3   Developing heterospecific Sterile Insect Technique for pest control: insights

4   from the spotted wing fly *Drosophila suzukii*

Flavia Cerasti [1], Massimo Cristofaro [2] Valentina Mastrantonio [1], Jessica Scifo[3], Adriano Verna[3],

Daniele Canestrelli[4], Daniele Porretta[1*]

[1]Department of Environmental Biology, Sapienza University of Rome, Rome, Italy

[2]Biotechnology and Biological Control Agency (BBCA), Rome, Italy

[3]FSN-FISS-SNI Laboratory, Italian National Agency for New Technologies, Energy and

Sustainable Economic Development (ENEA), Rome, Italy

[4]Department of Ecology and Biology, Tuscia University, Viterbo, Italy

[*]Correspondence: daniele.porretta@uniroma1.it

18   Abstract

BACKGROUND: Reproductive interference (i.e., sexual interaction between males of one

species and females of another species that reduce the fitness of one or both the interacting

individuals) is an important species interaction significantly affecting population dynamics and

persistence. However, its exploitation in pest control remains overlooked. Here, we investigated

the possible integration of reproductive interference into the Sterile Insect Technique (SIT) to

develop a heterospecific SIT (h-SIT). Under this approach, contrary to the classic SIT, sterile

heterospecific males from closely related, non-pest species are released to compete with the pest

population for mates. At this aim, we focused on the invasive pest species *Drosophila suzukii*

and used *D. melanogaster* as the control species. First, we investigated the effect of irradiation

on *D. melanogaster* sterility and longevity. Then, we tested the mating performance of irradiated

males and their ability to reduce the *D. suzukii* fitness.

RESULTS: We found by microcosm experiments that: i) irradiation induced high levels of *D. melanogaster* male sterility without reducing longevity; ii) irradiated *D. melanogaster* males court *D. suzukii* females as much as *D. suzukii* males and they couple, mate and fecund heterospecific females; iii) irradiated *D. melanogaster* males significantly reduce the offspring of *D. suzukii* females under two different species ratios.

CONCLUSIONS: Our results provide the first foundation to develop a heterospecific Sterile Insect Technique against *D. suzukii* and fuel to test this approach against other groups of pest species.

Keywords: Reproductive interference, pest control, Sterile Insect Technique, *Drosophila suzukii*, *Drosophila melanogaster*

1. Introduction

Species interactions, such as predation, parasitism, or competition, can significantly affect the population dynamics of the interacting species, affecting their abundance and persistence [1]. These interactions have been used for a long time to control pest species in agriculture and are the foundation of modern biological control approaches [2].

Along with the above species interactions, reproductive interference is now recognized as a major ecological process affecting population dynamics and species persistence [3-7]. Reproductive interference (or satyrisation in animals) consists of any sexual interaction between species that reduce the fitness of one or both interacting individuals [8]. It occurs due to incomplete mating barriers between species and can occur at any stage of mate acquisition, from courtship to mating and hybridization [3]. Reproductive interference has been documented under laboratory and field conditions in a wide variety of sexually reproducing taxa [5,6]. Theoretical and empirical studies showed that reproductive interference, as competition, is density-dependent and can result in population or species exclusion [4,9].

The first attempts to exploit reproductive interference for pest control date back to the first mid of the XX century. During the 1930s – 40s, in pioneering works, F. L. Vanderplank and colleagues applied reproductive interference to suppress tsetse fly, *Glossina swynnertoni* Austen

(reviewed in [10]). Laboratory experiments showed that offspring produced by cross-mating

between *Glossina swynnertoni* (the vector species) and *G. morsitans* Westwood (a nonvector

species) had low fertility, as all hybrid males and some hybrid females were sterile. Then, a large

field experiment was carried out in the Shinyanga area, Tanzania, by releasing fertile *G.*

morsitans pupae over seven months. After the mass release, the density of *G. swynnertoni* was

drastically reduced demonstrating the potential of hybrid sterility and sexual interference to

suppress tsetse flies [9,10].

Although the experience with tsetse flies showed the potential effectiveness of

reproductive interference for controlling a pest species, it did not lead to common exploitation of

this approach. A major concern about heterospecific sterility was that pre-mating isolation

mechanisms could constrain mating between released heterospecific males and wild females,

leading to failure to control the target species in the field [11,12]. A further concern has been the

risk of introducing non-native pest species. The mass release of fertile individuals of a closely

related species that is non-native or a pest itself, at least potentially, would lead to replacing one

pest species with another. These concerns and the success in the 1950s of Knipling and his team

in eradicating the screwworm *Cochliomyia hominivorax* (Coquerel) using ionizing radiation

[13], has led to the spread of homospecific Sterile Insect Technique (SIT) as the major approach

exploiting sterility in pest control during the last decades [14-18]. It consists of mass rearing,

sterilization by ionizing radiation, and massive release of conspecific individuals into the target

population. The unfertile mating between the released males and wild females leads to a

progressive decline of the target pest population [19,20].

However, a renewed interest has recently been in exploiting reproductive interference to

control pest species [8,21-24]. Mitchell et al. [8] reviewed the literature on reproductive

interference in natural populations. They highlighted the effects of these processes on population

decline in nature and proposed a framework for their use in pest control [8].

Interestingly, McInnis [21] tested the potential use of sterilized males of the oriental fruit

fly *Bactrocera dorsalis* (Hendel) against wild carambola fruit fly *B. carambolae*, suggesting that

the concern about using non-native or potential pest species to control another pest can be

overcome if the released heterospecific individuals are sterile, thus integrating reproductive

interference and SIT. More recently, Honma et al. [22] have proposed a framework for

incorporating reproductive interference into a classic SIT program. They argued that the sterile

males released in a SIT program to suppress the wild-type population of the same species could
also lead to suppression of a closely related pest species through reproductive interference (an
approach that they called sterile interference).

