Title:

High frequency bursts facilitate fast communication for human spatial

attention

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Abstract

Brain-wide communication supporting flexible behavior requires coordination between

sensory and associative regions but how brain networks route sensory information at fast

timescales to guide action remains unclear. Using human intracranial electrophysiology and

spiking neural networks during spatial attention tasks, where participants detected targets at cued

locations, we show that high-frequency activity bursts (HFAb) mark temporal windows of elevated population firing that enable fast, long-range communications. HFAbs were evoked by sensory cues and targets, dynamically coupled to low-frequency rhythms. Notably, both the strength of cue-evoked HFAbs and their decoupling from slow rhythms predicted behavioral accuracy. HFAbs synchronized across the brain, revealing distinct cue- and target-activated subnetworks. These subnetworks exhibited lead-lag dynamics following target onset, with cue-activated subnetworks preceding target-activated subnetworks when cues were informative. Computational modeling suggested that HFAbs reflect transitions to population spiking, denoting temporal windows for network communications supporting attentional performance. These findings establish HFAbs as signatures of population state transitions, supporting information routing across distributed brain networks.

Main text

Prioritizing information from the external environment to guide ongoing behavior and upcoming actions requires fast coordination of neural activity in large-scale networks distributed across distant brain areas.[1–6] This coordination allows information to be routed selectively from sensory to higher level executive brain networks.[7–9] Previous research, particularly in non-human primates, has shown that selective information routing emerges through dynamically changing neuronal interactions.[2,10,11] Such studies highlight the role of oscillatory dynamics and transient changes in inter-areal coherence in enabling attentional selection and the flexible reconfiguration of neural pathways according to task demands. Yet, most prior investigations have focused on pairwise

interactions between a few brain areas, leaving open the question of how fast neural dynamics

emerge and enable flexible, large-scale information routing across distributed networks.

Investigating these questions in the human brain is challenging due to spatial or temporal

constraints of available techniques. Studies examining network-level interactions have largely

been based on connectivity maps derived from functional magnetic resonance imaging (fMRI).

While fMRI studies provide valuable insights into functional networks, the temporal resolution

cannot capture sub-second routing dynamics in attention tasks.[12–15] Electroencephalography

(EEG) and magnetoencephalography (MEG) offer high temporal resolution but have limited

spatial resolution.[16,17] Lastly, single unit recordings provide both fine temporal and spatial signals

but lack the broad coverage for addressing questions of brain-wide network communication.[2,6,18]

Human intracranial electroencephalography (iEEG), offers a unique opportunity to address these

challenges by providing spatially localized and temporally precise neural signals from multiple

brain regions.[19,20] High-frequency activity detected in iEEG signals correlates with different

cognitive functions, including attention,[19–24] and has been reported to index aggregated spiking

activity, dendritic post-synaptic activity, state transitions into spiking regimes, or spike current

leakage to local field potentials (LFPs).[25–29] Additionally, high-frequency activities show long-

range phase synchronization with different frequency bands, making them candidates for studying

fast brain-wide communications.[21,30–33] However, iEEG studies of attention have predominantly

focused on fast-slow interactions, such as cross-frequency coupling between high- and low-

frequency rhythms, which may not fully capture the dynamics of fast-fast interactions between

high-frequency activities.[21,30,34–37]

Here, we address this gap by identifying transient high-frequency activity bursts (HFAbs) in

human iEEG data from epilepsy patients performing spatial attention tasks, hypothesizing that

HFAbs reflect periods when neural populations establish temporal alignment across distributed

regions, creating windows for selective information routing. HFAbs during sensory cue processing

predicted successful detection of upcoming targets. They were locally phase-locked to slow

rhythms (4-25 Hz), and transiently decoupled during cue and target processing, with decoupling

associated with correct performance. Across brain networks, HFAbs temporal relationships were

largely characterized by zero-lag synchronization, identifying functionally specialized

subnetworks with distinct topographical distributions and temporal activation sequences.

Specifically, HFAbs in cue-subnetwork preceded target-subnetwork activity following target

onset, when sensory cues provided relevant information about the target location. Using

computational modeling, we found that HFAbs mark state transitions in neural populations into a

spiking state, characterized by bouts of elevated neuronal firing, indicating enhanced population

excitability enabling synchronized and temporally ordered activation patterns across regions.

Results

HFAbs track spatial attention and predict behavioral accuracy

We analyzed iEEG data from two spatial attention tasks in which patients were cued either

exogenously or endogenously to a spatial location to detect visual targets (**Fig. 1a**, **Extended Data**

Fig. 1a). Throughout the paper, the main figures focus on the results of experiment 1, while the

results of experiment 2 are presented as supplementary information, unless mentioned otherwise

(for brain group-average heatmap plots, we combined both experiments to improve 3D rendering

coverage).

In experiment 1, patients performed a spatial attention task.[23] Each trial started with

the presentation of two vertical or horizontal bar stimuli. Patients were instructed to fixate their

gaze at the center of the display. A transient spatial cue appeared at the end of one bar, exogenously

cueing an upcoming target location. Following a delay period, a target (i.e., luminance changes at

perceptual threshold) appeared at the cued location or infrequently at equally distant non-cued

locations. Patients responded when they detected the target (**Fig. 1a**). In experiment 2, patients

were endogenously cued to a hemifield and reported a target if it appeared in the cued hemifield[3]

(**Extended Data Fig. 1a**, see **Methods,** see **Extended Data Fig. 1b** for detailed performance

breakdowns).

First, we used an adaptive method[6] and detected bursts of high frequency activity at each electrode

(HFAbs, 65-115 Hz, intermittent high amplitude events lasting more than 2.5 frequency cycles,

average burst length 36.2 ms; **Fig. 1b**). The frequency band was selected based on average spectral

peaks observed across subjects (91.2 ± 20.9 Hz, n = 12, see **Methods**). HFAbs occurred frequently

(0.1 ± 0.01 Hz) on average in our data, approximately 15-fold higher than recently described

cortical ripples (0.006 ± 0.0008 Hz),[33] suggesting distinct characteristics of HFAbs despite

overlapping spectral properties (see **Methods**). We then calculated the HFAb density (HFAb

events per unit of time) for each electrode to examine their evoked response during different task

epochs. HFAbs showed higher density averaged across all electrodes in response to both cue and

target (**Fig. 1c**). We measured HFAb responses following cue and target onsets across different

trial outcomes (hit, reject, miss, and false alarm). At the population level, HFAbs activated to cue

onsets in correct hit and reject trials (Wilcoxon signed-rank test, $p < 0.001$) and differed

significantly from missed and false alarm trials (Kruskal-Wallis test, $p = 0.001$, Dunn's test, $p =$

0.008 and $p = 0.01$, respectively; **Fig. 1d**). Target responses were also enhanced in correct hit trials

compared to other outcome conditions (Kruskal-Wallis test, $p < 0.001$; Dunn's test, $p < 0.001$; **Fig.**

1d). Similar results were observed in experiment 2 (**Extended Data Fig. 1c,d**). Overall, $10.7 \pm$

2% of channels (n = 12) showed significant activation to cues, whereas $33.2 \pm 3\%$ of channels (n

= 12) were responsive to targets (Wilcoxon signed-rank test, $p < 0.05$). The broader activation to

targets is in accord with greater complexity of target processing, engaging brain regions involved

in sensory processing, motor planning, and decision-making. Among channels that responded to

cues and/or targets, HFAb responses to targets were negatively correlated with those to the cue

(Spearman correlation, $p < 0.001$, $r = -0.36$, **Fig. 1e**), suggesting distinct electrode populations

process cues and targets. Cue responses were prominent in occipital and parietal regions, including

areas in extrastriate cortex, intraparietal sulcus (IPS), temporoparietal junction (TPJ), superior

parietal lobule (SPL), and inferior parietal lobule (IPL) (**Fig. 1f**), whereas target responses were

more widely distributed including superior, middle, and inferior frontal gyrus, precentral and

postcentral gyrus, IPL, SPL, and TPJ (**Fig. 1g**). This activation profile was evident only for correct

trials both for cue- and target responses (**Extended Data Fig. 1e,f**). It is worth noting that for

experiment 2, we also observed enhanced cue responses in the frontal gyrus consistent with more

top-down attentional control in endogenous cueing. See **Extended Data Table 1** for detailed

electrode positions of the cue.

We investigated the effect of cue validity (targets at cued vs uncued location) and laterality (visual

field ipsilateral versus contralateral to electrodes) on cue and target responses using a Generalized

Linear Mixed Effect (GLME) model (see **Methods**). We found a main effect of outcome on both

cue response ($t = 2.8$, $p = 0.005$) and target response ($t = 15.2$, $p < 0.001$), a main effect of validity

on the target response (**Fig. 1i**, $t = 4.7$, $p < 0.001$), and a main effect of laterality on the cue response

(**Fig. 1j**, $t = 5.1$, $p < 0.001$). No significant main effect was observed for laterality on target (**p** =

0.13), or validity on cue responses (**Extended Data Fig. 1g**, $p = 0.32$, See **Extended Data Fig. 1h**

for experiment 2).

Given that HFAb activation to sensory cues was associated with outcome accuracy, we examined

if HFAb density following cue onset in cue responsive electrodes (Cue+) predicted correct trials.

A classifier was trained on the grand mean of HFAb density across selected electrodes using a 350

ms sliding window around the cue onset, with trial-level outputs labeled as correct or incorrect

(see **Methods**). In Cue+ electrodes, HFAb density within 500 ms post-cue predicted correct

performance above baseline and chance levels (binomial test, FDR corrected for dependent

samples, $p < 0.05$, **Fig. 1k**). This prediction was consistent across both experiments (**Extended**

Data Fig. 1i) and was not dependent on an individual subject.

These findings indicate that HFAbs occur frequently in response to sensory cues and targets and

exhibit distinct spatial profiles across the brain. HFAbs predicted performance accuracy following

cues on a trial-by-trial basis. Together, these spatially selective and behaviorally predictive

responses support HFAbs as neural signatures of attention-related processing.

HFAbs are coupled to slow rhythms and decouple in response to cues and targets

High-frequency activity dynamics in brain networks are linked to low-frequency dynamics,

especially through phase-locking relationships with theta rhythms (4-8 Hz).[30] We asked whether

HFAbs synchronized to the phase of low-frequency rhythms, and whether these cross-frequency

dynamics were associated with task variables. For each electrode, we extracted the LFP around

the HFAb centers and measured both the HFAb-triggered LFP average and the phase locking value

(PLV) of HFAb peaks to low-frequency LFP dynamics. Spectral analysis revealed HFAb-triggered

spectral peaks (**Fig. 2a**, top) and phase locking peaks (**Fig. 2a**, bottom) in theta (4–8 Hz), alpha

(8–14 Hz), and beta (15–25 Hz) frequency bands. Phase-locking peaks occurred at one or more

narrow frequency bands, showing different mean phases (**Fig. 2b**; see **Extended Data Fig. 2** for

additional single-electrode examples, mean phase distributions in theta/alpha and beta bands

across subjects, and spectral and phase-locking peak details). Phase-locking was evident in most

electrodes across all subjects (reliable peaks in $97.8\% \pm 0.8\%$ of electrodes; mean peak frequency

9 ± 0.24 Hz; significant peaks marked by black dots, **Fig. 2c**; see **Extended Data Fig. 3a,b** for

topography of phase-locking peak frequencies).

To test whether sensory cues or target processing affected HFAb phase-locking, we analyzed time-

resolved PLV in sliding windows around cue and target onsets (HFAb count controlled per

window; see **Methods**). HFAbs showed a transient decrease in their phase-locking (decoupling)

in theta/alpha and beta frequency bands following cue and target onsets (**Fig. 2d, Extended Data**

Fig. 3c, randomization test, $p < 0.05$, FDR corrected for dependent samples). This decoupling was

not attributable to event-triggered potentials or the measurement approach (**Extended Data Fig.**

3d,e see **Methods**). Furthermore, changes in HFAb coupling strength were only observed in

correct trials, not in incorrect trials (**Fig. 2e,f,** HFAb counts controlled across trial conditions, see

Methods, see **Extended Data Fig. 3f** for individual examples). Similar results were observed in

experiment 2 (**Extended Data Fig. 3g**).

Next, we examined whether changes in electrodes' HFAb coupling to low frequencies were related

to their responsiveness to cues and targets. For each subject, we correlated coupling ratios (changes

in HFAb coupling to <25 Hz after cues or targets relative to baseline; see **Methods**) with burst

density. Coupling ratios at cue and target onsets were negatively correlated with burst density

following these events suggesting that electrodes with strong HFAb responses also showed

decoupling (**Fig. 2g,h**, Spearman correlation, r = -0.22, p < 0.001, and r = -23, p<0.001,

respectively). GLME models showed a main effect of cue response on coupling ratio after cue

onset (**Fig. 2g** t = -6.17, p < 0.001), and of target response after target onset (**Fig. 2h,** t = -5.05, p

< 0.001). No significant cross-effects were observed (cue response after target onset: p = 0.15;

target response after cue onset: p = 0.21; **Extended Data Fig. 3h,i**).

Overall, HFAbs were predominantly phase-locked to theta, alpha, and beta rhythms. However,

their coupling strength to slower rhythms decreased during cue and target processing, specifically

in correct trials. This selective decoupling during correct trials suggests that releasing HFAbs from

their default phase-locked state may facilitate the transition from internally regulated network

activity to externally driven sensory processing required for successful task performance.

