Single molecule Lipid Biosensors Mitigate Inhibition of Endogenous Effector Proteins

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Genetically encoded lipid biosensors are essential cell biological tools. They are the only technique that provide real time, spatially resolved kinetic data for lipid dynamics in living cells. Despite clear strengths, these tools also carry significant drawbacks; most notably, lipid molecules bound to biosensors cannot engage with their effectors, causing inhibition. Here, we show that although PI 3-kinase (PI3K)-mediated activation of Akt is not significantly reduced in a cell population transfected with a PH-Akt1 PIP_3/$PI(3,4)P_2$ biosensor, single cells expressing the PH-Akt at visible levels (used for live-cell imaging) have no activated Akt at all. Tagging endogenous AKT1 with neonGreen at its genomic locus reveals its EGF-mediated translocation to the plasma membrane, accumulating at densities of ~0.3 molecules/μm^2. Co-transfection with the PH-Akt biosensor or other PIP_3 biosensors completely blocks this translocation, despite robust recruitment of the biosensors. A partial inhibition is even observed with $PI(3,4)P_2$-selective biosensor. However, we found that expressing lipid biosensors at low levels, comparable with those of endogenous AKT, produced no such inhibition. Helpfully, these single-molecule biosensors revealed improved dynamic range and kinetic fidelity compared with over-expressed biosensor. This approach represents a less invasive way to probe spatiotemporal dynamics of the PI3K pathway in living cells.

Introduction

Genetically encoded lipid biosensors are a transformative tool for studying the role of specific lipids in cellular physiology (Yang et al., 2018; Wills et al., 2018). They are uniquely able to report both the organellar location of cytosolic leaflet-localized lipids, as well as their dynamic changes in intact, living cells. Although a mainstay of experimental techniques for interrogating lipid function, these biosensors, like all experimental tools, have specific drawbacks and limitations. One of the most prominent is the fact that lipid engagement by a biosensor occludes the lipid's headgroup, blocking its interaction with proteins that mediate biological function. This suggests that large fractions of lipid may be effectively titrated by the biosensor, inhibiting the associated physiology. We have argued that, in most cases, this is unlikely because the total number of lipid molecules outnumbers expressed biosensors by one to two orders of magnitude (Wills et al., 2018). However, for more transiently generated second messenger lipids, total copy numbers of lipids may be in the order of tens to hundreds of thousands, making titration by biosensors a real possibility.

One of the most famous second messenger lipids is phosphatidylinositol 3,4,5-trisphosphate (PIP_3). Generated by class I phosphoinositide 3-OH kinases (PI3Ks), PIP_3 activates numerous effector proteins involved with stimulating cell metabolism, migration, growth and survival, along with activation of the immune system (Fruman et al., 2017; Vanhaesebroeck et al., 2021; Madsen and Toker, 2023). Its best-known effector is the serine-threonine kinase, AKT (a.k.a. PKB). This protein is recruited to the membrane and allosterically activated by PIP_3 binding to its plekstrin homology (PH) domain, facilitating phosphorylation of the enzyme at threonine 308 and serine 473 (Truebestein et al., 2021; Bae et al., 2022). The selectivity and affinity of the PIP_3 interaction with PH domains of proteins like AKT led to their development as some of the first biosensors for lipid signaling (Venkateswarlu et al., 1998; Watton and Downward, 1999; Várnai et al., 1999; GRAY et al., 1999).

PIP_3 is synthesized by 3-OH phosphorylation of the substrate lipid, $PI(4,5)P_2$, of which cells contain approximately 10 million in their plasma membranes (Wills and Hammond, 2022). Yet only 3-5% of $PI(4,5)P_2$ is converted to PIP_3 (Stephens et al., 1993), resulting in no more than about 500,000 molecules per cell. PIP_3 effector proteins can be predicted based on sequence (defined as an RFC score greater than 10; Park et al., 2008). Inspecting OpenCell proteome data for the number of these binding proteins predicts approximately 442,000 PIP_3 effectors in a HEK293 cell (Cho et al., 2022). Therefore, PIP_3 production is well matched to engage this particular cell type's effector proteins. Translating these numbers into cellular concentrations (assuming 13 pL for a 15 μm spherical HEK293 cell) gives ~500 nM concentration, which is again well-matched by the 100-500 nM dissociation constant of these effector proteins (Wills et al., 2018). However, now consider that an over-expressed PH-domain based biosensor has been measured at up to 10,000 nM (Xu et al., 2003): these domains could easily dominate binding to PIP_3 and thus titrate out

