Complex genomic ancestry in southern regions and drivers of continental-level genetic diversity in the wolves of Asia

Authors

Lauren M. Hennelly[1,2], Barbara R Parreira[1], Ash Noble[3], Camilla Scharff-Olsen[1], M. Çisel Kemahlı Aytekin[4], Çağan H. Şekercioğlu[4,5,6], Pavel Kosintsev[7,8], Ladislav Paule[9], Pavel Hulva[10,11], Hans K. Stenøien[12], Bilal Habib[13], Hira Fatima[14], Ghulam Sarwar[15], Samara P. El-Haddad[16], Frank Hailer[3,17], Xin Sun[1], Nuno Filipes Gomes Martins[1], M Thomas P Gilbert[1,6], Mikkel-Holger S. Sinding[18], Benjamin N Sacks[19], Shyam Gopalakrishnan[1]

[1]Section for Evolutionary Hologenomics, Globe Institute, University of Copenhagen, Denmark

[2]Center for Conservation Genomics, Smithsonian's National Zoo and Conservation Biology Institute, Washington, DC, USA

[3]Organisms and Environment, School of BiosSciences, Cardiff University, Cardiff, UK

[4]Department of Molecular Biology and Genetics, Koçc University, İlstanbul, Türkiye

[5]School of Biological Sciences, University of Utah, Salt Lake City, 84112, UT, USA

[6]Kuzey Doğa Society, Kars, Türkiye

[7]Ural Federal University, Yekaterinburg, Russia

[8] Institute of Plant and Animal Ecology, Ural Branch of Russian Academy of Science, Yekaterinburg, Russia

[9]Technical University in Zvolen, Zvolen, Slovakia

[10]Department of Zoology, Charles University, Prague, Czechia

[11]Biology and Ecology, University of Ostrava, Ostrava, Czechia

[12]NTNU University Museum, Norwegian University of Science and Technology,

Trondheim, Norway

[13]Wildlife Institute of India, Dehradun, India

[14]University of Education – Attock Campus, Attock, Pakistan

[15]University of the Punjab, Lahore, Pakistan

[16]Lebanese Wildlife, El Metn, Lebanon

[17]Cardiff University-Institute of Zoology Joint Laboratory for Biocomplexity Research

(CIBR), Beijing, China

[18]Department of Biology, University of Copenhagen, Copenhagen, Denmark

[19] Mammalian Ecology and Conservation Unit, Veterinary Genetics Laboratory,

Department of Population Health and Reproduction, School of Veterinary Medicine,

University of California, Davis, California, USA

38
39   **Corresponding author**: Lauren M. Hennelly laurenmaehennelly@gmail.com

56
57 **Abstract**

58
Gray wolves (*Canis lupus*) in Asia encompass most of the species' global genetic

diversity and many endangered populations. However, a clear understanding of the

evolutionary history of wolves from many parts of Asia, especially southern regions, is

lacking. We used 98 whole genomes of wolves sampled across Eurasia to better

resolve their evolutionary history by investigating phylogenetic and gene flow histories

across the genome, and to characterize their demographic history and genetic diversity.

The strongest barriers to gene flow coincided with boundaries separating the three

major extant wolf lineages - Indian, Tibetan, and Holarctic. Wolves in the central Asian

mountain ranges belonged to the Holarctic lineage, and despite their geographic

adjacency only shared minimal ancestry with the Tibetan lineage. In contrast, wolves

from eastern Asia share population-wide ancestry with the Tibetan lineage, which may

reflect an unsampled lineage similar, but not exactly to, the modern-day Tibetan

lineage. Wolves from southwestern Asia also share population-wide ancestry with the

Indian lineage, likely due to old (>6kya) admixture events. Long-term historical declines

over the last 100,000 years, geographic isolation, and recent inbreeding have resulted

in the Indian and Tibetan wolves having some of the lowest levels of genetic diversity

and highest realized genetic loads. In contrast, adjacent populations exhibit some of the

highest genetic diversity, due in part to admixture along contact zones. Our study

illustrates how using multiple approaches that consider heterogenous signals across the

genome can more fully resolve the historical and contemporary processes that have led

to present-day species' diversity.

Keywords: *Canis lupus,* demographic history, genetic diversity, gray wolf,

phylogenomics

83
84
Introduction

86
Understanding the processes that shape genetic variation within a species is

fundamental in evolutionary biology. Genetic variation is often unevenly distributed

across a species' range, and assessing where and how it varies provides insight into

underlying evolutionary processes. Because multiple evolutionary processes impact a

species' genetic variation, it has long remained a methodological challenge to

distinguish between these processes, such as intrapopulation and interspecies gene

flow, genetic drift, and incomplete lineage sorting. The contribution of these processes

also varies temporally over a species' evolutionary history, further challenging

inferences. Recent methodological advances now consider genomes as mosaics of

evolutionary history whereby different genomic regions have distinct evolutionary

histories (Martin et al. 2017, Excoffier et al. 2021, Harris et al. 2022). Carefully

interpreting these distinct signatures across the genome has recently improved our

understanding of the evolutionary histories of notoriously complex groups (Figuiro et al.

2019, Lescroart et al. 2023, Jensen et al. 2023).

101
Widely distributed species offer a unique opportunity to study how genetic variation is

shaped by historical and contemporary environmental changes. Because they are so

widespread, their populations experience different histories of isolation and

interpopulation or interspecific gene flow, often in response to past and present

environmental change. Applying new methodological approaches to widely distributed

species can better resolve their evolutionary histories and offer additional insight into

how periodic gene flow and isolation in different parts of their distribution has given rise

to present-day patterns in genetic variation within species.

110
Gray wolves are one of the most widely distributed and best-studied large mammals,

making them a promising system to explore how historical and contemporary processes

shape present-day genetic variation. Due to their scientific and public appeal, they were

one of the first large carnivores to be studied using genomic tools (vonHoldt et al. 2011,

Stronen et al. 2013). Over the years, genomic work of wolves has grown into large-

scale studies using numerous modern and ancient genomes. This has led to critical

insights on recognizing the role of hybridization in speciation and impacts of the late

Pleistocene on species' persistence (Gopalakrishnan et al. 2018, Bergstrom et al.

2022). However, most such studies have been on North American and European

portions of the range. Fewer genomic studies have focused on wolves in Asia, which

today holds most of their phylogenetic diversity and many endangered populations.

Gray wolves are composed of three extant divergent lineages – the Indian, Tibetan, and

Holarctic – all of which occur in Asia (Wang et al. 2020, Hennelly et al. 2021, Wang et

al. 2022). These lineages diverged > 100,000 years ago after which the Indian and

Tibetan lineages became isolated within separate glacial refugia (Wang et al. 2020,

Wang et al. 2022). In contrast, wolves of the Holarctic lineage, spanning northern

latitudes from northern Europe to Siberia, as well as North America, were highly

connected by gene flow during the past 100,000 years (Bergstrom et al. 2022, **Figure**

S1). This high gene flow resulted in genome-wide homogenization of wolves in northern

Eurasia during the late Pleistocene (Bergstrom et al. 2022). While this has provided a

broad picture of wolf evolutionary history in Eurasia, few studies have examined how

ancestral composition and genetic diversity is distributed in wolves across Eurasia,

especially in more southern regions of Asia where lineages occur in close proximity.

136
For example, earlier work based on from a handful of wolf genomes from southwestern

Asia (defined as the region spanning Iran to the Arabian Peninsula to Türkiye) largely

reflected Holarctic ancestry, with some minor ancestry from the Indian lineage, as well

as from another canid species, the African wolf (*Canis lupaster*) (Gopalakrishnan et al.

2018, Hennelly et al. 2021). However, a fuller understanding of this population's history

has been hampered by a lack of geographically spread samples from across

southwestern Asia. For wolves in Japan and parts of China, previous work found they

contain partial ancestry similar to ancient Pleistocene wolves from ~35-50kya, all of

which fell within the Holarctic lineage based on mitochondrial DNA (Niemann et al.

2020, Madrigal et al. 2021, Segawa et al. 2022). In addition, some of their partial

ancestry was found to be ancestral to late Pleistocene wolves from northern Eurasia,

including ancient wolves dating back to 100 kya (Bergstrom et al. 2022). Together, this

suggests various wolf populations in the southern regions of Asia may retain diversity

that is ancestral to northern Eurasian wolves, and thus, were not completely

homogenized by gene flow from northern wolves during the late Pleistocene (Bergstrom

et al. 2022). It remains unclear what the geographical extent of ancestral diversity

(defined as ancestral to Holarctic wolves in the northern latitudes) within these wolf

populations is, and the spatiotemporal origin of this ancestry in their genomes. To

resolve this, additional samples and analyses are needed to clarify the ancestry

composition of wolves in southern regions of Asia, and what historical processes have

given rise to modern-day wolves in these southern regions.

We sought to address these four gaps in our knowledge about wolves in Asia through

building a genomic dataset that includes 20 newly sequenced wolves sampled from key

regions and 81 previously published wolf genomes from Eurasia and North America.

Our first objective was to characterize continental-level population structure and gene

flow patterns of wolves across Eurasia, with a focus on the southern regions of Asia.

Our second objective was to evaluate the presence and clarify the origin of ancestral

diversity (i.e. ancestral to Holarctic wolves in the northern latitudes) for wolf populations

in the central Asian mountain ranges and eastern regions of Asia by incorporating

recombination rate variation into phylogenetic analyses. Our third objective was similar

to objective two, but evaluating ancestral diversity within the southwestern Asian wolf

population using phylogenetic analyses and demographic modeling. For our fourth

objective, we use our genomic dataset to characterize the genetic diversity of wolves

throughout Eurasia, as well as inbreeding and genetic load.

172
Results

For this study, we resequenced new whole genomes of 20 wolves from various regions

in Eurasia. These regions included southwestern Asia (Lebanon, n=1; Israel, n=1,

Türkiye=1), Indus plains of Pakistan (n=2), central Asian mountain ranges (Sulieman,

n=2; Karakoram, n=2; Hindu Kush, n=3), the Eurasian steppe (Kazakhstan, n=3;

Russia, n=3), and in Europe (Slovakia, n=1; Ukraine, n=1). We also conducted

additional (i.e., deeper) sequencing of a previously sequenced Indian wolf from Central

India. We compiled these newly sequenced genomes with 81 previously sequenced

wolves from North America (n=3) and Eurasia (n=78), as well as dogs (n=5), and

related canid species (n=12) into a dataset, which we then aligned to the domestic dog

genome assembly (canFam3.1) (**Table S1**).

