Visually-evoked activity and variable modulation of

auditory responses in the macaque inferior colliculus

Abbreviated title:

How visual cues affect auditory activity in the IC

Meredith N. Schmehl[1-3], Jesse L. Herche[1-3], and Jennifer M. Groh[1-5]

1. Department of Neurobiology, Duke University, Durham, NC 27708

2. Center for Cognitive Neuroscience, Duke University, Durham, NC 27708

3. Duke Institute for Brain Sciences, Duke University, Durham, NC 27708

4. Department of Psychology & Neuroscience, Duke University, Durham, NC 27708

5. Department of Biomedical Engineering, Duke University, Durham, NC 27708

Corresponding Author: Meredith N. Schmehl (meredith.schmehl@duke.edu)

Abstract

How multisensory cues affect processing in early sensory brain areas is not well understood. The inferior colliculus (IC) is an early auditory structure that is visually responsive (Porter et al. 2007; Bulkin and Groh 2012a, 2012b), but little is known about how visual signals affect the IC's auditory representation. We explored how visual cues affect both spiking and local field potential (LFP) activity in the IC of two monkeys performing a task involving saccades to auditory, visual, or combined audiovisual stimuli. We confirm that LFPs are sensitive to the onset of fixation lights as well as the onset of visual targets presented during steady fixation. The LFP waveforms evoked by combined audiovisual stimuli differed from those evoked by sounds alone. In single-unit spiking activity, responses were weak when visual stimuli were presented alone, but visual stimuli could modulate the activity evoked by sounds in a stronger way. Such modulations could involve either increases or decreases in activity, and whether increases or decreases were observed was variable and not obviously correlated with the responses evoked by visual or auditory stimuli alone. These findings indicate that visual stimuli shape the IC's auditory representation in flexible ways that differ from those observed previously in multisensory areas.

New & Noteworthy

We find that the inferior colliculus, a primarily auditory brain area, displays distinct population-level responses to visual stimuli. We also find that visual cues can influence the auditory responses of individual neurons. Together, the results provide insight into how relatively early sensory areas may play a role in combining multiple sensory modalities to refine the perception of complex environments.

1   **Introduction**

Visual and auditory processing streams were once thought to remain initially separate in

the brain. Studies of how visual and auditory stimuli *interact* focused chiefly on late stages of

processing: association areas such as intraparietal cortex or motor output stages such as the

superior colliculus (SC) (e.g., (Witten and Knudsen 2005; Chandrasekaran 2017; Schmehl and

Groh 2021)). At such late processing stages, two key features characterize response patterns:

a) both modalities can lead to strong responses, i.e., spiking activity increases in response to

either visual or auditory stimuli; and b) combinations of visual and auditory stimuli lead to

greater increases in spiking activity than do either of the two stimuli when presented alone.

Indeed, in the SC, combined visual and auditory stimuli are thought to lead to nearly linear

summation of the responses evoked by each stimulus alone (Meredith and Stein 1986; Wallace

et al. 1998; Stanford et al. 2005; Alvarado et al. 2007).

However, interactions between these sensory systems are now known to begin earlier

than was previously appreciated, including at pre-cortical stages (Bulkin and Groh 2006;

Ghazanfar and Schroeder 2006; Schmehl and Groh 2021). What *kinds* of interactions between

visual and auditory signals occur in "early" areas is unknown, and may be different from those

observed at later stages. Early sensory areas are typically generalists, playing a role in many

aspects of sensory perception, whereas later areas such as the SC are thought to be focused

on specific roles such as generating orienting movements of the eyes to the location of a

sensory stimulus regardless of its modality. And only some of those perceptual functions are

likely to be under the influence of other sensory modalities. It is also not clear that sensory

signals should necessarily add linearly; rather, a variety of different types of modulation may

occur. Indeed, evidence for a diversity of types of interaction has been observed at the level of

core auditory cortex (Schroeder and Foxe 2002; Ghazanfar 2005; Bizley et al. 2007; Kayser et

al. 2007, 2008; Morrill and Hasenstaub 2018; Atilgan et al. 2018).

To gain insight into the properties of multimodal interactions in an even "earlier" sensory

brain region, we turned to a predominantly auditory area, the inferior colliculus (IC). The IC is

located two stops earlier on the auditory pathway than auditory cortex, to which it sends input

via the medial geniculate nucleus as an intermediary (Huffman and Henson 1990; Winer and

Schreiner 2005). Building on previous work establishing the presence of visual inputs to the IC

(retina: Itaya & Van Hoesen, 1982; Yamauchi & Yamadori, 1982; Zhang, 1984; Paloff et al.,

1985; superior colliculus: Harting, 1977; visual cortex: Cooper & Young, 1976) as well as visual

responses in both spiking and local field potential (LFP) activity (Porter et al. 2007; Bulkin and

Groh 2012a), we evaluated both single-unit and LFP signals in the IC during an audiovisual

localization task. Visually-evoked activity was robust in the LFP, but much weaker in spiking

activity. Nevertheless, visual stimuli could alter auditory-evoked responses in both measures of

activity. Intriguingly, the addition of a visual stimulus to an auditory stimulus could either

increase or decrease the response, and the direction of the effect was not predictable from the

unimodal responses. Together, these results indicate that the cross-modal interactions in this

early auditory brain area are considerably different from those reported in the adjacent SC,

suggesting that the computations supporting multisensory interactions may vary widely in the

brain.

Materials and Methods

Subjects

22   Subjects were two rhesus macaques (*Macaca mulatta*), one female age 13 years

23   (Monkey J) and one male age 4 years (Monkey D) at the start of training. All procedures were

1   conducted at Duke University and approved by the Duke University Institutional Animal Care

2   and Use Committee.

Surgical Preparation

Each monkey underwent two stereotactic surgeries using isoflurane anesthesia and

aseptic techniques, and post-surgical analgesics were administered. The first surgery served to

implant a device to limit head motion during experiments. The second surgery consisted of

implanting a recording cylinder (Crist Instruments, 2 cm in diameter) over a craniotomy to make

both the right and left inferior colliculi (IC) accessible at an angle of 26 degrees from vertical in

the coronal plane (approaching from the right for Monkey J and from the left for Monkey D).

Equipment

During experiments, monkeys were seated comfortably in custom primate chairs (Crist

Instruments) with heads restrained. The chair was situated in a dark single-walled sound

attenuation chamber (Industrial Acoustics Company) lined with sound attenuating foam (Sonex).

