Diel and seasonal rhythmicity in activity and corticosterone in an Arctic migratory

herbivore: A multifaceted approach

Margje E. de Jong * [1,3], Annabel Slettenhaar [2,3], Rienk W. Fokkema [4,5], Marion Leh [1], Elisabeth

Barnreiter [1], Larry Griffin [6], Maarten J.J.E. Loonen [3], Eva Millesi [1], Børge Moe [7], Isabella B.R.

Scheiber [1]

*corresponding author

(1) Dept. of Behavioral and Cognitive Biology, University of Vienna, Djerassiplatz 1, 1030 Vienna,

Austria

(2) Faculty of Biosciences and Aquaculture, Nord University, Kongens Gate 42, 7713 Steinkjer,

Norway

(3) Arctic Centre, Aweg 30, 9718 CW Groningen, The Netherlands

(4) Conservation Ecology Group, Groningen Institute for Evolutionary Life Sciences (GELIFES),

University of Groningen, Nijenborgh 7, 9747 AG, Groningen, The Netherlands

(5) BirdEyes, Centre for Global Ecological Change at the Faculties of Science & Engineering and

Campus Fryslân, University of Groningen, Zaailand 110, 8911 BN Leeuwarden, The Netherlands

(6) ECO-LG Ltd, Crooks House, Mabie, DG2 8EY, United Kingdom

(7) Norwegian Institute for Nature Research (NINA), PO Box 5685 Torgard, 7485 Trondheim,

Norway

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Abstract

Birds that migrate from temperate areas to the Arctic for breeding lose their strongest *Zeitgeber* of circadian organization when they cross the Arctic circle in spring – the 24h geophysical light-dark cycle. Under continuous daylight various behavioural and physiological patterns have been detected in both free-ranging and laboratory animals. To better understand the evolution of plasticity in circadian clocks, it is essential to study behavioural and physiological rhythmicity in the context of a species' ecology. Employing a multifaceted approach, which included wildlife cameras, accelerometers, and non-invasive sampling of hormone metabolites, we investigated activity patterns and corticosterone rhythmicity in a migratory herbivore, the Svalbard barnacle goose (*Branta leucopsis*), during its Arctic breeding season. We detected that females showed a combination of both ultradian and diel rhythmicity in incubation recesses and sleep, respectively. In both parents, rhythms in activity continued over the period, when goslings were present. During moult, many geese aligned activity with the tidal rhythm. Barnacle geese showed weak diel rhythmicity in excreted corticosterone metabolites. This suggests that while Arctic geese may adopt an alternative *Zeitgeber* during the Arctic summer to keep a diel rhythm, ultradian rhythmicity is crucial and geese are able to tune their rhythms flexibly to environmental conditions.

Keywords

Camera trap, GPS transmitter, circadian rhythmicity, hormones, continuous daylight, polar

Introduction

Various biological processes occur rhythmically and are restricted to specific times of the day or season [1, 2], which in many species is maintained by using photoperiod, the environmental period of daylight, as a *Zeitgeber* [3]. Upholding these rhythms in polar regions is challenging insofar as above the Arctic and below the Antarctic circle (66°N and S) natural light is completely absent in winter but continuously present during the summer [4]. Nevertheless, various Arctic species are able to retain endogenous and behavioural rhythms [reviewed in 1, 5].

Arctic vertebrates show plasticity in rhythmicity in behavioural activity within individuals, between individuals and between species. Patterns may vary seasonally within individuals, *e.g.* driven by annual changes of day length [5, 6]. For example, the Svalbard rock ptarmigan (*Lagopus muta hyperborea*), the sole permanent resident bird on Svalbard, exhibits ultradian activity patterns with periodic feeding during polar summer and winter days. In contrast, during spring and fall its activity patterns are diurnal and feeding mainly occurs within daylight hours [7]. Between individual differences exist, for example, in migratory, bi-parentally breeding shorebirds, where different conspecific pairs may show different incubation rhythms even in pairs which breed in the same area [5, 8]. Furthermore, Arctic vertebrate species are highly variable in their behavioural activity with some species (i) being completely *arrhythmic,* (ii) showing *circadian or diel rhythmicity, i.e.* displaying a recurring rhythm of approximately 24-hours even in the absence of light fluctuations, (iii) showing *ultradian cycles* of recurrent periods of foraging and resting bouts less than 24-hours, or (iv) showing *free running rhythmicity, i.e.* not synchronized with environmental time cues [1, 5]. For herbivore species, Bloch *et al*. suggested that ultradian activity patterns in polar regions are linked with feeding behaviour and digestive processes, because even at lower latitudes continuous activity is common in herbivores [6]. The diversity in behavioural responses to continuous daylight underline the plasticity of the circadian system and indicate the importance of life history and social contexts [5, 9].