Population structure and gene flow patterns of wolves in Eurasia

Principal component analysis (PCA) and inferred individual admixture proportions of 98

Eurasian wolves primarily supported the presence of three major clusters corresponding

to the three major extant wolf lineages: the Indian, Tibetan, and Holarctic wolves

(**Figure 1A,1B, S2**). Some wolves fell in-between these main clusters, forming clines

between Tibetan and Holarctic as well as between Indian and Holarctic clusters. In both

cases, these intermediate individuals came from areas nearby the corresponding

contact zones between the lineages. Intermediate individuals on the Tibetan-Holarctic

cline included two wolves from Tajikistan (approximately 24% Tibetan ancestry at K=6)

and two wolves from Ladakh, which is a region on the southwestern edge of the Tibetan

plateau. Intermediate individuals on the Indian-Holarctic cline included only wolves from

Pakistan. In contrast, all 13 wolves from southwestern Asia clustered tightly nearby

other Holarctic wolves and exhibited ~15-20% Indian wolf ancestry at K=6. A few

wolves, most sampled near Africa, showed basal canid ancestry, likely reflecting

admixture with originating from the African wolves.

To better characterize geographic patterns of gene flow, we inferred the estimated

effective migration surface (EEMS), which indicates regions of low or high gene flow

relative to an isolation-by-distance model (Marcus et al. 2021). Mirroring the PCA and

admixture results, the most significant EEMS barriers to gene flow in Eurasia

corresponded to divisions among the three major lineages (**Figure 1C**). The strongest

barrier was along the Himalayan mountain range that separates the Indian and Tibetan

lineages. A second strong barrier surrounds the Tibetan plateau, in agreement with our

PCA and admixture results showing reduced gene flow between the Tibetan lineage

and wolves in central and eastern Asia. The positioning of another barrier west of

Pakistan was less clear, but may be artifactual due to our lack of sampling on the

Eurasian steppe, such as in Turkmenistan and Uzbekistan. Otherwise, the population

structure of Holarctic wolves across northern Eurasia can be modeled as isolation-by-

distance (**Figure 1B,C).**

Tibetan-like ancestry is rare in the central Asian mountains yet found

population-wide in eastern Asia

To explore the evolutionary history of wolves in the central Asian mountain ranges and

eastern Asia, we investigated the presence and potential source of ancestral diversity

(i.e. ancestral to wolves in northern Eurasia) within these two populations. We first used

D statistics to test for excess derived allele sharing with various wolf populations in Asia

(Green et al. 2010). For wolves in the central Asian mountains, we observed a wide

variance of allele sharing with the Tibetan lineage (**Figure 2A**). Apart from a few

individuals, most wolves in Pakistan, Tajikistan and Kyrgyzstan show no statistically

significant excess of derived alleles shared with the Tibetan lineage (**Figure 2A; Figure**

S3). A large inter-individual variance in Tibetan derived alleles may suggest that

admixture with Tibetan wolves occurs occasionally or is recent, rather than arising from

long-term admixture. In contrast, we observe a consistent and significant excess of

Tibetan wolf derived alleles across wolves from eastern China and Mongolia (**Figure**

2A, Figure S3, S4).

231
It can be difficult to identify the actual source using D statistics because any ancestry

more closely related to the Tibetan wolf will result in a negative D statistic. To better

understand the source and historical processes that may have resulted in Tibetan-like

ancestry, we inferred a set of phylogenetic trees from different regions of the genome.

We first inferred a multispecies coalescent tree in ASTRAL using 5,000 regions with a

length of 20kb across the autosomes using all 113 wild canid samples (Zhang et al.

2018). Tibetan wolves formed a well-supported branch basal to other Eurasian gray

wolves (**Figure 2B**). The rest of the wolves form weakly supported clades, including

Indian wolves which are clustered with wolves from Pakistan, and sister to those from

southwestern Asia (**Figure S5**).

242
Because the relationships inferred by genome-wide phylogenies can be impacted by

many processes, such as gene flow and incomplete lineage sorting (ILS), we leveraged

how genome architecture relates to phylogenetic signal. Briefly, the X chromosome and

low recombination regions are more likely to retain the historical relationship due to

higher selection against introgressed ancestry and higher lineage sorting (Schumer et

al. 2018, Martin et al. 2019). In particular, we took advantage of the negative

relationship between recombination and historical phylogenetic relationship that has

been demonstrated across multiple taxa, including wolves (Li et al. 2019, Hennelly et al.

2021, vonHoldt et al. 2021, Feng et al. 2023, Jiang et al. 2024, Monthey and Spellman

2024).

Our phylogenetic tree based on low-recombination regions (<0.2 cM/.Mb) of the X

chromosome demonstrated that one of the two wolves from Tajikistan that showed the

highest allele sharing with Tibetan wolves clustered within the Tibetan lineage, and all

other wolves from Tajikistan and Kyrgyzstan clustered with other Holarctic wolves

(**Figure 2C, Figure S6, Figure S7**). Wolves from eastern Asia formed a well-resolved

clade that branched basally to the rest of the Holarctic lineage. Our phylogeny using low

recombination regions (<0.2 cM/Mb) of the autosomes and the entire X chromosome

recapitulates this pattern (**Figure S8, Figure S9**). In addition, topology weighting

analyses show low recombination regions of the X chromosome are more resolved for

topologies where wolves from eastern Asia form the basal branch within the Holarctic

lineage (**Figure 2D, Figure S10**). Thus, while wolves in Tajikistan, Pakistan, and

eastern Asia have similar D-values, they have contrasting topological patterns. This is

consistent with wolves from Tajikistan and Pakistan having experienced recent and/or

periodic gene flow from Tibetan wolves near the secondary contact zone, yet this

Tibetan ancestry has not yet spread widely through the population. In contrast, wolves

in eastern China and Mongolia have retained population-wide ancestry that is more

ancestral than wolves in northern Eurasia, yet appears distinct from modern-day

Tibetan wolves.

274 **Population-wide Indian ancestry within wolves of southwestern Asia**

We detected complex ancestry patterns in wolves across southwestern Asia. Estimated

admixture proportions and D statistics suggest that wolves in this region shared

ancestry similar to the Indian lineage and another canid species, the African wolf

(**Figure 1A,B, Figure 3A,B**). We detected nine wolves that showed significant derived

allele sharing with the African wolf, which were largely confined to areas most

geographically close to Egypt (Saudi Arabia, Syria, Lebanon) (**Figure S11, S12**).

281
Interestingly, we find ancestry shared with the Indian lineage was nearly equal in all

wolves in southwestern Asia (**Figure 3A, S12, S13, S14**). There appeared to be no

geographic decline in the admixture proportions of assigned Indian ancestry spanning

the contact zone in Pakistan until reaching the Caucasus mountain range and from

Türkiye into Europe, where no Indian wolf ancestry was detected (**Figure 3A, Figure**

S14). There was also a lesser, yet significant, signal of ancestry similar to the Indian

lineage on parts of the Eurasian steppe in Central Asia, such as in Kazakhstan (**Figure**

S14). This geographically widespread signal contrasts what we observed with Tibetan

and African wolf ancestry, which declines quickly as distance increases from their

respective distributions (**Figure 3A**).

The balanced Indian wolf-like ancestry across southwestern Asia could be consistent

with an admixture event between Indian and Holarctic lineages. To investigate the origin

of this Indian-like ancestry, we used FASTSIMCOAL2 (Excoffier et al. 2021) to test three

demographic scenarios where (A) there is no introgression event between Indian

wolves and southwestern Asian wolves, (B) southwestern Asian wolves experience an

introgression event with Indian wolves, and (C) southwestern Asian wolves are formed

as a hybrid lineage between Indian and central Asian wolves (**Figure S15**). We

obtained similar likelihood scores for the three models, suggesting no one model is

strongly favored over the others (**Table S2**). However, topologies that included

bifurcation (Divergence and Introgression models) had the highest likelihoods. The

model that included introgression from Indian wolves to southwestern Asian wolves was

preferred, with a relative likelihood of 62% (**Table S2**). Estimates suggest a relatively

high introgression proportion (0.11) from India to southeast Asia, consistent with our D

statistic results (**Figure S14**). We estimate the introgression occurred around 6,000

years ago, relatively close to the divergence time between southwest and central Asian

populations about 10,300 years ago. Therefore, if introgression did occur, it likely

happened near the time of the split between southwest and central Asian populations,

explaining the similar likelihood between the divergence (A) and introgression (B)

models. Importantly, both models estimate similar dates for the split between

southeastern and central Asian lineages and all models date the divergence of Indian

wolves from Eurasian wolves to around 100,000 years before present, providing

robustness to time estimates. Additionally, both introgression and divergence models

produced similar estimates for population sizes, suggesting a small effective population

size (N_E) for India and southwest Asia, whereas central Asian populations and the

ancestral Asian population had a larger N_E (**Figure S15**). Our results indicate that the

Indian wolf ancestry within the genomes of southwestern Asian wolves reflects older

admixture (>6kya) and/or shared demographic history rather than appearing within the

past few last thousands of years.

Finally, to better assess how this Indian wolf-like ancestry relates to modern-day Indian

wolves, we inferred a set of phylogenetic trees to investigate the topological positions of

Indian and southwestern Asian wolves. In almost all inferred autosomal and X

chromosome trees, we observe the Indian lineage and wolves from Pakistan cluster

together, with almost all southwestern Asian wolves as a sister clade – all within the

Holarctic clade (**Figure S5, S6, S7, S8**). The clustering of the Indian lineage with wolves

in Pakistan implies they share more recent or prevalent ancestry, consistent with this

region being a prominent contact zone (Hennelly et al. 2023). When excluding wolves

from Pakistan and southwestern Asia, Indian wolves form the most basal branch of the

gray wolf phylogeny, in line with southwestern Asian wolves being the admixed

population (Wang et al. 2022, **Figure S17**). To investigate Indian-like segments within

southwestern Asian wolf genomes in more detail, we identified genomic regions across

the autosomes where Indian and southwestern Asian wolves were sister to each other

using topology weighting with Twisst (Martin and Belleghem 2017). For this, we

quantified the frequency of 105 possible topologies using 100-SNP windows across the

genome for five populations with the Andean fox (*Lycalopex culpaeus*) as an outgroup

(**Table S3**). The topology with the highest average weight was one where southwestern

Asian wolves were sister to Indian wolves, suggesting some of their genome shares a

more recent common origin rather than with other Holarctic wolves (**Figure 3C, Table**

S3).

