Mating strategies explain sex-biased infections in an emerging fungal disease

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<u>Abstract</u>

1) Mating dynamics can govern species impacts from novel pressures by influencing

demographic processes that affect mortality risk. Understanding how mating systems

shape interactions among individuals can improve predictions of the effects of

environmental change on populations.

2) Here, we examined sex differences in mating phenology in populations where mating

activity coincides with exposure to *P. destructans*, a lethal fungal pathogen, to understand

how reproductive strategies contribute to disease impacts. As *P. destructans* can replicate

only at the cool temperatures at which bats hibernate, we expected differences in activity

among sexes to modify disease dynamics.

3) We used passive antenna systems installed at the entrances of hibernacula to characterize

activity patterns of bats impacted by white-nose syndrome. We also measured pathogen

loads on bats during autumn mating and early hibernation to assess how infection

severity changed according to host phenology.

4) We found that females spent fewer days active during autumn, arrived after males, and

were primarily active on the warmest nights. Males remained highly active throughout

the mating period and later in autumn than females. Importantly, these differences in

mating phenology likely led to more severe infections in females during early hibernation

as male activity and thus warm body temperatures inhibited pathogen growth.

5) Differences in activity between sexes and in the transition from swarm to hibernation

likely reflects males maximizing their mating opportunities while females conserve

energy to meet the cost of spring migration and reproduction. More broadly, our results

show how mating phenology can contribute to sex-biased impacts of a novel disease and

highlight the value of understanding species' mating systems to anticipate the impacts of

environmental change.

1. **Introduction**

Global change has imposed novel pressures that threaten many species with extinction,

including habitat loss, pollution, overutilization and emerging infectious disease (Dirzo et al.,

2014; Mazor et al., 2018; Otto, 2018; Tilman et al., 2017). The resilience of populations to global

changes can vary based on the species' life history strategies, including the mating system

(Capdevila et al., 2022; Péron, 2013; Plesnar-Bielak et al., 2012; Sæther et al., 2004). Mating

system dynamics govern how quickly populations recover and adapt by influencing demographic

and evolutionary processes, such as rates of recruitment and genetic mixing (Bessa-Gomes,

Legendre, & Clobert, 2004; Legendre et al., 1999). Accordingly, traits associated with mating

systems are often shared within demographic classes (e.g., sex, age, social status, etc.) and can

result in mortality on only some demographic classes that can restructure or destabilize

populations (Clutton-Brock et al., 2002; Grüebler et al., 2008; Martin et al., 2022; Oro, Álvarez,

& Velando, 2018; Rosa et al., 2019; Wells et al., 2017). Therefore, understanding how aspects of

the mating system influence interactions between individuals and novel pressures should

improve the ability to predict responses of populations to rapid changes.

Sexes use different strategies for maximizing their fitness that give rise to variation in

morphology, behaviors, or physiology (Kolata, 1977; D. Promislow, E. L., Montgomerie, &

Martin, 1992; Trivers, 1972), which collectively can affect their mortality risk. In many species

under sexual selection, males invest in quantity of offspring through copious mating, whereas

females, who are more limited by gametic production, invest in ensuring survival of few high-

quality offspring (Bateman, 1948; Lehtonen, Parker, & Schärer, 2016). Accordingly, males may

be more likely to engage in risky behaviors to increase their competitive mating advantages

whereas females, whose fitness relies on surviving to carry and care for young, may be more

risk-averse (Bateman, 1948; Bonduriansky et al., 2008; D. E. Promislow, 1997). Additionally,

asynchrony between sexes in their reproductive investments can also contribute to differences in

how females and males respond to the same environmental pressures (Cornwallis & Uller, 2010;

Forsgren et al., 2004; Kasumovic et al., 2008), including when their peak vulnerability to

stressors occurs (Williams et al., 2022). While sex specific responses to novel pressures are

expected, the proximate factors that influence differences in impacts between females and males

are overlooked for many novel threats (Connallon, Débarre, & Li, 2018; Connallon & Hall,

2018).