Behavioural and physiological processes are regulated through the rhythmic excretion of hormones by an endogenous network of one or several 'Master' clocks, which then coordinate and synchronise more peripheral clocks [reviewed in 1, 2]. Crucial for tracking environmental cycles is

the neurohormone melatonin. Its diel secretion pattern, namely an extended peak at night and basal

secretion during the day, provides the paramount hormonal signal transducing day length for

peripheral clocks [3]. Melatonin secretion drives enduring changes in the hypothalamic-pituitary

adrenal axis (HPA), where low levels of melatonin culminate in the increased secretion of the adrenal

glucocorticoids. *i.e.* cortisol in mammals and corticosterone (CORT) in birds [1, 10, 11].

Glucocorticoids are involved in correlating peripheral clocks [12], are best known as one of the two

major stress response systems [13], and interrelate with feeding, modulation of energy storage and

mobilisation as well as activity [10]. They display robust diel rhythmicity in latitudes with year-round

day-night cycles [14-16, but see 17]. In temperate zones, the diel rhythmic pattern of baseline CORT

correlates with activity with low levels during inactive phases followed by a rise just before activity

begins, to remain high as long as birds are active [10]. In polar species with 24 hours of continuous

natural light during the summer, findings of a diel CORT rhythm are ambivalent, however. Common

murres (*Uria aalge*) maintain a diel CORT rhythm in the polar summer [18]. Contrary, the link

between activity, time of day and corticosterone was not evident in thick-billed murres (*U. lomvia*), a

species, which lacks the diel CORT rhythm [10]. Other such species without a diel CORT rhythm are

Adélie penguins (*Pygoscelis adeliae*) [4] and common eiders (*Somateria mollissima*) [19]. This

arrhythmicity was attributed to the suppression of melatonin secretion when photoperiod as *Zeitgeber*

is absent [5, 10, 19] and in common eiders it was linked to CORT interfering with the need to constant

forage [5], while in thick-billed murres it was ascribed to a stable modulation of energy storage and

mobilisation [10].

Many bird species, which show rapid migration to their Arctic breeding grounds, experience

large changes in light conditions en route, before they arrive in the polar regions with 24-hours of

continuous daylight. Latitudinal migrants may benefit from longer photoperiods at higher latitudes as

for diurnal animals daily activity and foraging time can be increased [20]. Nevertheless, such quick

fluctuations in light conditions present a significant challenge for circadian physiology [21-23]. There

is a surprising variation in the response of migrants to continuous daylight [*e.g.* 5, 8], despite that

changing to a different rhythm can be costly [24]. To increase our understanding of the evolution of

plasticity in circadian clocks in migratory birds, it is necessary to study rhythmicity in behaviour and

physiology in relation to the ecology of the species [*e.g.* 6, 8, 9].

Here we aim to quantify behaviour- and CORT-based rhythmicity in the barnacle goose, a

migratory herbivore, during their breeding season in the Arctic using a multifaceted approach. Earlier

findings on rhythmicity in barnacle goose behaviour and physiology are somewhat ambiguous. A

recent study showed that barnacle geese lost diel rhythmicity in body temperature once they

encountered continuous daylight during their migration to the Arctic breeding sites [23]. During

incubation in the Arctic, however, a diel rhythm in incubation recesses was described earlier [25-27].

In addition, we showed that young barnacle geese retained a diel, albeit time-shifted, pattern of

corticosterone metabolites (CORTm) determined from droppings, over one 24-hour period in their

first summer [28]. Here, we investigated potential diel and seasonal rhythmicity in activity and daily

excretion patterns of CORTm, using three different approaches; (i) activity and sleep, *i.e.* sleep

posture of females indicative of sleep [29], during incubation were determined from wildlife camera

photos placed in the vicinity of the nest, (ii) activity across the breeding season of females and males

was assessed using data from accelerometers (ACC), and (iii) CORTm concentration was determined

from individually assigned droppings. Contrary to mammalian herbivores, where digestive processes

dictate ultradian activity patterns [30, 31], barnacle geese do not have complex digestive systems.

Their summer food retention time is increased by interrupting feeding with longer loafing spells [30],

however. We expect that barnacle geese may feed around the clock [*sensu* 23], which has been

suggested to be a possible adaption of Arctic herbivores that allows for optimally utilizing the peak in

food quality during the short summer [32]. As we predict ultradian rhythmicity in activity then to be

important, thus, we expect to find no or only a weak diel rhythm of CORTm. In contrast, if barnacle

geese strongly depend on an evolutionary-based endogenous clock to schedule their biology or

respond directly to diel external cues, we expect diel rhythmicity in activity and CORTm throughout

the breeding season, similarly to some other polar species [*e.g.* 18, 33, 34].

