# A genome-wide association screen for genes affecting leaf trichome development and epidermal metal accumulation in Arabidopsis

Radek Bezvoda, Yazmín Mónica Landeo-Ríos, Zdeňka Kubátová, Eva Kollárová, Ivan Kulich, Wolfgang Busch, Viktor Žárský, Fatima Cvrčková*

*Corresponding author

## Author information:

**Radek Bezvoda**
Department of Experimental Plant Biology, Faculty of Sciences, Charles University, Prague, Czechia
E-mail: radek.bezvoda@natur.cuni.cz
ORCID ID: 0000-0002-9540-7422

**Yazmín Mónica Landeo-Ríos**
Department of Experimental Plant Biology, Faculty of Sciences, Charles University, Prague, Czechia
E-mail: landeoyazmin@gmail.com
ORCID ID: 0000-0001-5391-7005

**Zdeňka Kubátová**
Department of Experimental Plant Biology, Faculty of Sciences, Charles University, Prague, Czechia
E-mail: z.vojtikova@seznam.cz
ORCID ID: 0000-0002-7931-3047

**Eva Kollárová**
Department of Experimental Plant Biology, Faculty of Sciences, Charles University, Prague, Czechia
E-mail: eva.slikova@gmail.com
ORCID ID: 0000-0003-2709-9461

**Ivan Kulich**
Department of Experimental Plant Biology, Faculty of Sciences, Charles University, Prague, Czechia
Present address: Institute of Plant Molecular Biology, Biology Centre, Czech Academy of Sciences, České Budějovice, Czechia
E-mail: ivan.kulich@umbr.cas.cz
ORCID ID: 0000-0002-0458-6470

**Wolfgang Busch**
Plant Molecular and Cellular Biology Laboratory, and Integrative Biology Laboratory, Salk Institute for Biological Studies, La Jolla, CA, USA
Gregor Mendel Institute (GMI), Austrian Academy of Sciences, Vienna Biocenter (VBC), Vienna, Austria
E-mail: wbusch@salk.edu
ORCID ID: 0000-0003-2042-7290

**Viktor Žárský**
Department of Experimental Plant Biology, Faculty of Sciences, Charles University, Prague, Czechia
Institute of Experimental Botany, Czech Academy of Sciences, Prague, Czechia
E-mail: viktor.zarsky@natur.cuni.cz
ORCID ID: 0000-0002-5301-0339

**Fatima Cvrčková**
Department of Experimental Plant Biology, Faculty of Sciences, Charles University, Prague, Czechia
E-mail for correspondence: fatima.cvrckova@natur.cuni.cz
ORCID ID: 0000-0002-5560-8473

## Abstract

We characterized the phenotypic variability of rosette leaf epidermis of 310 sequenced *Arabidopsis thaliana* accessions, focusing on trichome shape and distribution patterns, compositional characteristics of the trichome cell wall (callose contents, mechanical attachment strength and autofluorescence), and on histologically detectable metal ion distribution. A novel metal deposition pattern in stomatal guard cells was observed in some accessions. Subsequent GWAS analysis identified 1546 loci with protein sequence-altering SNPs associated with one or more traits, including five genes with previously reported relevant mutant phenotypes and 80 additional genes with known or predicted roles in relevant developmental and cellular processes. Several large gene families were overrepresented among the candidates, suggesting epidermal development-related functions. In particular, DUF1262, DUF3741 and receptor-like kinases associated with trichome shape traits, cytochrome P450 paralogs with cell wall composition, Cys/His-rich C1 domain proteins with trichome shape and metal deposition, and formin, DUF674, DUF784 and DUF1985 gene families with metal deposition. A possible participation of formins in guard cell metal deposition was supported by observations in available loss of function

mutants. Screening of candidate gene lists against the STRING interactome database uncovered several predominantly nuclear protein interaction networks with possible novel roles in epidermal development.

**Keywords:** *Arabidopsis thaliana,* GWAS, phenotypic variability, trichome, guard cell, metal accumulation

## Introduction

The plant above-ground organs epidermis is a sophisticated interface between the plant and its environment, providing mechanical protection, deterring pathogens and predators and allowing for a controlled gas exchange via stomata. Multiple cell types act in a concerted manner to fulfil these functions, and even subtle disruption of their differentiation may cause observable phenotypic alterations (Zuch et al., 2022). Unconspicuous epidermal defects, well tolerated under laboratory conditions, can indicate perturbation of fundamental cell differentiation or cell morphogenesis processes, as documented, e.g., by identification of mutants defective in actin nucleation based on a distorted trichome phenotype (see Szymanski, 2005).

The most abundant cell type in the leaf epidermis of the model plant *Arabidopsis thaliana* are the puzzle piece-shaped pavement cells. Their morphogenesis is driven by microtubule rearrangements that constrict cell surface expansion to generate indentations and allow actin-dependent intrusion of neighboring cell´s lobes. Coordination of these processes involves cell wall-mediated mechanical feedback and is orchestrated by a regulatory network including ROP small GTPases, their effectors, localized exo- and endocytosis and auxin signaling (see Sapala et al., 2018; Lin and Yang, 2020; Liu et al., 2021; Igisch et al., 2022). Leaf epidermis also contains stomatal complexes, hydathodes and trichomes, whose patterning is controlled by complex regulatory networks (see Torii, 2021). Differentiation of these specialized cell types involves rearrangement of gene expression patterns, well documented in *A. thaliana* for both stomatal guard cells (Zhao et al., 2008; Bates et al., 2012; Xia et al., 2022) and trichomes (Jakoby et al., 2008; Huebbers et al., 2022, Huebbers et al., 2023).

Trichomes contribute to anti-herbivore defense by acting as mechanical deterrents of herbivore feeding; their ecological relevance is well documented (e.g. Sato et al., 2019; Qu et al., 2022). They also engage in detoxification of heavy metals (e.g. Harada et al., 2010; Yaashikaa et al.,.2022) and may sense vibrations and mechanical signals (Zhou et al., 2017; Liu et al., 2017). Development of the branched unicellular *A. thaliana* trichomes is a multi-stage process (Mathur et al., 1999; Han et al., 2022) involving genome endoreduplication, initiation and expansion of trichome branches, and production of secondary cell wall with a characteristic structure and patterns of callose and metal ion deposition (Kulich et al., 2015; Kulich et al., 2018).

Knowledge regarding *A. thaliana* epidermal development comes mainly from studies on a handful of inbred lineages maintained for decades in the laboratory, such as the widespread Columbia accession. Nevertheless, for some epidermal traits, phenotypic variation among natural Arabidopsis accessions has been attributed to sequence polymorphism in specific

genes, such as the *GL1, ETC2* and *MYC1* transcription regulators affecting trichome density (Hauser et al., 2001; Hilscher et al., 2009; Symonds et al., 2011; see also Hauser, 2014) or the *STI* gene controlling trichome branch number (Ilgenfritz et al., 2003).

With the onset of high throughput sequencing and availability of genetically characterized resources linked to computational tools, genome-wide association studies (GWAS) identifying sequence polymorphisms correlated with distinct phenotypic features became a powerful tool for discovering new gene functions. While the GWAS methodology is well established in Arabidopsis (see, e.g., Brachi et al., 2011; Slovak et al., 2015), it found only limited use in studying epidermal development in this species. A screen for host-side genetic determinants of plant-associated microbiome variability revealed genomic regions enriched in genes participating in cell wall biogenesis, and somewhat surprisingly, also in trichome branching (Horton et al., 2014). A recent report applied GWAS to identify genes responsible for variation in aerial organ trichome density (Arteaga et al., 2022). Epidermal or trichome characteristics have, however, been addressed by GWAS in non-model plant species, including commercially important crops. Recent studies focused on trichome distribution patterns in *Brassica napus* (Xuan et al., 2020) and *Aegilops tauschii* (Mahjoob et al., 2022), on cuticular wax deposition in *B. napus* (Jin et al., 2020; Long et al., 2023) and on cotton trichome development (Li et al., 2020; Wang et al., 2021).

Here we report a GWAS for genes associated with phenotypic variability of selected epidermal traits of rosette leaves in 310 sequenced *A. thaliana* accessions from the 1001 Genomes collection (1001 Genomes Consortium, 2016), focusing on traits related to trichome development, cell wall composition and epidermal metal deposition. Besides identifying genes known or already suspected to affect these characteristics, our results suggest possible roles in epidermal development for several gene families with hitherto uncharacterized or only partly characterized functions.

## Results

### Natural variation of rosette leaf epidermis traits

We analyzed 310 natural *A. thaliana* accessions (Supplementary Table S1) to gain insight into the genetic basis of variability in trichome shape, size and density, epidermal autofluorescence and epidermal metal deposition patterns. We recorded 7 continuous quantitative and 11 categorical semiquantitative (ordinate) or qualitative parameters (Table 1; Bezvoda, 2023) that exhibited readily noticeable variability among the accession and were amenable to semi-high-throughput analysis with good inter-observer replicability.

The first group of parameters with continuous distribution (Figure 1) describes quantitative aspects of trichome shape, i.e. size-related parameters (area, perimeter, overall length and

stem length) and descriptors of shape complexity (circularity and solidity). Other morphological features, namely trichome density (Figure 2A, 2B) and branch number (Figure 2C), were evaluated semiquantitatively.

The next group of parameters reflects structural and chemical properties of the trichome cell walls, affecting trichome autofluorescence upon damage, estimated as a semiquantitative categorical variable (Figure 2D, 2E) and callose content, evaluated semiquantitatively on a continuous scale in a manner capturing also the spatial distribution of callose (Figure 2F, 2G). Obvious variability in the background epidermal autofluorescence outside trichomes was recorded as a categorical semiquantitative variable (Figure 2D). We introduced another categorical qualitative variable to describe visible differences in the color of trichome and background fluorescence (Figure 3A).

A third group of categorical semiquantitative parameters captures the spatial distribution of metal ion deposits, as detected by dithizone staining, in trichomes and other epidermal cell types (Figure 3B; for description of parameters see Table 1).

Since we noticed that trichomes of some accessions resist mechanical detachment during staining for callose, we recorded this feature as a qualitative parameter “shaveproof”.

All seven continuous variable parameters exhibit smooth distribution with a single roughly symmetrical maximum (Figure 1B, 1D, 1F; Figure 2G; Figure 4A), except callose content, where most accessions have very little if any callose in trichomes, resulting in an asymmetric distribution. A roughly symmetrical single maximum distribution, in some cases noisy, was observed also for the semiquantitative categorical variables of trichome autofluorescence, epidermis autofluorescence, trichome density and trichome branch number. The “noisy single maximum distribution” may be explainable by under-representation of visually attributed intermediate values, perhaps reflecting observer uncertainty rather than genuine differences. For several other parameters (metal gradient, metal surround, metal stomata) the distribution was asymmetric with most accessions exhibiting low or no signal. Remaining parameters either varied between only two values (autofluorescence color, shaveproof) or showed a possibly bimodal distribution, such as in the case of metals at trichome base and metal rings (Supplementary Figure S1).

Not surprisingly, some quantitative descriptors of trichome shape were mutually correlated **(**Figure 4A, Supplementary Figure S1). Trichome area, perimeter and overall length, which can all be viewed as measures of trichome size, exhibited strong or very strong positive correlation. Weak to moderate positive correlation with these parameters was found also for trichome stem length. Circularity (proportional to the ratio of an object´s area and squared perimeter) and solidity (ratio of the area of the examined object and its convex hull), which

both achieve higher values for less intricate shapes, were mutually moderately positively correlated, and negatively correlated with the size-related parameters. While there were some cases of non-negligible correlation among the categorical variables, or between categorical and continuous ones, such correlations were generally weak except of moderate positive correlation between the frequency of metal rings and intensity of metal gradients at the trichome stems, as well as moderate positive correlation between strength of mechanical attachment (the “shaveproof” trait) and trichome callose content, which was also moderately negatively correlated with trichome length. Remarkably, all the strong or moderate correlations were statistically significant **(**Supplementary Figure S1).

For a subset of traits including representatives of the mutually correlated trait groups, heritability was estimated from data collected separately for individual plants (Supplementary Table S2). The broad sense heritability values, ranging between 0.58 and 0.94, indicate a strong genetic contribution to the observed variability (Supplementary Table S3).

Principal component analysis (PCA) of the 18 recorded parameters did not identify any obvious clustering of accessions when considering the first three principal components (PCs), which, however, cumulatively explained only less than 43 % of total variability (Figure 4B). Analysis of PC loadings indicated that PC1 is positively correlated with the mutually correlated parameters reflecting trichome size, and negatively with circularity and solidity, PC2 correlates either positively or negatively with specific patterns of metal staining, and PC3 mostly reflects callose content and strength of mechanical attachment - i.e. two parameters related to cell wall structure (Figure 4C). The PCA results thus support the overall pattern of continuous variability of traits addressed in our study, and also suggest that trichome shape, metal staining and cell wall organization might be determined by distinct sets of genes.

**GWAS analysis identifies loci associated with some epidermal traits**

The phenotypic data were subjected to association analysis using the publicly available GWAPP GWAS platform (Seren et al., 2012) as described in Materials and Methods, aiming first towards identification of all single nucleotide polymorphisms (SNPs) within the transcribed section of genomic DNA (i.e., including the CDS, introns and the 5´ and 3´ UTRs). The numbers of identified significantly associated loci varied among traits on the scale from zero to several hundred (Table 2; Supplementary Table S4). No significant SNPs were found for trichome area, metal ring, metal staining of trichome bases (“metal base”), and resistance towards mechanical detachment (“shaveproof”), and only one or a few presumably silent polymorphisms (i.e. synonymous mutations or SNPs located in an intron or in untranslated mRNA regions) were identified for overall trichome length, number of

branches, and the presence of a metal gradient. These traits were therefore not included in subsequent analyses.

The distribution of SNP types for individual traits with multiple SNPs exhibits some differences, although they are not major **(**Figure 5). In particular, SNPs altering CDS length were found only for traits with overall large SNP numbers, reflecting low probability of polymorphisms affecting start and stop codons. Subsequently, we focused only on loci with SNPs changing the predicted protein product sequence (in most cases due to a missense mutation), and treated all such SNPs as a single category. The full list of genes affected by such SNPs for each trait is provided in Supplementary Table S5**.** In total, 1547 different loci exhibited association with at least one of the studied traits, corresponding to approximately 5.6 % of all Arabidopsis genes.

For further analysis, we divided the traits with significantly associated SNPs into four groups: those affecting trichome shape (circularity, solidity, perimeter and stem length), trichome distribution on the surface of the leaf (i.e. trichome density), wall composition-related parameters (epidermis and trichome autofluorescence, autofluorescence color, callose content) and metal accumulation (metal staining of stomata and trichome surroundings). While some loci associated with several of the mutually correlated cell shape traits, there was only a very limited, if any, overlap between candidate lists for traits from the wall composition-related and metal accumulation-related groups. However, polymorphisms in some loci were identified as associating with traits from more than one category **(**Figure 6A, Supplementary Table S6).

Assuming that SNPs in transposon-borne genes are unlikely to cause phenotypic differences, we examined what fraction of our candidates corresponds to CDSs from mobile genetic elements to gain insights into the extent of non-specific background. The fraction of transposon-derived loci was significantly depleted compared to whole genome values for the sum of all GWAS candidates, as well as for those associated with shape-related traits or trichome density. However, while depletion of transposon genes was also noticeable for wall-related traits, it was only marginally significant (possibly due to a relatively low number of loci), and practically no depletion was observed for metal-related traits (Table 3). Sets of genes associated with multiple loci usually contained fewer transposon-derived loci than those found on the basis of a single trait (Figure 6B).

**Multiple GWAS candidates encode proteins with known or suspected epidermal roles**

Next, we examined the traits related to trichome development (i.e. the shape traits, density, and the cell wall parameters that largely reflect trichome properties) for enrichment of genes abundantly expressed in trichomes either at the mRNA (Jakoby et al., 2008) or protein

(Huebbers et al., 2022) levels. While we did not detect significant enrichment (or depletion) of these genes in our candidates lists, we identified multiple genes associated with trichome related traits that were abundantly expressed in trichomes, consistent with their participation in trichome development. In some cases, available annotations suggest function in membrane trafficking, biogenesis of cell wall components or turgor generation and maintenance. This might point to possible mechanisms whereby variation in the responsible loci may affect cell growth or cell wall development and generate the observed phenotypic diversity, either by direct modification of trichome morphogenesis or by modulating epidermal cell expansion and hence trichome density (Table 4). Prompted by the finding of several genes engaged in cuticle or epidermal wax biosynthesis, i.e. traits often linked to trichome development (see Berhin et al., 2022) among these loci, we examined the rest of our candidates for overlap with a list of 87 putative cuticle and wax biosynthesis genes identified on the basis of their known activities and/or expression patterns (Suh et al., 2005). However, we did not find any additional loci in this manner beyond two genes that were also highly expressed in trichomes - namely AT1G01600/CYP86A4 and AT1G76690/OPR2 (see Table 4).

