1    Evolution of drought resistance strategies following the introduction of white clover (*Trifolium*

2                                            *repens* L.)

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ABSTRACT

**Background and Aims:** Success during colonization likely depends on growing quickly and

tolerating novel and stressful environmental conditions. However, rapid growth, stress

avoidance, and stress tolerance are generally considered divergent physiological strategies.

**Methods:** We evaluate how white clover (*Trifolium repens*) has evolved to a divergent water

regime following introduction to North America. We conduct RNAseq within a dry-down

experiment utilizing accessions from low and high latitude populations from native and

introduced ranges, and assess variation in dehydration avoidance (avoidance of wilting) and

dehydration tolerance (ability to survive wilting).

**Key Results:** Introduced populations are better at avoiding dehydration, but poorer at tolerating

dehydration than native populations. There is a strong negative correlation between avoidance

and tolerance traits and expression of most drought-associated genes exhibits similar tradeoffs.

Candidate genes with expression strongly associated with dehydration avoidance are linked to

stress signaling, closing stomata and producing osmoprotectants. However, genes with

expression linked to dehydration tolerance are associated with avoiding excessive ROS

production and toxic bioproducts of stress responses. Several candidate genes show differential

expression patterns between native and introduced ranges, and could underlie differences in

drought resistance syndromes between ranges.

**Conclusions:** These results suggest there has been strong selection following introduction for

dehydration avoidance at the cost of surviving dehydration.

## INTRODUCTION

Species introductions are occurring at a drastically increasing rate due to globalization and human commerce (Seebens *et al.* 2017; Bonnamour *et al.* 2021). Determining how and why some introduced species become invasive is a persistent goal for biologists that has had relatively limited success (Kolar and Lodge 2001; Pyšek and Richardson 2007). Introduced species almost certainly encounter some level of novel abiotic or biotic environments during initial colonization, and must both establish and spread to become invasive. While historically neglected as an important evolutionary force during invasion, selection is commonly observed during successful invasions - especially for human commensal species that are introduced repeatedly and/or have high genetic variation among colonists (Huey 2000; Maron *et al.* 2004; Colautti and Lau 2015; Battlay *et al.* 2024).

Selection is key for many of the hypotheses that describe why some species become successful invaders. Stress resistance and phenotypic plasticity have frequently been hypothesized as key attributes for successful invasive species (Richards *et al.* 2006; Gewing *et al.* 2019). Individuals that are able to avoid or tolerate stress can better establish and spread in novel environments. Key to this hypothesis is that selection acts to remove less stress resistant individuals from the populations. Such selection could also occur only in certain areas of the introduced range where selection pressures are strongest. Alternatively, hypotheses like the Evolution of Increased Competitive Ability (EICA; Blossey & Notzold, 1995) suggest that introduced plants are removed from key selective pressures in the native range following colonization and the strongest selection pressure is simply for rapid growth rate and high fecundity to outcompete native species.

One of the most important selection pressures for plants is water availability and associated

drought events (Siepielski *et al.* 2017). Three different non-mutually exclusive strategies are

hypothesized to evolve in environments with low or inconsistent water availability: drought

escape, dehydration avoidance and dehydration tolerance (Ludlow 1989; Kooyers 2015; Volaire

2018). Plants that have evolved to escape drought grow rapidly, reproduce quickly, and finish

their life cycles prior to or immediately following drought stress. Plants that are dehydration

avoiders actively evade drought conditions with physiological or morphological adaptation or

acclimation responses to limit desiccation and thus wilt at a lower soil volumetric water content.

While dehydration avoidance can be a constitutive strategy where certain individuals may have

high succulence, more trichomes, higher water-use efficiency, or lower stomatal density, it also

manifests as a response to reduced water availability by lowering stomatal conductance, raising

water-use efficiency, and producing osmoprotectants for osmotic adjustment. Plants with

dehydration tolerant strategies limit water-loss during drought conditions by limiting vegetative

growth, allocating resources to root growth, and producing antioxidants to limit cellular damage

from desiccation. These responses allow plants to recover following intense drought. A common

experimental design to tease apart variation in drought strategies between populations and/or

species is conducting dry-down experiments (e.g. Des Marais *et al.*, 2014; Bouzid *et al.*, 2019;

FitzPatrick *et al.*, 2023). In these experiments, variation in time to first flower can be considered

a drought escape metric, differences in soil relative water content at wilting can be considered a

measure of dehydration avoidance, and the ability to recover following wilting is a measure of

dehydration tolerance (Bouzid *et al.* 2019).

While drought strategies are not mutually exclusive, negative correlations between some

strategies are expected due to physiological constraints. For instance, the rapid growth expected

for drought escape requires opening stomata in order to increase photosynthetic rate (Geber and

Dawson 1990; Dudley 1996). However, open stomata increase transpiration and lower water use

efficiency – effectively neutralizing any dehydration avoidance strategy. Yet, the evidence for

this tradeoff is equivocal and seemly context dependent (McKay *et al.* 2003; Kooyers *et al.*

2015). Correlations between dehydration avoidance and tolerance strategies are less explored,

and are often viewed as compatible responses (Volaire 2018). However, resource allocation

tradeoffs could be hypothesized as the functions necessary for maintaining water balance in a

limited water environment are not the same resources necessary for surviving and recovering

from dehydration.

Although the molecular signals underlying the perception and integration of water stress are

becoming better resolved, variation in drought-related responses is rarely understood at the

molecular level (VanWallendael *et al.* 2019). For instance, it is largely unknown whether

variation in drought resistance arises through differences in sensitivity or perception of drought,

integration of signals, or re-wiring of connections within gene-regulatory networks. Molecular

responses to low water availability center around the activation of the abscisic acid (ABA)

dependent and ABA-independent signal transduction pathways in response to disruption in water

potential relative to the surrounding soil and/or atmosphere (Raghavendra *et al.* 2010; Dar *et al.*

2017; Aslam *et al.* 2022). Increases in ABA drive a signal cascade leading to guard cell

regulation and stomatal closure (Hsu *et al.* 2021). Notably, production of ABA also induces gene

regulatory cascades that lead to increases in antioxidants and osmoprotectants such as proline,

trehalose, and raffinose family oligosaccharides (Strizhov *et al.* 1997; Lunn *et al.* 2014). The

interactions of ABA with other plant hormones (specifically antagonistic interactions with

gibberellic acid and ethylene), lead to cessation of shoot growth and/or increased root growth

(Wilkinson and Davies 2010). Not surprisingly, this complex web of gene regulatory cascades

has substantial overlap – for instance, products of the SNF1-related kinases 2 (*SnRK2*) gene

family integrate growth rate changes, promote stomatal closure, and increase osmoprotectants

(Yoshida *et al.* 2015; Belda-Palazón *et al.* 2020). A key goal for the plant science community is

determining how these networks are manipulated to create the diversity of responses observed

within and among species (Kesari *et al.* 2012; Des Marais *et al.* 2014). Invasive species with

divergent responses between introduced and native areas can provide key natural experiments for

examining the evolution of drought resistance.