Materials and Methods

1. Study site and species

We collected data during the summers of 2020 – 2022 in a barnacle goose population nesting on islets in Kongsfjorden, Svalbard. Except on days with inclement weather when boating was not possible or polar bears (*Ursus maritimus)* were present, we monitored nests every other day during incubation and hatch on the two main breeding islands of the study area, Storholmen and Prins Heinrichøya from June to the beginning of July. We determined the exact nest location using a handheld GPS (Garmin GPSmap 64S), noted clutch size and identified parents by their individually recognizable engraved plastic leg rings when a researcher approached the nest [35]. To diminish disturbance, we got information on possible nests of geese with transmitters (see below) on other islands (Midtholmen, Juttaholmen, Observasjonsholmen) from colleagues who worked there regularly. Egg laying and incubation in barnacle geese takes approximately 29 days [36]. After hatching, many geese raise their goslings in or in direct vicinity of the village Ny-Ålesund (78°55′30″N 11°55′20″E) [37, 38]. Here, we observed geese at least twice-daily using binoculars or spotting scopes to identify individuals by their colour rings, to establish family size and record feather moult. From the first day of moult, the complete moulting process lasts 35 to 40 days on average, with a flightless period of ca. 25 days [39]. We used the flightless period from the first observation of loss of all flight feathers until 25 days later in our analyses (from now on called 'moult').

In 2020, we fitted one individual of 24 barnacle goose pairs (½ males and ½ females, sex based on cloacal inspection [35]) with black coloured OrniTrack-NL40 3G transmitter neckbands (Ornitela, UAB, Lithuania. Mass ~20 gram, height 21mm, diameter 38-40 mm). We caught ten geese on or near their nest on the islands Storholmen or Prins Heinrichøya during late incubation using two methods: (1) using an extendable pole with a small noose in fishing wire at one end, which was slid around the neck, quickly capturing the goose, or (2) by catching the geese by hand, which was mainly possible for aggressive males that came close. All remaining geese where caught during annually performed catches, during which flightless moulting geese are driven into corrals [30]. The 24 individuals were chosen following a suite of criteria, including that it would be desirable if both pair partners were

167   marked with unique colour rings previously (for details of criteria see [40]). During incubation, we

168   deployed three additional transmitters in 2021 and four in 2022.

2. Activity during incubation and across the breeding season

I.   Incubation patterns from wildlife camera photos

To investigate rhythmicity over the period of incubation, we set up wildlife cameras (Usogood TC30

Trail Camera) from 9/6/2021 to 16/7/2021 near nests of incubating geese, which were fitted with GPS

transmitters in 2020 (n = 15). Cameras, set in time-lapse mode, took two pictures every five minutes.

We ultimately retrieved the cameras either after hatching or when a nest was predated or abandoned

[40].

We analysed pictures with Timelapse2 Image Analyser (version 2.2.4.3, [41]). We quantified

whether the female was (i) sitting on the nest, (ii) in 'sleep posture', *i.e.* head resting on back with

often beak tucked under wing [29], (iii) standing right next to the nest or (iv) absent from the nest (n

photos = 110739). For further analyses on nest recesses, we pooled behaviours (iii) and (iv) as *active*

and behaviours (i) and (ii) as *inactive*. For the analyses of 'sleep', we contrasted being in the sleep

posture with the other behaviours. Males sometimes nest sit [40], but here we focussed on females

only. Instances, where males nest sat, while females were on incubation recesses, were classified as

(iv) absent from nest.

In total, wildlife cameras supplied data from 2 hours to 20 days. We only used data which

exceeded the recommended minimum duration of ten days for biological rhythm analyses [42, 43].

We could assess activity, *i.e.* incubation recesses, and sleep patterns, from photos taken by the

wildlife cameras of 11 out of 15 females. The other four cases were discarded, because data were

insufficient due to nest (n = 2) or camera failure (n = 2).

II.   Seasonal activity from transmitter accelerometer data

We used ACC data collected by the transmitters during the summers of 2021 and 2022 (2021: females

n = 8, males n = 9; 2022: females n = 9, males n = 9; same individuals across 2021 and 2022: n = 12).

When geese arrived within a geofence, *i.e.* a virtual boundary defined around a specific real-world

geographic area, that was put around their breeding area (rectangle latitude/longitude: top-left; 79.161

/ 10.95, bottom-right; 78.737 / 13.131), the transmitters were set to record a GPS-fix every 15 min

when battery voltage was 75-100%, 30 min at 50-74%, 60 min at 25-49% or 240 min at voltages

lower than 25%. Immediately after each GPS-fix, a 2-sec ACC burst was taken at a frequency of 20

Hz. Gravitational acceleration was measured in unit g/1000 in the three spatial axes [44].

We used ACC data to identify activity and inactivity of individual geese during the breeding

season [45, 46]. For each goose, we checked the number of bursts taken during each GPS-fix to

ensure a complete dataset. Then, we calculated the vectorial sum of dynamic body acceleration

(VeDBA) from the ACC data, which is a proxy for energy expenditure [47]. For this, we first

calculated static acceleration, *i.e.* the average raw measured acceleration for each dimension (x, y, z)

within the bursts. Second, we subtracted the static acceleration from the raw data for each dimension,

thus getting dynamic acceleration. Last, we calculated the vectorial sum of dynamic body acceleration

of each burst by taking the square root of the summed dynamic accelerations of each dimension [47].