Because the loci associated with metal accumulation-related traits mainly reflect stomatal metal deposition, we examined the metal-related candidate list for overlap with a published set of 63 genes upregulated in guard cells (Leonhardt et al., 2004), as well as a list of 1508 proteins repeatedly identified in guard cell proteome (Zhao et al., 2008). We found no overlap between our candidate list and the list of genes highly transcribed in guard cells, but identified 21 loci from the proteome set, all of them associated with metal accumulation in the guard cells and some encoding proteins engaged in ion or other solute transport, membrane trafficking, cell wall biogenesis or gene expression, i.e. processes possibly relevant for the observed trait (Table 5). However, there was no significant enrichment of guard cell-expressed genes among our candidates, similar to the trichome case.

To further verify the performance of our GWAS screen, we inspected the candidate gene lists for genes previously reported to affect the studied traits. While such a search cannot be exhaustive, we found at least five such previously characterized relevant genes (Table 6), including loci coding for two cytoskeletal proteins CLASP and SCAR/DIS3, as well as the EXO84B gene, encoding a subunit of the exocyst complex that regulates targeting of membrane vesicles. All three genes exhibit mutant phenotypes involving trichome shape alterations, and all associated with trichome shape traits. In addition, the gene for another exocyst subunit, EXO70H4, documented to participate in the deposition of metals in the trichome cell wall, was picked up as associating with guard cell metal deposits. The fifth gene, TT9, affects cell wall autofluorescence and was found in association with trichome

density and autofluorescence traits. Additional 80 genes with previously reported roles in cytoskeletal organization, membrane trafficking, cell wall biogenesis, tissue patterning or other relevant functions were also found among the candidates, including several genes affecting aspects of epidermal development unrelated to traits addressed in our study (Supplementary Table S7).

These findings suggest that, despite the lower efficiency of the metal trait- associated gene search (as evident from the lack of transposon gene depletion) the set of candidates found in our screen contains at least a sizeable proportion of genes biologically relevant in the context of leaf epidermis differentiation.

**Overrepresentation of some gene families among GWAS candidates suggests new gene functions**

We next examined the sets of loci associated with the four trait groups (trichome shape, trichome density, wall-related traits and metal-related traits) for Gene Ontology term association. No enrichment or depletion of any GO categories was found on the P = 0.01 significance level, while on the P = 0.05 level, a 2.73 x enrichment of the Transmembrane signal receptor protein class was found among shape-associated loci, and a 12.22 x enrichment of the snoRNA-binding molecular function category was observed among metal trait-associated loci. The biological significance of the latter, however, remains unclear in the light of the apparent low reliability of metal-related candidate detection (see above).

While inspecting our candidate gene lists, we noticed the presence of multiple members of some gene families, including those encoding evolutionarily conserved domains of unknown function (DUF). We thus examined several such multigene families for possible enrichment among candidates for individual trait groups. Our enrichment analyses included all seven DUF families represented by two or more genes in at least one trait group, as well as five large gene families containing multiple candidates associated with at least one trait group, namely the F-box proteins (568 paralogs in Arabidopsis), receptor-like protein kinases (307 paralogs), cytochrome P450 isoforms (256 paralogs), cysteine/histidine-rich C1 domain proteins (153 paralogs) and formins (FH2 proteins, 21 paralogs).

For nine out of these 12 gene families, we found in at least one category of GWAS candidates a reliable, significant at least 2x enrichment compared to whole genome abundance (Figure 7; Supplementary Table S8).These include receptor-like kinases (minor but significant enrichment in shape traits association), cytochrome P450 (enrichment in cell wall fluorescence-related traits, compare also Table 4), cysteine/histidine-rich C1 domain proteins (enrichment in shape and metal accumulation-related traits), formins, DUF674,

DUF784 and DUF1985 proteins (enrichment in metal accumulation traits), as well as the DUF1261 and DUF3741 families that were found in association with trichome shape traits.

Making use of available previously characterized loss of function mutants affecting all three formin genes associating with the stomatal metal deposition, i.e. *FH1*, *FH13* and *FH14*, we performed single-blinded evaluation of epidermal metal deposition in these mutants and corresponding wild type plants. The results suggest a tendency towards increased stomatal metal accumulation especially in the *fh1-1* T-DNA insertion mutant of the formin gene *FH1*, and possibly also in the CRISPR-generated loss of function allele of the same gene **(**Figure 8). Although none of the observed between-genotype quantitative differences was statistically significant, this observation is consistent with a possible contribution of formins to epidermal metal deposition that would deserve further attention, and suggests that also the other observed gene family enrichments may be relevant.

**Associations among candidate genes indicate possible functional modules**

To uncover functional relationships among genes associating with the studied traits, we scanned the candidate lists for mutual links recorded in the STRING database (Sklarczyk et al., 2023) that registers both experimentally documented and predicted interactions among genes (such as, e.g., transcriptional co-regulation, clustering on the chromosomes, epistasis) or proteins (such as, e.g., mutual binding).

Groups of associated genes were found for all four trait groups (Figure 9; Supplementary Figure S2, Supplementary Table S9), but only for trichome density and metal accumulation were the numbers of interactions significantly greater than predicted for a random gene selection (Supplementary Table S9). Inspection of the found clusters, however, reveals biologically meaningful associations also among candidates linked to the remaining trait groups (e.g., among the shape-associated genes, cluster 15 consists of Exocyst complex subunits). For density and metal deposition, the clusters notably included mainly genes participating in nuclear functions such as chromatin organization, gene expression, RNA processing or nuclear transport.

Since we noticed that some genes are shared by clusters found for two or more trait groups, we next performed an analogous search for inter-gene associations among candidates associating with multiple phenotypic trait groups. With somewhat less stringent criteria than those used in the initial interaction searches, this group of genes also exhibited significantly more mutual associations than a random gene selection (Figure 10; Supplementary Table S9), with one large and two smaller clusters comprising mostly genes associated with shape traits, as well as a small cluster consisting of genes linked to trichome density. We believe

that these findings may provide a starting point for exploration of novel regulatory pathways engaged in Arabidopsis epidermal differentiation.

## Discussion

We employed the GWAS approach to explore the genetic basis of *A. thaliana* phenotypic variability, focusing on epidermal traits including trichome development, secondary cell wall composition, production of autofluorescent substances (including those deposited in the cell wall) and metal accumulation. To our knowledge, no similar study has been conducted in Arabidopsis so far, except of a recent report (Arteaga et al., 2022) focusing on trichome patterning and trichome density (the latter also included in our study).

We studied 18 quantitative, semiquantitative or qualitative epidermal traits from 310 predominantly Scandinavian *A. thaliana* accessions from the 1001 Genomes collection (1001 Genomes Consortium, 2016). These traits can be divided into four groups (trichome shape, trichome density, cell wall composition and metal deposition-related). All quantitative parameters, as well as most semiquantitative categorical ones, exhibited continuous variability. As expected, traits reflecting trichome size, as well as those describing trichome shape complexity, were mutually positively correlated.

Surprisingly, we noticed strong dithizone staining, presumably detecting zinc and other heavy metal ion accumulation (see, e.g., Frederickson, 2003; Srivastava et al., 2014; Peco et al., 2020), in stomatal guard cells of some accessions (but not the common Col-0 line). A similar pattern was reported for Cd accumulation in cadmium-stressed Col-0 plants (Zeng et al., 2017). In metazoan cells and in yeast, zinc accumulation is associated with increased secretory activity (Yuan, 2011), and this also may be the case in the guard cells, which exhibit vigorous membrane turnover.

The outcome of GWAS studies always presents a compromise between specificity and sensitivity (see e.g. Brachi et al., 2011). We found statistically significant missense SNPs associated with phenotypic variability for 10 out of the 18 parameters examined, affecting over 5 % of all *A. thaliana* protein-coding genes. Moreover, none of our candidate genes corresponded to any of the 15 trichome density-associated loci found in the recent GWAS report (Arteaga et al., 2022), which was based on phenotyping 191 predominantly Iberian *A. thaliana* accessions. This raises concerns whether our screen was specific and sensitive enough to be informative. However, genotype collection size in both studies was obviously far below the saturation threshold. For the similarly sized human genome, GWAS saturation was achieved only with population size several orders larger (Yengo et al., 2022). Our list of trichome density-associated genes also did not contain any previously characterized trichome patterning genes from the list compiled by Arteaga et al., 2022), nor did we observe

significant enrichment of genes encoding most abundant trichome transcripts (Jakoby et al., 2008) or proteins (Huebbers et al., 2022). In case of the metal accumulation traits, where most candidates reflected the guard cell staining, there was no enrichment of the most abundant guard cell transcripts (Leonhardt et al., 2004), or proteins (Zhao et al., 2008). It is not uncommon that candidate genes from GWAS studies are not enriched in, or even are depleted of, known pathway genes, which would be consistent with the assumption that mutations in large effects genes (which typically tend to be the known genes) might not be beneficial in natural environments (Ristova et al., 2018). Taken together, we believe that at least some of our candidates are biologically relevant, because the candidate genes set, both as a whole and for each trait group except metal accumulation, was significantly (or, in one case marginally significantly) depleted of transposon-borne loci, whose SNPs (unlike polymorphisms concerning transposon presence or position – see, e.g., Yan et al., 2022) are rather unlikely to have phenotypic effects. Thus, the sets of trichome shape, density and probably also wall trait-associated GWAS candidates are likely to contain biologically meaningful genes, while the relevance of candidates found on the basis of metal-related traits remains questionable.

We recovered five loci with well documented mutant phenotypes consistent with the associations identified in our study. First of them is the AT2G20190/CLASP gene coding for a microtubule-associated protein whose loss reduces trichome branch number (Pietra et al., 2013), In this gene, a SNP causing a conserved M1399L substitution in the C-terminal Armadillo-like repeat correlated with trichome stem length polymorphism. The second gene is AT2G38440/DIS3, encoding a subunit of the WAVE complex regulating Arp2/3 mediated actin nucleation, whose mutation causes a characteristic distorted trichome phenotype (Basu et al., 2005). Here, a conserved I1265V substitution in an area immediately preceding the WASP homology motif associated with polymorphism in trichome circularity and solidity, as well as trichome density. The third gene, AT5G49830/EXO84B, codes for an exocyst complex subunit with a pleiotropic mutant phenotype that includes trichome deformation (Kulich et al., 2015; Fendrych et al., 2010). A non-conservative E123G substitution at a variable sequence position of this locus (see Cvrčková et al., 2012) associated with trichome circularity and solidity. A non-conservative S93N substitution in a variable segment of another exocyst subunit, AT3G09520/EXO70H4, involved in metal deposition in the trichome cell wall (Kulich et al., 2015), associated with guard cell metal staining. Last but not least, we found two conservative substitutions (S626A and D671E) associating with trichome autofluorescence and also trichome density in AT3G28430/TT9, which codes for a Golgi-associated peripheral membrane protein involved in membrane trafficking, vacuole

development and flavonoid accumulation whose mutation results in altered autofluorescence (Ichino et al., 2014; Ichino et al., 2020).

Among the loci associated with trichome shape-related traits and also highly expressed in trichomes were three genes participating in the biosynthesis of cutin and/or cuticular waxes. AT1G01600/CYP86A4 encodes a cytochrome P450 paralog, associates also with the trichome autofluorescence trait. Its expression is controlled by a transcription network that regulates also trichome branching (Camoirano et al., 2020; see also Berhin et al., 2022). Remarkably, the cytochrome P450 gene family is significantly over-represented among cell wall trait-associated candidates, also consistent with its role in cell wall biogenesis. The second trichome shape-related candidate with high trichome expression, AT1G12570, encodes a glucose-methanol-choline oxidoreductase related to the maize IPE1 gene participating in pollen exine development, which is co-regulated with CYP86A4 (Kannangara et al., 2007). Polymorphisms in both loci correlate also with variation in trichome density. Remarkably, trichome patterning is altered in some cuticle biogenesis mutants (Berhin et al., 2022), suggesting cuticle contribution to the mechanical cell coupling and tissue patterning (Reynoud et al., 2021). The third locus implicated in cuticle biogenesis is AT1G76690/OPR2, a 12-oxophytodienoic acid reductase co-regulated with the previous two loci (Suh et al., 2005), which only associated with variation of trichome stem length.

Relevant candidates were also found among guard cell-expressed genes. The loci associated with guard cell metal accumulation and highly expressed in guard cells include AT3G13772/TMN7, encoding a membrane protein whose heterologous expression causes yeast to accumulate copper (Hegelund et al., 2010), and AT5G39040/ALS1, coding an ABC transporter whose loss leads to increased aluminum sensitivity (Larsen et al., 2007). Another gene implicated in stomatal metal accumulation, AT3G13460/ECT2, encodes a mRNA stability-controlling protein whose mutational loss affects trichome morphogenesis (Scutenaire et al 2018; Wei et al., 2018). Identification of this locus, which is intensively expressed in guard cells (Zhao et al., 2008) in the context of a guard cell-related phenotype indicates its broader role in epidermal development.

Eighty further candidates with likely roles in biologically relevant processes such as membrane trafficking, cytoskeletal organization or cell wall biogenesis were recovered. Among them are several members of the trichome birefringence-like (TBL) gene family whose founding member was discovered due to altered trichome cell wall optical properties (Bischoff et al., 2010), an additional exocyst subunit, multiple myosin- or kinesin-related proteins, several extensins and expansins, and about 20 cell wall modifying enzymes.

While these observations can be considered confirmatory, our candidate gene lists are significantly enriched in members from several multigene families not yet reported to participate in relevant aspects of epidermal development. The cysteine/histidine-rich C1 domain proteins, a group of nucleic acid binding proteins with a possible role in plant defense (Hwang et al., 2014) were enriched among trichome shape and metal accumulation-linked candidates, and receptor-like kinases (RLKs) associated with trichome shape traits, although no overlap with a published list of trichome-expressed LRR-RLKs (Wu et al., 2016), representing one of the RLK family branches, was detected. Formins, evolutionarily conserved cytoskeletal organizers that also engage in endomembrane dynamics (see Cvrčková et al., 2024), surprisingly associated with stomatal metal accumulation, including the main housekeeping Class I formin FH1/AT3G25500 (Rosero et al., 2016; Oulehlová et al., 2019; Cifrová et al. 2020). Two candidate conservative amino acid substitutions were found in FH1 (L5F within the membrane insertion signal and D502E in a variable cytoplasmic segment). Observations in two loss of function *fh1* mutants suggest a trend towards increased metal accumulation in guard cells. Last but not least, members of five hitherto uncharacterized “domain of unknown function” (DUF) families associated with either trichome shape or metal accumulation traits, including the prolamin-related DUF784 (Zhang, 2009), suggesting their participation in relevant developmental pathways.

Although our Gene Ontology term enrichment analyses did not indicate links between the candidates and specific cellular functions, we observed a significant increase in mutual genetic, physical and functional interactions recorded in the STRING database (Sklarczyk et al., 2023) among candidates for the trichome density and metal deposition traits, as well as for candidates associated with multiple traits, compared to a random gene sample. Remarkably, the interacting gene clusters consisted predominantly of genes participating in nuclear functions. A prominent cluster of candidates associated with two or more trait groups comprising mainly signaling and regulatory proteins was centered on a homolog of the mammalian ATM kinase engaged in DNA damage response (Lee and Paull, 2021). In plants, ATM homologs participate in DNA repair and UV stress response (Shi and Liu, 2021), but also in reaction to exogenous DNA (Vega-Muñoz et al., 2023). Among genes associated with more than one trait category, we also found smaller clusters of shape-associated genes presumably engaged in cytoskeletal and cell wall-related functions, as well as an additional cluster comprising several nuclear proteins and containing genes linked to trichome density.

In summary, our screen identifies several candidate gene families and functional gene clusters that would deserve experimental study to verify their possible roles in epidermal development, possibly leading to uncovering new genetic players in cell morphogenesis and

cell differentiation, but also in functions possibly relevant for environmental adaptation under natural conditions.

## Materials and methods

### Plant material

A set of 310 natural *Arabidopsis thaliana* accessions from the 1001 Genomes collection (1001 Genomes, 2016), has been included in our study (Supplementary Table S1). Predominantly, genotypes of North-European origin were chosen to reduce the effect of geographical differences in population structure and related parameters reducing GWAS efficacy (see Gloss et al., 2022).

Seeds were sterilized with chlorine gas, stratified in tubes at 4 °C for 3 days in the dark and sown into standard soil substrate pre-treated with Confidor to prevent blackfly infestation. Plants were grown in growth chambers under controlled long-day conditions (16 h light, 23 °C/8 h darkness, 18 °C; relative air humidity 60 %). Three seedlings per genotype were planted into individual pots at the first true leaf stage. Leaves were collected at the age of 5 weeks (at which point plants of some genotypes were beginning to bolt).