We then inferred a maximum likelihood tree using only genomic regions that are most

reflective of Indian and southwestern Asian wolves being sister clades (>0.3 topology

weight for the highest supported topology; **Figure 3C**). In this maximum likelihood

phylogeny, we find the most basal wolf lineage consists of Indian and southwestern

Asian wolves, where southwestern Asian wolves fall outside of the diversity within the

modern-day Indian lineage (**Figure 3D, 3E, S18, S19**). One hypothesis for this finding is

that Indian-like ancestry within southwestern Asia may derive from a once more diverse

Indian lineage rather than arising directly from modern-day Indian wolves within the

Indian subcontinent. Alternatively, there could be some Holarctic ancestry within these

100-SNP windows, thus placing southwestern Asian wolves outside of modern-day

Indian wolf diversity. Our work emphasizes a complex evolutionary history of wolves in

southwestern Asia where additional studies are necessary to fully resolve it.

355
356
357
358   **Historical and present-day impacts on genetic diversity**

359
Our whole-genome dataset allowed us to assess patterns of genome-wide diversity of

wolves across Eurasia and North America. We calculated autosomal heterozygosity

using genotype likelihoods for 101 wolves from Eurasia (n=98) and North America

(n=3), and found wolves in Asia encompass a wide range of some of the lowest and

highest levels of genetic diversity among wolves in our dataset (**Figure 4A**). Along with

the well-known depleted heterozygosity of Mexican and Iberian wolves, we found

wolves in India and on the Tibetan plateau have some of the lowest documented

heterozygosity in our dataset. As observed in our Pairwise Sequentially Markovian

coalescent (PSMC) analysis, this remarkably low heterozygosity is in part due to their

long-term population declines in the last >100 kya rather than, or in addition to,

anthropogenic-related declines (Li and Durbin 2011) (**Figure 4B**). We also observe that

the wolf from Shanxi, China, had a lower population size throughout the Pleistocene

compared to other Holarctic wolves (**Figure 4B**). This may suggest the Shanxi wolf and

other wolves in southern China were less connected to northern wolf populations,

however, a greater number of higher coverage genomes are needed to rule out the

effects of low coverage. Otherwise, there is a shift to elevated heterozygosity coinciding

with secondary contact zones between the Holarctic lineage and the Indian and Tibetan

lineages, respectively. For example, admixed wolves in lowland Pakistan have higher

genome-wide heterozygosity than wolves of Central India, even though both

populations are similarly endangered and declining. We find the highest heterozygosity

among wolves in Asia is geographically clustered in Iran and Xinjiang of China (**Figure

4A**). These findings are concordant with our estimated inbreeding coefficients using

genotype likelihoods (Hanghoj et al. 2019) (**Figure S20).**

383
To further disentangle what historical and present-day processes may explain patterns

of genetic diversity in wolves, we inferred runs of homozygosity (ROH) for genomes with

moderate coverage (>15x) using BCFtools/RoH (Narasimhan et al. 2016). The lengths

of ROHs allowed additional insight into the timing of inbreeding. Combined with

knowledge of the generation time and recombination rate, we estimated the timing of

inbreeding, where, for example, a ROH length of 5Mb suggests inbreeding took place

~7 generations ago (Thompson 2013). We find that Indian and Tibetan wolves show the

most extensive inbreeding between 800-102 years compared to all other wolves in our

study (**Figure S21**). Along with the highest number of short ROHs (0.1-1Mb), Indian and

Tibetan wolves show evidence of larger ROH as well (5-10Mb), suggesting their low

heterozygosity is a combination of long-term isolation over the last 200kya and

anthropogenic pressures in the last hundreds of years (**Figure 4, Figure 3B**). We find

the well documented and dramatic population declines of Mexican and Iberian wolves

occurred more recently over the course of the last century, in line with previous work

(**Figure S21**, Taron et al. 2021, Gomez-Sanchez et al. 2018). Likewise, we observe a

few individuals in Asia possess long ROHs (5-100Mb), implying the slightly reduced

genetic variation in these wolves is due to recent declines in the last 100 years (**Figure

S21**). Thus, in contrast to genetic diversity patterns in Europe, our results suggest much

of the variation in genetic diversity in Asia can be explained by historical processes,

coinciding with glacial refugia and secondary contact zones.

404
Because Indian and Tibetan wolves show low genetic diversity, we explored how

another component of genetic diversity – functional genetic diversity – compares to

other wolves across Asia. We were specifically interested in quantifying patterns of

deleterious variation that is expected to have a negative impact on fitness. Using SnpEff

to estimate functional impact of mutations, we quantified the total genetic load (total

number of derived alleles classified as high or moderate impact), realized load

(homozygous state of derived alleles in high or moderate impact), and masked load

(heterozygous state of derived alleles in high or moderate impact) for five selected wolf

populations with assuming the derived alleles was the deleterious one (Cingolani et al.

2012, Bertorelle et al. 2022). Wolves from historically large population sizes – central

and southwestern Asia – had higher masked and total genetic load than wolves from

historically smaller populations (Indian, Tibetan, Mexican wolves) (**Figure 5A,B, S22,**

S23). This is consistent with theoretical predictions that larger populations with higher

genetic diversity should also possess more deleterious alleles in a heterozygous state

(Mathur and DeWoody 2021). In contrast, we found that Indian, Tibetan, and Mexican

wolves have a higher realized load and a lower total genetic load (**Figure 5,C**) than the

other surveyed populations. A higher realized load in these small populations is

expected, because genetic drift and inbreeding will result in an increased proportion of

homozygous genotypes, including deleterious ones.

A lower total genetic load in the Indian and Tibetan wolves is intriguing, as this may

reflect a loss of derived variants due to stronger genetic drift in historically small

populations and/or purging (Dussex et al. 2023). Purging predominantly occurs when

recessive deleterious variation is exposed to purifying selection in a homozygous state.

To look for explore evidence of stronger genetic drift versus purging, we asked whether

deleterious alleles are lost at a higher frequency than the background neutral alleles

(mutations in non-coding regions) for each individual wolf. In other words, does the fact

that derived alleles are categorized as deleterious result in more frequent loss compare

to neutral derived alleles within an individual, as expected for purging via purifying

selection? Or are deleterious derived alleles lost at the same frequency as neutral

derived alleles, suggesting the loss in deleterious derived alleles found in our total

genetic load results could be due to genetic drift? We assessed the prevalence of sites

associated with four impact categories: high impact (loss of function, LOF), moderate

impact (missense), low impact (synonymous), and modifier (non-coding; here assumed

to be selectively neutral). To account for differences in total derived alleles found in wolf

individuals, we divided the total number of LOF, missense and synonymous derived

mutations by the total number of neutral derived alleles assumed to be under neutral

processes (mutations in non-coding regions) for each individual.

443
We found Indian, Tibetan, and Mexican wolves have proportionally fewer missense and

synonymous mutations per 100,000 derived neutral mutations compared to wolves in

central and southwest Asia. A reduced frequency of synonymous mutations may reflect

stronger genetic drift associated with a smaller long-term population size rather than

purifying selection via purging, as synonymous mutations are assumed to be nearly

neutral (**Figure 5F,G**). We emphasize that it is difficult to differentiate evidence of

genetic drift and purging, and further work is needed to verify the functional impacts of

our missense and synonymous mutations. For high impact mutations, we observe no

clear pattern among individuals (**Figure 5E**). One reason for this could be incorrectly

annotated LOF mutations in the short-read assembled dog reference, as suggested by

Smeds et al. (2023). In addition, among the ones that may be correct, there are so few

LOF sites across the genome (n=145 on average in our dataset) and being such a small

number, their frequency may be more strongly influenced by genetic drift, as seen with

killer whales (Kardos et al. 2023). Overall, we suggest caution when interpreting our

genetic load results. Further work is needed to assess the accuracy of annotations from

a short-read assembled genome, functional impacts of mutations, and to increase the

number of individuals included in each population.

Discussion

In this study we conducted a comprehensive whole genome analysis on population

structure, ancestral composition, and genetic diversity of wolves in Asia. We find that

the strongest barriers to gene flow in Eurasia coincides with the Indian, Tibetan, and

Holarctic lineage distributions. We show that Holarctic wolf populations found in

southern regions of Asia possess genetic variation ancestral to wolves in Northern Asia

and that it is similar in the degree of its antiquity to that of the Tibetan and Indian

lineage. Despite evidence of recurrent gene flow across northern Eurasia during the late

Pleistocene, it appears this process of homogenization did not completely reach areas

in southern regions of Asia (Bergstrom et al. 2022). Lastly, for wolves in Asia we find

genetic diversity largely corresponds to historical processes, such as long-term isolation

within glacial refugia, contact zones, and long-standing environmental differences,

followed by anthropogenic processes.

Asia is the biogeographic cradle for gray wolf evolution, so it is not too surprising that

Asia harbors ancestral diversity in its southern regions. Previous work has also shown

that southern regions contain deeper local ancestry than both modern and some

Pleistocene wolf genomes from Northern Eurasia. Wolves from southwestern Asia,

eastern parts of China, and Japan contain ancestry that is more ancestral than Siberian

wolves living during the Pleistocene from 12 kya and even 100 kya (Bergstrom et al.

2022). In addition, a single wolf from Shanxi of eastern China had significant allele

sharing with ancient Siberian wolves from 50kya, 48kya, and 33kya (Ramos-Madrigal

2021). Recent work has emphasized the importance of carefully interpreting

Introgression analyses and to consider the possibility of "ghost lineages", that is extinct

or unsampled lineages (Tricou et al. 2022). For eastern Asian wolves, we detected

significant introgression with the Tibetan lineage as P3, but not with the older diverged

Indian lineage or African wolf as P3. While this suggests the origin of this partial

ancestry within eastern Asian wolves is similar to modern-day Tibetan lineage, the

degree to which the partial ancestry is similar to the Tibetan lineage remains unclear.