HFAbs synchronized brain-wide and their network-level synchronization identified

functionally specialized subnetworks

We found that HFAbs evoked by cue and target demonstrated different topographical distributions,

largely non-overlapping responsive electrode populations. Additionally, HFAbs transiently

decoupled from local low-frequency rhythms during task events, suggesting reduced local

rhythmic constraints during active processing. The distinct spatial patterns and release from local

phase-locking raise the possibility that HFAbs may instead synchronize with HFA in distant

regions processing similar information. We therefore asked whether brain networks could be

divided to distinct subnetworks based on the temporal relationships of their HFAbs, and whether

subnetworks with similar synchronization patterns are functionally specialized.

We considered HFAbs outside the cue/delay and target/response periods to avoid stimulus-driven

synchronization effects and measured the power of high frequency activity (65-175 Hz, HFA, to

capture broader spectral contents) in one electrode aligned to the center of HFAb events in another

electrode (HFAb-triggered HFA). HFAb-triggered HFA between electrodes showed synchronized

high-frequency amplitude increases (**Fig. 3a,** similar results were observed for HFAb-HFAb).

These synchronized patterns exhibited rhythmic modulation at low-frequency rhythms, with

spectral peaks predominantly in the theta band (4-8 Hz) (**Fig. 3b**, top). These patterns were

consistent across subjects and experiments (**Fig. 3b**, bottom, **Extended Data Fig. 4a,b**). HFAb

synchronization strength (the sharpness of HFAb-triggered HFA) was inversely related to inter-

electrode distance, with closer sites showing stronger synchronization (**Fig. 3c**, **Extended Data**

Fig. 4c, Spearman correlation between HFAb-triggered HFA kurtosis and electrode distances, $r =$

$-0.25, p < 0.001$).

To extract temporal relations of high-frequency activity across electrode pairs, we used Principal

Component Analysis (PCA) to reduce the dimensionality of HFAb-triggered HFA. The first

component (PC1) reflected zero-lag synchronized and near-symmetric distributions of high-

frequency activity in all subjects, explaining over 20% of the total variance (**Fig. 3d,** see **Extended**

Data Fig. 4d for experiment 2). We projected HFAb-triggered HFAs onto PC space and used a

resampling-based consensus K-means clustering technique to identify robust clusters of electrodes

based on their scores on the synchronized PC (**Extended Data Fig. 4f**, see **Methods**). The

clustering algorithm identified the most stable clusters (electrodes consistently grouped together

across multiple runs) for each cluster number (K = 2-8, **Fig. 3e, Extended Data Fig. 5**). A series

of accuracy metrics was used to determine the optimal number of clusters for each subject through

a voting poll, selecting the cluster number that outperformed others across the majority of metric

(**Extended Data Fig. 6a-c**).

Next, we examined whether these clusters were functionally specialized, revealing functional

subnetworks within the larger brain networks. All subjects (except one excluded due to insufficient

electrodes) showed clusters with distinct cue- and target-evoked activation profiles (**Fig. 3f,** see

Extended Data Fig. 6d for individual examples). Clusters activated by cues and targets were

labeled as *cue-* or *target-subnetworks*, respectively (Wilcoxon signed-rank test, $p < 0.05$; FDR

corrected). Cue-subnetworks were predominantly located in occipital and parietal regions (e.g.,

IPS, TPJ, SPL, IPL), whereas target-subnetworks were more widely distributed across different

brain areas including parietal, motor, premotor, and frontal cortices (**Fig. 3g**, see **Extended Data**

Table 1 for individual subject electrode locations).

Lastly, similar to **Fig. 1h**, we tested whether cue responses in cue- and target-subnetworks could

predict trial outcomes. HFAb density averaged across electrodes in cue-subnetworks within 98-

374 ms post-cue predicted successful detection of upcoming targets (binomial test, $P < 0.001$, FDR

corrected for dependent samples). This prediction was specific to cue-subnetwork (**Fig. 3h**) and

was not driven by a single subject (see **Extended Data Fig. 6e-g** for classifier details in both

experiments). It is worth noting that HFAbs in both cue- and target-subnetworks, predicted trial

outcome following target onset (**Extended Data Fig. 6h**).

As a control, we re-referenced datasets from common average referencing to local composite

referencing (LCR, each electrode was referenced to its nearest neighbors, see **Methods**), This re-

referencing did not alter any of the main results. Together, these findings suggest that large-scale

brain networks show synchronized HFAb patterns, with their synchronization identifying

functionally specialized subnetworks that exhibit distinct temporal dynamics. Furthermore,

HFAbs in cue-subnetworks following cue onset were predictive of performance accuracy.

HFAbs in cue-subnetworks precede target-subnetworks

Our observation that HFAb responses in cue-subnetworks predict performance suggests their

involvement in target detection, raising a key question: do cue-subnetworks route information to

target-subnetworks during target processing, implying directional flow that supports attentional

performance? To address this question, we used two different approaches (only subjects whose

electrode coverage included both cue- and target-subnetworks were considered; n = 3 in

experiment 1, and n = 3 in experiment 2). First, we measured HFAb-triggered HFA during target-

to-response period and tested whether HFAbs showed temporally ordered activity between cue

and target-subnetworks within this window. On average, HFAbs in cue-subnetworks preceded

HFA in target-subnetworks during target processing (**Fig. 4a**, permutation test, p < 0.05, see

Extended Data Fig. 7a for individual example). We further quantified the distribution of HFAb-

triggered HFA peaks for individual subjects. The HFA peak distribution in target-subnetworks

showed a mean at 39.7 ± 4.4 ms (n = 3) after HFAbs in cue-subnetworks and was significantly

different from the opposite direction (**Fig. 4b**, Wilcoxon rank-sum test, p < 0.05). Similar results

were found in experiment 2 (**Extended Data Fig. 7b**). For comparison, no lead-lag patterns were

observed during the cue-to-target interval **(Extended Data Fig. 7c).** Topographically, and at the

level of individual subjects, HFAbs in the occipital, posterior parietal, and frontal areas led over

the motor/premotor areas during target processing (**Fig. 4c**, see **Extended Data Fig. 7d** for more

examples).

Next, we used delayed mutual information (DMI) to further quantify temporal dependencies

between cue and target-subnetworks. DMI revealed (i) when cue and target-subnetworks shared

maximum information relative to target onset, and (ii) time-lags of maximum inter-predictability.

DMI peaked 285.4 ± 18 ms after target onset (**Fig. 4d**; n = 3). Time-lag distributions showed cue-

subnetworks preceded target-subnetworks by 48.8 ± 23 ms during target processing (**Fig. 4d,**

Wilcoxon signed-rank test, $p < 0.001$, n = 3; see **Extended Data Fig. 7e** for individual heatmaps).

The temporal precedence occurred only after target onset, not during cue processing (**Extended**

Data Fig. 7f), and specific to valid-cue trials, indicating it is not merely visual hierarchy activation,

but reflects routing only when cue information is relevant for target detection (**Fig. 4e**). This

pattern was consistent across individuals and both experiments (**Extended Data Fig. 7e-h**).

Overall, these results demonstrate that subnetworks, identified by their network-level HFAb

synchronization patterns show temporal sequences during task events. Specifically, when cues

provide valid spatial information about the target, HFAbs in cue-subnetworks precede those in

target-subnetworks following target onset.

Computational modeling of HFAbs through spiking neural networks

We first developed a computational model using spiking neural networks to investigate both the

mechanisms underlying HFAbs and their functional role in attentional performance. We simulated

two interconnected networks, each consisting of 1000 neurons (80% excitatory, 20% inhibitory;

Fig. 5a, Extended Data Fig. 8a, see **Methods**). Each network was fed by external input currents

primarily to the excitatory population.

We implemented two recording sites on top of each network measuring electrical field dynamics

of postsynaptic and transmembrane potentials (**Fig. 5a,b**, **Extended Data Fig. 8b,c,** see **Methods**).

Simulated iEEG signals showed low-frequency spectral peaks corresponding to network resonance

frequencies and high-frequency (65–115 Hz) peaks, consistent with experimental data (**Fig. 5c**).

To investigate the neural mechanisms underlying HFAbs, we calculated the spike density for the

population of neurons, and the HFAb density from each recording site. HFAb density correlated

with spike density compared to random timings (randomization test, $p < 0.05$) consistent with

previous empirical findings.[25,28,38] When feeding the network different levels of external non-

rhythmic current coherence (**Extended Data Fig. 8c**), we observed a linear increase in the

correlation between HFAb and spike densities as input coherence increased (Spearman correlation,

$r = 0.87$, $p < 0.001$, **Fig. 5d**). These findings suggest that HFAbs emerge when coherent external

inputs drive neural populations into elevated firing states, indicating that HFAbs reflect input-

driven population excitability rather than spontaneous fluctuations in neural activation.

Next, we examined HFAb phase-locking to slower rhythms. HFAbs triggered spectral peaks and

showed phase locking to theta, alpha and beta bands (4-25 Hz, **Fig. 5e**), depending on synaptic

time constants, external input strength, and neural connectivity strength, consistent with previous

computational findings.[39] To test network mechanisms underlying decoupling during cue and

target processing, we modeled feedforward networks and measured time-resolved PLV in both

networks after feeding one with a brief input pulse (~50% of neurons receiving brief in-phase

inputs; **Fig. 5f, Extended Data Fig. 8e,f**). The external impulse increased HFAb rate and

desynchronized both networks, resulting in transient decoupling of HFAbs from low-frequency

rhythms (Wilcoxon rank-sum test, $p < 0.001$, **Fig. 5i, Extended Data Fig. 8g,h**). These results

reveal that external inputs shift neural networks from internally regulated states (HFAbs phase-

locked to slow rhythms) to externally driven states, mirroring the empirical decoupling observed

during cue and target processing.

We also examined HFAb synchronization patterns between networks. Reciprocally connected

networks and networks with correlated external inputs showed synchronized high frequency

activity (**Fig. 5g**), while feedforward networks showed lead-lag patterns within 150 ms of HFAbs

(Wilcoxon rank-sum test, $p < 0.001$, **Fig. 5h**). Overall, these inter-network HFAb patterns showed

intrinsically generated rhythmic fluctuations, though rhythmic external inputs could also entrain

its rhythms to HFAb synchronization patterns between networks (Spearman correlation, $r = 0.83$,

$p < 0.001$, **Fig. 5i**). These findings suggest that inter-network HFAb patterns reflect both structural

connectivity and shared input statistics, providing mechanistic support for the zero-lag

synchronization within subnetworks and lead-lag patterns between subnetworks observed

empirically.

To link network dynamics to behavioral performance, we extended our model to a four-network

architecture with spatially selective cue networks connected to target networks. All networks were

activated by a shared decaying input initiated at trial onset. Cue networks responded transiently to

cue onset and decayed thereafter. Target presentation was modeled as an additional input to the

target networks. If targets appeared at cued locations, cue networks showed weak reactivation (**Fig.**

5j, Extended Data Fig. 8i,j, see **Methods**). We ran the model for 150 trials with valid and invalid

cue conditions. Valid cues increased burst response in target networks following target onsets

(Wilcoxon rank-sum test, p < 0.001, **Fig. 5k**). Delayed mutual information analysis revealed that

cue networks temporally preceded target networks specifically for valid trials, with maximal

information transfer occurring after target onset (**Fig. 5l, Extended Data Fig. 8k**), mirroring our

empirical findings of cue-subnetwork precedence. We implemented a detection threshold model

to test how cue-driven input enhances target detection. When fitting detection thresholds to invalid

trials, valid cues consistently increased HFAb rates above threshold, resulting in higher detection

probability (**Fig. 5m, Extended Data Fig. 8l**). This provides a mechanistic account of how cue

information enhances behavioral performance through increased state transition probability by

inputs from cue networks.

Our modeling results demonstrate that HFAbs reflect transitions in neural populations into

spiking states. HFAbs are phase locked to low-frequency rhythms, but transient inputs disrupt this

coupling. HFAb synchronization patterns between networks depend on connectivity and task

structure, with behavioral relevance demonstrated through enhanced detection rates when cue-

driven inputs increase target network burst rates.

Discussion

Here we present a framework for understanding mechanisms of large-scale information routing in

brain networks at fast timescales. We characterize high-frequency activities as discrete burst events

(HFAbs), rather than continuous fluctuations, supported by computational and animal

electrophysiology studies emphasizing their circuit-level origins and roles in long-range

communication.[6,40–44] This quantification allows a millisecond-resolution, network-level account

of selective attention that extends beyond traditional pairwise interareal interactions. Our findings

revealed that HFAbs were synchronized across distributed cortical networks, identifying

functionally specialized subnetworks for cue and target processing. Importantly, HFAbs in cue-

subnetworks following sensory cues predicted trial-by-trial performance accuracy and temporally

preceded HFAbs in target-subnetworks during target processing when cues were informative.

Complementing these findings, our computational modeling showed HFAbs emerge from bouts

of elevated excitatory drive to local networks, marking fast state transitions in neural populations

that support attention processes.

Our analysis of both exogenous and endogenous attention tasks revealed conserved mechanisms

despite distinct control pathways. In both tasks, valid cue information enhanced target-triggered

HFAb responses, and cue subnetworks showed temporal precedence over target subnetworks only

when the cue was informative. These preserved findings across both tasks suggest that information

routing signified by HFAb dynamics represents a fundamental mechanism for transforming cue

information into spatial priority signals, regardless of whether attention is stimulus-driven or top-

down controlled.[45,46] However, task-specific differences emerged in the timing of lateralized

processing; exogenous attention showed laterality effects during cue processing, reflecting

stimulus-driven nature of peripheral cues, while endogenous attention showed laterality only

during target processing, consistent with the internal maintenance and deployment of spatial

attention from symbolic cues.[46] These findings support the argument that while attentional control

signals may rely on different cortical pathways, more posterior for exogenous and involving frontal

regions for endogenous attention, they converge onto common selection mechanisms in parietal

and sensory areas.[47,48]

A main finding of our study revealed that the most prominent pattern of high-frequency dynamics

in the brain network was zero-lag synchronization (**Fig. 3d**), consistent with previous studies

showing phase locking of high-frequency dynamics in brain networks.[31–33] Zero-lag

synchronization patterns are thought to enhance large-scale information processing and facilitate

representational states of sensory information in the brain.[49–51] While our results cannot directly

identify the origins of these synchronized interactions, two mechanisms might explain these

patterns: subcortical inputs,[52–55] particularly higher-order thalamic nuclei,[52,56–58] or direct

intracortical interactions.[29,59–61] Our modeling supports both these possibilities showing correlated

inputs to distinct neural networks, or reciprocal connections between spiking networks could drive

such long-range synchronization (**Fig. 5g**).