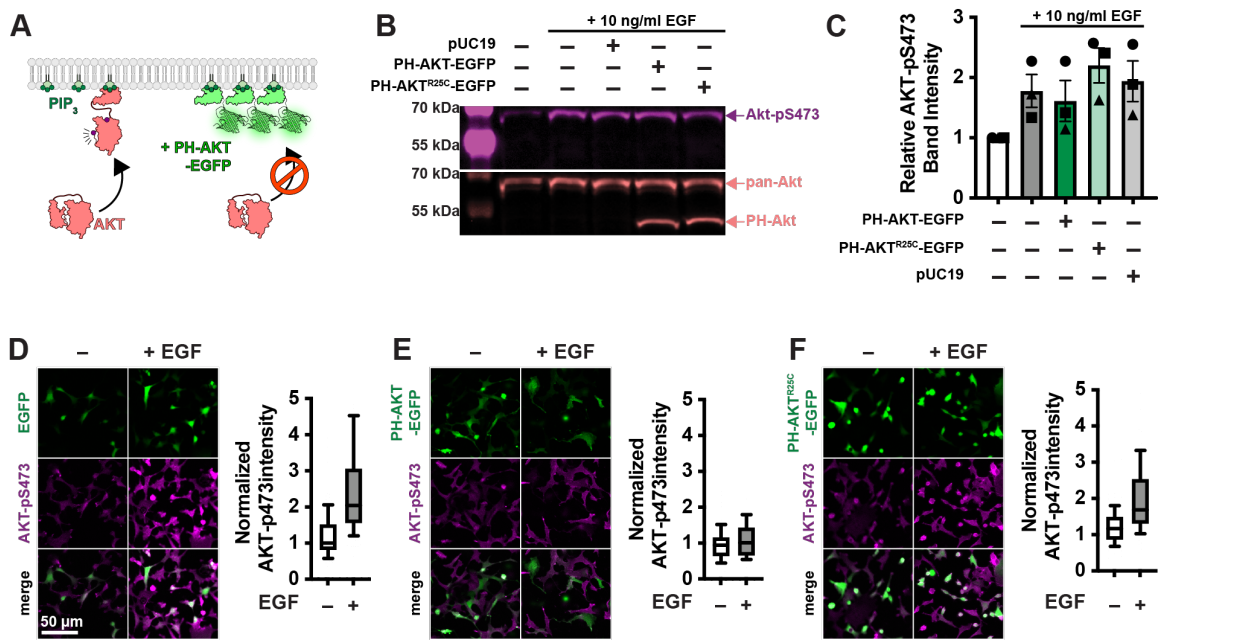

Figure 1: Single cell analysis reveals profound inhibition of Akt by PH-Akt PIP_3 biosensor. (**A**) Akt is autoinhibited by its PIP_3 binding PH domain. PIP_3 production alleviates this steric inhibition, facilitating activating phosphorylations at S473 and T308. Over-expression of PH-Akt is hypothesized to out-compete the endogenous Akt's PH domain. (**B**) Inhibition of Akt S473 phosphorylation is not apparent at the population level. HEK293A cells were serum starved and then treated (where indicated) with 10 ng/ml EGF to activate PI3K. After 5 min, cells were lyzed and analyzed by Western Blot for total Akt and Akt-pS473. (**C**) Quantification of blots from 3 experiments like that shown in B. Data are means ± s.e. (**D-F**) Inhibition of Akt activation (via S473 phosphorylation) is apparent through analysis of single cells. HEK293A cells were stimulated with EGF as in B, then fixed and stained with antibodies against pS473. (**D**) In EGFP control transfected cells, a large increase in pS473 staining induced with EGF is obvious. (**E**) in PH-Akt-EGFP expressing cells, pS473 staining is blocked in EGFP-positive cells. (**F**) in cells transfected with PIP_3-binding deficient PH-Akt^{R25C}-EGFP, EGF still induces increases in pS473. Data in panels D-F are box and whisker plots denoting median and interquartile range (boxes) and 10-90th percentile (whiskers); data are from 82-160 cells pooled from three experiments.

endogenous effector proteins, blocking physiological signaling. Indeed, there have been prior reports of over-expressed PIP_3 biosensors inhibiting PI3K signaling (Várnai et al., 2005). However, in this case it was unclear how much this was due to simple titration of the lipid, versus titration of alternative protein binding partners of the PH domains in a lipid-dependent manner. Furthermore, it remains possible that cells may respond to biosensor PIP_3 titration by increasing PIP_3 production by yet-to-be defined feedback mechanisms. Such feedback has been reported in the case of $PI(4,5)P_2$ biosensors (Traynor-Kaplan et al., 2017).

In this manuscript, we have addressed the question of whether over-expressed biosensors titrate PIP_3 to an extent that effector protein translocation is inhibited. To this end, we generated gene-edited cell lines incorporating a neonGreen fusion protein driven from a native AKT1 allele in HEK293A cells. We show that a variety of PIP_3 effector proteins very efficiently block AKT1 translocation and inhibit signaling, despite robust PIP_3 production. This titration can be eliminated by reducing biosensor concentrations (through reduced transfection times and/or using weaker promoters) to reach approximate concentrations of the endogenous effectors, though this requires single molecule sensitivity. As well as alleviating inhibition of PI3K signaling, single molecule biosensors show improved dynamic range and report more accurate kinetics than their over-expressed counterparts.

Results

Inhibition of AKT activation by PH-AKT1 PIP_3 biosensor.

PIP_3 mediated activation of AKT1 occurs via the initiation of both allosteric and localization-based mechanisms (Truebestein et al., 2021; Bae et al., 2022). Firstly, PIP_3 binding to the AKT PH domain disrupts an autoinhibitory interaction with the kinase activation loop. Secondly, the enzyme is localized to the membrane where it is concentrated with upstream activating kinases PDK1 and TORC2, which phosphorylate and activate the enzyme at residues T308 and S473, respectively (**Fig 1A**). Both mechanisms critically depend on PIP_3, so we reasoned that high concentrations of PIP_3 biosensor might disrupt AKT1 activation by titrating out the lipid.

To test for such an effect, we overexpressed the most commonly utilized PIP_3 biosensor, the isolated PH domain of AKT1. As a control, we used the