Following [46], we created probability density histograms to identify peaks for activity and inactivity

and used the *mix* function in the R-package *mixdist* to decompose the distribution into two gamma

distribution components for active and inactive behaviour [48]. We found the threshold between the

active and inactive behaviour distributions by calculating the intersection point between the two [Fig.

S1, 45, 46].

In addition, we estimated the nesting period based on the method described by [44] using

VeDBA and GPS data. Nesting is defined as the entire period of egg laying, nest building, incubation,

and hatching. For females, we took daily median VeDBA < 1 for motionless days, as this

corresponded best with observed periods of nesting (n = 14: nesting period or part of the nesting

period could be estimated, n = 3: nesting period could not be estimated due to transmitter attachment

during hatching or when the goose was likely not breeding).

Most males, on the other hand, were never motionless during nesting (n = 14). Four males,

however, did show motionlessness. The immobility of these males could plausibly be used to

establish the nesting period. In addition, for nine males we established nesting period based on known

nest location. For six males we could not establish nesting. In total, for 14 geese (males and females)

we obtained both an observed nest location and an estimated nest location based on the transmitter

data; the distance between these two was on average 2.37 meters (SD = 0.98, range 0.85 – 3.67,

excluding an outlier of 25.45 meters where the handheld GPS was likely turned on too late).

Later breeding stages, *i.e.* when geese had goslings and were moulting, were based on

observations (see above). For 13 geese we obtained data on the period with goslings, for 11 geese

when they were moulting and for 3 geese that had goslings while moulting ('goslings & moult'

period). Since these different breeding stages pose different demands on the geese [30], we tested for

rhythmicity within these three periods. Once again, we only used data, which exceeded the

recommended minimum duration of ten days for biological rhythm analyses. Therefore, we excluded

four nesting periods, five gosling periods, one gosling & moult period and one moult period. We

pooled the remaining two goslings & moult periods with the moult period data to increase sample

size. For the gosling periods used in the analyses, goslings were observed until they were on average

27 days old. Because of limited sample size of geese with goslings due to high polar bear egg

predation and Arctic fox (*Vulpes lagopus*) gosling predation, we did not take gosling age into account

in our analyses.

3. Sample collection and corticosterone metabolite assay

To investigate diel rhythmicity of CORTm excretion, we collected droppings (faeces) of ringed geese

in the village of Ny-Ålesund from 1/7/2020 to 13/8/2020 (n = 349 droppings. 26 individuals; 14

females, 12 males) and 16/6/2021 to 31/7/2021 (n = 333 droppings. 52 individuals; 28 females, 24

males). Of 17 individuals we collected samples in both years. We attempted to collect a minimum of

three samples [49] per 3-hour time intervals, *i.e.* from 0:00 – 2:59, 3:00 – 5:59, …, 21:00 – 23:59 and

intended to cover one complete 24-hour cycle every week for each individual. This was not always

possible, as pairs, which lost their offspring, sometimes left the area and could not be located.

Logistically, collecting samples at a finer temporal scale from specific individuals proved not

possible. Therefore, we were able to study diel patterns in CORTm only and could not determine

possible ultradian patterns or changes in rhythmicity over the season. Only unambiguously assigned

samples were collected and frozen at -20°C within 1 h (for methodology on how to collect droppings,

see electronic supplementary material). Droppings were shipped frozen to the Dept. of Behavioural

and Cognitive Biology, University of Vienna, Austria, for analyses.

Dropping samples (n = 679) for determining CORTm were quantified using an enzyme

immunoassay validated for barnacle geese [50] and applied successfully in previous studies [*e.g.* 28].

Concentration limits ranged from 7.49 to 320.90 ng CORTm/g droppings (median concentration

32.13 ng/g) in 2020 and from 1.02 to 583.65 ng CORTm/g droppings (median concentration 28.56

ng/g) in 2021. Determined from homogenized pool samples, intra- and inter-assay coefficients of

variation (% CV) were 11.91% (< 15%) and 13.03% (< 25%), respectively.

4.   Statistical analyses

All analyses were performed in R version 4.4.0 (The R Foundation for Statistical Computing 2024).

I.   Rhythmicity in activity

We plotted actograms to visualize overall activity patterns during incubation, as assessed from

wildlife camera data, and across the entire breeding season as determined from transmitter data

(*ggetho* package [51]). Actograms, which show rhythmicity or a lack thereof, were double-plotted to

facilitate inspection. We tested for periodicity in actograms using Lomb-Scargle periodogram (LSP)

analysis [*periodogram* function, *zeitgebr* package: 51, 52] during specific periods: *i.e.* the incubation

period for wildlife camera data to the point when goslings hatched, and the nesting, gosling rearing

and moult periods for the transmitter data. The LSP analysis was very suitable to discover periodicity

in our dataset, because it can handle unequally sampled time-series with missing data [52].