White clover (*Trifolium repens*) is perennial plant that is native to Eurasia but has been

distributed across the world as a cover crop and forage legume during the past 300 years

(Kjærgaard 2003). Despite relatively recent introductions, there is little decline in genetic

diversity in the introduced ranges of white clover (Wu *et al.* 2021) and extensive evidence for

selection and adaptation following introduction (Kooyers and Olsen 2012, 2013; Wright *et al.*

2018, 2021; Santangelo *et al.* 2022; Battlay *et al.* 2024). While white clover is most often

associated with temperate habitats, there is extensive variation in aridity across both the native

and introduced ranges, and water availability seems to be a key selective pressure (Kooyers *et al.*

2014). Escape, avoidance, and tolerance responses have been noted by different groups. Drought

escape via more annualized life history is favored in lower latitude populations in North America

(Wright *et al.* 2021). Dehydration avoidance may be favored in these same populations as clines

in cyanogenic glucosides, potentially key sources of nitrogen during drought conditions, have

evolved following introduction to several different continents. In each introduced region, the

proportion of plants possessing cyanogenic glucosides increases in more arid areas (Kooyers &

Olsen 2013, 2014). Finally, agricultural breeders frequently target persistence following drought

as a key trait for selection in agricultural lines (Marshall *et al.* 2001; Jahufer *et al.* 2002).

However, differences in drought resistance strategies have not been systematically examined

across native and introduced populations.

Here we use a manipulative dry-down experiment to examine how populations across the native

and invasive ranges of white clover resist drought. Specifically, we examine the relative degree

of dehydration avoidance and tolerance among populations from low and high latitude

populations in the native and invasive range using volumetric water content at wilting as a

measure of dehydration avoidance and ability to recover following wilting as a measure of

dehydration tolerance. We also examine differential expression between well-watered and dry

down plants at a timepoint when the dry down lines begin to experience drought stress. We use

this data to address four questions: First, do populations from introduced areas or from more arid

areas have more substantial drought resistance strategies? The stress tolerance hypothesis

suggests that introduced plants should be better stress tolerators, but EICA may suggest the

opposite. Second, are there tradeoffs that exist among dehydration avoidance and tolerance

strategies? That is, are plants that are good dehydration avoiders also poor dehydration

tolerators? Third, do native and introduced populations exhibit different patterns of gene

expression in response to early drought stress? That is, are differentially expressed genes

between well-watered and drought treatments also variable among ranges and populations?

Fourth, how do expression differences control variation in dehydration avoidance and tolerance

strategies? Together these experiments provide insight on the molecular mechanisms underlying

the evolution of drought resistance across a rapid invasion.

MATERIALS AND METHODS

We performed a manipulative experiment to examine variation among drought strategies and the

transcriptomic drivers of such variation within and among the native and introduced ranges of

white clover, *Trifolium repens*. We planted seed collected from 3-4 populations from low

latitude and high latitude in the native and invasive range (14 total populations; Fig. S1; Innes *et*

*al.*, 2022). Seeds for each population had been pooled from >25 different maternal lines. For

each population, we used latitude and longitude to extract WorldClim2 variables (Fick and

Hijmans 2017) and annual aridity and evapotranspiration data from CGIAR (Zomer *et al.* 2008).

Seeds were scarified and planted into 4" pots filled with PRO-MIX LP-15 soil pre-saturated with

water. Pots were placed in AR-66L2 growth chambers (Percival Scientific; Perry, IA, USA) and

misted daily to promote germination. Growth chambers were set to a constant 22°C with a

14hr:10hr day:night cycle. Germinants were randomized within well-watered and dry-down

treatments (well-watered = 26 individuals, dry-down = 25 individuals; Table S1). Plants were

rotated within chambers every three days to minimize microenvironmental variation. Plants were

grown for six weeks to accumulate above and below ground biomass. At six weeks, all pots were

saturated with water by bottom-watering. Number of leaves on each plant were counted and one

pot was excluded from further analysis because it was substantially larger than the other plants.

Plants in the well-watered (control) treatment received periodic watering to saturation. Plants in

the dry-down treatment did not receive additional water. Each day, we assessed volumetric water

content in each pot using a SMT150T soil moisture meter (Dynamax; Houston TX, USA). Well-

watered and dry-down treatments functioned as expected with separation in volumetric water

content (VWC) between treatments after control plants were resaturated on day 9 (Fig. S2)

To examine dehydration avoidance, we recorded the day and the soil moisture at which each

plant in the dry-down treatment wilted (hereafter, "wilt VWC"). To determine dehydration

tolerance, we determined the propensity to recover following wilting (hereafter "wilt survival").

Forty-eight hours after wilting, we resaturated the pots of each wilted plant by bottom watering

for an extended period of time (~1hr). We then resurveyed plants for survival three days (72hrs)

after resaturation. Survival was measured as a discrete variable (yes/no) with any revived green

tissue or new growth counting as survival.

**Manipulative Experiment Statistics**

All statistics were conducted in R v4.2.2 (R Foundation for Statistical Computing, Vienna,

Austria) with univariate linear models (LM) and generalized linear models (GLM) implemented

with the lm() or glm() functions, respectively. First, to examine whether plant size influenced

either dehydration avoidance or tolerance, we examined associations between either wilt VWC

and wilt survival against leaf number at the beginning of the experiment. Neither wilt VWC or

survival was significantly associated with leaf number (Fig. S3). Models below produce

qualitatively similar results whether or not leaf number at day 1 is included as a covariate.

We examined whether variation in dehydration avoidance and tolerance was associated with

range (native/introduced), latitude (low/high) or an interaction using generalized linear models.

