AI-enabled Alkaline-resistant Evolution of Protein to Apply in Mass Production

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Abstract: Artificial intelligence (AI) models have been used to study the compositional regularities of proteins in nature, enabling it to assist in protein design to improve the efficiency of protein engineering and reduce manufacturing cost. However, in industrial settings, proteins are often required to work in extreme environments where they are relatively scarce or even non-existent in nature. Since such proteins are almost absent in the training datasets, it is uncertain whether AI model possesses the capability of evolving the protein to adapt extreme conditions. Antibodies are crucial components of affinity chromatography, and they are hoped to remain active at the extreme environments where most proteins cannot tolerate. In this study, we applied an advanced large language model (LLM), the Pro-PRIME model, to improve the alkali resistance of a representative antibody, a VHH antibody capable of binding to growth hormone. Through two rounds of design, we ensured that the selected mutant has enhanced functionality, including higher thermal stability, extreme pH resistance and stronger affinity, thereby validating the generalized capability of the LLM in meeting specific demands. To the best of our knowledge, this is the first LLM-designed protein product, which is successfully applied in mass production.

1. Introduction

Protein engineering, situated at the nexus of molecular biology, bioinformatics, and biotechnology, focuses on the design of proteins to introduce novel functionalities or enhance existing attributes[1-3]. With the exponential growth of biological data and computational power, protein engineering has experienced a significant shift towards advanced computational methodologies, particularly deep learning, to expedite the design process and unravel complex protein-function relationships[4-9]. However, a significant challenge in industrial protein engineering is designing proteins with inherent resistance to extreme conditions, such as high temperature and extreme pH environments (acidic or alkaline)[10, 11]. Unlike proteins in natural ecosystems, those used in industrial processes often encounter harsh physical and chemical conditions, necessitating exceptional resilience to maintain functionality[12, 13]. Previous efforts to enhance protein resistance have often relied on rational design and mutant library screening. These methods are typically labor-intensive, inefficient, and yield limited improvements[14-17]. Consequently, the industrial demand for proteins resilient to harsh environments poses a notable absence within the training datasets of Artificial Intelligence (AI) models. Exploring whether AI can achieve the evolution of protein resistance to extreme environments is crucial for broadening protein applications and improving modification efficiency.

Recent advances in large-scale protein language models (LLMs) have enabled zero-shot predictions of protein mutants based on self-supervised learning from natural protein sequences[18-21]. Although AI-guided protein design has been applied to predict the mutants with greater thermostability and higher activity[22-24], it is unexplored whether these models based on the natural protein information can find the mutants that adapt the unnatural extreme environments, such as the alkaline solution with the pH value higher than 13.

Here, we employed a LLM (large language model) developed by our group, the Pro-PRIME model[25], to predict dozens of mutants of a nano-antibody against growth hormone (a VHH antibody), and examined their fitness,

including alkali resistance and thermostability, to evaluate their performance under extreme environments. We

utilized the Pro-PRIME model to score saturated single-point mutations of the VHH in a zero-shot setting, and

selected the top 45 mutants for experimental testing. Some mutants exhibited improved alkali resistance, while

others demonstrated higher thermal stability or affinity. Subsequently, we fine-tuned the Pro-PRIME model to predict

dozens of multi-point mutations. As a result, we obtained three multi-point mutants with enhanced alkali resistance,

higher thermostability, as well as strong affinity to the targeted protein. Also, the dynamic binding capacity of the

selected mutant did not show significant decline after more than 100 cycles, making it suitable for practical

application in industrial production. The selected mutant has been used in practical production and lower the cost for

over one million dollars in a year. To the best of our knowledge, this is the first protein product developed by a LLM

that has been successfully applied in mass production. Due to the Pro-PRIME model's ability to achieve precise

predictions of multi-point mutations with reliance on a small amount of experimental data, our two-round design

process involved experimental validation of only 65 mutants in two months, demonstrating remarkable high

efficiency. Furthermore, we performed a systematic analysis of these findings and determined that the model can

yield more valuable predictive outcomes while remaining consistent with rational design principles. Specifically,

within the framework of multi-point combinations, the model's incorporation of negative single-point mutations into

the combinatorial space led to exceptional results, showcasing its capacity to capture epistatic interactions. Notably,

in striving for global optimum, deep learning methods offer distinct advantages over traditional rational design

approaches.