Our phylogenetic trees provide some clarity. None of our trees nor most frequent

topologies using Twisst show eastern Asian wolves clustering within or sister to the

Tibetan lineage. Instead, low recombination regions on the X chromosome show they

form their own lineage that diverges before the rest of the Holarctic wolves, yet after the

Tibetan lineage. Thus, it is possible that wolves in southern China may contain ancestry

of an unsampled wolf lineage that was present in southern China and was distinct from

the Tibetan lineage.

Consistent with this, previous work shows historical genomes of wolves in Southern

China and Tibetan wolves occupy opposite ends of the PC2 axis, suggesting distinct

partial ancestries (Wang et al. 2019). Fossils of wolves have been present across most

regions of China over the last 100,000 years, including 12,000 year old remains from

the Southern province of Jiangxi (Wang 2016). During this time, much of southern

China was drier with mixed broadleaf forests occupying where monsoon rain forests are

now (Pinxian and Xiangjun 1994). Thus, it is plausible that ancestry that was distinct

from the Tibetan lineage could have existed within a glacial refugium in Southern China,

and has not been completely replaced by expansion waves from northern Eurasian

wolves. An interesting future avenue of research is whether partial ancestry of extinct

Honshu wolves may also be from this glacial refugia, and thus share a common origin

with wolves of southern China. Incorporating ancient genomes from eastern China and

the newly sequenced 35 kya Japanese wolf genome in future analyses would be

needed to place this ancestry into a spatiotemporal context during the Pleistocene.

514
An unexpected result in our study was uncovering population-wide ancestral diversity

present across southwestern Asia. Our analyses point towards the widespread

ancestral component is likely related to the Indian lineage, rather than from the African

wolf. This fits well with previous work suggesting part of the deep ancestry within

southwestern Asia is at least 100,000 years old, which is before the estimated

divergence time of the Indian lineage (Bergstrom et al. 2022, Wang et al. 2022). When

and how the present-day population structure formed in southwestern Asia remains

unclear, but our study offers some insights. Combined with the fact there have been no

Indian mitogenomes detected outside of the Indian subcontinent, our demographic

history results from fastsimcoal2 and PSMC finds no strong evidence that the Indian-like

ancestry within southwestern Asia is due to recent (pre-6kya) gene flow from the Indian

lineage. In addition, it appears that the X chromosome, including low recombination

(<0.2 cM/Mb) regions, of southwestern Asian wolves are more similar to the Holarctic

lineage. Along with our PSMC results, the major genomic background of southwestern

Asian wolves does seem to be that of the broader Holarctic lineage, possibly tracing

back to gene flow from northern latitudes. At the same time, we find the Indian-like

genomic regions across the autosomes that do not fit into the modern-day diversity of

the Indian lineage. These could be remnants of a once more diverse Indian lineage, or,

despite our efforts, these genomic regions could contain some Holarctic ancestry that

places southwestern Asian wolves nearby but not within Indian wolves. A more

thorough analysis that uses multiple local ancestry approaches to classify Indian-like

ancestry would be insightful, as the length distribution and its phylogenetic relationships

can be more thoroughly assessed. More broadly, ancient remains of wolves have been

documented in southwest Asia throughout the late Pleistocene (Kurten 1965, Dayan et

al. 1992, Mashkour et al. 2008, Wang et al. 2016). Ancient genomes from these wolf

remains in southwestern Asia can directly test hypotheses on whether there were shifts

in ancestral composition over the late Pleistocene and if the Indian lineage was once

more widespread and genetically diverse.

543
544

Together, our work uses a combination of methodological approaches to better resolve

the historical and contemporary processes that shape genetic variation within a wide

ranging species. For many species during the late Pleistocene, their populations

underwent distinct evolutionary trajectories of isolation and expansion in response to

shifting climates. The process in which populations don't or do come together, and how

often, is a key force in speciation and evolution. Our study's findings of divergent

ancestry persisting in southern regions may be common pattern in widespread species,

where red foxes (*Vulpes vulpes*), Eurasian otters (*Lutra lutra*), and brown bears (*Ursus*

arctos) all show evidence of cryptic divergent ancestry in southern regions of Asia

(Statham et al. 2014, Jong et al. 2023, du Plessis et al. 2023). While our study

addresses gaps in our knowledge on wolves, many questions remain to be answered.

For example, why some wolf populations seem to have been more susceptible to gene

flow from northern populations (southwestern Asia, eastern Asia) during the late

Pleistocene, while others show little gene flow and prominent contact zones (Indian and

Tibetan) is an open question. With so many ancient and modern genomes now

available, wolves are poised to serve as a valuable model to study how intra-population

gene flow, interspecies hybridization, and isolation during the late Pleistocene has

contributed to present-day species' diversity.

563
564 **Conservation Implications**
565

Our study has several implications for the conservation of various wolf populations in

Asia. For taxonomy, our study supports previous calls that a taxonomic revision is

necessary for wolves in Asia (Alvares et al. 2019, Werhahn et al. 2022, Krofel et al.

2022). While wolves from India to Türkiye are collectively described as *Canis lupus*

pallipes, our study emphasizes past work that the ancestral Indian lineage is confined to

India and parts of Pakistan (Hennelly et al. 2021, Hennelly et al. 2023). For the wolves

in southwestern Asia that are considered "*Canis lupus pallipes*", we find this distinct

population may be geographically limited by the Bosphorus strait in Türkiye and the

Caucasus mountain range (Figure 1AB, Figure 2A). Further work is necessary to

understand the population genetic structure in the arid areas of southern Central Asia,

such as the Karakum desert in Turkmenistan and Kyzylkum desert in Uzbekistan. We

suggest the morphological and genomic distinctiveness of wolves in southwest Asia

warrants subspecies status, in line with recent recommendations (Werhahn et al. 2022).

For Indian and Tibetan wolves, the presence of gene flow barriers and likely

reproductive barriers coinciding with lineage boundaries further supports the recognition

of subspecies or species status (Werhahn et al. 2022).

582
Our study underscores the importance of robustly describing wolves in parts of China.

Wolves have been documented as South as Guangxi in China, a province that borders

Vietnam (Wang et al. 2016, **Figure S1**). In 1936, wolves were reported to exist in parts

of Central and South China, but were already less common compared to Northern

China (Sowerby 1936). A report in 2003 concluded wolves have largely been extirpated

from regions of Guangxi that once occurred there in the 1990s, but they were possibly

still present in Nanling National Nature Reserve (Fellowes et al. 2003). In response to

their decline in the country, the wolf was recently added in 2021 as a protected species

under Class II of the List of Wildlife under Special State Protection of China. To the

authors' knowledge, we are not aware of published estimates of the population size and

status, or a morphological description, of wolves in Southern parts of China. Given their

evolutionary distinctiveness, our work highlights the need for more research on their

distribution, genetics, morphology, and population status to conserve what populations

remain.

597
Overall, our study emphasizes that southern regions of Asia hold most of the ancestral

genetic diversity within gray wolves, and are also the most endangered. The Indian

lineage numbers at ~2,700 individuals in India and the Tibetan lineage only ~3,000

individuals (Jhala et al. 2022, Werhahn et al. 2023). There are less than ~750 wolves

left on the Arabian peninsula, a genomically unique but understudied population where

only one genome has been sequenced (Bonsen et al. 2024). It is possible that wolves in

Southern China are already extinct or very close to it. Wolves in these regions are

impacted by numerous human-induced threats, especially for the remaining populations

in Southern China and India. Our study highlights the need to prioritize these

populations for conservation to preserve the full spectrum of extant wolf diversity.

609
610 **Methods**

611
This study includes 20 new whole genomes from wolves from Pakistan (n=8), Israel

(n=1), Lebanon (n=1), Afghanistan (n=1), Türkiye (n=1), Ukraine (n=1), Slovakia (n=1),

Kazakhstan (n=3), and Russia (n=3). For all wolf samples from Pakistan and Israel, we

extracted DNA from these samples using DNAeasy Blood and Tissue Kit (Qiagen)

following the manufacturer's protocol. For the rest of the wolf samples, they were

extracted using Thermo Scientific KingFisher instrument following manufacturer's

protocol. The Indian wolf from Maharashtra originated from a previous study and was

resequenced to a deeper depth following protocols in Hennelly et al. 2021. For the gray

wolves from Pakistan and Israel, libraries were sequenced on an Illumina NovaSeq

6000 S4 flowcell and paired end at 150-bp (Illumina). For the rest of the newly

sequenced wolf samples, they were sent to BGI Copenhagen for library build and

sequenced on ⅛ lane each on DNBSEQ at paired end 150. We constructed a dataset

comprising an additional 105 previous published canid genomes, resulting in a total of

118 canid genomes in the study (**Table S1**).

Alignment, Variant Calling, and Filtering

We used the Paleomix v.1.2.13.2. pipeline to align our raw reads to the dog reference

genome Canfam3.1 (Schubert et al. 2014). Specifically, we used the BWA mem

algorithm and used the GATK indel realigner to generate an indel-realigned BAM file for

each sample. Variant calling was performed using the Genome Analysis Toolkit (GATK)

version 4.2.5.0 (McKenna et al. 2010). First, we used GATK Haplotype Caller to perform

variant calling on each sample, then used the resulting GVCF files to perform joint

genotyping with all canid individuals combined using GenomicsDBImport and

GenotypeGVCF in GATK. We obtained a set of high quality single nucleotide

polymorphisms (SNPs) using the following filtering steps in GATK: QD < 2.0 || SOR >

3.0 || FS > 60.0 || MQ < 40.0 || MQRankSum < -12.5 || ReadPosRankSum < -8.0. Sites

with a mean depth of > 1800 for all individuals were removed to exclude paralogues

from our dataset. Lastly, we removed indels from our dataset using SelectVariants in

GATK (flag -select-type SNP). We excluded genomes that were derived from museum

samples in the genotype calling, which was only the wolf from Afghanistan.