For mutant genotype verification for selected formin-encoding genes, homozygous plant lines carrying previously characterized loss of function mutants *fh1-1* (Rosero et al., 2013), *fh1:CRISPR* (Cifrová et al., 2020), *fh13-1* (Kollárová et al., 2021), and *fh14-1* (Li et al., 2010) with corresponding congenic wild type plants were used.

### Leaf sample processing

Three approximately fingernail-sized mature, non-senescent rosette leaves per plant (i.e. nine leaves per genotype) were harvested, resulting in three pooled samples, each containing one leaf from each individual. Samples were stored for at least two days before further processing or observation.

For trichome shape evaluation and for determination of callose content in trichomes, leaves were harvested into a tube containing 1 x PBS (phosphate-buffered saline) and 100 mM EGTA. Trichomes were isolated and stained for callose by aniline blue as described previously (Kulich et al., 2015). In genotypes where trichomes failed to detach from the leaves (further referred to as “shaveproof”), whole leaves were stained for callose using an analogous procedure (Kulich et al., 2018). This *in situ* staining was also used for additional documentation, such as the photos shown in Figure 1.

For visualization of autofluorescence and for determination of additional morphological parameters, leaves were pressed adaxial side up between two microscope slides and left to

dry up to induce breakage of trichomes resulting in increase of autofluorescence (Kulich et al., 2018).

For visualization of metal content, leaves were collected into a tube containing 3 ml of acetone and stored for at least several days. On the day of observation, they were stained by addition of diphenylthiocarbazone (dithizone) reagent freshly prepared from 1.5 mg of dithizone, 1 ml of distilled water and 1-2 drops of glacial acetic acid (Seregin and Ivanov, 1997). After at least 1 h but not more than 6 h of staining, leaves were briefly washed in distilled water and mounted in water on microscopy slides for observation.

**Microscopy and imaging**

Microscopic images were acquired using a Nikon Eclipse 90i fluorescence microscope equipped with PlanApo 4x/0.2 objective and Nikon DsFi 2 camera as described previously (Kulich et al., 2018). For autofluorescence and callose fluorescence detection, UV-B excitation was employed; in case of callose staining, an additional image was acquired in polarized light for trichome visualization. Leaves stained for metal detection were imaged using bright field settings.

If whole leaves were photographed, tilling images covering at least a half of the leaf lengthwise were automatically stitched from multiple frames, which were generated as maximum projections of 3 images with 50 µm of Z distance, using the Nikon Imaging Software (NIS Elements AR). Additional image processing was performed using the Fiji platform (Schindelin et al., 2012).

**Epidermal phenotypes determination**

An overview of phenotypic trauts analyzed in our screen is provided in Table 1.

The first group of parameters was determined from photos of callose stained trichomes. At least 25 interactively selected clearly visible trichomes from 3 leaves of 3 plants (detached, or, in shaveproof genotypes, 8-9 in situ trichomes per leaf, selected across its diagonal) were evaluated. Trichome shape parameters (area, circularity, solidity, length and perimeter) were determined from binary images generated by thresholding polarized light photos using the Yen method as implemented in the Fiji software; the procedure was partially automated using recorded macros to increase throughput, with individual trichome selection being the only manual step. Trichome length was measured manually as the longest trichome dimension. Average parameter values from at least 25 trichomes were recorded for these parameters. Callose content was measured as the fraction of trichome area stained for callose, determined using built-in functions of Fiji from binary images generated by thresholding fluorescence and polarized light photos using the Yen method, again with the

aid of recorded macros and manual selection of individual trichomes. The reported value for each accession is the sum of percentage values from 25 trichomes.

The second group of parameters was determined from fluorescence microscopy images of dried whole leaves, taken in the autofluorescence channel. Trichome stem length was measured using the NIS elements software with manual endpoint identification; average values from at least 25 clearly visible trichomes from 3 leaves, selected 7-9 per leaf across its diagonal, are reported. Additional parameters (trichome autofluorescence, overall leaf epidermis autofluorescence, autofluorescence color, trichome density and typical number of trichome branches) were visually categorized (see Table 1).

A third group of parameters reflects visually detectable patterns of metal accumulation, categorized as described in Table 1. This group includes the presence of a metal-enriched Ortmannian ring at the bottom part of the trichome stem (Kulich et al., 2015), presence of a visible metal staining gradient in the trichome stem, presence of metal staining at the trichome base, diffuse epidermal staining outside trichomes, and staining of stomatal guard cells.

The final categorical parameter, denoted as “shaveproof”, reflects the resistance of trichomes against mechanical detachment during staining for callose.

All categorical values were based on the typical appearance of the epidermis of three leaves. Glabrous leaves were attributed the value “0” for all categorical variables. In theexceptional cases where noticeable differences were seen among the three leaves of a given accession, the majority phenotype was recorded.

Phenotype data for a subset of accessions and traits have been independently rescreened by different team members for the purpose of broad sense heritability estimation (Supplementary Table S2); this also documented good inter-observer replicability of phenotype determination (Supplementary Figure S3).

**Phenotypic data processing and initial analyses**

Primary phenotypic data were assembled into a structured spreadsheet and deposited in the public AraPheno database (Togninalli et al, 2020) as Study No. 126 (Bezvoda, 2023). Broad sense heritability (H2 = VG/VP), i.e. the proportion of phenotypic variation (VP) due to genetic variation (VG) estimated from the between- and within-line phenotypic variance, was calculated from trait values of all individuals in this dataset (Alonso-Díaz et al., 2021).

Statistical analysis of phenotypic data was carried out by the Pandas data analysis toolbox (Reback et al., 2021) and pair plots of scatterplots were created by the Seaborn visualization

tool (Waskom, 2021) v. 0.11.2. Correlation strength is reported using criteria from Schober et al. (2018).

Principal component analysis (PCA) has been performed using the PAST software (Hammer et al., 2001) v. 4.11 using the correlation matrix method. All quantitative or semi-quantitative parameters were treated as ordinal variables, the qualitative “autofluorescence color” and “shaveproof” traits were handled as nominal.

**GWAS analyses and genomic data processing**

Phenotype values for each trait were uploaded to the online GWAS application (Seren et al., 2012; Seren, 2024) and correlation analyses were run at 5 % FDR threshold with all possible combinations of input data transformations and statistical methods available with default settings except the minor allele count (MAC) value that has been lowered to 5. The 1001 Full sequence Dataset (TAIR v. 9) was used as the source of genotype information. After initial manual exploration of resulting Manhattan plots, the process of analyzing data was automated.

Raw result data files were downloaded manually and used for subsequent steps. All SNPs identified as significant by at least one method were considered. Additional information about each significant hit was acquired from the GWAS application by web scraping using the Python programming language v. 3.8 (Python Software Foundation, 2024) and driver for Google Chrome web browser (Google, 2024). Locus annotation was based on TAIR9.0 to maintain compatibility with the 1001 genomes dataset. All intergenic hits were discarded from subsequent steps of data processing. Hits were categorized according to the position of the SNP in corresponding gene (INTRON, NON_SYNONYMOUS_CODING, START_LOST, STOP_GAINED, STOP_LOST, SYNONYMOUS_CODING, SYNONYMOUS_STOP, UTR_3_PRIME, UTR_5_PRIME).

Annotation of loci that were subjected to further investigation was individually updated by manual searches of the Araport 11 genome annotation (Cheng et al., 2011) accessed via the BAR Thalemine portal (Pasha et al., 2020).

**Enrichment and depletion analyses of GWAS candidate gene lists**

To determine whether lists of candidate genes identified in the GWAS are enriched or depleted for specific gene groups, we used the Arabidopsis genome (Araport 11 version) and compared our data lists after manual annotation with the following gene lists.

As mobile element-derived genes, those with annotations containing the word “transposable” were considered.

As genes with high transcript levels in trichomes, we considered the 164 genes listed as encoding 5 % most abundant transcripts in the mature trichome transcriptome (Jakoby et al., 2008).

As genes with guard cell-specific transcription patterns, we considered the list of loci specifically upregulated in the guard cells compared to the mesophyll (Leonhardt et al., 2004).

As genes encoding trichome-abundant proteins, we selected genes from the recently published Arabidopsis trichome proteome study (Huebbers et al., 2022) using the following criteria. A protein had to be significantly enriched in at least two of the four reported proteomic experiments, at least in one case by a factor of two and more, while it was not depleted in any experiment. These criteria produced a list of 455 loci.

For genes encoding guard cell- abundant proteins, we considered the list from a published guard cell proteome study (Zhao et al., 2008).

For gene family enrichment, we included all gene family members as identified by keyword search of candidate gene annotations. Family member counts were based either on literature, as in the case of F-box proteins (Kuroda et al., 2002), on TAIR (Reiser et al., 2024) Gene Families annotations (as in the case of receptor-like protein kinases and FH2 proteins), or estimated by keyword searches of the Araport 11 annotations (for all DUFs).

Genes from each list were identified among the GWAS candidates and significance of any observed enrichment or depletion was evaluated using pairwise Chi-square test (Stagroom, 2024) with Benjamini-Hochberg correction for multiplicity performed using an online calculator (Radua et al., 2024).

**Verification of candidates**

Loss-of-function mutants in candidate genes were grown alongside corresponding wild type plants and leaf samples were harvested, processed and imaged as described above for the main ecotype screen. To minimize effects of observer´s expectation bias, a single blinded experimental design was employed, with one member of the team performing the imaging and others quantitatively evaluating microphotographs with coded labels without knowledge of the plant´s genotype. After decoding the identity of the samples, significance of between-genotype differences in the relevant categorical parameters was estimated by the Chi square test (or Fisher's exact test in cases where some categories exhibited zero counts) using online calculators (Stagroom, 2024; Vasavada, 2016).

**Mapping of protein-protein interactions among candidature gene products**

To gain initial insight into the interactions among candidate genes, the full network STRING protein-protein interaction database v. 11.5 (Szklarczyk et al, 2023) was searched with non-redundant lists of genes associated with the indicated trait groups as queries, using high confidence (score = 0.7) and high stringency (FDR = 1 %) settings. Statistical significance of interaction enrichment was obtained during this search. Resulting interaction networks were exported both as tables (further processed in Excel to generate cluster and node lists that were subsequently annotated as described above) and as vector images that were manually edited (to remove unlinked modes, tidy up the layout), annotated and colored.
Mapping of interactions among the subset of candidates associated with multiple trait groups was performed analogously except that the confidence threshold was lowered to medium (score = 0.4), i.e. to the default settings of the STRING database search tool.

## Acknowledgements

We thank Christian Göschl and Samantha Krasnodebski (Gregor Mendel Institute) for help with experiment setup and cultivating plants for the GWAS screen, and Marta Čadyová for technical support.

We acknowledge continuous support from the Charles University Progres Q43 and COOPERATIO programs. The initial stages of this work have been supported by the CSF/GACR/FWF project GF16-34887L (V.Ž., W.B., Y.M.L-R., F.C.), the formin mutant verification experiments by the CSF/GACR 22-33471S grant (E.K. and F.C.), and the finalization of this report by the project TowArds Next GENeration Crops, reg. no. CZ.02.01.01/00/22_008/0004581 of the ERDF Programme Johannes Amos Comenius.

## Author Contributions

R.B. analysed the primary phenotypic data, co-ordinated data management during the project, and drafted most tables, data-based illustrations, and parts of the manuscript, Y.M.L-R, Z.K., E.K., I.K. and F.C. collected phenotypic data during the initial screen, Z.K. and Y.M.L.-R drafted photograph-based figures and parts of the manuscript, F.C. and R.B. rescreened primary data for replicability and heritability estimation, E.K. and F.C. performed phenotypic characterization of formin mutants, W.B. and V.Ž. conceived the project, developed the study design and participated in data analyses and interpretation, F.C. participated in developing the study design, performed PCA, gene family enrichment and interaction network analyses, participated in other data analyses and results interpretation, drafted the manuscript and performed final editing of figures, tables and supplementary materials.

## References


1001 Genomes Consortium (2016). 1,135 genomes reveal the global pattern of polymorphism in *Arabidopsis thaliana*. *Cell*, *166*(2), 481–491. https://doi.org/10.1016/j.cell.2016.05.063

Alonso-Díaz, A., Satbhai, S. B., de Pedro-Jové, R., Berry, H. M., Göschl, C., Argueso, C. T., Novak, O., Busch, W., Valls, M., & Coll, N. S. (2021). A genome-wide association study reveals cytokinin as a major component in the root defense responses against *Ralstonia solanacearum*. *Journal of Experimental Botany*, *72*(7), 2727–2740. https://doi.org/10.1093/jxb/eraa610

Arteaga, N., Méndez-Vigo, B., Fuster-Pons, A., Savic, M., Murillo-Sánchez, A., Picó, F. X., & Alonso-Blanco, C. (2022). Differential environmental and genomic architectures shape the natural diversity for trichome patterning and morphology in different Arabidopsis organs. *Plant, Cell & Environment*, *45*(10), 3018–3035. https://doi.org/10.1111/pce.14308

Basu, D., Le, J., El-Essal, S.el-D., Huang, S., Zhang, C., Mallery, E. L., Koliantz, G., Staiger, C. J., & Szymanski, D. B. (2005). DISTORTED3/SCAR2 is a putative arabidopsis WAVE complex subunit that activates the Arp2/3 complex and is required for epidermal morphogenesis. *The Plant Cell*, *17*(2), 502–524. https://doi.org/10.1105/tpc.104.027987

Bates, G. W., Rosenthal, D. M., Sun, J., Chattopadhyay, M., Peffer, E., Yang, J., Ort, D. R., & Jones, A. M. (2012). A comparative study of the *Arabidopsis thaliana* guard-cell transcriptome and its modulation by sucrose. *PloS One*, *7*(11), e49641. https://doi.org/10.1371/journal.pone.0049641

Berhin, A., Nawrath, C., & Hachez, C. (2022). Subtle interplay between trichome development and cuticle formation in plants. *New Phytologist*, *233*(5), 2036–2046. https://doi.org/10.1111/nph.17827

Bezvoda, R. (2023). AraPheno Study: trichome_development_epidermis_metal_traits. https://doi.org/10.21958/study:126

Bischoff, V., Nita, S., Neumetzler, L., Schindelasch, D., Urbain, A., Eshed, R., Persson, S., Delmer, D., & Scheible, W. R. (2010). TRICHOME BIREFRINGENCE and its homolog AT5G01360 encode plant-specific DUF231 proteins required for cellulose biosynthesis in Arabidopsis. *Plant Physiology*, *153*(2), 590–602. https://doi.org/10.1104/pp.110.153320

Brachi, B., Morris, G. P., & Borevitz, J. O. (2011). Genome-wide association studies in plants: the missing heritability is in the field. *Genome Biology*, *12*(10), 232. https://doi.org/10.1186/gb-2011-12-10-232

Brocker, C., Vasiliou, M., Carpenter, S., Carpenter, C., Zhang, Y., Wang, X., Kotchoni, S. O., Wood, A. J., Kirch, H. H., Kopečný, D., Nebert, D. W., & Vasiliou, V. (2013). Aldehyde dehydrogenase (ALDH) superfamily in plants: gene nomenclature and comparative genomics. *Planta*, *237*(1), 189–210. https://doi.org/10.1007/s00425-012-1749-0

Camoirano, A., Arce, A. L., Ariel, F. D., Alem, A. L., Gonzalez, D. H., & Viola, I. L. (2020). Class I TCP transcription factors regulate trichome branching and cuticle development in Arabidopsis. *Journal of Experimental Botany*, *71*(18), 5438–5453. https://doi.org/10.1093/jxb/eraa257

Cheng, C. Y., Krishnakumar, V., Chan, A. P., Thibaud-Nissen, F., Schobel, S., & Town, C. D. (2017). Araport11: a complete reannotation of the *Arabidopsis thaliana* reference genome. *The Plant Journal*, *89*(4), 789–804. https://doi.org/10.1111/tpj.13415

Cifrová, P., Oulehlová, D., Kollárová, E., Martinek, J., Rosero, A., Žárský, V., Schwarzerová, K., & Cvrčková, F. (2020). Division of labor between two actin nucleators - the formin FH1 and the ARP2/3 complex - in *Arabidopsis* epidermal cell morphogenesis. *Frontiers in Plant Science*, *11*, 148. https://doi.org/10.3389/fpls.2020.00148

Crowell, D. N., Huizinga, D. H., Deem, A. K., Trobaugh, C., Denton, R., & Sen, S. E. (2007). Arabidopsis thaliana plants possess a specific farnesylcysteine lyase that is involved in detoxification and recycling of farnesylcysteine. *The Plant Journal*, *50*(5), 839–847. https://doi.org/10.1111/j.1365-313X.2007.03091.x

Cvrčková, F., Grunt, M., Bezvoda, R., Hála, M., Kulich, I., Rawat, A., & Žárský, V. (2012). Evolution of the land plant exocyst complexes. *Frontiers in Plant Science*, *3*, 159. https://doi.org/10.3389/fpls.2012.00159

Cvrčková, F., Ghosh, R., & Kočová, H. (2024). Transmembrane formins as active cargoes of membrane trafficking. *Journal of Experimental Botany*, *75*(12), 3668–3684. https://doi.org/10.1093/jxb/erae078

Dettmer, J., Hong-Hermesdorf, A., Stierhof, Y. D., & Schumacher, K. (2006). Vacuolar H+-ATPase activity is required for endocytic and secretory trafficking in Arabidopsis. *The Plant Cell*, *18*(3), 715–730. https://doi.org/10.1105/tpc.105.037978

Eggers, R., Jammer, A., Jha, S., Kerschbaumer, B., Lahham, M., Strandback, E., Toplak, M., Wallner, S., Winkler, A., & Macheroux, P. (2021). The scope of flavin-dependent reactions and processes in the model plant Arabidopsis thaliana. *Phytochemistry*, *189*, 112822. https://doi.org/10.1016/j.phytochem.2021.112822

Fendrych, M., Synek, L., Pečenková, T., Toupalová, H., Cole, R., Drdová, E., Nebesářová, J., Šedinová, M., Hála, M., Fowler, J. E., & Žárský, V. (2010). The Arabidopsis exocyst complex is involved in cytokinesis and cell plate maturation. *The Plant Cell*, *22*(9), 3053–3065. https://doi.org/10.1105/tpc.110.074351

Frederickson C. (2003). Imaging zinc: old and new tools. *Science's STKE: Signal Transduction Knowledge Environment*, *2003*(182), pe18. https://doi.org/10.1126/stke.2003.182.pe18

Gloss, A. D., Vergnol, A., Morton, T. C., Laurin, P. J., Roux, F., & Bergelson, J. (2022). Genome-wide association mapping within a local *Arabidopsis thaliana* population more fully reveals the genetic architecture for defensive metabolite diversity. *Philosophical transactions of the Royal Society of London. Series B, Biological sciences*, 377(1855), 20200512. https://doi.org/10.1098/rstb.2020.0512

Google (2024). ChromeDriver. *Chrome for Developers*, https://chromedriver.chromium.org/ (accessed August 20, 2024).