Specifically, we modeled wilt VWC and wilt survival as response variables within a GLM with

range, latitude, and range:latitude interaction as factors. Wilt VWC was modeled with a gaussian

distribution with an identity link and wilt survival was modeled with a binomial distribution with

a logit link. Statistical significance was determined through ANOVA using a type II sum of

squares implemented with the *Anova()* function (*car* library; Fox *et al.*, 2013)

Latitude across continental ranges may not be strongly related to water availability or aridity of

populations and thus may not be the main factor driving variation in drought strategies. To

examine whether variation in drought strategies was related to differences in precipitation or

aridity, we examined associations between wilt VWC and wilt survival with annual aridity index,

potential evapotranspiration, and coefficient of variation in annual precipitation (bio15).

Distributions and links of models as well as assessment of statistical significance were the same

as the above models. Leaf number at day 1 of the experiment was included as covariate in each

analysis as there were weak associations between each climatic variable and initial plant size

(Fig. S4).

To explore potential tradeoffs between dehydration avoidance and tolerance, we examined

associations between wilt VWC and wilt survival using a GLM (binomial distribution, logit

link). Ability to recover from wilting was the response variable and volumetric water content at

wilting the predictor variable. Significance was tested with an ANOVA as above and model fit

was assessed using the *adjR2()* function (glmToolbox package; Vanegas *et al.*, 2024) .

**RNAseq Analysis**

We used an RNAseq analysis to examine differential expression underlying variation in drought

strategies. During the manipulative experiment, leaf tissue from two healthy adult leaves was

flash frozen in liquid nitrogen ten days after the dry down treatment began from plants in both

the well-watered and dry-down treatment. Tissue sampling was done during a two-hour period in

the afternoon with order of collection randomized within each treatment. Total RNA was

extracted with Direct-zol RNA Miniprep (Zymo Research; Irvine, CA, USA). RNAseq libraries

were constructed with QuantSeq 3' mRNA-Seq FWD prep kit (Lexogen; Vienna, Austria) using

an input of ~415ng total RNA/sample following manufacturers protocols. Two samples had low

RNA concentrations and we followed Lexogen's modified procedure for low input samples.

Four samples from the manipulative experiment did not have RNAseq libraries constructed due

to loss of labels during shipping or near-zero read counts (RNAseq N = 47; Table S2). All

individuals were barcoded and multiplexed in a single tube. We sequenced this library on a

single HiSeqX lane (PE 150bp reads) through Novogene (Sacramento, CA, USA). A second

round of multiplexing and sequencing was conducted for a subset of samples that had low data

yield the first round, and files from both rounds were subsequently merged. Raw sequenced

reads per individual averaged 37,241,613 reads (SD 16,534,847; Table S2)

Fastp v.023.4; (Chen *et al.* 2018) was used to trim adapters and poly-a tails. Microbial RNA

contamination was removed by aligning each sample to all fungal and bacterial assembly

genomes within the NCBI database using bowtie2 v2.5.1(Langmead and Salzberg 2012). The

bacterial and fungal genome database was indexed with the 2.2.4 release of bowtie2. The

remaining reads that did not align to the contamination database were presumed to be white

clover RNA reads. A transcriptome was created from the *T. repens* reference genome

(Santangelo *et al.* 2023) using gffread v0.12.7 (Pertea and Pertea 2020). A full decoy-aware

transcriptome was constructed to mitigate specious mapping of reads that occur from

unannotated genomic loci that are similar to the annotated transcriptome, and a mapping-based

index was then constructed using a k-mer hash over k-mers of length 31 using Salmon v1.10.2

(Patro *et al.* 2017). Read mapping to the transcriptome and quantification was then performed

with Salmon v1.10.2 in mapping-based mode using the *T. repens* transcriptome and RNA

sequences. There was a clear issue with DNA contamination as only 3.83% of reads per

individual aligned to the transcriptome (SD 1.0%; Table S2); however, this should not bias our

comparative analysis (differential expression or transcript abundance-phenotype associations)

that rely on read counts of each transcript.

**Quantifying expression across treatment, range, and latitude**

We used DESeq2 (Love *et al.* 2014) to test for differences in transcript abundance between dry-

down and well-watered treatment groups, between the North American and European range, and

between high and low latitude populations. We controlled for the VWC at the time of tissue

collection by treating it as a covariate in the DESeq2 model (i.e., Transcript ~

Treatment*Latitude*Range + SoilVWC). False discovery rate (FDR) for each gene was

calculated and a transcript was then categorized as differentially expressed if FDR was < 0.1.

Concordance of gene expression responses to drought between ranges and latitudes was

determined by comparing the coefficient estimates of protected one-way ANOVAs. That is, we

conducted ANOVAs for each gene found to be differentially expressed between the well-watered

and dry-down treatments with relative expression as the response variable and either range or

latitude as the predictor variable.

All transcripts were used for weighted correlation network analysis (WGCNA) to determine gene

modules and co-expression networks within the WGCNA v1.72-5 library (Langfelder and

Horvath 2008). To achieve homoskedasticity amongst transcript abundance, the variance

stabilization transformation from the fitted dispersion-mean relations was calculated with

DESeq2, which was then used to transform transcript count data by dividing by the size factor.

Normalized transcript counts were then analyzed with WGCNA using a soft-thresholding power

of 9 and assuming that biological networks follow a scale-free structure (Barabási and Albert

1999). The sets of genes within each resulting module were then cross referenced with the

'biological process' gene ontology terms (GO; Ashburner *et al.*, 2000) of *Arabidopsis thaliana*

and used for gene enrichment analysis with an FDR cutoff of 0.1. Fold enrichment, $-\log_{10}(FDR)$,

and number of genes for each GO term was then calculated. Genes belonging to the same

module were then compared to the Kyoto Encyclopedia of Genes and Genomes (KEGG;

Kanehisa & Goto, 2000) to visualize expression pathways.

**Identifying genes associated with wilt VWC and wilt survival**

GLMs were constructed to quantify the correlation of transcript abundance with either wilt VWC

or wilt survival. First, transcripts of drought treatment samples were standardized to acquire

relative expression for each gene. Univariate GLMs included relative expression as the response

variable and either wilt VWC (gaussian distribution, identify link) or wilt survival (binomial

distribution, logit link) as predictor variables with VWC at the start of the experiment as a

covariate. Significance was then tested using ANOVA as above. To determine the set of

phenotypically significant genes that are co-expressed, genes significantly associated with each

phenotype were compared to the previously calculated WGCNA network. We also determined

the location of these genes within the white clover genome (Santangelo *et al.* 2023) via *blastn*

(Altschul *et al.* 1990).