 2. Results and Discussion

 2.1. Wet experimental testing of single-point mutants

Currently, the prevailing approach in protein engineering involves rational design or high-throughput screening to

identify positive single-point mutations, followed by their combination into more effective multi-point mutations

through greedy search methods[26]. However, these approaches are inclined to trap in a local optimum and unable

to avoid the negative epistasis[27, 28]. To overcome these limitations, we employed the Pro-PRIME model to screen

single-point mutations in the first round, enabling rapid inference of scores for saturated mutations. An efficient

strategy involves validating only a select number of top-ranked mutants, thereby saving considerable time and

reducing economic costs compared to rational design and high-throughput experiment. Moreover, we posit that

deep learning models can capture nuanced features during training, and recommend single-point mutations that

expert experience may not cover. These mutants expand possibilities for subsequent combination of multi-point

mutations and mitigate the risk of converging into local optima.

Specifically, we made use of the Pro-PRIME model to score the single-site saturated mutants, and selected the top

45 single-point mutants of the VHH antibody for testing. Pro-PRIME is a deep learning-based methodology developed

to guide protein engineering, utilizing masked language modeling (MLM) and multi-task learning to study and

comprehend the semantic and grammatical features inherent in protein sequences, and further capture the

temperature traits associated with these sequences. A higher score corresponds to a greater appearance probability

of the residue at this site in the Pro-PRIME model[25]. The alkali resistance of a mutant was determined by its half

maximal effective concentration (EC50) after treatment with alkali. A lower EC50 represents that antibodies have

stronger affinity. Firstly, the 45 single-point mutants were treated with 0.3 M NaOH for 24 hours, and evaluated by

their thermal stability and affinity. The results revealed that 15 mutants exhibited higher alkali resistance, and 35

mutants displayed higher melting temperatures (T_m) (Figure 1A). We also noted that 8 out of 45 single-site mutants

designed by Pro-PRIME showed improved affinity as compared to the WT before alkali treatment (Figure S1). It is

worth noting that six mutants (A57D, A15P, V113D, P117Q, R20T, and L12K) exhibited enhancements in all three

properties, i.e., higher alkali resistance, greater thermal stability, and stronger affinity before alkali treatment.

Additionally, eight mutants (P29T, N85Q, G134R, N78S, R110E, L49V, T58K, and W112F) exhibited improved alkali

resistance and T_m, albeit at the moderate cost of affinity. This trade-off is acceptable, as excessively high affinity could

complicate the separation of growth hormone from the VHH. Furthermore, we subjected single-point mutants with

remarkable alkali resistance to treatment with 0.5 M NaOH, and measured their affinity after 24 hours. Our findings

demonstrated that four mutants (L49V, A15P, W112F, and R20T) exhibited higher affinity than the wild type following

the same treatment protocol (Figure 1B). Interestingly, these four single-point mutations all showed improvements in

T_m, and their affinities before alkali treatment were either similar to or lower than those of the wild type. However,

this does not imply that enhancing thermal stability will concurrently improve alkali resistance. The spearman

correlation between the EC50 values after 0.3 M NaOH treatment and T_m values of single-point mutants is -0.29,

indicating that these two properties of the antibody are only weakly correlated (Figure S3). Therefore, achieving

multi-point mutants that enhance both alkali resistance and thermal stability remains a challenge.

Figure 1. The experimental results from two rounds of design. (A) Experimental results of single-point mutants. The melting temperatures of mutants are shown as red squares. The affinity of mutants after treatment with 0.3 M NaOH for 24 hours are shown as blue bars. (B) Experimental results of multi-point mutants and single-point mutants. The melting temperatures of mutants are shown as red squares. The affinity of single-point mutants after treatment with 0.5 M NaOH for 24 hours are shown as blue bars, while those of multi-point mutants are shown as green bars. The affinity values were normalized in the graph, with the wild-type EC50 set as 1.