Hammer, Ø., Harper, D.A.T., & Ryan, P.D. (2001.). Past: Paleontological statistics software package for education and data dnalysis. *Palaeontologia Electronica*, 4(1), 4. http://palaeo-electronica.org/2001_1/past/issue1_01.htm.

Han, G., Li, Y., Yang, Z., Wang, C., Zhang, Y., & Wang, B. (2022). Molecular mechanisms of plant trichome development. *Frontiers in Plant Science*, *13*, 910228. https://doi.org/10.3389/fpls.2022.910228

Harada, E., Kim, J. A., Meyer, A. J., Hell, R., Clemens, S., & Choi, Y. E. (2010). Expression profiling of tobacco leaf trichomes identifies genes for biotic and abiotic stresses. *Plant & Cell Physiology*, *51*(10), 1627–1637. https://doi.org/10.1093/pcp/pcq118

Hauser M. T. (2014). Molecular basis of natural variation and environmental control of trichome patterning. *Frontiers in Plant Science*, *5*, 320. https://doi.org/10.3389/fpls.2014.00320

Hauser, M. T., Harr, B., & Schlötterer, C. (2001). Trichome distribution in *Arabidopsis thaliana* and its close relative *Arabidopsis lyrata*: molecular analysis of the candidate gene GLABROUS1. *Molecular Biology and Evolution*, *18*(9), 1754–1763. https://doi.org/10.1093/oxfordjournals.molbev.a003963

Hegelund, J. N., Jahn, T. P., Baekgaard, L., Palmgren, M. G., & Schjoerring, J. K. (2010). Transmembrane nine proteins in yeast and Arabidopsis affect cellular metal contents without changing vacuolar morphology. *Physiologia Plantarum*, *140*(4), 355–367. https://doi.org/10.1111/j.1399-3054.2010.01404.x

Hilscher, J., Schlötterer, C., & Hauser, M. T. (2009). A single amino acid replacement in ETC2 shapes trichome patterning in natural Arabidopsis populations. *Current Biology : CB*, *19*(20), 1747–1751. https://doi.org/10.1016/j.cub.2009.08.057

Horton, M. W., Bodenhausen, N., Beilsmith, K., Meng, D., Muegge, B. D., Subramanian, S., Vetter, M. M., Vilhjálmsson, B. J., Nordborg, M., Gordon, J. I., & Bergelson, J. (2014). Genome-wide association study of *Arabidopsis thaliana* leaf microbial community. *Nature Communications*, *5*, 5320. https://doi.org/10.1038/ncomms6320

Huebbers, J. W., Büttgen, K., Leissing, F., Mantz, M., Pauly, M., Huesgen, P. F., & Panstruga, R. (2022). An advanced method for the release, enrichment and purification of high-quality *Arabidopsis thaliana* rosette leaf trichomes enables profound insights into the trichome proteome. *Plant Methods*, *18*(1), 12. https://doi.org/10.1186/s13007-021-00836-0

Huebbers, J. W., Mantz, M., Panstruga, R., & Huesgen, P. F. (2023). Proteomics dataset on detached and purified *Arabidopsis thaliana* rosette leaf trichomes. *Data in brief*, *46*, 108897. https://doi.org/10.1016/j.dib.2023.108897

Hwang, I. S., Choi, D. S., Kim, N. H., Kim, D. S., & Hwang, B. K. (2014). The pepper cysteine/histidine-rich DC1 domain protein CaDC1 binds both RNA and DNA and is required for plant cell death and defense response. *New Phytologist*, *201*(2), 518–530. https://doi.org/10.1111/nph.12521

Ichino, T., Fuji, K., Ueda, H., Takahashi, H., Koumoto, Y., Takagi, J., Tamura, K., Sasaki, R., Aoki, K., Shimada, T., & Hara-Nishimura, I. (2014). GFS9/TT9 contributes to intracellular membrane trafficking and flavonoid accumulation in *Arabidopsis thaliana*. *The Plant Journal*, *80*(3), 410–423. https://doi.org/10.1111/tpj.12637

Ichino, T., Maeda, K., Hara-Nishimura, I., & Shimada, T. (2020). Arabidopsis ECHIDNA protein is involved in seed coloration, protein trafficking to vacuoles, and vacuolar biogenesis. *Journal of Experimental Botany*, *71*(14), 3999–4009. https://doi.org/10.1093/jxb/eraa147

Igisch, C. P., Miège, C., & Jaillais, Y. (2022). Cell shape: A ROP regulatory tug-of-war in pavement cell morphogenesis. *Current Biology*, *32*(3), R116–R118. https://doi.org/10.1016/j.cub.2021.12.028

Ilgenfritz, H., Bouyer, D., Schnittger, A., Mathur, J., Kirik, V., Schwab, B., Chua, N. H., Jürgens, G., & Hülskamp, M. (2003). The Arabidopsis STICHEL gene is a regulator of trichome branch number and encodes a novel protein. *Plant Physiology*, *131*(2), 643–655. https://doi.org/10.1104/pp.014209

Jakoby, M. J., Falkenhan, D., Mader, M. T., Brininstool, G., Wischnitzki, E., Platz, N., Hudson, A., Hülskamp, M., Larkin, J., & Schnittger, A. (2008). Transcriptional profiling of mature Arabidopsis trichomes reveals that NOECK encodes the MIXTA-like transcriptional regulator MYB106. *Plant Physiology*, *148*(3), 1583–1602. https://doi.org/10.1104/pp.108.126979

Jin, S., Zhang, S., Liu, Y., Jiang, Y., Wang, Y., Li, J., & Ni, Y. (2020). A combination of genome-wide association study and transcriptome analysis in leaf epidermis identifies candidate genes involved in cuticular wax biosynthesis in *Brassica napus*. *BMC Plant Biology*, *20*(1), 458. https://doi.org/10.1186/s12870-020-02675-y

Kannangara, R., Branigan, C., Liu, Y., Penfield, T., Rao, V., Mouille, G., Höfte, H., Pauly, M., Riechmann, J. L., & Broun, P. (2007). The transcription factor WIN1/SHN1 regulates Cutin biosynthesis in *Arabidopsis thaliana*. *The Plant Cell*, *19*(4), 1278–1294. https://doi.org/10.1105/tpc.106.047076

Kollárová, E., Baquero Forero, A., & Cvrčková, F. (2021). The *Arabidopsis thaliana* class II formin FH13 modulates pollen tube growth. *Frontiers in Plant Science*, *12*, 599961. https://doi.org/10.3389/fpls.2021.599961

Kulich, I., Vojtíková, Z., Glanc, M., Ortmannová, J., Rasmann, S., & Žárský, V. (2015). Cell wall maturation of Arabidopsis trichomes is dependent on exocyst subunit EXO70H4 and involves callose deposition. *Plant Physiology*, *168*(1), 120–131. https://doi.org/10.1104/pp.15.00112

Kulich, I., Vojtíková, Z., Sabol, P., Ortmannová, J., Neděla, V., Tihlaříková, E., & Žárský, V. (2018). Exocyst subunit EXO70H4 has a specific role in callose synthase secretion and silica accumulation. *Plant Physiology*, *176*(3), 2040–2051. https://doi.org/10.1104/pp.17.01693

Kuroda, H., Takahashi, N., Shimada, H., Seki, M., Shinozaki, K., & Matsui, M. (2002). Classification and expression analysis of Arabidopsis F-box-containing protein genes. *Plant & Cell Physiology*, *43*(10), 1073–1085. https://doi.org/10.1093/pcp/pcf151

Larsen, P. B., Cancel, J., Rounds, M., & Ochoa, V. (2007). Arabidopsis ALS1 encodes a root tip and stele localized half type ABC transporter required for root growth in an aluminum toxic environment. *Planta*, *225*(6), 1447–1458. https://doi.org/10.1007/s00425-006-0452-4

Lee, J. H., & Paull, T. T. (2021). Cellular functions of the protein kinase ATM and their relevance to human disease. *Nature Reviews. Molecular Cell Biology*, *22*(12), 796–814. https://doi.org/10.1038/s41580-021-00394-2

Leonhardt, N., Kwak, J. M., Robert, N., Waner, D., Leonhardt, G., & Schroeder, J. I. (2004). Microarray expression analyses of Arabidopsis guard cells and isolation of a recessive abscisic acid hypersensitive protein phosphatase 2C mutant. *The Plant Cell*, *16*(3), 596–615. https://doi.org/10.1105/tpc.019000

Li, Y., Shen, Y., Cai, C., Zhong, C., Zhu, L., Yuan, M., & Ren, H. (2010). The type II Arabidopsis formin14 interacts with microtubules and microfilaments to regulate cell division. *The Plant Cell*, *22*(8), 2710–2726. https://doi.org/10.1105/tpc.110.075507

Li, Z., Wang, P., You, C., Yu, J., Zhang, X., Yan, F., Ye, Z., Shen, C., Li, B., Guo, K., Liu, N., Thyssen, G. N., Fang, D. D., Lindsey, K., Zhang, X., Wang, M., & Tu, L. (2020). Combined GWAS and eQTL analysis uncovers a genetic regulatory network orchestrating the initiation of secondary cell wall development in cotton. *New Phytologist*, *226*(6), 1738–1752. https://doi.org/10.1111/nph.16468

Lin, W., & Yang, Z. (2020). Unlocking the mechanisms behind the formation of interlocking pavement cells. *Current Opinion in Plant Biology*, *57*, 142–154. https://doi.org/10.1016/j.pbi.2020.09.002

Liu, S., Jiao, J., Lu, T. J., Xu, F., Pickard, B. G., & Genin, G. M. (2017). Arabidopsis leaf trichomes as acoustic antennae. *Biophysical Journal*, *113*(9), 2068–2076. https://doi.org/10.1016/j.bpj.2017.07.035

Liu, E., MacMillan, C. P., Shafee, T., Ma, Y., Ratcliffe, J., van de Meene, A., Bacic, A., Humphries, J., & Johnson, K. L. (2020). Fasciclin-like arabinogalactan-protein 16 (FLA16) is required for stem development in *Arabidopsis*. *Frontiers in Plant Science*, *11*, 615392. https://doi.org/10.3389/fpls.2020.615392

Liu, S., Jobert, F., Rahneshan, Z., Doyle, S. M., & Robert, S. (2021). Solving the puzzle of shape regulation in plant epidermal pavement cells. *Annual Review of Plant Biology*, *72*, 525–550. https://doi.org/10.1146/annurev-arplant-080720-081920

Long, Z., Tu, M., Xu, Y., Pak, H., Zhu, Y., Dong, J., Lu, Y., & Jiang, L. (2023). Genome-wide-association study and transcriptome analysis reveal the genetic basis controlling the formation of leaf wax in Brassica napus. *Journal of Experimental Botany*, *74*(8), 2726–2739. https://doi.org/10.1093/jxb/erad047

Mahjoob, M. M. M., Kamal, N. M., Gorafi, Y. S. A., & Tsujimoto, H. (2022). Genome-wide association study reveals distinct genetic associations related to leaf hair density in two lineages of wheat-wild relative *Aegilops tauschii*. *Scientific Reports*, *12*(1), 17486. https://doi.org/10.1038/s41598-022-21713-3

Mathur, J., Spielhofer, P., Kost, B., & Chua, N. (1999). The actin cytoskeleton is required to elaborate and maintain spatial patterning during trichome cell morphogenesis in *Arabidopsis thaliana*. *Development*, *126*(24), 5559–5568. https://doi.org/10.1242/dev.126.24.5559

Mitsuda, N., Enami, K., Nakata, M., Takeyasu, K., & Sato, M. H. (2001). Novel type *Arabidopsis thaliana* H(+)-PPase is localized to the Golgi apparatus. *FEBS letters*, *488*(1-2), 29–33. https://doi.org/10.1016/s0014-5793(00)02400-5

Oulehlová, D., Kollárová, E., Cifrová, P., Pejchar, P., Žárský, V., & Cvrčková, F. (2019). Arabidopsis class I formin FH1 relocates between membrane compartments during root cell ontogeny and associates with plasmodesmata. *Plant & Cell Physiology*, *60*(8), 1855–1870. https://doi.org/10.1093/pcp/pcz102

Parsons, H. T., Christiansen, K., Knierim, B., Carroll, A., Ito, J., Batth, T. S., Smith-Moritz, A. M., Morrison, S., McInerney, P., Hadi, M. Z., Auer, M., Mukhopadhyay, A., Petzold, C. J., Scheller, H. V., Loqué, D., & Heazlewood, J. L. (2012). Isolation and proteomic characterization of the Arabidopsis Golgi defines functional and novel components involved

in plant cell wall biosynthesis. *Plant Physiology*, *159*(1), 12–26. https://doi.org/10.1104/pp.111.193151

Pasha, A., Subramaniam, S., Cleary, A., Chen, X., Berardini, T., Farmer, A., Town, C., & Provart, N. (2020). Araport lives: an updated framework for Arabidopsis bioinformatics. *The Plant Cell*, *32*(9), 2683–2686. https://doi.org/10.1105/tpc.20.00358

Peco, J. D., Higueras, P., Campos, J. A., Olmedilla, A., Romero-Puertas, M. C., & Sandalio, L. M. (2020). Deciphering lead tolerance mechanisms in a population of the plant species *Biscutella auriculata* L. from a mining area: Accumulation strategies and antioxidant defenses. *Chemosphere*, *261*, 127721. https://doi.org/10.1016/j.chemosphere.2020.127721

Pietra, S., Gustavsson, A., Kiefer, C., Kalmbach, L., Hörstedt, P., Ikeda, Y., Stepanova, A. N., Alonso, J. M., & Grebe, M. (2013). Arabidopsis SABRE and CLASP interact to stabilize cell division plane orientation and planar polarity. *Nature Communications*, *4*, 2779. https://doi.org/10.1038/ncomms3779

Python Software Foundation (2024). *Python.* https://www.python.org/ (accessed August 20, 2024).

Qu, J., Bonte, D., & Vandegehuchte, M. L. (2022). Phenotypic and genotypic divergence of plant-herbivore interactions along an urbanization gradient. *Evolutionary Applications*, *15*(5), 865–877. https://doi.org/10.1111/eva.13376

Radua, J., Albajes-Elsagirre, A., Fortea. L. (2024). FDR online calculator. *Seed-based d Mapping*, https://www.sdmproject.com/utilities/?show=FDR (accessed August 20, 2024).