Finally, we then determined which of the genes associated with phenotype also were

differentially expressed between ranges or across latitudes to identify genes likely to be involved

in adaptation following introduction or to divergent climatic conditions. We conducted GLMs

modeling relative expression as a function of traits and either range or latitude. Specifically

relative expression was the response variable while trait value (wilt VWC or wilt survival),

spatial group (range or latitude), and the interaction between trait:group were included as

predictors. Genes with relative expression that is explained by trait:group interaction are good

candidates for producing the phenotypic variation observed in nature.

RESULTS

There was substantial variation in dehydration avoidance and tolerance throughout the native and

introduced ranges of white clover. Wilt VWC was lower in North American populations than in

European populations ($F_{1,23}$ = 4.8, p = 0.03; Fig. 1A) indicating greater drought avoidance in

introduced populations. However, the effect size of this difference was low as the difference in

mean wilt VWC was only 1.3% lower water content at wilting in the North American

populations. There were no differences among low and high latitude populations or any

interaction between introduction and latitude impacting dehydration avoidance (Table S3).

Dehydration tolerance also differed among native and introduced ranges with native regions able

to better survive wilting ($X^2 = 7.09$, p = 0.012; Fig. 1B). All but one of the plants from the native

range were able to recover following wilting (11/12 plants), but only 46.7% (7/15) plants

recovered from the introduced North American range. There were no differences among low and

high latitude populations or any interaction between introduction and latitude impacting

dehydration tolerance (Table S2).

There are strong associations between the abiotic environment of site where each line originated

and wilt survival, but little correlation between abiotic variables and wilt VWC. Dehydration

tolerance was associated with aridity index, with plants from areas with greater annual aridity

index being more likely to survive wilting ($X^2 = 6.2$, p = 0.013; Fig. S5). This pattern was driven

by the low latitude populations from the native range that all recovered following wilting. These

low latitude populations, largely from Spain, had relatively high evapotranspiration, low annual

precipitation, and higher variation in precipitation. Dehydration avoidance was not significantly

associated with any abiotic variables (Fig S5.; Table S4).

There was a strong tradeoff between dehydration avoidance and tolerance. Plants that wilted at

lower VWC were less likely to recover following wilting (Adj. $r^2 = 0.53$, p = 0.005; Fig. 2). The

four plants wilting at the lowest volumetric water contents all did not survive following wilting.

Moreover, six of the seven plants with the lowest wilt VWC also did not survive wilting.

***Differential expression across the native and introduced ranges***

Many genes were differentially expressed between dry down and well-watered treatments, high

and low latitude populations, and native and introduced ranges (Table S5). Across predictor

variables, there were 19,344 differentially expressed transcripts out of 86,723 sequenced

transcripts corresponding to 2,253 of 5,001 unique genes. Most differentially expressed

transcripts were differentially expressed within all three contrasts (Treatment, Range, Latitude;

35.3%; Fig. S6). An additional 29.4% of transcripts were differentially expressed both in Range

and Latitude contrasts, not in the Treatment contrast. A far lower percentage of differentially

expressed transcripts were differentially expressed in only one contrast (Latitude: 13.4%;

Treatment: 5.6%; Range: 10.0%). The large union between the three contrasts in differentially

expressed transcripts indicates different drought responses are likely occurring across latitudinal

gradients and ranges.

We examined co-expression between differentially expressed genes to determine how gene

expression networks are influenced by drought treatment and vary across range and latitude.

Eight modules were discovered from WGCNA including 5.77% of transcripts (Table 1, Table

S5). Expression profiles of normalized transcript counts show the "turquoise" module as having

elevated transcript counts in samples within the dry-down treatment in contrast to all other

modules that exhibit relative uniformity across sample types (Table 1; Fig. S7). Gene enrichment

analysis of the "turquoise" module reveals marginal or significant enrichment for many GO

terms typically associated with drought resistance including: hyperosmotic stress, proline

metabolic process, response to oxygen-containing compound, and response to water deprivation

(Table S6). Within the "turquoise" module, 41 of the 53 genes (77.4%) exhibit a significant

latitude by range interaction (24 homologues exhibit significantly different range responses

between high and low latitudes) – substantially higher than the genes not in modules (18.2%;

Table 1). These results suggest that genes within the turquoise module may underlie adaptation

to climate across the native and introduce ranges.

**Differentially expressed genes are associated with dehydration avoidance and tolerance.**

Expression of a large number of genes was associated with wilt VWC and wilt survival (635 and

392, respectively; 198 shared; Table S6). The overlap between this group and genes that are

differentially expressed within the dry-down vs. well-watered contrast represent the best

candidate genes for impacting dehydration avoidance and tolerance strategies. A promising

number of genes are found in this overlapping group – 47 differentially expressed genes were

associated with wilt VWC and 17 differentially expressed genes with wilt survival (7 shared

between phenotypes). These genes disproportionately were found in the 'turquoise' module: 80%

of gene homologs for wilt VWC and 50% of gene homologs for wilt survival.

From this initial group, we identified several strong candidate genes by examining genes with

extreme effect sizes on each phenotype and are also known drought-associated genes in other

species. We first examined genes which were more highly expressed in plants with low wilt

VWCs as genes potentially involved in dehydration avoidance. Several genes from the turquoise

co-expression network had the greatest negative beta estimates for wilt VWC. These included

multiple homologs of *NAC4* and *BCAT2* (Fig. 3AB). Notably, expression of these genes was also

strongly associated with death following wilting, i.e. the most negative beta estimates for

dehydration tolerance. Several other strong candidate genes had this same strong tradeoff

between phenotypes including homologs of *AlaAT2*, *GOLS2*, *SDIR1* and *P5SC1* (Fig. 3CDEF).

In fact, all transcripts that had significant associations with both wilt VWC or wilt survival had

either negative beta for both wilt VWC and wilt survival or positive betas for both traits ($\alpha$

=0.05, 582 transcripts & 170 homologues; Fig. 4). We discuss gene functions in more depth in

the discussion, but wilt VWC-associated genes are strongly related to stress signaling via both

ABA-dependent and independent pathways, stomatal responses, and producing various

osmoprotectants.