We further showed that network-level baseline synchronization of high-frequency dynamics

reliably identifies distinct functional brain subnetworks. While the baseline state did not contain

cue or target processing periods, it could identify subnetworks with unique activation patterns in

response to those behavioral events (**Fig. 3**). This approach diverges from conventional methods

for delineating brain network organizations and communications, such as fMRI, which primarily

focuses on low-frequency BOLD signal fluctuations.[62,63] By leveraging high-frequency dynamics,

we demonstrate a link between baseline neural activity and task-induced network activation

patterns, suggesting that intrinsic network fluctuations can affect behavioral responses at

millisecond timescales. Identifying these subnetworks at fast time scales is critical to capture rapid

state transitions in brain networks, which may underlie the temporal structures necessary for

adaptive information routing during behaviorally relevant events.[64] Our findings demonstrate that

brain subnetworks, identified by their network-level HFAb synchronization, exhibit lead-lag

patterns during behavioral events. Specifically, cue-subnetworks preceded target-subnetworks

during target presentation when spatial cues provided relevant information about target locations

(**Fig. 4**). Our modeling provided a mechanistic account for this, showing that inputs from cue

networks during target processing increase the probability of state transitions in target networks,

reflected by elevated HFAb rates (**Fig. 5k,m**). Overall, these observations uniquely bridge the gap

between studies focusing on local and cross-regional mechanisms of fast information

routing[6,36,41,42,44,56] and those examining network dynamics beyond pairwise interactions,[15,65]

providing new insight into the mechanisms of selective attention.

Another key observation was the coupling dynamics of HFAbs and local low-frequency activities.

We found that phase-locking of HFAbs to low-frequency dynamics occurs even with random

external input patterns in modeling, and this is a widespread phenomenon in iEEG signals,

consistent with previous studies.[30,66,67] However, our findings further showed that HFAbs

transiently decouple from local low-frequency phases in response to both cue and target events

(**Fig. 2**). Our modeling supports this observation, showing that transient inputs interrupt the default

state coupling of HFAbs to low frequencies (**Fig. 5f**). These network interferences, characterized

by brief and strong responses, can temporarily induce local cross-frequency decoupling, possibly

by increasing response heterogeneity and disrupting the timing of inhibitory and excitatory neuron

activation.[68] This desynchronization may enhance cognitive processes such as selective attention,

perception, and memory retrieval by facilitating the processing of new information and

suppressing internally regulated activity states.[68–71] We observed that this decoupling,

accompanied by increased HFAb density following cue and target events, resembled a

desynchronized up-state, a brain state characterized by activated but desynchronized neural

activity across cortical networks. Importantly, this decoupling was more pronounced in correct

trials during both cue and target processing, suggesting that desynchronization correlates with

accurate cognitive performance (**Fig. 2f**).

It is worth noting that the HFAbs identified in this study share spectral features with cortical ripples

recently described in human studies,[33,38,72] However, HFAbs exhibit different characteristics

suggesting specialized functions. While both show synchronization and correlate with population

spiking, HFAbs in our data occurred ~15-fold more frequently than ripples, with shorter durations

(~36 vs 50-100 ms).[33,72] Ripples are largely described in support of memory maintenance with

consistent phase-locking to slow waves,[72,73] whereas HFAbs show performance-dependent

decoupling from oscillations and directional lead-lag patterns between functional subnetworks.

These distinctions suggest that ripple-band high-frequency bursts constitute a general population-

level mechanism that can flexibly switch between binding (memory) and routing (attention)

modes, with HFAbs instantiating the routing mode during active sensory processing.

In summary, our study provides a multilevel approach to understanding large-scale cortical

communications, showing that HFAbs are markers of fast population state transitions that enable

selective information routing through zero-lag synchronization within subnetworks and lead-lag

temporal sequences between subnetworks.

Acknowledgments

This work was supported by the C.V. Starr Fellowship (KBB), German Research Foundation,

Emmy Noether Program (DFG HE8329/2-1; RFH), NIBIB (P41-EB018783; PB), NINDS

(R01NS021135; RTK), and NEI (2R01EY017699; SK), NIH (1R01MH137624; SK), NIMH

(2R01MH064043, P50MH132642; SK). The funders had no role in study design, data collection

and analysis, the decision to publish, or the preparation of this manuscript.

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457 **Author Contributions**

458 Conceptualization, K.B.B. and S.K.; Methodology, K.B.B.; Formal Analysis, K.B.B.; Modeling,

459 K.B.B.; Investigation, R.F.H., R.T.K. and J.J.L.; Software, R.F.H. and I.C.F.; Visualization,

460 K.B.B; Writing – Original Draft, K.B.B.; Writing – Review & Editing, K.B.B., R.F.H., I.C.F.,

461 J.N.B., P.B., J.J.L., R.T.K., and S.K..; Supervision, S.K.

Competing Interest: The authors declare no competing interests.

Figure Legends

Fig. 1. HFAb activation profile predicts behavioral outcome on a trial-by-trial basis. a, Experiment 1 task outline: two bars appear on the screen, followed by a transient cue indicating the likely target location. After a variable delay, a near-threshold contrast change appears at the cued or, less often, at an equidistant uncued location. In 10% of trials no target appears. Subjects report the change detection. **b,** An example trial showing detected high-frequency bursts (HFAbs, in red) around the cue onset. The brain shows localization of electrodes across all subjects in

experiment 1. **c**, Normalized HFAb density for individual subjects and the group average, aligned
to the cue (left) and target onset (right); shaded error bars denote mean $\pm$ SEM. **d**, HFAb responses
to cues (top) and targets (bottom) grouped by trial outcome (n=6, median $\pm$ SEM, each line
represents one subject, with thick lines showing group average, horizontal lines showing
significant differences between groups (Dunn's test, $p < 0.05$), and asterisks denoting non-zero
responses, Wilcoxon signed-rank test, *: $p < 0.001$). **e**, Correlation of HFAb responses to cue and
target, for channels with significant activation to either or both cue and target (each point represents
one channel, each line represents regression for one subject, the orange line shows regression
across all channels). **(e, g)** Group average heatmap 3D rendering of HFAb responses to **(a)** cues
and **(g)** targets for correct trials. **h**, An individual subject example of HFAb responses to cue and
target, for correct and incorrect trials (black circles indicate electrode locations). **(i,j)**, HFAb
responses to **(i**, GLME, $t = 4.7$, *: $p < 0.001$) targets for valid and invalid cue conditions and **(j**,
GLME, $t = 5.1$, *: $p < 0.001$) to cues ipsi- and contra-lateral to electrodes (n=6, median $\pm$ SEM).
k, Classifier accuracy in predicting trial outcome based on HFAb density around the cue onset for
cue responsive electrodes and cue unresponsive electrodes (error bars: SEM across realizations
and cross validations). Red lines indicate points significantly above baseline and chance (two-
sided binomial test, $p < 0.05$, FDR-corrected for multiple dependent comparisons).
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Fig. 2. HFAbs dynamically phase lock to low-frequency LFPs and decouple transiently after
cue and target onsets. a, HFAb-triggered LFP spectrum (top) and phase locking values (PLV,
bottom) across all electrodes in both experiments (each line represents one subject). **b**, Examples
of phase-frequency distribution of HFAbs for an electrode with theta phase locking (top) and beta
phase locking (bottom). Polar histograms of phase distributions at the phase-locking frequencies
are shown in the right-hand panels. **c**, PLVs for individual electrodes (each row)**.** Black dots
indicate the maximum peak of PLV for each electrode, and white dashed lines indicate subject
boundaries. Phase-locking peak distribution is shown above. **d**, Top: time-resolved PLV analysis
averaged across all subjects aligned to cue (left) and target (right) onsets in experiment 1. Bottom:
The proportion of time points where phase locking dropped significantly below the baseline
averaged across all electrodes (two-sided random permutation test, $p < 0.05$). Thick lines indicate
segments significantly different from the chance level (two-sided binomial test, $p < 0.05$, FDR-
corrected for multiple dependent comparisons; see **Methods**). **e**, Example subject brain heatmap
showing coupling ratio between HFAb and low frequency (4-25 Hz) LFPs after cue (top) and
target onsets (bottom) for correct and incorrect trials. **f**, Proportion of time points with significant
decoupling from low-frequency LFPs (4-25 Hz) after cue and target onsets for correct and incorrect
trials. **g, h**, Regression plots showing correlation of coupling ratios following cue onset with cue
responses (green, **g**), and coupling ratios following target onset with target responses (purple, **h**).

Scatter points denote electrodes, lines indicate individual subjects with orange line showing the
regression across all subjects.
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Fig. 3. Network-level synchronization of HFAbs identifies functionally specialized subnetworks. a, HFAb-triggered HFA measured between electrode pairs during baseline periods (outside cue/delay and target/response epochs). Top: example electrode pair; bottom: group average across all pairs for individual subjects. **b**, Normalized power spectral density (PSD) of HFAb-triggered HFA for each electrode relative to all others (white dashed lines indicate subject boundaries), bottom: mean PSD across all electrodes for individual subjects (red dots denote peaks). **c**, HFAb-triggered HFA HFAb-triggered HFA as a function of inter-electrode distance quartiles (25, 50, 75, and 100 mm), colored from green (short) to red (long). **d**, The first two principal components of HFAb-triggered HFA for individual subjects. **e**, Example subject showing pairwise grouping probability matrix for optimal K=8 clusters (top, white lines indicate cluster boundaries, clusters are ordered by stability, see **Methods**) and corresponding electrode topography (bottom). **f**, HFAb density around cue and target onsets for each cluster from panel **e** (mean ± SEM; thick lines indicate significant responses, $p < 0.05$, two-sided Wilcoxon test, FDR-corrected for multiple dependent comparisons). **g**, Group topography of electrodes, in clusters with significant cue responses ("cue-subnetworks") and target responses ("target-subnetworks") across all subjects. **h**, Performance classifier trained on mean HFAb density across electrodes in cue-subnetworks (purple) and target-subnetworks (green) around cue onset (error bars: SEM across

535 realizations and cross-validations). Red lines indicate points significantly above baseline and
536 chance (two-sided binomial test, $p < 0.05$, FDR-corrected for multiple dependent comparisons).

Fig. 4. Cue-subnetworks precede target-subnetworks during target processing. a, Group-average HFAb-triggered HFA during target processing showing temporal relationships between subnetworks (mean ± SEM): HFA in target-subnetworks aligned to HFAbs in cue-subnetworks (purple) versus HFA in cue-subnetworks aligned to HFAbs in target-subnetworks (green). Shaded regions indicate significant lead-lag patterns (two-sided permutation test, $p < 0.05$; red: cue leads, [65 313] ms, blue: target leads, [-250 -37] ms). **b,** Time-lag distributions of HFAb-triggered HFA peaks for individual subjects (medina ± SEM; n=3, two-sided Wilcoxon rank-sum test between directions; *: $p < 0.05$, **: $p < 0.01$, ***: $p < 0.001$). **c,** An individual example of topographic visualization of lead-lag patterns between cue and target subnetwork electrodes averaged across cluster numbers during target processing period (see **Extended Data Fig. 7** for more examples). The lead and lag temporal patterns are shown in a color gradient from red to blue. **d,** Group average heatmaps of normalized delayed mutual information (DMI) heatmaps showing temporal dependencies when shifting cue-subnetwork relative to fixed target-subnetwork (left) and vice versa (right). The white line indicates the time-lag distribution of DMI peaks across electrode pairs. The cross signs indicate the median of DMI peak time-lags ± SEM. **e,** Comparisons of DMI peak distributions between the two opposite directions (green for shifting the cue-subnetwork and purple for shifting the target-subnetwork) for valid (right) and invalid cue (left) trials. Individual subjects shown as crosses (n=3, median ± SEM across electrode pairs).

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Fig. 5. Computational modeling of HFAbs in spiking neural networks. a, Two interconnected
cubic networks of point source 1000 spiking neurons 1000 (80% excitatory, 20% inhibitory) with
recording electrodes measuring field potentials above each network (bottom: raster plots) **b**,
Example trial showing simulated iEEG signal, population spike density with raster, detected
HFAbs (red), and resulting HFAb density. **c**, The power spectral density (PSD) of iEEG signals.
d, Correlation between HFAb and spike density increases linearly with external input coherence.
e, HFAb phase distribution at different frequencies (bottom), and Phase-locking value (PLV)
across all HFAbs (top). **f**, PLV of HFAbs to low frequencies following transient input (thick black
line). **g,h**, HFAb-triggered HFA between reciprocally connected (**g**) and feedforward (**h**) networks
with varying connectivity strengths (purple and green traces show HFAb-triggered HFA when
HFAbs are extracted from N1 and N2, respectively). **i**, PSD over the HFAb-triggered HFA for
rhythmic inputs of varying frequencies. **j,** Four-network model simulating spatial attention task:
two spatially selective cue networks (N1, N2) with dominant feedforward projections to target
networks (N3, N4); cue and target networks activate to cue and target onsets, respectively, cue
networks reactivate when targets appear at cued locations. **k**, HFAb responses in cue networks to
cue onset (left) and target networks to target onset (right) for valid (purple) and invalid (green)
trials (mean ± SEM). **l,** Delayed mutual information heatmaps between cue and target networks

for valid (left) and invalid (right) conditions showing temporal precision of cue over target
networks following target onset. **m,** Detection rates as a function of HFAb threshold demonstrating
enhanced detection for valid versus invalid trials.