In this paper, we aimed to explore the potential use of reproductive interference to
develop a heterospecific SIT (h-SIT) approach against the spotted wing fly *Drosophila suzukii*
Matsumura. This approach, contrary to the classic SIT, is based on using sterile heterospecific
males from closely related species to compete with the pest population for mates. *D. suzukii* is an
invasive pest that has spread in the last few decades from its native range in East Asia throughout
North America, Europe, and South America [25,26]. Unlike most Drosophilidae, *D. suzukii* can
lay eggs in unripe and healthy fruits, causing severe economic losses for fruit industries
worldwide [27,28]. We selected the fruit fly *D. melanogaster* as the species inducing
reproductive interference. Previous studies showed that post-mating isolation between *D.*
melanogaster and *D. suzukii* is complete, while there is an incomplete pre-mating isolation
[29,24]. Wolf and colleagues, have recently assessed the potential for hybridization between gene
drive-modified *D. suzukii* individuals and non-target *Drosophila* species in Europe. They found
by male mating behavior tests that *D. melanogaster* males frequently showed interest in *D.*
suzukii females but did not achieve copulation [29]. Accordingly, in our previous study, by
investigating reproductive interference between non-irradiated *D. melanogaster* males and *D.*
suzukii, we found that *D. melanogaster* males successfully courted *D. suzukii* females.
Furthermore, they can inseminate *D. suzukii*, leading to egg deposition; however, no hybrids are
produced as these eggs did not progress to larval development. Finally, the presence of non-
irradiated *D. melanogaster* males under different species ratio also imposed fitness costs on *D.*
suzukii females, resulting in reduced *D. suzukii* offspring production [24]. These results are a
baseline for exploiting irradiated *D. melanogaster* males in a h-SIT approach.

Here, we specifically aimed: *i*) to assess the effect of irradiation doses on the sterility degree
and longevity of *D. melanogaster* males. To this end, we irradiated virgin *D. melanogaster* males
with two gamma-ray doses (60 and 80 Gy) and assessed their fertility, through mating trials with
D. melanogaster females, and their longevity; *ii*) to analyze the mating performance of irradiated
D. melanogaster males in courting and mating with *D. suzukii* females; *iii*) to evaluate if
irradiated *D. melanogaster* males can reduce *D. suzukii* fitness with whom they mated. To this

end, we analyzed the effect of irradiated *D. melanogaster* males on the fertility of *D. suzukii*

females using different species ratios.

2. Material and methods

2.1 Laboratory colonies

Drosophila suzukii and *D. melanogaster* individuals from the laboratory rearing facilities of the

Sapienza University of Rome were used in this study [24]. Both species were reared on an

artificial diet consisting of agar (7 g), table sugar (16 g), precooked ground maize (72 g), mother

yeast (18 g), soy flour (10 g), and methylparaben (2.5 g) [30]. The colonies were maintained in

entomological cages (30 x 30 x 30 cm), in a climate chamber at 25 ±1°C, 14:10 h light:dark

cycle.

2.2 Effect of irradiation dose on sterility and longevity of *D. melanogaster* males

To evaluate the effect of irradiation doses on the sterility degree of *D. melanogaster* males,

mating trials between irradiated males and fertile *D. melanogaster* females were carried out.

Virgin *D. melanogaster* adults were obtained by checking the adult emergence from the pupae

confined in rearing falcons (50 ml) every thirty minutes, where they completed their full larval

development. As soon as new individuals emerged, we divided them into cages (15 x 15 x 15

cm) according to the gender. This procedure allows us to be sure to use only virgin males and

females. After selection, *D. melanogaster* males were sterilized using gamma radiation. Males

(48-72-96 hours old) were placed in 50 ml falcon tubes containing wet cotton to avoid

dehydration and transported to the Calliope Facility at ENEA Casaccia Research Centre (Rome).

The Calliope plant is a pool-type irradiation facility equipped with a ^{60}Co (mean energy 1.25

MeV) radio-isotopic source array in a high volume (7.0 m ´ 6.0 m ´ 3.9 m) shielded cell. The

irradiation cell can provide different dose rates by placing the samples in specific positions and

exposing them for varying periods [31]. We provided irradiation doses of 60 Gy and 80 Gy to *D.*

melanogaster males, according to Nelson et al. [32] and Henneberry et al. [33]. The dose rate

was 175.03 Gy/h (2.92 Gy/min). After irradiation, five irradiated males were placed in falcon (50

ml) with five non-irradiated *D. melanogaster* females. Each falcon contained food substrate to

allow females to lay eggs. We assessed two control treatments: a control treatment called

"Home", where we used non-irradiated *D. melanogaster* males, which have not undergone

transport stress but have always been kept in the climate chamber conditions; a control treatment
called "Trip", where we used non-irradiated *D. melanogaster* males and females previously
transported to the Calliope facility without receiving any irradiation dose. In this way, we
evaluated the impact that the stress due to transport could have on their fertility. We performed
twelve to sixteen replicates for each irradiation dose (60 and 80 Gy) and the control treatment (0
Gy). The couples were left together for six days and then removed. New individuals that
emerged were removed and counted, and oviposition substrates were checked daily until no
newborn individuals were observed. We further evaluated the impact of the highest irradiation
dose (80 Gy) on the sterility of *Drosophila melanogaster* females, by carrying out mating trials
between irradiated females with fertile *D. melanogaster* males as described above. This
information can be useful in developing a h-SIT approach since, in mass-rearing conditions,
sexing may not be completely accurate, and females could inadvertently be released into the
field.

To evaluate the effect of irradiation doses on male longevity, we compared the average
lifespan between irradiated and non-irradiated *D. melanogaster* males. Virgin *D. melanogaster*
males were selected and irradiated as described above. A cage (30 x 30 x 30 cm) with twenty
males was set up for each dose treatment (60 and 80 Gy). Two more cages were set up as control,
one as "Control home" and one as "Control trip", as in the previous experiment. Mortality was
recorded every morning until all flies had died.

2.3 Mating performance of irradiated *D. melanogaster* males
All the following assays were carried out using only the *D. melanogaster* males irradiated with
an 80 Gy irradiation dose, which led to higher sterility without affecting male longevity (see
Results section).

First, the mating performance of *D. melanogaster* males was investigated in no-choice
and choice tests. In no-choice tests, we set up four experimental conditions by placing in 15 ml
falcons: 1) one no-irradiated *D. melanogaster* male and one *D. melanogaster* female; 2) one
irradiated *D. melanogaster* male at 80 Gy and one *D. melanogaster* female; 3) one *D. suzukii*
male and one *D. suzukii* female; 4) one irradiated *D. melanogaster* male at 80 Gy and one *D.*
suzukii female. In this way, it has been possible to understand the average courtship time of *D.*
melanogaster males with conspecific and heterospecific females and investigate if the irradiated

D. melanogaster males court *D. suzukii* females as much as *D. suzukii* males. In the choice test, we set up an experimental condition by placing in 15 ml falcons one *D. suzukii* female, one *D. suzukii* male, and one irradiated *D. melanogaster* male and analyzing the courtship behavior of *D. melanogaster* males and *D. suzukii* males separately when they co-occurred.

