsos.231747)

81. Janvier P. 1996 *Early vertebrates*. Oxford, U.K.: Clarendon Press.

82. Ehlman SM, Trimmer PC, Sih A. 2019 Prey responses to exotic predators: Effects of old
risks and new cues. *The American Naturalist* **193**, 575–587. (doi:10.1086/702252)

83. Schakmann M, Korsmeyer KE. 2023 Fish swimming mode and body morphology affect
the energetics of swimming in a wave-surge water flow. *Journal of Experimental Biology*
226, jeb244739. (doi:10.1242/jeb.244739)

84. Kelley JL, Fitzpatrick JL, Merilaita S. 2013 Spots and stripes: Ecology and colour pattern
evolution in butterflyfishes. *Proceedings of the Royal Society B: Biological Sciences* **280**,
20122730. (doi:10.1098/rspb.2012.2730)

85. Wilson MVH, Caldwell MW. 1993 New Silurian and Devonian fork-tailed 'thelodonts'
are jawless vertebrates with stomachs and deep bodies. *Nature* **361**, 442–444.
(doi:10.1038/361442a0)

86. Girard C, Renaud S. 2012 Disparity changes in 370 Ma Devonian fossils: The signature
of ecological dynamics? *PLOS ONE* **7**, e36230. (doi:10.1371/journal.pone.0036230)

87. Friedman M, Sallan LC. 2012 Five hundred million years of extinction and recovery: A
phanerozoic survey of large-scale diversity patterns in fishes. *Palaeontology* **55**, 707–
742. (doi:10.1111/j.1475-4983.2012.01165.x)

88. Henderson S, Dunne EM, Fasey SA, Giles S. 2023 The early diversification of ray-finned
fishes (Actinopterygii): Hypotheses, challenges and future prospects. *Biological Reviews*
98, 284–315. (doi:10.1111/brv.12907)

89. Richardson LE, Graham NAJ, Pratchett MS, Hoey AS. 2017 Structural complexity
mediates functional structure of reef fish assemblages among coral habitats.
Environmental Biology of Fishes **100**, 193–207. (doi:10.1007/s10641-016-0571-0)

90. Majchrzyk A, Jakubowicz M, Berkowski B, Król JJ, Zatoń M, Zapalski MK. 2024
Modern-type reef in ancient time - Palaeoecology of a Middle Devonian coral
community from Madène el Mrakib (Anti-Atlas, Morocco). *Palaeogeography,*
Palaeoclimatology, Palaeoecology **633**, 111876. (doi:10.1016/j.palaeo.2023.111876)

91. Benton MJ, Emerson BC. 2007 How did life become so diverse? The dynamics of
diversification according to the fossil record and molecular phylogenetics. *Palaeontology*
50, 23–40. (doi:10.1111/j.1475-4983.2006.00612.x)

92. Hill JJ, Puttick MN, Stubbs TL, Rayfield EJ, Donoghue PCJ. 2018 Evolution of jaw

disparity in fishes. *Palaeontology* **61**, 847–854. (doi:10.1111/pala.12371)

93. He S *et al.* 2023 Genetic innovations underpin morphological diversity and radiation in
teleost fish. (doi:10.21203/rs.3.rs-3148441/v1)

94. Witten PE, Hall BK. 2015 Teleost skeletal plasticity: Modulation, adaptation, and
remodelling. *Copeia* **103**, 727–739. (doi:10.1643/CG-14-140)

95. Long JA, Choo B, Clement A. 2018 The evolution of fishes through geological time. In
Evolution and development of fishes (eds Z Johanson, C Underwood, M Richter), pp. 3–
29. Cambridge University Press. (doi:10.1017/9781316832172.002)

96. Peres-Neto PR. 2004 Patterns in the co-occurrence of fish species in streams: The role of
site suitability, morphology and phylogeny versus species interactions. *Oecologia* **140**,
352–360. (doi:10.1007/s00442-004-1578-3)

97. Grossman GD, Ratajczak Jr. RE, Crawford M, Freeman MC. 1998 Assemblage
organization in stream fishes: Effects of environmental variation and interspecific
interactions. *Ecological Monographs* **68**, 395–420. (doi:10.1890/0012-
9615(1998)068[0395:AOISFE]2.0.CO;2)

98. Gratwicke B, Speight MR. 2005 The relationship between fish species richness,
abundance and habitat complexity in a range of shallow tropical marine habitats. *Journal*
of Fish Biology **66**, 650–667. (doi:10.1111/j.0022-1112.2005.00629.x)

