Endophilin A1 facilitates organization of the GABAergic postsynaptic machinery to maintain excitation-inhibition balance

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Abstract

The assembly and operation of neural circuits in the brain rely on the coordination and balance of excitatory and inhibitory activities. Inhibitory synapses are key regulators of the functional balance of neural circuits. However, due to the diversity of inhibitory presynaptic neurons, the complex composition of postsynaptic receptor subunits and the lack of typical postsynaptic dense structure, there are relatively few studies on the regulatory mechanisms for inhibitory synaptic structure and function, and insufficient understanding of the cellular and molecular abnormalities of inhibitory synapses in neurological and neuropsychiatric disorders. Here, we report a crucial role for endophilin A1 in inhibitory synapses. We show that endophilin A1 directly interacts with the inhibitory postsynaptic scaffold protein gephyrin in excitatory neurons, and promotes organization of the inhibitory postsynaptic density and synaptic recruitment/stabilization of the γ-aminobutyric acid type A receptors via its plasma membrane association and actin polymerization promoting activities. Loss of endophilin A1 by gene knockout in mouse hippocampal CA1 pyramidal cells weakens inhibitory synaptic transmission and causes imbalance in the excitatory/inhibitory function of neural circuits, leading to increased susceptibility to epilepsy. Our findings identify endophilin A1 as an iPSD component and provide new insights into the organization and stabilization of inhibitory postsynapses to maintain E/I balance as well

as the pathogenesis of epilepsy.

Key words: endophilin A1, E/I balance, epilepsy, $GABA_AR$, gephyrin, inhibitory synapse

Introduction

The excitatory and inhibitory synaptic activities of neural circuits are tightly orchestrated in the central nervous system (Sakimoto et al., 2021). The imbalance between excitatory and inhibitory neuronal activity is implicated in many neurological and neuropsychiatric diseases including schizophrenia, autism spectrum disorder and epilepsy (Canitano and Pallagrosi, 2017; Canitano and Palumbi, 2021; Foss-Feig et al., 2017; Fritschy, 2008). In the brain, in contrast to asymmetric excitatory synapses formed between axonal boutons and dendritic postsynaptic structures with thick disc-shaped PSD, inhibitory synapses are symmetric and are constituted of the presynaptic terminal from diverse interneurons and the postsynaptic plasma membrane with thin sheet-like PSD where receptor complexes are anchored (Bai et al., 2021; Favuzzi et al., 2019; Liu et al., 2020; Mody and Pearce, 2004; Tao et al., 2018). Although the formation, maturation and plasticity of both excitatory and inhibitory synapses are vitally important for neural circuit functioning, much more research efforts have been focused on the regulatory mechanisms governing excitatory synaptic structure and function. The mechanisms by which inhibitory synapses are constructed in the brain and fine-tuned in response to neural activity remain largely unexplored.

The γ-aminobutyric acid type A receptors ($GABA_ARs$), major targets of antiseizure drugs, are heteropentameric chloride channels that mediate fast synaptic transmission at inhibitory synapses in the brain (Scott and Aricescu, 2019). Several neuropathologies are characterized by alterations in $GABA_A$ receptor trafficking, synaptic E/I ratio and neuronal excitability (Sakimoto et al., 2021). At the inhibitory postsynaptic site, synaptic localization and functioning of $GABA_ARs$ requires trans-synaptic cell adhesion molecules, submembranous scaffold proteins and intracellular signaling proteins. Gephyrin is a master organizer of iPSD, which self-assembles into

planar lattices beneath the plasma membrane and anchors $GABA_A$Rs or glycine receptors (GlyRs) to the postsynaptic sites (George et al., 2021; Kasaragod and Schindelin, 2019; Kirsch et al., 1991; Tyagarajan and Fritschy, 2014). Collybistin, a membrane lipid-binding protein, functions in recruitment of gephyrin and $GABA_A$Rs to the plasma membrane by interacting with gephyrin, the synaptic adhesion molecule neuroligin-2 (NL2) and the α2 subunit of $GABA_A$Rs (Kalscheuer et al., 2009; Kins et al., 2000; Poulopoulos et al., 2009; Saiepour et al., 2010). The transmembrane protein Shisa7 binds $GABA_A$Rs and regulates their trafficking, synaptic abundance and functional properties via binding to protein(s) other than gephyrin (Han et al., 2019; Liu et al., 2020). Nevertheless, mechanisms governing the assembly, recruitment and maintenance of various inhibitory synaptic components remain largely enigmatic.

The majority of inhibitory synapses are formed on the dendritic shaft, or around the cell body of excitatory principal neurons where both actin and microtubule cytoskeletons are present underneath the plasma membrane. Through direct interaction with polymerized tubulin, gephyrin organizes the iPSD by anchoring the GlyRs and $GABA_A$Rs to the microtubule cytoskeleton (Parato and Bartolini, 2021). Notably, although small F-actin patches are observed within the dendritic shaft where the majority of inhibitory synapses are assembled (van Bommel et al., 2019), role(s) of the actin cytoskeleton in the organization and maintenance of inhibitory synapses is still unclear.

Endophilin A1 (EndoA1) is a member of the endophilin A protein family characterized by an amino (N)-terminal membrane-binding N-BIN/amphiphysin/Rvs (BAR) domain and a carboxyl (C)-terminal Src homology 3 (SH3) domain. The gene encoding endophilin A1 (*EndoA1*, gene name *sh3gl2*) is almost exclusively expressed in the brain (Ringstad et al., 1997), and has been implicated in epilepsy, schizophrenia, depression, Parkinson's disease, and Alzheimer's disease (Chang et al., 2017; Corponi et al., 2019; Germer et al., 2019; Ren et al., 2008; Yu et al., 2018a; Yu et al., 2018b). In excitatory synapses, endophilin A1 functions in synaptic vesicle recycling and autophagosome formation at the axon terminal (Bademosi et al., 2023; Milosevic et al.,

2011; Murdoch et al., 2016; Ringstad et al., 1999; Schuske et al., 2003; Soukup et al., 2016; Verstreken et al., 2003; Watanabe et al., 2018), and regulates the morphogenesis, maturation, and acute structural plasticity of dendritic spines by promoting actin polymerization via its effector protein p140Cap (Yang et al., 2021; Yang et al., 2018; Yang et al., 2015).

Previous studies reported that the double knockout (KO) mice of *EndoA1* and *EndoA2* exhibit spontaneous epileptic seizures (Milosevic et al., 2011). However, *EndoA1* depletion only causes a slight decrease in excitatory synaptic transmission in hippocampal CA1 neurons (Yang et al., 2021; Yang et al., 2018). As E/I imbalance is caused by changes in either excitatory or inhibitory activity, or both, we were prompted to investigate the biological function of endophilin A1 in the inhibitory synapses. In this study, we show that endophilin A1 interacts with gephyrin and localizes to GABAergic postsynapses. Via its membrane-binding and actin polymerization-promoting activities, endophilin A1 facilitates organization of the inhibitory postsynaptic machinery. Loss of endophilin A1 causes impairment of inhibitory synaptic transmission, E/I imbalance and aggravation of epileptic susceptibility.

Results

Ablation of endophilin A1 causes E/I imbalance and increases epilepsy susceptibility

Previous studies found that *EndoA1* and *EndoA2* double knockout (KO) mice exhibit spontaneous epileptic seizures (Milosevic et al., 2011), suggesting that endophilin A1 and A2 are involved in maintaining the E/I balance. However, another study reported that the protein levels of endophilin A1 were higher in temporal neocortex neurons of temporal lobe epilepsy (TLE) patients (Yu et al., 2018b). Moreover, in a chemical kindling mouse model of epilepsy, in which mice are chronically induced with the $GABA_AR$ antagonist pentylenetetrazole (PTZ) (Dhir, 2012), endophilin A1 was also upregulated in the hippocampus and adjacent temporal cortex, and silencing its expression in the hippocampal CA3 region reduced PTZ-induced seizure susceptibility

and severity (Yu et al., 2018b). To investigate the role of endophilin A1 in epilepsy, we determined the impact of *EndoA1* gene ablation on chemical kindling of pan-neural *EndoA1* KO mice (Yang et al., 2018) with PTZ at a sub-convulsive dose of 25 mg/kg. Compared with the wild-type, PTZ-induced epileptic behaviors were significantly exacerbated in *EndoA1* KO mice (Figure 1A-C and Video S1).

We previously found that *EndoA1* KO caused a slight decrease in the amplitude of evoked excitatory postsynaptic currents (eEPSC) and a significant attenuation of long-term potentiation (Yang et al., 2018). As imbalance between excitatory and inhibitory signals in neuronal circuits is implicated in epilepsy, we then determined whether loss of endophilin A1 in excitatory neurons affects the E/I ratio by electrophysiological analysis. To this end, we eliminated the *EndoA1* gene in pyramidal neurons by stereotaxic injection of adeno-associated virus (AAV) expressing CaMKIIα promoter-driven Cre recombinase into the hippocampal CA1 region of *EndoA1*$^{fl/fl}$ mice (Figure 1D). Whole-cell patch clamp recording of CA1 pyramidal cells indicated that, while the eEPSC and its paired-pulse ratio were not significantly changed, the evoked inhibitory postsynaptic current (eIPSC), not its PPR, was decreased in the *EndoA1* KO neurons, resulting in a significant increase in the E/I ratio (Figure 1E-I). Of note, neither the frequency nor the amplitude of miniature EPSCs (mEPSC) or miniature IPSCs (mIPSC) was altered in *EndoA1* KO neurons (Figure 1J-O). Moreover, neither fast mIPSCs (perisomatic events: < 2.8 ms) nor slow mIPSCs (distal dendritic events: > 2.8 ms) (Miles et al., 1996) were affected by *EndoA1* KO (Figure S1), indicating that presynaptic quantal release of soma- or dendrite-targeting inhibitory synapses is normal. Taken together, these data indicate that endophilin A1 in hippocampal pyramidal neurons functions in inhibitory synaptic transmission to maintain E/I balance.

Endophilin A1 is required for the formation and stabilization of inhibitory synapses

Diminished eIPSC in *EndoA1*$^{-/-}$ pyramidal neurons prompted us to determine the role of endophilin A1 in inhibitory synapses by immunofluorescence staining of brain

sections from *EndoA1*$^{-/-}$ mice. Analysis of confocal microscopy images revealed that, compared with the wild-type, there was a significant decrease in fluorescent signals of both VGAT and gephyrin, the inhibitory pre- and post-synaptic markers, or VGLUT1 and PSD95, the excitatory pre- and postsynaptic markers, in all subregions of the hippocampus of the *EndoA1*$^{-/-}$ mice on postnatal day 21 (P21) (Figure 2A,C,D,F, and S2A,C,D,F), suggesting defects in synapse development. While a slight reduction in excitatory presynaptic protein was observed, the decreases in inhibitory but not excitatory postsynaptic proteins were also detected in the *EndoA1*$^{-/-}$ mice at P60 (Figure 2B,C,E,F, and S2B,C,E,F), indicating that the assembly and/or stabilization of inhibitory postsynapses was impaired throughout brain development. Notably, gephyrin signals in all layers of the different hippocampal subregions were reduced in the *EndoA1*$^{-/-}$ mice (Figure S2G,H). In line with the data from immunofluorescent labeling (Figure 2 and S2), decreased endogenous PSD95 or gephyrin clusters at synapses were also visualized by the recombinant probes, PSD95.FingR-eGFP or mRuby2-gephyrin.FingR, respectively (Bensussen et al., 2020; Gross et al., 2013), in *EndoA1* KO neurons in the CA1 region at P21 (Figure S3A-D). As different types of interneurons synapse with hippocampal excitatory neurons at distinct subcellular compartments (soma, axon initial segment, proximal and distal dendrites) indicated by the layered structures of the hippocampus (Spruston, 2008), these data suggest that endophilin A1 functions in the assembly/stabilization of inhibitory synapses formed between various presynaptic neurons and postsynaptic neurons.

Next we analyzed the $GABA_A$ receptors and found that, consistent with changes in inhibitory postsynaptic marker, signals of the $GABA_AR$ γ2 subunit were decreased in the hippocampi of both juvenile and adult *EndoA1*$^{-/-}$ mice (Figure 2G-I). These data collectively indicate that endophilin A1 orchestrates the assembly of inhibitory synapses, the architecture of inhibitory postsynapses in particular, and functioning of GABAergic synapses.

Endophilin A1 localizes to the iPSD and augments the organization of the

inhibitory postsynaptic machinery

To substantiate the role of postsynaptic endophilin A1 in the assembly of inhibitory synapses and exclude the impacts of presynaptic endophilin A1 and its depletion on early embryonic development, we determined the effects of endophilin A1 KO on inhibitory and excitatory synapses in cultured hippocampal pyramidal neurons during late developmental stages. As the expression levels of $GABA_AR$ α1 are highly heterogeneous in individual cultured neurons (Figure S4A), we determined the effect of Cre-mediated *EndoA1* KO on surface expression of $GABA_AR$ γ2 in *EndoA1*$^{fl/fl}$ hippocampal neurons by immunofluorescence staining. Whilst no changes in surface GluA1 was detected, the fluorescence signal intensity and number of surface $GABA_AR$ γ2 clusters were largely lower in dendrites of *EndoA1* KO neurons, which was rescued by re-expression of endophilin A1 (Figure 3A-D). We further examined the total and surface levels of synaptic proteins by immunoblotting. Biotinylation assay of surface proteins revealed that, compared with wild-type, although there was no change in total levels of synaptic proteins in *EndoA1* KO pyramidal neurons, the surface expression of $GABA_ARs$ but not NL2 or the NR1 subunit of the NMDA-type glutamate receptor was significantly reduced (Figure 3E,F). Collectively, these data indicate that postsynaptic endophilin A1 enhances the surface clustering of $GABA_ARs$ and facilitates the assembly of inhibitory postsynapses in pyramidal neurons.

Next we sought to determine the molecular function of endophilin A1 in inhibitory synapses at the cellular level. By immunofluorescence staining and super-resolution microscopy of hippocampal neurons in dissociated culture, we observed partial colocalization of endophilin A1 with gephyrin as well as surface $GABA_AR$ γ2 in dendrites of hippocampal neurons, and juxtaposition or partial colocalization of endophilin A1 and VGAT puncta (Figure 3G-I and Video S2-4). We further detected the localization of endophilin A1 to inhibitory synapses by co-immunostaining with both pre- and post-synaptic markers (Figure. S4B and Video 5). Quantitative analysis of super-resolution localization maps revealed that ~ 47 % puncta of gephyrin or Bassoon were proximal to endophilin A1 (Figure. S4G, n = 14), with a mean distance between

endophilin A1- and gephyrin-positive pixels of ～ 120 nm, or between endophilin A1- and Bassoon-positive pixels of ～ 130 nm (Figure. S4C-F). These results indicate that endophilin A1 localizes to not only ePSD but also iPSD. Therefore, we speculated that endophilin A1 acted as a component of iPSD. Previous studies reported that synaptic $GABA_A$Rs contain the γ2 subunit along with α and β subunits (e.g., α1β2γ2) (Scott and Aricescu, 2019). The γ2 subunit is essential for receptor clustering at the synapse, mediated by interactions with scaffold proteins (Essrich et al., 1998). Indeed, co-immunoprecipitation (coIP) of endogenous proteins in membrane fractions from mouse brain lysates with antibodies against $GABA_A$R α1 detected not only the known iPSD components γ2, NL2 and gephyrin, but also endophilin A1, and vice versa (Figure 3J). Of note, loss of endophilin A1 caused decreases in levels of $GABA_A$R γ2 and gephyrin but not NL2 in anti-$GABA_A$R α1 immunoprecipitates (Figure 3K). Further, immunoblotting of synaptosome fractions revealed that the levels of $GABA_A$R γ2 and gephyrin were reduced at synapses in adult *EndoA1* KO mice (Figure 3L). Together, these data support that endophilin A1 functions at the iPSD to promote the assembly and/or stabilization of the GABAergic postsynaptic machinery.

Endophilin A1 directly interacts with the scaffold protein gephyrin and promotes its clustering and localization of $GABA_A$Rs to iPSD

We subsequently sought to investigate the mechanistic role of endophilin A1 in the clustering of inhibitory postsynaptic proteins. To this end, we performed coIP from mouse brain lysates with antibodies against endophilin A1 (Figure 4A) and analyzed the immunoprecipitates by mass spectrometry. Gene Ontology (GO) analysis revealed that, in addition to proteins participating in known cellular processes involving endophilin A1, such as synaptic vesicle recycling, actin cytoskeleton remodeling, endocytosis and long-term potentiation (Gad et al., 2000; Ringstad et al., 1999; Schuske et al., 2003; Verstreken et al., 2003; Yang et al., 2021; Yang et al., 2018; Yang et al., 2015), proteins related to not only glutamatergic but also dopaminergic and GABAergic synapses were also identified, including gephyrin (Figure 4B and Table S1).