Novel pressures like infectious diseases frequently interact with mating systems resulting

in differential impacts to sexes (Herrera & Nunn, 2019; Stephenson, 2019; Stephenson et al.,

2016; Zuk, 2009; Zuk & McKean, 1996) that can influence the severity of epidemics and

coevolution between hosts and pathogens (Hall & Mideo, 2018). Typically, male behaviors or

physiological states optimize fitness at the costs of higher pathogen exposure or growth,

including territory defense or aggression, dispersal, and reduced immunity (Ferrari et al., 2003;

Grear, Luong, & Hudson, 2012; Luong et al., 2010; Sauer et al., 2024; Stephenson, 2019). For

example, adult male badgers had higher contacts with conspecifics and higher risks of acquiring

tuberculosis compared to females (Delahay et al., 2006; Graham et al., 2013; McDonald et al.,

2014). Male-biased disease is more frequently documented, but female-biased disease has been

shown to arise seasonally (Cattadori et al., 2005; Sweeny et al., 2021). For example, during

parturition adult female European rabbits had the sharpest peak in nematode intensities relative

to non-reproductive females and males (Cattadori et al., 2005), illustrating that the magnitude

and direction of sex-biases can vary temporally to affect the dynamics between hosts and

pathogens (Cattadori et al., 2005; Sweeny et al., 2021). However, the behavioral and

physiological mechanisms contributing to sex-biases in impacts for introduced pathogens are

often unclear, thus limiting opportunities for management.

White-nose syndrome is a recently introduced disease of bats that is caused by the

psychrophilic fungal pathogen, *Pseudogymnoascus destructans*, and has led to population

declines exceeding 95% (Frick et al., 2015; Langwig et al., 2012; Langwig et al., 2016).

Euthermic bats that are active on the landscape maintain higher body temperatures (37-41°C)

(O'Shea et al., 2014) that are above the upper critical limit of *P. destructans* growth (20°C)

(Verant et al., 2012). However, when bats reduce their body temperature during torpor (short

periods of inactivity that comprise hibernation) to near ambient (1-16°C) (Perry, 2013; Webb,

Speakman, & Racey, 1996) the pathogen is able to grow, making it dependent on seasonal

hibernation cycles (Jackson, Willcox, & Bernard, 2022; Jackson et al., 2022; Langwig et al.,

2015; Langwig et al., 2021). Pathogen exposure occurs during autumn when bats return annually

to contaminated hibernation sites (Hoyt et al., 2020; Laggan et al., 2023) but infection and

mortality peak over winter (Hoyt, Kilpatrick, & Langwig, 2021; Langwig et al., 2015; Reeder et

al., 2012).

Bats mating occurs during autumn, known as swarm, when large numbers of individuals

aggregate near hibernacula entrances, then enter the site to form smaller groups (typically pairs

or triads), and copulate after landing on walls or in crevices (Fenton, 1969; Fraser & McGuire,

2023; Thomas, Brock Fenton, & Barclay, 1979; Wai-Ping & Fenton, 1988). While females mate

during this period, they store sperm through hibernation until spring emergence, when they

migrate to summer maternity colonies, become pregnant, and rear pups cooperatively with other

females (Fenton & Robert, 1980; Kurta & Kunz, 1987; Wimsatt, 1942, 1944, 1945). Males

segregate from females during parturition and pup rearing (Davis & Hitchcock, 1965; Fenton &

Robert, 1980). To that end, females' reproductive investments are primarily during spring and

summer, and are more extensive in magnitude and duration than males, whose reproductive

investments are highest during autumn (P. Racey et al., 2000). While autumn mating is critical

for both sexes, asynchronous allocation of energy for reproduction between females and males

could lead to extensive behavioral and physiological differences that contribute to disease

patterns and impacts. Importantly, the congregation of sexes during autumn swarm and

hibernation provide a unique opportunity to explore how sex-specific mating strategies and

phenology influence infection between sexes.

Figure 1. Hypothesized reproductive investment of females and males with different mating strategies developed from Fenton & Robert, 1980, P. Racey et al., 2000, and Wimsatt, 1945. Males are expected to invest most towards their reproductive fitness during autumn mating when they mate polygynously. Females likely require less energy during autumn mating, but considerable investment is made following hibernation as they singly parent their young in maternity colonies and require energy for gestation, parturition, and lactation.