2.2. Multi-point mutation design driven by experimental data

In the process of combining multiple single-site mutations, common greedy algorithms typically proceed by sequentially adding the most effective single-point mutations and gradually increasing the number of stacked mutations. However, this approach often demonstrates low efficiency and is susceptible to becoming trapped in local optima. Importantly, the most effective multi-point mutations in practical scenarios may not necessarily include the most effective single-point mutations[28]. Moreover, in the traditional approach of incrementally stacking mutations from single points to multiple points, only paths that yield better results at each step are retained[27]. Nevertheless, combinations that are discarded due to decreased effectiveness may significantly enhance performance in subsequent stacking step, a phenomenon unforeseeable in traditional methods.

Although the model inference is fast, it is not feasible to explore all possible mutations when designing

multi-point mutants due to the exponential increase in the number of potential combinations. To manage this

challenge, we constructed a mutant library based on a two-stage design process. In the first stage, we scored all

single-point mutations using the model, and in the second stage, we combined experimentally validated single-point

mutations to create the multi-point mutant library. This approach ensures that even when designing multi-point

mutants (e.g., five-point mutants), the number of mutants to score remains in the millions, which is computationally

efficient and practical. The number of single-point mutations selected for the multi-point mutant library is a key

factor influencing both the computational load and the scope of the design space. To maintain a balance between

efficiency and accuracy, we limited the number of single-point mutations to between 30 and 50. This strategic

approach allows us to achieve both scalability and precision in our protein engineering tasks.

To better search the global optimium, we chose the Pro-PRIME model to predict the results of multi-point mutants

due to its remarkable performance on the design of diverse proteins[25]. We trained two Pro-PRIME models using

the alkali resistance and T_m data of single-point mutants, respectively, to score these two properties of mutants.

During the prediction of multi-point mutations, we prioritized mutants with higher alkali resistance scores while

ensuring T_m scores were not lower than those of the wild type. Ultimately, we selected 20 multi-point mutants for

experimentation (see details in Supporting Information, SI). Experimental results indicated that the alkali resistance

of five multi-point mutants (A57D;P29T, A57D;D114Y;W112F, A15P;R20T, A15P;P117Q, and A57D;G134R;D114Y)

surpassed that of the best single-point mutant. The T_m of multi-point mutants was generally higher, with the highest

(P29T;A15P) exceeding that of the wild type by approximately 10 °C (Figure 1B). Although we did not optimize for the

affinity of the VHH antibody, certain multi-point mutants (A15P;R20T, A15P;G134R, and P29T;G134R) exhibited

affinity levels close to or even exceeding those of the wild type (Figure S2). Although experimental results from

single-point mutations suggested that simultaneously improving the alkali resistance and thermal stability of VHH

might be challenging, we successfully designed multi-point mutants that balance multiple properties. This

demonstrates the excellent multi-objective optimization capability of the Pro-PRIME model.

Note that we employed different strategies for designing single-point and multi-point mutants, specifically using a

zero-shot approach for single-point mutations and fine-tuning the model for multi-point mutations. These choices

were made based on the distinct characteristics of the two tasks and the availability of experimental data. For

single-point mutations, the number of possible mutations is relatively limited, and at the outset, there were no

experimental data available. In such cases, the zero-shot setting was chosen because it allows the model to predict

the fitness of mutants based solely on the information learned during pre-training on a large protein sequence

dataset. Since single-point mutations are computationally manageable, this approach was deemed appropriate to

generate initial predictions for protein engineering. However, when designing multi-point mutants, the situation

changes significantly. The potential combinations of mutations increase exponentially, and without prior data, it

becomes computationally infeasible to evaluate every possible combination within a reasonable timeframe.