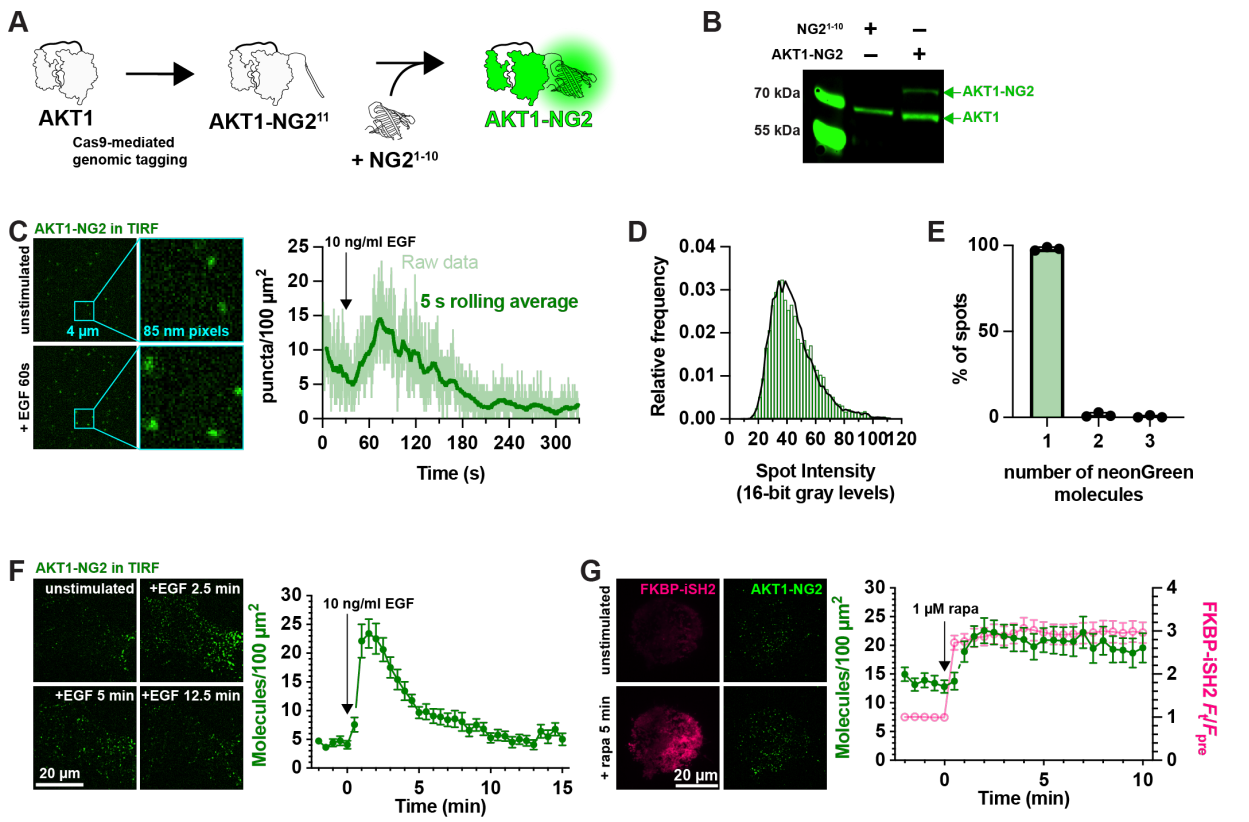

Figure 2: fluorescent tagging of AKT1 at its genomic locus. (**A**) CRISPR/Cas9 directed cutting at the 3' end of the AKT1 ORF is coupled to homology-directed repair to integrate an in-frame NG2[11] tag, encoding the 11th strand of neonGreen2. The edit is made in a HEK293A cell line stably expressing NG2[1-10], the remainder of the neonGreen protein. The two neonGreen2 protein fragments assemble in the cell to generate fluorescent NG2 protein. (**B**) Western blot of AKT protein from a sorted, polyclonal population of edited cells shows the appearance of a second molecular weight band, consistent with the 81.9 kDa complex between AKT1 (55.7 kDa) and NG2 (26.2 kDa). (**C**) TIRF imaging of AKT1-NG2 cells exhibit discrete fluorescent spots at the cell surface, which increase in density after PI3K activation with 10 ng/ml EGF. The light lines are data points from images recorded at 20 Hz; the thicker, darker line is the 5 s rolling average. (**D-E**) AKT1-NG2 spots are single molecules: (**D**) The intensity of fluorescent AKT1 spots pooled from 2 cells from a representative experiment shows a mono-modal lognormal distribution (green). The data are fit with a model assuming the intensity is derived from a mixture of monomeric, dimeric or trimeric fluorescent proteins calibrated against a known monomeric mNeonGreen fluorescent protein distribution. The fit predicts 98.1% monomers with a reduced χ^2 of 1.06. (**E**). The results of this analysis pooled across three experiments yields consistent results with mean χ^2 of 1.12 ± 0.22 (s.e.) (**F**). Extended TIRF imaging of AKT-NG2 cells with reduced duty cycle to minimize photobleaching reveals robust recruitment of AKT1 after EGF stimulation. Data are means ± s.e. of 20 cells pooled from two independent experiments. (**G**) PI3K activation is sufficient to recruit AKT1 to the plasma membrane. AKT1-NG2 cells were transfected with PM-targeted Lyn N-terminal 11 residues fused to FRB and the PI3K p110 catalytic subunit-binding iSH2 fused to FKBP and mCherry. 1 μM rapamycin was added to cells to induce dimerization of FRB and FKBP and hence recruitment of iSH2/p110 to the plasma membrane, inducing PIP_3 synthesis. AKT1-NG2 is further increased on the PM by this maneuver. Data are means ± s.e. of 32 cells pooled from three independent experiments.

non PIP_3-binding mutant R25C (Watton and Downward, 1999). The most commonly employed method to assay AKT activation is by immunoblotting for the activated phosphorylated species. Therefore, we transiently transfected HEK293A cells with EGFP control, PH-AKT1-EGFP or the R25C mutant, then stimulated them for 5 minutes with a moderate experimental dose of epidermal growth factor (EGF; 10 ng/ml). Blotting for pS473 revealed no drastic inhibition of pS473 levels by PH-AKT1 expression (**Fig 1B & C**). However, since transient transfection produces expression in a subset of cells, with those cells displaying wide a distribution of expression levels, we reasoned that analysis of the whole cell population might be misleading, since this includes transfected cells or cells with low expression of the biosensor that would not be selected for imaging. We therefore took a single cell approach, immunostaining the cells for pS473 after stimulation, and quantifying the staining intensity in EGFP positive cells. As shown in **Fig. 1D**, EGFP positive cells showed a large increased in pS473 staining in response to EGF. However, cells positive for PH-AKT1-EGFP showed a complete block in pS473 (**Fig. 1E**). In the non PIP_3 binding PH-AKT1[R25C]-EGFP positive cells, we still observed an increase in pS473 intensity, though it was somewhat