Periodicity, *i.e.* the regular recurrence of behaviour or an event over fixed, equal time intervals, is a

common but not universal trait of rhythmicity, which in itself emphasizes the repetitive pattern of

changes rather than precise timing [53]. We scanned the data at an oversampling rate of 100 to

distinguish periodicity between 1 and 18 hours to focus on ultradian rhythmicity, and between 18 and

36 hours to focus on diel rhythmicity [following 33, 43, 54]. Data were re-sampled every 5 minutes in

case of data from nest cameras and every 15 minutes for transmitter data. We used the *find_peaks*

function from the *zeitgebr* package to extract significant peaks ($p < 0.05$). Time-series data can

contain two separate peaks, which may point to a true combination of ultradian and diel rhythmicity

or it can be a data artefact [55]. We followed the methods of [54] to distinguish between these two: if

we found two peaks in the normalized power for the same individual with an 18 h window we

rejected the smaller peak if it was less than one-third the height of the larger one.

We investigated possible differences in ultradian and diel peak periods between breeding stages

(nesting, goslings and moult) using linear mixed-effects models (*lme* from the package *nlme,*[56, 57])

with peak period in hours as the response variable and breeding stage (categorical) as a predictor

variable. We controlled for the factor sex (categorical: female, male), the interaction between sex and

breeding stage and year (categorical). We accounted for repeated measures by adding ID as a random

effect. We used an automated model selection using the function *dredge* (package *MuMIn*) and AICc

[58]. We give the 95% confidence set of models in the electronic supplementary material [59] and

information including 85% confidence intervals for the full model in the text [60]. The 85% interval is

consistent with how variables are selected using AIC. When present in the top model, we computed

estimated marginal means for specific factors and comparisons among levels [package *emmeans*, 61].

II.   Diel patterns of immune-reactive corticosterone metabolites

To investigate the association between predictor variables and CORTm concentration over the day,

we fitted linear fixed-effects models (*lme*). We log-transformed CORTm concentration to adhere to

model assumptions. The predictor variables in all models include time of day (continuous) and

controlled for year (categorical), day of the year (continuous) and sex (categorical). As time of day is

a circular variable, we changed it into two linear variables by first transforming hour of day to radians

and then calculating the sine and cosine of those radians. We included sine and cosine as continuous

predictor variables in our models [18, 62]. To account for repeated measures of the same individuals,

we added individual identity as random factor in all models. A more complex random slope model

could not be explored, because this fitted mixed model was singular. As above, we used model

selection using the function *dredge* (package *MuMIn*) and AICc and give the same model information.

Results

5. Rhythmicity in activity

I. Rhythmicity during incubation

All females showed rhythmicity in incubation recesses (Figure 1A, for actograms of all females see

electronic supplementary material Figure S2). Ten females showed a combination of both ultradian

and diel rhythmicity while one female showed ultradian rhythmicity only (electronic supplementary

material Table S1, Figure S3). The mean ultradian period in incubation recesses was 3.21 h (95% CI:

2.41, 3.89) and the mean diel period was 24.03 h (95% CI: 23.73, 24.38). Most incubation recesses

occurred during the conventional day (Figure 1B). In addition, nine females had both ultradian and

diel rhythmicity when resting in a sleep posture (Figure 1C, for actograms of all females see

electronic supplementary material Figure S4), while two females showed diel rhythmicity only

(electronic supplementary material Table S2, Figures S5). The mean peak ultradian period was 8.17 h

(95% CI: 5.14,10.64) and the diel period was 24.96 h (95% CI: 23.57,1 26.12). Overall, geese were

more often observed in the sleeping posture during conventional night time (Figure 1D).

    II.   Seasonal rhythmicity in activity

We detected rhythmicity in all geese over the course of the entire breeding season (for examples of

actograms of two individuals see electronic supplementary material Figure S6). With the exception of

three females, that showed no rhythmicity in activity patterns during nesting in 2022, most geese

showed diel rhythmicity or a combination of ultradian and diel rhythmicity during nesting and when

they had goslings (Figure 2 left and centre panel). Furthermore, we also detected either ultradian

rhythmicity only or a combination of ultradian and diel rhythmicity during moult (Figure 2 right

panel). We found evidence that breeding stage correlated with ultradian peak rhythmicity, as this

variable was present in all models within the 95% confidence set (Table 1A, electronic supplementary

material Table S3). The top model included breeding stage only (Table 1A) and post-hoc testing

revealed that the ultradian peak period during moult was significantly higher than when geese nested

or had goslings (Figure 3, goslings – moult; estimate = -6.05, SE = 1.75, p = 0.015, goslings – nesting;

estimate = 0.35, SE = 1.80, p = 0.9794, moult – nesting; estimate = 6.40, SE = 1.49, p = 0.004). There

was no strong indication that breeding stage influenced diel peak period, with the top model being the

null model (Table 1B, Figure 3, see also electronic supplementary material Table S4). The fraction of

time when the geese were active, varied between males and females during nesting, with females

being more active around noon while males were less active during that time (Figure 4A). When

geese had goslings or were moulting, females and males did not differ in the fraction of time active

(Figure 4B & C). During the period geese had goslings, there was a sharp decrease in activity after

midnight until early morning. This pattern was not present during moult.