Genes with expression strongly associated with wilt survival had functions more explicitly

associated with detoxifying reactive oxygen species and negative products of photorespiration as

well as continued light harvesting. The gene with the strongest association with wilt survival was

a homolog of *DegP5* – a gene that dampens $Ca^{+2}$ signaling associated with stress responses and

is associated with reduced ROS (Fig. 5A). This gene also had the strongest positive association

with wilt VWC of any gene, that is, high expression of this gene is linked with wilting at a high

VWC. Expression of a *CUT1* (*Cutinase 1*) homolog, a gene that impacts cuticle wax

biosynthesis, was also strongly associated with greater survival following wilting. One additional

candidate gene, a homolog of *SAPK2*, also had among the highest associations with increased

wilt survival and higher wilt VWC (Fig. 5C). Other genes, including homologs of *PGLP2*,

*CAB7*, and *LHCA6*, with expression strongly associated with wilt survival are more weakly

associated with wilt VWC. These genes have less clear associations with known drought

responses. Together, these candidates suggest that genes associated with lower wilt VWC

include those typically associated with dehydration avoidance, while genes associated with wilt

survival are associated with increasing photosynthesis and protecting tissues against the harmful

bioproducts of stress responses.

**Expression patterns associated with phenotypic differences between ranges.**

While many genes are associated with dehydration avoidance and tolerance phenotypes, a

limited subset of these candidates have the potential to drive the differences in dehydration

avoidance or tolerance between ranges. We examined which of the candidate genes linked to

dehydration avoidance or tolerance also had differential expression across ranges or latitudes. As

is the case with many genes in the turquoise network, relative expression of *BCAT2* in the

drought treatment is greater in high latitude populations, exhibits variable expression in low

latitude North American populations, and has very limited expression in the low latitude

European populations (Range:Latitude: $\log_2$FoldChange = 30, P = 4.83e-9; Fig. 3, Fig. S8).

Similar patterns exist for both *P5CS* (Range:Latitude: $\log_2$FoldChange = 29.9, P = 8.28e-11) and

*GOLS2* (Range:Latitude: $\log_2$FoldChange = 18.5, P = 0.002). To determine other genes that may

create phenotypic differences between ranges or across latitudes, we identified genes whose

relative expression is best explained by an interaction between wilt VWC or wilt survival and

range. Many genes (22.8% of genes surveyed; Table S8) exhibited such interactions including 46

of the 52 genes in the turquoise network. These results suggest the molecular variation

underlying phenotypic differences between ranges either involves many loci of varying effect

sizes or are large effect variants that impact substantial hormonal stress responses and many gene

expression networks.

DISCUSSION

During invasions, plants encounter novel environments and may need to acclimate or adapt to

succeed. Here we identify differences in drought resistance strategies that have evolved

following the introduction of white clover to North America from its native range in Europe.

North American populations have stronger dehydration avoidance responses while European

populations have stronger dehydration tolerance responses. These differences appear to be

shaped by the abiotic conditions in each region as well as a strong tradeoff between dehydration

avoidance and tolerance. We also identify a number of strong candidate genes underlying each of

these phenotypes with a significant proportion falling into a single co-expression network. Most

of these genes have expression profiles where higher expression improves either dehydration

avoidance or tolerance while decreasing the other phenotype. Additionally, the limited subset of

genes that are associated with both a drought resistant phenotype and exhibit differential

expression between native and introduced ranges represent the best candidates for identifying a

molecular basis of drought adaptation following introduction. We discuss these results below in

the context of invasion biology and the evolutionary genetics of adaptation to plant abiotic

stresses: two fields that overlap but are rarely synthesized.

**Evolution of Drought Resistance During Invasions**

The roles of selection and adaptation during invasions is now widely documented with many

species, including white clover, rapidly evolving climate-associated clines following introduction

(e.g. Kooyers & Olsen, 2012; Colautti & Barrett, 2013; Battlay *et al.*, 2023). However,

generalization of which traits are favored during invasions is less clear (Kolar and Lodge 2001).

Different trait-based hypotheses suggest that rapid growth (EICA; Blossey & Notzold, 1995) or

stress tolerance may be favored in the introduced regions (Gewing *et al.* 2019). Our results are

partially consistent with both hypotheses, but are far more nuanced as drought resistance is

conferred through many different mechanisms. Introduced North American populations wilt at

lower volumetric water contents than populations from the native European range, but also are

less able to tolerate wilting than native populations (Fig. 1). Thus, North American populations

are likely more dehydration avoidant than European populations. Greater dehydration avoidance

may allow plants to maintain growth during periods of low water and high evapotranspiration.

This would be consistent with devoting more resources to rapid growth within the introduced

range as well as being more resistant to initial water stress. This nuance may account for some of

the mixed evidence for the EICA within the literature (Rotter and Holeski 2018).

We suggest that the evolution of greater stress avoidance to facilitate rapid growth may be a

pattern frequently observed across invasions of perennial plants. Introduced species are likely to

encounter different abiotic regimes in their new range. Invasive populations of annual plants

often evolve more rapid life histories (Colautti and Barrett 2013; Battlay *et al.* 2023), but

perennial plants must survive stresses in order to establish and reproduce. There are few plant

systems where the evolution of drought avoidance or tolerance have been assessed across native

and introduced regions. These examples largely consist of species where it is clear that they have

a very different drought resistance strategy in the introduced range. For instance, camphorweed

(*Heteroteca subaxcillaris*) has drastically greater root growth in its invasive range in Israel than

in its native range, North America, that likely is a benefit both for low water availability and

structural integrity in extremely arid dune environments in Israel (Sternberg 2016). Jerusalem

artichokes (*Helianthus tuberous*) have evolved extreme clonality phenotypes in their introduced

range (Europe) to accommodate a shift to a more aquatic environment (Bock *et al.* 2018).

Outside of plants, a greater tolerance or plasticity to stressful environments is often observed –

for instance, anoles (*Anolis cristatellus)* from the introduced range survive colder temperatures

than from the native range (Leal and Gunderson 2012). We suggest that invasive species should

be more widely used for identifying natural phenotypic and genomic variation in drought

resistance.