Visual stimuli were presented via green light-emitting diodes (LEDs), and sounds were

presented from loudspeakers (Cambridge SoundWorks) calibrated to equal volume across

speakers (see "Stimuli"). Monkeys performed a saccade task involving looking at the lights

and/or sounds as described in detail below (see "Task"). Eye position was monitored using an

Oculomatic Pro eye tracking camera and software (500 Hz sampling rate). On correctly-

performed trials, juice rewards were delivered using a custom mouth piece and a PVC tube

(ClearFLEX, Finger Lakes Extrusion) controlled with a solenoid valve (Parker Hannifin

Corporation).

Neural recordings were conducted with 125 mm tungsten microelectrodes (FHC). A 19

mm diameter grid of holes (Crist Instruments) spaced 1 mm apart was placed into the recording

cylinder to allow electrode penetrations, which were controlled by a microdrive (NAN Instruments LTD). Neural activity was collected with a Multichannel Acquisition Processor (MAP system, Plexon Inc.), passed through appropriate amplifiers, and sent to SortClient software for online spike sorting. Single unit spiking activity was extracted from signals bandpass filtered between 150 Hz and 8 kHz. Local field potential (LFP) signals were filtered from 0.7 to 300 Hz and stored at a sampling rate of 20 kHz (Plexon, Inc.). Spike times, eye positions, and task parameters were also stored for offline analysis (Plexon, Inc.; Ryklin Software, Inc.).

Stimuli

As noted above, stimuli consisted of sounds and lights originating from loudspeakers and LEDs located on a horizontal row at the monkey's eye level. Speakers and lights were located at -12 and +12 horizontal degrees in the monkey's visual field. Sounds were bandpass filtered noise at center frequencies of 420 or 2000 Hz (11 kHz sampling rate, 55 dB SPL, ±100 Hz bandwidth, 10 ms on ramp). Sounds were frozen for a given neuron, meaning a single bandpass stimulus was generated for each frequency at the beginning of the session and the same pattern was used throughout the session. Sounds were generated using custom software (Ryklin Software, Inc.). Lights were green light-emitting diodes (LEDs) placed on the front face of each speaker and were amplified with a 5.2 volt power source.

Task

Monkeys performed tasks involving saccade targets at either one or two locations. The current paper reports results from the single location trials; the two-location trials allowed us to explore the contribution of multisensory information to the processing of two objects (Caruso et al. 2018; Mohl et al. 2020; Glynn et al. 2021; Jun et al. 2022; Schmehl et al. 2024; Chen et al. 2024; Groh et al. 2024) and will be the subject of a separate publication. To initiate a trial, the

monkey fixated on a central light (0 horizontal degrees from the midline, at eye level). After 600-

700 ms, a visual, auditory, or combined visual and auditory stimulus was presented at either -12

or 12 degrees while the fixation light remained on. After a further 600-800 ms of fixation, the

fixation light was extinguished to cue the monkey to make a saccade to the target (which itself

remained on until completion of the saccade; Figure 1). The reinforcement window around the

target was 16 degrees wide by 60 degrees tall, to account for low precision in sound localization

in the vertical domain (Gnadt et al. 1991; Barton and Sparks 2001). Monkeys then maintained

fixation on the target for 200-250 ms to receive a fluid reward (typically Welch's 100% grape

juice diluted 30% in water). The additional two-location trials not analyzed here were very

similar, except that two target locations were used simultaneously and the monkeys were

required to make sequential saccades to both target locations. All conditions were randomly

interleaved.

Figure 1: Task structure. Each trial began with the onset of a central light, and the monkey
initiated the trial by fixating on the light for a variable period between 600 and 700 ms. A target
was then presented (auditory, visual, or both), and the monkey waited 600-800 ms for fixation
offset before making a saccade to the target (green arrow). After localizing the target, the
monkey received a fluid reward. Trials continued in succession, and conditions were randomly
interleaved. Conditions involving two target locations (two sounds with varying numbers of
accompanying lights) were also included but are not analyzed here.
21
22   Modalities were weighted such that 30% of the trials were visual, 30% of the trials were

23   auditory, and 40% of the trials were audiovisual. Performance was >74% correct for both

24   monkeys for all modalities, with performance on visual-only and audiovisual trials slightly

25   exceeding that for auditory-only trials (Supplementary Figure 1).

Neural Recordings

The left and right IC in each monkey were localized using magnetic resonance imaging scans conducted at Duke University Hospital: sagittal localizing scan, coronal MPRAGE (T1 weighted 3D image, 0.5 mm), coronal T2 1.0 mm 55 slices with phase set right to left, coronal T2 0.5 mm 55 slices with phase set right to left (through the cylinder), and coronal T2 0.5 mm 55 slices with phase set anterior to posterior (through the cylinder). The location of the IC could be reconstructed accurately in reference to the coordinate system of a grid for holding electrodes in the recording cylinder (Crist Instruments). We recorded from the IC using a single tungsten microelectrode (FHC) while the monkey performed the task and neural data were collected as described above as well as previously (e.g., Willett & Groh, 2022). Neural data was recorded relative to a ground wire attached to the guide tube. Most recording sites were found in the low frequency-responsive region of the IC (Bulkin and Groh 2011).

Data Analysis

All analyses were conducted on correctly performed trials, in which the monkey successfully localized the target. Only recording sites where the neuron was responsive to at least one modality (outlined below) and that had a minimum of 250 correct trials were included for analysis.

Single Unit Activity

We recorded a total of 106 well-isolated single neurons, 82 from the right IC (Monkey J), and 24 from the left (Monkey D). Results were similar across monkeys and were combined for subsequent analysis.

Auditory responsiveness was assessed by comparing the activity during a time window

after sound onset to a corresponding period of steady fixation immediately prior to sound onset.

Two windows were used, either 0-100 ms or 0-500 ms after sound onset, to account for the

presence of primarily transient vs. transient+sustained response patterns, which are known to

occur in IC neurons (Ryan and Miller 1978; Willott and Urban 1978; Aitkin et al. 1994; Syka et

al. 2000; Bulkin and Groh 2011). Units that exhibited a significant change in activity relative to

baseline in either of these periods, or that passed visual inspection of peri-stimulus time

histograms pooled across all auditory conditions, were considered auditory-responsive (pooled

across all sound-only trials regardless of sound location or frequency, paired two-tailed t-test

comparing firing rates before and after stimulus onset, $p < 0.025$ for each tail, equivalent to

$p < 0.05$ for a one-tailed test). 106 neurons met these criteria.

Visual responsiveness was also assessed in several ways. Since each trial involved a

visual fixation stimulus, activity was analyzed in a time window 50-250 ms after the onset of that

fixation stimulus in comparison to a comparable baseline prior to fixation stimulus onset. This

time window was chosen based on prior work, which showed that visual responses in the IC

occur with a slight delay relative to stimulus onset (Gutfreund et al. 2002; Porter et al. 2007).