The experiments were carried out with virgin individuals selected as described above. After 5 minutes of acclimatation, we recorded 10 minutes of the individual's behavior with an Olympus Tough TG-6 camera. The videos were used to analyze the courtship behavior elements of *D. melanogaster* and *D. suzukii* males [34,35] using the BORIS software [36]. Twenty replicates were carried out.

Second, we investigated if irradiated *D. melanogaster* males were able to mate with *D. suzukii* females and fecund them. We set up two experimental conditions: - a control condition where we placed in 50 ml falcons with food substrates one virgin *D. suzukii* female alone, to investigate if females lay unfertilized eggs; - an experimental condition where we placed in 50 ml falcons with food substrates, one virgin *D. suzukii* female and one irradiated *D. melanogaster* male at 80 Gy as described previously. We left the individuals inside the falcons for 6 days, to allow the females to oviposit. After that time, we removed the individuals and, through a stereomicroscope Leica EZ4W at magnification 5x we checked daily the possible occurrence of eggs laid by *D. suzukii* females in food substrates. If present, eggs were photographed with a stereomicroscope digital camera and monitored for eclosion and for eventual larval development. Forty replicates were carried out in both conditions.

2.4 Effect of irradiated *D. melanogaster* males on *D. suzukii* fitness

We evaluated whether the reproductive interactions between *D. melanogaster* males and *D. suzukii* individuals result in fitness costs for *D. suzukii* females. At this aim, we set up three experimental setting by placing in entomological cages (15 x 15 x 15 cm): 1) five pairs of no-irradiated virgin *D. suzukii* (selected as described above) without *D. melanogaster* males; 2) 40 irradiated *D. melanogaster* males and five pairs of virgin *D. suzukii* to test an 8:1 over-flooding ratio (sterile:wild males, OFR); 3) 60 irradiated *D. melanogaster* males with five pairs of virgin *D. suzukii* to test an OFR of 12:1 [20]. Then, we compared the number of newborn individuals that emerged from cages with only *D. suzukii* pairs and from cages where *D. suzukii* pairs plus irradiated *D. melanogaster* males co-occurred.

2.5 Data analysis

To evaluate the effect of the irradiation doses on the sterility degree of *D. melanogaster* adults, a GLM model (generalized linear model; package MASS) [37] was applied. We used the irradiation dose of *D. melanogaster* adults as a fixed effect (0-60-80 Gy) on the offspring produced by *D. melanogaster* females (the response variable), which was recorded as an event with a continuous distribution. We analyzed the offspring produced with a negative binomial distribution applied to the GLM model. The model family was selected comparing the AIC and BIC estimators and the likelihood ratio test. Multiple Comparisons of Means Tukey Contrasts was performed as a post-hoc test using the *multcomp* package in R [38].

To evaluate the effect of the irradiation doses on the longevity of *D. melanogaster* males, survival distributions of the four *D. melanogaster* groups ('Control cages', 'Control trip', '60 Gy', '80 Gy') were computed using the Kaplan–Meier method [39]. At this end, the *survival* and *survminer* R packages were used [40]. The differences between survival distributions were estimated using the Log-Rank Test.

To analyze the courtship behavior data, we used a GLM model with a negative binomial distribution, according to model selection estimators, for comparing the courtship time among no-choice conditions. As a post-hoc test, we performed the Tukey test. To compare the time spent by *D. melanogaster* and *D. suzukii* males in courting *D. suzukii* females in the choice test, we used the Wilcoxon Mann–Whitney U test (*dplyr* package).

Regarding the effect of *D. melanogaster on D. suzukii* fitness, a GLMM analysis was performed using the *D. suzukii* offspring as a response variable with a continuous distribution. In the analysis, the number of *D. melanogaster* males (0, 40, or 60 individuals) in each replicate was considered a fixed effect (the explanatory variables). The age of the experimental individuals (48, 72, and 96 hours old) was considered a random effect since they are a sampling of infinite possible combinations, and we were not interested in studying them as such but in identifying whether they constituted a source of significant variability. We applied a negative binomial distribution to the GLMM effect, comparing the models based on the optimal parsimony principle (AIC and BIC estimators) and the likelihood ratio test. We performed the Multiple Comparisons of Means Tukey Contrasts as a post-hoc test.

246   All analyses were performed using the software R version 3.6.2. [41].

3. Results

3.1 Effect of irradiation dose on sterility and longevity of *D. melanogaster*

In the mating trials aimed to assess the effect of irradiation on male sterility, the mean number of offspring produced by *D. melanogaster* females was 128.66 ($\pm$ 10.67) (mean $\pm$ SE) in the control "Home" and 149.9 ($\pm$ 18.07) in the control "trip" condition. In the treatments at 60 and 80 Gy, the number of offspring produced by *D. melanogaster* females was 35.81 ($\pm$ 4.08) and 29.84 ($\pm$ 3.62), respectively (Fig. 1). The GLM model showed a significant effect of the irradiation dose on the number of offspring produced by *D. melanogaster* females when the males were irradiated at 60 Gy and 80 Gy (Table 1). The Tukey Multiple Comparison tests showed a significant reduction in *D. melanogaster* offspring produced when *D. melanogaster* males were irradiated at 60 Gy ($z = -6.268$, $p = < 0.001$) and 80 Gy ($z = -9.335$, $p = < 0.001$) than in the control "home" condition. There was a significant reduction also with irradiated males at 60 Gy ($z = -6.784$, $p = < 0.001$) and 80 Gy ($z = -9.702$, $p = < 0.001$) than in the Control "trip" condition. There were no significant differences between the two control conditions ($p > 0.05$) (Fig. 1).

In the mating trials aimed to assess the effect of irradiation on female sterility, the mean number of offspring produced by *D. melanogaster* females was 72.6 ($\pm$ 10.34) (mean $\pm$ SE) in the control "Home" and 81.4 ($\pm$ 13.86) in the control "trip" condition. In the treatments at 80 Gy, the number of offspring produced by *D. melanogaster* females was 10.08 ($\pm$ 1.39) (Fig. 2). The GLM model showed a significant effect of the irradiation dose on the number of offspring produced by irradiated *D. melanogaster* females at 80 Gy (Table 1). The Tukey Multiple Comparison tests showed a significant reduction in *D. melanogaster* offspring produced when *D. melanogaster* females were irradiated at 80 Gy ($z = -9.358$, $p = <1e-04$) than in the control "home" condition. There was a significant reduction also with irradiated females at 80 Gy ($z = -9.917$, $p = <1e-04$) than in the Control "trip" condition. There were no significant differences between the two control conditions ($p > 0.05$) (Fig. 2).