Gephyrin is the major scaffold protein which interacts with the intracellular domains of $GABA_A$Rs and anchors them at the inhibitory postsynaptic membrane (Essrich et al., 1998; Kneussel et al., 1999; Mukherjee et al., 2011; Tretter et al., 2008). The cell adhesion molecule NL2 drives postsynaptic assembly at perisomatic inhibitory synapses via interaction with gephyrin and the membrane lipid-binding protein collybistin (Poulopoulos et al., 2009). We speculated that endophilin A1 functions in the assembly of iPSD via interaction with the scaffold and/or intercellular adhesion proteins. To determine which inhibitory synaptic protein(s) interacts with endophilin A1, we performed coIP from HEK293T cells transiently expressing endophilin A1 and NL2, or gephyrin, and detected interaction of endophilin A1with gephyrin but not NL2 (Figure 4C and 4D). Moreover, coIP from mouse brain lysates corroborated interaction between endophilin A1 and gephyrin but not $GABA_AR$ α1 or γ2 (Figure 4E). Further, neither endophilin A2 nor A3 interacted with gephyrin (Figure 4F).

Notably, although the expression levels of gephyrin were not affected by ablation of *EndoA1*, its distribution in the PSD fraction was decreased (Figure 3L). In dendrites of mature cultured hippocampal neurons following *EndoA1* depletion, the signal intensity and number of gephyrin.FingR-eGFP clusters were also significantly attenuated, with no obvious changes in PSD95.FingR-eGFP clusters (Figure S5A-D), which was consistent with the reduced gephyrin but unaltered PSD95 labeled by antibodies in brain slice of mature *EndoA1* KO mice (Figure 2 and S2), Moreover, immunofluorescence staining revealed a decrease in the clustering of gephyrin across the soma, the axon initial segment (AIS), and dendrites (Figure S5E-H). The gephyrin clustering phenotype was rescued by expression of endophilin A1 but not endophilin A2 or A3 (Figure 4G,H), suggesting that their functions at the inhibitory postsynapses are not redundant. Conversely, overexpression of endophilin A1 caused an increase in the mean signal intensity and number of gephyrin puncta in the soma and dendrites (Figure 4I,J). Collectively, these data demonstrate that endophilin A1 interacts with the scaffold protein gephyrin and facilitates its clustering at inhibitory postsynapses.

At the submembraneous postsynaptic density, gephyrin self assembles via the N-

terminal G and C-terminal E domains to provide scaffold for inhibitory postsynapses and anchors $GABA_A$Rs to the postsynaptic sites. Consistent with the effect of *EndoA1* KO on the clustering of gephyrin (Figure 4G,H) and $GABA_A$R γ2 (Figure 3A-D), immunofluorescence staining and confocal microscopy analysis showed that, depletion of endophilin A1 also caused a decrease in the localization of $GABA_A$R γ2 to gephyrin puncta in *EndoA1* KO neurons (Figure 5A-C). Moreover, overexpression of endophilin A1 also enhanced clustering of $GABA_A$R γ2 in the soma and dendrites (Figure 5D,E). Together these data indicate that endophilin A1 promotes anchoring of $GABA_A$Rs to the iPSD.

To further understand the mechanism by which endophilin A1 regulates the assembly and function of inhibitory postsynapses, next we attempted domain mapping of the endophilin A1-gephyrin interaction. To this end, we generated a series of truncation and internal deletion mutants for endophilin A1 or gephyrin (Figure 5F). For those which did not express in mammalian cells, we determined their interaction with gephyrin by in vitro binding assays with recombinant proteins. None of the truncation or internal deletion mutants lost the ability to bind to gephyrin, and deletion of neither the BAR nor the SH3 domain weakened the binding (Figure 5G,H), suggesting that they are involved in the interaction with gephyrin. Pull-down assay showed both G and E domains but not C domain of gephyrin were involved in the interaction with endophilin A1 (Figure 5I). Indeed, the decreases in clustering of either gephyrin or $GABA_A$R γ2 in *EndoA1* KO neurons were not rescued by expression of endophilin A1 fragments (Figure 5J-M), indicating that all functional domains of endophilin A1 are required for the assembly of the GABAergic postsynaptic machinery.

Actin polymerization is required for stabilization of inhibitory postsynapses

Endophilin A1 functions in the morphogenesis and structural plasticity of dendritic spines by promoting actin polymerization underneath the plasma membrane (Yang et al., 2021; Yang et al., 2018; Yang et al., 2015). Of note, previous studies reported controversial results about the role of the actin and microtubule cytoskeleton in gephyrin clustering (Allison et al., 2000; Charrier et al., 2006; Kirsch and Betz, 1995).

To further investigate the mechanism underlying endophilin A1-mediated assembly of inhibitory postsynapses, we asked whether actin polymerization contributes to the stabilization of iPSD and $GABA_AR$ clustering in dendrites. Consistent with previous studies (van Bommel et al., 2019), we observed large patches of F-actin labeled by LifeAct-mCherry in spines and small puncta in dendritic shafts that exhibited partial colocalization with or adjoined gephyrin in cultured hippocampal neurons by super-resolution microscopy (Figure S6A and Video S6). We then determined the impact of F-actin depolymerization on gephyrin-labeled iPSD and γ2-containing $GABA_ARs$ in cultured hippocampal neurons. Treatment of mature neurons with nocodazole, a microtubule depolymerizing reagent, for one hour (short-term) or two hours (long-term), caused decreases in the number of both PSD95 and gephyrin puncta (Figure 6A-D), which was in good agreement with previous findings that microtubule dynamics regulates the spatial organization of both excitatory and inhibitory postsynaptic structures (Gu et al., 2008; Jaworski et al., 2009; Kirsch and Betz, 1995; Kirsch et al., 1991). Short-term or long-term incubation with latrunculin A, which disrupts actin polymerization, also caused decreases in the number of both ePSD and iPSD puncta (Figure 6A-D), indicating that actin polymerization also contributes to the stabilization of the iPSD. We then tested the effect of actin depolymerization on $GABA_AR$ clustering and found that, long-term but not short-term latrunculin A application caused a reduction in the number of $GABA_AR$ γ2 puncta in dendrites (Figure 6E-H), this finding supported the observation that the decrease in gephyrin cluster preceded the reduction in $GABA_AR$ γ2 receptor cluster following latrunculin A treatment (Figure 6I). In addition, colocalization analysis revealed less recruitment of γ2 to gephyrin puncta even after short-term drug application, without changes in γ2 clustering (Figure 6J,K). Together, these data demonstrate that actin polymerization is essential for the stabilization of iPSD and anchoring of $GABA_ARs$ to the postsynaptic sites.

Both the membrane-binding and p140Cap-binding capacities of endophilin A1 are indispensable for its function at the inhibitory postsynapses

At the excitatory synapse, the plasma membrane-associated endophilin A1 regulates the morphogenesis, maturation and structural plasticity of dendritic spines by promoting branched actin polymerization underneath the postsynaptic membrane via its interaction with the cytoskeleton regulator p140Cap (Yang et al., 2021; Yang et al., 2018; Yang et al., 2015). Having established the requirement for both endophilin A1 and the actin cytoskeleton in the assembly and stabilization of the inhibitory postsynaptic structure, we speculated that regulation of inhibitory postsynapse assembly by endophilin A1 necessitated its plasma membrane-binding and actin polymerization-promoting activities. Silencing p140Cap expression in postsynaptic CA1 pyramidal cells by shRNA-mediated knockdown caused decreases in the clustering of gephyrin puncta in dendrites, which was restored by overexpression of p140Cap (Figure S6B, C), suggesting that p140Cap-mediated actin polymerization is required for the assembly of iPSD in postsynaptic neurons. However, compared with mere depletion of endophilin A1, p140Cap knockdown did not further attenuate the clustering of gephyrin in *EndoA1* KO neurons (Figure S6D, E). Moreover, overexpression of p140Cap did not rescue the iPSD phenotype in *EndoA1* KO neurons (Figure 7A,B), suggesting that p140Cap-enhanced actin polymerization promotes the organization of iPSD through endophilin A1-mediated pathway. Indeed, overexpression of neither the membrane-binding deficient (KKK-EEE) nor the p140Cap-binding deficient (Y343A) mutant of endophilin A1 (Yang et al., 2021) could rescue the iPSD defect (Figure 7A,B). Consistently, neither p140Cap nor the endophilin A1 loss-of-function mutants could rescue the $GABA_AR$ γ2 clustering phenotype in *EndoA1* KO neurons (Figure 7C,D). Collectively, these data indicate that both plasma membrane association and actin polymerization-promoting activities are required for endophilin A1 to promote the assembly of the inhibitory postsynaptic machinery.

Next, we determined the mechanistic role of endophilin A1 in vivo by rescuing the epilepsy susceptibility caused by *EndoA1* KO. Indeed, when AAVs expressing the Cre recombinase and endophilin A1 were co-injected into the hippocampal CA1 region of *EndoA1*$^{fl/fl}$ mice, only the wild-type but not the membrane-binding or p140Cap-binding

deficient mutant of endophilin A1 rescued the early onset and severity of PTZ-induced seizures (Figure 7E,F and Video S7). Consistently, electrophysiological recording in acute brain slices indicated that only wild-type endophilin A1 but not the mutants restored the amplitude of eIPSC and E/I balance in *EndoA1* KO neurons (Figure 7G-K). All together, these data indicate that endophilin A1 contributes to the structure and function of inhibitory synapses by promoting the submembrane assembly of iPSD and synaptic localization of the $GABA_A$Rs, thereby maintaining the balance of excitatory and inhibitory activities in neural circuits (Figure 8).

Discussion

Although collybistin is implicated in plasma membrane recruitment and clustering of gephyrin and $GABA_A$R α2 (Saiepour et al., 2010), studies of collybistin KO mice showed that the $GABA_A$R α3 subunit promotes gephyrin-mediated formation of inhibitory synapses in the absence of collybistin (Wagner et al., 2021), raising the possibility that formation of a subset of GABAergic synapses requires gephyrin-interacting membrane-associated proteins other than collybistin. In the present study, we identify endophilin A1 as a novel binding partner for gephyrin and demonstrate that it promotes the assembly of inhibitory synapses and clustering of $GABA_A$Rs at the iPSD. Loss of endophilin A1 activity causes impairment of inhibitory synaptic transmission in neuronal circuits and increases susceptibility to seizures. In the central nervous system, inhibitory interneurons are highly diverse in morphologies, molecular profiles and physiological functions. Presynaptic terminals from different types of interneurons target distinct subcellular compartments in postsynaptic pyramidal neurons where $GABA_A$ receptors of specific subunit compositions are located. Parvalbumin-expressing (PV+) interneurons usually contact the soma and proximal dendrites, whereas somatostatin-expressing (SST+) interneurons mainly innervate the dendrites of pyramidal cells (Klausberger and Somogyi, 2008). Although different inhibitory synapses localized in different subcellular compartments of postsynaptic neurons have distinct properties and express a diversity of synaptic proteins, gephyrin

379 remains a core postsynaptic component (Favuzzi et al., 2019). Our study revealed that
380 endophilin A1 promotes the clustering of gephyrin in all subcellular compartments of
381 pyramidal neurons, indicating that endophilin A1 functions in the organization of
382 inhibitory postsynapses innervated by various presynaptic interneurons.

383 Among the endophilin A proteins, endophilin A1 is the most abundant in the brain
384 (Milosevic et al., 2011). Although they have redundant roles in physiological or
385 pathological functions (Milosevic et al., 2011), endophilin A1, but not A2 or A3,
386 promotes spine morphogenesis and excitatory postsynaptic organization (Yang et al.,
387 2018; Yang et al., 2015; Milosevic et al., 2011). Here, we uncover a previously
388 unanticipated role for endophilin A1 at GABAergic synapses. By deleting endophilin
389 As in specific neuron subtypes, we found that endophilin A1 regulates GABAergic
390 postsynaptic compartments. Endophilin A1, rather than A2 or A3, interacts with
391 gephyrin and modulates the inhibitory postsynaptic machinery. Although endophilin A1
392 might function redundantly with other endophilin As in synaptic vesicle recycling at
393 the presynaptic site, it participates in the organization or reorganization of postsynaptic
394 structures through specific binding to postsynaptic proteins, a function that cannot be
395 fully replaced by other endophilin As at both excitatory and inhibitory postsynapses.

396 The membrane lipid-binding protein collybistin interacts with the trans-synaptic
397 cell adhesion molecule NL-2 to recruit gephyrin and $GABA_AR$ α2 to the postsynaptic
398 membrane (Kalscheuer et al., 2009; Kins et al., 2000; Poulopoulos et al., 2009; Saiepour
399 et al., 2010). Adamtsl3 secreted from presynapses binds to the transmembrane protein
400 DCC (Deleted in Colorectal Cancer) and controls its synaptic localization, which is
401 required for the maintenance of GABAergic synapses in the adult brain (Cramer et al.,
402 2023). Anchoring of the α5 subunit of $GABA_AR$ to the actin cytoskeleton through
403 radixin, an actin-binding protein, facilitates its clustering (Loebrich et al., 2006). These
404 findings underscore the necessity for investigation of mechanism(s) regulating the
405 plasma membrane-iPSD-actin cytoskeleton interaction. As membrane-binding and
406 actin polymerization-promoting activities of endophilin A1 are both required for its
407 function in enhancing iPSD formation and γ2–containing $GABA_AR$ clustering to iPSD,

408 we propose that membrane-bound endophilin A1 promotes postsynaptic assembly by
409 coordinating the plasma membrane tethering of the postsynaptic protein complex and
410 its stabilization with the actin cytomatrix (Figure 8). In contrast, clustering of β2/3
411 subunit-containing $GABA_A$Rs is independent of the actin or microtubule cytoskeleton
412 in hippocampal neurons (Allison et al., 2000; Kasaragod et al., 2022). Given that
413 synaptic and extrasynaptic $GABA_A$Rs are composed of distinct subunits (Kasaragod et
414 al., 2022; Koh et al., 2023), these findings indicate that the actin cytoskeleton plays
415 distinct roles in the clustering and possibly dynamic changes in the subunit composition
416 and subcompartmental localization of different types of $GABA_A$Rs. Of note, the
417 subcellular distribution of gephyrin, which has a tubulin-binding motif, is not affected
418 by depolymerization of F-actin or microtubule (Allison et al., 2000). Overall, the
419 discrepancy in data about the involvement of the actin cytoskeleton in inhibitory
420 synapses may be due to differences in drug concentration, treatment time, receptor
421 subtypes and neuronal cell types (Mody and Pearce, 2004; Sexton et al., 2021). In the
422 present study, the impaired clustering of gephyrin and $GABA_A$ γ2 by F-actin
423 depolymerization underscores the essential role of F-actin in the assembly and
424 stabilization of the inhibitory postsynaptic machinery. Membrane-bound endophilin A1
425 promotes F-actin polymerization beneath the plasma membrane through its interaction
426 with p140Cap, an F-actin regulatory protein, thereby facilitating and/or stabilizing the
427 clustering of gephyrin and γ2-containing $GABA_A$ receptors at postsynapses.

428 The number of synaptic $GABA_A$ receptors is subject to alterations not only during
429 development and neuronal plasticity but also during pathological processes such as
430 epilepsy (Nusser et al., 1998). Synaptic $GABA_A$ receptors are recruited directly from
431 extrasynaptic pools following exocytosis, and their endocytosis occurs exclusively at
432 extrasynaptic sites (Bogdanov et al., 2006). Therefore, lateral diffusion of $GABA_A$Rs
433 between synaptic and extrasynaptic sites and their iPSD anchoring are very important
434 for their synaptic distribution (Thomas et al., 2005). In good agreement with the finding
435 that $GABA_A$Rs stabilization at synapses depends on gephyrin clustering (Jacob et al.,
436 2005), we found that decreases in gephyrin clustering precedes the reduction in surface

437 $GABA_ARs$ following F-actin depolymerization (Figure 6). As the core scaffold protein
438 in iPSD, gephyrin is highly enriched at the inhibitory postsynaptic sites and drives the
439 formation of iPSD sheets via phase separation (Bai et al., 2021). As binding of gephyrin
440 by its partner proteins inhibits the iPSD condensate formation via autoinhibition of the
441 multivalent interactions between its E domain and $GABA_ARs$ (Bai et al., 2021;
442 Hoffmann and Milovanovic, 2021; Liu et al., 2020), it is conceivable that endophilin
443 A1 binding not only tethers the scaffold protein to the plasma membrane, but also
444 releases it from the autoinhibitory conformation to facilitate the phase separation of the
445 gephyrin-receptor complexes. Whether the activity of endophilin A1 is regulated by
446 neuronal activity to participate in the structural and functional remodeling of iPSD
447 awaits further investigation.

448 At the presynaptic site of excitatory synapses, endophilin A1 functions in synaptic
449 vesicle recycling by recruiting the clathrin-uncoating factor synaptojanin (Milosevic et
450 al., 2011; Schuske et al., 2003; Verstreken et al., 2003). At the excitatory postsynapses,
451 plasma membrane-associated endophilin A1 facilitates the morphogenesis, stabilization,
452 maturation and plasticity of dendritic spines by promoting actin polymerization via its
453 interaction with p140Cap (Yang et al., 2021; Yang et al., 2018; Yang et al., 2015). In
454 this study, we demonstrate that endophilin A1 functions upstream of p140Cap in not
455 only excitatory but also inhibitory postsynapses of excitatory neurons. Intriguingly,
456 recent studies reported that p140Cap is present in the inhibitory presynaptic
457 compartment and regulates GABAergic synaptogenesis and neurotransmission (Russo
458 et al., 2019), raising the possibility that endophilin A1 and p140Cap also coordinate to
459 regulate the organization and activity of GABAergic synapses at the inhibitory
460 presynaptic site. Remodeling of inhibitory synapses and dendritic spines are spatially
461 clustered by sensory inputs in the central nervous system (Chen et al., 2012). Notably,
462 the turnover of inhibitory synapses outpace that of excitatory synapses (Chen et al.,
463 2012). Therefore, it is conceivable that deficits in inhibitory synapses are much more
464 severe in *EndoA1* KO mice, leading to perturbation of the E/I balance and increased
465 epileptic vulnerability.