Moreover, by the time we reached the multi-point mutation design stage, experimental data for several single-point

mutations had already been obtained. This data enabled us to fine-tune the model to better capture the specific

structural and functional features that contribute to protein stability and resistance, especially in the context of

multiple interacting mutations. Fine-tuning improves the model's accuracy by adjusting its parameters to align more

closely with the experimental data, ensuring that the predicted multi-point mutants are more likely to meet the

desired engineering goals. After the second round of design, the fitness of the mutants was further improved. In

improving alkali resistance, experimental results showed that 15 of the 45 designed mutants exhibited positive

responses, yielding a success rate of 30%, close to the 35% success rate achieved in the second round. Compared to

the wild type, the best single-point mutant improved alkali resistance by approximately 44.7%, while the best

multi-point mutant achieved a 67.7% increase. For thermal stability enhancement, the success rate in the first round

was 77.8%, rising to 100% in the second round. The top single-point mutant exhibited a Tm increase of 6.37°C over

the wild type, while the best multi-point mutant had a Tm increase of 10.02°C. We also tested the performance of

the zero-shot approach for multi-point mutants, and the results showed that this method did not yield satisfactory

predictions. The Spearman correlation coefficient between the zero-shot predictions and experimental results for

multi-point mutants was -0.71, indicating a significant discrepancy. This further highlights the importance of

fine-tuning the model for multi-point mutations, as the fine-tuned model provided more accurate and reliable results.

In summary, the choice of zero-shot for single-point mutations and fine-tuning for multi-point mutations was driven

by practical considerations regarding computational feasibility and the availability of experimental data. Fine-tuning

the model significantly enhances its predictive performance, particularly for complex multi-point mutations where

multiple residues interact. We believe this strategy strikes an optimal balance between computational efficiency and

predictive accuracy, making it well-suited for practical protein engineering applications.

2.3. Complex epistatic effects generated by the combination of single-point mutations

To understand the principle of the evolution, we analyzed the improvement in multi-point mutants across three

different indicators. As illustrated in Figure 2B, the evolution of thermal stability in the second round was successful,

with nearly all multi-point mutants exhibiting an increase in T_m compared to the best single-point mutant they

contain. Three multi-point mutants demonstrated higher affinity than the wild type, but only P29T and G134R

30 exhibited positive epistasis, resulting in the creation of a double-point mutant with superior affinity. Although

31 A15P;R20T and A15P;G134R showed higher affinity than the wild type, these combinations did not contribute to

32 affinity improvement because the A15P single-point mutation they contained exhibited stronger affinity compared to

33 these double-point mutants.

Figure 2. Schematic diagram illustrating the combined effects of single-point mutations. The colors (outer contours representing multi-point mutations and inner solid circles representing the included single-point mutations) and numerical values represent the EC50 values after alkali treatment with 0.5 M NaOH for 24 hours, the EC50 values before alkali treatment, and the Tm values. Blue indicates mutations inferior to the wild type, while red indicates mutations superior to the wild type. The EC50 values before and after alkali treatment are normalized, with the wild type set to 1. (B) Experimental elvalutation of multi-point mutations. Different colors represent the proportions of improvement or decline in T_m, affinity, and alkali resistance of multi-point mutations compared to the corresponding best single-point mutations they include. (C) Distribution of the occurrences of single-point mutations included in the 20 multi-point mutation variants. The length of the bar represents the frequency of occurrence of each mutation.

As shown in Figure 2A, single-point mutations produced complex results during the combination process due to epistatic effects, which were unpredictable using traditional methods. A57D;P29T exhibited remarkably high alkali resistance, despite A57D and P29T being negative mutants individually. This "double negative yields positive" phenomenon also occurred in terms of affinity, where the EC50 value of P29T;G134R was approximately one-third of that of the wild type, despite both P29T and G134R caused a decrease in affinity individually, with the EC50 value of

P29T being larger than that of the wild type by a factor of five. These mutation pairs are distantly located on the VHH

antibody structure, making it challenging to infer the reasons for the enhanced properties after combination (Figure

3B). Not all combinations resulted in improved mutant effects. For instance, while A15P showed high affinity and

alkali resistance, adding R20T or P29T led to decrease in affinity or alkali resistance, respectively. Additionally,

combining A15P with P117Q, two single mutations capable of individually enhancing affinity, resulted in a decrease

in affinity.