The Kaplan–Meier survival curves showed significant differences in the lifespan of *D. melanogaster* males among treatments (60 Gy, 80 Gy, the Trip cage control and Home cage control groups) (Mantel-Cox log-rank; $\chi2 = 17.8$, d.f.= 3, $P = 5e-04$) (Fig. 3). The pairwise comparisons test showed that the individuals irradiated with 60 and 80 Gy had higher survival

probability than control individuals. Significant differences were indeed observed between the

males irradiated at 60 Gy and those of the "Control home" (p= 0.0308) and the "Control trip"

(p=0.0151) groups, as well as between the males irradiated at 80 Gy and those of the two control

groups (control home p=0.0053; control trip p= 0.0028). No significant differences were

observed between the two control groups (p=0.7859) and between the two experimental

conditions (p=0.8541).

3.2 Mating performance of the irradiated *D. melanogaster* males

The mating performance of irradiated *D. melanogaster* males toward *D. suzukii* females was

investigated by courtship and mating trials.

First, we investigated if irradiated *D. melanogaster* males were able to court *D. suzukii* females

as much as *D. suzukii* males in no-choice and choice tests. Under all experimental conditions the

typical behavior elements during courtship were observed, including "orientation" (i.e., the male

approaches the female, quivering the abdominal and scissoring its wings), "tapping," (i.e., the

male hits the female abdomen, or middle and hind legs by stretching his foreleg); "wing

spreading", "wing scissoring" (i.e., the male is oriented toward the female front, quivers with the

abdomen and scissors his wings keeping them at 180° for seconds to expose the upper side and

wing spot toward to female) [34,35].

In the no-choice tests, the average courtship time spent by no-irradiated *D. melanogaster* males

courting no-irradiated *D. melanogaster* females was 23.37 % (± 5.19)(mean ± standard error);

the average courtship time spent by irradiated *D. melanogaster* males at 80 Gy courting no-

irradiated *D. melanogaster* females was 20.84 % (± 3.61); the average courtship time spent by

irradiated *D. melanogaster* males courting *D. suzukii* females was 26.41% (± 5.90); the average

courtship time spent by *D. suzukii* males courting *D. suzukii* females was 35.36% (± 8.17). The

GLM model showed no significant differences in the average courtship time among conditions.

The Tukey Multiple Comparison tests showed no significant differences among the four

conditions (p> 0.05) (Fig. 4).

In choice tests, where *D. melanogaster* males and *D. suzukii* males were placed together in 15 ml

falcon with *D. suzukii* females, the average courtship time spent by *D. melanogaster* males was

19.43% (± 6.16) and the average courtship time spent by *D. suzukii* males was 9.52% (± 2.29).

There are no significant differences between the courtship time of *D. suzukii* and *D. melanogaster* males (Wilcoxon Mann–Whitney test W = 194.5, p-value = 0.9105) (Fig. 5).

Second, we investigated if irradiated *D. melanogaster* males could mate and fecund *D. suzukii* females. At this end, we analyzed the occurrence of oviposited eggs by vergin *D. suzukii* females confined alone and with irradiated *D. melanogaster* males. In monitoring the food substrates of each replicate daily, we found no eggs in 39 out of 40 substrates in the condition with only virgin *D. suzukii* females. In the condition with virgin *D. suzukii* females paired with irradiated *D. melanogaster* males, eggs were found in 12 out of 40 replicates (30%), but no subsequent larval development was observed.

3.3 Effect of irradiated *D. melanogaster* males on *D. suzukii* fitness

The mean number of the individuals originating from five pairs of *D. suzukii* was 37.95 (± 3.62) (± standard error) in the control tests, while it was 16.11 (± 5.58) and 13.89 (±3.69), when 40 and 60 irradiated *D. melanogaster* males were added, respectively (Fig. 6). The GLMM model showed a significant effect on the *D. suzukii* offspring (Table 2). The Tukey Multiple Comparison tests showed a significant reduction of *D. suzukii* offspring when 40 (z = -2.758, p = 0.01601) and 60 (z = -3.218, p = 0.00376) irradiated *D. melanogaster* males were placed with *D. suzukii* couples (Fig. 6), while no significant differences were observed between treatments with 40 and 60 *D. melanogaster* irradiated males (z = -0.408, p = 0.91176).

4. Discussion

4.1 Irradiation dose, *D. melanogaster* male sterility and longevity

Heterospecific SIT and SIT approaches can be developed under similar theoretical frameworks. Irradiation dose is an important factor affecting the sterility degree, male longevity, and performance [42]. It is, therefore, necessary to find a high enough dose to induce sterility but that has a minimum impact on the biological quality of the irradiated males [43].

We found high levels of sterility in both *D. melanogaster* males and females (Fig. 1 and 2). As observed also in other insect species [44-46], we found that females showed higher sensibility to irradiation than males. While affecting fertility, irradiation did not reduce male longevity. Interestingly, irradiated males exhibited a longer lifespan compared to control individuals. This finding contrasts with some results reported in the literature on Diptera. For

example, Lanouette et al. [47] observed no significant differences in the longevity of *D. suzukii* males exposed to increasing irradiation doses (30–120 Gy). In contrast, Chen et al. [48] reported that longevity in *D. suzukii* began to decrease at doses exceeding 90 Gy. Similarly, in *Aedes aegypti*, survival of irradiated males was significantly reduced compared to controls [41,49]. Despite the disparity between our findings and those of other studies, the phenomenon of increased longevity following irradiation has been documented in other species, including some *Anopheles* species [50,51] and the stink bug pest *Bagrada hilaris* [52], where irradiated males at 80 Gy exhibited a longer average lifespan than control individuals. The reasons for such results could be several, since molecular and cellular changes to biological, physical, and human factors [53,54]. Understanding the mechanisms underlying this result would certainly warrant further in-depth analysis. However, within the context of this study, sterile males with stable or extended longevity are likely to remain in the environment longer, effectively competing with fertile males and contributing to population suppression over a prolonged period.

