Practical media formulations for rapid growth of *Lactobacillus iners* and other vaginal bacteria

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Abstract

Vaginal microbiome composition is closely tied to host health. A microbiome dominated by specific anaerobes (e.g., *Gardnerella vaginalis*) is termed bacterial vaginosis (BV) and is associated with negative health outcomes, while colonization by *Lactobacillus* species is thought to protect against BV. However, the role of the species *Lactobacillus iners* in vaginal health is controversial, with evidence that some strains may not protect against BV while others do. To better characterize *L. iners* strains, their interactions with vaginal bacteria and human cells need to be investigated *in vitro*, but this has been impeded by the lack of liquid media for rapid *L. iners* growth. We have developed three liquid media formulations for *L. iners* growth: Serrador's Lactobacilli-adapted Iscove's Medium (SLIM) which leads to robust *L. iners* growth, a vaginally adapted version of SLIM (SLIM-V) and a chemically defined medium (SLIM-CD). SLIM and SLIM-V lead to dramatically improved *L. iners* growth compared to previously published formulations and support growth of other vaginal bacteria, including *L. crispatus, L. jensenii, L. gasseri* and *G. vaginalis*. SLIM-CD leads to slower growth but could prove useful for characterizing *L. iners* nutrient requirements or metabolite production. A modified version of SLIM-V supports growth of human cervical epithelial cells and provides a base for future co-culture work. Here, we present the formulations of SLIM, SLIM-V and SLIM-CD, and compare the growth of bacterial strains and human cells in the media.

Introduction

The composition of the vaginal microbiome is closely tied to host health. Colonization by *Lactobacillus* bacterial species, like *L. crispatus, L. gasseri* and *L. jensenii*, is considered optimal while colonization by other anaerobes, like *Gardnerella* and *Prevotella* species, can lead to bacterial overgrowth, resulting in bacterial vaginosis (BV)[1]. BV is the most common cause of vaginal complaints among reproductive-age women, affecting an estimated 23-29% globally[2]. In addition to vaginal symptoms, BV is associated with increased risk of preterm delivery, low birth weight, cervical cancer, and acquisition of several sexually transmitted infections, including HIV and herpes[3-7]. After standard antibiotic treatment, BV will recur in over 50% of patients[8]. A better understanding of the dynamics of the vaginal microbiome are needed to improve treatment and prevention efforts.

Lactobacillus species are thought to protect against BV by directly or competitively inhibiting growth of BV-associated bacteria (BVAB)[9]. A significant proportion of vaginal microbiomes are dominated by one of four *Lactobacillus* species: either *L. crispatus, L. iners, L. gasseri,* or *L. jensenii,* with the former two being more prevalent[10-13]. *L. iners* is estimated to be the most prevalent vaginal bacteria globally[14]. Unfortunately, its role in vaginal health is controversial, with some labeling *L. iners*-dominated microbiomes as a 'transitional' state, between the other 'healthy' *Lactobacillus* and BV[15, 16]. *L. iners*-dominated microbiomes transitioned to BV more frequently on average than those dominated by the other *Lactobacillus*, though notably some *L. iners*-dominated women were stably colonized by *L. iners* while others had frequent BV transitions[17]. Unlike the other *Lactobacillus*, *L. iners* frequently co-colonizes the vagina along with BVAB during BV[11]. Additionally, *L. iners* produces a pore-forming toxin, inerolysin, capable of lysing human epithelial cells, which has elevated expression in patients with BV[18, 19]. However, there is also evidence that some *L. iners* strains may be able to inhibit growth of BVAB, as some encode lanthipeptides, one of which inhibited growth of *Gardnerella vaginalis* when expressed in *Escherichia coli*[20].

Several have proposed that some strains of *L. iners* may stably colonize the vaginal microbiome and protect against BV, while others are permissive to BV[14-16]. To investigate this, the impact of *L. iners* strains on BVAB, other *Lactobacillus*, and host cells *in vitro* needs to be determined. While these analyses have been performed with the other *Lactobacillus* species, *L. iners* is frequently omitted[21, 22]. One study examining interactions of *Lactobacillus* and BVAB in co-culture included only a single *L. iners* strain, and another study comparing how supernatant from strains of *Lactobacillus* impacted *G. vaginalis* and *Prevotella bivia* growth excluded *L. iners* completely[21, 22]. This may have been due to the lack of liquid media for robust *L. iners* growth.

L. iners does not grow in de Man-Rogosa-Sharpe (MRS) broth, the standard cultivation media for *Lactobacillus* species[23, 24]. Recently, Bloom *et al.* published a medium for *L. iners* growth that consisted of MRS with additional cysteine and glutamine (MRS-CQ)[25]. In MRS-CQ, they found peak *L. iners* growth required 36 hours of incubation[25]. This is

suboptimal for co-culture growth assays with other *Lactobacillus*, which grow to high density after just 18 hours in MRS. MRS-CQ was not designed to mimic the native vaginal environment and does not support growth of human cells, which would be useful for creating better models of the vaginal environment. Additionally, MRS is not a chemically defined medium[24]. It is composed of beef extract, yeast extract, and peptone, which is a tryptic meat digest, which vary by batch or manufacturer and make analyzing nutrient requirements or produced metabolites difficult[24].