Temperate bats reduce activity seasonally and use torpor to conserve energy even outside of winter hibernation (Alston et al., 2022). However, females and males balance torpor and activity differently throughout the year (Grinevitch, Holroyd, & Barclay, 1995; Hoff et al., 2023). These sex specific differences in activity and torpor could strongly influence WNS dynamics if sex-specific torpor use influences growth of the cool-temperature dependent fungus. Yet, empirical

investigations are limited by the challenge of obtaining individually based data on mating

dynamics through time. Previous research has shown that females can be more infected and

suffer greater impacts (Grieneisen et al., 2015; Kailing et al., 2023), but see (Johnson et al.,

2014). However, the mechanisms driving the difference between sexes remain unknown. We

hypothesize that pre-hibernation mating activity drives highly disparate infections in early winter

since the pathogen is unable to grow on bats when they are active. As such, we investigated how

traits associated with mating vary between females and males during seasons important for

reproduction (swarm mating) and disease (swarm mating and early hibernation) and contribute to

sex-biased disease. Specifically, we aimed to characterize sex-specific mating periods, examine

environmental and host factors associated with mating activity, and explore links between

phenology and disease.

2. <u>Methods</u>

2.1 Field data collection and monitoring

We studied the activity of *Myotis lucifugus* in Wisconsin, USA during autumn mating

(i.e., July 01 to November 01) each year from 2020 to 2023 at three large hibernacula consisting

of 10 total entrances. At every entrance, we installed an RFID system consisting of an antenna

with an automated data logger (IS10001; Biomark) that recorded a time-stamped datapoint each

time a passive integrated transponder (PIT) tag was in proximity. Each system was scheduled to

run continuously by a direct or solar source of power. Systems were programmed to record

unique tags every minute to help to maximize detections as >1 tag cannot be simultaneously

recorded and bats sitting on top of antenna would otherwise block detections of all other

individuals. In addition, systems recorded test tags at least once every 12 hours enabling us to

determine when systems were operational.

We captured bats at each site during autumn using a harp trap (Harptrap; Faunatech

Austbat) and mist-nets (Polyester 38mm mesh; Avinet) near the entrances where individuals

were actively swarming. For every individual, we weighed, injected a PIT tag, categorized sex

based on external morphology, and attached an aluminum-lipped band (2.9 mm; Porzana,

Icklesham, UK) as secondary identification. Capture efforts at each site concluded after one

night, unless the target number of bats (approximately 100) was not achieved, in which we

trapped a second night. We also sampled bats in early winter (November) to estimate disease

severity at the same sites that we characterized autumn activity. In early hibernation, we walked

each section of the hibernacula to collect a stratified sample of up to 20 bats and targeted

previously banded individuals. As in autumn, we recorded sex based on external morphology,

PIT tag ID, and band number of every individual. All bat handling procedures followed the

approved IACUC protocols [IACUC Protocol# 20-150] and decontamination followed USFWS

protocols as well state recommendations.

2.2 Characterizing individual activity and climate

We determined individuals' mating phenology, including swarm period and nightly

activity, from PIT tag detections (See Supplemental Table 1 for replication statement). An

individual was considered active if it was detected at least once in a night. We only included bats

detected on at least two nights to properly capture a full swarm window (beginning to end). We

classified bats that were detected between the hours of 00:00 and 07:00 as active on the previous

date to be consistent with bat ecology. For each bat observed in a year, we assigned a beginning

swarm date as the first date detected after July 01. Only individuals that returned from a previous

year were assigned a beginning swarm date, so results were not biased by the time of tagging.

Therefore, only adults were included (Brunet-Rossinni & Wilkinson, 2009). We assigned a

swarm ending date based on the last date detected before Nov 01 of the same year. For

consistency with the beginning of swarm, we only assigned ending dates for adults, but included

individuals tagged the same year since our other criteria (detected at least two nights) accounted

for the bias that would arise from timing of tagging efforts.

We extracted climate data using the R package 'daymetr' (Hufkens et al., 2018) and GPS

coordinates of hibernation sites, which provided continuous, grid-based estimates of daily

weather variables based on location. To reflect bat ecology, and because mean temperatures were

not available in the climate data, we calculated a mid-point temperature (Jackson et al., 2022) for

each date as: (minimum temperature on the following day + maximum temperature of current

day)/2.