99. Passos CVB, Fabré NN, Malhado ACM, Batista VS, Ladle RJ. 2016 Estuarization
increases functional diversity of demersal fish assemblages in tropical coastal
ecosystems. *Journal of Fish Biology* **89**, 847–862. (doi:10.1111/jfb.13029)

100.Potter IC, Tweedley JR, Elliott M, Whitfield AK. 2015 The ways in which fish use
estuaries: A refinement and expansion of the guild approach. *Fish and Fisheries* **16**, 230–
239. (doi:10.1111/faf.12050)

101.Cardoso P, Pekár S, Jocqué R, Coddington JA. 2011 Global Patterns of Guild
Composition and Functional Diversity of Spiders. *PLOS ONE* **6**, e21710.
(doi:10.1371/journal.pone.0021710)

102.Mouillot D *et al.* 2014 Functional over-redundancy and high functional vulnerability in
global fish faunas on tropical reefs | PNAS. *PNAS* **111**, 13757–13762.
(doi:10.1073/pnas.1317625111)

103.Pigot AL, Trisos CH, Tobias JA. 2016 Functional traits reveal the expansion and packing
of ecological niche space underlying an elevational diversity gradient in passerine birds.
Proceedings of the Royal Society B: Biological Sciences **283**, 20152013.
(doi:10.1098/rspb.2015.2013)

104.Myers EMV, Anderson MJ, Liggins L, Harvey ES, Roberts CD, Eme D. 2021 High
functional diversity in deep-sea fish communities and increasing intraspecific trait
variation with increasing latitude. *Ecology and Evolution* **11**, 10600–10612.
(doi:10.1002/ece3.7871)

105.Stuart-Smith RD *et al.* 2013 Integrating abundance and functional traits reveals new
global hotspots of fish diversity. *Nature* **501**, 539–542. (doi:10.1038/nature12529)

106.Berke SK, Jablonski D, Krug AZ, Valentine JW. 2014 Origination and immigration drive
latitudinal gradients in marine functional diversity. *PLOS ONE* **9**, e101494.
(doi:10.1371/journal.pone.0101494)

107.Schumm M, Edie SM, Collins KS, Gómez-Bahamón V, Supriya K, White AE, Price TD,
Jablonski D. 2019 Common latitudinal gradients in functional richness and functional
evenness across marine and terrestrial systems. *Proceedings of the Royal Society B:*
Biological Sciences **286**, 20190745. (doi:10.1098/rspb.2019.0745)

108.Jarzyna MA, Quintero I, Jetz W. 2021 Global functional and phylogenetic structure of
avian assemblages across elevation and latitude. *Ecology Letters* **24**, 196–207.
(doi:10.1111/ele.13631)

109. Lamanna C *et al.* 2014 Functional trait space and the latitudinal diversity gradient. *Proceedings of the National Academy of Sciences* **111**, 13745–13750. (doi:10.1073/pnas.1317722111)

110. Hillebrand H. 2004 On the generality of the latitudinal diversity gradient. *The American Naturalist* **163**, 192–211. (doi:10.1086/381004)

111. Ford BM. 2018 Latitudinal gradient of dispersal and niche processes mediating neutral assembly of marine fish communities. *Marine biology* **165**. (doi: 10.1007/s00227-018-3356-5)

112. Villéger S, Novack-Gottshall PM, Mouillot D. 2011 The multidimensionality of the niche reveals functional diversity changes in benthic marine biotas across geological time: long-term functional diversity changes. *Ecology Letters* **14**, 561-568. (doi: 10.1111/j.1461-0248.2011.01618.x)

113. McKitrick MC. 1993 Phylogenetic constraint in evolutionary theory: Has it any explanatory power? *Annual Review of Ecology and Systematics*, 307–330.

114. Champagnat N, Ferrière R, Méléard S. 2006 Unifying evolutionary dynamics: From individual stochastic processes to macroscopic models. *Theoretical Population Biology* **69**, 297–321. (doi:10.1016/j.tpb.2005.10.004)

115. Fritz SA, Schnitzler J, Eronen JT, Hof C, Böhning-Gaese K, Graham CH. 2013 Diversity in time and space: Wanted dead and alive. *Trends in Ecology & Evolution* **28**, 509–516. (doi:10.1016/j.tree.2013.05.004)