While our results are consistent with classic invasion biology theory, they also make sense in the

context of environmental characteristics of the populations. The populations involved in this

experiment have very different growing seasons and historical climatic features (Table S1).

Southern European populations come from a Mediterranean climate with cool and temperate

winters and hot dry summers. Relative to other locations, there is little precipitation throughout

the year and almost no rainfall during summer months. A long summer with little rain is the ideal

conditions for the evolution of dehydration tolerance in a perennial plant that likely needs to

survive across multiple growing seasons (Kooyers 2015). Alternatively, eastern North America

has more substantial rainfall throughout the growing season with drought stress likely coming

during shorter stretches without rain (Table S1). These conditions may favor the evolution of an

avoidance strategy as additional rainfall is likely coming in the near future. Interestingly, we

observe very little difference in drought resistance strategy between high and low latitude

populations of white clover in the native or introduced ranges. In both ranges, on average, lower

latitude populations are less dehydration avoidant but more dehydration tolerant. This is

surprising as previous work documents differentiation in life history strategy evolving across this

gradient with warmer southern populations employing a more annualized life history (Wright *et*

*al.* 2021). We suggest our small sample size only has the power to detect major effects and future

larger studies may be able to parse differences in stress responses across latitudinal gradients.

**Molecular basis of drought tolerance and avoidance variation**

The molecular basis of drought resistance has been examined extensively in crop plants and

model systems, but there has been limited success in determining the genes that underlie

variation in natural populations (Des Marais *et al.* 2013; VanWallendael *et al.* 2019). Here we

find several candidate genes that likely influence dehydration avoidance and tolerance responses

across both invasive and native ranges despite relatively few mapped reads from only a single

tissue type (Fig. 3). Notably the turquoise co-expression network is strongly enriched for gene

ontologies related to drought responses (i.e. response to abiotic stress, response to water and

proline synthesis). A high proportion of the genes within this network are also associated with

wilt VWC including some of the strongest candidate genes. Interestingly, these genes represent a

diverse set of drought-related functions in other organisms.

*NAC4* and *SDIR* homologs are likely involved in stress signaling pathways. NAC4 (nascent

polypeptide-associated complex) is a transcription factor that is strongly upregulated in response

to drought, salt, and water stress in many species (Xia *et al.* 2010; Shao *et al.* 2015; Yu *et al.*

2016) and confers drought tolerance when transgenically expressed in *Arabidopsis* (Mei *et al.*

2021). More generally, the NAC gene family is involved in stomatal closure and in lateral root

development in rice (Hu *et al.* 2006; Zheng *et al.* 2009). SDIR1 (Salt and Drought-Induced Ring

Finger1) is a RING-finger E3 ligase that is upregulated in *Arabidopsis* in response to drought

stress and modulates both the ABA response and stomatal closure (Zhang *et al.* 2007). While

upregulation of both genes in response to drought stress in our system is consistent with findings

in other species and expected for dehydration avoidant plants, *sdir1* mutants in *Arabidopsis* had

greater mortality following wilting than wild type plants. This suggests an additional dehydration

tolerance role for SDIR1 that we do not observe.

Expression of multiple genes that promote the synthesis of osmoprotectants (i.e. branched chain

amino acids or BCAAs, proline, and raffinose family oligosaccharides) are also strongly

associated with lower wilt VWC. *BCAT2* (BCAA transaminase 2) catalyzes the final

transamination step in production of BCAAs as well as the first degradation step. Under stress,

BCAAs accumulate as osmoprotectants in an ABA-dependent manner, with BCAT serving as

the main regulator in *Arabidopsis*. P5CS, delta 1-pyrroline-5-carboxylate synthase, controls the

rate-limiting step of proline biosynthesis from glutamate during osmotic and salt stress (Strizhov

*et al.* 1997). Transgenic overexpression of P5CS consistently increases tolerance to drought and

salt stress across several plant species by increasing proline content (Su and Wu 2004; Yamchi *et*

*al.* 2007; Vendruscolo *et al.* 2007). *GOLS2* (galactinol synthase 2) encodes galactinol synthase 2,

a key enzyme that regulates the first step in biosynthesis of raffinose family oligosaccharides

resulting in the creation of galactinol (Panikulangara *et al.* 2004). While the oligosaccharides act

as osmoprotectants and as antioxidants, both galactinol and the oligosaccharides may also act as

signaling molecules (Song *et al.* 2016). In other species, GOLS activity is triggered by abiotic

and biotic stress (*Arabidopsis*, Coffee, Rice) and confers drought tolerance (Dos Santos and

Vieira 2020). Unexpectedly, our results indicate that expression of *GOLS2* is linked with lower

wilt VWC (greater dehydration avoidance) and with lower survival following wilting (lower

dehydration tolerance). Increased expression of *GOLS2* in *Arabidopsis* does reduce transpiration

rate (Taji *et al.* 2002).

We also note a third process that may enhance dehydration avoidance and tolerance responses:

the recycling and efficient use of nitrogen. *AlaAT2* (alanine aminotransferase 2) is strongly

upregulated in plants that wilt at low VWC. We propose that this upregulation links drought

response to nitrogen metabolism. Specifically, AlaAT2 catalyzes the reversible conversion of

alanine and α-oxoglutarate to pyruvate and glutamate, facilitating the synthesis of other amino

acids as needed (Miyashita *et al.* 2007). *AlaAT* homologs are upregulated during hypoxia in

several species, including *Arabidopsis*, Poplar, Medicago, and Corn, and are regulated by ABA

in other stressful contexts (Ricoult *et al.* 2005; Miyashita *et al.* 2007; Xu *et al.* 2017). Drought

stress typically decreases the activity of nitrate reductase and nitrile reductase (Foyer *et al.*

1998), impairing the plant's ability to accumulate amino acids due to limited nitrogen resources.

Greater AlaAT2 activity could convert alanine into glutamate, which may then be used to

produce BCAAs or proline as osmoprotectants. In sum, we hypothesize that the production of

osmoprotectants requires carbon and nitrogen resources that may be most easily procured

through recycling of existing metabolites.