To facilitate further *in vitro* characterization of *L. iners*, we have developed three inexpensive and easy to make liquid media formulations for improved *L. iners* growth. Serrador's *Lactobacillus*-adapted Iscove's Medium (SLIM) leads to rapid and robust growth of several *L. iners* strains, as well as vaginal *Lactobacillus* and *Gardnerella* strains. *L. iners* growth is slightly improved by a vaginally adapted medium (SLIM-V). A modified version of SLIM-V can be used for culturing cervical epithelial cells. Finally, a fully defined medium (SLIM-CD) grows several *L. iners* strains and will prove useful for metabolite characterization or analyzing the impact of specific nutrients on *L. iners* growth.

Methods

Media preparation

To create MRS-CQ, MRS broth (NutriSelect® Basic, Sigma-Aldrich, St. Louis, MO, 69966-500G) with 0.1% TWEEN® 80 (see table 1) was autoclaved before addition of L-Cysteine and L-Glutamine (see table 1) at concentrations used by Bloom *et al* (4 mM and 1.1 mM, respectively)[25]. Media was passed through a 0.2 µm filter (Nalgene™, ThermoFisher, Waltham, MO, 595-5420) and stored at 4 °C until use.

SLIM is created by adding nutrients to Iscove's Modified Dulbecco's Medium (IMDM), a chemically defined, commercially available cell culture medium[26]. To create SLIM, IMDM is supplemented with peptone, L-Cysteine, L-Glutamine, a trace mineral supplement and TWEEN® 80 (Table 1). L-Cysteine and L-Glutamine concentrations were increased as used by Bloom *et al*[25].

SLIM formulations were prepared by creating the liquid base, then adding an additional amino acid source and, if desired, vaginal adaptations. After preparation, media was passed through a 0.2 µm filter (Nalgene™, ThermoFisher, Waltham, MO, 595-5420) and stored at 4 °C until use.

A vaginal-adapted version of SLIM (SLIM-V) was developed by adding urea, glycerol and acetic acid, which were present in a simulated vaginal fluid developed by Owen & Katz[27] (Table S1). To create SLIM-V, urea, glycerol, and acetic acid, at concentrations used in the vaginal fluid simulant, were added to SLIM (Table 1).

We observed that *L. iners* grew in SLIM-V when peptone was replaced by a nucleotide supplement (Table 2) and casamino acids (Bio Basic, Markham, ON, CB3060), an acid-hydrolysis of the milk protein casein (Figure S1). To create a chemically defined medium

114 (SLIM-CD), we replaced peptone with the nucleotide supplement and amino acids
115 mimicking the composition of casein supplements reported by Rasmussen *et al*[28] (Table
116 1).

117 A 10X nucleotide stock was prepared by dissolving nucleotides in ultra-pure water, then
118 passing the solution through a 0.2 µm filter (Nalgene™, ThermoFisher, Waltham, MO,
119 595-5420). The stock was stored at -20 °C until use. Concentrations were based on 10X
120 ACGU solution (TekNova, Hollister, CA) (Table 2).

121 Bacterial cultivation

122 All bacterial cultivation and growth assays were performed in an anaerobic chamber
123 (AS-580, Anaerobe Systems, Morgan Hill, CA) using a gas mixture of 10% CO_2, 10%
124 H_2, balance N_2 (Linde, Mississauga, ON). Liquid media was allowed to deoxygenate in
125 the chamber for at least 24 hours prior to use, and agar plates were deoxygenated for at
126 least 7 hours prior to use. *L. iners* and *Gardnerella* species were grown on New York
127 City III (NYC III) agar plates[29]. All other lactobacilli, including vaginal *Lactobacillus*, were
128 grown on MRS agar plates. Bacterial stocks were maintained at -80 °C in brain-heart
129 infusion broth (BHI) (Millipore, Darmstadt, Germany, 75917-500G) with 20% glycerol
130 (BioShop, Burlington, ON, GLY002.4). After streaking frozen stocks onto plates, plates
131 were incubated at 37 °C for 48-66 hours.