Materials and Methods

Ethics statement

All animal experiments were approved by and performed in accordance with the guidelines of the Animal Care and Use Committee of Institute of Genetics and Developmental Biology, Chinese Academy of Sciences (Approval code: AP2022008). All animals were housed in standard mouse cages at 22-24°C on a 12 h light/dark cycle with access to food and water freely.

Animals

Generation of *EndoA1*$^{fl/fl}$ and *EndoA1*$^{-/-}$ (KO first) mice on the C57BL/6J background were as previously described. *EndoA1*$^{+/+}$ and *EndoA1*$^{-/-}$ mice were bred as littermates from heterozygous breeders. Genotyping of mouse lines was performed by genomic PCR of tail prep DNA from offspring with the following primer pairs: loxPF/loxPR: 5'-CAAGGACTCCCAGAGACCTAGCATC-3' and 5'-GAGATGGCGCAACGCAATTAAT-3' (A PCR product of 375 base pairs in *EndoA1* KO first mice but none in wild-type mice). zptF/zptR: 5'-gtaagcggctctagcgcatgttct-3' and 5'-GCAGGGGCATGTAGGTGGCTCAAC-3' (A PCR product of 466 base pairs in wild-type mice, none in *EndoA1*$^{-/-}$ mice, and of 627 base pairs in *EndoA1*$^{fl/fl}$ mice).

Constructs and viruses

pCMV-Tag2B-EndoA1, pCMV-Tag2B-p140Cap, pCMV-Tag2B-EndoA1 Y343A, pCMV-Tag2B-EndoA1 KKK-EEE, pCMV-Tag2B-EndoA1 (aa172-352), pGEX-4T-1-SH3 (aa295-352), pET-28a (+)-EndoA1 and LifeAct-mCherry constructs were described previously (Yang et al., 2021; Yang et al., 2015). pAOV-CAMKIIα-mCherry-2A-3FLAG were purchased from OBiO Technology (Shanghai) Corp. Ltd., (Shanghai, China). pNICE-NL2(-) was a gift from Peter Scheiffele (Addgene plasmid # 15246) (Chih et al., 2006). pCR3-FLAG-Gephyrin P1 was a gift from Shiva Tyagarajan (Addgene plasmid # 68816) (Tyagarajan et al., 2013). pCAG_PSD95.FingR-eGFP-

CCR5TC and pCAG_GPN.FingR-eGFP-CCR5TC were gifts from Don Arnold (Addgene plasmid # 46295 & #46296) (Gross et al., 2013). The pCMV-Tag3B-GPN construct was generated by cloning gephyrin cDNA amplified from pCR3-FLAG-gephyrin into pCMV-Tag3B (Clontech Laboratories, Inc.). The pGEX-4T-1 EndoA1 ΔBAR (Δaa6-242) and pGEX-4T-1 EndoA1 ΔSH3 (Δaa1-352) constructs were generated by cloning fragments amplified from pCMV-Tag2B-EndoA1 into pGEX-4T-1. The pGEX-4T-1 GPN G (aa15-169), pGEX-4T-1 GPN C (aa170-326) and pGEX-4T-1 GPN E (aa327-733) constructs were generated by PCR amplification from full-length constructs and subcloned into pGEX-4T-1. All constructs were verified by DNA sequencing. Viral particles of adeno-associated virus (AAV) carrying mCherry, EGFP, the Cre recombinase, EndoA1 WT or mutants were purchased from OBiO Technology (Shanghai) Corp. Ltd., (Shanghai, China). Viral particles of rAAV-EF1a-PSD95.FingR-eGFP-CCR5TC and rAAV-EF1a-mRuby2-Gephyrin.FingR-IL2RGTC (Bensussen et al., 2020) were purchased from brain case Corp. Ltd., (Shenzhen, China).

Antibodies

The following antibodies were obtained from commercial sources: rabbit anti-endophilin A1 (159 002, Synaptic Systems GmbH, Germany), mouse anti-Gephyrin (147 111, Synaptic Systems), guinea pig anti-gephyrin (147 318, Synaptic Systems), mouse anti-neuroligin 2 (129 511, Synaptic Systems), mouse anti-GluA1 (MAB2263, Millipore), rabbit anti-$GABA_AR$ α1 (06-868, Millipore Sigma), rabbit anti-$GABA_AR$ γ2 (224 003, Synaptic Systems), mouse anti-SYP (D-4) (sc-17750, Santa Cruz), rabbit anti-FLAG (F3165, Sigma-Aldrich), rabbit and mouse anti-RFP which recognizes DsRed, mCherry and mRuby2 (PM005 and M165-3, Medical & Biological Laboratories, Naka-ku Nagoya, Japan); mouse anti-α-tubulin (T9026, Sigma-Aldrich), mouse anti-PSD95 (610495, BD Biosciences, San Diego, CA) and mouse anti-GST (M071-3, MBL) for western blotting (WB) and rabbit anti-VGAT (131 004, Synaptic Systems), mouse anti-PSD95 (75-028, NeuroMab, Davis, CA), guinea pig anti-Ankyrin G (386 004, Synaptic Systems), guinea pig anti $GABA_AR$ γ2 (224 004, Synaptic

Systems) and mouse anti-Bassoon (sc-58509, Santa Cruz) for immunofluorescence staining; Rabbit anti-p140Cap was described previously (Yang et al., 2015). Alexa Fluor dye-conjugated secondary antibodies for immunofluorescence staining were from Molecular Probes (Invitrogen, Carlsbad, CA, USA).

Pentylenetetrazole (PTZ)-kindling and Seizure Behavior Scoring

To evaluate seizure susceptibility, 8-10-week-old male and female *EndoA1*$^{+/+}$ or *EndoA1*$^{-/-}$ littermates or *EndoA1*$^{fl/fl}$ littermates were intraperitoneally administered with PTZ (25 mg/kg; K0250, Sigma-Aldrich) or saline (control) for 8-12 days, and the resulting seizure behaviors were video-recorded for the next 30 min. Seizure susceptibility was measured blindly by rating seizures on a scale of 0 to 7 as follows (Van Erum et al., 2019): no abnormal behavior (0), reduced motility and prostate position (1), facial jerking (Schulz et al.), neck ierks (Huo et al.), clonic seizure (sitting) (4), clonic, tonic-clonic seizure (lving on belly) (5), clonic, tonic-clonic seizure (lving on side) and wild jumping (6), tonic extension, possibly leading to respiratory arrest and death (7).

Stereotaxic injection

For virus injection, 8-9-week-old naive male and female littermates were anesthetized and stereotactically injected with viral particles in the hippocampal CA1 region as described (Yang et al., 2018). Briefly, mice were weighed and anesthetized with isoflurane (2% 2,2,2-tribromoethanol, Sigma), and secured in a stereotaxic apparatus. The scalp of mouse was cut along the midline between the ears after sterilized with iodophors and 75% (vol/vol) alcohol. Holes for inserting needle were drilled in the bilateral skull. Viral particles carrying pAOV-CAMKIIα-MCS-mCherry-3FLAG or pAOV-CAMKIIα-mCherry-2A-Cre and pAAV-CaMKIIα-EGFP-2A-MCS-3FLAG or pAAV-CaMKIIα-EGFP-2A-MCS-3FLAG-EndoA1 WT or mutants (1.2 μL, 2.0× 10^{12} viral genomes/mL) (OBiO Technology Corp., Ltd) were injected bilaterally into the hippocampal CA1 regions using the following coordinates relative to bregma: 2.0 mm

553 posterior, 1.8 mm lateral, and 1.4 mm ventral at a rate of 0.125 μL/min using a
554 microinjection system (World Precision Instruments). The needle was kept in place for
555 2 min before withdrawal, and the skin was sutured with surgical line. The virus-injected
556 mice were tested for behavior two weeks later.
557
558 **Immunofluorescence staining of brain sections**
559 Male and female littermates (P21 or P60) were anesthetized and immediately perfused,
560 brains were dissected out, fixed overnight in 4% (vol/vol) paraformaldehyde, incubated
561 overnight with 30% sucrose in phosphate-buffered saline (PBS), and then sliced into 30
562 μm-thick coronal sections using a cryotome (Model CM-1950; Leica Biosystems). For
563 immunostaining of brain sections, floating 30 μm-thick slices were rinsed with PBS and
564 permeabilized in 0.4% Triton X-100 in 0.01M PBS for 30 min. Cyrosections were
565 blocked with 2% BSA in PBS containing 0.4% Triton X-100 for 2 h at room temperature
566 (RT), then incubated with primary antibodies overnight at 4 °C. Secondary antibodies
567 conjugated with Alexa Fluor 488 or 555 were used for detection. Sections were then
568 incubated with DAPI (Roche, Grenzach-Wyhlen, Germany) for nuclear staining for 3
569 min at RT. Following rinsing, cyrosections were mounted on gelatin-coated slides and
570 covered with coverslip with mounting medium. Confocal images were collected using
571 the Spectral Imaging Confocal Microscope Digital Eclipse A1Si (Nikon, Tokyo, Japan)
572 with a 40× Plan Fluo (NA 1.30) oil objective (Yang et al., 2018).
573
574 **Cell culture, transfection and drug treatment**
575 Primary hippocampal neurons were cultured as previously described (Yang et al., 2021).
576 Briefly, Hippocampi of female or male pups (P0) were rapidly dissected under sterile
577 conditions, kept in cold DMEM (4°C) with high glucose. Tissue was then digested with
578 0.25% trypsin solution at 37°C for 18-20 min, triturated in DMEM supplemented with
579 10% F12 (11765054, Gibco) and 10% FBS (Gibco, Carlsbad, CA, USA). Then the
580 tissue was mechanically dissociated into single cells by gentle pipetting up and down.
581 Neurons were plated at a density of ~3.0-3.5× 10^5 cells/well in poly-D-lysine (P6407,

582 Sigma-Aldrich) -coated 6-well plate and at a density of 2.5–3.0×10^4 cells/well in 24-
583 well plate. Four hours later, media was changed to Neurobasal A (NB-A) growth
584 medium supplemented with 2% B27 supplement (17504-044, Gibco), 1% GlutaMAX
585 (35050-061, Gibco) and 33 mM glucose. Culture media were changed by half volume
586 with Neurobasal maintenance media once a week. The cultures were maintained at
587 37°C with 5% CO_2. Experiments were performed at DIV 14-16 unless otherwise stated.
588 For neuronal transfection, hippocampal neurons (DIV11) were transfected using
589 Lipofectamine-LTX (15338100, Invitrogen, ThermoFisher Scientific). For 12 mm
590 coverslips in individual wells of a 24 well plate, 0.5 μg of DNA was mixed with 0.5 μL
591 PLUS reagent in 50 μL of NB-A and incubated for 5 min at RT, then gently mixed with
592 1 μL Lipofectamine LTX in 50 μL NB-A and incubated for 20 min at RT. The DNA-
593 Lipofectamine LTX mixture was then added to the neurons in NB-A at 37°C in 5% CO_2
594 for 1 h. Neurons were then rinsed with NB-A and incubated in the original medium at
595 37°C in 5% CO_2 for 3-4 days.
596 For HEK293T cell culture, DMEM supplemented with 10% FBS were used. Cell
597 transfections were performed using Lipofectamine-2000 (11668019, Invitrogen,
598 ThermoFisher Scientific) according to the manufacturer's instructions. Cells were
599 harvested 24-48 h post transfection.
600 For inhibitor treatment, primary neurons cultured on coverslips were pre-incubated
601 with Latrunculin A (1 μM, 76343-93-6, Sigma-Aldrich), Nocodazole (1 μM, 31430-18-
602 9, Sigma-Aldrich) for 60 min or 120 min.
603
604 **Immunofluorescence staining of cultured neurons**
605 Hippocampal neurons were fixed in 4% paraformaldehyde/4% sucrose in PBS at RT
606 for 15 min, then permeabilized in 0.4% Triton X-100 in PBS for 10 min. After blocking
607 with 2% BSA in PBS containing 0.4% Triton X-100 for 30 min at RT, neurons were
608 incubated with primary antibodies for 1 h at RT or overnight at 4°C, and appropriate
609 secondary antibodies conjugated with Alexa Fluor 488, Alexa Fluor 555, or Alexa Fluor
610 647 were used for detection. The primary antibody was diluted in a PBS-2% BSA

solution and incubated overnight at 4°C. After washing with PBS three times, samples were incubated with Alexa Fluor secondary antibodies (1:2000, Thermo Fisher) for 1 h at RT, and mounted without bubbles.

Biotinylation of cell surface proteins

Surface biotinylation experiments were performed as previously described (Guo et al., 2022). Briefly, Neurons were washed with ice cold extracellular solution (ECS) (in mM: 125 NaCl, 2 $CaCl_2$, 2.5 KCl, 5 HEPES, 33 glucose, pH, 8.0) and incubated for 30 min with 0.8 mg/ml NHS-SS-Biotin (21327, Thermo Fisher Scientific) in ECS buffer at 37°C. After incubation, cells were washed once with PBS and the unbound biotin was quenched via two 8 min incubations with quenching buffer (20 mM glycine in PBS). Lysis was performed by mechanical scraping in ice-cold lysis buffer (50 mM Tris-Cl pH 7.4, 150 mM NaCl, 1% TritonX-100, 0.1% SDS). After rotating for 30 min at 4°C, lysates were centrifuged for 10 min at 14,000× *g* at 4°C. Balanced Pierce Streptavidin Agarose beads (25 μL, 20353, Thermo Fisher Scientific) were added to the supernatant. The mixture of lysates and beads were rotated at 4°C overnight and then washed twice with ice-cold lysis buffer. Beads were incubated with 2× SDS sample buffer for 10 min at 37°C and biotinylated proteins were analyzed by immunoblotting.

Protein expression and purification

GST-tagged EndoA1 or gephyrin fragments was expressed in *E. coli* BL21 (DE3). Cells were grown at 37°C in LB (g/L: tryptone 10, yeast extract 5, NaCl 10) supplemented with ampicillin. Cells were induced at OD600 of ~0.6 with 0.4 mM isopropyl-D-1-thiogalactopyranoside (IPTG, A600168-0005, Bio Basic Inc) for 4 h at 30°C or 16 h at 16°C. Cell pellets were resuspended in PBS supplemented with 0.2% Triton X-100 and 0.1 mM PMSF. Proteins were purified with Glutathione Sepharose 4B (GE17-0756-01, Sigma-Aldrich, St. Louis, MO, USA) according to the manufacturer's instructions.

GST-pull down, co-immunoprecipitation (IP) and immunoisolation

For GST-pull down assays, GST-fused protein EndoA1 and its fragments were expressed and purified from *E. coli*. Myc-tagged GPN constructs were expressed in HEK293 cells for 48 h and lysates were prepared in a buffer (in mM: Tris-HCl 50, pH 7.4, NaCl 150, 1% NP-40, EDTA 2) plus protease inhibitors cocktail. Cell lysates were centrifuged at 12,000× *g* for 15 min at 4°C and the supernatant was incubated with individual GST-fused proteins conjugated with Glutathione-Sepharose beads for overnight at 4°C. Beads were washed five times with lysis buffer and boiled in 2× SDS sample buffer. Bound proteins were analyzed by immunoblotting.

For co-IP from brain lysates, the whole brain from 8-10-week-old WT and KO littermates were dissected and homogenized in the pre-chilled lysis buffer (in mM: Tris-HCl 20, pH 7.4, NaCl 150, 1% NP-40, EDTA 5) plus protease inhibitor cocktail, followed by incubation on ice for 15 min. The lysates were then centrifuged at 12,000× *g* for 15 min and the supernatant was collected and incubated with anti-EndoA1 antibody (2 μg) or control IgG overnight at 4°C, followed by incubation with 20 μL Dynabeads Protein A (ThermoFisher, 10007D for rabbit antibody) for 2 h at 4°C. After washing the beads with the same lysis buffer four times, the bound proteins were incubated 2× SDS sample buffer at 37°C for 10 min and subjected to SDS-PAGE and Western blot with indicated antibodies.

For co-IP of proteins in HEK293T cells (Cat. No. 632180, Takara), cells were lysed with lysis buffer (in mM: 0.5% NP-40, Tris-HCl 50, pH 7.4, NaCl 150, EDTA 5) supplemented with protease inhibitors. Lysates were then centrifuged at 16,000× *g* for 15 min at 4°C. Cell lysates were incubated with anti-Flag Affinity Gel (A2220, Sigma-Aldrich) overnight at 4°C on a roller mixer. Immunoprecipitates were washed four times in lysis buffer and boiled in 2× SDS sample buffer, then subjected to SDS-PAGE and immunoblotting.