Reback, J., jbrockmendel,. McKinney, W., Van den Bossche, J., Augspurger, T., Cloud, P., Hawkins, S.,Roeschke, M., gfyoung, Sinhrks, Klein, A., Petersen, T., Hoefler, P., Tratner, J., She, C., Ayd, V., Naveh, S., Garcia, M., Darbyshire, J.H.M., Schendel, J., …, Seabold, S. (2021) pandas-dev/pandas: Pandas 1.3.5 (v. 1.3.5). *Zenodo*, https://doi.org/10.5281/zenodo.5774815

Reiser, L., Bakker, E., Subramaniam, S., Chen, X., Sawant, S., Khosa, K., Prithvi, T., & Berardini, T. Z. (2024). The Arabidopsis Information Resource in 2024. *Genetics*, *227*(1), iyae027. https://doi.org/10.1093/genetics/iyae027

Reynoud, N., Petit, J., Bres, C., Lahaye, M., Rothan, C., Marion, D., & Bakan, B. (2021). The complex architecture of plant cuticles and its relation to multiple biological functions. *Frontiers in Plant Science*, *12*, 782773. https://doi.org/10.3389/fpls.2021.782773

Ristova, D., Giovannetti, M., Metesch, K., & Busch, W. (2018). Natural genetic variation shapes root system responses to phytohormones in Arabidopsis. *The Plant*, *96*(2), 468–481. https://doi.org/10.1111/tpj.14034

Rosero, A., Žárský, V., & Cvrčková, F. (2013). AtFH1 formin mutation affects actin filament and microtubule dynamics in *Arabidopsis thaliana*. *Journal of Experimental Botany*, *64*(2), 585–597. https://doi.org/10.1093/jxb/ers351

Rosero, A., Oulehlová, D., Stillerová, L., Schiebertová, P., Grunt, M., Žárský, V., & Cvrčková, F. (2016). Arabidopsis FH1 formin affects cotyledon pavement cell shape by modulating cytoskeleton dynamics. *Plant & Cell Physiology*, *57*(3), 488–504. https://doi.org/10.1093/pcp/pcv209

Sapala, A., Runions, A., Routier-Kierzkowska, A. L., Das Gupta, M., Hong, L., Hofhuis, H., Verger, S., Mosca, G., Li, C. B., Hay, A., Hamant, O., Roeder, A. H., Tsiantis, M., Prusinkiewicz, P., & Smith, R. S. (2018). Why plants make puzzle cells, and how their shape emerges. *eLife*, *7*, e32794. https://doi.org/10.7554/eLife.32794

Sato, Y., Shimizu-Inatsugi, R., Yamazaki, M., Shimizu, K. K., & Nagano, A. J. (2019). Plant trichomes and a single gene GLABRA1 contribute to insect community composition on field-grown *Arabidopsis thaliana*. *BMC Plant Biology*, *19*(1), 163. https://doi.org/10.1186/s12870-019-1705-2

Schapire, A. L., Voigt, B., Jasik, J., Rosado, A., Lopez-Cobollo, R., Menzel, D., Salinas, J., Mancuso, S., Valpuesta, V., Baluska, F., & Botella, M. A. (2008). Arabidopsis synaptotagmin 1 is required for the maintenance of plasma membrane integrity and cell viability. *The Plant Cell*, *20*(12), 3374–3388. https://doi.org/10.1105/tpc.108.063859

Schindelin, J., Arganda-Carreras, I., Frise, E., Kaynig, V., Longair, M., Pietzsch, T., Preibisch, S., Rueden, C., Saalfeld, S., Schmid, B., Tinevez, J. Y., White, D. J., Hartenstein, V., Eliceiri, K., Tomancak, P., & Cardona, A. (2012). Fiji: an open-source platform for biological-image analysis. *Nature Methods*, *9*(7), 676–682. https://doi.org/10.1038/nmeth.2019

Schober, P., Boer, C., & Schwarte, L. A. (2018). Correlation coefficients: appropriate use and interpretation. *Anesthesia and Analgesia*, *126*(5), 1763–1768. https://doi.org/10.1213/ANE.0000000000002864

Scutenaire, J., Deragon, J. M., Jean, V., Benhamed, M., Raynaud, C., Favory, J. J., Merret, R., & Bousquet-Antonelli, C. (2018). The YTH domain protein ECT2 is an $m^6A$ reader required for normal trichome branching in Arabidopsis. *The Plant Cell*, *30*(5), 986–1005. https://doi.org/10.1105/tpc.17.00854

Seregin, I.V., & Ivanov, V.B. (1997). Histochemical investigation of cadmium and lead distribution in plants. *Russian Journal of Plant Physiology*, 44(6):791-796. https://www.researchgate.net/publication/279705125_Histochemical_Investigation_of_Cadmium_and_Lead_Distribution_in_Plants

Seren, Ü., Vilhjálmsson, B. J., Horton, M. W., Meng, D., Forai, P., Huang, Y. S., Long, Q., Segura, V., & Nordborg, M. (2012). GWAPP: a web application for genome-wide association mapping in Arabidopsis. *The Plant Cell*, *24*(12), 4793–4805. https://doi.org/10.1105/tpc.112.108068

Seren, Ü. (2015). *The GWA-Portal Resource for phenotypes and GWAS studies.* https://gwas.gmi.oeaw.ac.at/ (accessed August 20, 2024).

Shi, C., & Liu, H. (2021). How plants protect themselves from ultraviolet-B radiation stress. *Plant Physiology*, *187*(3), 1096–1103. https://doi.org/10.1093/plphys/kiab245

Szklarczyk, D., Kirsch, R., Koutrouli, M., Nastou, K., Mehryary, F., Hachilif, R., Gable, A. L., Fang, T., Doncheva, N. T., Pyysalo, S., Bork, P., Jensen, L. J., & von Mering, C. (2023). The STRING database in 2023: protein-protein association networks and functional enrichment analyses for any sequenced genome of interest. *Nucleic Acids Research*, *51*(D1), D638–D646. https://doi.org/10.1093/nar/gkac1000

Slovak, R., Göschl, C., Seren, Ü., & Busch, W. (2015). Genome-wide association mapping in plants exemplified for root growth in *Arabidopsis thaliana*. *Methods in Molecular Biology, 1284*, 343–357. https://doi.org/10.1007/978-1-4939-2444-8_17

Srivastava, R. K., Pandey, P., Rajpoot, R., Rani, A., & Dubey, R. S. (2014). Cadmium and lead interactive effects on oxidative stress and antioxidative responses in rice seedlings. *Protoplasma*, *251*(5), 1047–1065. https://doi.org/10.1007/s00709-014-0614-3

Stagroom, J. (2024). Chi-Square test calculator. *Social Science Statistics*. https://www.socscistatistics.com/tests/chisquare2/default2.aspx (accessed August 20, 2024).

Suh, M. C., Samuels, A. L., Jetter, R., Kunst, L., Pollard, M., Ohlrogge, J., & Beisson, F. (2005). Cuticular lipid composition, surface structure, and gene expression in Arabidopsis stem epidermis. *Plant Physiology*, *139*(4), 1649–1665. https://doi.org/10.1104/pp.105.070805

Symonds, V. V., Hatlestad, G., & Lloyd, A. M. (2011). Natural allelic variation defines a role for ATMYC1: trichome cell fate determination. *PLoS Genetics*, *7*(6), e1002069. https://doi.org/10.1371/journal.pgen.1002069

Szklarczyk, D., Kirsch, R., Koutrouli, M., Nastou, K., Mehryary, F., Hachilif, R., Gable, A. L., Fang, T., Doncheva, N. T., Pyysalo, S., Bork, P., Jensen, L. J., & von Mering, C. (2023). The STRING database in 2023: protein-protein association networks and functional enrichment analyses for any sequenced genome of interest. *Nucleic Acids Research*, *51*(D1), D638–D646. https://doi.org/10.1093/nar/gkac1000

Szymanski D. B. (2005). Breaking the WAVE complex: the point of Arabidopsis trichomes. *Current Opinion in Plant Biology*, *8*(1), 103–112. https://doi.org/10.1016/j.pbi.2004.11.004

Togninalli, M., Seren, Ü., Freudenthal, J. A., Monroe, J. G., Meng, D., Nordborg, M., Weigel, D., Borgwardt, K., Korte, A., & Grimm, D. G. (2020). AraPheno and the AraGWAS Catalog 2020: a major database update including RNA-Seq and knockout mutation data for *Arabidopsis thaliana*. *Nucleic Acids Research*, *48*(D1), D1063–D1068. https://doi.org/10.1093/nar/gkz925

Torii K. U. (2021). Stomatal development in the context of epidermal tissues. *Annals of Botany*, *128*(2), 137–148. https://doi.org/10.1093/aob/mcab052

Vasavada, N. (2016). Fisher test of exact count data. *Online Web Statistical Calculators*. https://astatsa.com/FisherTest/ (accessed August 20, 2024).

Vega-Muñoz, I., Herrera-Estrella, A., Martínez-de la Vega, O., & Heil, M. (2023). ATM and ATR, two central players of the DNA damage response, are involved in the induction of systemic acquired resistance by extracellular DNA, but not the plant wound response. *Frontiers in Immunology*, *14*, 1175786. https://doi.org/10.3389/fimmu.2023.1175786

Vukašinović, N., & Žárský, V. (2016). Tethering complexes in the Arabidopsis endomembrane system. *Frontiers in Cell and Developmental Biology*, *4*, 46. https://doi.org/10.3389/fcell.2016.00046

Wang, X., Miao, Y., Cai, Y., Sun, G., Jia, Y., Song, S., Pan, Z., Zhang, Y., Wang, L., Fu, G., Gao, Q., Ji, G., Wang, P., Chen, B., Peng, Z., Zhang, X., Wang, X., Ding, Y., Hu, D., Geng, X., … Du, X. (2021). Large-fragment insertion activates gene GaFZ (Ga08G0121) and is associated with the fuzz and trichome reduction in cotton (*Gossypium arboreum*). *Plant Biotechnology Journal*, *19*(6), 1110–1124. https://doi.org/10.1111/pbi.13532

Waskom, M. L., (2021). seaborn: statistical data visualization. *Journal of Open Source Software*, *6(60)*, 3021, https://doi.org/10.21105/joss.03021.

Wei, L. H., Song, P., Wang, Y., Lu, Z., Tang, Q., Yu, Q., Xiao, Y., Zhang, X., Duan, H. C., & Jia, G. (2018). The $m^6A$ reader ECT2 controls trichome morphology by affecting mRNA stability in Arabidopsis. *The Plant Cell*, *30*(5), 968–985. https://doi.org/10.1105/tpc.17.00934

Wienkoop, S., Zoeller, D., Ebert, B., Simon-Rosin, U., Fisahn, J., Glinski, M., & Weckwerth, W. (2004). Cell-specific protein profiling in *Arabidopsis thaliana* trichomes: identification of trichome-located proteins involved in sulfur metabolism and detoxification. *Phytochemistry*, *65*(11), 1641–1649. https://doi.org/10.1016/j.phytochem.2004.03.026

Xia, K., Sun, H. X., Li, J., Li, J., Zhao, Y., Chen, L., Qin, C., Chen, R., Chen, Z., Liu, G., Yin, R., Mu, B., Wang, X., Xu, M., Li, X., Yuan, P., Qiao, Y., Hao, S., Wang, J., Xie, Q., … Xu, X. (2022). The single-cell stereo-seq reveals region-specific cell subtypes and transcriptome profiling in Arabidopsis leaves. *Developmental Cell*, *57*(10), 1299–1310.e4. https://doi.org/10.1016/j.devcel.2022.04.011

Xu, J. J., Fang, X., Li, C. Y., Zhao, Q., Martin, C., Chen, X. Y., & Yang, L. (2018). Characterization of *Arabidopsis thaliana* hydroxyphenylpyruvate reductases in the tyrosine conversion pathway. *Frontiers in Plant Science*, *9*, 1305. https://doi.org/10.3389/fpls.2018.01305

Xuan, L., Yan, T., Lu, L., Zhao, X., Wu, D., Hua, S., & Jiang, L. (2020). Genome-wide association study reveals new genes involved in leaf trichome formation in polyploid oilseed rape (*Brassica napus* L.). *Plant, Cell & Environment*, *43*(3), 675–691. https://doi.org/10.1111/pce.13694

Yaashikaa, P. R., Kumar, P. S., Jeevanantham, S., & Saravanan, R. (2022). A review on bioremediation approach for heavy metal detoxification and accumulation in plants. *Environmental Pollution (Barking, Essex: 1987)*, *301*, 119035. https://doi.org/10.1016/j.envpol.2022.119035

Yan, H., Haak, D. C., Li, S., Huang, L., & Bombarely, A. (2022). Exploring transposable element-based markers to identify allelic variations underlying agronomic traits in rice. *Plant Communications*, *3*(3), 100270. https://doi.org/10.1016/j.xplc.2021.100270

Yengo, L., Vedantam, S., Marouli, E., Sidorenko, J., Bartell, E., Sakaue, S., Graff, M., Eliasen, A. U., Jiang, Y., Raghavan, S., Miao, J., Arias, J. D., Graham, S. E., Mukamel, R. E., Spracklen, C. N., Yin, X., Chen, S. H., Ferreira, T., Highland, H. H., Ji, Y., … Hirschhorn, J. N. (2022). A saturated map of common genetic variants associated with human height. *Nature*, *610*(7933), 704–712. https://doi.org/10.1038/s41586-022-05275-y

Yuan D. S. (2011). Dithizone staining of intracellular zinc: an unexpected and versatile counterscreen for auxotrophic marker genes in *Saccharomyces cerevisiae*. *PloS One*, *6*(10), e25830. https://doi.org/10.1371/journal.pone.0025830

Wu, Y., Xun, Q., Guo, Y., Zhang, J., Cheng, K., Shi, T., He, K., Hou, S., Gou, X., & Li, J. (2016). Genome-wide expression pattern analyses of the Arabidopsis leucine-rich repeat receptor-like kinases. *Molecular Plant*, *9*(2), 289–300. https://doi.org/10.1016/j.molp.2015.12.011

Zeng, L., Zhu, T., Gao, Y., Wang, Y., Ning, C., Björn, L. O., Chen, D., & Li, S. (2017). Effects of Ca addition on the uptake, translocation, and distribution of Cd in *Arabidopsis thaliana*.

*Ecotoxicology and Environmental Safety*, *139*, 228–237. https://doi.org/10.1016/j.ecoenv.2017.01.023

Zhang D. (2009). Homology between DUF784, DUF1278 domains and the plant prolamin superfamily typifies evolutionary changes of disulfide bonding patterns. *Cell Cycle*, *8*(20), 3428–3430. https://doi.org/10.4161/cc.8.20.9674

Zhao, Z., Zhang, W., Stanley, B. A., & Assmann, S. M. (2008). Functional proteomics of *Arabidopsis thaliana* guard cells uncovers new stomatal signaling pathways. *The Plant Ccell*, *20*(12), 3210–3226. https://doi.org/10.1105/tpc.108.063263

Zhou, L. H., Liu, S. B., Wang, P. F., Lu, T. J., Xu, F., Genin, G. M., & Pickard, B. G. (2017). The Arabidopsis trichome is an active mechanosensory switch. *Plant, Cell & Environment*, *40*(5), 611–621. https://doi.org/10.1111/pce.12728

Zuch, D. T., Doyle, S. M., Majda, M., Smith, R. S., Robert, S., & Torii, K. U. (2022). Cell biology of the leaf epidermis: Fate specification, morphogenesis, and coordination. *The Plant Cell*, *34*(1), 209–227. https://doi.org/10.1093/plcell/koab250

**Table 1**. Epidermal phenotype parameters included in the GWAS screen.

| Parameter | Refers to | Variable type | Categorical variable values | Sample preparation |
|---|---|---|---|---|
| Area | Trichomes | Continuous | | Callose staining |
| Circularity | Trichomes | Continuous | | |
| Solidity | Trichomes | Continuous | | |
| Length | Trichomes | Continuous | | |
| Perimeter | Trichomes | Continuous | | |
| Callose content | Trichomes | Continuous | | |
| Stem length | Trichomes | Continuous | | Drying (autofluorescence observed under UV light) |
| Trichome autofluorescence | Trichomes | Categorical ordinate | 0 (none); 0,5; 1; 2; 3 (strongest) | |
| Epidermis autofluorescence | Epidermis outside trichomes | Categorical ordinate | 0 (none); 1; 2; 3; 4; 5 (strongest) | |
| Autofluorescence color | Epidermis | Categorical qualitative | 0 (none, whitish or colorless); 1 (bluish or brownish hue) | |
| Trichome density | Epidermis | Categorical ordinate | 0 (none); 0.5; 1; 1.5; 2; 2.5; 3; 4 (densest) | |
| Number of branches | Trichomes | Categorical ordinate | values roughly correspond to typical (or average) trichome branch number | |
| Metal ring | Trichomes | Categorical ordinate | 0 (no rings); 1 (rings in some trichomes); 2 (rings in most or all trichomes) | Metal staining |
| Metal gradient | Trichomes | Categorical ordinate | scale reflects gradient length - 0 (no gradient); 0.5; 1; 1.5; 2; 2.5; 3 (longest) | |
| Metal base | Trichomes | Categorical ordinate | 0 (no staining of trichome base; 1 (in some trichomes); 2 (in most or all trichomes) | |
| Metal surround | Epidermis around trichome base | Categorical ordinate | 0 (no staining around trichome base; 1 (around some trichomes); 2 (around most or all trichomes) | |
| Metal stomata | Guard cells | Categorical ordinate | 0 (no staining; 1 (in some stomata); 2 (in many stomata); 3 (in most or all stomata) | |
| Shaveproof | Epidermis | Categorical qualitative | 0 (trichomes detachable); 1 (shaveproof) | Detachment of trichomes during callose staining |

**Table 2.** Numbers of loci and polymorphism types identified as significantly associated with individual parameters.

| Parameter | Start/stop change | Missense | UTR or intron | Synonymous |
|---|---|---|---|---|
| Area | 0 | 0 | 0 | 0 |
| Circularity | 0 | 44 | 86 | 25 |
| Solidity | 0 | 33 | 66 | 16 |
| Length | 0 | 0 | 1 | 0 |
| Perimeter | 0 | 6 | 15 | 3 |
| Callose content | 0 | 1 | 2 | 1 |
| Stem length | 26 | 657 | 1197 | 574 |
| Trichome autofluorescence | 0 | 21 | 39 | 12 |
| Epidermis autofluorescence | 0 | 11 | 16 | 4 |
| Autofluorescence color | 1 | 52 | 95 | 35 |
| Trichome density | 2 | 216 | 357 | 182 |
| Number of branches | 0 | 0 | 1 | 0 |
| Metal ring | 0 | 0 | 0 | 0 |
| Metal gradient | 0 | 0 | 2 | 1 |
| Metal base | 0 | 0 | 0 | 0 |
| Metal surround | 0 | 16 | 30 | 17 |
| Metal stomata | 27 | 627 | 970 | 425 |
| Shaveproof | 0 | 0 | 0 | 0 |

**Table 3.** Distribution of mobile element-derived genes among candidate loci associated with ORF-changing SNPs.

| Trait group | All loci | Transposon loci | Fraction from transposons | P-value |
|---|---|---|---|---|
| Whole genome (Araport) | 27655 | 3879 | 14% | N.A. |
| All traits | 1547 | 129 | 8% | < 0 .0001 |
| Shape traits | 715 | 43 | 6% | < 0 .0001 |
| Density | 216 | 5 | 2% | < 0 .0001 |
| Wall-related traits | 82 | 6 | 7% | 0.10 |
| Metal-related traits | 654 | 82 | 13% | 0.25 |

P-values indicate significance of the transposon-derived locus depletion compared to the whole genome dataset (pairwise Chi-square test with Benjamini-Hochberg correction for multiplicity). N.A. - not available.