The candidate genes associated with wilt survival generally do not have strong functional ties to

drought responses in other systems with two exceptions. *DegP5* (degradation of periplasmic

proteins 5) belongs to a family of proteases that is typically upregulated in response to abiotic

stress. Overexpression of *DegP5* has been linked to increased dehydration tolerance in *Nicotiana*

*tabacum* via the suppression of $Ca^{+2}$ and flagellin signaling leading to a reduced accumulation of

reactive oxygen species (Chen *et al.* 2023). In our study, increased expression of *DegP5* is also

associated with wilting at higher soil moistures. This suggests that the increase wilt survival

comes at a cost of a less active drought response. The second exception is *SAPK2*, a plant-

specific protein kinase in the family of SNF1-related protein kinase 2 (*SnRK2*) genes. Expression

of the *SAPK2* homolog is positively associated with wilt survival (Fig. 5C). This role is similar to

that documented in rice, where SAPK2 not only upregulates genes involved in osmoprotection

but also induces genes involved in enzymatic ROS detoxification (Lou *et al.* 2017). However, a

role in dehydration avoidance has not been documented. Other SnRK2s do regulate cation

channels (KAT1) and impact stomatal closure in *Arabidopsis* (Sato *et al.* 2009). Other genes

strongly associated with high wilt survival in our study function in other organisms in the

detoxification of photorespiration byproducts (*PGLP2*; Schwarte & Bauwe, 2007) and in

efficient light harvesting and photosynthesis (*LHCA6*, *CAB7*; Otani *et al.*, 2017). Together, these

results suggest that improved survival after wilting is achieved through a combination of

dampening responses to drought, protecting cells from harmful bioproducts of drought stress,

and maintaining metabolic functions.

**A tradeoff between drought avoidance and tolerance strategies?**

Drought resistance strategies are often presented as non-mutually exclusive with the recognition

that they represent different physiological mechanisms for a plant to acclimate to low water

availability. The relationship between drought escape and dehydration avoidance has been the

most extensively scrutinized as there are clear physiological tradeoffs for C3 plants – closing

stomata for better dehydration avoidance leads to lower carbon uptake and the inability to

synthesize more sugars for rapid growth and reproduction (Geber and Dawson 1990; Dudley

1996). Theoretical and empirical identification of tradeoffs between dehydration avoidance and

tolerance is rare – potentially because there is not a true physiological tradeoff that exists or

because these strategies may be expressed sequentially as water stress increases (Volaire 2018).

We document relatively strong tradeoffs between wilt VWC and wilt survival (Fig. 2) and

expression responses for wilt VWC and wilt survival are strongly correlated (Fig. 4). However,

the functions of genes associated with each of these phenotypes suggest that our experiment is

not truly measuring separate dehydration avoidance and tolerance responses. Rather, our wilt

VWC treatment is likely catching both dehydration avoidance and tolerance responses – plants

that survive to lower wilting *are* effectively tolerating dehydration for longer than plants that

wilted earlier, likely by increasing osmoprotectants as well as closing stomata to regulate gas

exchange. Correlated dehydration avoidance and tolerance responses make sense as both

functions are at least partly regulated by ABA. However, since dehydration tolerance responses

often involve the production of various osmoprotectants and closing stomata limits $CO_2$ (and

useable plant nitrogen) uptake, correlated physiological responses must require recycling carbon

and nitrogen.

The strong correlation between wilt VWC and wilt survival more likely represents a tradeoff

between different elements of dehydration tolerance. Wilting occurs when the turgor pressure

within cells falls to zero, leading to potential cellular damage. This damage is exacerbated under

prolonged drought stress, as increased ROS production can overwhelm the plant's antioxidant

defenses and, coupled with stomatal closure, can also increase photorespiration flux (Noctor *et*

*al.* 2002). To survive wilting, plants must be able to protect cellular function and integrity. Our

study found that plants with better wilting survival had higher expression of genes associated

with antioxidant defense (*DegP5*) and detoxification of photorespiration byproducts (*PGLP2*).

While expression of each of these genes was associated with high wilt VWC, the only truly

strong correlation was for *DegP5* – a gene linked to diminishing $Ca^{+2}$ signaling as well as lower

ROS production in tobacco (Chen *et al.* 2023). Expression of this gene may represent a true

tradeoff between different physiological strategies. Generally, our results suggest that a

dehydration tolerance strategy actually represents multiple different types of tolerance – one

strategy that provides resistance against dehydration and another that allows longer-term survival

following wilting (i.e. extreme dehydration). We hypothesize that dehydration tolerance related

to drought survival may have overlapping physiology with desiccation tolerance strategies

occurring in plants like resurrection fern. In sum, connecting variation between physiological

processes and plant performance will provide additional insight into the evolution of different

ecological strategies in specific environments.

**Loci underlying drought response adaptation following introduction**

Identifying the causative molecular variants that underlie natural intraspecific variation in

drought responses has proven challenging with limited of successes (Juenger 2013;

VanWallendael *et al.* 2019). This objective is largely beyond the scope of our study. We do

identify many genes that are both linked with wilt VWC or wilt survival phenotypes and also

show variable expression across ranges, conditions and latitudes. This overlap includes a

substantial portion of the turquoise gene co-expression network, suggesting that variation in

dehydration avoidance and tolerance phenotypes is physiological manifested through broad shifts

across gene expression networks (e.g. Lovell *et al.*, 2018) rather than through simple, large effect

variants that directly cause lower stomatal conductance or greater osmoprotection (e.g. Kesari *et*

*al.*, 2012; Des Marais *et al.*, 2014).

Recent work has identified several haploblocks that underlie climate adaptation following

introduction to North America as well as several candidate genes within haploblocks that were

differentially expressed across drought treatment, range and latitude (Battlay *et al.* 2024). Within

the three haploblocks on Chr04_Occ (syn. Chromosome 7), 26 differentially expressed

transcripts were found (9 on hb7a1; 14 on hb7a2; and 3 hb7b, Table S9). On hb7a1, 4 genes

within the turquoise submodule were differentially expressed. Two turquoise submodule genes,

*P5CS1 and FER1*, were strongly correlated with wilt VWC and wilt survival. In addition,

*BRR2A* and *DWF5* were correlated significantly with wilt survival whereas *IAA6* and *DWF5*

were correlated significantly with wilt VWC. On hb7b, transcripts belonging to the gene *CCL12*

are significantly correlated with wilting. Multiple of these genes have allele variation associated

with higher fitness in North American common gardens (Albano et al. in prep; Battlay *et al.*,

2024). These results are a first step to tying locally adaptive genetic variation to variation in gene

regulatory networks, physiology and ecological strategies.