The position of this fixation stimulus on the retina naturally varied based on where the eyes

happened to be when the fixation stimulus appeared. We also examined responsiveness in a

similar time period following entry of the eye into the fixation window, again in comparison to

baseline (50-250 ms prior to fixation). Finally, on trials in which a visual saccade target was

presented, we examined responsiveness 50-250 ms following target onset. As with the tests of

auditory responsiveness, we used a paired two-tailed t-test, $p < 0.025$ for each tail, deployed on

all the trials involving visual stimuli regardless of their locations. Detailed results will be

presented further below.

Local Field Potentials

3   Local field potential (LFP) signals were successfully analyzed from 83 of the 106

recording sites (the remainder were excluded due to technical problems affecting alignment of

these data with the trial event data). As noted above, LFP signals were filtered from 0.7 to 300

Hz and stored at a sampling rate of 20 kHz (Plexon, Inc.), and were recorded with respect to a

ground wire attached to the guide tube. Signals were normalized by subtracting the mean

voltage during a 200 ms baseline period prior to fixation light onset across all trials for each

recording site. Recording sites were included based on the significance of the associated single

unit activity rather than LFP responsiveness.

Results

Local Field Potential

14   Given that the IC is fundamentally an auditory structure, visually-evoked activity is

expected to be weaker and rarer than auditory-evoked activity. Accordingly, the LFP is a

particularly useful measure because it represents aggregate activity from a population of

neurons, and can include pre- and post-synaptic activity in that population regardless of whether

the threshold for triggering a spike was crossed. Therefore, we first probed the LFP, and we

considered three points in time during the trial: the onset of the visual fixation light, the time of

entry of the eyes into the reinforcement window around the fixation light, and the onset of the

visual target (for trials with visual saccade targets).

22   Figure 2 shows LFP signals aligned to each of these time points. Figure 2a shows

results aligned on fixation light onset for each recording site (individual gray traces). A distinct

wave-like response occurs after fixation light onset for nearly every recording site. This wave

exhibits a positive, then negative, peak before settling at a slightly negative plateau. In Figure

2b, the traces are aligned on the eye's entry into the window around the fixation light, and a

fresh visual response can be observed as the saccade brings the fixation light to a new (more

foveal) retinal location. This response is most evident in the mean across sites (red trace).

Finally, Figure 2c shows the responses to a visual target presented at either -12° or 12° during

steady fixation (data are combined across both locations). These responses appear slightly

weaker than those seen in connection to the onset of the visual fixation light. This could be

because (a) they are added on top of any activity evoked by that target, so the deviation from

the voltage level prior to target onset is modest, (b) they are presented from only 2 retinal

locations, only 1 of which is in the contralateral hemifield, or (c) visually-evoked activity in the IC

is context dependent and depends on the role or timing of the stimulus in the task. We will

discuss these possibilities further in the Discussion.

Figure 2: Local field potentials display responses to fixation and target lights in addition to sounds. Local field potential (LFP) aligned to various stages of the trial. a) LFP aligned on the onset of a fixation light. b) LFP aligned on the time of the monkey's eye entry into a spatial window surrounding the fixation light. Note that sometimes the monkey anticipated the onset of the fixation light and no saccade was required; such trials are excluded from this analysis. c) LFP aligned on the onset of a visual target during steady fixation elsewhere. d) LFP aligned on the onset of an auditory target. e) LFP aligned on the onset of an audiovisual target. f) Difference in LFP shown in (d-e). In all subpanels, gray traces represent individual recording sites, and thick red traces represent the mean across all recording sites. Traces are averaged across all relevant stimulus conditions (e.g., traces aligned on auditory target onset in panel d include all sound frequencies and locations; those in e include all sound frequencies and locations that were paired with visual stimuli). Note the change in y-axis scale in (d-e) compared to (a-c), reflecting stronger auditory and audiovisual than visual responses. See Supplementary Figure 2 for a depiction of the mean traces accompanied by standard error for easier comparison to baseline responses.

In contrast, the sound-evoked LFP is quite strong. Figure 2d shows the signals on auditory trials (pooled across both frequencies and locations). Note the y-axis is adjusted to accommodate these much larger signals. Other than the greater magnitude, their character was similar to the visually-evoked LFPs, again usually beginning with a short-latency and short-duration positive peak, followed by a steadier and longer-lasting negative plateau. In other words, the peak sequences were similar, but occurred much faster on auditory than on visual trials.

The LFP evoked on audiovisual trials (again pooled across all stimulus combinations) is grossly similar to that observed on auditory-only trials (Figure 2e). However, when the mean audiovisual LFP for each site is subtracted from the corresponding auditory one (Figure 2f), a modest influence of the inclusion of the visual stimulus appears, particularly when averaged across sites (red trace deflects away from zero, first negative, then positive peaks).

Together, these findings reveal that visual signals are indeed reaching the IC in the context of this task, and that, as previously shown (Porter et al. 2007; Bulkin and Groh 2012a), the most obvious visual influences can be seen at the onset of the fixation light. Even so, the

1   presentation of a visual target later in the trial does cause a measurable signal and a

2   measurable modulation of auditory signals.

Spiking Responses

We next considered individual neurons' spiking responses to the onset of the fixation

light, as previously reported by (Porter et al. 2007). Generally, these responses were quite

weak. Figure 3a shows the individual (gray) and average peri-stimulus time histograms (PSTHs)

(red) across sites. A few sites show distinct excitatory or inhibitory responses, but, in general,

any visual effect on actual spiking was relatively uncommon in our dataset: the difference in the

number of spikes observed 50-250 ms after fixation light onset was usually not very different

from the number of spikes observed in the same window before fixation light onset (Figure 3b,

Z-scores calculated by dividing the difference between the mean light-evoked and baseline

firing rates by the standard deviation of baseline firing rates). In total, 8% of IC neurons showed

significant excitatory responses (green bars) and 17% showed significant inhibitory responses

(red bars; paired two-tailed t-test, $p < 0.025$ for each tail). One possible explanation for the low

prevalence of visual responses is that the visual receptive fields found in this population of IC

neurons may be poorly probed by "chance" variation in eye position at the time of fixation

stimulus onset. Figure 3c-f show the individual PSTHs (c,d) and receptive field maps (e,f) for the

two most responsive neurons in the sample – while they are strongly responsive, their receptive

fields are quite small and centered on the fovea. The fixation light only appeared at one location,

and the monkeys' eyes often lingered at or near that location while waiting for the next trial to

start. This may have privileged detecting spike responses in a small set of foveally-sensitive IC

neurons while disadvantaging the detection of spiking responses at other receptive field

locations. The LFP signals may have been less sensitive to this limitation, by virtue of pooling

over a larger population of neurons and/or inclusion of subthreshold signals.