Methods

<u>**Experimental model and subject details**</u>

Participants. Intracranial recordings were obtained from 12 epilepsy patients who underwent pre-

surgical monitoring with implanted grid electrodes. Study 1 included seven patients (35.99 ± 12.42

years; mean $\pm$ SD; 5 females; see[23] for further details). Patients were recruited from the University

of California, Irvine Medical Center, USA (n = 6) and California Pacific Medical Center (CPMC),

San Francisco, USA (n = 1). Study 2 included 5 patients (30.20 ± 1186 years; mean $\pm$ SD; 1 female,

3 patients were excluded from the original study due to their limited electrode coverages; see[3] for

further details) from Johns Hopkins Hospital in Baltimore, MD, USA (N = 1) and Stanford

Hospital, CA, USA (N = 4). The electrode placement was entirely guided by clinical

considerations, and all patients provided written informed consent to participate in the study. All

procedures were approved by the Institutional Review Board at each site, as well as the Committee

for Protection of Human Subjects at the University of California, Berkeley (Protocol number:

2010-02-783) and were in accordance with the Declaration of Helsinki.

<u>**Experimental design and procedure**</u>

Behavioral tasks. Participants performed a spatial attention task in each experiment. In

experiment 1, participants performed a variant of the Egly-Driver task;[74,75] see[23] for further details)

Behavioral data were collected with Presentation Software (Neurobehavioral Systems). Subjects

were seated approximately 60 cm from the laptop screen. Subjects started each trial by pressing a

left mouse button. On each trial, after appearance of a fixation cross, two bars appeared vertically

or horizontally on the screen, followed by a brief spatial cue (100 ms), presented after 400-800 ms

at one of the four corners of the bar stimuli. The spatial cue indicated the location where the target

was most likely to appear (72% cue validity) and occurred pseudo-randomly in any of the four

quadrants. A variable cue-to-target interval (500 – 1700 ms) was introduced after the cue during

which participants sustained spatial attention at the cued location. Targets could randomly appear

at any point during the cue-target interval, and participants released the mouse button to report a

detected target. Infrequent catch trials (10%) during which no actual target appeared were used to

track the false alarm rate. Auditory feedback indicated whether the trial was performed correctly.

The target luminance was adjusted every 15 trials, if necessary, by increasing/decreasing the RGB

value, in order to achieve an overall approximate accuracy of 80%. The experimenter monitored

continuous fixation. All participants responded by using the hand contralateral to the implanted

grid, except for participant S5 who had bilateral grids and responded by using the left hand.

Participants performed up to 5 blocks of 60 trials each (190 trials ± 67; mean ± SD).

Experiment 2 (Extended Data Fig. 1) used EPrime software (Psychology Software Tools) to

control stimulus presentation (see Szczepanski et al., 2014 for more details). Subjects were seated

approximately 60 cm away from the laptop screen. Each trial began with red circles (distractors)

dynamically switching on and off on a dark background. A spatial cue guided participants to the

right or left hemifield, and the cue remained on the screen throughout the trial. Subjects were

instructed to maintain fixation and only covertly shift their attention to the cued hemifield. Through

the trial, the experimenter monitored eye movements and ensured central fixation. After a variable

cue-target interval (1000 – 2000 ms), a blue square target appeared on the screen (~62/38% on the

cued/uncued hemifields). The target remained on the screen until the subject responded or the trial

ended (2000 ms timed out). Subjects were asked to report a target seen in a cued hemifield by

pressing a button while withholding a response if the target was seen in a non-cued hemifield.

Targets appeared randomly during the cue-target interval. Three out of five subjects responded

with the hand ipsilateral to the grid. Each participant completed six blocks (each 42 trials). The

experimenter monitored eye movements and ensured central fixation throughout both experiments.

ECoG data acquisition. Electrophysiological and peripheral (photodiode) data were collected

using a Nihon Kohden recording system at UC Irvine, CPMC and Children's Hospital (128/256

channel, 1000/5000 Hz sampling rate), a Tucker Davis Technologies recording system at Stanford

(128 channel, 3052 Hz sampling rate), or a Natus Medical Stellate Harmonie recording system at

Johns Hopkins (128 channel, 1000 Hz sampling rate).

Electrode localization. In experiment 1, the electrodes were localized by transforming both the

pre-implant MRI and the post-implant computed tomography CT into Talairach space. For all

subjects, MNI coordinates were also calculated for each electrode location, which was used for

group-level visualizations.[23] In experiment 2, post-implant CT was aligned to the pre-implant MRI

and all were transformed into MNI space across subjects.[3] Strip or grid electrodes were implanted

with 1 cm spacing. One participant (S5) had an additional 8 contact depth probe inserted into the

occipital cortex. Electrode positions were primarily determined using the VTPM atlas.[76] For

electrodes without an assigned label, the process was repeated using the AFNI atlas.[77] The assigned

positions were manually verified and adjusted based on electrode reconstructions visualized in

native Talairach space. Electrodes near the Temporoparietal Junction (TPJ) were manually

localized, as TPJ definitions were not available in either the VTPM or AFNI atlases (see Helfrich

et al., 2018 for more details).

iEEG Data. Preprocessing: All intracranial EEG channels were manually examined by a

neurologist for epileptiform activity and artifacts. Affected channels and epochs were excluded.

The raw data was preprocessed using the EEGLAB and Fieldtrip toolbox Field Trip toolbox[78]

Version 20230118 in MATLAB, The Mathworks Inc. R2023b; https://www.mathworks.com/.

Preprocessing included notch filtering at 60 Hz and all harmonics, as well as referencing the data

to the common mean of electrodes.[3,23] Then, the data was time locked to individual trials. Trials

were 8 seconds long, -3 to +5 seconds around cue onsets in the experiment 1 and -2 to +6 seconds

around cue onsets in the experiment 2. As a control, we re-referenced the datasets from common

average referencing to local composite referencing (LCR), a spatial Laplacian estimate relative to

nearest neighbors. This re-referencing did not alter any of the main results.

<u>**Analysis of high frequency activity bursts (HFAbs)**</u>

HFAb detection and density analysis. We adopted an adaptive burst detection approach, similar

to previous work,[6] to identify high-frequency oscillatory bursts (65-115 Hz) at each electrode. The

frequency band was selected based on spectral peaks observed across all individuals (87 Hz trials

± 14; mean ± SD) and in the modeling. Similar analyses were conducted for a broader band (65-

175 Hz) and the low gamma band (35-65 Hz). While the broader band produced similar results,

the low gamma band yielded inconsistent responses and noisy clustering. We selected the

frequency band based on spectral peaks across individuals and in the modeling. First, we applied

a zero-phase Butterworth bandpass filter to the padded signal and then calculated the analytic

signal x(t) using the Hilbert transform. We extracted the instantaneous amplitude as the real part

of the analytic signal $z(t)$ following eq. 1:

$z(t) = x(t) + iy(t) = a(t)e^{i\theta(t)}$ eq. 1

where $y(t)$ is the Hilbert transform of $x(t)$:

$y(t) = \frac{1}{\pi} \int_{-\infty}^{+\infty} \frac{x(\tau)}{t-\tau} d\tau.$ eq. 2

The instantaneous energy [79] $IE(t)$ of the signal is calculated from its Hilbert transform:

$IE(t) = \int_0^{\omega_N} H^2(t, \omega) d\omega$ eq. 3

$\omega(t) = \frac{d\theta(t)}{dt}$ eq. 4

where $\omega(t)$ corresponds to instantaneous angular velocity. If the frequency band is narrow and if

the instantaneous frequency (eq. 4) is small enough, it can be approximated by squared $a(t)$, which

is the instantaneous amplitude. The marginal mean energy of the signal then can be estimated as

eq. 5:

$\overline{IE} = \frac{1}{T} \int_0^T a(t)^2 \, dt$ eq. 5

where T represents the duration of the signal. For skewed energy distributions in empirical data,

we determined a median absolute deviation (MAD) threshold by setting it to 3.3 times plus the

median energy.

To qualify as a burst, the energy level had to exceed this threshold for at least 1.5 cycles of the

upper bond frequency, and the instantaneous amplitude needed to surpass $\sqrt{2}$ times the RMS of

the peaks. Burst boundaries were marked by the closest points to a burst peak where either the

instantaneous energy fell below the signal's mean energy or the deviation in instantaneous

frequency exceeded the mean change plus two standard deviations. This was to exclude multi-

component or noisy events. Bursts were considered significant if their duration was at least 2.5

times the upper bond frequency cycle and exceeded the average span of adjacent local minima of

the energy function. Finally, when bursts were too close (less than five frequency cycles apart),

only the burst with the higher energy peak was kept. For comparison to ripple, we also quantified

ripples in our data by calculating z-score of instantaneous energy, and finding events lasting longer

than 3 cycles above a value of 2.5.[33] We observed extremely rare events in our data with ripple

characteristics.

To estimate how HFAb events are distributed over time, we calculate the HFAb density by

convolving a vector of burst events with a gaussian window of 500 ms and a standard deviation of

100ms. After estimating the HFAb density for each channel and trial, we investigated whether the

cue and target stimuli affected burst density. For each electrode, the average burst density aligned

to the cue and target onsets were calculated separately. Following that, we normalized timeseries

to the baseline (1000 ms before each event onset) by subtracting the mean and dividing it by the

standard deviation (see **Fig. 1c**). This is a similar approach to the peri-stimulus time histogram

(PSTH) for spike trains [80].

Visualization of individual and group average responses on 3D brains. To visualize how HFAb

responses were topographically organized in the 3D brains, we plotted both individual (**Extended**

Data Fig. 1f) and group responses to cue (**Fig. 1f**) and target onset (**Fig. 1g**). Each electrode value

was calculated as the mean of Z-scores within 500 ms of cue and target onset. For individual

subjects, the plotted value for each electrode was linearly attenuated with distance from the

electrode and for a sphere of 1 cm radius (illustrating correct versus incorrect trials separately, **Fig.**

1h, **Extended Data Fig. 1f**). The group average plot (**Fig. 1f,g, Extended Data Fig. 2e**) was

calculated using electrode locations from both experiments to better cover the entire brain. The

MNI coordinates of electrodes were used for rendering on a template brain (subject S4 from

experiment 1 and subject S12 from experiment 2 were excluded due to suboptimal wrapping of

Tal to MNI spaces). We set the value of all mesh surfaces to zero for each subject. Then, similar

to the plotting for individual data, we calculated the value for mesh surfaces by using linear

attenuation (the sphere radius was set to 2.5 cm in order to achieve a smoother visualization of the

whole brain). As a last step, we averaged the surface values across all subjects to find consistent

patterns of activations for the cue and target. We plotted the data onto 3D brain for correct and

incorrect trials separately.

Statistical analysis of HFAb responses to cues and targets events. After calculating the burst

density and finding PSTH for each channel, we examined whether the HFAb responses evoked by

cues (within a window of 500 ms after cue onset) and targets (750 ms after target) were

significantly different from baseline. This analysis was done at the network level. The average

responses of electrodes within the defined time window were determined using the normalized

HFAb density for each channel. Wilcoxon signed-rank tests were then used to examine if there

was a non-zero response at the network level to cue and target onsets. Also, we performed a linear

regression analysis of the cue and target responses in electrodes under the null hypothesis that the

cue and target responses are independent (**Fig. 1e**).

To test whether cue and target responses were different when grouped by trial outcomes, we used

the Kruskial Wallis test under the null hypothesis that cue, and target responses do not differ by

outcome condition. For pairwise comparisons between different groups, we used Dunn's test [81]

with Tukey-Kramer multiple comparison correction (**Fig. 1d**).

Identifying cue-responsive electrodes. We identified Cue+/- electrodes by determining increased

HFAb rate or density profile within window of 500 ms after cue onsets. For each electrode, we

compared the average HFAb density following cue onset with that at baseline using Wilcoxon

rank-sum test. Electrodes with significant increases in HFAb rate after cue onset were labeled as

Cue+ electrodes, while the remaining electrodes were labeled as Cue- electrodes, which was

subsequently used in the classification (**Fig. 1k**).

Generalized Linear Mixed Effect Models. We used generalized linear mixed effects (GLMEs)

models to examine the effects of each task variable on HFAb response to cue and target events.

Three main predictors were used for the independent variables: *Laterality* (if the cue was either

ipsilateral or contralateral to the electrode) with two levels (*Ipsi* and *Contra*), *Validity* (if the target

appeared at the cued location) with two levels (*valid* and *invalid*), and trial outcome with four

levels (*Correct hit, False alarm, Correct Rejection, Miss*). Since not all subjects had trials for all

four outcome conditions, an alternative analysis also considered outcome as a binary variable

(*Correct* and *Incorrect*) with a logit link function. The random effect was determined by

considering groups as subject and channel labels. The response variable was either the mean burst

density in response to the target or the cue. The GLME is then formalized as shown in eq. 6:

$Response_{cue/target} = Laterality + Validity + Outcome + (1|Subject) + (1|Channel) +$

$b + \varepsilon$ eq. 6

Training classifiers for predicting trial outcome based on HFAb density around cue onset. A

time-resolved classification approach was used to determine how accurately HFAb responses

evoked by sensory cues could predict trial outcomes. First, we selected electrodes that showed a

significant HFAb responses to the cue presentation (*Cue+* electrodes, see above), for which we

then calculated the average HFAb density for *Cue+* electrodes for each trial and subject. Next, we

tested whether the average burst density in sliding windows around the cue onset could predict the

outcome at the trial level with a sliding window of 350 ms and a step of 25 ms. We used binary

Support Vector Machine (SVM) with one-to-one comparisons of HFAb density in sliding windows

with five folds of cross-validation. For training each SVM, a vector of the HFAb density in each

sliding window was used along with a vector of outcome labels (*"Correct"* = 1, *"Incorrect"* = 0).