Principal Component Analysis and Individual Admixture Proportions

To assess genome-wide genetic structure of the autosomes, we conducted a principal

component analysis (PCA) using PCangsd (Meisner and Albrecthtsen 2018) with our

total set of Eurasian gray wolf genomes. We inferred the PCA using genotype

likelihoods from ANGSD, in which we used the SAMtools model (-GL 1) and filtered the

dataset by including only properly paired reads (-only_proper_paired 1) from autosomal

bam files, excluded reads with excess of mismatches (-C 50) or mapping quality lower

than 20 (-minMapQ20), removed transitions (-noTrans 1), and keep bases with a

sequence quality above 20 (-minQ 20) (Korneliussen et al. 2014). We also retained

SNPs that were present in at least 90% of individuals present (>90 out of xx total; -

minInd 90). We obtained the covariance matrix from PCAngsd and used the R function

eigen to infer the PCA.

To estimate individual ancestry proportions, we inferred genotype likelihoods using

ANGSD and ran NGSadmix (Skotte et al. 2013) using K=2 to K-9 genetic clusters. We

used the canids within the genus *Canis* in this analysis, which included an additional 3

North American gray wolves, 3 golden jackals, 2 coyotes, 4 African wolves, 1 Ethiopian

wolf, 5 dogs (**Table S1**). We used the same criteria in ANGSD as to infer the PCA,

however, we retained SNPs that were present in at least 90% of individuals. This

resulted in 578,343 SNPs after filtering.

Assess barriers to gene flow for wolves across Eurasia

We inferred the estimated effective migration surface (EEMS) to assess spatial variation

in rates of gene flow among gray wolf populations in Eurasia (FEEMS; Marcus et al.

2012). The FEEMS approach utilizes a stepping-stone model to calculate genetic

dissimilarities between individuals based on spatial and genetic data. These genetic

dissimilarities can be visualized to illustrate departures from strict isolation by distance,

thereby detecting regions of high (i.e. corridors) and low (i.e. barriers) to gene flow. For

the FEEMS, we only used our dataset of 95 Eurasian wolves, where we filtered the

dataset in which we kept sits with a minimum allele count of 3 (--mac 3), all of

individuals represented at a site (--geno 0), and pruned sites in high linkage

disequilibrium by removing each SNP with a r_2 value of greater than 0.5 (--indep-

pairwise 50 10 0.5). This resulted in ~8.7 million autosomal SNPs across our Eurasian

wolves. For gathering the geographic data for each wolf sample, we had some gray wolf

samples that only had location information as the country or province within a country.

In these cases, we took the center coordinate within that country or province.

To run FEEMs, we first constructed a dense spatial grid using an edge width of 5 that

covered the entire Eurasian continent. We initially performed a random initialization over

the map to fix the estimate of the residual variance. We selected the tuning parameter,

lambda, using leave-one-out cross-validation using 20 folds where the lambda with the

lowest cross-validation was selected. Our cross-validation test found a lambda value of

100 to be the optimal value of lambda (**Figure S24**).

Phylogenomic relationships

We constructed the autosomal and X chromosome phylogeny that included all wild

canid species except dogs and the Ethiopian wolf (**Figure 2B, S5, S6**). We filtered the

autosomal and X chromosome VCF to exclude indels, keep sites with at least minimum

quality of 30 (--minQ30), keep only biallelic sites, and kept sites that had at least 90%

individuals called (-max-missing 0.9). We then converted each vcf file into the phylip

format using the script vcf2phylip (Ortiz et al. 2019). We randomly selected 500 and

1,000 regions with a length of 20 kb across the autosomes and X chromosome,

respectively. We inferred a maximum-likelihood phylogenetic tree for each 20kb region

using IQ-Tree 1.6.12 where we estimated the best model using ModelFinder and used

1,000 ultra-fast bootstraps to infer each tree (Kalyaanamoorthy et al. 2017, Nguyen et

al. 2014). The inferred phylogenetic trees were used as input into ASTRAL 5.7.8, which

inferred a species tree from a set of gene trees while taking into account gene tree

discordance (Zhang et al. 2018).

In addition, we inferred phylogenetic trees of the X chromosome and autosomes using

only the low recombination regions across the genome. For the X chromosome, we

inferred five different runs containing different sets of canid individuals. These runs were

of: (1) including all wild canid species except dogs and the Ethiopian wolf, (2) all wild

canid species and wolf individuals excluding dogs, the Ethiopian wolf, wolves from

Pakistan, and wolves from Ladakh (3) all wild canid species, and wolf individuals

excluding dogs, the Ethiopian wolf, wolves from Pakistan and Ladakh, Indian wolves,

(4) all wild canid species and wolf individuals excluding dogs, the Ethiopian wolf, wolves

from Pakistan, and wolves from southwestern Asia, and (5) all wild canid species and

wolf individuals excluding dogs, the Ethiopian wolf, wolves from Pakistan, and two

wolves from that clustered with Indian wolves (wolf from Zanjan and Hamadan in Iran).

All VCF files were filtered for sites with a minimum quality of 30 (-minQ 30), only biallelic

sites, and at least 90% individuals present at each site (--max-missing 0.9). We

downloaded the recombination map of the domestic dog from Auton et al. 2013 and

used the recombination rate (cM/Mb) in our analyses. We averaged the recombination

rates within 100-SNP windows across the autosomes and X chromosome, as generated

in a previous study (Hennelly et al. 2021). We selected genomic regions that had an

average recombination rate of below 0.2 cM/Mb, and used these positions to create a

bed file, which we then extracted these genomic regions from our VCF.

We inferred low recombination phylogenies for each of the five runs with different

samples with IQ-Tree 1.6.12 (**Figure S6, Figure S16, Figure S17A,B,C**). We converted

the VCF containing all low recombination regions to a phylip file format using

vcf2phylip.py (https://github.com/edgardomortiz/vcf2phylip). We then ran IQ-Tree 1.6.12

with estimating the best model using ModelFinder and used 1,000 ultra-fast bootstraps

to infer each tree (Kalyaanamoorthy et al. 2017, Nguyen et al. 2014). Second, we

inferred low recombination autosomal and X chromosome phylogenies using ASTRAL

5.7.8 (**Figure 2C, Figure S7, Figure S8**). We split each SNP dataset into 10 kb regions,

and converted each of these regions to a phylip file (Zhang et al. 2018). We then

inferred a tree for each 10 kb region using IQ-Tree 1.6.12 with estimating the best

model using ModelFinder and used 1,000 ultra-fast bootstraps.

Assessing genome-wide gene admixture

To test for gene flow among various canid lineages and species, we performed D-

statistic analyses using AdmixTools (Patterson et al. 2012). We filtered our dataset to

exclude missing data (--max-missing 1), kept biallelic sites, and a minimum quality filter

of 30 (--minQ30), along with the filters we applied using GATK. We converted plink files

into eigenstrat format using the convertf script within ADMIXTOOLS. We calculated the

D-statistic and Z-score for modern populations for three different topologies in the

format (((P1, P2),P3),P4): (((Norwegian wolf MW005, X wolf); Indian wolf BH123);

Andean fox), (((Norwegian wolf MW005, X wolf); Tibetan wolf TI32; Andean fox),

(((Norwegian wolf MW005, X wolf); African wolf); Andean fox). The D-statistic value will

be negative for an excess of shared derived alleles between P2 and P3, and/or P1 and

P4. Alternatively, the D-statistic will be positive for an excess of shared derived variants

between P2 and P4, or P3 and P1. For African wolves, we used multiple individuals as

a population for P3, which consisted of all four of our African wolf samples from Kenya,

Ethiopia, Morocco, and Algeria.

748
We also quantified the frequency of different topologies in genomic windows across the

genome using Twisst (Martin and Van Belleghem 2017). To prepare the dataset for the

Twisst analysis, we first phased the filtered dataset that contained no indels, biallelic

sites with a minimum quality of 30 and that were present in at least 90% of individuals

using Shapeit v2.r904 (Delaneau et al. 2013). We used the dog genetic map from Auton

et al. 2013 as input and a window size of 0.5 Mb. We converted the phased VCF to a

geno file using the parseVCF.py script within

https://github.com/simonhmartin/genomics_general. We then estimated phylogenies in

100-site windows using phyml with a General Time Reversible (GTR) substitution model

with phyml_sliding_windows.py within

https://github.com/simonhmartin/genomics_general. We summarized the relative

prevalence of topologies using the twisst.py script. The set of topologies we

summarized consisted of four population groups with Andean fox as an outgroup

focused on wolves in Central Asia and Eastern Asia, and five population groups with

Andean fox as an outgroup focused on southwestern Asian wolves. These resulting in

15 possible topologies for the four population groups, and 105 possible topologies using

five population groups. We also quantified the frequency in which topologies occurred in

windows of low (<0.2cM/Mb) recombination rate regions. To do this, we partitioned our

our averaged recombination rates within 100-SNP windows inferred in Hennelly et al.

2021 across the autosomes and X chromosome datasets.

769
Finally, we inferred phylogenetic trees using genomic regions within wolves of

southwestern Asia that were designated to be Indian ancestry from our twisst results.

First, we normalized the Twisst results for all 105 topologies and calculated the mean

for each topology. The topology with the highest mean of 0.33 was topology 73 where

Indian and southwestern Asian wolves form sister clades as the most well-resolved

topology across the autosomes (**Table S3**). We extracted genomic regions that had a

topology weight of over 0.2 and 0.3 for topology 73 from a VCF containing all autosomal

SNPs using bedtools. This resulted in 299,560 SNPs and 40,887 SNPs found within

genomic regions that had over 0.2 and 0.3 topology weight for topology 73, respectively.

We converted two VCF datasets, one with genomic regions that had over 0.2 and 0.3

topology weight for topology 73, to a phylip file using vcf2phylip.py

(https://github.com/edgardomortiz/vcf2phylip). We then ran IQ-Tree 1.6.12 with

estimating the best model using ModelFinder and used 1,000 ultra-fast bootstraps

(Kalyaanamoorthy et al. 2017, Nguyen et al. 2014) (**Figure 3D,E, Figure S18, Figure**

S19).

Demographic Modeling with fastsimcoal2

In order to better understand the origin of southwestern Asian wolves, we compared

three alternative demographic models and inferred demographic parameters under the

FASTSIMCOAL2 (v2.7 Excoffier et al 2021), a composite-likelihood method that

estimates historical introgression, divergence times and population topologies from the

site frequency spectrum using coalescent simulations.