132 Bacterial strains

133 Experiments included previously reported strains, as well as 6 *L. iners* isolates, 1 *L.*
134 *crispatus* isolate and 1 *Gardnerella* isolate cultivated from cervicovaginal secretions
135 (Table 3). Cervicovaginal secretion samples were provided by the lab of Dr. Rupert Kaul
136 (Immunology, University of Toronto). In brief, female sex workers in Nairobi, Kenya were
137 recruited through the Sex Worker's Outreach Program (SWOP) clinics, and they were
138 provided informed, written consent for immune and microbial studies (REB approval in
139 protocol #37046). Consenting participants provided cervicovaginal secretions collected
140 by SoftCup (Evofem, San Diego, CA). Secretions were diluted 10-fold in sterile PBS,
141 then centrifuged at 1730 x g for 10 minutes, after which supernatant was extracted and
142 pellets were resuspended in 500 µL PBS. Both were frozen at -80 °C and transported to
143 the Kaul lab at the University of Toronto for analysis.

144 To isolate bacteria from the cervicovaginal secretions, the pellets were thawed and 10-
145 fold serial dilutions in sterile 1X PBS were created. Dilutions were spread onto agar
146 plates (NYC III for *L. iners* and *Gardnerella* isolations, MRS for *L. crispatus* isolations)
147 and incubated for 48 hours at 37 °C. Single colonies were picked and restruck onto agar
148 plates, then incubated for 48 hours at 37 °C. This was repeated twice, at which point
149 colony morphology for each isolate was consistent, and frozen stocks were created.

150 Each isolate was identified using PCR with species-specific primers (Table S2). Isolates
151 NVM020S08 and NVM020S14 were isolated from the same vaginal sample, and
152 isolates NVM025S01, NVM025S03, NVM025S06 and NVM025S07 were also isolated

153 from one sample. Thus, it is possible within each set of isolates, there is the same
154 strains cultured multiple times. However, we believe them to be different strains as
155 random amplified polymorphic DNA typing yielded differing banding patterns for these
156 isolates[30].

157 Growth curve measurements

158 In the anaerobic chamber, for each media type, three 13 mL polypropylene tubes
159 (Sarstedt, Nümbrecht, Germany, 62.515.006) per bacterial strain tested were prepared
160 with 4 mL of the deoxygenated media. Tubes were inoculated with 2-3 bacterial colonies
161 from fresh agar plates, then incubated anaerobically at 37°C. At each timepoint, cultures
162 were removed from the incubator, resuspended, and 200 µL from each culture, as well
163 as media blanks, was aliquoted into a 96-well plate (Sarstedt, Nümbrecht, Germany,
164 83.3924), after which cultures were returned to the incubator. The 96-well plate was
165 removed from the anaerobic chamber, and optical density at 600 nm (OD_{600}) was
166 measured using a SpectraMax Plus 384 Microplate Reader (Molecular Devices, San
167 Jose, CA).

168 Cell Culture and Viability Assay

169 Human endometrial carcinoma cell line (HEC-1-B) was acquired from Dr. Scott Gray-
170 Owen (Molecular Genetics, University of Toronto). The epithelial cells were maintained
171 in DMEM (Wisent, Saint-Jean-Baptiste, QC, 319-005-CL) supplemented with 10% Fetal
172 Bovine Serum (Gibco, Waltham, MO, 12483-020) and 1% Penicillin-streptomycin
173 (Gibco, Waltham, MO, 15140-122). Cells were grown at 37 °C in 5% CO_2. The viability
174 of the HEC-1-B cells in the developed media were tested using a resazurin assay,
175 where the fluorescence of metabolized resazurin (resorufin) is indicative of the number
176 of viable cells[31]. The HEC-1-B cells were harvested from T25 flasks using a standard
177 trypsinization protocol and passaged in DMEM into a 96-well plate (100µl /well).
178 Following overnight growth (50-60% confluency), the media was removed from the wells
179 and the adherent cells were washed with 1X PBS. Experimental media (SLIM, SLIM-V
180 and SLIM-CD) and control media (DMEM and IMDM) were aliquoted into the
181 appropriate wells in triplicate. A set of wells without cells were reserved as media-only
182 controls. After a 3-hour incubation the cells were imaged using a light microscope (Evos
183 FLoid, ThermoFisher, Waltham, MO, 4471136) before 20 µl of a 0.15 mg/ml resazurin
184 solution was added to each well and incubated for 4 hours. An Infinite M Nano$^+$
185 microplate reader (Tecan, Mannedorf, Switzerland, 30190087) was used to determine
186 cell viability by measuring fluorescence at 560 nm excitation and 590 nm emission.

187 **Results**

188 *L. iners* growth in SLIM formulations

189 For all *L. iners* strains tested, growth was significantly greater in SLIM and SLIM-V
190 compared to MRS-CQ, with all strains but *L. iners* ATCC 55195 showing no growth in
191 MRS-CQ after 66 hours (Figure 1). This is in accordance with previous experiments in

our lab, which saw turbid growth in MRS-CQ for *L. iners* SPIN 2503V10-D only after 6 days of incubation (Figure S2). For all strains, growth was slightly (though sometimes not significantly) improved in SLIM-V compared to SLIM (Figure 1). SLIM-CD grew all strains except NVM020S08 and NVM020S14, though peak growth did not occur until 66 hours, unlike SLIM and SLIM-V with peak growth at 24 or 42 hours (Figure 1).