The classifier used a Gaussian radial basis function kernel with a scaling factor of one. We assured

that the number of correct and incorrect trials used in the training was identical, to prevent sample

size biases. As the number of incorrect trials were lower than the number of correct trials, we

randomly sampled correct trials with a sampling size equal to the number of incorrect trials in each

fold. For each sliding window, we trained the classifier 1000 times and calculated the average

accuracy and confusion matrix across all iterations and folds. (**Fig. 1k).**

To determine whether HFAb density after cue onset could more accurately predict trial outcome

than chance and baseline, we used a binomial test on the accuracy of each window against the

maximum value of baseline and chance-level prediction accuracy. After obtaining a p-value for

each binomial test, we corrected the p-values using false discovery rate (FDR) for dependent

samples with a 0.05 alpha level [82] to correct for multiple comparisons error rate across all windows.

We performed this procedure separately for each experiment (**Fig. 1k**). In our main analysis, we

pooled data across all subjects in each experiment due to the low number of trials per subject.

However, to ensure that the results were not solely dependent on one subject, we ran a control

analysis in which one subject was left out each time and the classifier was trained and tested.

Controlled analyses in both experiments demonstrated that our observation was not based solely

on one subject. We used a similar approach for training the classifier on cue and target-

subnetworks to train the classifier on cue and target-subnetworks, as further detailed in the

'Identifying synchronous subnetworks' section.

HFAb-triggered spectrum and Phase Locking Value (PLV) analysis. To understand the

spectral dynamics of HFAbs and local population activities, we performed HFAb-triggered

spectrum analyses using HFAb centers as discrete points. We estimated the PLV of points (burst

centers) to their local field activity dynamics (both in the modeling and electrophysiological data)

in order to investigate the phase synchronization of HFAbs within their local networks. We

calculated the HFAb-triggered spectrum using an adaptive window around HFAbs. We extracted

a window centered around each selected point (HFAb center), which covered 2.5 cycles of the

frequency of interest before and after the selected point. This window was then multiplied by a

Hanning window. We estimated the power spectrum for each window using the Fast Fourier

Transform (FFT) for frequencies ranging from 1 Hz to 100 Hz. To account for trial-wise power

variations, triggered spectrum estimates were normalized by dividing by total power. We averaged

spectrum estimates across all points and trials to calculate the final HFAb-triggered spectrum.

Following the analysis of the HFAb-triggered spectrum, a PLV calculation was performed to

quantify the level of phase consistency of the HFAbs across trials for the frequencies of interest.

The phase angle for each frequency was calculated based on the FFT results from the HFAb-

triggered spectrum analysis. The PLV of HFAbs at each frequency was then calculated as the mean

resultant in eq. 7:

$\text{PLV(f)} = \left| \frac{1}{N} \sum_{n=1}^{N} e^{i\phi_{f,n}} \right|$ eq. 7

where N is the total number of HFAbs and $\phi_{f,n}$ denotes the phase angle for the nth HFAb at

frequency f. The statistical significance of the observed PLV values was determined using a non-

parametric permutation test with 1000 permutations and the Rayleigh test corrected for multiple

comparisons, at 0.05. Peaks that passed both tests and had a prominence of 25% higher than the

PLV range were considered significant (shown in black dots in **Fig. 2c**). We further quantified the

phases at peak frequencies (**Extended Data Fig. 2a**), peak width (**Extended Data Fig. 2d**), and

the power ratio of peak frequency relative to baseline (**Extended Data Fig. 2e,f**) to ensure that

HFAb phase-locking occurred within narrow frequency bands.

Time-resolved PLV analysis. To quantify the temporal dynamics of the phase synchronization of

HFAbs with the low frequency LFP during cue and target processing, we extracted the phase of

HFAbs at each frequency as explained above. Then, we used a sliding window of 500 ms with a

step of 25 ms between -1500 and 1500 ms around the cue and target onsets separately. PLV was

calculated for each sliding window and across all trials (different types of trials were analyzed

separately, e.g., correct trials and incorrect trials). We set a minimum number of data points of 50

bursts for the analysis (to achieve reliable statistics on circular data,). We measured the PLV for

1000 samples of 25 HFAbs drawn randomly from a population of HFAbs in each time bin to

control for the number of bursts. The PLV for each time bin was calculated as the average PLV

across all randomly drawn samples. The average PLV value for each subject and for all subjects

in each experiment was calculated separately.

To test whether HFAb phase synchronization differed during cue and target processing, we used a

channel-specific randomization test. For each recording channel and time bin, 1000 subsamples of

HFAbs were randomly selected from the baseline period (within 1 second before each event). For

each channel, each time bin, and each frequency, we calculated the upper and lower bond CIs

(2.5% most extreme PLVs). For multiple comparison correction, we repeated this procedure 1000

time and found the most extreme 2.5% value across all CIs for time bins under the null hypothesis

that the PLV for each time bin and frequency is not significantly different from the baseline value.

Each time-frequency bin was considered significant if it differed from the critical values

(**Extended Data Fig. 3c**). Each electrode and frequency band (theta/alpha (4-14 Hz) and beta (15-

25 Hz)) for a time bin was considered significant if it showed a significantly different PLV from

the baseline in more than 25% of the frequency points in that frequency band. Next, we computed

the mean number of electrodes with significantly different PLVs than the randomized PLV

distribution for each subject. Using the binomial test, we determined whether the proportion of

electrodes with significantly lower PLV after the cue and target onset was different from the

baseline level as well as the chance level (5%, **Fig. 2d**). To adjust for multiple comparisons, we

used FDR for dependent samples and an alpha level of 0.05.

We also performed a control analysis to ensure that event-evoked iEEG signals are not

confounding the variation in synchronization. First, we extracted -1.5 to 1.5 seconds around cue-

and target-aligned iEEG signals for each electrode and trial. We averaged the data across all trials

and removed the average event-triggered iEEG signal from individual trials. The same

synchronization and statistical analyses were then performed on the trial with event-evoked iEEG

subtracted. Subtraction of event-evoked iEEG did not change the main results pattern (**Extended**

Data Fig. 3d). While our main analyses controlled for HFAb counts, we also conducted a control

analysis using pairwise phase consistency (PPC), a measure commonly used to quantify spike-

field synchronization that is unbiased by event count. Results were consistent with our PLV

analysis (**Extended Data Fig. 3e**).[83]

Analysis of Coupling ratio. A Coupling Ratio (CR) index was defined to compare PLV after cue

and target events as compared to baseline. We used this CR index also to visualize coupling

variation in both 3D brain renderings (**Fig. 2e**), and the average network level coupling analysis

(**Fig. 2g,h**). We calculated the coupling ratio by:

$CR = \dfrac{PLV_{event} - PLV_{baseline}}{PLV_{baseline}}$ eq. 8

Where the $PLV_{baseline}$ is the average PLV for each electrode within 1 second before the event, and

PLV_{event} is the average PLV for each electrode within 0.5 seconds after the event. In this context,

network decoupling is defined as a negative CR value indicating a reduction in PLV relative to the

baseline. We show the CR value at both cue and target events for each subject.

We also used GLME models to investigate the effect of cue and target response on the coupling

ratio following cue and target onsets (**Fig. 2g,h**). The GLME is formalized as shown in eq. 9:

$Coupling\ Ratio_{cue/target} = Response_{cue} + Response_{target} + (1|Subject) + b + \varepsilon$ eq. 9

<u>Quantification and Spectral analysis of HFAb-triggered HFA</u>

We calculated the interaction between HFAbs and HFA between each pair of electrodes. To

estimate HFA, first we used a zero-phase Butterworth filter with cut-off frequencies of 65 Hz and

175 Hz to bandpass filter the signal. We chose the HFA frequency band broader to capture larger

spectral content (similar results were obtained by choosing 65-115 Hz band). Next, we performed

a Hilbert transform and used the real part of the analytical signal as HFA amplitude (eq. 1). We

extracted a duration of one second around each HFAb from the HFA signal. We then calculated

the HFAb-triggered HFA for each individual burst event as well as the average value for all bursts

between each pair of electrodes. Our main analyses were restricted to HFAb-triggered HFA that

occurred outside of the main behavioral epochs; either before cues or after target detection

(**Extended Data Fig. 4e**, for those without a response 1 second following target presentation). To

control for any burst-independent correlations of iEEG signals across electrodes, we first

calculated the HFAb-triggered HFA using randomly assigned burst times. Each burst was jittered

with a random time lag of ±1000 ms. We then subtracted this jittered HFAb-triggered HFA from

the original for further analysis.

We calculated the spectral power of HFAb-triggered HFA across all electrode pairs to assess how

HFA is organized relative to HFAbs recorded on other channels. The power spectral density was

computed over the ±500 ms time window using a hanning taper. Additionally, we analyzed peak

prominence for all pairs and plotted the distribution of spectral peaks. For each pair of electrodes,

we extracted the prominent spectral peaks and plotted their distribution.

Dimensionality reduction of high-frequency temporal relation patterns. To identify prominent

temporal patterns of high-frequency activities within the brain network, we analyzed a high-

dimensional space of HFAb-triggered HFA across all electrode pairs for each subject, with each

electrode pair representing a single dimension. Using the HFAb-triggered HFA population, we

calculated the covariance matrix C by:

$$C = \frac{1}{n-1}\sum_{i=1}^{n}(X_i - \bar{X})(X_i - \bar{X})^T \qquad \text{eq. 10}$$

where X_i shows each HFAb-triggered HFA time series, n is the total number of electrode pairs, $\bar{X}$

is the marginal mean, and X^T is the transpose matrix of X. We found the eigenvectors of C as in

eq. 11:

$$C = VDV^T \qquad \text{eq. 11}$$

where V columns are the eigenvectors and principal components (PCs), and D contains

eigenvalues indicating how much variance each component explains (**Fig. 3d**). In all participants,

PC1 showed a near-symmetric activity state and explained more than 10% of variance in HFAb-

triggered HFA dynamics over the population of electrode pairs (we used multistep Wilcoxon rank-

sum test to examine if there was asymmetry of HFA within mirrored time windows around the

HFAbs). By projecting the original data to the PC vector space, we calculated the HFAb-triggered

HFA scores as:

$Score = XC$ eq. 12

We then used the scores of electrode pairs on the first component as their loading values on the

synchronous component of the population (**Extended Data Fig. 4**).

Identifying synchronous subnetworks. Based on the scores of each electrode pair on the

synchronous principal component, we generated a network synchrony matrix that describes

synchronous inter-electrode interactions. For each electrode, we then defined a vector of variables

with a dimension equal to the total number of electrodes. Each element of this vector describes the

score of an electrode pair on the synchronous component. This vector was defined for all

electrodes, which generated a matrix in which each row represented one observation (electrode)

and each column represented the score of the observed electrode on another electrode.

$S_i = Score\,(X_{i,\,1:n})$ eq. 13

Using this matrix, we clustered the electrodes using resampling-based consensus K-Means

algorithm with a correlational defined distance as in eq. 14:

$d_{i,j} = 1 - r(S_i, S_j)$ eq. 14

where $d_{i,j}$ is the correlation distance between two electrodes i and j. The correlational distance

between two electrodes is low when they show synchronous activity, and similar synchronization

patterns to the rest of the network. First, we used several clustering indices, including Hubert,

Silhouette, Davies-Bouldin, Calinski-Harabasz, Hartigan, Homogeneity, and Gap, to find an

optimal range of cluster numbers (between 2 and 8 for all subjects) [84–87].

For each cluster number, we ran the K-means algorithm 1000 times. The sample size was

subsampled and only 25 percent of electrodes were randomly selected for each clustering (we

ensured that each cluster had at least an average of 5 data points). After running the clustering

1000 times, a probability matrix for electrode pairs was defined as the ratio of numbers that

electrode pairs clustered together, divided by the number of electrode pairs in the same random

sampling for K-Means clustering. This ratio was calculated for all electrode pairs and used to

create a matrix of pair-wise grouping probabilities. We then ran a second K-means clustering

algorithm on this matrix to identify clusters that were similar in their network-level pair-wise

grouping likelihood. Using a similar approach and random sampling of electrodes, we generated

another pair-wise grouping probability matrix indicating how often electrodes were clustered

together based on their pair-wise grouping likelihood (**Extended Data Fig. 4e**). Using this

hierarchically defined pair-wise clustering likelihood, we ran a final K-means clustering on all

electrodes, labeling them according to the probability of being stably grouped together for each

cluster number. For each K (cluster number), we calculated clustering accuracy, confusion

(probability of non-diagonal clusters), a confusion rank, and a ratio from dividing median accuracy

(diagonals) by median confusion. We normalized each of the measures between 0 and 1 (1 showing

the best, and 0 showing the worst performance between all clusters). We used these four measures

and in a non-parametric vote, we chose the cluster number that outperformed others in this voting

pool. This non-parametric measure shows how well each K performs to detect more stable clusters

with high accuracy and low confusion level and estimate the optimal number of clusters. While

there is no definitive answer on what number of clusters is the best, we selected the optimal number

of clusters using this method to conduct further analysis. In summary, this clustering technique

reduces clustering biases caused by outlier electrode pairs in all clustering realizations as well as

stabilizes clusters.