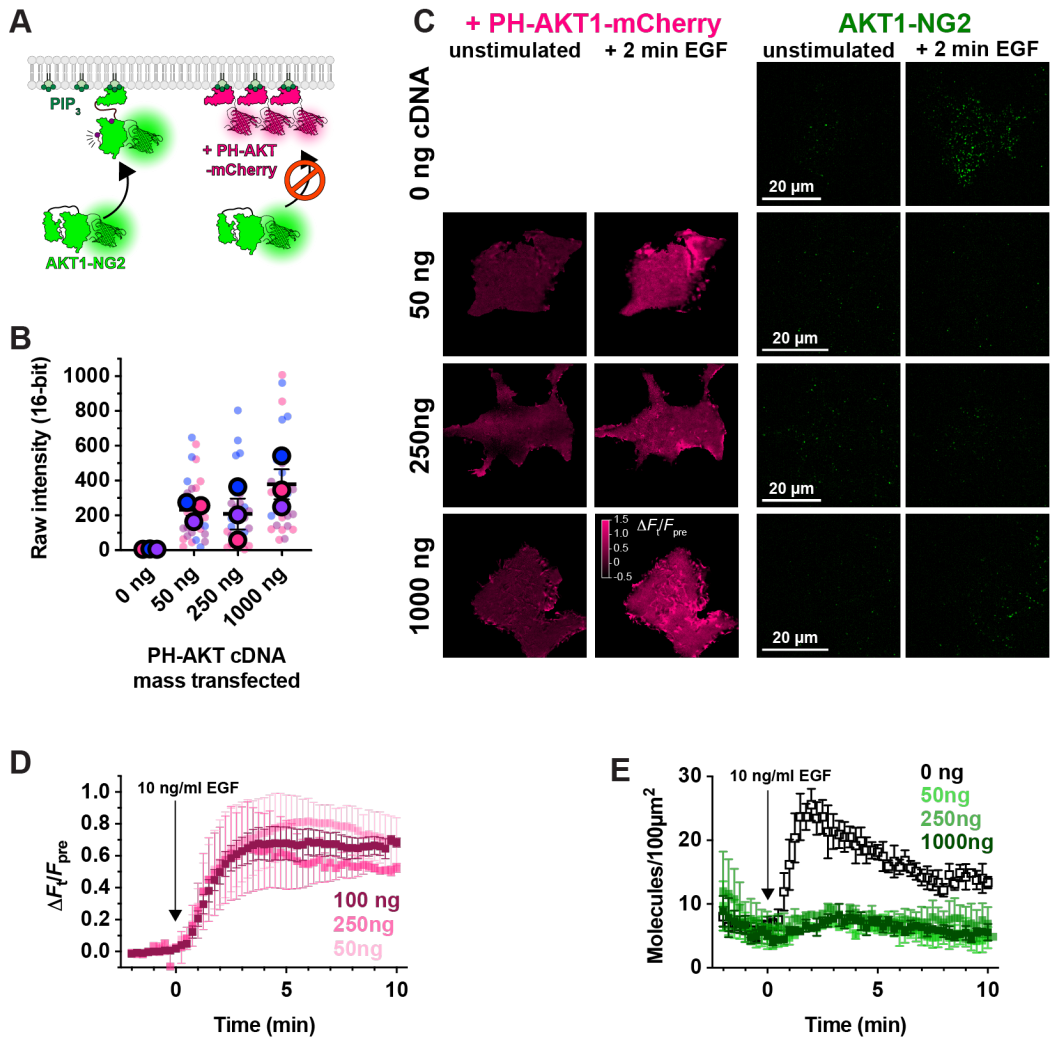

Figure 3: PH-AKT1 PIP_3 biosensor abolishes endogenous AKT recruitment to the PM. (**A**) Hypothesized competition for PIP_3 between the overexpressed PH domain and endogenous AKT1-NG2. (**B**) Raw 16-bit intensity levels of mCherry fluorescence in HEK293A cells transiently transfected with the indicated mass of PH-AKT1-mCherry for 24 hours. Small data points represent measurements of individual cells, whereas large points are the means of each of three independent experiments. Points are color matched by experiment. Grand means ± s.e. are also indicated. (**C**) TIRF imaging of AKT1-NG2 cells transiently transfected as described in (B) and stimulated with 10 ng/ml EGF. (**D**) Relative PM fluorescence of PH-AKT1-mCherry during time lapse imaging and stimulation with 10 ng/ml EGF. (**E**). As in (D), except the density of endogenous AKT1-NG2 molecules are counted. Data in (D) and (E) are grand means of three experiments ± s.e. imaging 8-10 cell each.

reduced compared to EGFP. Together, these experiments showed that at the population level, PH-AKT1 over-expression has little impact, but at the level of individual cells expressing high levels of PH-AKT1 (that would be used for imaging experiments), inhibition of AKT activation was profound.

Genomic tagging of AKT1 as an assay for PIP_3 interactions in cells

Whilst inhibition of AKT1 S473 phosphorylation by PH-AKT1 over-expression is consistent with titration of that target lipid, it is not a direct demonstration of the phenomenon. We therefore wanted to devise an experiment where we could quantify competition for PIP_3 between the biosensor and endogenous AKT1. To that end, we used gene editing to incorporate a bright, photostable neonGreen tag to the C-terminus of AKT1 via gene editing using a split fluorescent protein approach (Kamiyama et al., 2016). Here, CRISPR/Cas9 is used to cut the 3' end of the AKT1 open reading frame. Homology-directed repair incorporates the 11th strand of the neonGreen2 protein. This is performed in cells stably expressing the other neonGreen2-1-10 fragment, leading to complementation and fluorescence (**Fig. 2A**). Western blotting with AKT1-specific antibodies revealed the presence of the neonGreen2 fusion in edited cells but not parental controls (**Fig. 2B**).

Imaging these cells by total internal reflection fluorescence microscopy (TIRFM) revealed diffraction limited spots at the cell surface, which increased in

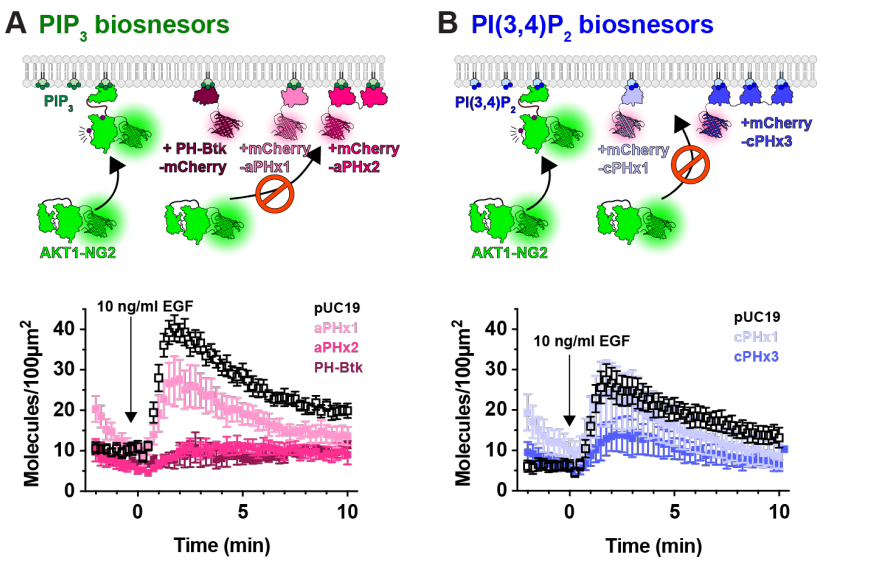

Figure 4: PIP_3 and $PI(3,4)P_2$ biosensors inhibit endogenous AKT recruitment to the PM. (**A**) Hypothesized competition for PIP_3 between PIP_3 biosensors and endogenous AKT1-NG2. AKT-NG2 cells were transiently transfected with the indicated PIP_3 biosensors, or pUC19 as an inert control, and imaged after 24 h by TIRFM. Data quantify AKT-NG2 density at the cell surface, are grand means ± s.e. of 3-4 independent experiments imaging 9-10 cells each. (**B**) As in (A), except high and low avidity versions of a $PI(3,4)P_2$-selective biosensor were tested. Data are grand means ± s.e. of 3-5 independent experiments imaging 6-10 cells each.

number with EGF stimulation (**Fig. 2C**). We analyzed the intensity of these spots and calibrated them against intensity distributions from a known monomeric protein localized to the plasma membrane (PM) and expressed at single molecule levels, namely a myristoylated and palmitoylated neonGreen protein (produced by fusion to the N-terminus of Lyn kinase). Fitting a basis histogram to this known distribution allows the AKT1 data to be fit to a function that will differentiate between single, dimeric, trimeric or higher fluorescent molecule intensities in the sample (Mutch et al., 2007), as shown in **Fig. 2D**. This revealed that 98.1% (95% confidence interval from three experiments = 95.6-100.5%) of the AKT1-NG2 spots were single molecules (**Fig. 2E**).

Although we could resolve single molecules of AKT1, video-rate imaging caused significant photobleaching (note the decline in molecule numbers prior to stimulus addition in **Fig. 2C**). To circumvent this, we dropped our imaging rate from an image every 50 ms to one every 30 s. Stimulation with 10 ng/ml EGF then revealed a roughly 5-fold increase in AKT1 localization at the PM within about 90 s, which then declines with first order kinetics over the subsequent 10 minutes of the experiment (**Fig. 2F**). This is consistent with the typical evolution of AKT activation in response to growth factor stimulation. As a further test of PI3K activation, we stimulated PI3K using chemically induced dimerization to recruit the isolated inter-SH2 domain of the PI3K regulatory subunit to the PM. This in turn recruits endogenous, constitutively active p110 catalytic subunits of PI3K, stimulating PIP_3 synthesis (Suh et al., 2006). iSH2 generated a robust and sustained recruitment of AKT1-NG2 molecules to the plasma membrane after recruitment of iSH2 (**Fig. 2G**).