Growth of other bacteria in SLIM

SLIM and SLIM-V support growth of strains of *L. crispatus* and *L. jensenii*, as well as a strain of *L. gasseri, L. paragasseri, G. vaginalis,* and an unclassified *Gardnerella* species (Figure 2). Growth in SLIM and SLIM-V was similar (Figure 2).

SLIM additionally supports growth of several non-vaginal lactobacilli, including *Limosilactobacillus reuteri, Lactiplantibacillus plantarum, Ligilalactobacillus murinus, Ligilalactobacillus ruminis, Logiolactobacillus coryniformis, Lacticaseibacillus rhamnosus, Companilactobacillus farciminis, Lactobacillus intenstinalis, Lactobacillus psittaci* and *Lactobacillus johnsonii* (Table S3, Figure S3).

Growth of Human Cells

The HEC-1-B epithelial cells were viable in both SLIM and SLIM-V; however, viability was significantly dampened in SLIM-CD (Figure 3). The inclusion of TWEEN® 80 in all media formulations, except for SLIM-CD, resulted in increased cell viability based on higher levels of resazurin metabolism (Figure 3). However, the morphology of the cells in media formulations containing TWEEN® 80 displayed considerably more vacuoles compared to media without TWEEN® 80 (Figure 4), suggesting that HEC-1-B cells are under stress in the presence of TWEEN® 80. Comparing both viability and morphology, SLIM-V without TWEEN® 80 is the condition most like standard DMEM (Figures 3 & 4).

Discussion

The role of *L. iners* in the vaginal microbiome is currently poorly understood, in large part due to difficulties in cultivation. Here, we present three media formulations, two leading to robust growth of several *L. iners* strains and other vaginal bacteria, and one that is chemically defined for nutrient and metabolite analysis. These are not minimal media, containing only the nutrients necessary for growth, as we aimed to grow a range of *L. iners* strains as well as other vaginal bacteria and human cells. The trace mineral supplement could likely be replaced with fewer components, and it is probable that not all amino acids added to SLIM-CD are necessary, especially as IMDM already contains a complete set of amino acids. Initial testing suggested that a mixture of several amino acids needed to be added to the IMDM to support growth in SLIM-CD, rather than a single amino acid being required in high amounts.

It is surprising that *L. iners* growth is improved in SLIM compared to MRS-CQ (Figure 1), which is a nutritionally richer medium. There are two potential explanations: that a component of MRS is inhibitory to *L. iners* growth and absent from SLIM, or that components of SLIM that are absent in MRS lead to improved growth. In preliminary

testing, *L. iners* ATCC 55195 and SPIN 2503V10-D growth in a 1:1 mixture of MRS-CQ and SLIM was similar to growth in SLIM (Figure S4). This suggests the latter hypothesis is more likely. One possibility is the presence of free amino acids in IMDM, as opposed to amino acids in peptides, in MRS. Bloom *et al.* suggested *L. iners* has a limited capacity to use L-Cysteine from complex sources, which was why additional free L-Cysteine needed to be added to MRS to grow *L. iners* in MRS-CQ[25]. If this is occurring with additional amino acids, IMDM with free amino acids makes a more suitable media base than MRS. Our observed growth in MRS-CQ (Figure 1, Figure S2) was significantly slower than that observed by Bloom *et al.,* where growth was seen in 35 hours[25]. This is likely due to differences in cultivation techniques. Larger volumes require more time for dense bacterial growth, and Bloom *et al.* grew 250 µL of culture, while here we grew 5 mL[25]. Additionally, in Bloom *et al.* starting inoculums were bacteria grown on agar plates resuspended in PBS, while here we used 2-3 bacterial colonies[25].

Growth was slightly improved for both *L. iners* and epithelial cells in SLIM-V compared to SLIM (Figure 1), which was designed to be more similar to the native vaginal environment. Future work could determine the individual impacts of urea, glycerol and acetic acid. One component absent from SLIM-V that is present in the vaginal tract is glycogen, which has been linked to *Lactobacillus* presence and improves *L. crispatus* growth[21, 32]. Future investigations into the impact of glycogen on *L. iners* could use SLIM-V with additional glycogen or create a modified version of SLIM-CD where glycogen is the primary carbon source instead of glucose. While SLIM-CD grew most *L. iners* strains, it did not grow *L. iners* NVM020S08 and *L. iners* NVM020S14 within 66 hours (Figure 1). As both isolates are from the same participant, this could suggest that isolates from the same microbiome share nutrient preferences. It is also possible that both isolates are clonal.