3   Similarly, Figure 3g shows the individual (gray) and average PSTHs (red) across sites

for spiking responses to the onset of the target light during fixation. Like responses to the

fixation light, these responses show visual effects on spiking activity in about 17% of the

neurons (Figure 3h; 4% excitatory, green bars; 13% inhibitory, red bars; paired two-tailed t-test,

$p<0.025$). In total, 37% of neurons exhibited significant responses in one or the other of these

two time periods, with 6% responsive to both. When a neuron was responsive during both time

periods, the direction of modulation (excitatory vs. inhibitory) was typically the same in these two

cases. When separating out by target location, slightly more neurons were responsive to the

contralateral stimulus (4% excited, 4% inhibited, or 8% overall) than to the ipsilateral target (2%

excited, 4% inhibited, or 6% overall). (The lower proportion of responsive neurons when limiting

to just one target location may be due to the impact of cutting the data in half on the odds of

reaching statistical significance.) The prevalence of visual responsiveness was also similar in

both monkeys: 23% of neurons in Monkey J and 33% of neurons in Monkey D were responsive

to the fixation light, while 13% of neurons in Monkey J and 29% of neurons in Monkey D were

responsive to the visual target. In summary, spiking modulation occurred in response to both

fixation and target lights, but was rarer in this IC dataset in comparison to previous reports

(Porter et al. 2007; Bulkin and Groh 2012a), perhaps due to the more sparse sampling of visual

space.

Figure 3: Spiking activity in the IC reflects weak and rare responses to fixation and target lights during a localization task. a) PSTHs of individual units (gray) and mean PSTH (red) for responses aligned to the onset of a fixation light presented at the horizontal midline and at eye level. b) Z-scores calculated as a difference between the mean light-evoked and baseline firing rates divided by the standard deviation of baseline firing rates. Green and red bars indicate units with a score significantly different from zero. c-d) Example responses of two individual neurons to the fixation light. Baseline firing rates were subtracted, and PSTHs were smoothed by convolving with a 5-point sliding triangle window and cubic spline interpolation. Shaded areas represent standard error. e-f) Spatial receptive fields of the same neurons as in (c-d), showing each neuron's response to an LED fixation light falling on various horizontal and vertical locations on the retina during passive looking at the start of a trial. Baseline firing rates are indicated by the background color. Activity patterns were smoothed both horizontally and vertically using a 5-degree window. g) PSTHs of individual units (gray) and mean PSTH (red) for responses aligned to the onset of a target light. h) Z-scores calculated as a difference between the mean light-evoked and baseline firing rates divided by the standard deviation of baseline firing rates, using a spike counting window 50-250 ms after stimulus onset. Green and red bars indicate units with a score significantly different from zero. Results in panels g, and h are pooled across both visual target locations for individual neurons. Responses were also evaluated separately for the contralateral and ipsilateral targets (significant responses to the contralateral target: 4% excitatory, 4% inhibitory; for the ipsilateral target: 2% excitatory, 4% inhibitory; results not shown).

We next asked how visual stimuli affect IC spiking activity when presented in

combination with sounds. Specifically, we examined responses aligned to the onset of visual,

auditory, and audiovisual targets following fixation onset during the task. Figure 4 shows the

responses of four example neurons to all combined presentations of visual (blue), auditory

(orange), and audiovisual (purple) targets at single locations. Although these neurons were not

responsive to visual input (blue), pairing that same visual cue with a sound (orange) produced a

different auditory response (purple) – either by enhancing (Figure 4a) or suppressing (Figure

4b,c) the neuron's auditory response. However, this was not always the case: Figure 4d shows

an example of a neuron whose auditory responses appeared to be unaffected by the

simultaneous presence of a visual stimulus.

Figure 4: A visual cue can modulate a neuron's auditory response. Example responses of
four individual neurons to pooled combinations of all visual (blue), auditory (orange), and
audiovisual (purple) conditions presented at a single location. Within each panel, the left
subpanel is a peri-stimulus time histogram (PSTH) of each modality, and the right subpanel is a
quantification of the average responses either in the first 500 ms after stimulus onset (0-500 ms)

for sustained responses, or in the first 100 ms after stimulus onset (0-100 ms) for transient
responses. Baseline firing rates were subtracted, and PSTHs were smoothed by convolving with
a 5-point sliding triangle window and cubic spline interpolation. Shaded areas and error bars
represent standard error. a) The presence of a visual cue augments the auditory response, and
both the auditory and audiovisual responses are sustained throughout the stimulus
presentation. b) The presence of a visual cue dampens the auditory response, and both the
auditory and audiovisual responses are sustained throughout the stimulus presentation. c) The
presence of a visual cue dampens the auditory response only immediately after stimulus onset,
and the auditory and audiovisual responses are similar later in the stimulus presentation
window. d) Auditory and audiovisual responses are similar, indicating no effect of the visual cue
on auditory responsiveness. Note low visual responsiveness in all panels, indicating that the
presence of a visual cue can modulate a neuron's auditory response even if the neuron is
unresponsive to visual input alone.
14
Given the low magnitude and low prevalence of visually-driven responses in these

neurons (Figure 3), the visually-induced modulation of auditory responsiveness (Figure 4) is

quite striking. These results suggest that although visual input may not drive a neuron's spiking

activity individually, pairing the same visual cue with a driving auditory input can be sufficient to

modulate a neuron's response, potentially to inform perceptual output.

We next quantified these effects at the population level. Figure 5a shows the differences

between the responses to an audiovisual stimulus and the corresponding auditory stimulus

alone. About 15% of conditions showed a significant decrease in responsiveness (red/green

bars, paired two-tailed t-test, $p < 0.025$ for each tail), whereas 7.5% showed a significant

increase in responsiveness.