2.   Diel patterns of immune-reactive corticosterone metabolites (CORTm)

Our results revealed that CORTm concentration was marginally influenced by time of day (Table 2),

with both cosine and sine of time of day being present in the top model as well as in many models

within the 95% confidence set (electronic supplementary material Table S5). CORTm concentration

increased during conventional night time and decreased during day time hours (Figure 5). CORTm

concentrations were slightly different between the years (Table 2; 2020 median: 32.13 ng/g dropping,

range: 7.49 – 320.9 ng/g; 2021 median: 28.56 ng/g dropping, range: 1.02 – 583.56 ng/g). In addition,

over the course of the season CORTm concentration slowly declined (Table 2). Collected mostly late

at night on one specific day, several samples contained very high levels of CORTm, *i.e.* >200 ng

CORTm/g droppings (n=7). Omitting these data points in a second analysis revealed that the outcome

did not differ substantially, except that the top model now only included the cosine of time of day and

year (electronic supplementary material Tables S6, S7).

Discussion

Svalbard barnacle geese maintain rhythmicity in physiology and behaviour during their Arctic

breeding season. We detected that most females showed a combination of both ultradian and diel

rhythmicity in incubation recesses and sitting in a sleep posture on the nest, respectively. Most

females and males also showed a combination of ultradian and diel rhythmicity or only diel

rhythmicity in activity, either when they were nesting or had goslings. Geese displayed either

ultradian rhythmicity only or a combination of ultradian and diel rhythmicity in activity during moult,

suggesting the adoption of a tidal rhythm. We detected no differences in peak periodicity between

females and males across the various breeding stages. However, during the nesting phase, females and

males exhibited contrasting activity patterns, with females leaving their nest more often during the

day and males being more active during the conventional night. Furthermore, the geese showed weak

diel rhythmicity in excreted corticosterone metabolites with concentrations increasing during the night

and decreasing during the day.

Our results indicate the importance of ultradian rhythmicity in activity behaviour over the

breeding season, but also the preservation of diel rhythmicity despite the continuous daylight of the

Arctic summer. During nesting and when geese had goslings, they showed ultradian activity rhythms

with a period of ~ 5 h (Figure 3). This ultradian rhythm might display a foraging activity – resting

pattern, as is also shown by other Arctic herbivores such as Svalbard ptarmigan, Svalbard reindeer

(*Rangifer tarandus platyrhynchus*) and muskoxen (*Ovibos moschatus*) during the summer solstice

[*e.g.* 43, 54, 63, 64]. Such an activity pattern could be endogenously generated by ultradian oscillators

[64] or controlled by the interplay of feeding and digestion [6, 65]. In addition, many geese showed

diel activity patterns in behaviour throughout the breeding season. The presence of a diel rhythm does

not inherently imply, however, that circadian systems are functional and/or entrained as the geese

could be reacting directly to external cues, also known as "masking" [1, 66]. For example, during

nesting female geese could respond adaptively to temperature differences between conventional day

and night, with slightly higher temperatures during the day despite 24h daylight. This might limit egg

cooling rates during incubation recesses, as shown in snow geese [*Anser caerulescens*, 67]. Males do

not incubate, although they sometimes sit on the nest [40], but often stand guard close by the nest

when the female is on incubation recesses [27]. Nests, however, are more vulnerable to predation if

the female is on recess [68] with the most significant egg predators, glaucous gulls (*Larus*

hyperboreus) and Arctic skuas (*Stercorarius parasiticus*), being more active during daytime [25].

This conflicts with females being on recesses then. Arctic foxes, on the other hand, are usually more

active at night [25, pers. obs.]. For more than a decade, however, no foxes have been observed on

those islands, where nests were monitored (pers. obs.). By contrast, foxes have been present in the

gosling rearing areas during the study years. Here, males and females usually rested close to water

bodies, into which they could escape to be safe from the fox [38, pers. obs.]. Although barnacle geese

may not be able to protect their nest against foxes even in the presence of both parents [69, 70], it was

suggested that fox predation might be the selective force of the observed diel rhythmicity during

incubation [25], *i.e.* a 'ghost of competition past'. If the day-active behaviour of female geese during

nesting is shaped by an expected daily rhythm in fox predation risk, it suggests the activity pattern is

regulated by an endogenous, innate circadian mechanism [71]. It is feasible that the geese' circadian

clock continues to function by employing alternative potential *Zeitgeber*s, such as diurnal changes in

light intensity, polarization patterns, solar azimuth, UV radiation, changes in the spectral composition

of light or slight changes in ambient temperature, which exist under polar summer conditions [72].