2.3 Estimating pathogen loads

For every bat captured during autumn (late August to early September) and early

hibernation (November), we collected an epidermal swab using a standardized swabbing

technique (Frick et al., 2015; Langwig et al., 2015) to measure infection severity. Immediately

upon collection, we stored swabs in RNAlater until testing was performed. We tested in

accordance with protocols specific to *P. destructans* that used Qiagen DNEeasy Blood and

Tissue Kits (Qiagen, USA) to extract fungal DNA followed by qPCR to quantify fungal loads

(Langwig et al., 2015; Muller et al., 2013).

2.4 Statistical analyses

2.4.1 Summary

We tested our hypotheses using generalized (i.e., GLMM) and linear (i.e., LMM) mixed

effects models to characterize autumn mating activity and disease by sex with R package 'lme4'

(Bates et al., 2015). We extracted model estimates using R package 'emmeans' (Lenth, 2022).

See Appendix with code for description of all statistical models and output.

196   *2.4.2 Autumn mating period*

197   We determined individually based active periods using LMMs. First, we estimated when

198   sexes start autumn swarm using beginning date (in monthly units then scaled to start at 0) as the

199   response variable, sex as a fixed effect (F|M), and site-year and individual as random effects.

200   Then, we used the same model structure, but with ending date as a response to estimate when

201   sexes end swarm.

202   *2.4.3 Effect of temperature on nightly activity*

To test whether environmental conditions differently affected nightly activity between

sexes, we used a GLMM with a binomial distribution to test the probability of an individual

being active (=1) or undetected (=0) with temperature and sex as interactive fixed effects and

individual and site-year as random effects. For the response variable, we treated each bat as a

series of binomial trials using all the possible dates between the first and last detection where an

individual was detected (=1) or undetected (=0). We excluded nights where the minimum

temperature was less than 10°C to minimize outlier effects as bat activity is greatly reduced

(Gorman et al., 2021).

211   *2.4.4 Effect of time since arriving to swarm on ending activity*

We examined how swarm duration affected the probability of ending activity. To do so,

we tested whether the days since arriving to swarm contributed to sex-based phenological

transitions using a GLMM with a binomial distribution that predicted individuals ending activity.

For the response variable, each bat was treated as a series of binomial trials where an individual

was detected for the last time (=1) or was detected on a subsequent date (=0). We included

interactive effects of sex and days since first detection, binned over 3-day intervals, with

individual ID and site-year as random effects. Days since first detection was calculated for

individuals as the date of activity minus the beginning date of swarm, whereby dates were treated

as monthly units and scaled to start at zero as above.

221   *2.4.5 Seasonal infection severity*

222   To understand disease dynamics according to mating phenology, we tested differences in

223   pathogen loads between sexes and across seasons using a LMM with $\log_{10}$ *P. destructans* loads

224   as our response variable, sex and monthly date units as fixed effects, and site-year as a random

225   effect.

227   **3. Results**

228   We characterized autumn activity of 148 female and 606 male *M. lucifugus* and measured

infection on 151 female and 368 males between 2020 and 2023. All statistical results are

reported with males as the intercept or reference level and thus coefficient +/- SE are in reference

to the value above or below the male references. Females began swarm activity later (Female

coef +/- SE = 0.249 +/- 0.08, t-value = 3.249; Figure 2, top panel; Appendix 1.1) but also ended

activity earlier than males (-0.387 +/- 0.06, t-value = -6.280; Figure 2, top panel; Appendix 1.2).

Male activity fully encompassed the activity period of females (Figure 2, bottom panel;

Appendix 1.3) and the number of individuals detected was overwhelmingly male biased

throughout autumn, given their higher abundance and activity overall (Supplemental Figure 1;

Appendix 4.1), and this finding was consistent between sampling events (Supplemental Table 2;

Appendix 4.2).