As observed in previous studies [32,33], we also obtained residual fertility in our results. It will be interesting to test if higher irradiation doses will lead to higher male sterility without reducing male performance. Notably, the purpose of male sterilization is different in SIT and h-SIT approaches. In the former, male sterilization is the way to introduce sterility into the wild pest population. Residual fertility could hamper the SIT effectiveness as wild females could mate with released unsterilized males and produce fertile offspring [20,55,56]. Conversely, in the h-SIT, the introduction of sterility in the target pest population is ensured by the reproductive post-zygotic isolation mechanisms between the released heterospecific males and the wild females of the target species [10,21]. Male sterilization avoids potential adverse environmental effects if non-native or potentially pest species are used in the h-SIT. In the case of h-SIT using *D. melanogaster*, residual fertility would not reduce the released males' effectiveness because not irradiated males would induce sterility in *D. suzukii* females [24]. Furthermore, there are no concerns about the potential release of fertile *D. melanogaster* males. Indeed, *D. melanogaster* is not considered an agricultural pest as the female oviposits on rotten fruits [57].

4.2 Mating performance of the irradiated *D. melanogaster* males and effect on *D. suzukii* fitness

Heterospecific SIT is based on the sterility between heterospecifics. The unfertile mating

between the released heterospecific males and wild females leads to a progressive decline of the

target pest population [19,20]. Therefore, the potential use of h-SIT as a control method strictly

depends on the mating ability of the released irradiated males [3]. Our results supported that the

irradiated *D. melanogaster* males at 80 Gy can be effective at different stages of mate

acquisition, from courtship to mating. The analysis of the courtship behavior showed that *D.*

melanogaster males courted *D. suzukii* females as much as *D. suzukii* males (Fig. 4 and 5)

Notably, irradiated *D. melanogaster* males exhibited comparable average courtship time toward

D. suzukii females and conspecific females (Fig. 4). This behavior may be driven by reproductive

interference, primarily through misdirected courtship by *D. melanogaster* males. Male insects,

including *D. melanogaster*, often exhibit less selective mate choice due to their lower

reproductive investment compared to females [58]. This tendency can lead to heterospecific

courtship behaviors. Additionally, the larger body size of *D. suzukii* females, often associated

with higher fecundity, could make them appear more attractive to *D. melanogaster* males, as

body size is commonly used as a proxy for reproductive potential [59,60].

The results of mating trials showed that in the control condition with virgin *D. suzukii*

females alone, there were no oviposit eggs in 39 food substrates except for one; larval

development and production of *D. suzukii* individuals were observed in that replicate, suggesting

we inadvertently selected a *D. suzukii* female that had likely mated before the experiment.

Notably, we found that in the test conditions (one *D. suzukii* female placed with one irradiated *D.*

melanogaster male), *D. suzukii* females oviposited eggs in 30% of food substrates, and no larval

development was observed in any tests. This result confirms that irradiated *D. melanogaster*

males could couple, mate, and fecund *D. suzukii* females, but the post-zygotic isolation between

D. suzukii and *D. melanogaster* is complete [24].

The results of our study clearly show that irradiated *D. melanogaster* males can lead to

reproductive interference in *D. suzukii*. Indeed, *D. suzukii* had significantly reduced offspring in

the presence of irradiated *D. melanogaster* males, irrespective of the species ratio used (Fig. 6).

The reproductive interference on *D. suzukii* by irradiated *D. melanogaster* males is likely due to

interference during courtship and mating, as described above. However, other factors could

contribute to the offspring reduction of *D. suzukii* by *D. melanogaster* males. First, it has been

shown that *D. melanogaster*, during courtship, produces cis-vaccenyl acetate (cVA), which acts

as a repellent to *D. suzukii* female for laying eggs [61,62]. Furthermore, *D. melanogaster* males

release substances through seminal fluid that reduce female remating in homospecific matings

[63,64]. This latter phenomenon deserves future studies. Indeed, if it also occurs in heterospecific

matings between *D. suzukii* and *D. melanogaster*, it would be of great interest for heterospecific

SIT, preventing *D. suzukii* females from remating with co-specific wild males.

4.3 "The importance of being *melanogaster*"

The above arguments make us optimistic about using *D. melanogaster* in h-SIT against *D.*

suzukii and fuel to test this approach under well-isolated confined-field facilities.

However, let us highlight a further issue that makes h-SIT particularly exciting to

explore: *D. melanogaster* is a model species. The huge amount of data available on the genetics

and biology of *D. melanogaster* can be exploited to address classical problems in developing SIT

[65,66,67]. Despite considerable research, for most species where SIT is used, we lack a deep

understanding of the effects of radiation, rearing conditions (diet, light, density), as well as the

effects of environmental conditions on individual biology and male performance in the field.

Furthermore, some authors emphasized the need to build "a better male" [68] to overcome the

prevailing approach based on overflooding ratios (sterile: wild males) in the field, which can

hamper the economic feasibility of the SIT. In this context, *D. melanogaster*, whose biology has

been dissected at the molecular, cellular, and physiological levels, could significantly contribute

to unveiling these critical issues, making, in perspective, *D. melanogaster-suzukii* a model

system for the development of heterospecific SIT.

5. Conclusions

Reproductive interaction between heterospecific individuals can have multiple effects on pest

management. When pre-mating barriers between interfertile species are weak, hybridization can

occur and significantly threaten pest control. It can favor pest outbreaks by improving population

fitness and adaptation to environmental conditions or by leading to control failures as a

consequence of the introgression of pesticide resistance alleles from one species to another

[69,70]. On the other hand, reproductive interaction can be beneficial for pest management

[8,22]. Here, we highlight the potential use of reproductive interference in pest management by

heterospecific SIT. This approach would broaden our control toolbox by offering additional

options to control important agricultural pests. In the last decades, the new sequencing

technologies combined with classical reproductive incompatibility studies have improved our knowledge of pre- and post-zygotic isolation in sibling pest species, allowing us to identify even more numerous potential systems for the development and application of heterospecific SIT control approach [71,72].

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Data availability

Raw data will be permanently archived if the paper is accepted for publication.

Author contributions

DP and MC conceived the study. DP, MC and DC designed the study. FC, VM, JS and AV collected the data. FC and VM analyzed the data. DP wrote the first draft of the manuscript. All authors contributed critically to the drafts and approved the final manuscript.

Conflict of Interest Statement

The authors declare no conflicts of interest.

References

1. Cain, ML, Bowman WD, Hacker SD. Ecology 2nd edition, ed. by Oxford University Press, Incorporate, Oxford, pp. 648 (2011).

2. Hoddle MS and Van Driesche RG, Biological control of insect pests. Encyclopedia of
insects, ed. by Academic Press, Cambridge MA, pp. 91-101 (2009).