In traditional methods, only positive single-point mutations are selected to compose the multi-point mutants and

negative single-point mutations are typically avoided in multi-point combinations. However, the number of negative

mutations often far exceeds that of positive mutations, thus limiting the sequence space that can be explored by

traditional methods. Large language models can leverage negative mutations to generate positive multi-point

mutants, surpassing the capabilities of rational design and significantly expanding the design space in protein

engineering. Therefore, the Pro-PRIME method manifests significant advantages in exploring sequence space, being

less susceptible to local optima, and having greater potential to find the global optimum. As depicted in Figure 2C,

the 20 multi-point mutations we identified consisted of combinations of nine different single-point mutations.

Although the distribution was biased, we incorporated some negative mutations that experts might consider

disadvantageous, implying that the Pro-PRIME method balances the exploitation of local optima and the exploration

of global optimum.

2.4. Reevaluating the AI predictions from a rational design perspective

In addition to interpret the multi-point mutations based on the single-site mutations, we investigated the design of

VHH antibodies from a rational design perspective to understand the AI predictions. The experimental results from

SDS-PAGE revealed a significant degradation of the VHH antibody following alkali treatment, with small bands

accounting for up to 61.1%. This phenomenon could be attributed to the poor alkali resistance of the VHH antibody.

Mass spectrometry analysis identified specific breakage sites at Q2, Q4, G9, G10, G16, S22, S23, S26, A57, N78, N85,

F101, and G134, highlighting these as ideal targets for rational design. Introducing mutations at these sites to prevent

the breakdown of VHH antibodies could effectively enhance their alkali resistance. The predictions of the Pro-PRIME

model align with this rationale, with 4 out of the top 10 performing single-point mutations occurring at these

breakpoint sites. Experimental evidence showed that our single-point mutations could effectively reduce the

proportion of small bands, and not all positive mutations locate at the breakage sites, such as P29T and A15P (Figure

3A). Mutations identified by the model at other locations may have non-local effects, contributing to overall protein

stability. In the second round of experiments, multi-point mutants exhibited an overall lower proportion of small

bands, reaching as low as 13.6% (A57D;G134R;D114Y), though we did not use the experimental results of SDS-PAGE

to train the model and make predictions. Thus, the improvement in the second round suggests a positive correlation

between the alkali resistance of the VHH antibody and the degree of breakage after alkali treatment. Employing a

rational design approach, it is plausible to identify single-point mutants with improved alkali resistance at these

breakpoints. However, determining the optimal mutated residues requires conducting multiple single-point

saturation mutation experiments, which is time consuming and costly. Deep learning methods can directly predict

mutation types at each site, enhancing design efficiency while remaining aligned with rational design principles.

Furthermore, we analyzed the VHH antibody and homologous sequences from the perspective of evolutionary

sequence alignment (Figure 3C). The results revealed that some critical single-point mutations (P29T, L49V) changed

original residues to more conservative ones, aligning with the "consensus sequence" method commonly used in

rational design[29]. The Pro-PRIME model's ability to predict such mutations is expected, given its training set

contains information from homologous proteins. However, the advantage of deep learning methods extends beyond

providing suggestions aligned with rational design principles; they can predict mutations contrary to expert

experience. For instance, both A15 and R20 are conservative residues according to homologous sequence alignment

(see Figure 3C). Moreover, mutating A15 to proline reduces hydrogen bonds, and changing R20 to threonine

contradicts empirical physicochemical properties, since arginine is more likely to form salt bridges and hydrogen

bonds, benefiting stability. However, both A15P and R20T contribute to increased alkali resistance and play positive

roles in subsequent multi-point combinations. This exceptional capability beyond the traditional rational-design

principles expands the exploration space of deep learning models, aiding to approach the global optimum.