Within their respective mating periods, the probability of being active in a night or not for

each sex was differently affected by temperature (detected=0|1; Female*temperature = 0.151 +/-

0.03, p-value < 0.001, Figure 3A; Appendix 2.1), such that the probability of females being

active was lower overall but increased more quickly with increasing temperatures (Temperature

slope estimated using 'emtrends': Female slope = 0.152 +/- 0.03 vs Male slope 0.001 +/- 0.01,

Figure 3A; Appendix 2.1), notably above 16°C. The sex and temperature interaction was also

evident when we used a balanced dataset of males and females (see Supplemental Statistical

Methods, Supplemental Figure 2A; Appendix 4.3.1). We also found support for an interaction

between the number of 3-day intervals since first detection and sex on the probability of ending

swarm (Date intervals since arrival*female = 0.367 +/- 0.07, p-value < 0.001, Figure 3B;

Appendix 2.2), as well as an interactive effect of temperature and sex on the probability of

ending activity, such that females were significantly more likely to end their active period as

temperatures decreased than males (Temperature*female = -0.166 +/- 0.08, p-value = 0.031,

Supplemental Figure 3; Appendix 4.3.3).

There was no clear sex bias in pathogen loads throughout autumn swarm (Female = -

0.044 +/- 0.09, t-value = -0.502; Appendix 3.1), however, following swarm mating, female

pathogen loads increased more rapidly, and females became significantly more infected than

males by early hibernation (Female*date = 0.293 +/- 0.08, t-value = 3.876; Figure 4; Appendix

3.2). Ending of autumn activity of males and females generally varied among sites and years and

earlier end dates were associated with higher pathogen loads on individuals (See Supplemental

Statistical Methods) (Median last date of activity slope on pathogen loads: -1.333 +/- 0.44, t-

value = -3.306; Supplemental Figure 4a; Appendix 4.4.1) as well as site-level pathogen loads in

early hibernation (Median last date of activity slope on mean pathogen loads: -1.400 +/- 0.38, t-

value = -3.715; Supplemental Figure 4b; Appendix 4.4.2).

Figure 2. *Top:* Differences in the autumn swarm period by sex. Each point represents the date of

beginning (first detection) and ending (last detection) autumn mating for an individual bat. Black

circles represent model estimated dates with standard errors shown with horizontal bars. ***Bottom:***

Distribution of activity by sex. Each density plot shows the proportion of all detections grouped

by sex that occurred on each date. An individual bat can be represented on multiple nights if

activity occurred. Asymmetry in the peak of male activity (based on the total number of

observations) relative to their estimated mating period in the top panel demonstrates that while

most males arrive early in autumn, they maintain activity into later swarm dates.

Figure 3A. Effect of temperature on the probability that an individual was active in a night. Each

point represents the proportion of individuals from a site and year that were active at each

temperature by sex. Females are shown by purple and males in orange and lines represent the

model predicted probabilities of being active across the temperature gradient. **Figure 3B.**

Relationship between number of days between starting swarm and the probability of ending

activity for females in purple and males in orange. Actual data are binary (1|0) such that 1 = ends

activity and 0 = continues to swarm. For visualization, we show raw data as proportions where

each point represents the proportion of individuals since beginning swarm mating that were

detected for the last time. Proportions were calculated for each 3-day interval at a site and year.

Lines denote the model predicted probabilities of ending activity.

Figure 4: Pathogen loads measured on adults of each sex throughout autumn swarm mating and

early hibernation periods. Each point represents an individual bat with females in purple and

males in orange. Solid lines denote model predicted fungal loads over time for females and males

respectively. Dashed lines represent the model predicted end dates of activity as estimated in

Figure 2. There was no clear difference in infection between sexes during the active swarm

period (August and September), but female infections were nearly an order of magnitude more

severe than males by the beginning of hibernation (November).

4. Discussion

Sex specific mating strategies are common across species, yet how aspects of mating

systems give rise to sex-biased disease impacts are often unexplored. Our results suggest that

asynchronous mating phenology between sexes may be a key driver of seasonally sex-biased

infections, which are known to affect pathogen transmission and population impacts in other

host-pathogen systems (Ferrari et al., 2003; Graham et al., 2013; Grear, Luong, & Hudson, 2012;

Perkins, Ferrari, & Hudson, 2008; Sauer et al., 2024; Silk et al., 2018; Skorping & Jensen, 2004).

We show female *M. lucifugus* begin swarm activity later and end earlier, spend fewer nights

active after arriving to swarm, and have a more pronounced increase in activity on warmer nights

compared to males. Importantly, pathogen loads do not differ between sexes until after the active

period of autumn mating, when bats transition to hibernation, and earlier end dates of autumn

activity were associated with higher pathogen loads by early winter. Earlier use of torpor by

females likely contributes to female-biased disease as it allows an extended period for the

pathogen to replicate as opposed to males that remain active and can minimize pathogen

replication through euthermia.