For immunoisolation of membrane proteins, hippocampi were homogenized with lysis buffer (in mM: Tris-HCl 20, pH 7.4, HEPES 10, pH 7.4, NaCl 150, sucrose 250, EDTA 0.5, pH 8.0) supplemented with protease inhibitors and centrifuged at 800× *g* for 15 min. The supernatants were collected and subjected to high-speed centrifugation at

669 100,000× *g* for 1 h (TLS-55 rotor, OptimaTMMAX Ultracentrifuge; Beckman Coulter,
670 USA). The supernatants (S100) and pellets (P100, the membrane fraction) resuspended
671 in lysis buffer were subjected to immunoisolation with Dynabeads Protein A (100-4D,
672 Invitrogen, Carlsbad, CA, USA) coupled with 2 μg of rabbit anti-EndoA1 or rabbit anti-
673 $GABA_AR$ α1 antibody. Bound proteins were eluted by incubation with 2× SDS loading
674 buffer at 37°C for 10 min and subjected to SDS-PAGE and immunoblotting.
675
676 **PSD fractionation of brain lysates**
677 PSD fractions from adult mouse brain were prepared as previously described (Yang et
678 al., 2015). In brief, hippocampi were homogenized on ice with 20 strokes of a Teflon-
679 glass homogenizer in 1 mL of HEPES-buffered sucrose (0.32 M sucrose, 4 mM HEPES,
680 pH 7.4) containing freshly added protease inhibitors, then centrifuged at 800× *g* at 4°C
681 to remove the pelleted nuclear fraction (P1). Supernatant (S1) was centrifuged at
682 10,000× *g* for 15 min to yield the crude synaptosomal pellet (P2) and the pellet was
683 washed once in 1 mL HEPES-buffered sucrose. P2 was lysed by hypoosmotic shock in
684 900 μL ice-cold 4 mM HEPES, pH 7.4 plus protease inhibitors, homogenized by
685 pipetting and rotating for 30 min at 4 °C. The lysate was centrifuged at 25,000× *g* for
686 20 min to yield supernatant (S3, crude synaptic vesicle fraction) and pellet (P3, lysed
687 synaptosomal membrane fraction). To prepare the PSD fraction, P3 was resuspended in
688 900 μL of ice-cold 50 mM HEPES, pH 7.4, 2 mM EDTA, plus protease inhibitors and
689 0.5% Triton X-100, rotated for 15 min at 4 °C and centrifuged at 32,000× *g* for 20 min
690 to obtain the PSD pellet. PSD pellets were resuspended in 50 μL ice-cold 50 mM
691 HEPES, pH 7.4, 2 mM EDTA plus protease inhibitors. Proteins were subjected to SDS-
692 PAGE and immunoblotting.
693
694 **Protein identification with mass spectrometry and pathway analysis**
695 For identification of EndoA1-interacting proteins, mouse brain was homogenized with
696 lysis buffer (in mM: Tris-HCl 20, pH 7.4, HEPES 10, pH 7.4, NaCl 150, EDTA 1, 1%
697 NP-40) supplemented with protease inhibitors. Lysates were centrifuged at 12,000× *g*

698 for 15 min. The supernatants were incubated with rabbit anti-EndoA1 antibody
699 overnight at 4°C. Beads were rinsed with lysis buffer for five times and bound proteins
700 were resolved by SDS-PAGE. Proteins in the gels were reduced with 10 mM DTT at
701 37°C for 1 h, then alkylated with 25 mM iodoacetamide at RT for 1 h in the dark before
702 digestion with trypsin (T1426, Sigma-Aldrich; enzyme-to-substrate ratio 1:50) in 25
703 mM ammonium bicarbonate at 37°C overnight. Tryptic peptides were extracted from
704 gel by sonication with a buffer containing 5% trifluoroacetic acid and 50% acetonitrile.
705 The liquid was dried by SpeedVac, and peptides were resolubilized in 0.1% formic acid
706 and filtered with 0.45 μm centrifugal filters before analysis with a TripleTOF 5600 mass
707 spectrometer (AB SCIEX, Canada) coupled to an Eksigent nanoLC. Proteins were
708 identified by searching the MS/MS spectra against the *Mus musculus* SwissProt
709 database using the ProteinPilot 4.2 software. Carbamidomethylation of cysteine was set
710 as the fixed modification. Trypsin was specified as the proteolytic enzyme with a
711 maximum of 2 miss cleavages. Mass tolerance was set to 0.05 Da and the false
712 discovery rates for both proteins and peptides were set at 1%. For enrichment pathway
713 analysis: Only proteins with intensity to IgG ratio of > 1.5 were considered in the
714 analysis. These genes were analyzed by Gene Ontology (GO) Enrichment Analysis
715 (http://bioinfo.org/kobas).
716

717 **Electrophysiology**

718 To knockout *EndoA1* in the brain, *EndoA1*$^{fl/fl}$ mice were injected with high-titer AAV
719 stock carrying pAOV-CaMKIIα-mCherry-2A-Cre (AAV-mCherry-2A-Cre) (1~5 × 10^{13}
720 IU/ml) in the hippocampal CA1 region within 24 h after birth. Newborn *EndoA1*$^{fl/fl}$
721 littermates (male or female) were anesthetized on ice for 4-5 min and then mounted in
722 a custom ceramic mold to align the head along the X and Y axes, with lambda set as (X,
723 Y) = (0, 0). The zero point of the Z axis was the position at which the injecting needle
724 penetrated the skin. Approximately 10 nL of the viral solution was injected at each of
725 the seven sites (coordinates in mm, [X, Y, Z] = [1.2, 1.2, 1.4/1.0/0.6] and [1.5, 1.0,
726 1.7/1.3/0.9/0.5]) targeting the hippocampus in each cerebral hemisphere using a

microsyringe (Sutter Instrument, Novato, CA, USA) and beveled glass injection pipette (Yang et al., 2018). For the rescue experiment, P0 *EndoA1*$^{fl/fl}$ mice were injected with a mixture of two high-titer AAV stocks: AAV-EGFP and AAV-mCherry or AAV-mCherry-2A-Cre; AAV-mCherry-2A-Cre and AAV-EGFP-2A-FLAG-EndoA1, AAV-EGFP-2A-FLAG-EndoA1 Y343A or AAV-EGFP-2A-FLAG-EndoA1 KKK-EEE. Approximately 20 nL of the viral solution was injected at each of the seven sites. The injected pups were returned to the home cage and used for recording 2 weeks later.

To prepare acute brain slices for electrophysiological analyses, mice were anesthetized with tribromoethanol (330-390 mg/kg) (T48402, Sigma-Aldrich, St. Louis, MO, USA) and decapitated. Whole brains were removed promptly and placed in ice-cold oxygenated (95% O_2/5% CO_2) low-Ca^{2+}/high-Mg^{2+} high-sucrose cutting solution (in mM: KCl 2.5, $NaHCO_3$ 25, NaH_2PO_4 1.25, glucose 10, sucrose 210, Na-Ascorbate 1.3, $CaCl_2$ 0.5, $MgSO_4$ 7). Coronal 300 μm slices were obtained using a vibrating tissue slicer (Leica VT1200S; Leica Biosystems, Wetzlar, Deutschland). Slices were immediately transferred to standard artificial cerebrospinal fluid (ACSF) (in mM: NaCl 119, KCl 2.5, $MgSO_4$ 1.3, $CaCl_2$ 2.5, glucose 11, $NaHCO_3$ 26.2, NaH_2PO_4 1, pH 7.3-7.4) at 33°C and continuously bubbled with 95% O_2/5% CO_2. After 30 min of incubation, slices were transferred to a recording chamber with the same extracellular buffer at room temperature (RT, ~25°C).

For whole-cell patch clamp recordings, brain slices were continuously perfused with standard ACSF at RT under visual control with differential interference contrast illumination on an upright microscope (BX51WI, Olympus, Tokyo, Japan) with a 40× water-immersion objective and a digital camera. The mCherry- and/or EGFP-positive hippocampal CA1 pyramidal neurons were identified by an epifluorescence microscope (pE-300, CoolLED Ltd, Andover, UK). Whole-cell recordings were performed on an AAV-infected neuron and a neighboring uninfected neuron. Patch clamp data were obtained using a Multiclamp 700B amplifier (Molecular Devices, San Jose, CA, USA), and signals were low-pass filtered at 2 kHz and digitized at 10 kHz using an Axon Digidata 1550B analog-to-digital board (Molecular Devices, San Jose, CA, USA).

To analyze evoked excitatory postsynaptic currents (eEPSCs), evoked inhibitory postsynaptic currents (eIPSCs), and paired-pause ratio, cells were recorded with 3-7 MΩ borosilicate glass patch pipettes filled with the intracellular solution (in mM: $CsMeSO_3$ 135, HEPES 10, EGTA 1, QX-314 3.3, MgATP 4, NaGTP 0.3, and Na_2-phosphocreatine 8, pH 7.3) (Horn and Nicoll, 2018), following stimulation of Schaffer collaterals with a bipolar stimulation electrode placed in the stratum radiatum of the CA1 region. EPSCs were measured at a holding potential of -70 mV, and IPSCs were recorded at +10 mV. Paired-pulse ratios of EPSCs were measured at a holding potential of -70 mV by applying two electrical stimulations at a 50 ms interval and calculating the ratio of the peak amplitudes. Paired-pulse ratios of IPSCs were measured at a holding potential of +10 mV by applying two electrical stimulations at a 200 ms interval and calculating the ratio of the peak amplitudes. E/I ratios were measured by applying the same stimulation current and then calculating the ratio of the absolute values of the peak amplitudes of EPSCs and IPSCs. The miniature EPSCs and IPSCs were measured in the ACSF bath solution containing 1 μM tetrodotoxin (TTX, MSS0002, BGB Analytik AG, BL, Switzerland) at -70 mV and +10 mV, respectively, in the voltage-clamp mode. Data were collected and analyzed using the pClamp 10.6 software (Molecular Devices, San Jose, CA, USA).

All electrophysiological recordings were carried out blindly and all n numbers in the data figures indicate the number of recorded neurons. All chemicals were purchased from Sigma-Aldrich except otherwise noted.

Confocal and three-dimensional structured illumination microscopy (3D-SIM) image acquisition and analysis

Confocal images were collected using the Spectral Imaging Confocal Microscope Digital Eclipse A1Si (Nikon, Tokyo, Japan) with a 100× Plan Apochromat VC (NA 1.40) oil objective (4× optical zoom). Images were z projections of images taken at 0.25-μm step intervals. The number of planes, typically 5-7, was chosen to cover the entire dendrite from top to bottom. Images were captured using a 1024×1024 pixels screen

785 for neuronal cells. Laser power, digital gain, and offset settings were identical in each
786 experiment. Images were processed with the NIH ImageJ software and exported into
787 Adobe Photoshop. Only brightness and contrast were adjusted for the whole frame, and
788 no part of a frame was enhanced or modified in any way. Control and experimental
789 group neurons which were to be directly compared were imaged with the same
790 acquisition parameters.

791 For 3D-SIM, secondary antibodies conjugated with Alexa Fluor 568 were used
792 and images were captured as described by Niu et al (Niu et al., 2017) on the DeltaVision
793 OMX V4 imaging system (Applied Precision Inc, USA) with a 100× 1.4 oil objective
794 (Olympus UPlanSApo), solid state multimode lasers (488, 593 and 642 nm) and
795 electron-multiplying charge-coupled device (CCD) cameras (Evolve 512× 512,
796 Photometrics, USA). Serial Z-stack sectioning was done at 125 nm intervals. The
797 microscope is routinely calibrated with 100 nm fluorescent spheres to calculate both
798 the lateral and axial limits of image resolution. SIM image stacks were reconstructed
799 using softWoRx 5.0 (Applied Precision) with the following settings: pixel size 39.5 nm;
800 channel-specific optical transfer functions; Wiener filter 0.001000; discard Negative
801 Intensities background; drift correction with respect to first angle; custom K0 guess
802 angles for camera positions. Pixel registration was corrected to be less than 1 pixel for
803 all channels using 100 nm Tetraspeck beads. Three dimensional images were
804 reconstructed from z stacks. For clarity of display, small linear changes of brightness
805 and contrast were performed on three-dimensional reconstructions.

806 Quantification of immunostaining were carried out blindly. For quantitative
807 analysis of immunofluorescent signals in neurons, maximal projection images were
808 created with the ImageJ software (https://imagej.net.) from 6-7 serial optical sections.
809 Background was subtracted by using the "subtract background" function and the
810 background level was held identical for all cells within each experiment. To examine
811 the number and fluorescence intensity of signal puncta in soma or dendrites, the EGFP-
812 or DsRed-labeled soma or dendrites were outlined automatically by using the "Wand
813 Tool" function. The region of interest (ROI) in cultured neurons was defined along a

segment of the dendrite according to the fluorescence signal distinguished from the background. Average values of fluorescence intensities in ROI (the total fluorescent intensity divided by the total area of a dendritic segment) were calculated by Analyze > measure. Gephyrin clusters were thresholded and confirmed visually to select appropriate clusters following a minimal size cut-off by using the "Analyze Particles" plugin of the ImageJ software. For colocalization quantifications, NIS-Elements Software was used as follows: region of interest (ROI) of dendritic segments was manually chosen, and thresholded using the "Threshold" function to remove background signals. Pearson's correlation coefficient and Manders overlap coefficient were analyzed by using the "Analyze Control > colocalization" function.

For spatial correlation/colocalization analysis in super-resolution images, we using the method described in the reference (McCall, 2024). Briefly, channels in 3D images were split and followed by Gaussian blur performed on both GPN and Bassoon channels. Next masks of all possible localizations for the signal in GPN and Bassoon image channel were obtained. Then, the input images of the three channels were deconvolved using 10 iterations of the Richardson-Lucy algorithm in Deconvolution Lab 2 (version 2.1.2), with theoretical point spread functions (PSFs) generated by the PSF Generator plugin. PSFs were generated using the Born & Wolf model, with parameters matching the acquisition parameters: 1.518 immersion refractive index, 1.4 NA, 40 nm pixel size and 125 nm z-step, 512 × 512 × 14 output diameters, and 520 nm, 592 nm and 670 nm wavelengths. Finaly, select the two images (GPN and EndoA1 or Bassoon and EndoA1) and the mask images, then perform a cross-correlation analysis (CCC) between the two images using the CCC plugin (https://github.com/andmccall/Colocalization_by_Cross_Correlation).

Statistical analysis

For all biochemical, cell biological and electrophysiological recordings, at least three independent experiments were performed (independent cultures, transfections or different mice). Statistical analysis was performed in GraphPad Prism 9.0 (GraphPad

Software, La Jolla, CA). Data sets were tested for normality and direct comparisons between two groups were made using two-tailed Student's t test (t test, for normally distributed data) as indicated. To evaluate statistical significance of three or more groups of samples, one-way ANOVA analysis with a Tukey test was used or repeated measures ANOVA analysis with a Tukey test was used in behavior assays. Statistical parameters are reported in the figures and the corresponding legends. The statistical significance was defined as * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, respectively. All data are presented as scatterplots with mean ± S.E.M.

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Figure legends

Figure 1. Ablation of endophilin A1 in CA1 neurons disrupts the E/I balance and increases epilepsy susceptibility.

(A) Experimental protocol of PTZ kindling. Saline serves as negative control for PTZ.

(B) Representative images of convulsive behavior in each group.

(C) Seizures were monitored and scored after each injection. Nine 10-week-old mice were used in each condition. Seizure scores are shown as means ± S.E.M. Statistical test: repeated measures ANOVA followed by Tukey post-hoc test at each time point. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, *EndoA1*$^{-/-}$_PTZ *vs EndoA1*$^{+/+}$_PTZ at each time point.

(D) The representative mCherry-positive hippocampal CA1 neuron used for electrophysiological recording.

(E-F) Representative traces of evoked excitatory postsynaptic currents (eEPSC) and quantification of mean eEPSC amplitude (E), and paired-pulse recording (F) from 10 control (Ctrl, uninfected) and 10 *EndoA1* KO (mCherry and Cre) pyramidal neurons of 4 independent animals.

(G-H) Representative traces of evoked inhibitory postsynaptic currents (eIPSCs) and quantification of mean eIPSC amplitude (G), and paired-pulse recording (H) from 9 control and 9 *EndoA1* KO pyramidal neurons of 4 independent animals.

(I) Effect of *EndoA1* KO on the E/I ratio from 9 control and 9 *EndoA1* KO pyramidal neurons of 4 independent animals.

(J) Representative recordings of miniature excitatory postsynaptic currents (mEPSCs) traces (+10 mV) from control or *EndoA1* KO CA1 pyramidal cells.

(K) Cumulative probability and quantification of mEPSC amplitude from 7 control and 7 *EndoA1* KO pyramidal neurons of 3 independent animals.

(L) Cumulative probability and quantification of mEPSC frequency from 7 control and 7 *EndoA1* KO pyramidal neurons of 3 independent animals.

(M) Representative recordings of miniature inhibitory postsynaptic currents (mIPSCs) traces (–70 mV) from control or *EndoA1* KO CA1 pyramidal cells.

(N) Cumulative probability and quantification of mIPSC amplitude from 7 control and 7 *EndoA1* KO pyramidal neurons of 3 independent animals.