Studies on sleep patterns under natural conditions are rare due to challenges measuring this

behaviour in wild free-moving animals [73]. During incubation, we scored sleep posture, which has

been associated with rapid eye movement (REM) sleep and non-REM (NREM) sleep in domesticated

geese [29]. Sleep being scattered over the course of the day during summer has also been detected in

captive barnacle geese under a natural day - night cycle [74]. The authors proposed that barnacle

geese might have an attenuated circadian organization and may profit from becoming arrhythmic

during the polar summer. However, we found no strong evidence of arrhythmia.

During wing moult, barnacle geese are most vulnerable to fox predation [68]. At times, when

goslings are still present, parents are forced to forage in areas with high quality food even during

moult, thereby risking higher predation [38]. When goslings are depredated, however, adults can retire

to areas with lower quality food, but higher safety, *i.e.* tundra lake shores with a mossy vegetation

[38]. Over the past years, we increasingly noticed moulting geese without young to utilize the

intertidal area to rest and/or forage on algae (pers. obs.). The ~ 12 h ultradian rhythmicity patterns of

geese during moult reflect the falling and rising tides (Figure 3, electronic supplementary material

Figure S6A, [75]). The tides pose a constraint on an all-day-activity [76, 77], but the nearby sea

provides safety from foxes and the enhanced digestion during moult might also allow the geese to

utilize most of this food source, which is likely difficult to digest [31, 78]. Although geese were

shown to avoid salt-marsh vegetation in spring because presumably they are less capable

physiologically to cope with high salt loads in their environment [79], moulting barnacle geese

foraged on algae in our study, possibly benefitting from fresh river water nearby. Moult in geese is

energetically costly, because they moult wing coverts, some body feathers and flight feathers

simultaneously. To compensate, geese become less active overall (Figure 4C, [39]). In contrast, geese,

which still lead young, seem to retain their activity rhythm throughout moult. This should be

investigated in the future, because in our data set sample sizes of geese with goslings are too small to

confirm this finding conclusively.

Besides rhythmicity in activity, geese also maintained an, albeit weak, rhythmic pattern of

excreted CORTm (Figure 5), similar to what we found in human-raised barnacle goslings over 24

hours [28]. Glucocorticoids are integral in providing physiological signals to regulate biological

rhythms in sync with daily environmental cycles linking with activity and feeding [11, 12]. During the

gosling period, when we collected the majority of samples for CORTm measurements, our finding

contradicts with this general assumption, because values dropped from conventional midnight to

being lowest around midday, when geese are most active. As we discussed in our earlier study [28],

increased fox activity at night, during the period when the goose families preferentially rest, could

have affected the CORTm rhythm. In sum, whether the attenuated rhythm in CORTm is sufficient to regulate activity patterns and other functions during the polar summer or whether it plays only a minor role in endogenous timekeeping [18] needs to be investigated in the future.

The use of a multifaceted approach in this study, combining behavioural observations via wildlife cameras and accelerometers with corticosterone measurements, has allowed for a comprehensive understanding of rhythmicity in barnacle geese during their Arctic breeding season in 24h daylight. We found that accelerometers are a suitable alternative for investigating activity, where time consuming direct behavioural observations are difficult. They are a valid substitute also for pictures obtained from wildlife cameras, where a multiplicity of photos need to be analysed in detail afterwards. The peak period in activity of five females during incubation, identified from both wildlife cameras and accelerometers corresponded very well, for example. With the exception of one female, where we found diel rhythmicity retrieved from the ACC data, which was not detected in data from the wildlife cameras, the ACC-based activity patterns gave reliable results in the other four individuals, despite the slightly longer 15 min intervals of ACC measurements relative to the 5 min intervals of subsequent photos (ultradian: camera data; mean = 3.5 h, SD = 1.7, ACC data; mean = 3.8 h, SD = 1.7. diel: camera data; mean = 23.8 h, SD = 0.7, ACC data; mean = 24.2 h, SD = 0.1), This indicates that ACC-based data describe activity very reliably. This is supported by another study in captive barnacle geese, which investigated sleep – wake rhythms. Here, activity measurements from ACC data correlated well with patterns retrieved from electroencephalograms (EEG) [74]. Contrary to our study, where ACCs were mounted in neck collars, these ACCs were head-mounted. They took high-frequency measurements at a high sampling rate of 100 Hz, and could, thus, record even small head movements, whereas data provided in our study are less fine-tuned, because they were collected at a lower rate, *i.e.* every 15 min during 2-sec bursts of 20 HZ. Therefore, we cannot immediately make the same inferences at present. In a future study, however, it should be possible to estimate sleep from ACC data also in wild geese.