Co-culture experiments with human and bacterial cells are commonly done in a 3-hour window and it is essential to use a media that supports the growth of both organisms and/or does not inhibit their growth. Of the media formulations tested, SLIM-V without TWEEN® 80 is the most comparable to DMEM in supporting uterine HEC-1-B epithelial cells. TWEEN® 80 is reported to damage epithelial cells, so disrupted cell morphology seen in media with TWEEN® 80 (Figure 4) is not unexpected[33]. Cell stress may be increasing metabolic rate, resulting in the elevated resazurin metabolism seen from cells in media with TWEEN® 80 (Figure 3). TWEEN® 80 is necessary for *L. iners* growth, but TWEEN® 80 concentrations could be altered to attempt to reduce epithelial cell stress. Cells are still viable with TWEEN® 80, so SLIM-V could be used for short term assays. SLIM-V provides a base for future co-culture experiments between *L. iners* and vaginal epithelial cells.

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368 **Table 1: Formulations for SLIM, SLIM-V and SLIM-CD.** Recipes to create 1 L of
369 media. (-) indicates component not added. When a component's amount differs
370 between formulations, values are bolded and underlined.

		SLIM	SLIM-V	SLIM-CD	Supplier	CAT #
Liquid base	IMDM, no phenol red	989 mL	989 mL	**<u>889 mL</u>**	Gibco	21056-023
	Trace mineral supplement	10 mL	10 mL	10 mL	ATCC	MD-TMS
	TWEEN® 80	1 mL	1 mL	1 mL	BioShop	TWN507.100
	10X nucleotide stock	-	-	100 mL	*See Table 2*	
Amino acid source	Meat Peptone	10 g	10 g	-	Criterion	C7481
	L-Cysteine-HCl	0.64 g	0.64 g	0.64 g	Sigma	30120-50G
	L-Glutamine	0.16 g	0.16 g	0.16 g	BioShop	GLU102.25
	L-Glutamic acid	-	-	2.02 g	BioShop	GLU202.100
	L-Proline	-	-	1.02 g	BioShop	PRO222.25
	L-Leucine	-	-	0.83 g	Sigma	L8912-25G
	L-Lysine-HCl	-	-	0.74 g	Bio Basic	LB0171
	L-Valine	-	-	0.65 g	BioShop	VAL201.25
	L-Aspartic acid	-	-	0.64 g	BioShop	ASP003.25
	L-Serine	-	-	0.57 g	Aldrich	S2600-10G
	L-Tyrosine	-	-	0.57 g	BioShop	TYR333.50
	L-Isoleucine	-	-	0.55 g	BioShop	ISO910.10
	L-Phenylalanine	-	-	0.45 g	Bio Basic	PB0422
	L-Threonine	-	-	0.44 g	BioShop	THR002.25
	L-Arginine-HCl	-	-	0.37 g	BioShop	ARG006.100
	L-Histidine-HCl	-	-	0.28 g	Bio Basic	HB0330
	L-Alanine	-	-	0.27 g	BioShop	ALA001.25
	L-Methionine	-	-	0.25 g	Sigma	M9625-100G
	Glycine	-	-	0.24 g	Sigma	G8898-1KG
	L-Tryptophan	-	-	0.11 g	BioShop	TRP100.25
Vaginal adaptation	Urea	-	0.4 g	0.4 g	BioShop	URE001.500
	Acetic acid	-	960 µL	960 µL	Caledon	1000-1-29
	Glycerol	-	128 µL	128 µL	BioShop	GLY002.4

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372 **Table 2: Composition of nucleotide stock.** Recipe for 500 mL of 10X stock.
373 Concentrations based on 10X ACGU solution (TekNova, Hollister, CA).

Compound	Amount	Supplier	CAT #
ddH_2O	498.75 mL	-	-
Potassium hydroxide	0.421 g	BioShop	PHY202.500
Guanine	0.151 g	BioShop	GUA904.5
Adenine HCl	0.135 g	Sigma	A8751-5G
Uracil	0.112 g	Sigma	U-0750
Cytosine	0.111 g	BioShop	CYT301.5

374

375 **Table 3: Vaginal bacterial strains used in this study.**

Species	Strain	Source	Original Study
Lactobacillus iners	ATCC 55195	Human vagina	Human Microbiome Project
Lactobacillus iners	SPIN 2503V10-D	Patient with bacterial vaginosis	Human Microbiome Project
Lactobacillus iners	NVM020S08	Cervicovaginal secretions	This study
Lactobacillus iners	NVM020S14	Cervicovaginal secretions	This study
Lactobacillus iners	NVM025S01	Cervicovaginal secretions	This study
Lactobacillus iners	NVM025S03	Cervicovaginal secretions	This study
Lactobacillus iners	NVM025S06	Cervicovaginal secretions	This study
Lactobacillus iners	NVM025S07	Cervicovaginal secretions	This study
Lactobacillus paragasseri	JV-V03	Female urogenital tract	Human Microbiome Project
Lactobacillus gasseri	EX336960VC01	Human mid-vaginal wall	Human Microbiome Project
Lactobacillus crispatus	PSS7772C	Urine of a pregnant woman	Human Microbiome Project
Lactobacillus crispatus	NVM146S01	Cervicovaginal secretions	This study
Lactobacillus jensenii	269-3	Human vaginal mucosa	Human Microbiome Project
Lactobacillus jensenii	EX849587VC03	Human mid-vaginal wall	Human Microbiome Project
Gardnerella vaginalis	ATCC 14019	Vaginal secretions	Human Microbiome Project
Gardnerella sp.	NVM022C02	Cervicovaginal secretions	This study