(O) Cumulative probability and quantification of mIPSC frequency from 7 control and 7 *EndoA1* KO pyramidal neurons of 3 independent animals. Statistical test: two-tailed Student's t-test in E-I & K, L, N, O. ns, non-significant. All data shown are scatterplots with means ± S.E.M.

Figure 2. *EndoA1* KO causes defects in the formation and stabilization of inhibitory synapses.

(A-B) Immunofluorescence staining and confocal microscopy of gephyrin (GPN) in the hippocampal region of 3-week-old (P21) and 10-week-old (P60) *EndoA1*$^{+/+}$ or *EndoA1*$^{-/-}$ mouse brains. Scale bar, 50 μm.

(C) Quantification of the signal intensity of GPN in (A) and (B). n = 6 mice for each genotype.

(D-E) Same as (A-B) except that slices were immunostained for VGAT, (D) for P21, (E) for P60. Scale bar, 50 μm.

(F) Quantification of VGAT signal intensity in (D) and (E). n = 6 mice for each genotype.

(G-H) Same as (A-B) except that slices were immunostained for $GABA_AR$ γ2 (γ2). (G) for P21, (H) for P60. Scale bar, 50 μm.

(I) Quantification of γ2 signal intensity in (G) and (H). n = 6 mice for each genotype. Statistical test: unpaired two-tailed Student's t-test. ns, non-significant. All data are scatterplots with means ± S.E.M. Abbreviations: CA1-3, cornus ammonis region 1-3; DG, dentate gyrus; DAPI staining of cell nuclei is shown in blue.

Figure 3. Endophilin A1 is distributed in the iPSD and regulates the assembly of the inhibitory postsynaptic machinery.

(A) Cultured *EndoA1*$^{fl/fl}$ hippocampal neurons were transfected with construct expressing GFP or GFP-2A-Cre on DIV12, fixed and immunostained for surface GluA1 (sGluA1) or $GABA_AR$ γ2 (sγ2) on DIV16. Shown are representative confocal microscopy images. Lower panels are magnification of boxed regions in the upper panels. Scale bar, 10 μm in the upper panels and 5 μm in the lower panels.

(B) Quantification of the signal intensity (left) and cluster number (right) of GPN puncta in dendrites in (A). GFP: 15 neurons; Cre: 18 neurons, from 3 independent cultures. Statistical test: unpaired two-tailed Student's t-test. ns, non-significant.

(C-D) Cultured *EndoA1*$^{fl/fl}$ hippocampal neurons were co-transfected with constructs for GFP or GFP-2A-Cre and Flag vector or Flag-endophilin A1 (Flag-EndoA1) on DIV12, and immunostained for surface γ2 on DIV16. Shown are representative confocal images (C) and quantification of surface γ2 in dendrites (D). Scale bar, 2 μm. GFP: 18 neurons; Cre: 19 neurons; Cre + EndoA1: 18 neurons, from 3 independent cultures. Statistical test: one-way ANOVA by Tukey post hoc test.

(E-F) *EndoA1*$^{fl/fl}$ hippocampal neurons were infected with adeno-associated virus (AAV) carrying mCherry or mCherry-2A-Cre on DIV3 and surface biotinylated on DIV16. Total and surface proteins were detected by SDS-PAGE and immunoblotting. N = 3. Statistical test: unpaired two-tailed t-test.

(G) Hippocampal neurons were transfected with construct expressing GFP to label neuronal morphology on DIV12, then immunostained with antibodies against EndoA1 and GPN on DIV16. Shown are representative z stack maximum projection images captured by structured illumination microscopy (SIM). Lower panels are insets of boxed regions in the upper panel.

Dendrites and spines are outlined in white. Scale bars, 2 μm in upper panels and 0.5 μm in magnified images.

(H) Same as (G) except that neurons were immunostained for EndoA1 and surface γ2.

(I) Same as (G) except that neurons were immunostained for EndoA1 and VGAT.

(J) Immunoisolation from the membrane fraction of mouse brain tissues with antibodies against $GABA_AR$ α1 (α1) or EndoA1.

(K) Immunoisolation from *EndoA1*$^{+/+}$ and *EndoA1*$^{-/-}$ brain lysates with antibodies to $GABA_AR$ α1. Shown are representative immunoblot (left) and quantification of neuroligin 2 (NL2), GPN and $GABA_AR$ γ2 co-immunoisolated with α1 (right). N = 6 independent experiments. Statistical test: unpaired two-tailed Student's t-test.

(L) PSD fractions were isolated by differential centrifugation from mouse brain lysates and analyzed by immunoblotting with antibodies to EndoA1 and synaptic proteins. S1, homogenates; PSD, postsynaptic density. SYP, synaptophysin. N = 7 independent experiments. Statistical test: unpaired two-tailed Student's t-test. All data represent scatter plots with means ± S.E.M.

Figure 4. Endophilin A1 interacts with gephyrin and promotes its clustering in postsynaptic neurons.

(A) Silver staining of proteins co-immunoprecipitated from *EndoA1*$^{+/+}$ mouse brain lysates with antibodies to endophilin A1.

(B) Gene Ontology (GO) enrichment analysis of EndoA1-interacting proteins identified by mass spectrometry.

(C-D) HEK293T cells co-transfected with constructs expressing Myc-tagged GPN or HA-tagged NL2 and Flag-tagged EndoA1 for 48 h were lysed for Flag IP. Input and bound proteins were analyzed by immunoblotting with antibodies against Myc, HA and Flag.

(E) Co-IP from brain lysates with antibodies to EndoA1.

(F) HEK293T cells overexpressing Flag-tagged endophilin As and Myc-tagged GPN were lysed for co-immunoprecipitation.

(G) *EndoA1*$^{fl/fl}$ hippocampal neurons were co-transfected with constructs expressing GFP and Flag vector, or GFP-2A-Cre and Flag vector, Flag-EndoA1, Flag-EndoA2 or Flag-EndoA3 on DIV12, fixed on DIV16 and immunostained for GPN, GFP and Flag. Shown are representative confocal microscopy images. Scale bar, 2 μm.

(**H**) Quantification of GPN signal intensity (left) and cluster number (right) in dendrites in (G). 36 neurons for GFP + vector; 34 neurons for Cre + vector; 38 neurons for Cre + EndoA1; 29 neurons for Cre + EndoA2; 27 neurons for Cre + EndoA3, from 3 independent cultures. Statistical test: one-way ANOVA by Tukey post hoc test.

(I) Hippocampal neurons were co-transfected with constructs expressing GFP and vector or Flag-EndoA1 on DIV12, fixed and immunostained for GPN on DIV16. Shown are

representative confocal microscopy images of GPN in the soma (left) and dendrites (right). Scale bars, 5 μm in the upper panels and 2 μm in the lower panels.

(J) Quantification of GPN signal intensity (left) and cluster number (right) in the soma and dendrites. 34 neurons for GFP + vector; 37 neurons for GFP + EndoA1, from 3 independent cultures. Statistical test: unpaired Student's two-tailed t-test. All data represent scatterplots with means ± S.E.M.

Figure 5. Both BAR and SH3 domains are required for endophilin A1-mediated organization of iPSD.

(A) *EndoA1*$^{fl/fl}$ hippocampal neurons expressing GFP or Cre were immunostained for GPN, surface γ2 and GFP on DIV16. Shown are representative confocal microscopy images. Scale bars, 5 μm in left panels and 1 μm in magnified images.

(B-C) Quantification of the mean intensity of GPN (green) and surface γ2 (red) and their colocalization in (A). 21 neurons for GFP, 22 neurons for Cre, from 3 independent cultures. Statistical test: unpaired two-tailed Student's t-test. All data represent scatterplots with means ± S.E.M.

(D) DIV12 hippocampal neurons were co-transfected with constructs expressing GFP and vector or Flag-EndoA1, fixed on DIV16 and immunostained for surface γ2. Shown are representative confocal microscopy images of surface γ2 in the soma (left) and dendrites (right). Scale bars, 5 μm in the upper panels and 2 μm in the lower panels.

(E) Quantification of surface γ2 mean intensity (left) and cluster number (right) in the soma and dendrites. Soma: 20 neurons for GFP + vector, 20 neurons for GFP + EndoA1; Dendrites: 35 neurons for GFP + vector, 30 neurons for GFP + EndoA1, from 3 independent cultures. Statistical test: unpaired two-tailed Student's t-test. All data represent scatterplots with means ± S.E.M.

(F) Schematic representation of EndoA1 and GPN domain structure and diagrams of fragments used for interaction mapping.

(G) GST-tagged EndoA1 fragments conjugated to Glutathione-Sepharose beads were incubated with lysates of HEK293T cells overexpressing Myc-tagged GPN. Input and bound proteins were analyzed by immunoblotting with antibodies against Myc. Bottom panel: Coomassie blue-stained SDS-PAGE gel showing GST fusion proteins. FL, full length.

(H) HEK293T cells co-transfected with constructs encoding Myc-tagged GPN and Flag-tagged EndoA1 fragments were lysed for Flag IP. Input and bound proteins were analyzed by immunoblotting with antibodies against Myc and Flag.

(I) Binding of His-tagged EndoA1 to GST-tagged GPN fragments in pull-down assay. Bound proteins were analyzed by SDS-PAGE and immunoblotting with anti-His antibodies. Input and bound proteins were analyzed by immunoblotting with antibodies against His. Bottom panel: Coomassie blue-stained SDS-PAGE gel showing GST fusion proteins. FL, full length. Right

panel: Quantification of EndoA1 binding to GPN domain. N = 4 independent experiments. Statistical test: one-way ANOVA by Tukey post hoc test. All data represent scatterplots with means ± S.E.M.

(J) DIV12 *EndoA1*$^{fl/fl}$ hippocampal neurons were co-transfected with constructs expressing GFP and Flag vector or GFP-2A-Cre, and Flag vector or Flag-EndoA1 FL, Flag-EndoA1 ΔSH3, Flag-EndoA1 aa172-352, fixed and immunostained for GPN on DIV16. Shown are representative confocal microscopy images of GPN. Scale bar, 2 μm.

(K) Quantification of mean intensity (left) and cluster number (right) in dendrites are shown for GPN in (J). 41 neurons for GFP + vector; 44 neurons for Cre + vector; 35 neurons for Cre + FL; 24 neurons for Cre + ΔSH3; 27 neurons for Cre + aa172-352, from 3 independent cultures. Statistical test: one-way ANOVA by Tukey post hoc test. All data represent scatterplots with means ± S.E.M.

(L) Same as (I) except that neurons were immunostained for surface γ2. Scale bar, 2 μm.

(M) Quantification of mean intensity (left) and cluster number (right) in dendrites are shown for surface γ2 in (K). 38 neurons for GFP + vector; 31 neurons for Cre + vector; 26 neurons for Cre + FL; 33 neurons for Cre + ΔSH3; 36 neurons for Cre + aa172-352, from 3 independent cultures. Statistical test: one-way ANOVA by Tukey post hoc test. All data represent scatterplots with means ± S.E.M.

Figure 6. Actin polymerization is required for stabilization of inhibitory postsynapses.

(A-D) DIV12 cultured hippocampal neurons transiently expressing GFP were treated with DMSO (vehicle control), 1 μM Latrunculin A (LatA) or 1 μM Nocodazole (Noco) for 60 min (A) or 120 min (C) on DIV16, respectively. Changes in postsynaptic proteins GPN (red) and PSD95 (green) were detected by confocal microscopy. Quantitative data shown were the mean intensity (left) and cluster number (right) of GPN and PSD95 at 60 min (B) and 120 min (D). Ctrl: 20 neurons for 60 min; 29 neurons for 120 min; LatA: 20 neurons for 60 min; 33 neurons for 120 min; Noco: 23 neurons for 60 min; 22 neurons for 120 min, from 3 independent cultures. Statistical test: one-way ANOVA by Tukey post hoc test. Scale bar, 5 μm.

(E-H) Same as (A-D) except that neurons were immunostained for GPN (red) and surface γ2 (green). Changes in GPN and surface γ2 at 60 min (E) or 120 min (G) were detected by confocal microscopy. Quantitative data shown were the mean intensity (left) and cluster number (right) of GPN and surface γ2 at 60 min (F) and 120 min (H). Ctrl: 35 neurons for 60 min; 24 neurons for 120 min; LatA: 34 neurons for 60 min; 20 neurons for 120 min; Noco: 34 neurons for 60 min; 24 neurons for 120 min, from 3 independent cultures. Statistical test: one-way ANOVA by Tukey post hoc test. Scale bar, 5 μm.

(I) Levels of synaptic proteins normalized to control neurons following LatA or Noco treatment in (A,C,E,G).

(J-K) The colocalization between surface γ2 and GPN signals in dendrites in (E) and (G). Statistical test: one-way ANOVA by Tukey post hoc test. All data are scatterplots with means ± S.E.M.

Figure 7. Both the membrane-binding and p140Cap-binding capacities of endophilin A1 are required for iPSD assembly and E/I balance.

(A) Cultured hippocampal neurons from *EndoA1*$^{fl/fl}$ mice were co-transfected with construct expressing GFP and Flag vector or GFP-2A-Cre and Flag vector, Flag-EndoA1 wild-type (WT), mutants (KKK-EEE or Y343A) or p140Cap, and analyzed on DIV16 by immunofluorescence staining for GPN (green) and GFP (red). Scale bar, 2 μm.

(B) Quantification of GPN mean intensity (left) and cluster number (right) in (A). 35 neurons for GFP vector; 43 neurons for Cre + vector; 32 neurons for Cre + WT; 33 neurons for Cre + KKK-EEE; 36 neurons for Cre + Y343A; 33 neurons for Cre + p140Cap, from 3 independent cultures. Statistical test: one-way ANOVA by Tukey post hoc test.

(C) Same as (A) except that neurons were immunostained for surface γ2 on DIV16. Scale bar, 2 μm.

(D) Quantification of surface γ2 mean intensity (left) and number (right) in (C). 44 neurons for GFP + vector; 42 neurons for Cre + vector; 32 neurons for Cre + WT; 35 neurons for Cre + KKK-EEE; 35 neurons for Cre + Y343A; 32 neurons for Cre + p140Cap, from 3 independent cultures. Statistical test: one-way ANOVA by Tukey post hoc test.

(E) Experimental scheme for seizure scoring of *EndoA1*$^{fl/fl}$ mice stereotactically co-injected with AAVs carrying mCherry and Cre, and EGFP and EndoA1 or its mutants (KKK-EEE and Y343A) in the hippocampal CA1 region. Mice were intraperitoneally administered PTZ for 12 days after AAV injections, then seizures were scored.

(F) PTZ-induced seizures in mice injected with the indicated AAVs were scored for 30 min. Ctrl, 7 mice; Cre, 7 mice; Cre + WT, 7 mice; Cre + KKK-EEE, 6 mice; Cre + Y343A, 7 mice.; Statistical test: repeated measures ANOVA followed by Tukey post-hoc test at each time point. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, red color is for Cre *vs* Ctrl, green color is for Cre + WT *vs* Cre. All data are means ± S.E.M.

(G-K) AAVs expressing Cre (AAV-mCherry-2A-Cre) and GFP (AAV-EGFP) or Cre and EndoA1 WT or mutant (AAV-EGFP-2A-EndoA1, AAV-EGFP-2A-EndoA1 KKK-EEE, AAV-EGFP-2A-EndoA1 Y343A) were stereotaxically injected into the CA1 regions of *EndoA1*$^{fl/fl}$ mouse brain at P0. Acute hippocampal slices were prepared on P14-P21 for dual recording of evoked excitatory postsynaptic currents (eEPSC) and inhibitory postsynaptic currents (eIPSC). Shown are representative traces and pairwise comparisons of noninfected (Ctrl) and infected neurons in the same slice. Scale bar = 50/50, 50/50, 50/50, 50/50, 40/20 (pA/ms) in (G); Scale bar = 100/100, 100/400, 100/100, 100/200, 300/100 (pA/ms) in (H); Scale bar = 50/50, 50/50, 50/50, 50/50, 50/50 (pA/ms) in (I); Scale bar = 100/100, 200/100, 100/100, 100/200, 100/200 (pA/ms) in (J). Statistical test: unpaired two-tailed Student's t-test. ns, non-significant; All data are scatterplots with means ± S.E.M. All number of recording were from more than 7 pyramidal neurons of 3 independent animals.

Figure 8. Model for the role of endophilin A1 at the inhibitory postsynapses and E/I balance.

(A) Endophilin A1 directly interacts with the inhibitory postsynaptic scaffold protein gephyrin to promote organization of iPSD and synaptic recruitment/stabilization of the $GABA_A$Rs via its plasma membrane association and actin polymerization-promoting activities. **(B)** Loss of endophilin A1 impairs inhibitory synaptic neurotransmission and disrupts the balance of excitatory/inhibitory activities in the neural circuits, leading to enhanced epileptic seizure susceptibility.

Figure S1. Loss of endophilin A1 *in vivo* does not affect fast mIPSCs (perisomatic events: < 2.8 ms) and slow mIPSCs (distal dendritic events: >2.8 ms). Related to Figure 1

(A) Quantification of somatic and dendritic amplitudes in Ctrl and *EndoA1* KO neuron. n = 7 Ctrl and 9 *EndoA1* KO neurons from 3 independent mice. Data are scatterplots with mean ± S.E.M.