To summarize, over the course of their Arctic breeding season, barnacle geese show some degree of plasticity in their daily rhythms. Such intra-individual shifts in rhythms between sexes and breeding stages also exist in shorebirds [reviewed in 1, 5]. One possible explanation is that in barnacle geese

the circadian clock mechanism keeps ticking, but the control, which it exerts over behavioural output,

is plastic and is applied only when it provides some advantage [1, 2]. Alternatively, the observed

rhythms are not endogenously controlled and geese respond directly to environmental or social cues,

such as temperature, predation pressure, tides and/or their mates' activity. To investigate individual

plasticity and consistency in behavioural and hormonal rhythmicity, and their effects on fitness, are

novel avenues for future work in this Arctic herbivore.

Ethics

The study conforms to Directive 2010/63/EU and was conducted under FOTS ID 23358 from the

Norwegian Animal Research Authority and approved by the Governor of Svalbard (RIS ID 11237).

Declaration of AI use

As all but one author are none native English speakers, AI was occasionally used while writing to

improve language and readability. AI was sometimes used as 'search engine' to find suitable code for

graphics.

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Tables

Table 1: Intercept and coefficient estimates from the full models investigating differences in (A) ultradian peak period in hours and (B) diel peak period in hours with corresponding standard errors (SE), 85% Confidence intervals (lower, upper), and if the variable was selected in the top AIC model.

Variable	(A) estimate	SE	lower	upper	AIC top	(B) estimate	SE	lower	upper	AIC top
Intercept	7.30	2.42	4.11	10.49		24.03	1.12	22.52	25.53	
Breeding stage - moult	5.79	2.89	1.74	9.83	Yes	-2.26	1.62	-4.50	-0.03	No
Breeding stage - nesting	-2.18	2.96	-6.33	1.97	Yes	-0.43	1.29	-2.21	1.35	No
Sex - Male	-2.91	2.96	-6.82	0.99	No	-0.22	1.56	-2.32	1.89	No
Year - 2022	-3.53	1.93	-6.23	-0.84	No	-0.22	0.86	-1.39	0.96	No
Breeding stage - moult: sex - male	1.04	3.58	-3.97	6.05	No	3.84	2.21	0.79	6.89	No
Breeding stage - nesting: sex - male	4.49	3.82	-0.85	9.84	No	-0.22	1.80	-2.70	2.26	No

Table 2: Intercept and coefficient estimates from the full model investigating rhythmicity in log-transformed corticosterone metabolite concentration with corresponding 85% CIs, and if the variable was selected in the top AIC model.

Variable	estimate	standard error	lower	upper	AIC top
Intercept	5.03	0.45	4.38	5.67	
cosine	0.1	0.04	0.05	0.15	Yes
sine	0.06	0.04	0.003	0.11	Yes
sex - male	-0.08	0.09	-0.21	0.05	No
year - 2021	-0.2	0.06	-0.29	-0.11	Yes
year day	-0.007	0.002	-0.01	-0.004	Yes

Figures

Figure 1: Rhythmicity of females during incubation recesses [panels (A) and (B)] and 'sleep' [panels (C) and (D)]. Double

plotted actograms of female ID CA41113 as an example show in black when the female is away from the nest during

incubation recesses (A) or when she is on the nest in the sleeping posture (C), while transparency indicates inactivity on the

nest in (A) or any behaviour other than sleep in (C). The x-axis displays two consecutive days and these consecutive days are

also shown from top to bottom on the y-axis. On the y-axis t = 0 is the day the first pictures were taken by the wildlife

cameras. Population level graphs show the fraction of time, averaged over 24 hrs., spent on incubation recesses B) and in

sleep posture D) in percentages (y-axis). The x-axis displays time over 24 hours, and the black line represents the mean,

while in grey bootstrapped 95% confidence intervals are displayed.

Figure 2: Percentages of 18 individual geese in total over the course of two years, which over the three breeding stages

showed either ultradian or diel rhythmicity alone, a combination of ultradian and diel rhythmicity, or no rhythm. Numbers

above the stacked bars indicate sample size per breeding stage.

Figure 3: Diel and ultradian average peak periodicity during different breeding stages. The values of the peak period are for rhythmic geese only, i.e. a peak above the significance threshold at α = 0.05. Individual geese are shown by dots. The size of the dots represents the power of the peak period. The error bars are 95% bootstrap confidence intervals on the population mean.

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745

Figure 4: Population level graphs showing the fraction of time active, averaged over 24 hrs., during nesting (A), when geese

had goslings (B), and during moult (C) in percentages (y-axis). Females (f) and males (m) are indicated in different colours.

The x-axis displays time over 24 hours, and the solid lines represents the mean, while in transparent colour bootstrapped

95% confidence intervals are displayed.

Figure 5: Daily rhythmicity in log-transformed corticosterone metabolite concentration measured in barnacle goose

droppings. The solid line represents the predicted values from the model.