**Table 4.** Candidate loci associated with trichome development traits and highly expressed in trichomes.

| Locus | Gene name(s) | Trait(s) | Gene description | Screen |
|---|---|---|---|---|
| AT1G01600 | CYP86A4 | Stem length, Density, Trichome autofluorescence | Member of the CYP86A subfamily of cytochrome p450 genes. Involved in cutin and wax biogenesis (Camoirano et al., 2020). | T |
| AT1G08920 | ESL1 | Stem length, Density | ERD (early response to dehydration) six-like 1; ESL1, a transporter for monosaccharides. | T |
| AT1G12550 | HPR3 | Stem length | Glyoxylate/hydroxypyruvate reductase HPR3. Transcript restricted to trichomes (Xu et al 2018) | P |
| AT1G12570 | HTH-like | Stem length, Density | Glucose-methanol-choline (GMC) oxidoreductase family protein. Ortholog of maize IPE1 gene participating in pollen exine development. Involved in cutin and wax biogenesis (Kannangara et al., 2007). | P |
| AT1G15670 | KFB1, KMD2 | Stem length | F-box/Kelch-repeat protein. Member of d the KISS ME DEADLY (KMD) family, that targets type-B ARR proteins for degradation and is involved in the negative regulation of the cytokinin response. Also named as KFB1, a member of a group of Kelch repeat F-box proteins that negatively regulate phenylpropanoid biosynthesis. | T |
| AT1G26400 | FOX3, ATBBE5 | Stem length | Berberine bridge enzyme-like 5. Possible role in defense, related proteins participate in wall modification (Eggers et al., 2021). | P |
| AT1G53210 | NCL | Stem length | Sodium/calcium exchanger NCL. | P |
| AT1G76690 | OPR2 | Stem length | 12-oxophytodienoic acid reductase with possible role in cutin or wax synthesis (Suh et al., 2005). | P |
| AT1G78920 | AVPL1 | Stem length | Pyrophosphate-energized membrane proton pump 2. Located at Golgi, strong expression in trichomes confirmed using a GUS reporter (Mitsuda et al 2001). | P |
| AT2G20240 | | Stem length | GPI-anchored adhesin-like protein, putative (DUF3741). | P |

| | | | | |
|---|---|---|---|---|
| AT2G20990 | SYTA, SYT1 | Stem length | Synaptotagmin A . Regulates endosome recycling at the plasma membrane, but not membrane traffic along the secretory pathway. Strong expression in trichomes confirmed using a GUS reporter (Schapire et al., 2008). | P |
| AT2G21410 | VHA-A2 | Stem length | V-type proton ATPase subunit a2. Required for endocytic and secretory trafficking (Dettmer et al., 2006). | P |
| AT2G21870 | MGP1, PHI1 | Stem length | Probable ATP synthase 24 kDa subunit, mitochondrial. Essential for pollen formation. | P |
| AT2G35860 | FLA16 | Density | Fasciclin-like arabinogalactan protein. Mutant has reduced cell size in some cell types in the stem (Liu et al., 2020). | P |
| AT3G28500 | RPP2C, P2X | Density | 60S acidic ribosomal protein P2-3. | P |
| AT3G48000 | ALDH2B4 | Stem length | Aldehyde dehydrogenase family 2 member B4, mitochondrial. May participate in biosynthesis of ferulic acid and sinapic acid, modulating cell wall strength (Brocker et al., 2013). | P |
| AT3G59010 | PME35, PME61 | Density | Pectin methyl esterase, activity controls cell wall mechanic strength; identified in a trichome transcriptome study (Wienkoop et al., 2004) | P |
| AT4G02930 | TUFA | Stem length | Elongation factor Tu, mitochondrial. | P |
| AT4G24840 | COG2 | Stem length | Oligomeric Golgi complex subunit-like protein containing the COG2 domain, part of the membrane trafficking machinery (see Vukašinovič and Žárský, 2016) | P |
| AT5G28220 | | Stem length | Protein prenylyltransferase (uncharacterized). | P |
| AT5G63910 | FCLY | Density | Farnesylcysteine lyase (EC 1.8.3.5) involved in a salvage /detoxification pathway of farnesylcysteine (FC) residues that are liberated during the degradation of prenylated proteins (Crowell et al., 2007). | P |

Unless additional references are provided, annotation is derived from Araport 11 or the TAIR description (Reiser et al., 2024). T – transcriptome, P – proteome.

**Table 5.** Candidate loci associated with stomatal metal accumulation and expressed in the guard cell proteome.

| Locus | Gene name(s) | Gene description |
|---|---|---|
| AT1G31850 | | S-adenosyl-L-methionine-dependent methyltransferases superfamily protein; probable methyltransferase PMT20; found in a Golgi complex proteome study (Parsons et al., 2012) |
| AT1G42550 | PMI1 | Protein PLASTID MOVEMENT IMPAIRED 1, a plant-specific protein of unknown function conserved among angiosperms |
| AT1G59820 | ALA3 | Phospholipid-transporting ATPase (phospholipid translocase) involved in secretory vesicle formation from trans-Golgi. |
| AT1G59900 | E1 ALPHA | Pyruvate dehydrogenase E1 component subunit alpha-1, mitochondrial. |
| AT1G67490 | GCS1, KNF, KNOPF | Mannosyl-oligosaccharide glucosidase GCS1; an alpha-glucosidase I enzyme that catalyzes the first step in N-linked glycan processing. Localized to the endoplasmic reticulum. |
| AT3G10670 | ABCI6, NAP7 | ABC transporter I family member 6, chloroplastic. Plastidic SufC-like ATP-binding cassette/ATPase essential for Arabidopsis embryogenesis. Involved in the biogenesis and/or repair of oxidatively damaged Fe–S clusters. |
| AT3G13460 | ECT2 | Evolutionarily conserved C-terminal region 1; regulates the mRNA levels of the proteasome regulator PTRE1 and of several 20S proteasome subunits, resulting in enhanced 26S proteasome activity. |
| AT3G13772 | TMN7 | Transmembrane 9 superfamily member, localized in the secretory pathway. Overexpression of this protein in yeast alters copper and zinc homeostasis (Hegelund et al., 2010). |
| AT3G48000 | ALDH2B4, ALDH2, ALDH2A | Aldehyde dehydrogenase family 2 member B4, mitochondrial. |
| AT4G13360 | | 3-hydroxyisobutyryl-CoA hydrolase-like protein 3, mitochondrial; ATP-dependent caseinolytic (Clp) protease/crotonase family protein. |
| AT4G18290 | KAT2 | Member of the Shaker family potassium ion (K+) channel. Critical to stomatal opening induced by blue light and to circadian rhythm of stomatal opening. Involved in plant development in response to high light intensity. |
| AT4G18670 | LRX5 | Leucine-rich repeat extensin-like protein 5, involved in cell wall biogenesis and organization. Interacts with several members of the RALF family of ligand peptides. |
| AT4G20400 | JMJ14 | Histone H3K4 demethylase repressing floral transition. |
| AT4G21180 | ERDJ2B | DnaJ domain protein localized in ER membrane. |

| | | |
|---|---|---|
| AT5G35430 | NOT10 | Tetratricopeptide repeat (TPR)-like superfamily protein, found also in stress granules. |
| AT5G39040 | ABCB27, ALS1, TAP2 | Aluminum sensitive 1, ABC transporter B family member 27; a member of TAP subfamily of ABC transporters that is located in the vacuole. Mutants are hypersensitive to aluminum and the gene product may be important for intracellular movement of some substrate, possibly chelated Al, as part of a mechanism of aluminum sequestration. |
| AT5G43130 | TAF4B | Transcription initiation factor TFIID subunit 4b. |
| AT5G52470 | MED36B, FBR1, FIB1, SKIP7 | Probable mediator of RNA polymerase II transcription subunit 36b; encodes a fibrillarin, a key nucleolar protein in eukaryotes which associates with box C/D small nucleolar RNAs (snoRNAs) directing 2'-O-ribose methylation of the rRNA. |
| AT5G55040 | BRD13 | DNA-binding bromodomain-containing protein; interacts with core SWI/SNF complex components.Identified as a specific subunit of BRM-associated SWI/SNF (BAS) complexes |
| AT5G58100 | IEF7 | Encodes a membrane protein involved in pollen nexine and sexine development. |
| AT5G58140 | PHOT2 | Phototropin-2; membrane-bound protein serine/threonine kinase that functions as blue light photoreceptor in redundancy with PHO1. Involved in stomatal opening, chloroplast movement and phototropism. |

Unless additional references are provided, annotation is derived from Araport 11 or the TAIR description (Reiser et al., 2024).

**Table 6**. Candidate loci that were previously characterized as participating in the development of relevant epidermal traits.

| Locus | Gene name(s) | Trait(s) | Gene description |
|---|---|---|---|
| AT2G20190 | CLASP | Stem length | CLIP-associated protein; a microtubule-associated protein involved in both cell division and cell expansion. It likely promotes microtubule stability. Loss of function mutants have decreased trichome branch number (Pietra et al., 2013) |
| AT2G38440 | DIS3, ITB1, SCAR2, WAVE4 | Circularity, Solidity, Density | Subunit of the WAVE complex, required for activation of ARP2/3 actin nucleation complex. Mutations cause defects in both the actin and microtubule cytoskeletons that result in aberrant epidermal cell expansion. Loss of function mutants exhibit a distorted trichome phenotype (Basu et al., 2005). |
| AT3G09520 | EXO70H4 | Metal stomata | A member of the EXO70 gene family of putative exocyst subunits, conserved in land plants. Involved in trichome cell wall maturation, required for deposition of heavy metals in the cell wall (Kulich et al., 2015, though guard cell localization was not specifically observed). |
| AT3G28430 | GFS9, TT9 | Density, Trichome autofluorescence. | Peripheral membrane protein localized at the Golgi apparatus that is involved in membrane trafficking, vacuole development and flavonoid accumulation. Loss of function mutants exhibit abnormal autofluorescence, though trichomes were not specifically examined (Ichino et al., 2014). |
| AT5G49830 | EXO84B | Circularity, Solidity | Exocyst complex component 84B; involved in tethering vesicles to the plasma membrane. Interacts with Exo70H4 that controls trichome cell wall development; loss of function mutant has multiple defects including in trichome morphogenesis (Kulich et al., 2015; Fendrych et al., 2010). |

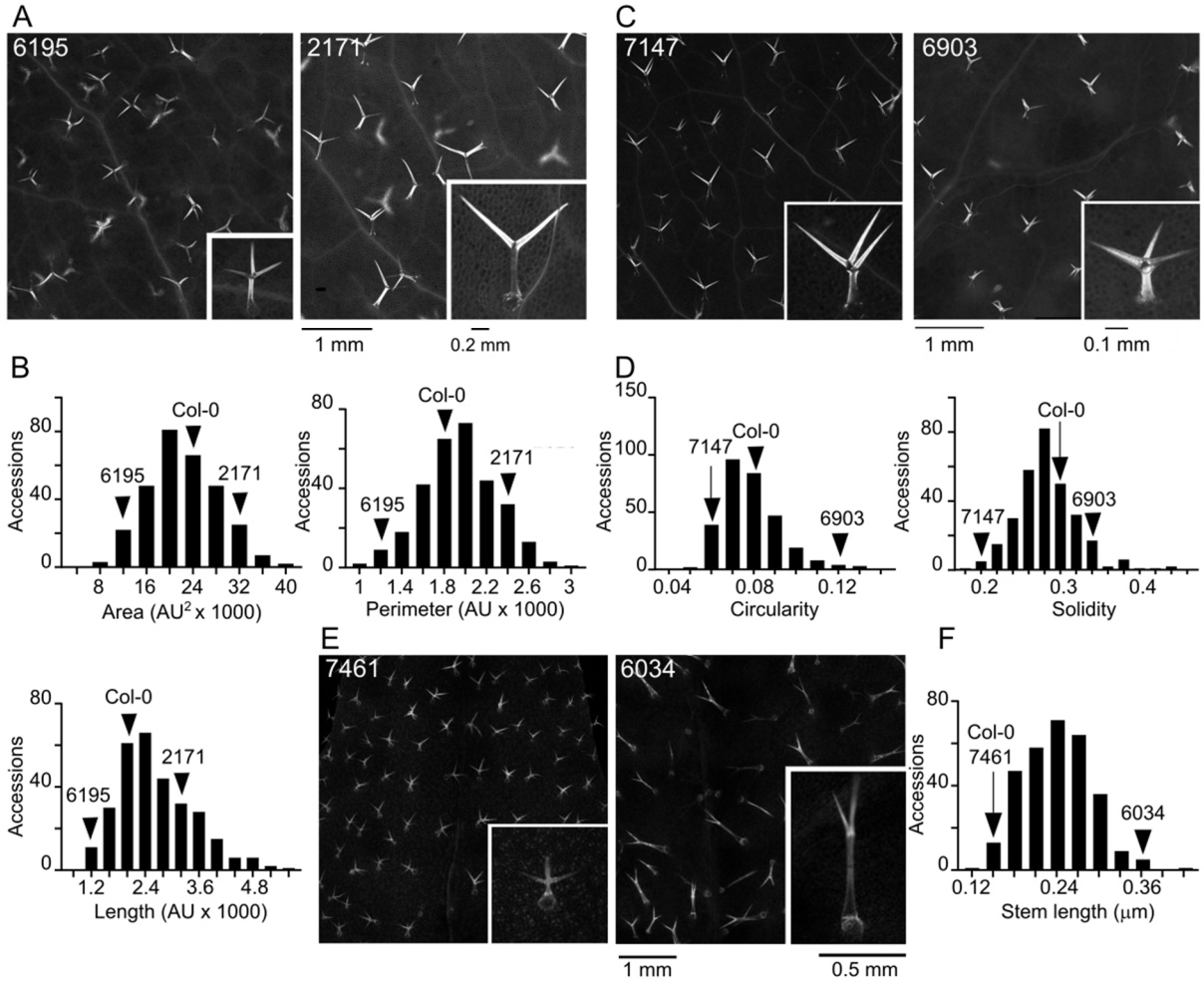


**Figure 1. Examples of continuous quantitative parameters of trichome shape.** (A) Detailed view of representative leaves of ecotypes with low (6195, TDr-9) and high (2171, Paw-26) trichome area, perimeter and length with close-up of a typical trichome. (B) Histograms showing distribution of area, perimeter and length among analyzed accessions. (C) Detailed view of representative leaves of ecotypes with low (7147, Gie-0) and high (6903, Bor-4) trichome circularity and solidity with close-up of a typical trichome. (D) Histograms showing distribution of circularity and solidity among analyzed accessions. (E) Detailed view of representative leaves of ecotypes with low (7461, H55) and high (6034, Hov1-7) trichome stem length with close-up of a typical trichome. (F) Histogram presenting distribution of stem length among analyzed accessions. Positions of listed accessions and Col-0 in the histograms are marked by arrowheads.AU - arbitrary units.