(B) Frequency of somatic and dendritic events from Ctrl and *EndoA1* KO mice. Ctrl: 7 neurons; *EndoA1* KO: 9 neurons from 3 independent mice. Data are scatterplots with mean ± S.E.M.

Figure S2. Effects of *EndoA1* knockout on excitatory and inhibitory synapses in the hippocampus. Related to Figure 2

(A-B) Immunofluorescence analysis of PSD95 clusters in the hippocampal region of 3-week-old (P21) (left) and 10-week-old (P60) (right) $EndoA1^{+/+}$ and $EndoA1^{-/-}$ mouse brains. Scale bar, 50 μm.

(C) Quantification of the mean intensity of PSD95 in the hippocampal region of 3-week-old (P21) and 10-week-old (P60) $EndoA1^{+/+}$ and $EndoA1^{-/-}$ mouse brains. n = 6 mice for each genotype. Statistical test: unpaired two-tailed Student's t-test. Data are scatterplots with mean ± S.E.M.

(D-E) Immunofluorescent signals of VGLUT1 in the hippocampal region of 3-week-old (P21) (left) and 10-week-old (P60) (right) $EndoA1^{+/+}$ and $EndoA1^{-/-}$ mouse brains. Scale bar, 50 μm.

(F) Quantification of the mean intensity of VGLUT1 in the hippocampal region of 3-week-old (P21) and 10-week-old (P60) $EndoA1^{+/+}$ and $EndoA1^{-/-}$ mouse brains. n = 6 mice for each genotype. Statistical test: unpaired two-tailed Student's t-test. Data are scatterplots with mean ± S.E.M.

(G) Immunofluorescent signals of GPN in distinct layers of P60 mouse hippocampus. Scale bar, 50 μm.

(H) Quantification of gephyrin (GPN) mean intensity in distinct layers of hippocampus. n = 6 mice for each genotype. Abbreviations: CA1-3, cornus Ammonis region 1-3; DG, dentate gyrus; slm, stratum lacunosum-moleculare; so, stratum oriens; sp, stratum pyramidal; sr, stratum radiatum. DAPI staining of cell nuclei is shown in blue. Statistical test: two-tailed Student's t-

test. ns, non-significant. Data are scatterplots with mean ± S.E.M.

Figure S3. Changes of endogenous postsynaptic proteins visualized by recombinant probes after depletion of EndoA1. Related to Figure 4

(A) AAVs expressing GFP (AAV-GFP) or GFP and Cre (AAV-GFP-2A-Cre) and mRuby2-GPN.FingR were stereotaxically injected into the CA1 regions of *EndoA1*$^{fl/fl}$ mouse brain at P0. Brain sections from P21 mice were fixed and immunostained for GFP and mRuby. Representative images at 40× magnification with 4× digital zoom showed the expression patterns of mRuby2-GPN.FingR. Scale bars, 10 μm.

(B) Quantification of signal intensity of mRuby2-GPN.FingR in (C). n = 8 mice for each genotype.

(C) AAVs expressing mCherry (AAV-mCherry) or mCherry and Cre (AAV-mCherry-2A-Cre) and PSD95.FingR-eGFP were stereotaxically injected into the CA1 regions of *EndoA1*$^{fl/fl}$ mouse brain at P0. Brain sections from P21 mice were fixed and immunostained for mCherry and GFP. Representative images at 40× magnification with 4× digital zoom showed the expression patterns of PSD95.FingR-eGFP. Scale bars, 10 μm.

(D) Quantification of signal intensity of PSD95.FingR-eGFP in (A). n = 6 mice for each genotype.

Figure S4. Expression of GABA$_A$R α1 in cultured hippocampal neurons and the localization of EndoA1 in inhibitory pre- and postsynaptic sites. Related to Figure 3

(A) Cultured hippocampal neurons were fixed on DIV16 and immunostained for surface GABA$_A$R α1. Shown are representative confocal microscopy images. Scale bar, 10 μm.

(B) Hippocampal neurons were fixed and immunostained with antibodies against EndoA1, Bassoon and GPN on DIV16. Shown are representative z stack maximum projection images captured by structured illumination microscopy (SIM). Lower panels are insets of boxed regions in the upper panel. Lower panels are insets of boxed regions in the upper panel. Scale bars, 5 μm in upper panels and 0.5 μm in magnified images.

(C-D) Color composite of the two input images (left) and composite image of the data that contributed to the Gaussian result (right) for spatial colocalization analysis in super-resolutional images. Scale bars, 5 μm.

(E-F) The blue line is the radial profle of original cross-correlation, the green line is the radial profle after the pixel randomization procedure, and the red dashed line is the fit Gaussian curve. CCC generates four statistical parameters (μ (the mean distance of the measured spatial correlation), σ (standard deviation distance for the spatial correlation), Confidence and R-squared) that was used to evaluate spatial correlation of GPN and EndoA1 in (E), and of Bassoon and EndoA1 in (F).

(G) The ratio of GPN or Bassoon cross-correlated with EndoA1 to total GPN or Bassoon,

respectively. n = 14, from 3 independent cultures. All data are scatterplots with means ± S.E.M.

Figure S5. Effect of *EndoA1* knockout on gephyrin clustering in cultured hippocampal neurons. Related to Figure 4

(A) *EndoA1*$^{fl/fl}$ hippocampal neurons were co-transfected with constructs expressing mCherry and GPN.FingR-eGFP or mCherry-2A-Cre and GPN.FingR-eGFP on DIV12, then fixed and immunostained for eGFP and mCherry on DIV16. Shown are representative confocal microscopy images of GPN.FingR-eGFP. The right panels are magnification of soma or boxed regions in the left panels. Scale bar, 10 μm in the left panels and 5 μm in the right panels.

(B) Quantification of GPN.FingR-eGFP mean intensity in the soma (left), mean intensity (center) and cluster number (right) in dendrites. Scale bar, 5 μm. 20 neurons for mCherry + GPN.FingR-eGFP; 20 neurons for Cre + GPN.FingR-eGFP. Statistical test: unpaired two-tailed Student's t-test. All data are scatterplots with means ± S.E.M.

(C) Same as (E) except that neuron were co-transfected with PSD.FingR-eGFP.

(D) Quantification of PSD95.FingR-eGFP mean intensity in the soma (left), mean intensity (center) and cluster number (right) in dendrites. Scale bar, 5 μm. 14 neurons for mCherry + PSD95.FingR-eGFP; 13 neurons for Cre + PSD95.FingR-eGFP. Statistical test: unpaired two-tailed Student's t-test. All data are scatterplots with means ± S.E.M.

(E-G) *EndoA1*$^{fl/fl}$ hippocampal neurons were transfected with construct expressing mCherry or mCherry-2A-Cre on DIV12 and immunostained for GPN and mCherry on DIV16. Shown are representative confocal microscopy images for GPN in the soma, axon initial segment (AIS) and dendrites 30 μm, 60 μm, 90 μm or120 μm from the soma, respectively. Scale bars, 5 μm.

(H) Quantification of GPN mean intensity normalized to the mCherry group in the soma (A), AIS (B) and dendrites at various distances from the soma (C). Number: 40 (mCherry), 47 (Cre) for soma; 26 (mCherry), 25 (Cre) for AIS; 55 (mCherry), 52 (Cre) for dendrites 30 μm; 52 (mCherry), 50 (Cre) for dendrites 60 μm; 48 (mCherry), 47 (Cre) for dendrites 90 μm; 49 (mCherry), 41 (Cre) for dendrites 120 μm. Ankyrin-G: Axon initial segment (AIS) labeling marker. Statistical test: unpaired two-tailed Student's t-test . All data are scatterplots with means ± S.E.M for each group.

Figure S6. p140Cap promotes gephyrin clustering via endophilin A1-mediated pathway. Related to Figure 6 and 7

(A) Cultured hippocampal neurons transiently expressing GFP and LifeAct-mCherry were immunostained for GPN, GFP and mCherry on DIV16. Shown are representative z stack maximum projection images captured by structured illumination microscopy (SIM). Right panels are insets of boxed regions in the left panel. Scale bars, 2 μm in left panels and 0.5 μm in magnified images.

(B) Cultured hippocampal neurons were co-transfected with Ctrl shRNA and vector or p140Cap

shRNA and vector or RNAi-resistant Flag-tagged p140Cap on DIV 11, followed by immunostaining with antibodies to GPN, Flag (blue) and RFP on DIV 16. Scale bar, 5 μm.

(C) Quantification of GPN mean intensity and puncta number in (B). 18 neurons for shCtrl + vector; 20 neurons for shp140Cap + vector; 22 neurons for shp140Cap + p140Cap, from 3 independent cultures. Statistical test: One-way ANOVA followed by Tukey post-hoc test. All data shown are scatterplots with means ± S.E.M.

(D) *EndoA1*$^{fl/fl}$ hippocampal neurons coexpressing GFP and Ctrl shRNA or p140Cap shRNA, or GFP-2A-Cre and Ctrl shRNA or p140Cap shRNA were immunostained for GPN on DIV16. Shown are representative confocal microscopy images. Scale bar, 5 μm.

(E) Quantification of GPN mean intensity and puncta number in (D). 27 neurons for GFP + shCtrl; 26 neurons for GFP + shp140Cap; 23 neurons for Cre + shCtrl; 26 neurons for Cre + shp140Cap, from 3 independent cultures. Statistical test: One-way ANOVA followed by Tukey post-hoc test. All data shown are scatterplots with means ± S.E.M.

Table S1. Proteins identified by mass spec analysis of endophilin A1 co-immunoprecipitates and their abundance. Related to Figure 4

Video S1. PTZ kindling behaviors of *EndoA1*$^{+/+}$ and *EndoA1*$^{-/-}$ mice. Related to Figure 1

Video S2. The spatial relationship between EndoA1 and GPN. **Related to Figure 3G.** 3D-SIM movie of an enlarged region of interest (ROI) from a hippocampal neuron dendrite shows the association of EndoA1 (red) and GPN (green).

Video S3. The spatial relationship between endophilin A1 and surface GABA$_A$R γ2. **Related to Figure 3H.** 3D-SIM movie of an enlarged region of interest from a hippocampal neuron dendrite shows the association of EndoA1 (red) and surface GABA$_A$R γ2 (green).

Video S4. The spatial relationship between endophilin A1 and VGAT. **Related to Figure 3I.** 3D-SIM movie of an enlarged region of interest from a hippocampal neuron dendrite shows the signals of EndoA1 (red) and VGAT (green).

Video S5. The spatial relationship between Bassoon, gephyrin and endophilin A1. Related to Figure S3B. 3D-SIM movie of an enlarged region of interest from a hippocampal neuron dendrite shows the signals of Bassoon (blue), GPN (green) and EndoA1 (red).

Video S6. The spatial relationship between gephyrin and LifeAct. Related to Figure 6 and Figure S5A. 3D-SIM movie of an enlarged region of interest from a hippocampal neuron dendrite shows the signals of LifeAct (red) and GPN (green).

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1468 **Video S7. PTZ kindling behaviors of *EndoA1*$^{fl/fl}$ mice injected with AAVs expressing Cre**