3. Gröning J, Hochkirch A, Reproductive Interference Between Animal Species. *Q Rev Biol*
83:257–82 (2008).

4. Kishi S, Nishida T, Tsubaki Y, Reproductive interference determines persistence and
exclusion in species interactions: Sexual interference governs competition. *J Anim Ecol*
78:1043–9 (2009).

5. Kyogoku D, Reproductive interference: ecological and evolutionary consequences of
interspecific promiscuity. *Popul Ecol* **57**:253–60 (2015).

6. Shuker DM, Burdfield-Steel ER, Reproductive interference in insects: Reproductive
interference in insects. *Ecol Entomol* **42**:65–75 (2017).

7. Gómez-Llano M, Germain RM, Kyogoku D, McPeek MA, Siepielski AM, When
Ecology Fails: How Reproductive Interactions Promote Species Coexistence. *Trends*
Ecol Evol **36**:610–22 (2021).

8. Mitchell C, Leigh S, Alphey L, Haerty W, Chapman T, Reproductive interference and
Satyrisation: mechanisms, outcomes and potential use for insect control. *J Pest Sci*
95:1023–36 (2022).

9. Ribeiro JMC, Can Satyrs Control Pests and Vectors? *J Med Entomol* **25**:431–40 (1988).

10. Klassen W, Curtis CF, Hendrichs J, History of the sterile insect technique. Sterile insect
technique, ed by CRC Press, Boca Raton, pp. 1-44 (2021).

11. Davidson G, The potential use of sterile hybrid males for the eradication of member
species of the *Anopheles gambiae* complex. *Bull World Health Organ* **40**:221–8 (1969).

12. Davidson G, Odetoyinbo JA, Colussa B, Coz J, Field attempt to assess the mating
competitiveness of sterile males produced by crossing 2 member species of the *Anopheles*
gambiae complex. *Bull World Health Organ* **42**:55–67 (1970).

13. Knipling EF, Screwworm eradication: concepts and research leading to the sterile-male
method. *In* Smithsonian Report for 1958. Smithsonian Inst., Washington DC, pp. 409–
418 (1959).

14. Enkerlin WR, Impact of fruit fly control programmes using the sterile insect technique.
Sterile insect technique, ed. by CRC Press, Boca Raton, pp. 979-1006 (2021).

15. Feldmann U, Dyck VA, Mattioli RC, Jannin J, Vreysen MJB, Impact of tsetse fly eradication programmes using the sterile insect technique. Sterile Insect Technique, ed. by CRC Press, Boca Raton, pp. 1051-1080 (2021).

16. Lees RS, Carvalho DO, Bouyer J, Potential impact of integrating the sterile insect technique into the fight against disease-transmitting mosquitoes». Sterile Insect Technique, ed by CRC Press, Boca Raton, pp. 1081-1118 (2021).

17. Simmons GS, Suckling DM, Carpenter JE, Addison MF, Dyck VA, Vreysen MJB, Improved quality management to enhance the efficacy of the sterile insect technique for lepidopteran pests. *J Appl Entomol* **134**:261–73 (2010).

18. Vargas-Terán M, Spradbery JP, Hofmann HC, Tweddle NE, Impact of screwworm eradication programmes using the sterile insect technique». Sterile insect technique, ed. by CRC Press, Boca Raton, pp. 949-978 (2021).

19. Bourtzis K, Vreysen MJB., Sterile Insect Technique (SIT) and Its Applications. *Insects* **12**:638 (2021).

20. Dyck VA, Hendrichs J, Robinson AS, Sterile Insect Technique: Principles and Practice in Area-Wide Integrated Pest Management, 2nd ed. by CRC Press, Boca Raton (2021).

21. McInnis DO, Rendon P, Jang E, Van Sauers-Muller A, Sugayama R, Malavasi A, Interspecific Mating of Introduced, Sterile *Bactrocera dorsalis* with Wild *B. carambolae* (Diptera: *Tephritidae*) in Suriname: a Potential Case for Cross-Species Sterile Insect Technique. *Annal Entomol Soc Am* **92**:758–65 (1999).

22. Honma A, Kumano N, Noriyuki S, Killling two bugs with one stone: a perspective for targeting multiple pest species by incorporating reproductive interference into sterile insect technique: Reproductive interference and SIT. *Pest Manag Sci* **75**:571–7 (2019).

23. Tsurui-Sato K, Fujimoto S, Deki O, Suzuki T, Tatsuta H, Tsuji K, Reproductive interference in live-bearing fish: the male guppy is a potential biological agent for eradicating invasive mosquitofish. *Sci Rep* **9**:5439 (2019).

24. Cerasti F, Mastrantonio V, Dallai R, Cristofaro M, Porretta D, Applying Satyrization to Insect Pest Control: The Case of the Spotted Wing Drosophila, *Drosophila suzukii* Matsumura. *Insects* **14**: 569 (2023).

25. Fraimout A, Debat V, Fellous S, Hufbauer RA, Foucaud J, Pudlo P, et al., Deciphering the routes of invasion of *Drosophila suzukii* by means of ABC random forest. *Mol Biol Evol* **34**:980-996 (2017).

26. Tait G, Mermer S, Stockton D, Lee J, Avosani S, Abrieux A, et al., *Drosophila suzukii* (Diptera: Drosophilidae): A Decade of Research Towards a Sustainable Integrated Pest Management Program. Brewer M, ed. by *J Econ Entomol* **114**:1950–74 (2021).

27. Bolda M, Goodhue RE, Zalom FG, Spotted wing drosophila: potential economic impact of a newly established pest. *Agric Resour Econ* **13**:5-8 (2010).

28. De Ros G, Conci S, Pantezzi T, Savini G, The economic impact of invasive pest *Drosophila suzukii* on berry production in the Province of Trento, Italy. *J Berry Res* **5**:89–96 (2015).

29. Wolf S, Collatz J, Enkerli J, Widmer F, Romeis J. Assessing potential hybridization between a hypothetical gene drive-modified *Drosophila suzukii* and nontarget *Drosophila* species. Risk Analysis. 43(10):1921–32 (2023).

30. Silva-Soares NF, Nogueira-Alves A, Beldade P, Mirth CK, Adaptation to new nutritional environments: larval performance, foraging decisions, and adult oviposition choices in *Drosophila suzukii*. *BMC Ecol* **17**:21 (2017).