We compared three demographic models (**Figure S15**) to better assess the

demographic history of southwestern Asian wolves. The included three populations

representing India, southwestern Asia and central Asia samples, and were defined as

(1) a divergent model with a strictly bifurcating topology separating Indian and Holarctic

Asian lineages and latter southwestern Asia from central Asia populations, (2) an

introgression model with the same strictly bifurcating topology followed by an

introgression event from the Indian to the southwestern Asian population (3) an

hybridization model, where southwestern Asian population was originated by

hybridization between Indian and central Asian populations. Input files including

template and parameter estimation files for the three models, are available at **Figure**

S27. A summary of all defined parameters and their search ranges are given in **Table**

S4.

We performed genotype calling using BCFtools specifically keeping monomorphic sites

(Danecek et al. 2021). We filtered our VCF containing all variant and monomorphic sites

by the following parameters, keeping only those loci with >10x coverage, and > 50%

missing data and removing sites SNPs violating Hardy-Weinberg equilibrium for

heterozygous excess (p < 0.01). After filtering, we retained 2,771,059 SNPs and

456,437 monomorphic sites, which we used to generate the joint 3D-SFS using the

(folded) minor allele frequency spectrum. We used the canFam 3.1 domestic dog

assembly as the reference, resulting in the reference being more closely related to

Holarctic wolves than to Indian wolves. Thus, we could not determine the ancestral

state of each allele and therefore used the minor allele frequency spectrum (folded

SFS). The mutation rate was set to 4.5×10^{-9} based on de novo mutations in a pedigree

of wolves (Koch et al. 2019). To generate the 3D-SFS while ensuring no missing data

and accounting for local linkage disequilibrium (LD) patterns, we down-sampled

individuals. Given an estimated recombination rate of 1.34cM/Mb using the

recombination map of the dog genome (Auton et al. 2013), we partitioned the genome

into independent and unlinked blocks of 2Mb, ensuring that sites within blocks were

linked, while those between blocks were unlinked. Within each block obtained, we

retained only SNPs without missing data by sampling three, six, and nine individuals

from the Indian, southwest, and central populations, respectively, ensuring that each

sampled individual had no missing data for that block. FASTSIMCOAL2 estimates a

composite likelihood, assuming independence between sites (i.e., it does not account

for linkage). Consequently, although the maximized likelihood converges to the correct

parameter values, the estimated likelihood approximates the true value when derived

from a set of unlinked loci (Excoffier et al., 2013). We utilized the parameter estimates

inferred from all retained SNPs to recalculate the likelihoods for each model using only

one SNP per block. This recalculation was based on a 3D-SFS derived from a set of

potentially independent SNPs, with one SNP selected per block. We used this newly

computed likelihood to apply the Akaike Information Criterion (AIC) to compare the

three models and computed the relative likelihood of models. For this we calculated

delta AIC according to Excoffier et al (2013), which represents the difference between

the AIC value of each model and the minimum AIC value among all models. Further we

thoroughly examined the parameter values that optimized the likelihood for each model.

Specifically, we compared divergence times between Holarctic and Indian lineages, the

effective population sizes of each population, and the admixture proportions estimated

under each model. This analysis aimed to determine whether the parameter estimates

that optimized each model were consistent with the most likely model as indicated by

the AIC. In addition, to evaluate whether the best-selected model could accurately

reproduce the observed data, we conducted a visual examination comparing the

expected and observed marginal one-dimensional SFS, as well as various two-

dimensional SFS pairs.

Long-term demographic history with PSMC

To infer the historical demography of wolves in Eurasia, we used the Pairwise

Sequential Markovian Coalescent (PSMC; Li and Durbin 2011). PSMC uses a

coalescent approach to estimate the history of change in effective population sizes over

time. We only included the autosomal sequences of each gray wolf individual. We

converted each bam file to a fasta-like consensus sequence by first using the mpileup

command with SAMtools and subsequently using BCFtools view –c to call variants and

vcfultils.pl vcf2fq to convert the vcf file to fastq format (Danecek et al. 2021). We

excluded any reads that were less than 20 for minimum mapping quality and minimum

base quality (-q 20 -Q 20) and excluded reads with excessive mismatches (-C 50). We

also removed sites with more than double or less than a third of the average depth of

coverage for each sample. We tested different combinations of parameters to infer the

PSMC, which were "`psmc -N25 -t15 -r5 -p 4+25*2+4+6`", "`psmc -N25 -t15`

`-r5 -p 2+2+25*2+4+6`" and "`psmc -N25 -t15 -r5 -p 1+1+1+1+25*2+4+6`"

following previous studies on gray wolves and recent recommendations for inferring

PSMC trajectories (Hilgers et al. 2024). Some samples showed a false peak when using

the parameter "`psmc -N25 -t15 -r5 -p 4+25*2+4+6`", therefore, we used the

parameter "`psmc -N25 -t15 -r5 -p 1+1+1+1+25*2+4+6`" for our final analyses

(**Figure S25**).

864
To account for our selected low coverage genomes (15-20x), we estimated the false

negative rate (FNR) by a downsampling high coverage gray wolf genome that were in

the same geographic region to the specific depth of the low coverage genome. To

determine the best FNR, we visually estimated the best correspondence between the

PSMC plots with the high coverage (>20x) regional wolf genome and downsampled

genomes with various FNR corrections (**Figure S26**). We then applied the best

estimated FNR to the low coverage gray wolf genomes to infer their demographic

history. We used a mutation rate of 4.5 x 10^{-9} (Koch et al. 2019) and a generation time

of 4.4 years (Mech and Barber-Meyer 2017).

874
Genetic Diversity and Runs of Homozygosity

We calculated the genome-wide heterozygosity for each wolf individual above 5x using

ANGSD (Korneliussen et al. 2014). We first estimated the folded site allele frequency

with the realSFS program in ANGSD (-doSaf) using genotype likelihoods with the

SAMtools model (-GL 1) and using the Andean fox genome to determine ancestral

state. We filtered each individual bam file to keep only autosomal chromosomes, only

properly paired reads (-only_proper_paired 1), excluded reads with excess of

mismatches (-C 50), excluded reads with a mapping quality (-minMapQ 20), excluded

reads with a depth less than 4 (-setMinDepth 4), and kept bases with a sequence

quality above 20 (-minQ 20). To calculate the genome-wide heterozygosity, we divided

the number of heterozygous sites by the total number of sites for each sample.

886
We inferred runs of homozygosity (ROH) using BCFtools/RoH (Narasimhan et al. 2016).

Along with the GATK filtering criteria, we included only biallelic SNPs that had all

individuals present per site (--max-missing 1, --min-alleles 2, --max-alleles 2). We also

included SNPs that had a depth of more than one third and less than double of the

average depth of coverage for each sample, where we chose samples that were >15x.

For the BCFtools/RoH analysis, we fixed the alternative allele frequency to 0.4 (-AFdflt

0.4) and used the dog recombination map from Auton et al. 201x to account for

recombination hotspots (--genetic-map). We kept ROHs that had a quality score of at

least 80 and excluded ROHs that were below 100 kb in length in our analysis.

896
To gain insight into the timing that inbreeding occurred, we calculated the expected

generations since inbreeding using the length of the ROHs, generation time of 4.4 years

(Mech and Barber-Meyer 2017), and an average recombination rate of 1.34 cM/Mb

calculated as the average recombination rate across the autosomes from the canfam3.1

genome assembly (Auton et al. 2013). Specifically, we used the formula: $g=100(2rL_{ROH})$

where g=generation interval, r=recombination rate (cM/Mb), and L=Length of ROH (Mb)

(Thompson et al. 2013). For example, a ROH length of 5Mb will have an expected time

of coalescence of 7.46 generations ago, corresponding to inbreeding occurring around

32 years ago using a wolf generation time of 4.4 years. Likewise, we estimate that a

ROH length of <0.16 Mb corresponds to inbreeding occurring >1,026 years ago.

Inbreeding coefficient

We inferred the per-individual inbreeding coefficient (F_{ind}) using genotype likelihoods,

where the inbreeding coefficient is the probability of identity by descent. We used

ANGSD to infer genotype likelihoods and allele frequencies for samples above 4x

coverage with the following parameters: -GL 2 -checkBamHeaders 0 -trim 0 -C 50 -baq

1 -skipTriallelic 1 -minMapQ 20 -minQ 20 -minInd 88 -uniqueOnly 1 -remove-bads 1 -

noTrans 1 -doGlf 3 -doMajorMinor 1 -doMaf 1 -minMaf 0.0001 -SNP_pval 1e-6. We then

used NgsRelate v2 with option -F1 to infer the inbreeding coefficients for each Eurasian

wolf individual (Hanghoj et al. 2019).

Genetic Load

We selected 14 individuals with > 20x sequencing coverage from India (n=2), the

Tibetan plateau (n=2), southwestern Asia (n=4), Central Asia (n=4), and the Mexican

wolf (n=1). We selected these wolves to assess the genetic load of Indian and Tibetan

wolves, in which we compared Indian and Tibetan wolf genetic load with two adjacent,

genetically diverse populations, and to a Mexican wolf individual with previously

documented very low genetic diversity. We used SnpEff to annotate a VCF, including

only biallelic non-indel sites with no missing data, minimum depth of 10, maximum

depth of 60, a minimum allele count of 2 and a minimum sequence quality (--minQ) of

30.

To polarize our dataset, we followed the method by Smeds et al. (2022) to map two

outgroups to the dog reference genome at only sites with a minimum depth of 4 at both

outgroup and included nonvariant sites. We used Dhole (*Cuon alpinus*) and Andean fox

(*Lycalopex culpaeus*) as our outgroups. We then performed pseudo-haploidization of

the outgroup genomes and kept only sites in which the allele agreed in both outgroups

(Smeds et al. 2022). We used this to add the ancestral allele information to our

annotated VCF, and assumed that the derived allele was the deleterious allele (Smeds

et al. 2022).

We counted the number of derived alleles and whether they were in a homozygous or

heterozygous state for variants predicted by SnpEff to have high, moderate, and low

impact on protein function. High impact is predicted to cause loss of function or protein

truncation (stop gain, frameshift mutation); moderate impact is predicted to change

protein effectiveness (missense variant), and low impact is assumed harmless, such as

a synonymous variant. We also calculated the following metrics: total load (the total

number of derived alleles classified as high or moderate impact), realized load

(homozygous state of derived alleles in high or moderate impact), and masked load

(heterozygous state of derived alleles in high or moderate impact).