In exploring within activity period differences, we found that females are far less active

during their shorter swarm period compared to males. This is probably best explained by the

need for females to conserve enough energy stores to survive hibernation. Females store sperm

overwinter and then give birth and raise offspring following emergence (Czenze, Jonasson, &

Willis, 2017; Jonasson & Willis, 2011; Wimsatt, 1945). We provide two new lines of

individually derived evidence that aligns with females using energy more conservatively than

males. First, we found a clear effect of temperature differently affecting the probability that

females and males are active on a given night within their swarm period. This suggests that

females may be prioritizing the limited days they spend active in autumn to forage on warmer

nights that average nearly 20°C (Appendix 4.3.2) likely because thermoregulating is more

efficient at warmer temperatures (Fjelldal et al., 2023; Rubalcaba et al., 2022) and prey resources

are more active (Grüebler, Morand, & Naef-Daenzer, 2008; Taylor, 1963). Second, we found

that females on average, regardless of their arrival date, begin transitioning into hibernation (i.e.,

remained in the site/stopped activity) after four days of activity, whereas males maintained

nightly activity until late in autumn regardless of the date they arrived to swarm, and averaged

over 13 active nights (Supplemental Figures 1 and 5, Appendix 4.1 and 4.5). Males that are less

constrained by reproductive energy budgets were active nearly four times as many nights as

females (Supplemental Figures 1 and 5; Appendix 4.1 and 4.5), and when temperatures are

relatively low, suggesting that males are optimizing mating opportunities over energy

conservation (Figure 3A, Appendix 2.1 and Supplemental Figure 2A-B, Appendix 4.3.1-4.3.2)

and supports previous hypotheses (Burns & Broders, 2015; Glover & Altringham, 2008; Rivers,

Butlin, & Altringham, 2006; Willis, 2017). Importantly, the disparate reproductive costs and

mating strategies between sexes is likely contributing to how each sex interacts with the same

novel pressure of an introduced pathogen.

The earlier arrival of males to mating locations than females, termed protandry, is

observed across many taxa, and two hypotheses have been posed to justify its ubiquity (Morbey

& Ydenberg, 2001). First, the 'rank advantage hypothesis' states the earliest arriving individuals

have access to the best resources or territories, providing a competitive advantage in acquiring

females. Second, the 'mate opportunity hypothesis' suggests that earlier arriving males can

increase the number of females mated, resulting in greater annual reproductive success. In

systems under strong sexual selection and in cases where females also have resource-based

advantages of arriving early, mate opportunity has more reliably explained protandry than rank

advantage, particularly in species exhibiting sperm competition and highly male-biased sex ratios

(Kokko et al., 2006), which is known of temperate bats (Paul A. Racey & Entwistle, 2000; P. A.

Racey et al., 1987). Sperm storage by females can also generate advantages for males that more

closely synchronize copulations with female receptivity, such that males that mate earlier can

secure storage sites in uteri that improve fertilization success (Scialli, 1993). Thus, we

hypothesize that the longer swarm period of males is driven by the advantage of increasing

copulations with more females in addition to elevating fertilization potential, as suggested by

other studies of temperate bats (Burns & Broders, 2015; Pfeiffer & Mayer, 2013). Other

environmentally mediated mechanisms likely function in concert with mating advantages.

Females migrate from summer maternity colonies and, because foraging to store fat for

hibernation is essential for bats in autumn (Kunz, Wrazen, & Burnett, 1998), timing the

departure from diminishing resources in summer habitats to optimal resource conditions at

swarm locations is likely a more important balance for females that have greater annual energetic

constraints than males.