31. Baccaro S, Cemmi A, Gamma irradiation CALLIOPE facility at ENEA – Casaccia Research Centre, ed. by ENEA, Rome, pp. 48 (2019).

32. Nelson FRS, *Drosophila melanogaster*: Effects of Gamma Radiation on Fecundity and Longevity. *J Econ Entomol* **66**:257–8 (1973).

33. Henneberry TJ, Effects of Gamma Radiation on the Fertility and Longevity of *Drosophila melanogaster*. *J Econ Entomol* **56**: 279–81 (1963).

34. Lasbleiz C, Ferveur JF, Everaerts C, Courtship behaviour of *Drosophila melanogaster* revisited. *Anim Behav* **72**:1001–12 (2006).

35. Revadi S, Lebreton S, Witzgall P, Anfora G, Dekker T, Becher P, Sexual Behavior of *Drosophila suzukii*. *Insects* **6**:183–96 (2015).

36. Friard O, Gamba M, BORIS: a free, versatile open source event logging software for video/audio coding and live observations. *Methods Ecol Evol* **7**:1325–30 (2016).

37. Ripley, B., Venables, B., Bates, D. M., Hornik, K., Gebhardt, A., Firth, D., & Ripley, M. B. Package 'mass'. *Cran r*, *538*, 113-120 (2013).

38. Hothorn T, Bretz F, Westfall P, Simultaneous Inference in General Parametric Models. *Biometrical J* **50**:346–63 (2008).

39. Kaplan E and Meier P, Nonparametric estimation from incomplete observations. *J Amer Statist Assoc* **53**:457–481 (1958).

40. Kassambara A, Kosinski M, Biecek P, survminer: Drawing Survival Curves using 'ggplot2. R package version 0.4.9. (2021).

41. R Core Team, A Language and Environment for Statistical Computing. R-Foundation for Computer Statistics, (2019).

42. Ernawan B, Anggraeni T, Yusmalinar S, Ahmad I, Investigation of Developmental Stage/Age, Gamma Irradiation Dose, and Temperature in Sterilization of Male *Aedes aegypti* (Diptera: Culicidae) in a Sterile Insect Technique Program. *J Med Entomol* **59**:320–7 (2022).

43. Lance DR, McInnis DO, Biological basis of the sterile insect technique». Sterile Insect Technique, ed. by CRC Press, Boca Raton, pp.113-142 (2021).

44. Sassù F, Nikolouli K, Pereira R, Vreysen MJB, Stauffer C, Cáceres C. Irradiation dose response under hypoxia for the application of the sterile insect technique in *Drosophila suzukii*. PLOS ONE. Dec 31;14(12):e0226582 (2019).

45. Arredondo J, Aguirre-Medina JF, Meza JS, Cancino J, Díaz-Fleischer F. Does the Effect of Irradiation Dose Vary Between Flies Selected and Non-selected to Resist Desiccation? The Case of *Anastrepha ludens* (Diptera: Tephritidae). Journal of Economic Entomology. Dec 1;113(6):2679–87 (2020).

46. Bakri, A., K. Mehta, and D. R. Lance. Sterilizing insects with ionizing radiation. pp. 233–269. In V. A. Dyck, J. Hendrichs, and A. S. Robinson (eds.), Sterile insect technique. principles and practice in area-wide integrated pest management. Springer, Dordrecht, the Netherlands (2005).

47. Lanouette G, Brodeur J, Fournier F, Martel V, Vreysen M, Cáceres C, et al. The sterile insect technique for the management of the spotted wing drosophila, *Drosophila suzukii*: Establishing the optimum irradiation dose. Ling E, editor. PLoS ONE. Sep 28;12(9):e0180821 (2017).

48. Chen Y, Pan H, Li J, Pan D, Liu P, Hu H. Effects of Irradiated Sterile Male and Mating
Sequence on the Fertility of *Drosophila suzukii* (Diptera: Drosophilidae). Journal of
Insect Science. Jan 1;22(1):22 (2022).

49. Yamada H, Maïga H, Kraupa C, Somda NSB, Mamai W, Wallner T, et al. Radiation
dose-fractionation in adult *Aedes aegypti* mosquitoes. Parasite.;30:5 (2023).

50. Abdel-Malek AA, Tantawy AO, Wakid AM, Studies on the Eradication of Anopheles
pharoensis Theobald by the Sterile-Male Technique Using Cobalt-60. I. Biological
Effects of Gamma Radiation on the Different Developmental Stages. *J Econ Entomol*
59:672–8 (1966).

51. Helinski ME, Parker AG, Knols BG, Radiation-induced sterility for pupal and adult
stages of the malaria mosquito *Anopheles arabiensis*. *Malar J* **5**:41 (2006).

52. Cristofaro M, Sforza RFH, Roselli G, Paolini A, Cemmi A, Musmeci S, et al., Effects of
Gamma Irradiation on the Fecundity, Fertility, and Longevity of the Invasive Stink Bug
Pest *Bagrada hilaris* (Burmeister) (Hemiptera: Pentatomidae). *Insects* **13**:787 (2022).

53. Bakri A, Heather N, Hendrichs J, Ferris I, Fifty Years of Radiation Biology in
Entomology: Lessons Learned from IDIDAS. Annals of the Entomological Society of
America. Jan 1;98(1):1–12 (2005).

54. Arthur, V., Machi, A., & Mastrangelo, T, Ionizing radiations in entomology. Evolution of
ionizing radiation research, 213-34 (2015).

55. Plá I, García de Oteyza J, Tur C, Martínez MÁ, Laurín MC, Alonso E, et al. Sterile Insect
Technique Programme against Mediterranean Fruit Fly in the Valencian Community
(Spain). Insects. May;12(5):415 (2021).

56. Oliva, C. F., Jacquet, M., Gilles, J., Lemperiere, G., Maquart, P.-O., Quilici, S.,
Schooneman, F., Vreysen, M. J. B., & Boyer, S. The Sterile Insect Technique for
Controlling Populations of *Aedes albopictus* (Diptera: Culicidae) on Reunion Island:
Mating Vigour of Sterilized Males. PLoS ONE, 7(11), e49414 (2012).

57. Markow TA, The secret lives of *Drosophila* flies. *eLife* **4**: e06793 (2015).

58. Arnqvist, G., & Rowe, L, Sexual conflict (Vol. 27). Princeton university press (2005).

59. Bonduriansky, R, The evolution of male mate choice in insects: A synthesis of ideas and
evidence. Biological Reviews, 76, 305–339 (2001).