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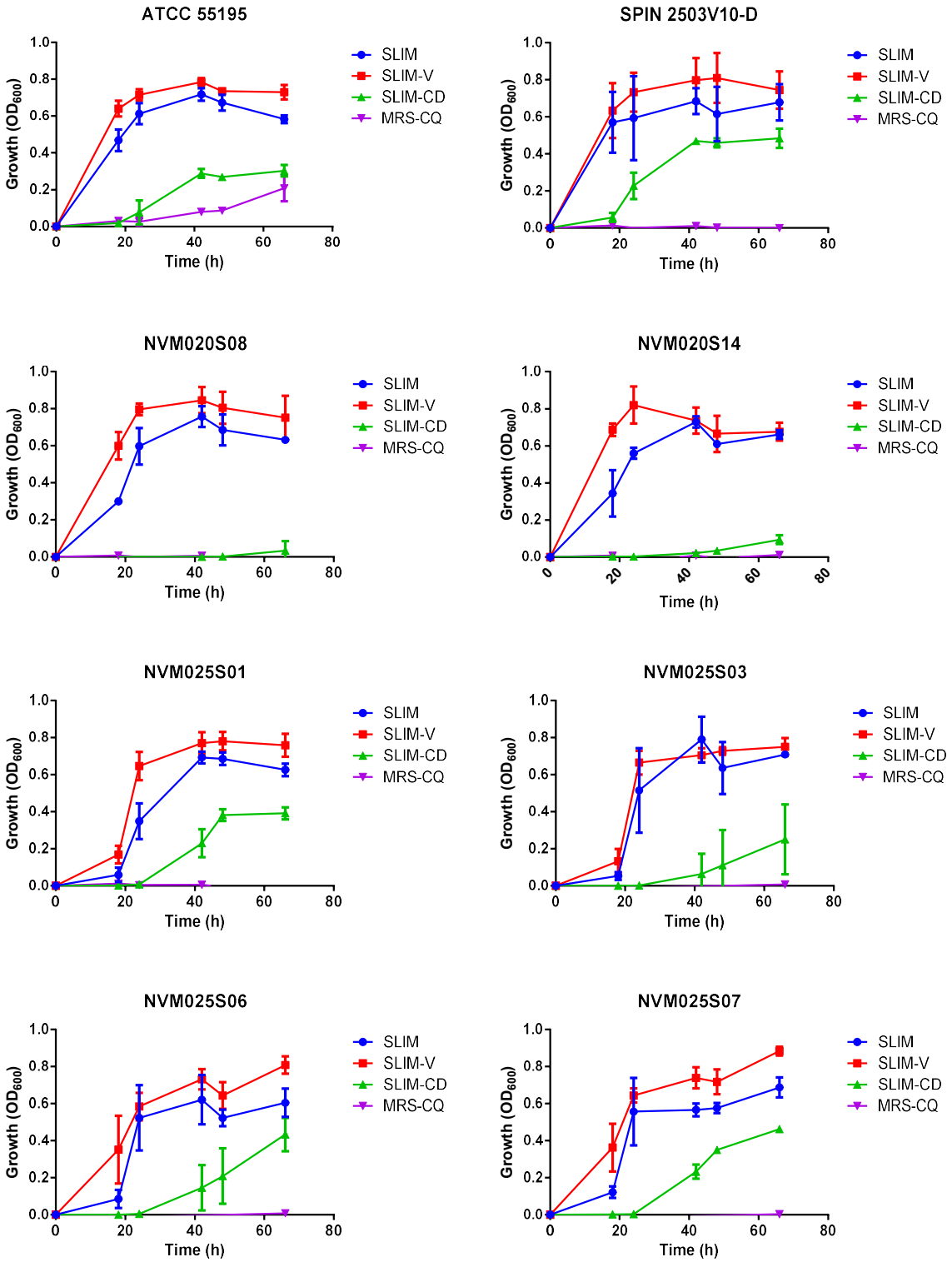

377

378 **Figure 1: Growth of *L. iners* strains in SLIM, SLIM-V, SLIM-CD and MRS-CQ.** OD_{600}
379 measured using SpectraMax Plus 384 Microplate Reader (Molecular Devices, San
380 Jose, CA). n = 3, mean ± SD.