1469 **and EndoA1 WT or mutants. Related to Figure 7**

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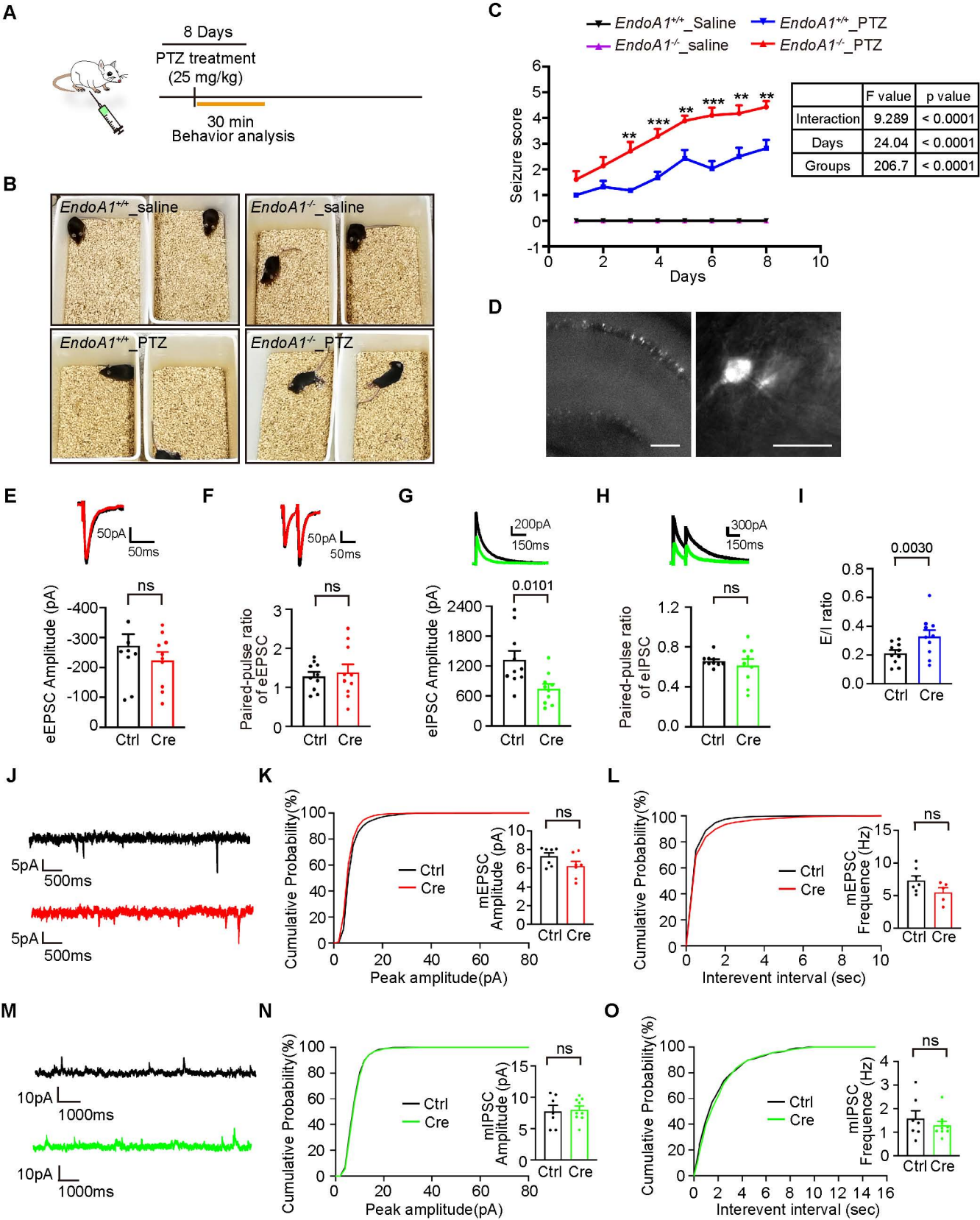
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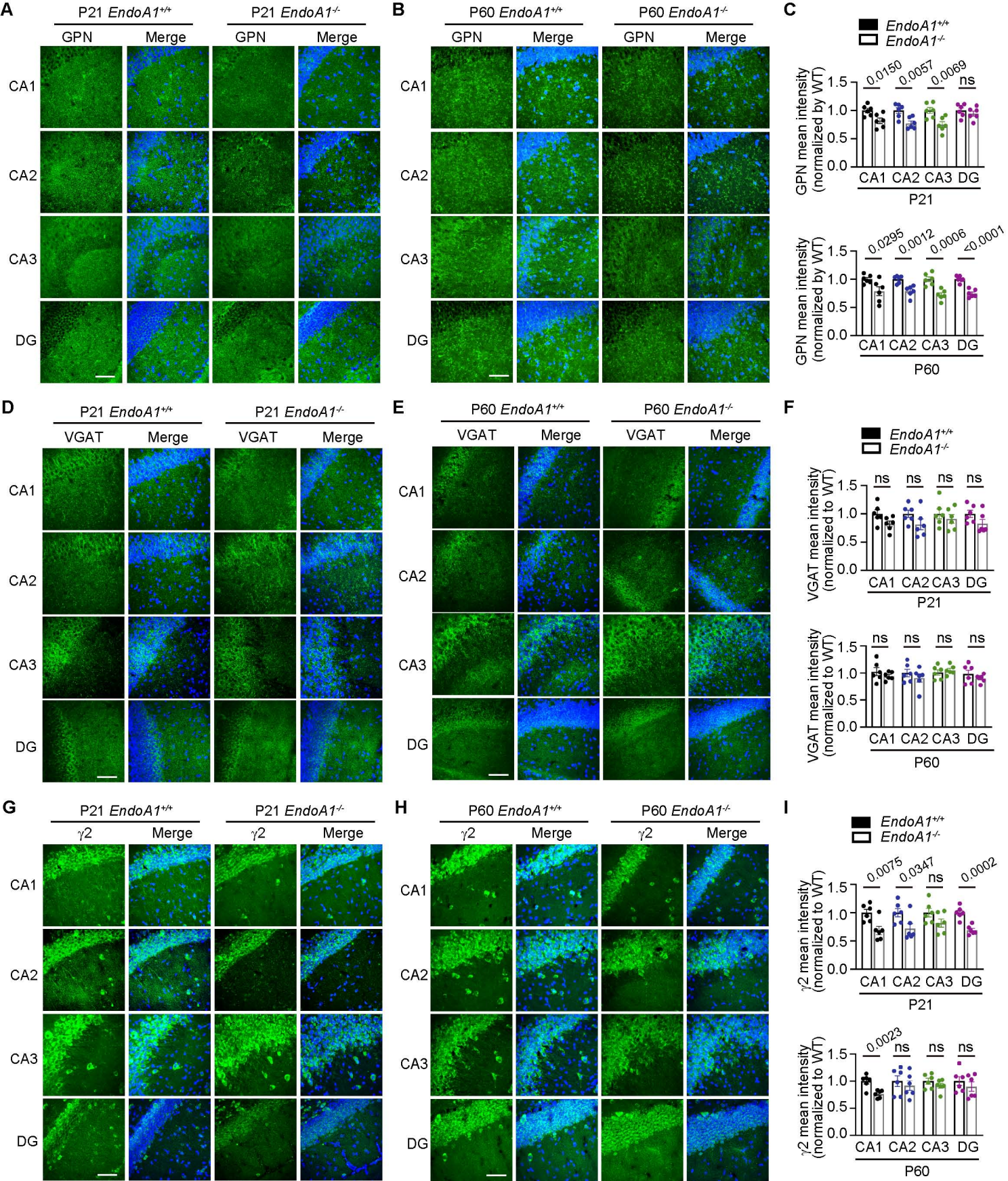
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GPN
Merge
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C
EndoA1+/+
EndoA1-/-
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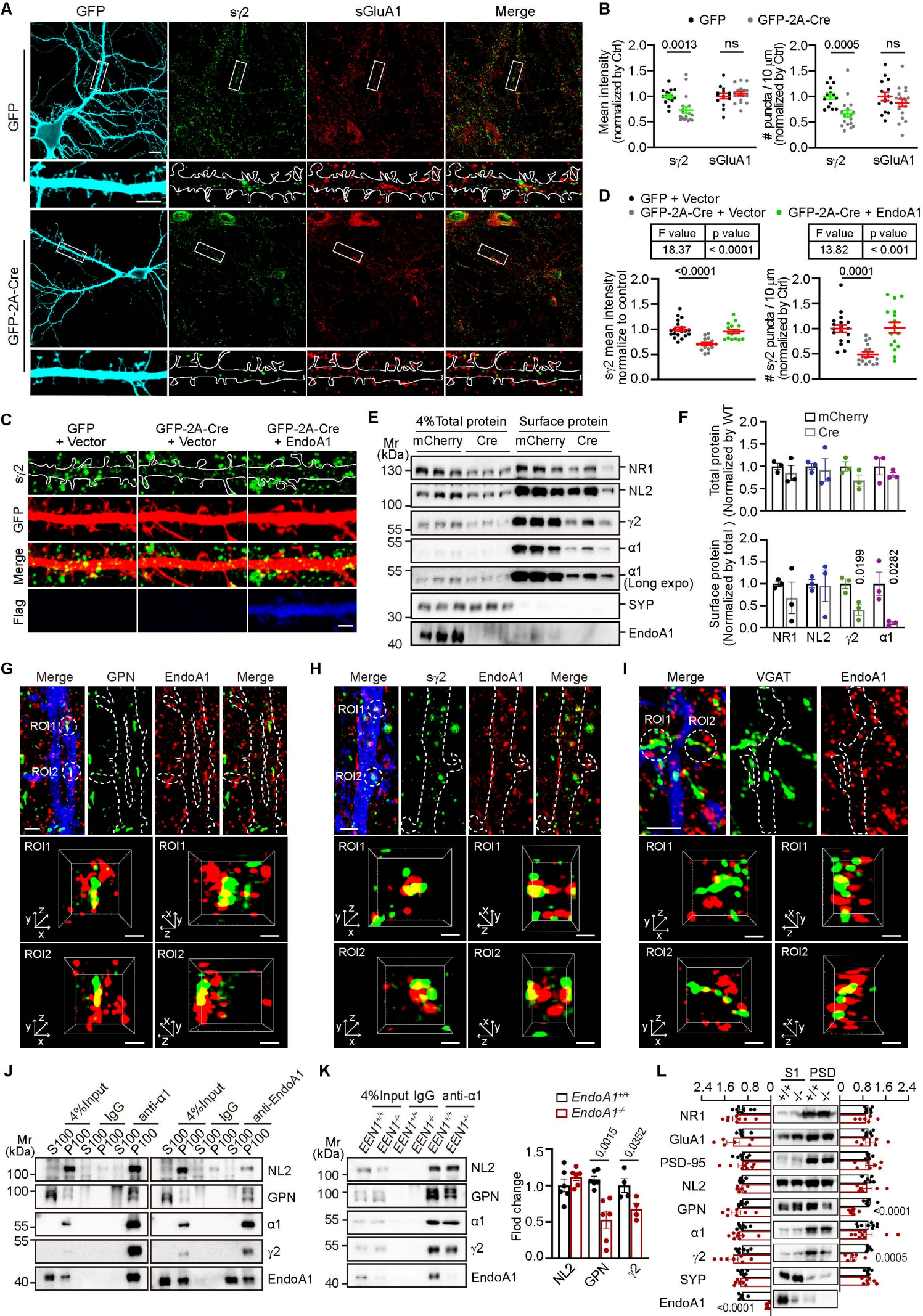
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GFP-2A-Cre + Vector
GFP-2A-Cre + EndoA1
sγ2
GFP
Merge
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E
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mCherry
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NL2
γ2
α1
α1 (Long expo)
SYP
EndoA1
F
mCherry
Cre
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Surface protein (Normalized by total)
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0.0282
NR1
NL2
γ2
α1
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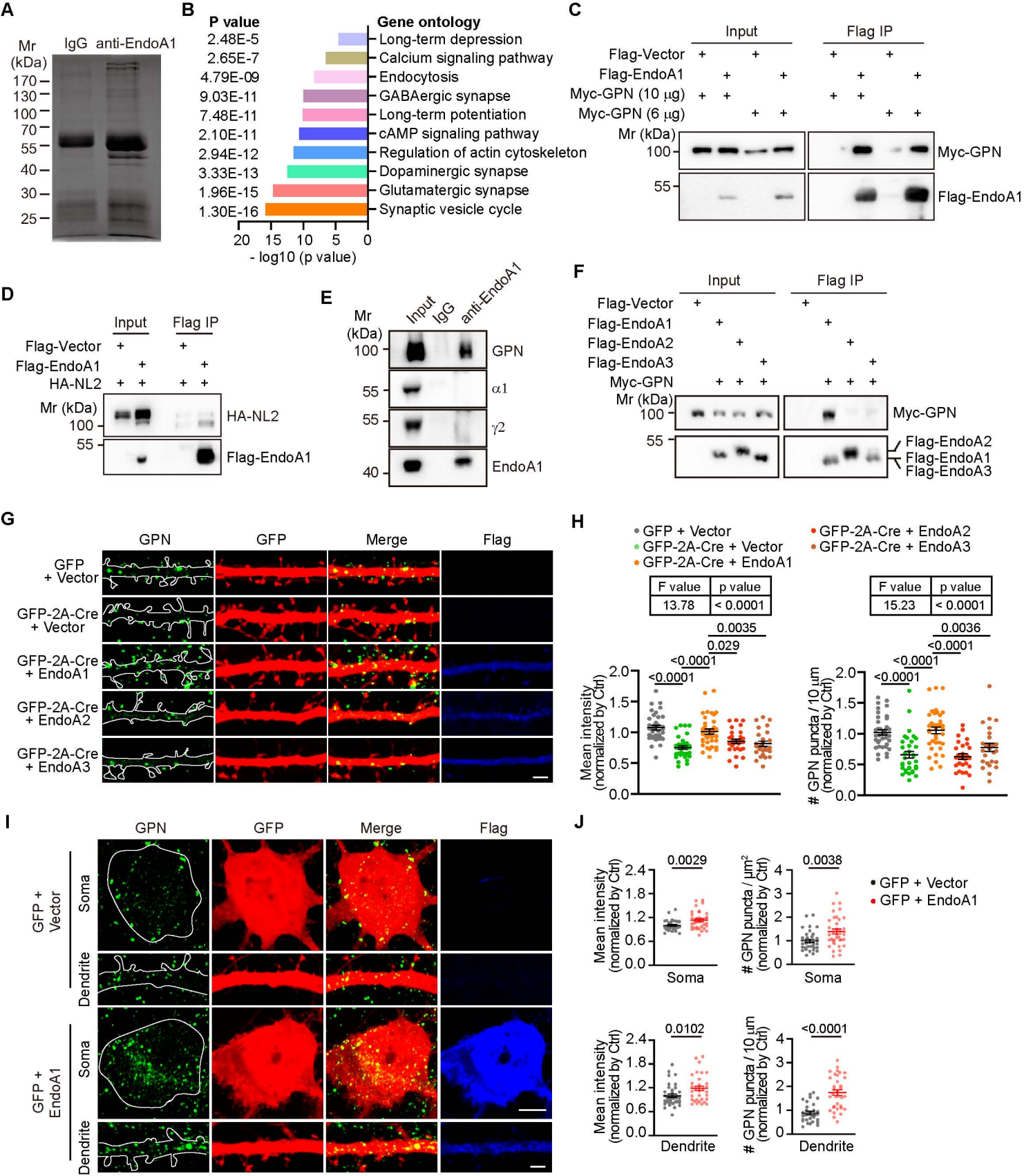
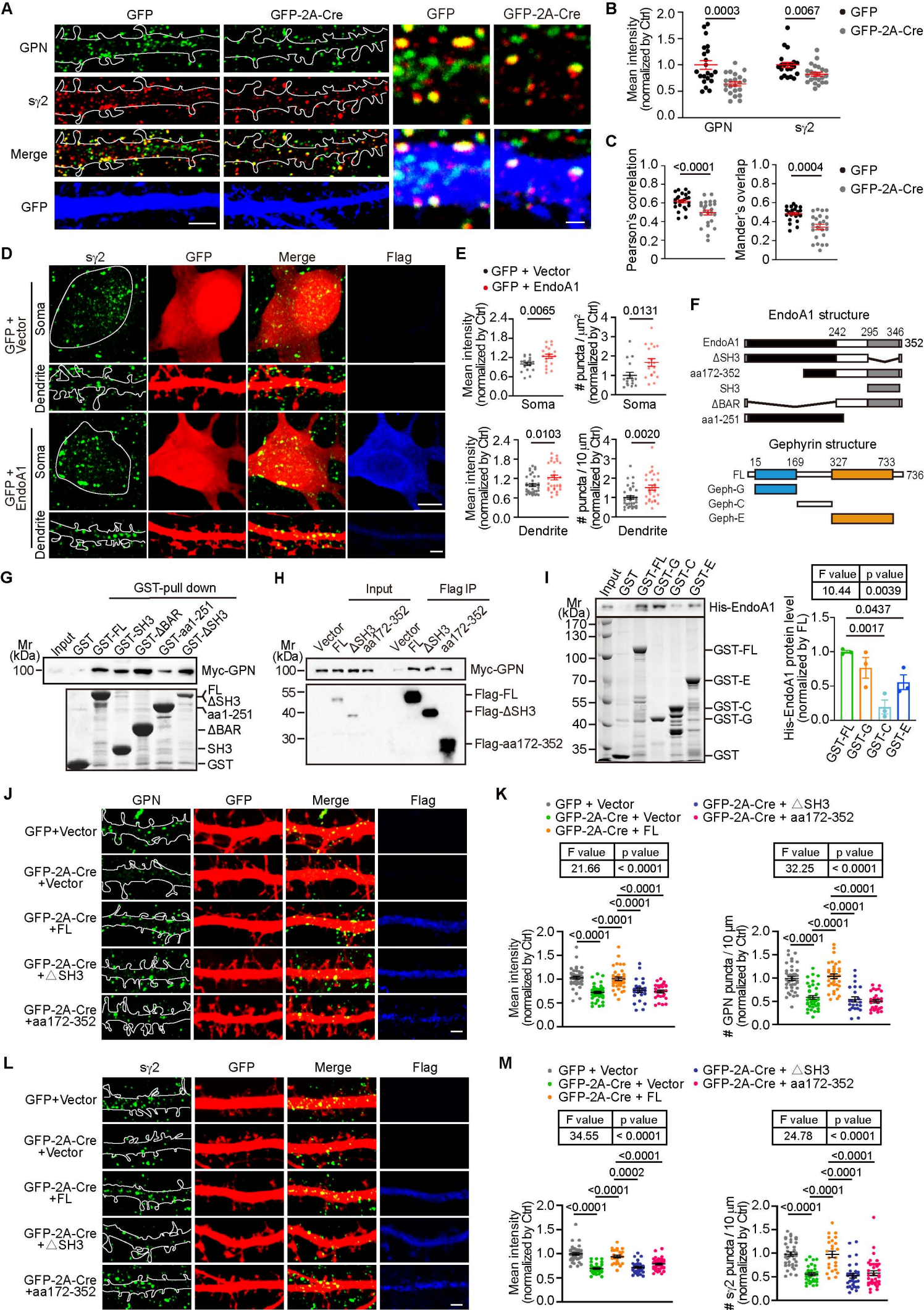
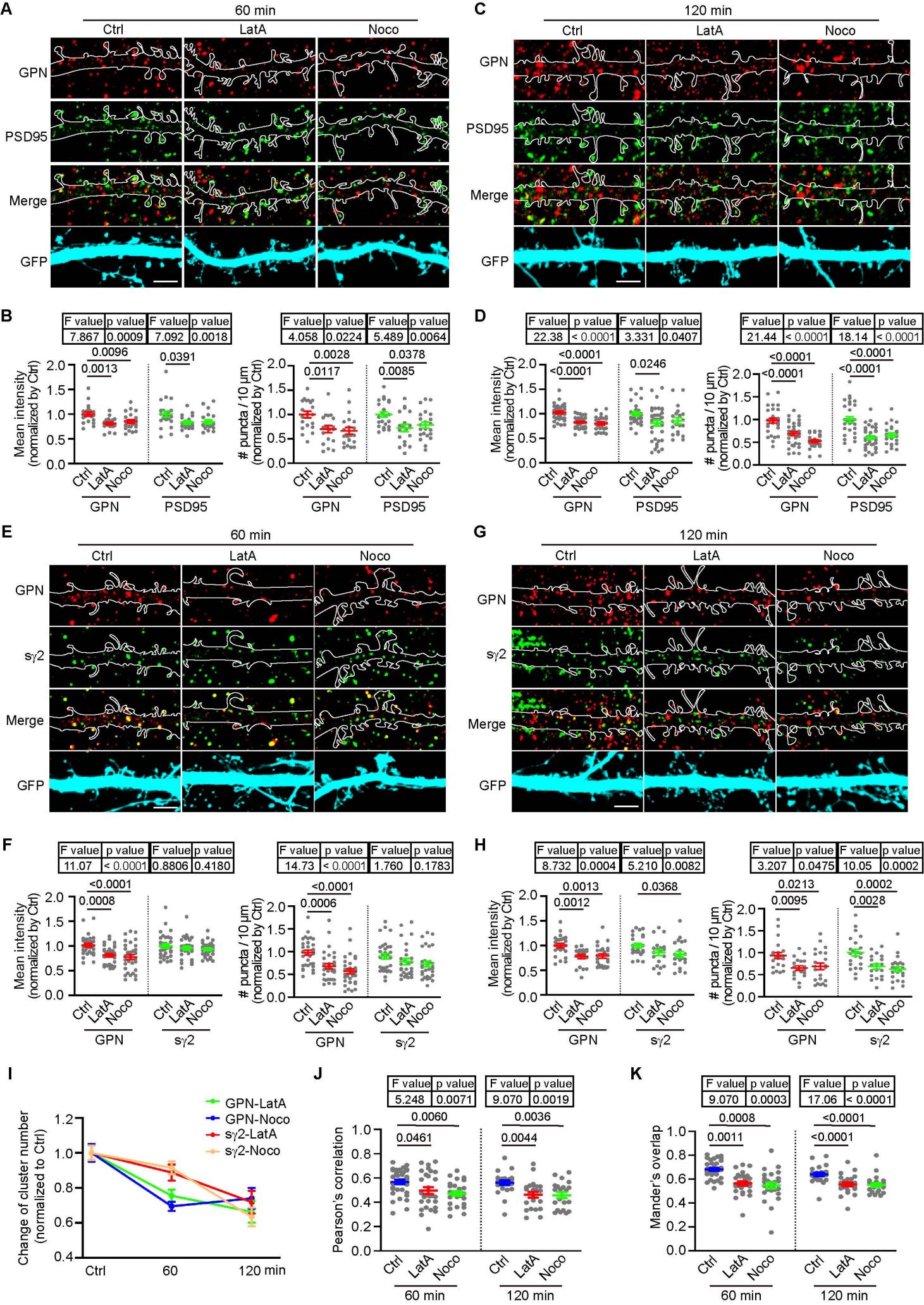
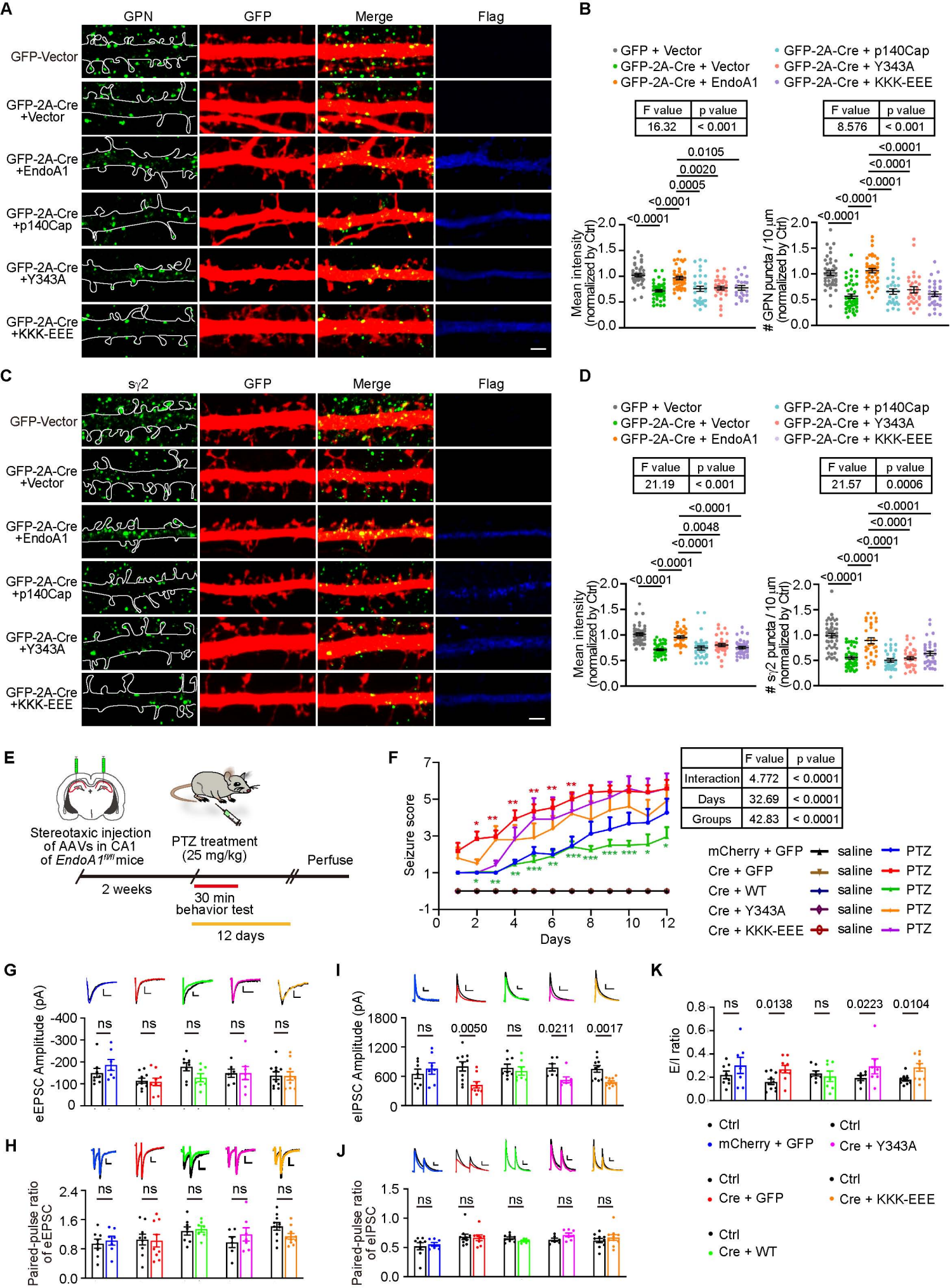
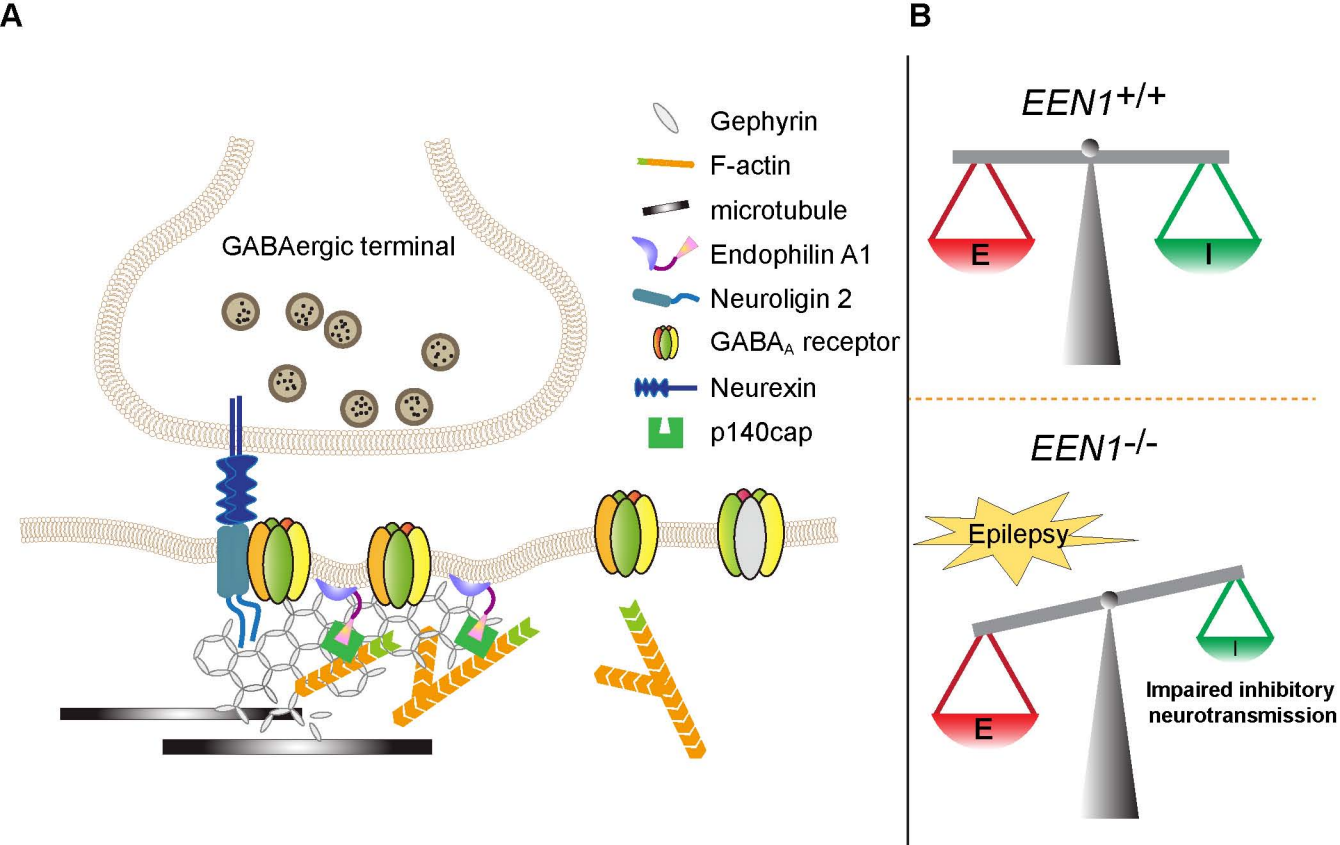
Merge
GPN
EndoA1
Merge
ROI1
ROI2
H
Merge
sγ2
EndoA1
Merge
ROI1
ROI2
I
Merge
VGAT
EndoA1
ROI1
ROI2
J
4%Input
IgG
anti-α1
anti-EndoA1
S100
P100
Mr (kDa)
100
100
55
55
40
NL2
GPN
α1
γ2
EndoA1
K
4%Input
IgG
anti-α1
EEN1+/+
EEN1-/-
Mr (kDa)
100
100
55
55
40
NL2
GPN
α1
γ2
EndoA1
EndoA1+/+
EndoA1-/-
Flod change
0.0015
0.0352
NL2
GPN
γ2
L
S1
PSD
+/+
-/-
NR1
GluA1
PSD-95
NL2
GPN
α1
γ2
SYP
EndoA1
<0.0001
0.0005
<0.0001