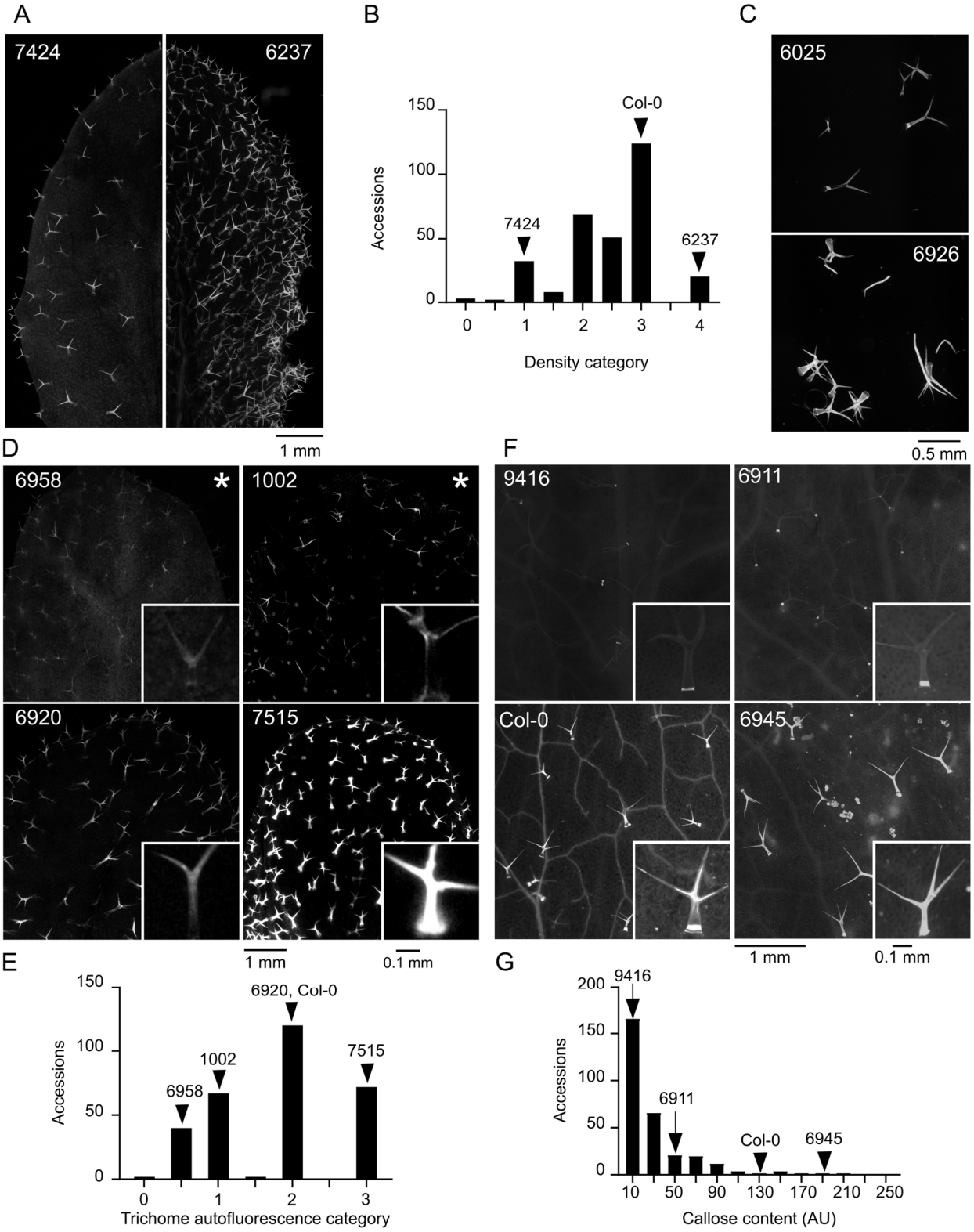


**Figure 2. Variability of trichome density, branch number, autofluorescence and callose content.** (A) Representative leaf photos of the indicated ecotypes with low (7424, JI-3) and high (6237, TOM 03) trichome density. (B) Histogram showing distribution of trichome density categories. (C) Typical isolated trichomes of an accession with low (6025, Gro-3)

and high (6926, Kin-0) branch numbers. (D) Detailed view of representative leaves of ecotypes with different levels of trichome cell wall autofluorescence. (6958, Ra-0; 1002, Ale-Stenar-64-24; 6920, Got-22; 7515, RRS-10), with close-ups of single typical trichomes. Ecotypes marked by asterisks also represent typical examples of low/absent (1002, Ale-Stenar-64-24) and high (6958, Ra-0) epidermal autofluorescence outside trichomes. (E) Histogram showing distribution of trichome autofluorescence categories. (F) Detailed view of representative leaves of ecotypes with different levels of callose deposition in trichomes (9416, Ktu-3; 6911, Cvi-0; 6909, Col-0; 6945, Nok-3), with close-ups of a single typical trichome. (G) Histogram showing distribution of callose content. Positions of listed accessions and Col-0 in the histograms are marked by arrowheads.

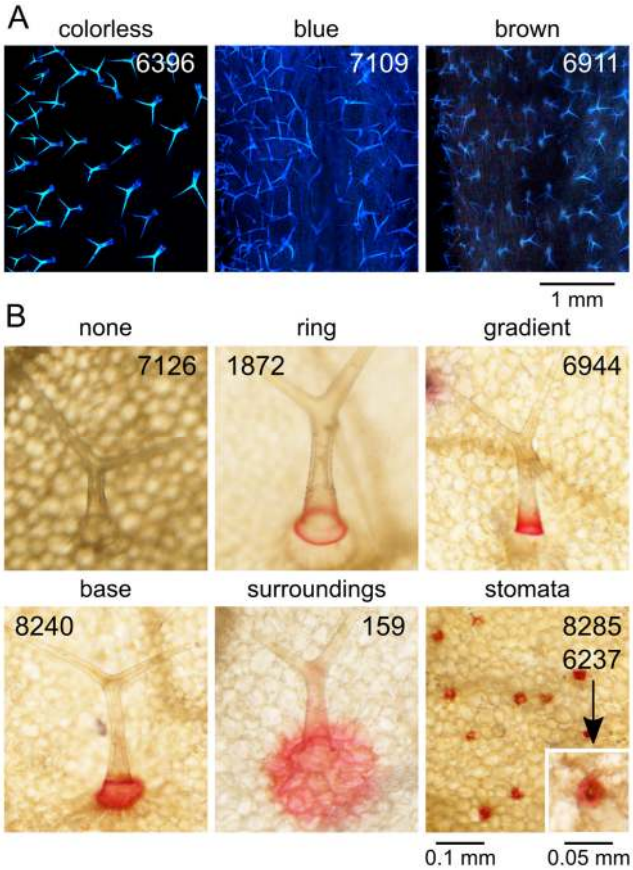


**Figure 3. Examples of autofluorescence colors and patterns of trichome and epidermal metal deposition.** (A) Representative examples of epidermal autofluorescence colors recorded as “colorless” (no background fluorescence, whitish trichomes - 6396, Udul 4-9) and “colored” (blue - 7109, Ema-1; brownish - 6911, Cvi-0). Contrast of the photographs has been uniformly adjusted using ImageJ´s “auto” settings. (B) Images of representative trichomes or epidermis areas for each pattern of metal deposition observed in the screen. Presented pattern is not necessarily the most typical or the only one found for the ecotype shown (none - 7126, Es-0; ring - 1872, MNF-Pot-75; gradient - 6944, NFA-8; base - 8240, Kulturen-1; surround - 159, MAR2-3; stomata - 8285, DraIII-1; stomata close-up - 6237, TOM 03).

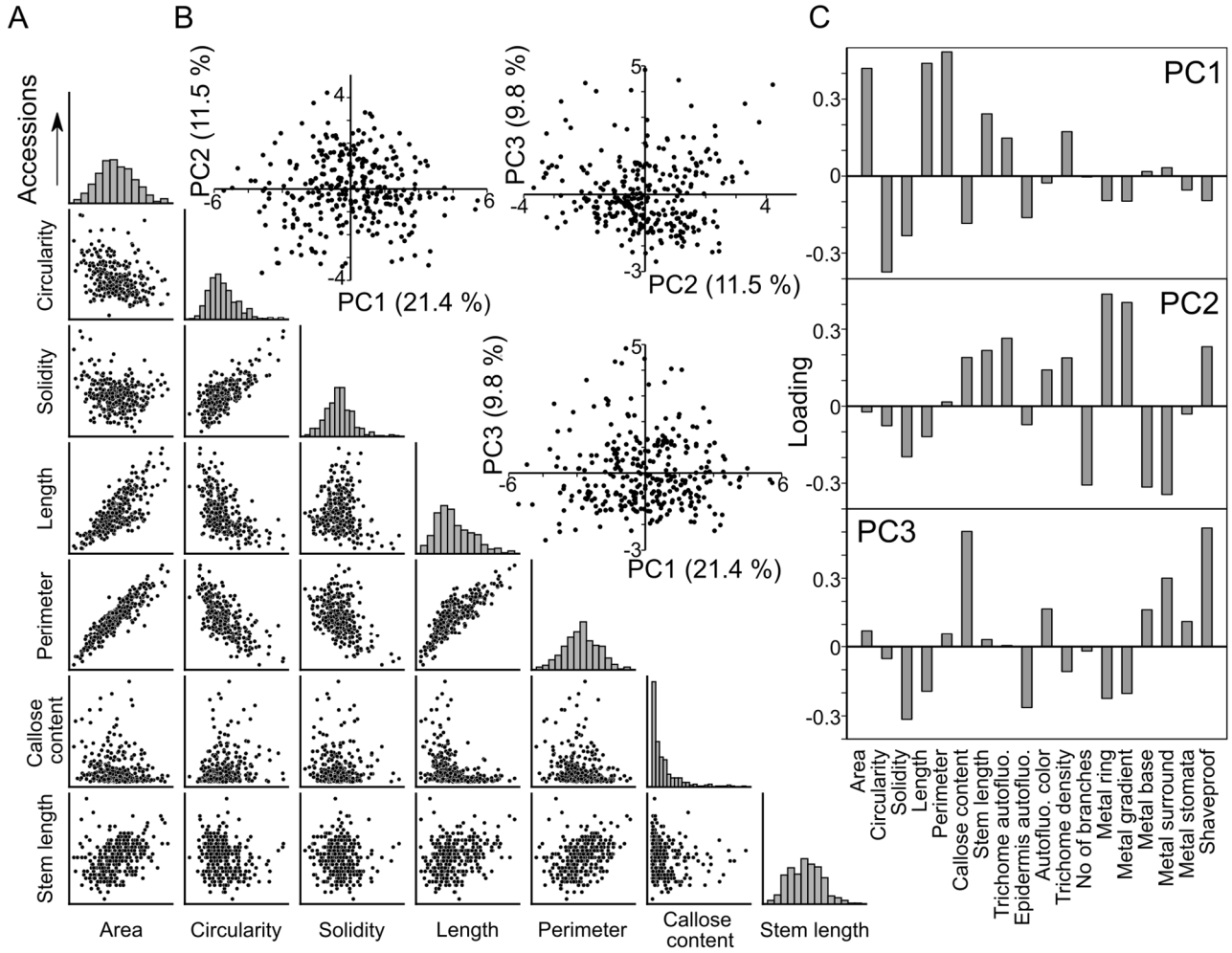


**Figure 4. Distribution of epidermal phenotypic variability among accessions.** (A) Mutual correlation of the 7 continuous-value quantitative parameters analyzed. Distribution of values is shown for each parameter at the diagonal of the matrix (scales are arbitrary, for quantitative values see Figure 1B, 1D, 1F and Figure 2G). (B) Scatterplot of PCA results for the complete epidermal phenotype dataset, with fraction of variance attributable to each PC shown in parentheses. (C) Loadings of the first three principal components.

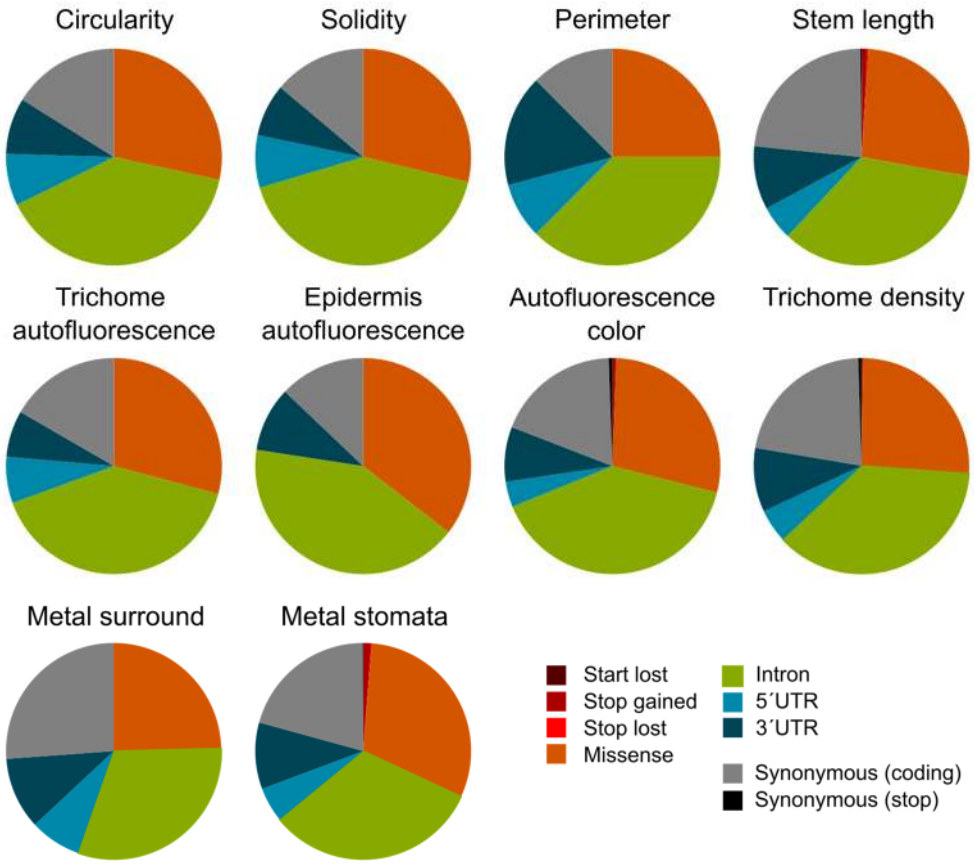


**Figure 5. Distribution of SNP types identified as significantly associated with individual parameters**. Only traits with detected polymorphisms in more than 5 loci are shown.

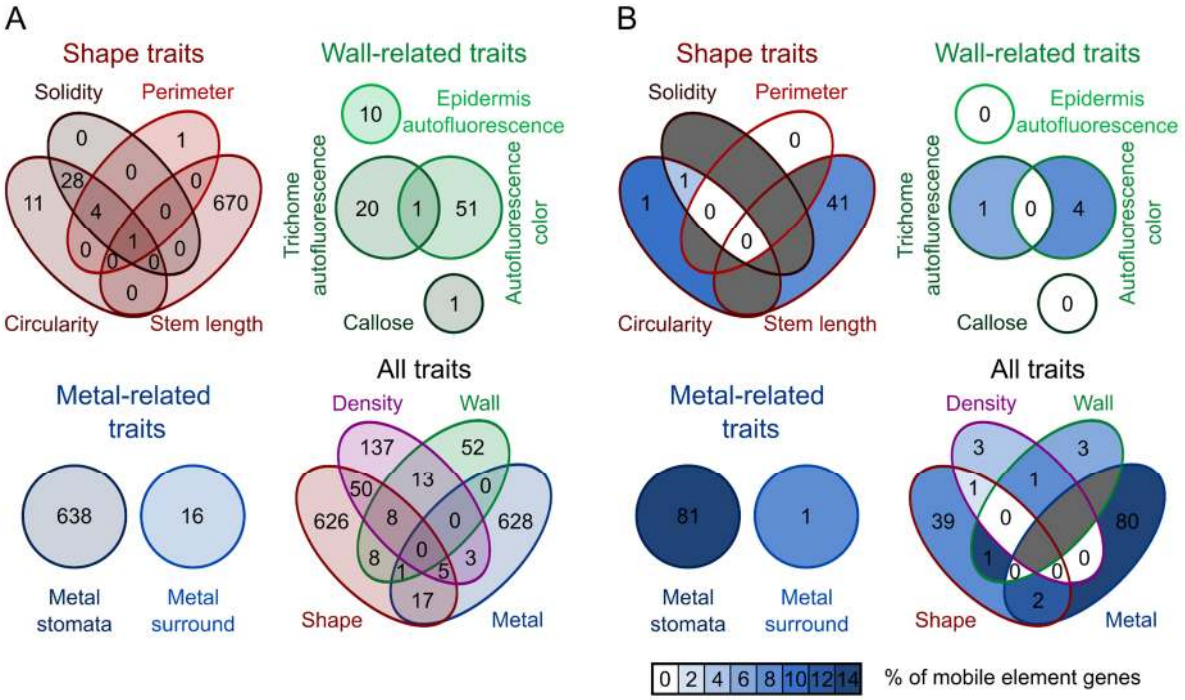


**Figure 6. Unique and shared SNPs among traits and trait groups.** Venn diagrams showing numbers of loci affected by ORF-changing SNPs associated with individual traits and trait groups and/or shared by multiple traits or trait groups. (A) All ORF-changing SNPs.. (B) ORF-changing SNPs in loci contained within transposons. Fields are color-coded according to proportion of transposon-derived loci among all loci detected for the given trait combination. Dark gray fields indicate trait combinations with no associated SNPs detected.