The direction of the effect was not strongly related to whether the visual response was

excitatory on its own, nor was it strongly related to the magnitude of the auditory response on its

own. Figure 5b shows the mean audiovisual response as a function of the mean auditory

response, with color indicating whether the visual response was excitatory (red) or inhibitory

(blue) relative to baseline. Both red and blue points lie both above and below the diagonal line

of slope one, indicating that responses could be either increased or decreased when the visual

and auditory stimuli were combined. Figure 5c shows the same pattern but from a different

perspective – here the differences between the audiovisual and auditory responses are plotted

as a function of the visual responses, with color indicating the magnitude of the auditory

responses. Points lie in all 4 quadrants, indicating that both enhanced and reduced responses

to audiovisual stimuli can occur regardless of whether the visual input is slightly excitatory or

slightly inhibitory when presented on its own. There is little evident relationship to the strength of

the auditory response: darker red points lie in all four quadrants. These results further support

the finding that the presence of a visual cue can modulate auditory responsiveness in the IC,

but reveal that the direction of the effect – increasing vs. decreasing activity – can vary across

the population and is not easily predicted from either the response to sound alone or the

response to the visual stimulus alone.

Figure 5: Visual influences on auditory responses are varied. a) Distribution of audiovisual
minus auditory responses. Visual stimuli added to sounds reduced activity in about 15% of
conditions (red bars) whereas they increased the activity in about 7% of conditions (green bars;
two-tailed t-test, p<0.025). In net, the population activity on audiovisual trials was slightly lower
than on their auditory counterparts (p = 0.0383, paired sample t-test). Asterisk indicates
distribution mean. Green and red bars indicate conditions with a value significantly different from
zero. b) Comparison of mean auditory and audiovisual firing rates for each neuron, with colors

indicating the corresponding visual firing rate for a visual cue presented at the same location. No strong relationship arises between visual firing rates and auditory or audiovisual firing rates, with both red and blue points falling both above and below the unity line. For clarity, visual responses are depicted relative to baseline, such that positive values indicate an excitatory response and negative values indicate an inhibitory one. No such baseline subtraction was applied to the auditory or audiovisual trials, but most were excitatory. c) Comparison of mean difference between audiovisual and auditory firing rates and baseline-subtracted visual firing rates, with colors indicating the corresponding auditory firing rate. Points arise in all four quadrants, indicating a wide degree of visual influences on auditory firing rates. In all subpanels, each neuron contributes 4 data points as sound frequencies and visual cue locations are counterbalanced (4 frequency-location pairs per neuron).

Discussion

Visual cues influence auditory perception at the behavioral level, but the role of early, predominantly unisensory areas in this process has remained relatively unexplored. Here we investigated how visual cues modulate activity in the IC at population and single-unit scales and in response to visual stimuli presented alone or with sounds. We found that fixation lights and visual targets can evoke distinct wave-like responses in local field potential recordings, and that single units exhibit visual responses that are weaker but statistically evident in a modest subpopulation of neurons. Intriguingly, individual units' responses to sounds can be modulated by the presence of a simultaneous visual cue, but in directions that are not easily accounted for by either the auditory or the visual responses alone. Indeed, modulation of auditory signals could be seen even when the visual stimulus did not itself evoke an overt spiking response. This suggests a complex form of interaction between visual and auditory signals in this nominally auditory structure.

These findings extend previous work that identified visual and eye movement-related responses (Porter et al. 2007; Bulkin and Groh 2012a) and eye position-dependent auditory responses (Groh et al. 2001; Zwiers et al. 2004; Porter et al. 2006) in the IC. The chief difference between our current study and the earlier studies from our group (Porter et al. 2007; Bulkin and Groh 2012a) is that the driven visual responses appeared to be less prevalent here.

It is not yet clear what may account for this difference. One possibility is that the current study

sampled visual space in a more limited way – there was only a single fixation location and only

two visual stimulus locations, limiting the sampling of retinal receptive fields. Another possible

factor is that the recordings in the current study were restricted to particular spatial areas within

the IC, whereas previous work intentionally sampled a wide range of locations spanning a large

portion of the IC (Bulkin and Groh 2012a). The difference in the variety of recording sites

sampled may mean that the recording sites in the current study had a lower overall degree of

visual input than in prior studies. It is known that the IC's subregions differ in the prevalence of

visual receptive fields with lower incidence in the low frequency-responsive region that we

tested here (Bulkin and Groh 2012a). Other attributes of the stimuli, tasks, or research subjects

may also contribute to differences in visual responsiveness across studies.

Our results are similar to those in auditory cortex, where a variety of multisensory

interactions have been identified. In auditory cortex, visual cues can elicit their own responses

as well as modulate auditory responses in both excitatory and inhibitory ways (Bizley et al.

2007; Kayser et al. 2007, 2008; Morrill and Hasenstaub 2018). Non-auditory cues can also

modulate current source density in auditory cortex, either by eliciting direct visual responses

(Schroeder and Foxe 2002) or by modulating an auditory response by resetting the phase of

neural oscillations (Lakatos et al. 2007). These effects are also evident for stimuli with high

behavioral relevance; images or videos of faces can elicit their own independent responses as

well as modulate LFP activity in response to vocalizations (Ghazanfar 2005). Finally, visual

stimuli can modulate auditory responsiveness in more complicated ways, such as influencing

which of two sounds is predominantly represented by individual neurons or the neural

population (Atilgan et al. 2018).

Together, our results and those of these other prior studies in auditory cortex differ from

related investigations in more multimodal areas such as the superior colliculus (SC). In the SC,

which is reciprocally connected to the IC (Woollard and Harpman 1940; Moore and Goldberg

1963; Powell and Hatton 1969; Harting 1977; Adams 1980; Covey et al. 1987; Coleman and

Clerici 1987; Sparks and Hartwich-Young 1989; Doubell et al. 2000; Hyde and Knudsen 2000;

Caño et al. 2006; Stitt et al. 2015), spiking responses to both visual and auditory stimuli are

common. Combinations of visual and auditory stimuli are generally thought to increase the

activity of individual neurons, most often in an additive fashion (Meredith and Stein 1986;

Wallace et al. 1998; Stanford et al. 2005; Alvarado et al. 2007). In contrast, audiovisual

responses in the IC can be either slightly higher or slightly lower than auditory-alone responses.

These patterns occurred even when a neuron had no visually-driven response, and rarely

appeared to be additive.