Titration of PIP_3 by lipid biosensors

Having developed the ability to visualize native AKT1 translocation to the PM in response to PI3K activation, we next designed experiments to test for competition of PIP_3 binding by over-expressed PH-AKT1-mCherry biosensor (**Fig. 3A**). We transfected HEK293A cells for 24 hours in 35 mm dishes with three different masses of plasmid (all adjusted to 1 μg total plasmid DNA by the addition of inert pUC19 plasmid carrier). When imaged at the single cell level in TIRFM, this led to a somewhat weak correlation of transfected plasmid mass with single cell mCherry fluorescence intensity (**Fig. 3B**). Nonetheless, when these cells were stimulated with EGF, cells transfected with any dose of PH-AKT1 showed robust recruitment of the biosensor to the PM (**Figs. 3C, D**). However, at all three doses, we observed complete ablation of endogenous AKT1-NG2 translocation to the plasma membrane (**Figs. 3C, E**). Thus, the presence of the biosensor seemed to prevent engagement of endogenous AKT1 with PIP_3.

Prior studies have reported inhibition of AKT signaling by the AKT1 PH domain, though this was attributed to secondary interactions of the PH domain with undefined effectors; it was not observed with other PIP_3 biosensors (Várnai et al., 2005). We therefore tested additional PIP_3 biosensors (**Fig. 4A**). We examined the commonly used Btk PH domain (Várnai et al., 1999), along with the 2G splice variant of the ARNO PH domain (Venkateswarlu et al., 1998), carrying an I303E point mutation to disrupt interactions with Arl-family GTPases (Goulden et al., 2019). This latter construct was expressed as a high-avidity tandem dimer (aPHx2), or else as a low-copy monomer (aPHx1) expressed from a truncated CMV promoter (Morita et al., 2012). Both PH-Btk and aPHx2 completely blocked AKT1-NG2 translocation, whereas the lower-affinity aPHx1 construct reduced AKT1-NG2 translocation by around 50% (**Fig. 4A**). Therefore, titration of PIP_3 occurred with a variety of PIP_3 biosensors.

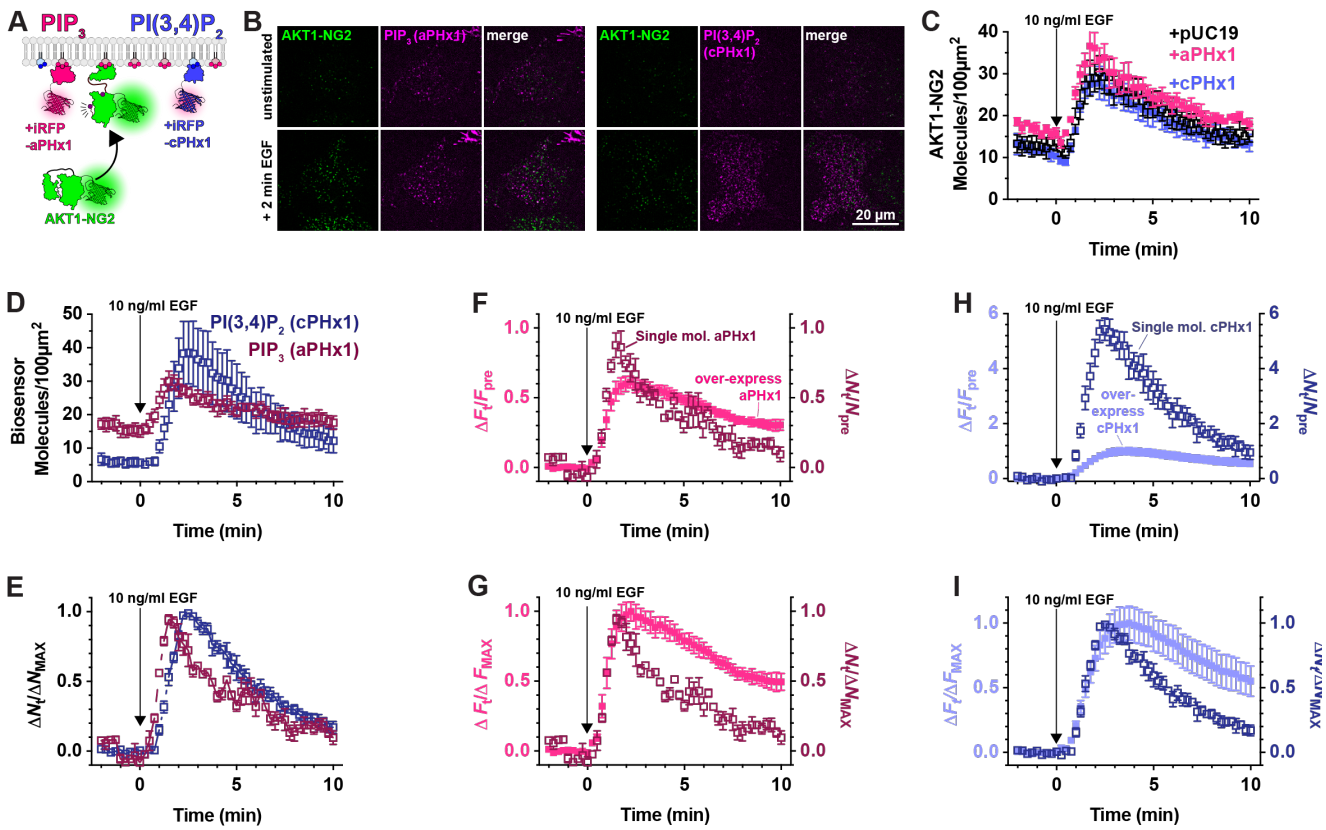

Figure 5: Weak expression of PIP_3 and $PI(3,4)P_2$ biosensors does not inhibit AKT1 translocation, and reveals improved dynamic range and kinetic fidelity. (**A**) Hypothesized weak expression of biosensors that do not sequester a large fraction of the available lipids. (**B**) Example TIRFM images of AKT1-NG2 cells transiently transfected with aPHx1 PIP_3 or cPHx1 $PI(3,4)P_2$ probes for 2-4 hours before imaging and stimulation with EGF. Note the single molecule expression of the biosensors. (**C**) Quantification of AKT1-NG2 translocation of aPHx1, cPHx1 or pUC19 (control)-transfected cells, showing no effect. (**D**) Quantification of the lipid biosensor density at the cell surface from the same experiments. (**E**) As in D, except data were normalized to the maximum density for each experiment, showing the characteristic lagging accumulation of $PI(3,4)P_2$ vs PIP_3. (**F-I**) Comparison of weakly (single-molecule level) expressed biosensors with the same constructs strongly over-expressed (24 h). (**F**) comparison of the change in fluorescence or molecule density vs baseline for weakly and strongly expressed PIP_3 biosensor, aPHx1. (**G**) as in F, but data are normalized to the maximum for each experiment. (**H**) comparison of the change in fluorescence or molecule density vs baseline for weakly and strongly expressed $PI(3,4)P_2$ biosensor, cPHx1. (**I**) As in I, but data are normalized to the maximum for each experiment. Data are grand means ± s.e. of 3-4 experiments analyzing 9-10 cells each. The data for the over-expressed biosensor were generated from the same experiment as presented in figure 4.