To determine whether identified clusters were functionally specialized, we compared their

responses evoked by cues and targets to their baseline activity levels. We calculated the averaged

baseline normalized burst density within windows of 500 ms after cue onset and 750 ms after target

onset. The Wilcoxon test was used to determine if this response was non-zero across the electrodes

in each subnetwork. We corrected for multiple comparisons by using FDR for dependent samples

with an alpha level of 0.05. Clusters that were significantly activated by cue events are referred to

as "cue-subnetworks" and clusters that were significantly activated by target events are referred to

as "target-subnetworks". We use "subnetworks" to indicate that these are functionally specialized

modules within the larger brain-wide network engaged during attention. We consider the

overarching network for each subject to be a network of all nodes. A subnetwork here is used to

describe "modules" that we observe within the larger brain networks. The cue and target-

subnetworks were found in each data set for the optimal number of clusters. For datasets, for which

the optimal number of clusters did not contain distinct cue and target-subnetworks, if existed, we

chose the next K (cluster number) with highest ranking in the clustering measures that contained

both subnetworks (only in subject S12 we observed this).

Time-lag analysis between cue and target-subnetworks

We calculated the HFAb-triggered-HFA for pairs of electrodes during the period between the

target onset and the manual response to understand how the HFAbs were temporally ordered after

the target onset in both the cue and target-subnetworks. Next, we compared HFAb-triggered HFA

between electrodes in cue- and target-subnetworks to determine whether there was any asymmetry

in the distribution of HFAbs between the two subnetworks. A total of six subjects (3 in experiment

1 and 3 in experiment 2) showed both stable cue and target-subnetworks. We then calculated once

the HFAb-triggered HFA when HFA in the target-subnetwork was measured around the HFAbs

in cue-subnetworks, and when the HFA in the cue-subnetwork was measured around the HFAbs

in target-subnetworks. A lead state was indicated by HFAbs followed by stronger HFA power; a

lag state was when HFAbs followed stronger HFA power.

First, we measured the maximum asymmetry around the HFAb onset across all subjects (by

calculating the absolute difference between windows of varying lengths, 25 ms to 250 ms every

25 ms). We then calculated the averaged HFAb-triggered-HFA within 150 ms of HFAb onsets

where asymmetry was at its maximum. Next, we asked if the value differed based on the

directionality of the two subnetworks. For both directions, we used the Wilcoxon test to determine

whether the asymmetry around HFAb onset is non-zero. Additionally, we tested whether HFAb-

triggered HFA differs between the two directions after and before HFAbs.

To assess differences in HFAb-triggered HFA between cue- and target subnetworks, we used a

non-parametric permutation testing approach. For each time bin (25ms) in the -500 to +500 ms

window relative to HFAb onset, we calculated the observed difference in mean HFA between

cue→target and target→cue electrode pairs. To test for significant differences between HFAb-

triggered HFA in different directions, we used a permutation test with 10000 iterations, in which

randomly shuffled electrode pair labels and generated a null distribution of mean differences. For

each bin, a p-value was computed by comparing the observed difference in the two opposite

directions, to the permutation distribution. To correct for multiple comparisons across time points,

we used FDR correction for dependent samples at α = 0.05 (**Fig. 4a**). Additionally, cluster-based

correction was implemented with a minimum cluster size threshold of 25 ms to identify temporally

contiguous significant effects. Peak analysis was performed by identifying the maximum HFA

amplitude and corresponding latency for each electrode pair after applying a 25 ms smoothing

window (to avoid spurious peaks). Peak times are reported as mean ± standard error across all

electrode pairs within each subject and task. We used Wilcoxon rank sum to test if the peak time

distributions were significantly different in the opposite directions (**Fig. 4b**).

For each subject, we visualized the lead/lag interactions between stable cue and target-

subnetworks. To achieve a better visualization of lead/lag interactions, we measured the HFA peak

time for each electrode in cue/target-subnetworks relative to HFAb onset across all cluster

numbers. Each electrode that was a member of the target-subnetwork or the cue-subnetwork was

analyzed to determine its median peak-time-lag relative to the other cluster members. A median

peak-time-lag of all electrodes satisfying this condition was then plotted, with red representing a

lead, and blue representing a lag (**Fig. 4c**).

1002

Delayed Mutual Information Analysis. We used mutual information (MI) which is a non-linear

metric used in information theory to estimate the shared information between different time

segments in two electrodes to find where their mutual predictability is maximized. For each

electrode, we extracted data (HFAb density) from -1500 ms to 1500 ms around the target onset

(and for the control analysis, around the cue onset). We normalized the data and calculated MI for

segments of 750 ms sliding every 50 ms. The MI between electrodes X and Y is given by eq. 15:

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1010 $MI(X,Y) = \sum_{x \in X_w} \sum_{y \in Y_w} p(x, y) \log \left(\frac{p(x,y)}{p(x)p(y)} \right)$ eq. 15

1011

where p(x,y) is the joint probability distribution function of each windowed segment of electrodes

X and Y, and p(x) and p(y) are their marginal probability distributions. We estimated the

probability distribution for each variable and for the joint distributions using a histogram-based

approach and binning the data (the number of bins was selected based on the Freedman-Diaconis

rule [88] to balance the trade-off between estimation resolution and statistical reliability.

To further quantify how information is directionally coupled between cue and target-subnetworks,

we calculated delayed mutual information (DMI). The DMI was calculated by comparing the

temporal dynamics and dependencies between these two subnetworks (**Fig. 4d**).

The DMI between X and Y was calculated similar to MI, except that one of the timeseries was

delayed incrementally to determine whether the past of one electrode is a better predictor of the

future of the other electrode. The delay time-lag ranged from -500ms to 500ms with a step of 25ms,

as in eq. 16.

$\text{DMI(X,Y)} = \sum_{x \in X_w} \sum_{y \in Y_w^{\tau}} p(x, y) \log\left(\frac{p(x,y)}{p(x)p(y)}\right)$ eq. 16

where X_w is segmented windows of electrode X, and Y_w^{τ} is segmented windows of electrode Y

shifted by a time lag of τ.

The DMI analysis of each electrode pair can inform us about (i) when the electrodes showed

maximum inter-predictability relative to an event onset (e.g., target), and (ii) at what time-lag the

inter-predictability was maximized. To address this question, we extracted the DMI peaks in a 2D

space, which gave us both the time-lag and the timepoint relative to the event onset where two

electrodes showed maximum DMI values. We then asked if the time-lag of this peak is different

between electrodes in the cue-subnetworks and electrodes in the target-subnetworks. For example,

when we shifted the timeseries of electrodes in target-subnetworks, for each electrode in the cue-

subnetwork, we measured its mean DMI relative to electrodes in target-subnetworks. We

determined at what time lag and at what time relative to target onset DMI was maximized. We

used the Wilcoxon test to see if the maximum DMI time-lag (τ_{max}) occurred between electrodes

in cue and target-subnetworks was non-zero. The lead and lag patterns in inter-predictability

between cue and target-subnetworks were then determined by the sign of the average over τ_{max} for

all electrodes in the shifting subnetworks.

1040

Modeling iEEG by spiking neural networks

We investigated high frequency activity in general and high frequency burst events in particular

using a limited network of two recording sites placed on two interconnected networks of spiking

neurons. For modeling the spiking networks, we used a framework described in[6]. Neuronal point-

source models can accurately simulate electrical fields in cortical neural networks.[89] Two cubic

neuronal structures were simulated in three dimensions. Synaptic dynamics and connectivity

patterns were implemented as explained in[6]. Each cubic network consisted of 1000 neurons (10 x

10 x 10, xyz). We modeled neurons as spherically symmetric points in a 3D grid (with r units on

each axis). A recording disk was implemented on top of each network at a distance three times

greater than the network depth (**Fig. 5a**). Pairwise connectivity was calculated based on anatomical

studies (discussed below) and a distant-dependent Gaussian rule.[90,91] The distant dependent

connectivity factor is:

$$P(d) = P_0\, e^{\frac{-(\frac{d}{r})^2}{\sigma^2}} \qquad\qquad eq.\,17$$

where r is the grid unit, d is the distance between two neurons, σ is the standard deviation of

distances between neurons (Barral and Reyes, 2016), and P_0 is a structural scaling factor

(**Extended Data Fig. 8a**) reflecting the maximum connection probability across the network (see

Extended Data Table 2).

Four interneuron types were used: Parvalbumin (PV), Calbindin (CB), Calretinin (CR), and

Cholecystokinin (CCK) expressing interneurons. Excitatory neurons were divided into regular

spiking neurons (70%), intrinsic low-threshold spiking bursting neurons (10%), and fast adapting

regular spiking neurons (20%).

Neuron models. We used Izhikevich neuron model [92] for simulating regular spiking, burst spiking,

fast spiking and low-threshold spiking neurons. Each neuron is modeled by a series of differential

equations as in eq 18.

$$\frac{dv}{dt} = 0.04v^2 + 5v + 140 - u + I \qquad\qquad eq. 18$$

$$\frac{du}{dt} = a(bv - u)$$

where variables v and u denote neural membrane potential and membrane recovery, respectively.

Parameters a and b define the recovery rate and sub-threshold fluctuations sensitivity,

respectively. For after-spike resenting the model uses two auxiliary equations as in eq. 19,

$$\text{if } v > 30 \ mV \text{ , then} \begin{cases} v = c \\ u = u + d \end{cases} \qquad\qquad eq. 19$$

the parameter c resets the value of membrane potential v after a spike, and the parameter d adjusts

the after-spike recovery variable u. Parameters for different neuron types were chosen as suggested

in [93] to approximately generate firing patterns of each neuron type (See **Extended Data Table 2**

for each neuron type parameter).

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Neural connectivity. Besides distance-based connectivity factors, neuron types had different

connection probabilities. We adapted the scaling factor for connectivity between excitatory

neurons and inhibitory interneurons from [91]. Additionally, rodent anatomical studies were

considered in determining the connectivity among different types of neurons. Generally, PV

interneurons inhibit themselves and VIP interneurons (likely similar to CR), whereas SOM

interneurons (likely similar to CB) do not inhibit each other, and VIP interneurons disinhibit SOM

interneurons preferentially [94,95]. The connectivity between networks was defined by a probability

and a rate of connection. This connectivity was attributed primarily to excitatory neurons (90%).

1084

Simulation of post-synaptic potentials. For each neuron the postsynaptic potentials were

modeled by a biexponential function as in eq. 20,

$$g(t) = e^{-\frac{t}{\tau_r}} - e^{-\frac{t}{\tau_d}} \qquad eq.\,20$$

where τ_r and τ_d denote the rise and decay time constant of postsynaptic current, respectively. The

biexponential function was implemented through a second-order ordinary differential equation as

in eq. 21.

1091

1092 $$\frac{d^2 g(t)}{dt^2} = -\frac{(\tau_d + \tau_r)}{\tau_d \tau_r} \frac{dg(t)}{dt} - \frac{g(t)}{\tau_d \tau_r} \qquad eq.\,21$$

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1094   Where t is time relative to spike, and g(t) is the synaptic conductivity. The rise time was set

1095   similarly to ~1ms for all neurons while the decay time for pyramidal neurons were and

1096   interneurons varied from ~6ms to 24ms[96–99] as shown in **Extended Data Table 2**.

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Network external inputs. Each network was fed external currents to generate firing rates similar

to cortical neurons. Each neuron received a cosine input. The objective was to first control

externally induced rhythmicity in network activation by frequency, as well as phase coherence

between the input function and neurons. On average, each network received external input

sufficient to generate a 5 Hz firing rate.[100] The external input to pyramidal neurons was three times

greater than the external input to inhibitory neurons. In addition, a Brownian noise ($\frac{1}{f^2}$) was added

to the input as per previous experimental observations.[101]

For nonrhythmic input, we considered slow and ultraslow oscillatory (<1 Hz) input to each neuron

with an initial phase lag. Then a coherence index was used to determine the distribution of phase

lags among neurons. Thus, the phase range was defined:

φ = 2π (1 - coherence)   *eq. 22*

For nonrhythmic input, we ran 100 simulations in which coherence varied from 0 to 1 with a step

size of 0.01 (**Fig. 5e**). For Networks with reciprocal connections, we fed the networks with

uncorrelated inputs, but both networks were reciprocally connected (**Fig. 5g**). The connectivity

strength was changed by one order of magnitude from 0.001 to 0.1 and the simulation was run 100

times for each value. For networks with feedforward connection, we simulated feedforward

networks in which the connections between the two networks are unidirectional. The connectivity

strength was changed from 0.001 to 0.1 by one order of magnitude (**Fig. 5h**). For the rhythmic

input, we incrementally increased the cosine input frequency up to 12 Hz for each simulation run

(**Fig. 5i**). For networks with shared input, we ran 100 simulations in which networks were not

connected but received correlated inputs (5-25 percent of neurons received the same input, not

shown).

To investigate the effect of an external stimuli on directional networks, we simulate a feedforward

network as explained before with an internetwork connectivity ratio of 9 to 1 (network 1 and 2

respectively, **Extended Data Fig. 8e**). We then fed network 1 with an external input pulse of 500

ms duration and 250 ms duty cycles (**Extended Data Fig. 8f**). The input was fed to 50 percent of

neurons in the network 1. We then calculated the time resolved PLV in both networks as explained

before (**Fig. 5f**, **Extended Data Fig. 8g**).