Figure 3. (A) The SDS-PAGE experimental results depict the proportion of small bands observed after alkali treatment for multi-point mutations and certain single-point mutations exhibiting relatively higher alkali resistance. (B) Structure of the VHH antibody. (C) The multiple sequence alignment of the VHH and several homologous sequences.

In order to gain deeper insights into the mechanisms by which the identified mutations enhance protein

properties, we performed molecular dynamics (MD) simulations on the best alkali-resistant mutant. The simulation

results revealed several key observations that help explain the observed improvements in protein stability and alkali

resistance. As shown in Figure S4A, the two-point mutant of A57D;P29T has a Tm increase of around 8 ℃ and a

much stronger binding affinity than the WT. Our analysis of the MD trajectories indicates that the A57D;P29T mutant

has a more rigid structure than that of WT due to its lower root mean squared deviation (RMSD) of protein (Figure

S4B). Furthermore, we calculated the root mean squared fluctuation (RMSF) for each residue, and realized that the

mutant displayed less fluctuation at residue 29 but similar flexibility at residue 57. Interestingly, residues at positions

10, 108 and 118 which spatially distant from residues 29 and 57 in the mutant exhibited remarkable weakened

fluctuations than those in the WT (Figure S1C), implying a more rigid structure of the mutant contributing to its

improved resistance on high temperature and strong alkalinity. However, Figure S4D shows the AlphaFold3 predicted

structures of the WT and the mutant are quite similar. To unveil the origin of change on structural flexibility, we

computed the intramolecular interactions, such as salt bridges and hydrogen bonds for both WT and the mutant. We

observed that the mutations increased the number of hydrogen bonds between the mutation sites and the rest of

the protein (Figure S4E). However, the overall structure of the mutant did not show significant changes, which is also

evident from the solvent-accessible surface area (SASA) analysis (Figure S4F). We also analyzed changes in salt

bridges and found that although residue 57 mutated to Histidine, no new salt bridges were formed. Additionally,

RMSF results showed that residues 10, 108, and 118 became more rigid, but further analysis revealed that there

were no significant changes in hydrogen bonds or other interactions in these regions. Taken together, these findings

suggest that the enhanced alkali resistance of the mutant is likely due to an overall increase in protein stability, rather

than a dramatic change in its structural conformation. The MD simulation results, which are detailed in Figure S4,

provide a deeper understanding of how specific mutations can improve protein properties and offer valuable insights

for future protein engineering applications.

2.5. Improved acid and salt resistance simultaneously

According to the rational design principles, a protein with enhanced intramolecular interactions can resist various

stress, including high temperature, strong alkali, strong acid, concentrated salt, etc.[30, 31] Hence, we expected that

the mutants with higher thermal stability and alkali resistance can tolerate acidic and saline environments as well.

We evaluated the binding affinity of the mutants to the target protein under elution with saline (1 M NaCl) or acidic

(20 mM citric acid) solutions to characterize their salt-induced and acid-induced dissociation abilities. Additionally,

we assessed the acid resistance of the mutants by measuring their affinity after a 48-hour treatment with 1 M

ethanoic acid (see details in SI). Figure 4 shows the acid and salt resistance of the selected multi-point mutants with

higher thermal stability and stronger alkali resistance. It is noteworthy that the Pro-PRIME model can provide us the

mutants, such as A57D;P29T and A15P;R20T, with strong resistance on acid and salt (Figure 4A) as well as high

temperature and alkali resistance (Figure 2B). Our design enhances protein stability while retaining other properties

such as affinity, thereby endowing the mutant with the potential for application in industrial production.