947
948

To investigate evidence of differential loss of derived variants for different types of

mutations, we assessed the relative amounts of mutational categories given by SnpEff.

These mutational categories are: Loss of Function (LOF) mutations that imply a high

predicted impact on protein structure, missense mutations (t moderate impact),

synonymous mutations (low impact), and "modifier" mutations found in non-coding

regions (no impact). We calculated the count of derived alleles and the number of

heterozygous and homozygous derived allele calls in each of the four mutational

categories for each wolf individual. To account for differences in overall derived allele

counts among individuals, we calculated the average derived allele count per 100,000

derived neutral alleles for total mutational load, LOF, missense, and synonymous

mutations. We considered the modifier category as neutral derived alleles. Specifically,

we calculated mutational load as: (total derived alleles in LOF and missense / total

derived alleles in modifier category) multiplied by 100,000.

References

964
965 Alvares F, Bogdanowicz W, Campbell LAD, Godinho R, Hatlauf J, Jhala YV, Kitchener
966 AC, Koepfli KP, Krofel M, Senn H, Sillero-Zubiri C, Viranta S, Werhahn G. 2019. Old
967 World Canis spp. With taxonomic ambiguity: workshop conclusions and
968 recommendations. Canid News 21
969
970 Auton A, Li YR, Kidd J, Oliveira K, Nadel J, Holloway JK, Boyko AR. 2013. Genetic
971 recombination is targeted towards gene promoter regions in dogs. Plos Genetics, 9,
972 e1003984.
973
974 Bergstrom A, Stanton DWG, Taron UH, Frantz L, Sinding MHS, Ersmark E, Pfrengle S,
975 Cassatt-Johnstone M, Lebrasseur O, Girdland-Flink L, Fernandes DM, Ollivier M, et al.
976 2022. Grey wolf genomic history reveals a dual ancestry of dogs. Nature 607: 313-320.
977
978 Bertorelle G, Raffini G, Bosse M, Bortoluzzi C, Iannucci A, Trucchi E, Morales ME,
979 Oosterhout C. 2022. Genetic load: genomic estimates and applications in non-model
980 animals. Nature Review Genetics 23: 492-503.

Bonsen GT, Wallach AD, Ben-Ami D, Keynan O, Khalilieh A, Dahdal Y, Ramp D. 2024. Navigating complex geopolitical landscapes: challenges in conserving the endangered Arabian wolf. Biological Conservation 296: 110655.

Danecek P, Bonfield JK, Liddle J, Marshall J, Ohan V, Pollard MO, Whitwham A, Keane T, McCarthy SA, RM Davies, H Li. 2021. Twelve years of SAMtools and BCFTools. GigaScience 10(2): giab008

Dayan, T, Simberloff, D, Tchernov E, Yom-Tov Y. 1992. Canine carnassial: character displacement in the wolves, jackals, and foxes of Israel. Biological Journal of the Linnean Society 45, 313–331

Delaneau O, Howie B, Cox AJ, Zagury JF, Marchini J. 2013. Haplotype estimation using sequencing reads. AJGH 93(4): P687-P696

Dussex N, Morales HE, Grossen C, Dalen L, Oosterhout C. 2023. Purging and accumulation of genetic load in conservation. Trends in Ecology and Evolution 38(10): P961-P969.

Exoffier L, N Marchi, DA Marques, R Matthey-Doret, A Gouy, VC Sousa. 2021. Fastsimcoal2: demographic inference under complex evolutionary scenarios. Bioinformatics 37(24): 4882-4885.

Fellowes JR, BPL Chan, CMN Lau, N Sai-Chit, GLP Siu. 2003. Report of rapid biodiversity assessment at Shiwandashan National Nature Reserve and National Forest Park, Southwest Guanxi, China, 2000-2001.

Feng C, Wang J, Liston A, Kang M. 2023. Recombination variation shapes phylogeny and introgression in wild diploid strawberries. Molecular Biology and Evolution 40:msad049.

Gomez-Sanchez D, Olalde I, Sastre N, Ensenat C, Carrasco R, Marques-Bonet T, Lalueza-Fox C, Leonard JA, Vila C, Ramirez O. 2018. On the path to extinction: inbreeding and admixture in a declining grey wolf population. Molecular Ecology 27:3599-3612.

Cingolani P, Platts A, Wang LL, Coon M, Nguyen T, Wang L, et al. A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff. Fly 6(2)

Gopalakrishnan S, Sinding MHS, Ramos-Madrigal J, Niemann J, Castruita JAS, Vieira FG, Caroe C, Montero MM, Kuderna L, et al. 2018. Interspecific gene flow shaped the evolution of the Genus *Canis*. Current Biology 28(21): P3441-3449.

1027　Green RE, Krause J, Briggs AW, Maricic T, Stenzel U, Kircher M, Patterson N, Li H,
1028　Zhai W, Fritz MH, Hansen NF. A draft sequence of the Neandertal genome. Science
1029　328:710-722.
1030
1031　Hanghoj K, Molke I, Andersen PA, Manica A, Korneliussen TS. 2019. Fast and accurate
1032　relatedness estimation from high-throughput sequencing data in the presence of
1033　inbreeding. GagaScience 1-9.
1034
1035　Harris AJ, Foley NM, Williams TL, Murphy WJ. 2022. Tree House Explorer: a novel
1036　genome browser for phylogenomics. Molecular Biology and Evolution 39(6):msac130
1037　Hennelly LM, Habib B, Modi S, Rueness EK, Gaubert P, Sacks BN. 2021. Ancient
1038　divergence of Indian and Tibetan wolves revealed by recombination-aware
1039　phylogenomics. Molecular Ecology 30:6687-6700.
1040
1041　Hennelly LM, Sarwar G, Fatima H, Werhahn G, Abbas FI, Khan AM, Mahmood T,
1042　Kachel S, Kubanychbekov Z, Waseem MT, et al. 2023. Genomic analysis of wolves
1043　from Pakistan clarifies boundaries among three divergent wolf lineages. 115:339-348.
1044
1045　Hilgers L, Liu S, Jensen A, Brown T, Cousins T, Schweiger R, Guschanski K, Hiller M.
1046　2024. Avoidable false PSMC population size peaks occur across numerous studies.
1047　BioRxiv.
1048
Jensen A, Swift F, de Vries D, Beck RMD, Kuderna LFK, Knauf S, Chuma IS, Keyyu
JD, Kitchener AC, Farh K, Rodgers J, Marques-Bonet T, Detwiler KM, Roos C,
Guschanski K. 2023. Complex evolutionary history with extensive ancestral gene flow in
an African primate radiation. Molecular Biology and Evolution 40(12): msad247
Jhala YV, Saini S, Kumar S, Qureshi Q. Distribution, status, and conservation of the
Indian Peninsula wolf. Front Ecol Evol. 2022:10:814966.
1055
1056　Jiang Z, Zang W, Ericson PGP, Song G, Wu S, Feng S, Drovetski SV, Liu G, Zhang D,
1057　Saitoh T, Alstrom P, Edwards SV, Lei F, Qu Y. 2024. Gene flow and an anomaly zone
1058　complicate phylogenomic inference in a rapidly radiated avian family (Prunellidae). BMC
1059　Biology 22:49
1060
1061　Jong MJ, Niamir A, Wolf M, Kitchener AC, Lecomte N, Serydokin IV, Fain SR, Hagen
1062　SB, Saarma Y, Janke A. 2023. Range-wide whole-genome resequencing of the brown
1063　bear reveals drivers of intraspecies divergence. Communications Biology 6: 153.
1064
1065　Kalyaanamoorthy S, Minh BQ, Wong TKF, Haeseler A, Jermiin LS. 2017. ModelFinder:
1066　fast model selection for accurate phylogenetic estimates. Nature Methods, 14, 587–589.
1067　Kardos M, Zhang Y, Parsons KM, Yunga A, Kang H, Xi X, Liu X, Matkin CO, Zhang P,
1068　Ward EJ, et al. 2023. Inbreeding depression explains killer whale population dynamics.
1069　Nature Ecology and Evolution 7: 675-686.
1070

1071 Koch EM, RM Schweizer, TM Schweizer, DR Stahler, DW Smith, RK Wayne, J
1072 Novembre. 2019. De novo mutation rate estimation in wolves of known pedigree.
1073 Molecular Biology and Evolution 36(11): 2536-2547.
1074
1075 Korneliussen TS, Albrechtsen A, Nielsen R. 2014. ANGSD: Analysis of next generation
1076 sequencing data. BMC Bioinformatics 15:356.
1077
1078 Krofel M, Hatlauf J, Bogdanowicz W, Cambell LAD, Godinho R. Towards resolving
1079 taxonomic lineages in wolf, dog, and jackal of Africa, Eurasia, and Australasia. J Zool.
1080 2022:316:155–168.
1081
1082 Kurten B. 1965. The carnivora of the Palestine caves. Acta Zooogical Fennica 07, 74.
1083 Lescroart J, Bonilla-Sanchez A, Napolitano C, Buitrago-Torres DL, Ramirez-Chaves
1084 HE, Pulido-Santacruz P, Murphy WJ, Svardal H, Eizirik E. 2023. Extensive
1085 phylogenomic discordance and the complex evolutionary history of the Neotropical cat
1086 Genus *Leopardus*. Molecular Biology and Evolution 40(12): msad255
1087
1088 Li G, Figueiro HV, Eizirik E, Murphy WJ. 2019. Recombination-aware phylogenomics
1089 reveals the structured genomic landscape of hybridizing cat species. Molecular Biology
1090 and Evolution 36(10): 2111-2126.
1091
1092 Li H, Durbin R. 2011. Inference of human population history from individual whole-
1093 genome sequences. Nature 475:493-496.
1094
1095 Marcus J, Ha W, Barber RF, Novembre J. 2021. Fast and flexible estimation of effective
1096 migration surfaces. eLife 10:e61927.
1097
1098 Martin S, Davey H, Salazar C, Jiggins CD. 2019. Recombination rate variation shapes
1099 barriers to introgression across butterfly genomes. PLoS Biology 17:e2006288.
1100 Martin SH, Belleghem SMV. 2017. Exploring evolutionary relationships across the
1101 genome using topology weighting. Genetics 1:429-438.
1102
1103 Mashkour M, Monochot H, Trinkaus E, Reyss JL, Biglari F, Bailon S, Abdi K. 2008.
1104 Carnivores and their prey in the Wezmeh Cave (Kermanshah, Iran): a Late Pleistocene
1105 refuge in the Zagros. International Journal of Osteoarchaeology 19, 678–694
1106
1107 Mathur S, DeWoody JA. 2021. Genetic load has potential in large populations but is
1108 realized in small inbred populations. Evolutionary Applications 14(6): 1540-1557.
1109
1110 McKenna A, Hanna M, Banks E, Sivachenko A, Cibulskis , Kernytsky A, Garimella K,
1111 Altshuler D, Gabriel S, Daly M, DePristo MA. 2010. The Genome Analysis Toolkit: a
1112 MapReduce framework for analyzing next-generation DNA sequencing data. Genome
1113 Research 20: 1297-1303.
1114