A likely factor in these differences is that IC is thought of as a general-purpose auditory

area, in contrast to the multimodal/oculomotor SC. The IC is located only a few synapses along

the auditory pathway from the cochlea, and nearly all ascending auditory input passes through

this brain region. Thus, it is likely that the overall purpose of this structure is to process sound,

and that any visual signals in this area serve a purpose of shaping those responses rather than

simply adding to them. Such shaping is likely to take multiple forms and not reduce easily to a

simple linear operation in which stronger input signals lead to more output (spiking) activity. Our

study provides a first look at the scope of the IC's visual-auditory interactions and the factors

that govern them. More work will be needed to explore such factors. For example, full mapping

of visual receptive fields should be combined with assessing the role of visual-auditory spatial

alignment on IC activity. This is a particularly interesting unexplored domain because the native

codes for visual and auditory space are so different. The optics of the eye cause light from

different locations in the environment to land at particular locations on the retinal surface,

leading neurons throughout the visual pathway to have circumscribed receptive fields. In

contrast, sound location is computed in part from binaural difference cues, which are

proportional to the horizontal eccentricity of a sound. Neurons in the primate IC (and SC) do not

exhibit circumscribed receptive fields, but rather appear to encode auditory space as a meter or

rate code in which a large population of neurons responds with a particular level of activity,

depending on where a sound is located along the azimuth (primate IC: Groh et al., 2003; Zwiers

et al., 2004; primate SC: Lee & Groh, 2014; see also: Middlebrooks et al., 1994, 1998; McAlpine

& Grothe, 2003; Werner-Reiss & Groh, 2008; Salminen et al., 2009). The brain must combine

these disparate visual and auditory codes to understand where visual and auditory signals arise

in space, and the IC is emerging as a site where such combination may occur.

In addition, further work is needed in the frequency domain. For example, at the

behavioral level, visual lip reading cues can modulate the perception of speech sounds (e.g.,

the McGurk effect, (Sams et al. 1991; Wright et al. 2003; Ghazanfar 2005; Kayser et al. 2007,

2008; Hoffman et al. 2008; Nath and Beauchamp 2012)). This perceptual phenomenon might

emerge from visually-induced changes in the representation of sound frequency within the

auditory pathway. Further exploration is needed to ascertain the properties that govern these

neural responses and their perceptual correlates.

Collectively, these experiments provide insight into the influence of visual cues on

auditory perception. Together with the other studies in related auditory regions, the findings

presented here shed new light on the connections between multiple sensory systems and the

ways in which context from another modality can inform processing of a brain region's

"dominant" modality. As the field of neuroscience moves forward, it will be critical to consider

how sensory systems interact to give rise to complex behaviors in varied sensory environments.

Supplementary Materials

Supplementary Figure 1: Monkeys accurately perform an audiovisual localization task. a-b) Mean and standard error of saccade endpoints during trials performed by each monkey for visual, auditory, and audiovisual stimuli presented at the two horizontal target locations in the task. Endpoints for all three modalities are largely overlapping and are aligned with the actual target location. c-d) Proportion of attempted trials of each modality in which each monkey correctly localized the targets within a 16-degree wide by 60-degree tall window. Visual and audiovisual conditions had a higher proportion of correct trials than auditory conditions, but all types of conditions were performed at an average accuracy of approximately 74% or greater.

Supplementary Figure 2: Mean local field potentials diverge from baseline in responses to fixation lights, target lights, and sounds. Same mean local field potential (LFP) responses as shown in Figure 2, with shaded areas representing standard error. As in Figure 2, responses are aligned to various stages of the trial. Standard error remains narrow and diverges from baseline in all cases.

References

Adams JC. Crossed and descending projections to the inferior colliculus. *Neuroscience Letters* 19: 1–5, 1980. doi:10.1016/0304-3940(80)90246-3.

Aitkin L, **Tran L**, **Syka J**. The responses of neurons in subdivisions of the inferior colliculus of cats to tonal, noise and vocal stimuli. *Exp Brain Res* 98: 53–64, 1994. doi:10.1007/BF00229109.

Alvarado JC, **Vaughan JW**, **Stanford TR**, **Stein BE**. Multisensory Versus Unisensory Integration: Contrasting Modes in the Superior Colliculus. *Journal of Neurophysiology* 97: 3193–3205, 2007. doi:10.1152/jn.00018.2007.

Atilgan H, **Town SM**, **Wood KC**, **Jones GP**, **Maddox RK**, **Lee AKC**, **Bizley JK**. Integration of Visual Information in Auditory Cortex Promotes Auditory Scene Analysis through Multisensory Binding. *Neuron* 97: 640-655.e4, 2018. doi:10.1016/j.neuron.2017.12.034.

Barton EJ, **Sparks DL**. Saccades to remembered targets exhibit enhanced orbital position effects in monkeys. *Vision Research* 41: 2393–2406, 2001. doi:10.1016/S0042-6989(01)00130-4.

Bizley JK, **Nodal FR**, **Bajo VM**, **Nelken I**, **King AJ**. Physiological and Anatomical Evidence for Multisensory Interactions in Auditory Cortex. *Cereb Cortex* 17: 2172–2189, 2007. doi:10.1093/cercor/bhl128.

Bulkin DA, **Groh JM**. Seeing sounds: visual and auditory interactions in the brain. *Current Opinion in Neurobiology* 16: 415–419, 2006. doi:10.1016/j.conb.2006.06.008.

Bulkin DA, **Groh JM**. Systematic mapping of the monkey inferior colliculus reveals enhanced low frequency sound representation. *Journal of Neurophysiology* 105: 1785–1797, 2011. doi:10.1152/jn.00857.2010.

Bulkin DA, **Groh JM**. Distribution of visual and saccade related information in the monkey inferior colliculus. *Front Neural Circuits* 6, 2012a. doi:10.3389/fncir.2012.00061.

Bulkin DA, **Groh JM**. Distribution of eye position information in the monkey inferior colliculus | Journal of Neurophysiology. *Journal of Neurophysiology* 107: 785–795, 2012b. doi:https://doi.org/10.1152/jn.00662.2011.

Caño GGD, **Gerrikagoitia I**, **Alonso-Cabria A**, **Martínez-Millán L**. Organization and origin of the connection from the inferior to the superior colliculi in the rat. *Journal of Comparative Neurology* 499: 716–731, 2006. doi:10.1002/cne.21107.

Caruso VC, **Mohl JT**, **Glynn C**, **Lee J**, **Willett SM**, **Zaman A**, **Ebihara AF**, **Estrada R**, **Freiwald WA**, **Tokdar ST**, **Groh JM**. Single neurons may encode simultaneous stimuli by switching between activity patterns. *Nature Communications* 9, 2018. doi:10.1038/s41467-018-05121-8.

Chandrasekaran C. Computational principles and models of multisensory integration. *Current*
Opinion in Neurobiology 43: 25–34, 2017. doi:10.1016/j.conb.2016.11.002.

Chen Y, **Groh JM**, **Tokdar ST**. Spike Count Analysis for MultiPlexing Inference (SCAMPI).
bioRxiv: 2024.09.14.613077, 2024.

Coleman JR, **Clerici WJ**. Sources of projections to subdivisions of the inferior colliculus in the
rat. *J Comp Neurol* 262: 215–226, 1987. doi:10.1002/cne.902620204.