60. Lev A, Pischedda A. Male size does not affect the strength of male mate choice for high☐quality females in *Drosophila melanogaster*. Journal of Evolutionary Biology. Sep 1;36(9):1255–65 (2023).

61. Dekker T, Revadi S, Mansourian S, Ramasamy S, Lebreton S, Becher PG, et al., Loss of *Drosophila* pheromone reverses its role in sexual communication in *Drosophila suzukii*. *Proc Roy Soc London Ser B* **282**:20143018 (2015).

62. Shaw B, Brain P, Wijnen H, Fountain MT, Reducing *Drosophila suzukii* emergence through inter-species competition: Reducing *D. suzukii* emergence through competition. *Pest Manag Sci* **74**:1466–71 (2018).

63. Manning A, A Sperm Factor Affecting the Receptivity of *Drosophila Melanogaster* Females. *Nature* **194**:252–3 (1962).

64. Gromko MH, Newport MEA, Kortier MG, Sperm Dependence of Female Receptivity to Remating in *Drosophila melanogaster*. *Evolution* **38**:1273 (1984).

65. Das SR, Maselko M, Upadhyay A, Smanski MJ. Genetic engineering of sex chromosomes for batch cultivation of non-transgenic, sex-sorted males. PLOS Genetics. Nov 2;16(11):e1009180 (2020).

66. Liu J, Rayes D, Akbari OS. A Fluorescent Sex-Sorting Technique for Insects with the Demonstration in *Drosophila melanogaster*. GEN Biotechnology. Feb;3(1):35–44 (2024).

67. Lutrat C, Giesbrecht D, Marois E, Whyard S, Baldet T, Bouyer J. Sex Sorting for Pest Control: It's Raining Men! Trends in Parasitology. Aug 1;35(8):649–62 (2019).

68. Pérez☐Staples D, Shelly TE, Yuval B, Female mating failure and the failure of 'mating' in sterile insect programs. *Entomol Exp Appl* **146**:66–78 (2013).

69. Corrêa AS, Cordeiro EM, Omoto C, Agricultural insect hybridization and implications for pest management. *Pest Manag Sci* **75**:2857–64 (2019).

70. Porretta D, Canestrelli D, The ecological importance of hybridization. *Trends Ecol Evol* **38**:1097–108 (2023).

71. Ivey V, Hillier NK, Hybridization in heliothine moths: impacts on reproduction, pheromone communication, and pest management. *Front Ecol Evol* **11**:1208079 (2023).

72. Saveer AM, Becher PG, Birgersson G, Hansson BS, Witzgall P, Bengtsson M Mate recognition and reproductive isolation in the sibling species *Spodoptera littoralis* and *Spodoptera litura*. *Front Ecol Evol* (2014).

Table 1. Effect of irradiation doses on sterility of *Drosophila melanogaster* males and females. GLM model values are shown. Values in boldface indicate significant differences

Fixed Effects	Estimate	±SE	z Value	*p*-Value
Males				
(Intercept)	4.8572	0.1306	37.191	**< 2e-16**
Control "Trip"	0.1527	0.1934	0.790	**0.43**
IRR60	-1.1004	0.1756	-6.268	**3.66e-10**
IRR80	-1.6960	0.1817	-9.335	**< 2e-16**
Females				
(Intercept)	4.2850	0.1428	30.009	**< 2e-16**
Control "Trip"	0.1144	0.2016	0.568	**0.57**
IRR80	-1.9741	0.2109	-9.358	**< 2e-16**

Table 2. Effect of irradiated *Drosophila melanogaster* males on *D. suzukii* fitness. GLMM model values are shown. Values in boldface indicate significant differences

Fixed Effects	Estimate	±SE	z Value	*p*-Value
(Intercept)	3.6361	0.1756	20,711	**< 2e-16**
40 *D. melanogaster* males	-0.8566	0.3105	-2.758	**-0.00581**
60 *D. melanogaster* males	-1.0050	0.3123	-3.218	**-0.00129**

Figure 1. Effect of irradiation on sterility of *Drosophila melanogaster* males. Comparison between the offspring originated from no-irradiated *D. melanogaster* females coupled with no-irradiated *D. melanogaster* males (green column and pink column, respectively) and irradiated at 60 Gy and 80 Gy (orange and blue column, respectively). Different letters mean significant differences by Tukey Multiple Comparison tests ($p < 0.05$).

Figure 2. Effect of irradiation on sterility of *Drosophila melanogaster* females. Comparison between the offspring originated from no-irradiated *D. melanogaster* females coupled with no-irradiated *D. melanogaster* males (green and pink columns, respectively) and irradiated females at 80 Gy (blue column). Different letters mean significant differences by Tukey Multiple Comparison tests (p<0.05).

Figure 3. Effect of different treatments on the longevity of *Drosophila melanogaster* males.
Kaplan-Meier survival curves for each condition are shown. The p-value of the Log-Rank Test is also shown.

Figure 4. Mating performance of irradiated *Drosophila melanogaster* males in no-choice tests. Time spent in conspecific and heterospecific courtship behavior by no-irradiated and irradiated *D. melanogaster* males and no-irradiated *D. suzukii* males. Percentage of the total time spent courting *D. melanogaster* females by no irradiated *D. melanogaster* males (orange column – conspecific courtship); percentage of the total time spent courting *D. melanogaster* females by irradiated *D. melanogaster* males (green column - conspecific courtship); percentage of the total time spent courting *D. suzukii* females by irradiated *D. melanogaster* males (blue column - heterospecific courtship); percentage of the total time spent courting *D. suzukii* females by no irradiated *D. suzukii* males (pink column - conspecific courtship). Tukey Multiple Comparison tests ($p > 0.05$). *D. suz = D. suzukii*; *D. mel = D. melanogaster*.

Figure 5. Mating performance of irradiated *Drosophila melanogaster* males in choice tests.
Time spent in courting *Drosophila suzukii* females by *D. suzukii* and irradiated *D. melanogaster* males. Percentage of the total time spent courting *D. suzukii* females by irradiated *D. melanogaster* males and *D. suzukii* males when placed together (green and orange column, respectively) Wilcoxon Mann–Whitney test p-value > 0.05.

Figure 6. Effect of irradiated *Drosophila melanogaster* males on *D. suzukii* fitness. Offspring originated from five pairs of *D. suzukii* without irradiated *D. melanogaster* males (green column) and with 40 (orange column) or 60 (blue column) males. Different letters mean significant differences by Tukey Multiple Comparison tests.