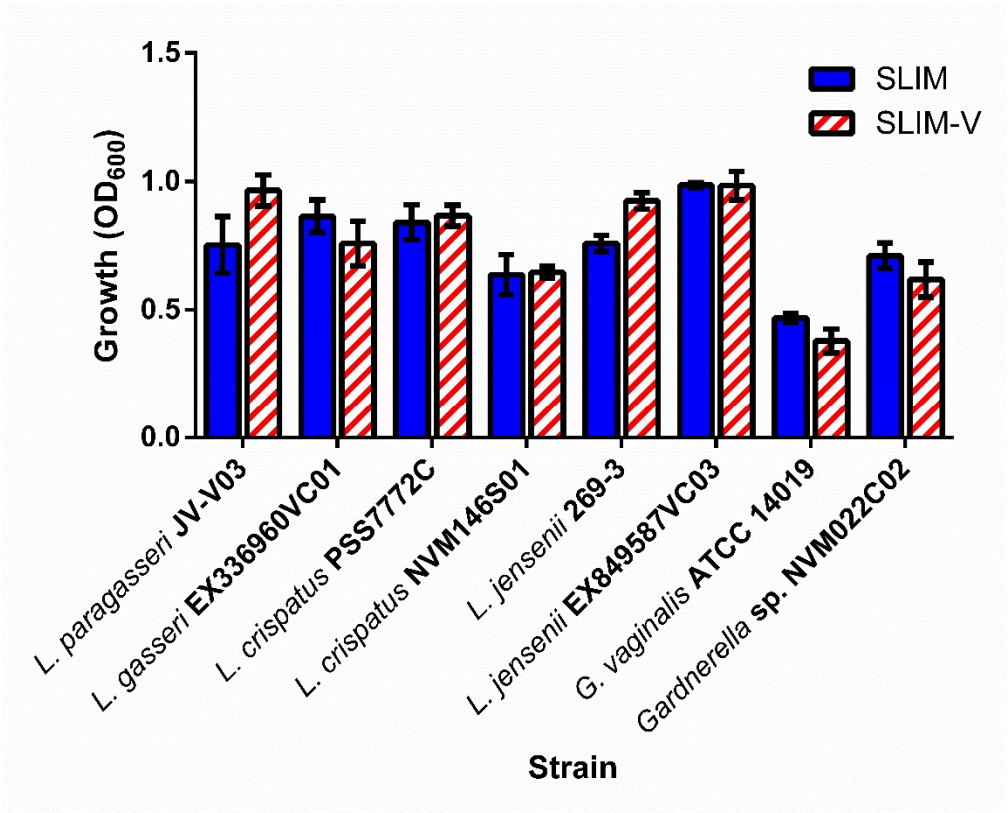

Figure 2: Growth of vaginal bacteria in SLIM and SLIM-V. Growth measured after 24 hours of incubation at 37 °C. OD_{600} measured using SpectraMax Plus 384 Microplate Reader (Molecular Devices, San Jose, CA). n = 3, mean ± SD.