Although PIP_3 is the principle activator of AKT, the enzyme can also be activated by the PIP_3 degradation product, $PI(3,4)P_2$ (Frech et al., 1997; Ebner et al., 2017). We therefore tested whether $PI(3,4)P_2$ biosensors could disrupt AKT1-NG2 translocation. A high avidity tandem trimer of the TAPP1 PH domain, cPHx3, is a sensitive reporter for $PI(3,4)P_2$ production (Goulden et al., 2019), and this construct produced a notable inhibition (but not ablation) of AKT1-NG2 recruitment (**Fig. 4B**). On the other hand, a single PH domain version of the same sensor, cPHx1, expressed from a truncated CMV promoter was without notable effect (**Fig. 4B**). Therefore, PIP_3 biosensors produced a more profound inhibition than $PI(3,4)P_2$ biosensors. This is not surprising, since complete $PI(3,4)P_2$ titration would not effect the ability of AKT1-NG2 to engage PIP_3, whereas PIP_3 biosensors might also reduce the ability of 5-OH phosphatases to convert PIP_3 to $PI(3,4)P_2$.

Mitigating PIP_3 titration using single molecule biosensors

The titration of native PIP_3 by over-expressed biosensors is not surprising when comparing the fluorescence intensity of endogenously labelled AKT1 with the biosensors. AKT1-NG2 expressed from a native allele produces fluorescence that is resolved as single molecules (**Fig. 2D, E**) and achieves densities of 25-35 molecules per 100 μm^2. On the other hand, PH-AKT1-mCherry produces an intense, even labelling of the PM when viewed in TIRFM (e.g. **Fig. 3C**). This profile is produced by the high densities of biosensor molecules, which cannot be resolved at the diffraction limit and instead convolve their fluorescence into a monolithic haze. Given an estimated Airy disc of an mCherry molecule of 210 nm diameter, or 0.035 μm^2 through our 1.45 NA optics, a minimum density to achieve such convolution would be around 3000

molecules per 100 μm^2. Therefore, the biosensor molecules clearly outnumber the endogenous effectors by at least two orders of magnitude. We then reasoned that if we dropped biosensor expression levels to be comparable with native AKT1, titration of PIP_3 might be prevented (**Fig. 5A**).

To accomplish this goal, we expressed aPHx1 and cPHx1 PIP_3 and $PI(3,4)P_2$ biosensors tagged with iRFP670 from truncated CMV promoters with less than 4 hours between transfection and imaging. Under these conditions, we selected cells with small numbers of individual fluorescent puncta with intensities consistent with single molecules (**Fig. 5B**). Typically, these cells exhibited 5-20 molecules per 100 μm^2 of PM prior to stimulation. Crucially, in AKT1-NG2 cells expressing these biosensors, we saw no inhibition of AKT1 translocation after EGF stimulation (**Fig. 5C**). In these experiments, both aPHx1 and cPHx1 showed robust translocation to the membrane, similar to AKT1-NG2 (**Fig. 5D**). Normalizing the data to the maximum response revealed a rapid onset of the signal followed by a first-order decay over about 10 minutes (**Fig. 5E**). The recruitment also revealed the lagging kinetics of the $PI(3,4)P_2$ reporter, owing to the synthesis of this lipid from PIP_3 (Hawkins et al., 1992).

The fact that single-molecule PIP_3 and $PI(3,4)P_2$ biosensors do not inhibit PI3K signaling justifies a use case for this approach. However, comparing performance with their over-expressed counterparts revealed additional benefits. Normalizing the aPHx1 PIP_3 sensor count to fold increase over baseline allows comparison with the strongly expressed aPHx1 data (from the experiments introduced in Fig. 4): this showed that single molecule biosensor has substantially better dynamic range, with a fold increase over baseline of nearly 100%, compared to 60% for the same probe over-expressed (**Fig. 5F**). Normalizing data from both expression modes to their maximum response (**Fig. 5G**) reveals the on-rate is remarkably similar. This is expected, since this phase is driven by synthesis of PIP_3 molecules that recruit biosensor. On the other hand, the off-rate occurs as PIP_3 degradation takes over; this is substantially slowed in the case of the over-expressed biosensor. Presumably, this is due to competition with PTEN and 5-OH phosphatases that degrade PIP_3. An even more exaggerated result is achieved with the cPHx1 $PI(3,4)P_2$ biosensor; this shows an increase in fold-change over baseline of 600% for single molecule expression levels, compared to only 100% in over-expressed cells (**Fig. 5H**). Again, the off-rate of the signal is substantially slowed by the over-expressed sensor (**Fig. 5I**). Collectively, these data show that single-molecule based PI3K biosensors show improved dynamic range and kinetic fidelity compared to the same sensors over-expressed.

Discussion

Inhibition of lipid effector protein function by lipid biosensors is one of the most commonly raised concerns with these probes (Balla et al., 2000; Maekawa and Fairn, 2014; Wills et al., 2018). Indeed, over-expression of lipid binding domains has been shown to inhibit lipid functions of PI3P, $PI(4,5)P_2$ and PIP_3 (Simonsen et al., 1998; Balla and Várnai, 2002; Várnai et al., 2005). What is not clear is whether this is due to titration of the lipid itself. An alternative is titration of a tertiary co-interactor by the lipid:biosensor complex, as for example observed between cytohesin family PH domains, PIP_3 and Arf family small GTPases (Cohen et al., 2007; Li et al., 2007; Hofmann et al., 2007). Notably, titration of a co-interactor would not necessarily prevent lipid-dependent localization of an endogenous lipid effector protein, whereas titration of the lipid itself would. Here, we unambiguously demonstrate direct titration of PIP_3 by lipid biosensors, preventing membrane localization of a key effector protein, AKT1 (**Figs. 3** and **4**) and leading to inhibition of AKT activation in cells (**Fig. 1**). Notably, we observed PIP_3 titration by three different PIP_3 binding modules, again arguing against co-interactors being critical for the inhibition.