As in other disease systems, sex-specific differences in behavior or physiology during

periods of pathogen exposure can result in sex-biased disease impacts and govern host-pathogen

dynamics (Guerra-Silveira & Abad-Franch, 2013). We showed that during the mating season

when bats are initially exposed to *P. destructans*, activity differs markedly between sexes,

however it did not promote differences in infection within that period (Supplemental Figure 2;

Appendix 3.1) as we expected based on other disease systems (Ferrari et al., 2003; Graham et al.,

2013; Guerra-Silveira & Abad-Franch, 2013; Klein, 2000). Rather, reproductive demands that

influence broader phenological patterns are more likely to be shaping sex-biased disease through

physiological mechanisms. Female infections rapidly increased almost two orders of magnitude

within the two months between the end of their active period and early hibernation (Figure 4),

suggesting the timing of exposure and pathogen growth have temporally decoupled the effects on

sex-specific disease in this host-pathogen system. We found that ending activity earlier was

associated with higher pathogen loads in early hibernation, which aligns with females being the

earliest to hibernate and develop the highest pathogen loads. Ending activity is significant in this

system because the pathogen can only replicate once bats begin hibernation, and hosts that

exceed an infection threshold earlier have lower over-winter survival (Langwig et al., 2016).

Consequently, females are likely predisposed to have higher disease than males because of their

need to conserve energy to achieve reproductive fitness.

Phenology evolves to synchronize important biological events, such as reproduction, to

optimal conditions and resources (Visser & Gienapp, 2019). However, rapid changes in the

biotic or abiotic environment can modify which traits are beneficial or maladaptive (Schlaepfer,

Runge, & Sherman, 2002). For bats affected by WNS, if swarm periods evolved to maximize

survival and recruitment, disease-induced mortality of early arriving and less active individuals

may be selecting for traits that are beneficial for surviving infection but potentially maladaptive

for recruitment. Alternatively, if phenology is a plastic trait in bats as it is in other taxa (de

Villemereuil et al., 2020), a shift to a later swarm window may minimize disease while still

supporting recruitment, thus ameliorating population impacts. Given the clear, primarily additive

effect of sex on periods of mating activity (Figure 2; Supplemental Table 2; Appendix 4.2), we

expect that any shifts in activity of one sex will be paralleled by the other. Because females have

a more restricted activity period and energy budgets, we expect females to be less flexible in

their phenology than males. As such, females will likely need to adapt alternative mechanisms to

overcome novel disease pressure or compensatory mechanisms to maintain recruitment if a later

swarm window is suboptimal for fitness. Previous work has implicated female-biased disease in

an initial reduction in the proportion of females following WNS invasion (Kailing et al., 2023),

however, many colonies where the pathogen invaded earliest are recovering (Grimaudo et al.,

2021; Hoyt, Kilpatrick, & Langwig, 2021; Langwig et al., 2017). Therefore, long-term

ecological and evolutionary responses of each sex to disproportionate disease impacts remain to

be explored but are warranted as they may contribute to how populations persist.

While sex-based differences in disease have received considerable attention, the

mechanisms often remain unexplored. We demonstrate that in the absence of clear behaviorally

mediated transmission differences, sex-specific physiology associated with phenological

transitions following exposure can facilitate sex-biased disease, providing a unique example in

which physiological asynchronies decouple impacts of the timing of acquiring a pathogen and

disease progression between sexes. Importantly, our findings suggest that asynchronous

reproductive energy allocation contribute to sex-biased infections, revealing a potentially

significant influence of mating dynamics on the trajectory of epidemics and pathways of host

persistence or extinction. Other novel pressures that similarly act more strongly on female

mating dynamics may impede population recoveries if sex-ratios are disrupted or if traits

beneficial for survival are antagonistic for recruitment. Given the importance of demographic

structure and recruitment in population impacts, determining how mating strategies influence

responses to novel stressors will be critical for conserving species in the face of increasing global

change.

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Conflict of interest

All authors declare no conflict of interest.

Author contributions

Macy J. Kailing, Kate E. Langwig, and Joseph R. Hoyt conceived the ideas and designed

methodology; Macy J. Kailing, Joseph R. Hoyt, J. Paul White, Jennifer A. Redell, Heather M.

Kaarakka, and Kate E. Langwig collected the data; Macy J. Kailing analyzed data with input

from Kate E. Langwig and Joseph R. Hoyt. Macy J. Kailing wrote the original manuscript with

Kate E. Langwig and Joseph R. Hoyt. All authors contributed critically to the drafts and gave

final approval for publication.

Data availability statement

The datasets and code will be deposited in Dryad Digital Repository upon final submission.

Exact site locations are not disclosed to protect endangered species and landowners.

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