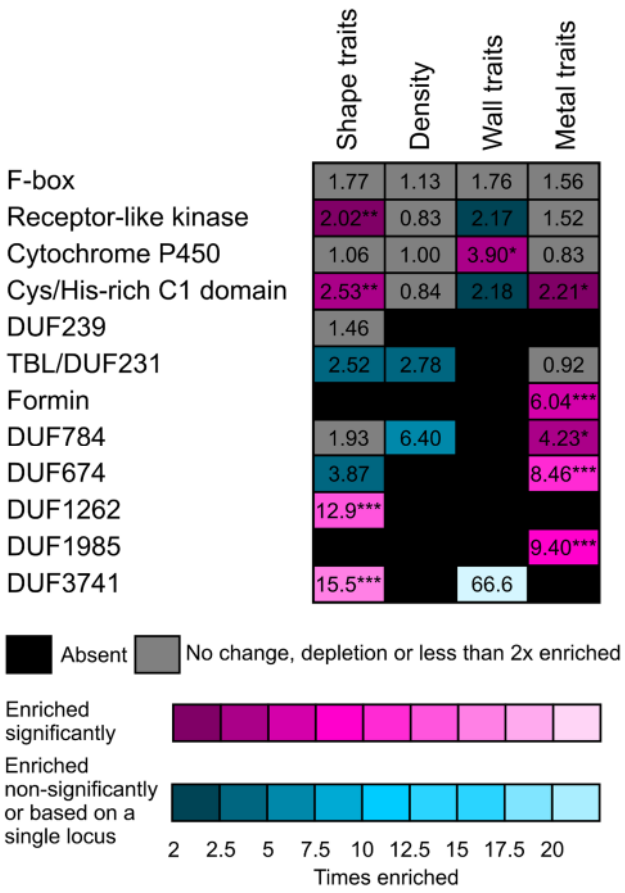


**Figure 7. Summary of gene family enrichment analysis for selected gene families.** The heatmap shows results of enrichment analysis for selected large gene families and all DUF (domain of unknown function) families that were represented by at least two candidates associated with at least one trait group. Numbers correspond to linear fold enrichment compared with the proportion of the family in question among all Arabidopsis loci (Araport11 genome annotation). Asterisks denote significant deviation from the proportion of the family in the whole genome (determined only for enrichment values of at least 2 using the Chi-square test with Benjamini-Hochberg correction for multiplicity; *** P<=0.001; ** 0.001<P<=0.01, * 0.01<P<=0.05).

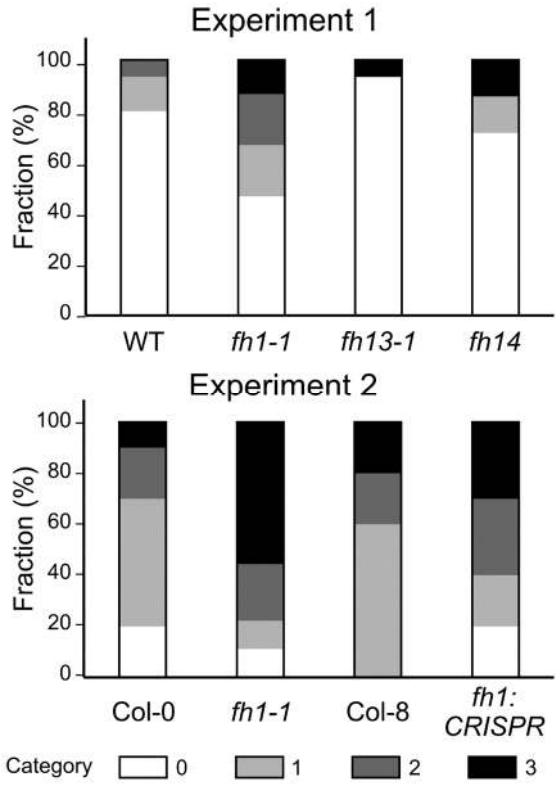


**Figure 8. Stomatal metal deposition patterns in selected formin mutants and wt plants.** Data from two independent experiments, involving plants grown at different times under imperfect temperature and humidity control, are shown, each of them based on single-blinded evaluation of metal deposition patterns in 9-15 plants per genotype. The wild type plants from experiment 1 were WT segregants from the Col-0 derived T-DNA mutant population that was the source of our *fh1-1*, *fh13-1* and *fh14* lines, while in the second experiment, *fh1-1* was compared to Col-0 and the *fh1:CRISPR* mutant to its progenitor genotype Col-8. While the between-phenotype differences of interest were not statistically significant (Fisher-Freeman-Halton test $P > 0.1$), there is a noticeable trend towards increased metal deposition in *fh1* loss of function mutants.

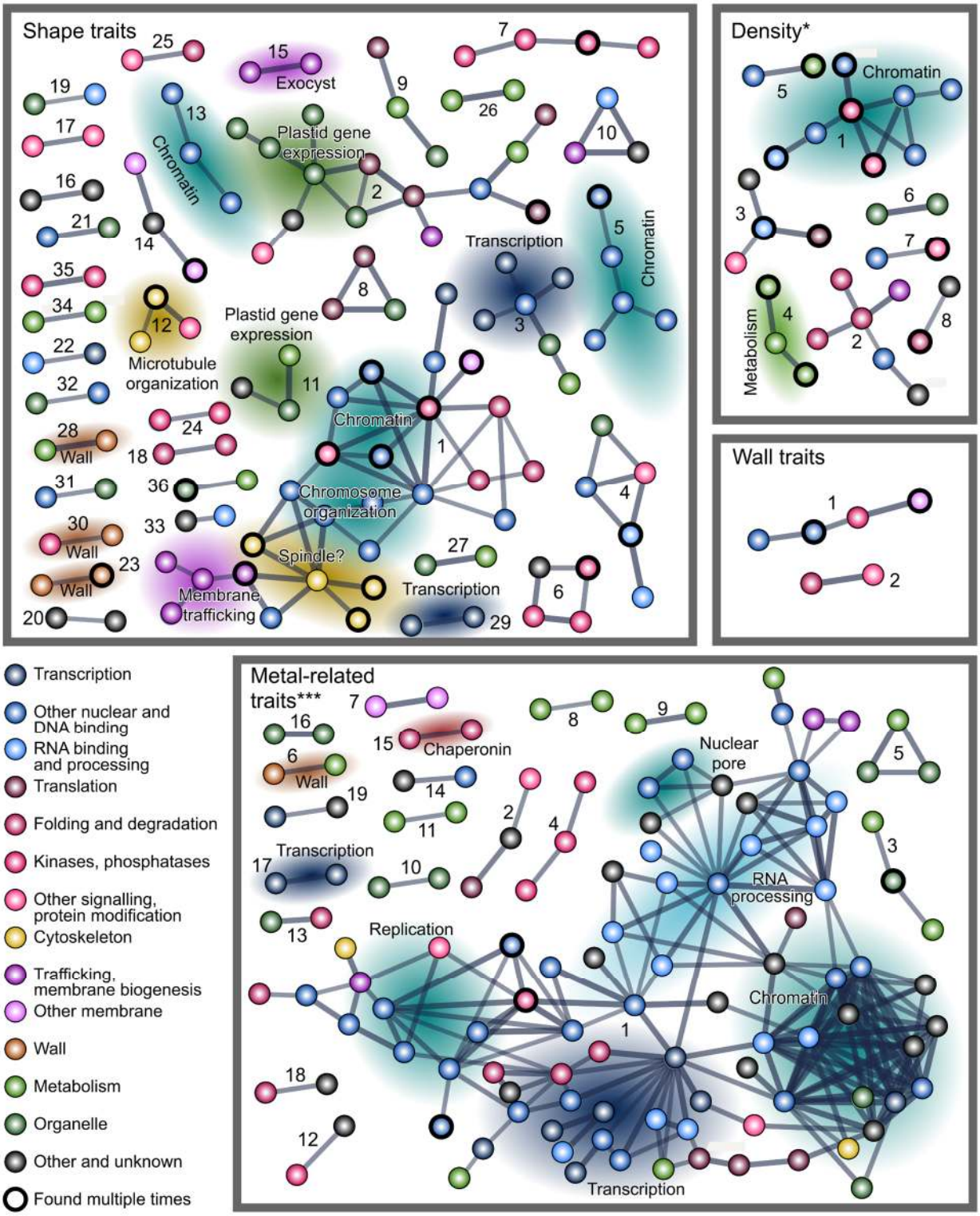


**Figure 9. A schematic map of predicted interactions among candidate genes for individual trait groups.** Thickness of graph edges reflects strength of evidence. Numbers correspond to the numbering of gene clusters in Supplementary Table S9. Asterisks denote significant enrichment of interactions compared to a random gene selection, as determined by the STRING algorithm (*** P<=0.001; * 0.01<P<=0.05).

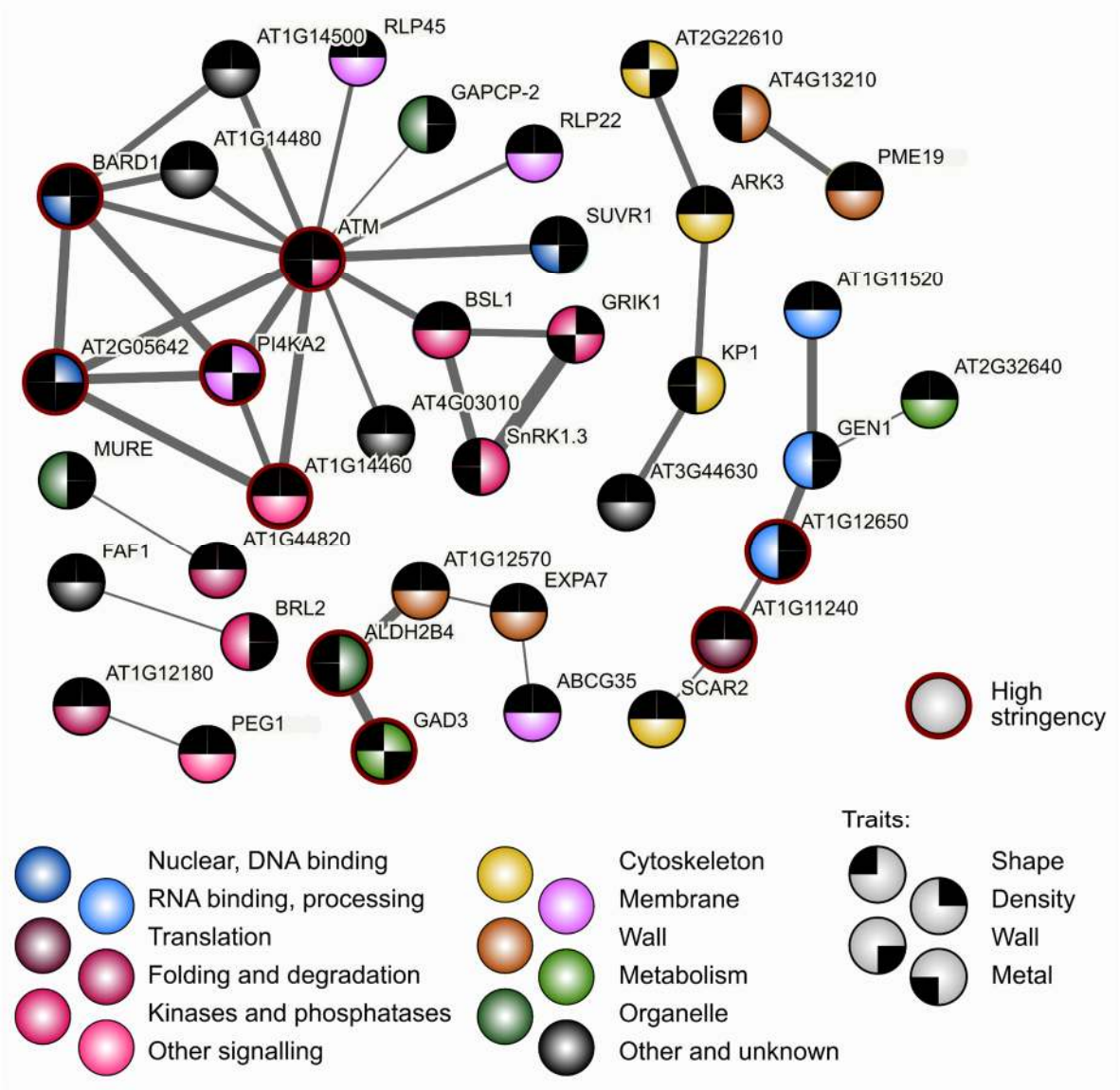


**Figure 10. Predicted interactions among candidate genes linked to two or more trait groups.** Black segments at graph nodes indicate the trait groups the gene was associated with. Note that the interactions were detected using lower stringency settings than those shown in Figure 9, but a part of the network was found also at high stringency settings (denoted by the red outline of participating nodes). Thickness of graph edges reflects strength of evidence.

**Supplementary tables** (MS Excel *.xlsx file).

**Table S1. List of *A. thaliana* accessions included in this study.** Accession numbers refer to the 1001 Genomes collection (1001 Genomes Consortium, 2016). Geographical coordinates of origin are listed for each accession.

**Table S2. Individual plant data for a subset of traits and accessions.** These data were used for replicability and heritability estimation.

**Table S3. Broad sense heritability of selected traits.** The values shown were calculated from data obtained in an independent rescreening of a subset of parameters (see Table S2 Materials and Methods).

**Table S4. List of all significant SNPs detected in this study.**

**Table S5**. **Annotated list of all loci containing ORF-changing SNPs significantly associated with individual traits.** Separate spreadsheets are provided for individual parameters.

**Table S6. Annotated lists of all loci with ORF-changing SNPs significantly associated with multiple traits and trait groups.** Separate spreadsheets are provided for individual trait combinations.

**Table S7. Candidate loci whose annotation suggests a possible role in known processes affecting the relevant traits.** Processes involved include, e.g., cell expansion, cell wall biogenesis, tissue patterning or metal accumulation. Only loci not identified by gene expression patterns (see Table 4 and Table 5) are listed.

**Table S8**. **Numbers and lists of candidates from selected gene families.** Known large gene families and all DUF (domain of unknown function) families that were represented by at least two loci associated with at least one of the studied trait groups are listed, together with results of enrichment analysis including statistical evaluation.

**Table S9. An annotated list of mutually associated gene clusters.** Clusters and genes participating in them are listed together with results of interaction enrichment analysis including statistical evaluation.

**Supplementary figures**

**Figure S1. Mutual correlation of all epidermal phenotype parameters analyzed.** Distribution of values for each parameter is shown at the diagonal of the matrix, with accession counts on the Y axis. Spearman's correlation coefficients (Rho), shown in the upper right triangle for all parameter pairs, are color-coded: gray for negligible correlation ($-0.1 < Rho < 0.1$), magenta for positive, cyan for negative correlation, with intensity of coloring reflecting correlation strength, while P-values for these correlations, shown in the lower left triangle, are color-coded from green (highly significant, $0 < P \leq 0.01$ with darker color corresponding to lower P) through yellow (significant, $0.01 < P \leq 0.05$) to orange ($P > 0.05$, not significant).

**Figure S2. A detailed map of predicted associations among candidate genes for individual trait groups.** Thickness of graph edges reflects strength of evidence. Numbers correspond to the numbering of gene clusters in Supplementary Table S9.

**Figure S3. Inter-observer replicability of selected parameters estimation**. For continuous variables, results from two independent screens of the same images are plotted against each other (X-axis values correspond to averages of three plants). For categorical ordinate variables, the replicability values correspond to the percentage of accessions where results for at least two out the three plants differed by no more than 1 between two independent visual screens of the same images.