How general a problem is lipid titration by biosensors likely to be? As we have argued here and elsewhere (Wills et al., 2018), this will depend on the bulk levels of the lipids and their effector proteins. Whereas we show here that PIP_3 biosensors outnumber individual effectors by orders of magnitude, this is unlikely to be the case even for a somewhat less scarce but still minor lipid like $PI(4,5)P_2$. HEK293 cells contain $PI(4,5)P_2$ effector proteins of the ERM and MARCKS family that are in the order of micromolar concentration (Cho et al., 2022), and therefore likely much less susceptible to “swamping” by biosensor expressed at comparable levels (Xu et al., 2003). In support of this, we observed that dropping biosensor expression to low levels matching that of AKT1 produced no inhibition of AKT1 recruitment **Fig. 5**). Therefore, standard techniques of transient transfection for strong expression of biosensors are likely only an issue for molecules present at exceptionally low levels, i.e. transiently generated signaling molecules such as PIP_3, and perhaps other scarce phosphoinositides such as PI5P and $PI(3,5)P_2$.

Intriguingly, we report a counter-intuitive finding that weaker expression of the PI3K lipid biosensors actually produces increased sensitivity of the probes (**Fig. 5**). This is almost certainly due to the failure of the lipids to become saturated by biosensor molecules (which causes titration of endogenous effectors). We also observed enhanced kinetic fidelity, perhaps because over-expressed biosensors prevent access of PIP_3 degrading phosphatases or effectors that stimulate negative feedback. Either way, this single molecule approach will permit precise and quantitative analysis of PI3K pathway activation in living cells with greater accuracy.

Materials and Methods

Plasmid	Backbone	Insert	Reference
EGFP	pEGFP-N1	None	
PH-AKT1-EGFP	pEGFP-N1	*AKT1(1-164)*:DPPVAT:*EGFP*	
PH-AKT1^{R25C}-EGFP	pEGFP-N1	*AKT1(1-164)-R25C*:DPPVAT:*EGFP*	
PH-PLCD1-mNeonGreen	pmNeonGreen-N1	*PLCD1(1-170)*:GVGG:*mNeonGreen*	
Lyn^{N11}-FRB-iRFP	piRFP713-N1	*LYN(1-11):MTOR(2021-2113):iRFP713*	
Lyn^{N11}-mNeonGreenx1	pNeonGreen-N1	*LYN(1-11)*:GVGG:*mNeonGreen*	
Lyn^{N11}-mNeonGreenx2		*LYN(1-11)*:GVGG:*mNeonGreen*:SPVAT:*mNeonGreen*	
Lyn^{N11}-mNeonGreenx3		*LYN(1-11)*:GVGG:*mNeonGreen*:SPVAT:*mNeonGreen*:SPVAT:*mNeonGreen*	
mCherry-FKBP-iSH2		*mCherry*: SGLRSRAALG:*FKBP1A(3-108)*: SA[GGSA]$_4$PRAQAS:*Mus musculusPIK3R2(420-615)*	Suh et al., 2006
PH-AKT1-mCherry	pmCherry-N1	*AKT1(1-164)*:DPPVAT:*mCherry*	
PH-Btk-mCherry	pmCherry-N1	*BTK(1-177)*:DPPVAT:*mCherry*	Varnai et al., 1999
NES-iRFP670-aPHx1	pCMVd3-iRFP670-C1	*X.leavis map2k1.L*(32-44):PVAT:*iRFP670*: SGLRSRAQASNSAVDM:*CYTH2i2(252-399)-I303E*	Goulden et al., 2019
NES-mCherry-aPHx2	pCMVd3-mCherry-C1	*X.leavis map2k1.L*(32-44):PVAT:*mCherry*: SGLRSRAQASNSAVDM:*CYTH2i2(252-399)-I303E*: GGGGGATCGGGTGGTGTCGACATG: *CYTH2i2(252-399)-I303E*	Goulden et al., 2019
NES-iRFP670-cPHx1	pCMVd3-iRFP670-C1	*X.leavis map2k1.L*(32-44):PVAT:*iRFP670*: SGLRSRAGGAGAILS: *PLEKHA1(169-329)*	Goulden et al., 2019
NES-mCherry-aPHx3	pCMVd3-mCherry-C1	*X.leavis map2k1.L(32-44)*:PVAT:*mCherry*: SGLRSRAQASNSTWKMSS:*PLEKHA1(169-329)*:GGSGGSGG: *PLEKHA1(169-329)*: GGSGGSGG: *PLEKHA1(169-329)*	Goulden et al., 2019

Table 1: Plasmids used in this study. All genes (except fluorescent proteins) are human unless otherwise stated. Amino acid linkers are indicated with single letter codes.

Cell Culture, Transfection and Gene Editing

HEK293A cells (ThermoFisher R70507) were cultured in low glucose DMEM (ThermoFisher 10567022) supplemented with 10% heat-inactivated fetal bovine serum (ThermoFisher 10438-034), 100 μg/ml streptomycin + 100 units/mL penicillin (ThermoFisher 15140122), and 0.1%(v/v) chemically defined lipid supplement (ThermoFisher 11905031) at 37˚C and 10% atmospheric CO_2 in humidified incubators. Passaging was performed 1:5 by rinsing with PBS and dissociating the cells in TrpLE (ThermoFisher 12604039) and diluting 1:5 in fresh media.

Gene editing was performed following previously published protocols (Kamiyama et al., 2016; Zewe et al., 2018). We used the strategy to combine CRISPR/Cas9-mediated targeting of an allele with homology-directed repair to tag AKT1 at the c-terminus with a split neonGreen variant (NG2-11) in an HEK293A cell line stably over-expressing the remainder of the neonGreen protein (NG2-1-10), as described by the OpenCell project (Cho et al., 2022). The guide RNA protospacer sequence was AGCGGCACGGCCTGAGGCGG (ordered as ThermoFisher custom gRNA) and the homology-directed repair template (HDRT) synthesized as a single stranded "ultramer" primer sequence (IDT) CAGCGAGCGCAGGCCCCACTTCCCCCAGTTCTC CTACTCGGCCAGCGGCACGGCCGGTGGCGGATT GGAAGTTTTGTTTCAAGGTCCAGGAAGTGGTACC GAGCTCAACTTCAAGGAGTGGCAAAAGGCCTTTA CCGATATGATGTGAGGCGGCGGTGGA CTGCGCTGGACGATAGCTTGGAGGGATGGAGAG GCGGCCT. A Neon® electroporation system (ThermoFisher) was used to introduce the components according to the manufacturer's instructions. Essentially, 10 pmol of gRNA was pre-incubated with 10 pmol TruCUT® Cas9 protein V2 in 5 μl buffer R for 20 min before adding 100 pmol of the HDRT. The mixture was then added to 200,000 cells in 5 μl buffer R and electroporated with a single 20 ms, 1500V pulse. Cells were seeded in complete media in 6-well plates and left to recover for 48 hours before screening for fluorescence by confocal microscopy. After estimating the fraction of edited cells at ~1%, the positive cells were sorted by FACS and expanded, resulting in a polyclonal population.