2.6. Application in industrial production

The Pro-PRIME method can predict multi-site mutants using a small amount of experimental data, and the entire

design process can be completed within two months. This method has the potential to empower industrial

production due to its relatively low economic and time costs. Our mutants have already been widely applied in the

purification process of growth hormones. Figure 4B shows the ratio of the residual dynamic binding capacity (DBC)

after 0.5 M NaOH treatment for 6 hours and 24 hours to the residual DBC before treatment for various mutants and

the wild type. The wild type experienced a 74.1% loss in the residual DBC after 6 hours of alkali treatment, and only

retained 15.2% of its residual DBC after 24 hours of alkali treatment. In contrast, the designed multi-site mutants

retained 60-90% of their residual DBC after 6 hours of alkali treatment, and even after 24 hours of treatment, some

mutants maintained over 50% of their residual DBC, which is remakably more stable than the wild type in the alkaline

environment of industrial production. Figure 4C shows the variation in yield during affinity chromatography across

multiple cycles after employing our mutant. After more than 140 cycles, the yield of our mutant did not exhibit a

significant downward trend. In contrary, the binding affinity of wild type is unable to sustain after 60 cycles.

Therefore, our designed mutants maintain activity after more cycles of reuse compared to the wild type,

substaintially reducing the production cost of growth hormones. Consequently, the selected mutant designed by the

LLM has been applied in mass production scale up to 5,000 liters.

Figure 4. (A) The tolerance of acid dissociation and salt dissociation, and the acid resistance of multi-point mutations with strong
alkali resistance. All EC50 values are normalized, with the wild type set to 1. (B) Experiment results of residual dynamic binding
capacity (DBC) at 10% breakthrough. The bars represent the ratio of the residual DBC after 0.5 M NaOH treatment for 6 hours
and 24 hours to the residual DBC before treatment for multi-site mutants and the wild type. (C) Yield in affinity chromatography
and the corresponding number of cycles. This figure illustrates the variation in yield during affinity chromatography across
multiple cycles.

3. Conclusions

Through two rounds of evolution, we successfully designed a VHH antibody with strong resistance to extreme

environments and enhanced affinity using the Pro-PRIME model. Although rare case can tolerate the extreme pH and

saline conditions in our pre-training dataset, the Pro-PRIME model showed impressive performance after supervised

learning with limited data, especially on capturing the epistatic effects. The analysis of these 65 mutants revealed

that the Pro-PRIME model is adept at exploring the large space of protein fitness, being less susceptible to local

optima, and having greater potential to find the global optimum. Our efficient method of designing mutants that

consider multiple properties improvement holds promise for industrial application of proteins. Specifically, the VHH

antibody has been deployed in practical production and significantly enhancing the efficiency of the entire

production line after our design. While the Pro-PRIME model itself has been reported[25], this work demonstrates its

first-time application to the challenge of designing proteins with alkali resistance and other extreme properties that

are not found in natural proteins, nor have previous studies addressed or provided data for such applications. This

shift from optimizing existing protein properties to engineering entirely new, unnatural traits is a significant advance

in the field. This study shows that the AI models, such as Pro-PRIME, can not only guide the evolution of protein

thermal stability, enzymatic activity, ligand affinity but also enable to develop the mutants adapting the harsh

unnatural environments, such as extreme pH and concentrated salt, largely expanding its application. The novelty of

this work lies in the ability to design and engineer proteins with novel properties, specifically alkali resistance, which

is an unprecedented achievement in AI-assisted protein engineering. The great potential of AI model is expected to

significantly accelerate the development of proteins for diverse applications in medicine, agriculture, bioengineering,

etc.

CRediT authorship contribution statement

Liqi Kang: Writing – original draft, Data curation, Conceptualization, Methodology. **Banghao Wu:** Data curation,

Methodology. **Bingxin Zhou:** Writing – review & editing. **Pan Tan:** Writing – review & editing. **Yun (Kenneth) Kang:**

Data curation. **Yongzhen Yan:** Data curation. **Yi Zong:** Data curation. **Shuang Li:** Data curation. **Zhuo Liu:** Writing –

review & editing, Supervision. **Liang Hong:** Writing – review & editing, Supervision.

Conflicts of interest

Patents have been filed in 2023 for all these mutations in the present work. GeneScience Pharmaceuticals, a company that discovered the native sequence of VHH, own the patent of VHH.

Data availability

Data will be made available on request.

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