Cooper MH, **Young PA**. Cortical projections to the inferior colliculus of the cat. *Experimental*
Neurology 51: 488–502, 1976. doi:10.1016/0014-4886(76)90272-7.

Covey E, **Hall WC**, **Kobler JB**. Subcortical connections of the superior colliculus in the
mustache bat, Pteronotus parnellii. *Journal of Comparative Neurology* 263: 179–197, 1987.
doi:10.1002/cne.902630203.

Doubell TP, **Baron J**, **Skaliora I**, **King AJ**. Topographical projection from the superior colliculus
to the nucleus of the brachium of the inferior colliculus in the ferret: convergence of visual and
auditory information. *European Journal of Neuroscience* 12: 4290–4308, 2000.
doi:10.1111/j.1460-9568.2000.01337.x.

Ghazanfar AA. Multisensory Integration of Dynamic Faces and Voices in Rhesus Monkey
Auditory Cortex. *Journal of Neuroscience* 25: 5004–5012, 2005.
doi:10.1523/JNEUROSCI.0799-05.2005.

Ghazanfar AA, **Schroeder CE**. Is neocortex essentially multisensory? *Trends in Cognitive*
Sciences 10: 278–285, 2006. doi:10.1016/j.tics.2006.04.008.

Glynn C, **Tokdar ST**, **Zaman A**, **Caruso VC**, **Mohl JT**, **Willett SM**, **Groh JM**. Analyzing second
order stochasticity of neural spiking under stimuli-bundle exposure. *The Annals of Applied*
Statistics 15: 41–63, 2021. doi:10.1214/20-AOAS1383.

Gnadt JW, **Bracewell RM**, **Andersen RA**. Sensorimotor transformation during eye movements
to remembered visual targets. *Vision Research* 31: 693–715, 1991. doi:10.1016/0042-
6989(91)90010-3.

Groh JM, **Kelly KA**, **Underhill AM**. A Monotonic Code for Sound Azimuth in Primate Inferior
Colliculus. *Journal of Cognitive Neuroscience* 15: 1217–1231, 2003.
doi:10.1162/089892903322598166.

Groh JM, **Schmehl MN**, **Caruso VC**, **Tokdar ST**. Signal switching may enhance processing
power of the brain. *Trends in Cognitive Sciences* 28: 600–613, 2024.
doi:10.1016/j.tics.2024.04.008.

Groh JM, **Trause AS**, **Underhill AM**, **Clark KR**, **Inati S**. Eye Position Influences Auditory
Responses in Primate Inferior Colliculus. *Neuron* 29: 509–518, 2001. doi:10.1016/S0896-
6273(01)00222-7.

Gutfreund Y, **Zheng W**, **Knudsen EI**. Gated Visual Input to the Central Auditory System.
Science 297: 1556–1559, 2002. doi:10.1126/science.1073712.

Harting JK. Descending pathways from the superior colliculus: An autoradiographic analysis in
the rhesus monkey (Macaca mulatta). *Journal of Comparative Neurology* 173: 583–612, 1977.
doi:10.1002/cne.901730311.

Hoffman KL, **Ghazanfar AA**, **Gauthier I**, **Logothetis NK**. Category-specific responses to faces
and objects in primate auditory cortex. *Front Syst Neurosci* 2, 2008.
doi:10.3389/neuro.06.002.2007.

Huffman RF, **Henson OW**. The descending auditory pathway and acousticomotor systems:
connections with the inferior colliculus. *Brain Research Reviews* 15: 295–323, 1990.
doi:10.1016/0165-0173(90)90005-9.

Hyde PS, **Knudsen EI**. Topographic projection from the optic tectum to the auditory space map
in the inferior colliculus of the barn owl. *J Comp Neurol* 421: 146–160, 2000.

Itaya SK, **Van Hoesen GW**. Retinal innervation of the inferior colliculus in rat and monkey.
Brain Research 233: 45–52, 1982. doi:10.1016/0006-8993(82)90928-3.

Jun NY, **Ruff DA**, **Kramer LE**, **Bowes B**, **Tokdar ST**, **Cohen MR**, **Groh JM**. Coordinated
multiplexing of information about separate objects in visual cortex. *eLife* 11: e76452, 2022.
doi:10.7554/eLife.76452.

Kayser C, **Petkov CI**, **Augath M**, **Logothetis NK**. Functional Imaging Reveals Visual
Modulation of Specific Fields in Auditory Cortex. *J Neurosci* 27: 1824–1835, 2007.
doi:10.1523/JNEUROSCI.4737-06.2007.

Kayser C, **Petkov CI**, **Logothetis NK**. Visual Modulation of Neurons in Auditory Cortex.
Cerebral Cortex 18: 1560–1574, 2008. doi:10.1093/cercor/bhm187.

Lakatos P, **Chen C-M**, **O'Connell MN**, **Mills A**, **Schroeder CE**. Neuronal Oscillations and
Multisensory Interaction in Primary Auditory Cortex. *Neuron* 53: 279–292, 2007.
doi:10.1016/j.neuron.2006.12.011.

Lee J, **Groh JM**. Different Stimuli, Different Spatial Codes: A Visual Map and an Auditory Rate
Code for Oculomotor Space in the Primate Superior Colliculus. *PLOS ONE* 9: e85017, 2014.
doi:10.1371/journal.pone.0085017.

McAlpine D, **Grothe B**. Sound localization and delay lines – do mammals fit the model? *Trends*
in Neurosciences 26: 347–350, 2003. doi:10.1016/S0166-2236(03)00140-1.

Meredith MA, **Stein BE**. Visual, auditory, and somatosensory convergence on cells in superior
colliculus results in multisensory integration. *Journal of Neurophysiology* 56: 640–662, 1986.
doi:10.1152/jn.1986.56.3.640.

Middlebrooks JC, **Clock AE**, **Xu L**, **Green DM**. A panoramic code for sound location by
cortical neurons. *Science* 264: 842–844, 1994. doi:10.1126/science.8171339.

Middlebrooks JC, **Xu L**, **Eddins AC**, **Green DM**. Codes for Sound-Source Location in
Nontonotopic Auditory Cortex. *Journal of Neurophysiology* 80: 863–881, 1998.
doi:10.1152/jn.1998.80.2.863.

Mohl JT, **Caruso VC**, **Tokdar ST**, **Groh JM**. Sensitivity and specificity of a Bayesian single trial
analysis for time varying neural signals. *NBDT* 3: 1–19, 2020.
doi:https://doi.org/10.48550/arXiv.2001.11582.

Moore RY, **Goldberg JM**. Ascending projections of the inferior colliculus in the cat. *Journal of*
Comparative Neurology 121: 109–135, 1963. doi:10.1002/cne.901210109.