*Conclusions*

Our work suggests that selection acts on rapid timescales following introduction to allow more

precise adaptation of drought responses to divergent environmental conditions. Specifically,

selection favors specific drought resistance strategies, with tradeoffs occurring between

strategies at both phenotypic and molecular levels. We identify a co-expression network that

underlies both dehydration avoidance and some elements of dehydration tolerance where many

of these loci also exhibit differential expression patterns between the native and introduced

ranges of white clover. We also identify tradeoffs at the molecular level among functions

(osmoprotection and detoxification) typically identified within the dehydration tolerance strategy

– a result that needs to be carefully considered in breeding efforts. This work stimulates more

studies examining and relating genetic variation and expression networks to phenotypic

differences within natural populations to create a broader consensus on mechanisms creating

variation in drought resistance and survival.

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DATA AVAILABILITY

Phenotypic data from the growth chamber experiment is available via dryad: link. RNAseq data

is available via NCBI's short read archive: link.

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TABLES

Table 1: Breakdown of the genes and module associations within phenotypes and across space

| Module | # Transcripts | Wilt VWC | Wilt Survival | Latitude | Range | Latitude X Range |
|---|---|---|---|---|---|---|
| No-Module | 45916 (4360) | 2066 (557) | 1201 (336) | 2994 (854) | 1076 (309) | 3895 (875) |
| In-Module | 4337 (662) | 346 (89) | 240 (62) | 603 (169) | 170 (51) | 592 (130) |
| *brown* | 2298 (428) | 158 (50) | 110 (29) | 426 (134) | 97 (28) | 221 (54) |
| *red* | 122 (36) | 8 (1) | 7 (2) | 20 (5) | 5 (1) | 15 (4) |
| *yellow* | 259 (45) | 3 (1) | 7 (1) | 16 (5) | 3 (2) | 50 (9) |
| *grey* | 821 (120) | 52 (6) | 28 (5) | 37 (8) | 34 (11) | 36 (6) |
| *turquoise* | 285 (53) | 86 (28) | 59 (15) | 70 (12) | 8 (0) | 196 (41) |
| *green* | 138 (31) | 5 (1) | 5 (2) | 8 (2) | 8 (4) | 7 (1) |
| *blue* | 324 (86) | 27 (5) | 21 (8) | 19 (8) | 15 (6) | 51 (20) |
| *black* | 91 (20) | 7 (0) | 3 (0) | 7 (1) | 0 (0) | 16 (5) |

907    Number outside of parentheses refers to the number of transcripts that are associated with the

908    phenotype or differentially expressed across Latitude or Range contrasts. The number within the

909    parentheses refers to gene homologues that are associated with the phenotype or differentially

910    expressed across Latitude or Range contrasts.

FIGURE LEGENDS

**Fig. 1** Variation in drought strategies between native and introduced populations of white clover.

(A). Boxplots of variation in wilt VWC within low and high latitude population in the native

(European) and introduced (North American) regions. Each point is an individual. Box edges

represent the interquartile range, the center line in the box is the median, and the whiskers

represent 1.5 times less or greater than the interquartile range. (B). Barplots identify the

proportion of individuals that survived the wilting treatment from each latitude:region contrast.

Numbers above the bars are the number of individuals surveyed.

**Fig. 2** Phenotypic tradeoff between dehydration avoidance and tolerance. Lower wilt VWC

indicates greater dehydration avoidance while higher wilt survival indicating greater dehydration

tolerance.

**Fig. 3** Association between candidate gene expression and dehydration avoidance and tolerance

phenotypes. Lower wilt VWC indicates greater dehydration avoidance while higher wilt survival

indicating greater dehydration tolerance. Points and bar are colored by latitude and range: low

latitude Europe = blue, high latitude Europe = red, low latitude North America = purple, High

Latitude North American = orange. Each point represents a different individual. Gene

Abbreviations: *NAC4* = nascent polypeptide-associated complex, *BCAT2* = branched chain

amino acid transaminase 2, *AlaAT2* = alanine aminotransferase 2, *GOLS2* = galactinol synthase

2, *SDIR1* = Salt and Drought-Induced Ring Finger1, *P5CS* = delta 1-pyrroline-5-carboxylate

synthase.

**Fig. 4** Correlations between model beta estimates from wilt VWC and wilt survival associations with RNAseq. Each point represents estimates for one transcript. Transcripts with purple dots have significant association of expression with both phenotypes, blue dots have significant association of expression with wilt VWC, red dots with wilt survival and black dots with neither phenotype.

**Fig. 5.** Candidate genes associated with wilt survival. Lower wilt VWC indicates greater dehydration avoidance while higher wilt survival indicates greater dehydration tolerance. Points and boxplots are colored by latitude and range: low latitude Europe = blue, high latitude Europe = red, low latitude North America = purple, High Latitude North American = orange. Each point represents a different individual. Gene Abbreviations: *DeGP5* = degradation of periplasmic proteins 5; *CUT1* = Cutinase 1; *SAPK2* = sucrose non-fermenting 1-related kinase 2.

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SUPPLEMENTAL INFORMATION

**Table S1**. Geographic and environmental characteristics of white clover populations.

**Table S2.** Mapping statistics from RNAseq experiment

**Table S3.** Univariate model results examining associations between drought strategy phenotypes

across ranges, latitudes, and treatments.

**Table S4.** Univariate model results examining associations between drought strategy phenotypes

with water-availability related variables of collection populations

**Table S5**. Summary of differentially expressed genes with associated modules and phenotypic

associations.

**Table S6.** Gene Ontology analysis of WGCNA modules

**Table S7.** Genes with variation in expression associated with wilt VWC or wilt survival

**Table S8.** Genes with significant range:phenotype interactions.

**Table S9.** Patterns of expression within haploblocks

**Fig. S1.** Map of sampling locations.

**Fig. S2.** Volumetric water content of pots within well-watered and dry down treatments

throughout the experiment.

**Fig. S3.** Relationships between drought strategy phenotypes and leaf number at the start of the

experiment.

**Fig. S4.** Associations between water-availability related variables of collection populations and

leaf number at the start of the experiment

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A. *DEGP5* B. *CUT1* C. *SAPK2*

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North America High Latitude
Europe Low Latitude
North America Low Latitude

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