A
Mr (kDa)
IgG
anti-EndoA1
170
130
100
70
55
40
30
25
B
P value
Gene ontology
2.48E-5 Long-term depression
2.65E-7 Calcium signaling pathway
4.79E-09 Endocytosis
9.03E-11 GABAergic synapse
7.48E-11 Long-term potentiation
2.10E-11 cAMP signaling pathway
2.94E-12 Regulation of actin cytoskeleton
3.33E-13 Dopaminergic synapse
1.96E-15 Glutamatergic synapse
1.30E-16 Synaptic vesicle cycle
20 15 10 5 0
- log10 (p value)
C
Input
Flag IP
Flag-Vector
Flag-EndoA1
Myc-GPN (10 μg)
Myc-GPN (6 μg)
Mr (kDa)
100
55
Myc-GPN
Flag-EndoA1
D
Input
Flag IP
Flag-Vector
Flag-EndoA1
HA-NL2
Mr (kDa)
100
55
HA-NL2
Flag-EndoA1
E
Mr (kDa)
Input
IgG
anti-EndoA1
100
55
55
40
GPN
α1
γ2
EndoA1
F
Input
Flag IP
Flag-Vector
Flag-EndoA1
Flag-EndoA2
Flag-EndoA3
Myc-GPN
Mr (kDa)
100
55
Myc-GPN
Flag-EndoA2
Flag-EndoA1
Flag-EndoA3
G
GPN
GFP
Merge
Flag
GFP + Vector
GFP-2A-Cre + Vector
GFP-2A-Cre + EndoA1
GFP-2A-Cre + EndoA2
GFP-2A-Cre + EndoA3
H
GFP + Vector
GFP-2A-Cre + Vector
GFP-2A-Cre + EndoA1
GFP-2A-Cre + EndoA2
GFP-2A-Cre + EndoA3
F value | p value
13.78 | < 0.0001
15.23 | < 0.0001
0.0035
0.029
<0.0001
<0.0001
0.0036
<0.0001
<0.0001
<0.0001
Mean intensity (normalized by Ctrl)
GPN puncta / 10 μm (normalized by Ctrl)
2.0
1.5
1.0
0.5
0.0
I
GPN
GFP
Merge
Flag
GFP + Vector
Soma
Dendrite
GFP + EndoA1
Soma
Dendrite
J
0.0029
0.0038
0.0102
<0.0001
Mean intensity (normalized by Ctrl)
GPN puncta / μm² (normalized by Ctrl)
GPN puncta / 10 μm (normalized by Ctrl)
Soma
Dendrite
GFP + Vector
GFP + EndoA1


A
GFP
GFP-2A-Cre
GFP
GFP-2A-Cre
GPN
sγ2
Merge
GFP
B
Mean intensity (normalized by Ctrl)
0.0003
0.0067
GFP
GFP-2A-Cre
GPN
sγ2
C
Pearson's correlation
<0.0001
Mander's overlap
0.0004
GFP
GFP-2A-Cre
D
sγ2
GFP
Merge
Flag
GFP + Vector
Soma
Dendrite
GFP + EndoA1
Soma
Dendrite
E
GFP + Vector
GFP + EndoA1
Mean intensity (normalized by Ctrl)
0.0065
puncta / μm² (normalized by Ctrl)
0.0131
Soma
Mean intensity (normalized by Ctrl)
0.0103
puncta / 10 μm (normalized by Ctrl)
0.0020
Dendrite
F
EndoA1 structure
242
295
346
352
EndoA1
ΔSH3
aa172-352
SH3
ΔBAR
aa1-251
Gephyrin structure
15
169
327
733
736
FL
Geph-G
Geph-C
Geph-E
G
GST-pull down
Mr (kDa)
Input
GST
GST-FL
GST-SH3
GST-ΔBAR
GST-aa1-251
GST-ΔSH3
100
Myc-GPN
FL
ΔSH3
aa1-251
ΔBAR
SH3
GST
H
Input
Flag IP
Mr (kDa)
Vector
FL
ΔSH3
aa172-352
Vector
FL
ΔSH3
aa172-352
100
Myc-GPN
55
40
30
Flag-FL
Flag-ΔSH3
Flag-aa172-352
I
Input
GST
GST-FL
GST-G
GST-C
GST-E
Mr (kDa)
His-EndoA1
170
130
100
70
55
40
35
GST-FL
GST-E
GST-C
GST-G
GST
F value
p value
10.44
0.0039
0.0437
0.0017
His-EndoA1 protein level (normalized by FL)
GST-FL
GST-G
GST-C
GST-E
J
GPN
GFP
Merge
Flag
GFP+Vector
GFP-2A-Cre +Vector
GFP-2A-Cre +FL
GFP-2A-Cre +△SH3
GFP-2A-Cre +aa172-352
K
GFP + Vector
GFP-2A-Cre + Vector
GFP-2A-Cre + FL
GFP-2A-Cre + △SH3
GFP-2A-Cre + aa172-352
F value
p value
21.66
< 0.0001
F value
p value
32.25
< 0.0001
<0.0001
<0.0001
<0.0001
<0.0001
Mean intensity (normalized by Ctrl)
<0.0001
<0.0001
<0.0001
<0.0001
GPN puncta / 10 μm (normalized by Ctrl)
L
sγ2
GFP
Merge
Flag
GFP+Vector
GFP-2A-Cre +Vector
GFP-2A-Cre +FL
GFP-2A-Cre +△SH3
GFP-2A-Cre +aa172-352
M
GFP + Vector
GFP-2A-Cre + Vector
GFP-2A-Cre + FL
GFP-2A-Cre + △SH3
GFP-2A-Cre + aa172-352
F value
p value
34.55
< 0.0001
F value
p value
24.78
< 0.0001
<0.0001
0.0002
<0.0001
<0.0001
Mean intensity (normalized by Ctrl)
<0.0001
<0.0001
<0.0001
<0.0001
sγ2 puncta / 10 μm (normalized by Ctrl)


A
60 min
Ctrl
LatA
Noco
GPN
PSD95
Merge
GFP
C
120 min
B
F value | p value | F value | p value
7.867 | 0.0009 | 7.092 | 0.0018
4.058 | 0.0224 | 5.489 | 0.0064
Mean intensity (normalized by Ctrl)
puncta / 10 µm (normalized by Ctrl)
GPN
PSD95
D
22.38 | < 0.0001 | 3.331 | 0.0407
21.44 | < 0.0001 | 18.14 | < 0.0001
E
sγ2
G
F
11.07 | < 0.0001 | 0.8806 | 0.4180
14.73 | < 0.0001 | 1.760 | 0.1783
H
8.732 | 0.0004 | 5.210 | 0.0082
3.207 | 0.0475 | 10.05 | 0.0002
I
GPN-LatA
GPN-Noco
sγ2-LatA
sγ2-Noco
Change of cluster number (normalized to Ctrl)
60
120 min
J
5.248 | 0.0071
9.070 | 0.0019
Pearson's correlation
60 min
K
9.070 | 0.0003
17.06 | < 0.0001
Mander's overlap


A
GPN
GFP
Merge
Flag
GFP-Vector
GFP-2A-Cre +Vector
GFP-2A-Cre +EndoA1
GFP-2A-Cre +p140Cap
GFP-2A-Cre +Y343A
GFP-2A-Cre +KKK-EEE
B
GFP + Vector
GFP-2A-Cre + Vector
GFP-2A-Cre + EndoA1
GFP-2A-Cre + p140Cap
GFP-2A-Cre + Y343A
GFP-2A-Cre + KKK-EEE
F value | p value
16.32 | < 0.001
8.576 | < 0.001
Mean intensity (normalized by Ctrl)
GPN puncta / 10 μm (normalized by Ctrl)
C
sγ2
GFP
Merge
Flag
D
F value | p value
21.19 | < 0.001
21.57 | 0.0006
Mean intensity (normalized by Ctrl)
sγ2 puncta / 10 μm (normalized by Ctrl)
E
Stereotaxic injection of AAVs in CA1 of EndoA1fl/fl mice
PTZ treatment (25 mg/kg)
Perfuse
2 weeks
30 min behavior test
12 days
F
Seizure score
Days
Interaction | 4.772 | < 0.0001
Days | 32.69 | < 0.0001
Groups | 42.83 | < 0.0001
mCherry + GFP saline PTZ
Cre + GFP saline PTZ
Cre + WT saline PTZ
Cre + Y343A saline PTZ
Cre + KKK-EEE saline PTZ
G
eEPSC Amplitude (pA)
H
Paired-pulse ratio of eEPSC
I
eIPSC Amplitude (pA)
J
Paired-pulse ratio of eIPSC
K
E/I ratio
Ctrl
mCherry + GFP
Cre + GFP
Cre + WT
Cre + Y343A
Cre + KKK-EEE


A
GABAergic terminal
Gephyrin
F-actin
microtubule
Endophilin A1
Neuroligin 2
GABA$_A$ receptor
Neurexin
p140cap
B
EEN1+/+
E
I
EEN1-/-
Epilepsy
E
I
Impaired inhibitory neurotransmission