HEK293A cells were seeded in either 6-well plates for immunofluorescence and Western blotting experiments or 35 mm dishes containing 20 mm #1.5 optical glass bottoms (CellVis D35-20-1.5-N) for live cell imaging. Dishes were pre-coated with 10 μg ECL cell attachment matrix (Sigma 08-110) for 60 minutes. Transfection used lipofectamine2000 (ThermoFisher 11668019) according to the manufacturer's instructions. Briefly, 1 μg total DNA was precomplexed with 3 μg lipofectamine 2000 in 0.2 ml Opti-MEM (ThermoFisher 51985091) for ≥ 5 min before adding to the cells in 2 ml media. Plasmids used for transfection are listed in **table 1**. After 4 hours, media was replaced to remove the transfection reagent.

24 h prior to stimulation with EGF, cells were serum starved in Fluorobrite (ThermoFisher A1896702) supplemented with 0.1% chemically defined lipid supplement (ThermoFisher 11905031) and 0.1% BSA.

Western Blotting

Cells in 6-well plates were treated as described in figure legends before lysis in 150 μl ice-cold RIPA buffer including protease (Sigma Millipore 539131-1VL) and phosphatase (Sigma Millipore 524627) inhibitor cocktails. After scraping and clearing at 10,000 *g*, 40 μl lysates were boiled for 5 min in Bolt LDS sample buffer (ThermoFisher B0008) before running on 12% bis-tris gels at 165 V, then transfered to nitrocellulose membrane (Novex LC2000) at 10V. After transfer, the membrane was blocked in Tris-buffered saline (50 mM Tris with 150 mM NaCl) with 0.05% tween-20) containing 1% nonfat dry milk (Cell Signaling Technology 9999S) for 1 hour. Blots were stained with primary antibodies pan AKT clone 40D4 (Cell Signaling Technology 2920), Phospho-AKT (Thr308) Monoclonal Antibody (ThermoFisher 200-301-269), Akt1 (Cell Signaling Technology 2938) or DM1A (ThermoFisher 62204) and secondary antibodies Alexa647 goat anti-rabbit (ThermoFisher A-21245), Alexa555 Goat anti-Mouse IgG1 Secondary Antibody (ThermoFisher A-21127) or Alexa800 Goat anti-Mouse IgG (ThermoFisher A32730).

Immunofluorescence

4 hours post transfection, cells were dissociated and re-seeded at 25% confluence in 40 μl onto 8-well multi-test slides (Electron Microscopy Sciences Hydrophobic PTFE Printed Microscope Slide 8 Round Wells 6 mm, 6342206) pre-coated with 0.8 μg ECL, and returned to the incubator for 24 h. Cells were serum starved 1.5 hours before stimulation with Fluorobrite supplemented with 0.1% chemically defined lipid supplement and 0.1% BSA, before stimulating where indicated with 10 ng/ml EGF (Corning 354052). After 5 minutes of stimulation, cells were fixed by the addition of formaldehyde (Electron Microscopy Sciences 16% Paraformaldehyde Aqueous Solution, EM Grade, Ampoule 10 ML 15710) in PBS to 4% final concentration for 15 minutes at room temperature. Cells were rinsed three times in 50 mM NH_4Cl in PBS before blocking and permeabilization in blocking solution (5% normal goat serum and 0.2% tritonX-100 in PBS) for 30 minutes. Cells were then stained in anti-PhosphoAKT (Thr308), Alexa647 goat anti-rabbit secondary and GFP-booster-Atto488 (Chromotek gba-488). Cells were rinsed with PBS then Millipore water, then mounted in ProLong Diamond (ThermoFisher P36961) before imaging on a Nikon A1R confocal microscope attached to a Nikon TiE inverted stand. Imaging used a 20x plan apochromatic 0.75 NA air immersion objective with the confocal pinhole fully open. Excitation was with 488 nm or 647 nm on a fiber-coupled 4-line excitation LU-NV laser combiner for Atto488/EGFP and Alexa647, respectively. Emission was collected using 500-550 nm or 663-737 nm band passes for each channel on separate line scans.

Live cell Imaging

Imaging was performed on a Nikon TiE inverted microscope stand with motorized TIRF illuminator (Nikon) fiber-coupled to a four line Oxxius laser launch equipped with 405, 488, 561 and 638 nm laser lines. A 100x 1.45 NA plan apochromatic oil-immersion objective was used combined with a 1.5x magnifier. Images were collected on a Hammamatsu Fusion-BT sCMOS camera in ultra-low noise mode with 2x pixel binning. mCherry (561 nm excitation) fluorescence was collected through a dual pass 420-480 & 570-620 nm filter (Chroma), neonGreen and EGFP (488 nm excitation) used a 500-550 nm bandpass filter (Chroma) and iRFP680 (638 nm excitation) used a dual pass 505-550 & 650-850 nm filter (Chroma).

Image analysis

All image analysis was performed using the open access ImageJ implementation, Fiji (Schindelin et al., 2012). For immunofluorescence, background was subtracted based on the modal intensity, which corresponded to the noise in the spaces between cells. For Western blot images, a rolling ball algorithm was used to subtract the somewhat non-uniform background from the gel image, using a ball diameter set to be just larger than the height of the bands. In either case, after background subtraction, regions of interest (ROI) were drawn around individual cells or bands, and mean pixel intensity was measured. A constant ROI size was used for all bands in an individual gel.

For analysis of over-expressed biosensor intensity, a similar approach to that used for immunofluorescence images was employed. For counting single fluorescent molecules, we employed a macro that took a manually-defined 100 μm^2 ROI for each cell footprint, and employed the Fiji plugin ThunderSTORM (Ovesný et al., 2014) to count single molecule localizations. After entering the pixel and camera calibrations to this plugin, we used the default

molecule detection parameters. Data were plotted in Graphpad Prism 9 or later.

Analysis of single molecule intensities

Polydispersity of fluorescent puncta was calculated by deconvolution of intensity distributions (Mutch et al., 2007). The intensity distribution of a population of N fluorescent puncta of unknown polydispersity can be expressed as

$$\rho(x) = \frac{1}{N}\sum_{c=1}^{M} A_c \rho_c(x)$$

Where A_c denotes the number of puncta containing c fluorophores; $\rho c(x)$ denotes the mean intensity distribution of a population of puncta with c fluorophores, and M is the highest number of fluorophores to be found in a single punctum.

The c=1 basis histogram was generated from intensity distributions acquired from in vivo single molecule tracking measurements of a known monomeric reporter (NGx1). Higher order basis histograms were calculated through a transformation of the c=1 histogram by:

$$\rho_c(x) = \rho_1(x/c)d(x/c)$$

Multimer population proportions were calculated by reduced chi-squared fitting of randomly generated intensity distributions derived from basis histograms to experimental intensity distributions.

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