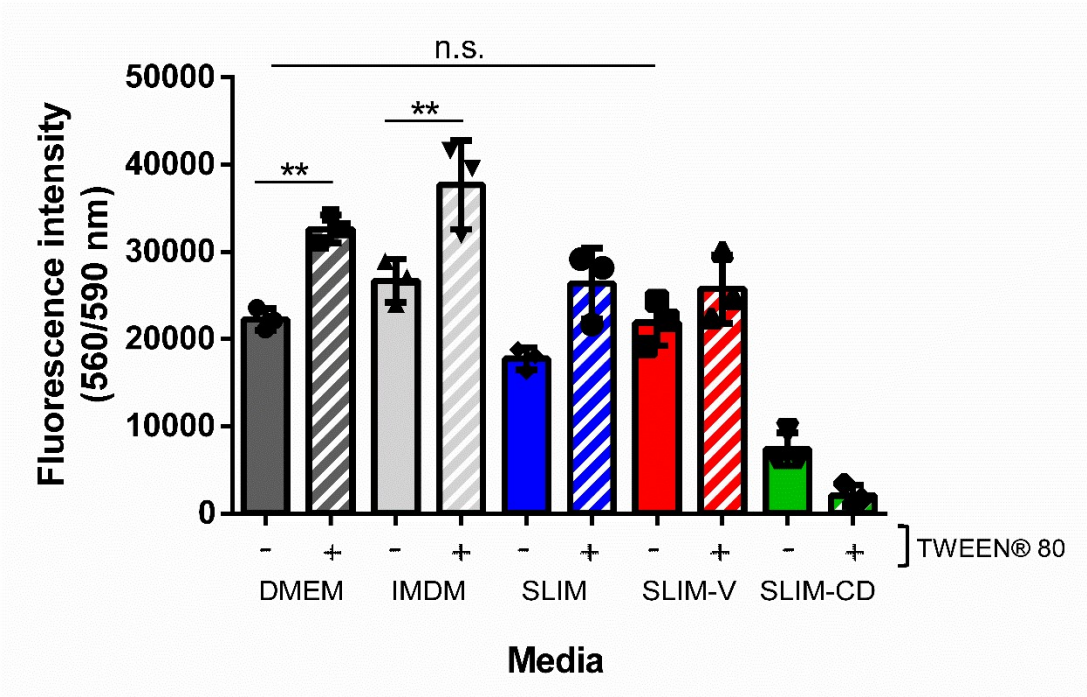

Figure 3: Resazurin-based cell viability of HEC-1-B cells in SLIM formulations. The viability of the cells is directly proportional to the fluorescence intensity of resorufin produced by live cells metabolizing resazurin[31]. Fluorescence intensity of the media-only controls is subtracted from the fluorescence intensity of the appropriate sample wells. Each bar represents the mean ± SD, n=3. Statistical significance was calculated using one-way ANOVA, *$p < 0.05$.

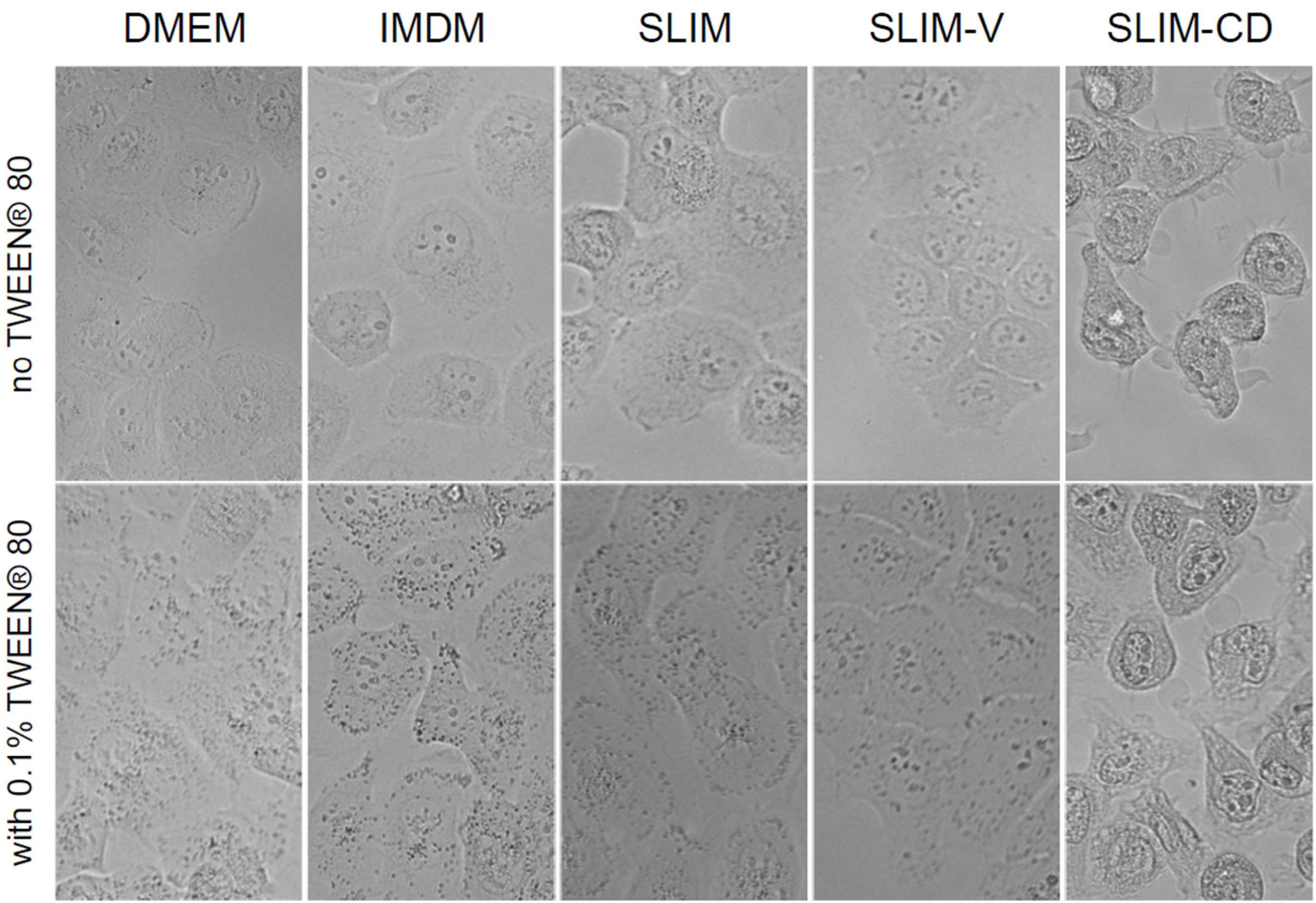

Figure 4: Light microscopy images of HEC-1-B cells in media after 3 hours of incubation.