1126

Integration of postsynaptic currents and neural activities at the recording sites. For each

simulation, we estimated the field potential at two recording sites (black disks **Fig. 5a**). By

assuming neurons as point-source field, we measured field dynamics at each recording site by

summing the attenuated membrane potentials and the postsynaptic currents from all neurons. We

estimated the voltages at the recording disk by considering membrane and post-synaptic potentials

as electrical dipoles (with negligible distance between poles relative to the recording disk). The

electric filed is then represented by $E(r, t)$, where r and t denote the distance from the source and

the time, respectively. The extracellular potential is then calculated by:

$$E = -\nabla V \qquad\qquad eq.\,23$$

Using Ohm's law, the electric field at distance r from each dipole with a current density of I_n (t)

is equal to:

$$E(r, t) = \frac{I_n(t)}{4\pi r^3 \sigma} \qquad\qquad eq.\,24$$

with $I_n(t)$ represented as:

$$I_n(t) = I_m(t) + I_{syn}(t) \qquad eq.\,25$$

where σ denotes the medium conductivity (which we assumed is independent of distance from

sources). $I_m(t)$ and $I_{syn}(t)$ are the transmembrane current and the synaptic current of each neuron

respectively. By integrating the electric field, we calculated the potential at the disk by:

$$V(r, t) = -\int_{\infty}^{r} E(r', t)dr' = -\int_{\infty}^{r} \frac{I_n(t)}{4\pi r'^2 \sigma} \, dr'$$

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1146
$$V(r,t) = \frac{I_n(t)}{4\pi r^2 \sigma} \qquad eq.\,26$$

1147 The voltage recorded at each site is calculated as in eq 27:

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$$V(t) = \frac{1}{4\pi r^2} \sum_{i=1}^{N} \frac{I_n(t)}{r_i} \qquad eq.\,27$$

where r_i is the distance between the recording disk and the ith point-source, and N is equal to the

total number of neurons in each network.

We used a non-ohmic filter to attenuate higher frequencies by getting insights from.[102] We

implemented an exponential attenuation in the frequency domain. First for each signal we

computed the FFT. We then applied an exponential attenuation factor to the magnitude of the

frequency components (**Extended Data Fig. 8b**). This factor exponentially decreases the

amplitude for higher frequencies as in eq. 28:

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$$A(f) = e^{-\alpha f} \qquad eq.\,28$$

1157

Where $A(f)$ is the attenuation factor for frequency f, and α is a parameter that controls the rate of

exponential decay which we set to 0.01. After multiplying the attenuation factor, we performed

the inverse FFT (iFFT) to transform the signal back to the time domain.

For each condition, we ran simulations 100 times, each simulating iEEG signals and neural activity

for three seconds. We detected HFAbs at each recording site and calculated the amplitude of the

analytical signal as explained in (eq. 1). Next, we examined whether there was a correlation

between HFAb density and aggregated spike density in each network. The spike density was

calculated and smoothed using a Gaussian window of 25 ms. We measured the correlation

coefficient between aggregated spike density and burst density for each simulation. On average,

HFAb events were significantly correlated with spike densities in each network.

In a control condition, we randomly assigned burst times and examined the correlation between

burst density and spike density. This randomization was performed 1000 times and we found the

95% confidence interval under the null hypothesis that burst density is not related to spike density.

All other analyses of the modeling results were conducted in a similar manner to those of the

experimental data.

1173

Four-Network Model of Spatial Attention

We extended our model to a four-network architecture mimicking the cue-target processing

hierarchy (**Fig. 5j**). The model contained two spatially-selective cue networks (N1, N2) and two

target networks (N3, N4), with dominant feedforward connections from cue to target networks (5

times stronger than feedback).[103] Intra-network connection strength was set 2 times greater than

the inter-network connectivity strength.[104] To simulate the spatial attention task, we implemented

three distinct external input functions:

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Trial initialization: A decaying exponential input ($\tau = 4$s) was delivered to all networks at trial

onset, decreasing to 25% of initial amplitude by trial end simulating baseline states maintained

throughout the trial.

Cue processing: Upon cue presentation, one cue network received spatially-selective input

modeled as a double-exponential function ($\tau_{rise} = 50$ms, $\tau_{decay} = 1$s).[56,105]

Target processing: Target networks received double-exponential input ($\tau_{rise} = 100$ms, $\tau = 500$ms)

at target onset. In valid trials, where targets appeared at cued locations, the previously activated

cue network received additional weak reactivation at 50% of original cue amplitude, simulating

either feedback or sustained attention signals from frontal-parietal networks or inputs from

subcortical structures such as pulvinar.[56,105,106]

We simulated 150 trials for each condition (valid and invalid cues) with 4s trial duration. HFAbs

were detected using identical parameters as empirical data, and we calculated HFAb responses in

each network and delayed mutual information (DMI) between cue-target network pairs.

1195

HFAb-based detection modelling. To link burst dynamics to behavioral performance, we

extracted HFAb rates from target networks within 500ms windows following target onset for both

valid and invalid conditions. Detection probability was modeled using a logistic function:

$$P(detect|x) = logit^{-1}(\beta_0 + \beta_1 x)$$

where x represents HFAb rate, and β_0, β_1 are model parameters estimated via maximum likelihood.

We fitted detection thresholds to invalid trials to achieve ~85% accuracy (matching empirical

performance), then evaluated whether valid trials exceeded this threshold. In addition, HFAb rates

were binned into 100 bins spanning the full range to calculate detection probabilities for each

condition (**Fig. 5m**).

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1207   **Data Availability**

1208   Preprocessed data used for generating these results are deposited at Figshare and will be accessible

1209   through https://doi.org/10.6084/m9.figshare.30434683.[107] Source data are provided with this

1210   paper.

1211

Code Availability

Custom programming codes for analysis and modeling are available from https://github.com/banaiek/Attentional_Routing/.

References

1. Saalmann, Y. B., Pigarev, I. N. & Vidyasagar, T. R. Neural Mechanisms of Visual Attention: How Top-Down Feedback Highlights Relevant Locations. *Science* **316**, 1612–1615 (2007).

2. Gregoriou, G. G., Gotts, S. J., Zhou, H. & Desimone, R. High-Frequency, Long-Range Coupling Between Prefrontal and Visual Cortex During Attention. *Science* **324**, 1207–1210 (2009).

3. Szczepanski, S. M. *et al.* Dynamic Changes in Phase-Amplitude Coupling Facilitate Spatial Attention Control in Fronto-Parietal Cortex. *PLOS Biology* **12**, e1001936 (2014).

4. Buschman, T. J. & Kastner, S. From Behavior to Neural Dynamics: An Integrated Theory of Attention. *Neuron* **88**, 127–144 (2015).

5. Fiebelkorn, I. C. & Kastner, S. Functional Specialization in the Attention Network. *Annu Rev Psychol* **71**, 221–249 (2020).

6. Banaie Boroujeni, K. & Womelsdorf, T. Routing states transition during oscillatory bursts and attentional selection. *Neuron* https://doi.org/10.1016/j.neuron.2023.06.012 (2023) doi:10.1016/j.neuron.2023.06.012.

7. Womelsdorf, T. & Fries, P. Neuronal coherence during selective attentional processing and sensory–motor integration. *Journal of Physiology-Paris* **100**, 182–193 (2006).

8.   Lakatos, P., Karmos, G., Mehta, A. D., Ulbert, I. & Schroeder, C. E. Entrainment of

Neuronal Oscillations as a Mechanism of Attentional Selection. *Science* **320**, 110–113

(2008).

9.   Siegel, M., Donner, T. H., Oostenveld, R., Fries, P. & Engel, A. K. Neuronal

Synchronization along the Dorsal Visual Pathway Reflects the Focus of Spatial Attention.

Neuron **60**, 709–719 (2008).

10. Buschman, T. J. & Miller, E. K. Top-Down Versus Bottom-Up Control of Attention in the

Prefrontal and Posterior Parietal Cortices. *Science* **315**, 1860–1862 (2007).

11. Siegel, M., Buschman, T. J. & Miller, E. K. Cortical information flow during flexible

sensorimotor decisions. *Science* **348**, 1352–1355 (2015).

12. Canolty, R. T. *et al.* Oscillatory phase coupling coordinates anatomically dispersed

functional cell assemblies. *PNAS* **107**, 17356–17361 (2010).

13. Braun, U. *et al.* Dynamic reconfiguration of frontal brain networks during executive

cognition in humans. *Proceedings of the National Academy of Sciences* **112**, 11678–11683

(2015).

14. Bassett, D. S. & Sporns, O. Network neuroscience. *Nat Neurosci* **20**, 353–364 (2017).

15. Avena-Koenigsberger, A., Misic, B. & Sporns, O. Communication dynamics in complex

brain networks. *Nat Rev Neurosci* **19**, 17–33 (2018).

16. Brookes, M. J. *et al.* Measuring functional connectivity using MEG: Methodology and

comparison with fcMRI. *NeuroImage* **56**, 1082–1104 (2011).

17. Hari, R. & Parkkonen, L. The brain timewise: how timing shapes and supports brain

function. *Philosophical Transactions of the Royal Society B: Biological Sciences* **370**,

20140170 (2015).

18. Higley, M. J. & Cardin, J. A. Spatiotemporal dynamics in large-scale cortical networks.

Current Opinion in Neurobiology **77**, 102627 (2022).

19. Lachaux, J.-P., Axmacher, N., Mormann, F., Halgren, E. & Crone, N. E. High-frequency

neural activity and human cognition: past, present and possible future of intracranial EEG

research. *Prog Neurobiol* **98**, 279–301 (2012).

20. Parvizi, J. & Kastner, S. Human Intracranial EEG: Promises and Limitations. *Nat Neurosci*

21, 474–483 (2018).

21. Voytek, B. *et al.* Oscillatory dynamics coordinating human frontal networks in support of

goal maintenance. *Nat Neurosci* **18**, 1318–1324 (2015).

22. Haller, M. *et al.* Persistent neuronal activity in human prefrontal cortex links perception and

action. *Nat Hum Behav* **2**, 80–91 (2018).

23. Helfrich, R. F. *et al.* Neural Mechanisms of Sustained Attention Are Rhythmic. *Neuron* **99**,

854-865.e5 (2018).

24. Martin, A. B. *et al.* Temporal Dynamics and Response Modulation across the Human

Visual System in a Spatial Attention Task: An ECoG Study. *J. Neurosci.* **39**, 333–352

(2019).

25. Ray, S. & Maunsell, J. H. R. Different Origins of Gamma Rhythm and High-Gamma

Activity in Macaque Visual Cortex. *PLOS Biology* **9**, e1000610 (2011).

26. Banaie Boroujeni, K., Tiesinga, P. & Womelsdorf, T. Adaptive spike-artifact removal from

local field potentials uncovers prominent beta and gamma band neuronal synchronization.

Journal of Neuroscience Methods **330**, 108485 (2020).

27. Leszczyński, M. *et al.* Dissociation of broadband high-frequency activity and neuronal

firing in the neocortex. *Science Advances* **6**, eabb0977 (2020).

28.   Leonard, M. K. *et al.* Large-scale single-neuron speech sound encoding across the depth of

human cortex. *Nature* **626**, 593–602 (2024).

29.   Zolnik, T. A. *et al.* Layer 6b controls brain state via apical dendrites and the higher-order

thalamocortical system. *Neuron* **112**, 805-820.e4 (2024).

30.   Canolty, R. T. & Knight, R. T. The functional role of cross-frequency coupling. *Trends in*

Cognitive Sciences **14**, 506–515 (2010).

31.   Arnulfo, G. *et al.* Long-range phase synchronization of high-frequency oscillations in

human cortex. *Nat Commun* **11**, 5363 (2020).

32.   Williams, N. *et al.* Modules in connectomes of phase-synchronization comprise

anatomically contiguous, functionally related regions. *NeuroImage* **272**, 120036 (2023).

33.   Garrett, J. C. *et al.* Binding of cortical functional modules by synchronous high-frequency

oscillations. *Nat Hum Behav* 1–15 (2024) doi:10.1038/s41562-024-01952-2.

34.   Jensen, O. & Colgin, L. L. Cross-frequency coupling between neuronal oscillations. *Trends*

in Cognitive Sciences **11**, 267–269 (2007).

35.   Jiang, H., Bahramisharif, A., van Gerven, M. A. J. & Jensen, O. Measuring directionality

between neuronal oscillations of different frequencies. *NeuroImage* **118**, 359–367 (2015).

36.   Kam, J. W. Y. *et al.* Default network and frontoparietal control network theta connectivity

supports internal attention. *Nat Hum Behav* **3**, 1263–1270 (2019).

37.   Johnson, E. L. *et al.* A rapid theta network mechanism for flexible information encoding.

Nat Commun **14**, 2872 (2023).

38.   Tong, A. P. S., Vaz, A. P., Wittig, J. H., Inati, S. K. & Zaghloul, K. A. Ripples reflect a

spectrum of synchronous spiking activity in human anterior temporal lobe. *eLife* **10**, e68401

(2021).

39.   Onslow, A. C. E., Jones, M. W. & Bogacz, R. A Canonical Circuit for Generating Phase-

1305   Amplitude Coupling. *PLOS ONE* **9**, e102591 (2014).

1306   40.   Colgin, L. L. *et al.* Frequency of gamma oscillations routes flow of information in the

1307   hippocampus. *Nature* **462**, 353–357 (2009).

1308   41.   Akam, T. & Kullmann, D. M. Oscillations and Filtering Networks Support Flexible Routing

1309   of Information. *Neuron* **67**, 308–320 (2010).

1310   42.   Palmigiano, A., Geisel, T., Wolf, F. & Battaglia, D. Flexible information routing by

1311   transient synchrony. *Nat Neurosci* **20**, 1014–1022 (2017).

1312   43.   Lundqvist, M., Herman, P., Warden, M. R., Brincat, S. L. & Miller, E. K. Gamma and beta

1313   bursts during working memory readout suggest roles in its volitional control. *Nat Commun*

1314   **9**, 394 (2018).

1315   44.   Hahn, G., Ponce-Alvarez, A., Deco, G., Aertsen, A. & Kumar, A. Portraits of

1316   communication in neuronal networks. *Nat Rev Neurosci* **20**, 117–127 (2019).