Morrill RJ, **Hasenstaub AR**. Visual Information Present in Infragranular Layers of Mouse
Auditory Cortex. *J Neurosci* 38: 2854–2862, 2018. doi:10.1523/JNEUROSCI.3102-17.2018.

Nath AR, **Beauchamp MS**. A neural basis for interindividual differences in the McGurk effect, a
multisensory speech illusion. *NeuroImage* 59: 781–787, 2012.
doi:10.1016/j.neuroimage.2011.07.024.

Paloff AM, **Usunoff KG**, **Hinova-Palova DV**, **Ivanov DP**. Retinal innervation of the inferior
colliculus in adult cats: Electron microscopic observations. *Neuroscience Letters* 54: 339–344,
1985. doi:10.1016/S0304-3940(85)80101-4.

Porter KK, **Metzger RR**, **Groh JM**. Representation of Eye Position in Primate Inferior
Colliculus. *Journal of Neurophysiology* 95: 1826–1842, 2006. doi:10.1152/jn.00857.2005.

Porter KK, **Metzger RR**, **Groh JM**. Visual- and saccade-related signals in the primate inferior
colliculus. *PNAS* 104: 17855–17860, 2007. doi:10.1073/pnas.0706249104.

Powell EW, **Hatton JB**. Projections of the inferior colliculus in cat. *Journal of Comparative*
Neurology 136: 183–192, 1969. doi:10.1002/cne.901360205.

Ryan A, **Miller J**. Single unit responses in the inferior colliculus of the awake and performing
rhesus monkey. *Exp Brain Res* 32: 389–407, 1978. doi:10.1007/BF00238710.

Salminen NH, **May PJC**, **Alku P**, **Tiitinen H**. A Population Rate Code of Auditory Space in the
Human Cortex. *PLOS ONE* 4: e7600, 2009. doi:10.1371/journal.pone.0007600.

Sams M, **Aulanko R**, **Hämäläinen M**, **Hari R**, **Lounasmaa OV**, **Lu S-T**, **Simola J**. Seeing
speech: visual information from lip movements modifies activity in the human auditory cortex.
Neuroscience Letters 127: 141–145, 1991. doi:10.1016/0304-3940(91)90914-F.

Schmehl MN, **Caruso VC**, **Chen Y**, **Jun NY**, **Willett SM**, **Mohl JT**, **Ruff DA**, **Cohen M**,
Ebihara AF, **Freiwald WA**, **Tokdar ST**, **Groh JM**. Multiple objects evoke fluctuating responses
in several regions of the visual pathway. *eLife* 13: e91129, 2024. doi:10.7554/eLife.91129.

Schmehl MN, **Groh JM**. Visual Signals in the Mammalian Auditory System. *Annu Rev Vis Sci*
7: 1–23, 2021. doi:10.1146/annurev-vision-091517-034003.

Schroeder CE, **Foxe JJ**. The timing and laminar profile of converging inputs to multisensory
areas of the macaque neocortex. *Cognitive Brain Research* 14: 187–198, 2002.
doi:10.1016/S0926-6410(02)00073-3.

Sparks DL, **Hartwich-Young R**. The deep layers of the superior colliculus. *Rev Oculomot Res*
3: 213–255, 1989.

Stanford TR, **Quessy S**, **Stein BE**. Evaluating the Operations Underlying Multisensory
Integration in the Cat Superior Colliculus. *J Neurosci* 25: 6499–6508, 2005.
doi:10.1523/JNEUROSCI.5095-04.2005.

Stitt I, **Galindo-Leon E**, **Pieper F**, **Hollensteiner KJ**, **Engler G**, **Engel AK**. Auditory and visual
interactions between the superior and inferior colliculi in the ferret. *European Journal of*
Neuroscience 41: 1311–1320, 2015. doi:10.1111/ejn.12847.

Syka J, **Popelář J**, **Kvašňák E**, **Astl J**. Response properties of neurons in the central nucleus
and external and dorsal cortices of the inferior colliculus in guinea pig. *Exp Brain Res* 133: 254–
266, 2000. doi:10.1007/s002210000426.

Wallace MT, **Meredith MA**, **Stein BE**. Multisensory Integration in the Superior Colliculus of the
Alert Cat. *Journal of Neurophysiology* 80: 1006–1010, 1998. doi:10.1152/jn.1998.80.2.1006.

Werner-Reiss U, **Groh JM**. A Rate Code for Sound Azimuth in Monkey Auditory Cortex:
Implications for Human Neuroimaging Studies. *J Neurosci* 28: 3747–3758, 2008.
doi:10.1523/JNEUROSCI.5044-07.2008.

Willett SM, **Groh JM**. Multiple sounds degrade the frequency representation in monkey inferior
colliculus. *European Journal of Neuroscience* 55: 528–548, 2022. doi:10.1111/ejn.15545.

Willott JF, **Urban GP**. Response properties of neurons in nuclei of the mouse inferior colliculus.
J Comp Physiol 127: 175–184, 1978. doi:10.1007/BF01352302.

Winer JA, **Schreiner C**, editors. *The inferior colliculus: with 168 illustrations*. New York, NY:
Springer, 2005.

Witten IB, **Knudsen EI**. Why Seeing Is Believing: Merging Auditory and Visual Worlds. *Neuron*
48: 489–496, 2005. doi:10.1016/j.neuron.2005.10.020.

Woollard HH, **Harpman JA**. The connexions of the inferior colliculus and of the dorsal nucleus
of the lateral lemniscus [Online]. *J Anat* 74: 441-458.3,
1940https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1252612/ [4 Feb. 2024].

Wright TM, **Pelphrey KA**, **Allison T**, **McKeown MJ**, **McCarthy G**. Polysensory Interactions
along Lateral Temporal Regions Evoked by Audiovisual Speech. *Cerebral Cortex* 13: 1034–
1043, 2003. doi:10.1093/cercor/13.10.1034.

Yamauchi K, **Yamadori T**. Retinal Projection to the Inferior Colliculus in the Rat. *CTO* 114:
355–360, 1982. doi:10.1159/000145608.

Zhang AB. Retinotectal pathways in rodents: particularly from the retinal ganglion cells to the
inferior colliculus. *Taiwan Yi Xue Hui Za Zhi* 83: 1–8, 1984.

Zwiers MP, **Versnel H**, **Opstal AJV**. Involvement of Monkey Inferior Colliculus in Spatial
Hearing. *J Neurosci* 24: 4145–4156, 2004. doi:10.1523/JNEUROSCI.0199-04.2004.

5