1317   45.   Corbetta, M. & Shulman, G. L. SPATIAL NEGLECT AND ATTENTION NETWORKS.

1318   *Annu Rev Neurosci* **34**, 569–599 (2011).

1319   46.   Chica, A. B., Bartolomeo, P. & Lupiáñez, J. Two cognitive and neural systems for

1320   endogenous and exogenous spatial attention. *Behavioural Brain Research* **237**, 107–123

1321   (2013).

1322   47.   Katsuki, F. & Constantinidis, C. Bottom-up and top-down attention: different processes and

1323   overlapping neural systems. *Neuroscientist* **20**, 509–521 (2014).

1324   48.   Vossel, S., Geng, J. J. & Fink, G. R. Dorsal and Ventral Attention Systems: Distinct Neural

1325   Circuits but Collaborative Roles. *Neuroscientist* **20**, 150–159 (2014).

1326   49.   König, P., Engel, A. K. & Singer, W. Relation between oscillatory activity and long-range

1327   synchronization in cat visual cortex. *Proceedings of the National Academy of Sciences* **92**,

1328   290–294 (1995).

1329   50.   Roelfsema, P. R., Engel, A. K., König, P. & Singer, W. Visuomotor integration is

1330   associated with zero time-lag synchronization among cortical areas. *Nature* **385**, 157–161

1331   (1997).

1332   51.   Uhlhaas, P. J. *et al.* Neural Synchrony in Cortical Networks: History, Concept and Current

1333   Status. *Front Integr Neurosci* **3**, 17 (2009).

1334   52.   Gollo, L. L., Mirasso, C. & Villa, A. E. P. Dynamic control for synchronization of

1335   separated cortical areas through thalamic relay. *NeuroImage* **52**, 947–955 (2010).

1336   53.   Gollo, L. L., Mirasso, C. R., Atienza, M., Crespo-Garcia, M. & Cantero, J. L. Theta Band

1337   Zero-Lag Long-Range Cortical Synchronization via Hippocampal Dynamical Relaying.

1338   *PLOS ONE* **6**, e17756 (2011).

1339   54.   Saalmann, Y. B. & Kastner, S. Cognitive and Perceptual Functions of the Visual Thalamus.

1340   *Neuron* **71**, 209–223 (2011).

1341   55.   Logothetis, N. K. *et al.* Hippocampal–cortical interaction during periods of subcortical

1342   silence. *Nature* **491**, 547–553 (2012).

1343   56.   Saalmann, Y. B., Pinsk, M. A., Wang, L., Li, X. & Kastner, S. The Pulvinar Regulates

1344   Information Transmission Between Cortical Areas Based on Attention Demands. *Science*

1345   https://doi.org/10.1126/science.1223082 (2012) doi:10.1126/science.1223082.

1346   57.   Vijayan, S. & Kopell, N. J. Thalamic model of awake alpha oscillations and implications

1347   for stimulus processing. *Proceedings of the National Academy of Sciences* **109**, 18553–

1348   18558 (2012).

58. Fiebelkorn, I. C., Pinsk, M. A. & Kastner, S. The mediodorsal pulvinar coordinates the

macaque fronto-parietal network during rhythmic spatial attention. *Nat Commun* **10**, 215

(2019).

59. Gollo, L. L., Mirasso, C., Sporns, O. & Breakspear, M. Mechanisms of Zero-Lag

Synchronization in Cortical Motifs. *PLOS Computational Biology* **10**, e1003548 (2014).

60. Zolnik, T. A. *et al.* Layer 6b Is Driven by Intracortical Long-Range Projection Neurons.

Cell Reports **30**, 3492-3505.e5 (2020).

61. Menara, T., Qin, Y., Bassett, D. S. & Pasqualetti, F. Relay Interactions Enable Remote

Synchronization in Networks of Phase Oscillators. *IEEE Control Systems Letters* **6**, 500–

505 (2022).

62. Smith, S. M. *et al.* Correspondence of the brain's functional architecture during activation

and rest. *Proceedings of the National Academy of Sciences* **106**, 13040–13045 (2009).

63. Srivastava, P. *et al.* Models of communication and control for brain networks: distinctions,

convergence, and future outlook. *Netw Neurosci* **4**, 1122–1159 (2020).

64. Engel, T. A., Schölvinck, M. L. & Lewis, C. M. The diversity and specificity of functional

connectivity across spatial and temporal scales. *NeuroImage* **245**, 118692 (2021).

65. Battiston, F. *et al.* Networks beyond pairwise interactions: Structure and dynamics. *Physics

Reports* **874**, 1–92 (2020).

66. Jensen, O., Gips, B., Bergmann, T. O. & Bonnefond, M. Temporal coding organized by

coupled alpha and gamma oscillations prioritize visual processing. *Trends in Neurosciences*

37, 357–369 (2014).

67.   Chapeton, J. I., Haque, R., Wittig, J. H., Inati, S. K. & Zaghloul, K. A. Large-Scale

1371   Communication in the Human Brain Is Rhythmically Modulated through Alpha Coherence.

1372   *Current Biology* **29**, 2801-2811.e5 (2019).

1373   68.   Harris, K. D. & Thiele, A. Cortical state and attention. *Nat Rev Neurosci* **12**, 509–523

1374   (2011).

1375   69.   Hanslmayr, S., Staudigl, T. & Fellner, M.-C. Oscillatory power decreases and long-term

1376   memory: the information via desynchronization hypothesis. *Front. Hum. Neurosci.* **6**,

1377   (2012).

1378   70.   Hanslmayr, S., Staresina, B. P. & Bowman, H. Oscillations and Episodic Memory:

1379   Addressing the Synchronization/Desynchronization Conundrum. *Trends in Neurosciences*

1380   **39**, 16–25 (2016).

1381   71.   Beaman, C. B., Eagleman, S. L. & Dragoi, V. Sensory coding accuracy and perceptual

1382   performance are improved during the desynchronized cortical state. *Nat Commun* **8**, 1308

1383   (2017).

1384   72.   Dickey, C. W. *et al.* Widespread ripples synchronize human cortical activity during sleep,

1385   waking, and memory recall. *Proc Natl Acad Sci U S A* **119**, e2107797119 (2022).

1386   73.   Verzhbinsky, I. A. *et al.* Co-occurring ripple oscillations facilitate neuronal interactions

1387   between cortical locations in humans. *Proceedings of the National Academy of Sciences*

1388   **121**, e2312204121 (2024).

1389   74.   Egly, R., Driver, J. & Rafal, R. D. Shifting visual attention between objects and locations:

1390   Evidence from normal and parietal lesion subjects. *Journal of Experimental Psychology:*

1391   *General* **123**, 161–177 (1994).

75.   Fiebelkorn, I. C., Saalmann, Y. B. & Kastner, S. Rhythmic Sampling within and between

Objects despite Sustained Attention at a Cued Location. *Current Biology* **23**, 2553–2558

(2013).

76.   Wang, L., Mruczek, R. E. B., Arcaro, M. J. & Kastner, S. Probabilistic Maps of Visual

Topography in Human Cortex. *Cereb Cortex* **25**, 3911–3931 (2015).

77.   Lancaster, J. l. *et al.* Automated labeling of the human brain: A preliminary report on the

development and evaluation of a forward-transform method. *Human Brain Mapping* **5**,

238–242 (1997).

78.   Oostenveld, R., Fries, P., Maris, E. & Schoffelen, J.-M. FieldTrip: Open Source Software

for Advanced Analysis of MEG, EEG, and Invasive Electrophysiological Data.

Computational Intelligence and Neuroscience **2011**, 156869 (2011).

79.   Boashash, B. Estimating and Interpreting the Instantaneous Frequency of a Signal – Part 1:

Fundamentals. in *Proceedings of the Ieee* 520–538 (1992).

80.   Hill, M. R. H., Fried, I. & Koch, C. Quantification and classification of neuronal responses

in kernel-smoothed peristimulus time histograms. *Journal of Neurophysiology* **113**, 1260–

1274 (2015).

81.   Dunn, O. J. Multiple Comparisons among Means. *Journal of the American Statistical*

Association **56**, 52–64 (1961).

82.   Benjamini, Y. & Yekutieli, D. False Discovery Rate–Adjusted Multiple Confidence

Intervals for Selected Parameters. *Journal of the American Statistical Association* **100**, 71–

81 (2005).

83.   Vinck, M., van Wingerden, M., Womelsdorf, T., Fries, P. & Pennartz, C. M. A. The

pairwise phase consistency: A bias-free measure of rhythmic neuronal synchronization.

NeuroImage **51**, 112–122 (2010).

84.   Milligan, G. W. & Cooper, M. C. An examination of procedures for determining the

number of clusters in a data set. *Psychometrika* **50**, 159–179 (1985).

85.   Jain, A. K. & Dubes, R. C. *Algorithms for Clustering Data*. (Prentice-Hall, Inc., USA,

1988).

86.   Halkidi, M., Batistakis, Y. & Vazirgiannis, M. On Clustering Validation Techniques.

Journal of Intelligent Information Systems **17**, 107–145 (2001).

87.   Vendramin, L., Campello, R. J. G. B. & Hruschka, E. R. Relative clustering validity

criteria: A comparative overview. *Statistical Analysis and Data Mining: The ASA Data*

Science Journal **3**, 209–235 (2010).

88.   Freedman, D. & Diaconis, P. On the histogram as a density estimator:L2 theory. *Z.*

Wahrscheinlichkeitstheorie verw Gebiete **57**, 453–476 (1981).

89.   Billeh, Y. N. *et al.* Systematic Integration of Structural and Functional Data into Multi-scale

Models of Mouse Primary Visual Cortex. *Neuron* **106**, 388-403.e18 (2020).

90.   Potjans, T. C. & Diesmann, M. The Cell-Type Specific Cortical Microcircuit: Relating

Structure and Activity in a Full-Scale Spiking Network Model. *Cerebral Cortex* **24**, 785–

806 (2014).

91.   Barral, J. & D Reyes, A. Synaptic scaling rule preserves excitatory–inhibitory balance and

salient neuronal network dynamics. *Nat Neurosci* **19**, 1690–1696 (2016).

92.   Izhikevich, E. M. Which model to use for cortical spiking neurons? *IEEE Transactions on*

Neural Networks **15**, 1063–1070 (2004).

93.   Izhikevich, E. M. Simple model of spiking neurons. *IEEE Transactions on Neural*

Networks **14**, 1569–1572 (2003).

94.   Pfeffer, C. K., Xue, M., He, M., Huang, Z. J. & Scanziani, M. Inhibition of inhibition in

visual cortex: the logic of connections between molecularly distinct interneurons. *Nat*

Neurosci **16**, 1068–1076 (2013).

95.   Karnani, M. M. *et al.* Cooperative Subnetworks of Molecularly Similar Interneurons in

Mouse Neocortex. *Neuron* **90**, 86–100 (2016).

96.   Medalla, M., Gilman, J. P., Wang, J.-Y. & Luebke, J. I. Strength and Diversity of Inhibitory

Signaling Differentiates Primate Anterior Cingulate from Lateral Prefrontal Cortex. *J.*

Neurosci. **37**, 4717–4734 (2017).

97.   Gilman, J. P., Medalla, M. & Luebke, J. I. Area-Specific Features of Pyramidal Neurons—a

Comparative Study in Mouse and Rhesus Monkey. *Cerebral Cortex* **27**, 2078–2094 (2017).

98.   Medalla, M. & Luebke, J. I. Diversity of Glutamatergic Synaptic Strength in Lateral

Prefrontal versus Primary Visual Cortices in the Rhesus Monkey. *J. Neurosci.* **35**, 112–127

(2015).

99.   Amatrudo, J. M. *et al.* Influence of Highly Distinctive Structural Properties on the

Excitability of Pyramidal Neurons in Monkey Visual and Prefrontal Cortices. *J. Neurosci.*

32, 13644–13660 (2012).

100.   Roxin, A., Brunel, N., Hansel, D., Mongillo, G. & Vreeswijk, C. van. On the Distribution of

Firing Rates in Networks of Cortical Neurons. *J. Neurosci.* **31**, 16217–16226 (2011).

101.   Milstein, J., Mormann, F., Fried, I. & Koch, C. Neuronal Shot Noise and Brownian 1/f2

Behavior in the Local Field Potential. *PLOS ONE* **4**, e4338 (2009).

102. Bédard, C., Kröger, H. & Destexhe, A. Modeling Extracellular Field Potentials and the

Frequency-Filtering Properties of Extracellular Space. *Biophysical Journal* **86**, 1829–1842

(2004).

103. Deco, G. & Rolls, E. T. Neurodynamics of Biased Competition and Cooperation for

Attention: A Model With Spiking Neurons. *Journal of Neurophysiology* **94**, 295–313

(2005).

104. Markov, N. T. *et al.* A Weighted and Directed Interareal Connectivity Matrix for Macaque

Cerebral Cortex. *Cerebral Cortex* **24**, 17–36 (2014).

105. Zhou, H., Schafer, R. J. & Desimone, R. Pulvinar-Cortex Interactions in Vision and

Attention. *Neuron* **89**, 209–220 (2016).

106. Buffalo, E. A., Fries, P., Landman, R., Liang, H. & Desimone, R. A backward progression

of attentional effects in the ventral stream. *Proceedings of the National Academy of*

Sciences **107**, 361–365 (2010).

107. Banaie Boroujeni, Kianoush; Helfrich, Randolph F.; Fiebelkorn, Ian C.; Bentley, Nicole;

Brunner, Peter; Lin, Jack J; et al. (2025). Preprocessed Burst-triggered HFA Datasets for individual

subjects. figshare. Dataset. https://doi.org/10.6084/m9.figshare.30434683