1115   Mech DL, Barber-Meyer S. 2017. Use of erroneous wolf generation time in
1116   assessments of domestic dog and human evolution Sci ELett. Available from:
1117   http://science.sciencemag.org/content/352/ 6290/1228/tab-e-letters
1118

1119   Meisner J, Albrechtsen A. 2018. Inferring population structure and admixture
1120   proportions in low-depth NGS data. Genetics 210:719-731.
1121

1122   Meisner J, Albrechtsen A. 2018. Inferring population structure and admixture
1123   proportions in low-depth NGS data. Genetics 2: 719-731.
1124

1125   Monthey JD, Spellman GM. 2024. Recombination rate variation shapes genomic
1126   variability of phylogeographic structure in a widespread North American songbird (Aves:
1127   Certhia americana). Molecular Phylogenetics and Evolution 196:108088.
1128

1129   Narasimhan V, Danecek P, Scally A, Xue Y, Tyler-Smith C, Durbin R. 2016.
1130   BCFtools/RoH: a hidden Markov model approach for detecting autozygosity from next-
1131   generation sequencing data. Bioinformatics 32:1749-2751.
1132

1133   Nguyen LT, Schmidt HA, Haeseler A, Minh BQ. 2014. IQTREE: a fast and effective
1134   stochastic algorithm for estimating maximum-likelihood phylogenies. Molecular Biology
1135   and Evolution, 32, 268–274.
1136

1137   Niemann J, Gopalakrishnan S, Yamaguchi N, Ramos-Madrigal J, Wales N, Gilbert
1138   MTP, Sinding MHS. 2020. Extended survival of Pleistocene Siberian wolves into the
1139   early 20th century on the island of Honshu. iScience 24: 101904.
1140

1141   Ortiz EM. 2019. Vcf2phylip v2.0: convert a VCF matrix into several matrix formats for
1142   phylogenetic analysis. DOI: 10.5281
1143

1144   Patterson N, Moorjani P, Luo Y, Mallick S, Rohland N, Zhan Y, Reich D. 2012. Ancient
1145   admixture in human history. Genetics, 192, 1065–1093.
1146

1147   Pinxian W, Xiangjun S. 1994. Last glacial maximum in China: comparison between land
1148   and sea. Catena 23: 341-535.
1149

1150   Plessis SJ, Blaxter M, Koepfli KP, Chadwick EA, Hailer F. 2023. Genomics reveals
1151   complex population history and unexpected diversity of Eurasian otters (*Lutra lutra*) in
1152   Britain relative to genetic methods. Molecular Biology and Evolution 40(11): msad207
1153

1154   Ramos-Madrigal J, Sinding MHS, Caroe C, Mak SST, Niemann J, Castruita JAS,
1155   Fedorov S, Kandyba A, Germonpre M, Bocherens H, et al. 2021. Genomes of
1156   Pleistocene Siberian wolves uncover multiple extinct wolf lienages. Current Biology
1157   31:198-206.
1158

1159   Schubert M, Ermini L, Sarkissian CD, Jonsson H, Ginolhac A, Schaefer R, Martin MD,
1160   Fernandez R, Kircher M, et al. 2014. Characterization of ancient and modern genomes

1161   by SNP detection and phylogenomic and metagenomic analysis using PALEOMIX.
1162   Nature Protocols 9: 1056-1082.
1163

1164   Schumer M, Xu C, Powell DL, Durvasula A, Skov L, Holland C, Przeworki M. 2018.
1165   Natural selection interacts with recombination to shape the evolution of hybrid genomes.
1166   Science 360:565-660.
1167

1168   Segawa T, Yonezawa T, Mori H, Kohno A, Kudo Y, Akiyoshi A, Wu J, Tokanai F,
1169   Sakamoto M, Kohno N, Nishihara H. 2022. Paleogenomics reveals independent and
1170   hybrid origins of two morphologically distinct wolf lineages endemic to Japan. Current
1171   Biology 32:2494-2504.
1172

1173   Skotte L, Korneliussen TS, Albrechtsen A. 2013. Estimating individual admixture
1174   proportions from Next Generation sequencing data. Genetics 3: 693-702.
1175

1176   Smeds L, Ellegren H. 2022. From high masked to high realized genetic load in inbred
1177   Scandinavian wolves. Molecular Ecology 32(7): 1567-1580.
1178

1179   Sowerby AC. 1936. China's natural history: a guide to the Shanghai Museum. Royal
1180   Asiatic Society of Great Britain and Ireland, North China Branch.
1181

1182   Statham MJ, Murdoch J, Janecka J, Aubry KB, Edwards CJ, Soulsbury CD, Berry O,
1183   Wang Z, Harrison D, Pearch M, et al. 2014. Range-wide multilocus phylogeography of
1184   the red fox reveals ancient continental divergence, minimal genomic exchange, and
1185   distinct demographic histories. Molecular Ecology 23(19): 4813-4830.
1186

1187   Stronen AV, Jedrzejewska B, Pertoldi C, Demontis D, Randi E, Niedzialkowska M, Pilot
1188   M, Sidorovich VE, et al. 2013. North-south differentiation and a region of high diversity
1189   in European wolves (*Canis lupus*). PLoS ONE 8(10): e76454
1190

1191   Taron UH, Salado I, Escobar-Rodriguez M, Westbury MV, Butschkau S, Paijmans JLA,
1192   vonHoldt BM, Hofreiter M, Leonard JA. 2021. A sliver of the past: the decimation of the
1193   genetic diversity of the Mexican wolf. Molecular Ecology 30:6340-6354.
1194

1195   Thompson EA. 2013. Variation in meiosis, across genomes, and in populations.
1196   Genetics 194:301-326.
1197

1198   Tricou T, Tannier E, Vienne DM. 2022. Ghost lineages highly influence the
1199   interpretation of introgression tests. Systematic Biology 71(5): 1147-1158.
1200

1201   vonHoldt B, Pollinger JP, Earl DA, Knowles JC, Boyko AR, Parker H, Geffen E, Pilot M,
1202   Jedrzejewski W, Jedrzejewska B, Sidorovich V, Greco C, et al. 2011. A genome-wide
1203   perspective on the evolutionary history of enigmatic wolf-like canids. Genome Research
1204   21(8): 1294-1305.
1205

vonHoldt BM, Brzeski KE, Aarema ML, Schell CJ, Rutledge LY, Fain SR, Shutt AC, Linderholm A, Murphy WJ. 2021. Persistence and expansion of cryptic endangered red wolf genomic ancestry along the American Gulf coast. Molecular Ecology 31:5440-5454.

Wang GD, Zhang M, Wang X, Yang MA, Cao P, Liu F, Lu H, Feng X, Skoglund P, Wang L, Fu Q, Zhang YP. 2019. Genomic approaches reveal an endemic subpopulation of gray wolves in Southern China. iScience 20:110-118.

Wang L, Ma YP, Zhou QJ, Zhang YP, Savolainen P, Wang GD. 2016. The geographical distribution of grey wolves (Canis lupus) in China: a systematic review. Zoological Research 37(6): 315-326.

Wang MS, Thakur M, Jhala Y, Wang S, Srinivas Y, Dai SS, Liu ZX, Chen HM, Green RE, Koepfli KP, Shapiro B. 2022. Genome sequencing of a gray wolf from Peninsular India provides new insights into the evolution and hybridization of gray wolves. Genome Biology and Evolution 2:evac012.

Wang MS, Wang S, Li Y, Jhala Y, Thakur M, Otecko NO, Si JF, Chen HM, Shapiro B, Nielsen R, YP Zhang, DD Wu. 2020. Ancient hybridization with an unknown population facilitated high-altitude adaptation of canids. Molecular Biology and Evolution 37:2616-2629.

Werhahn G, Hennelly LM, Lyngdoh S, Habib B, Viranta S, Shrotriya S. 2023. Himalayan wolf: Canis lupus spp chanco. The IUCN Red List of Threatened Species
Werhahn G, Senn H, Macdonald DW, Sillero-Zubiri C. The diversity of genus Canis challenges conservation biology: a review of available data on Asian wolves. Front Ecol Evol. 2022:10:10.3389.

Zhang C, Rabiee M, Sayarri E, Mirarab S. 2018. ASTRAL-III: polynomial time species tree reconstruction from partially resolved gene trees. BMC Bioinformatics 19:153

Acknowledgements

L.M.H thanks the National Science Foundation Postdoctoral Research Fellowship

(award number 2208950) for funding and support. The Norwegian Environment Agency

(project 18088069) provided funding and support for sequencing efforts on newly

sequenced wolf genomes. Ç.H.Ṣ. thanks Fondation Segré and the Sigrid Rausing Trust

for providing the majority of the funding for this project, H. Batubay Özkan and Barbara

Watkins for their support of the Biodiversity and Conservation Ecology Lab at the

University of Utah, and Bilge Bahar, Seha İşmen, Ömer Koç, Ömer Külahçıoğlu, Burak

Över, Emin Özgür, Suna Reyent and Ceren Sağlamer for supporting this project.

Türkiye's Department of Nature Conservation and National Parks and the Ministry of

Agriculture and Forestry granted the permit for Türkiye (No. 72784983-488.04-114100).

We thank the Museum of the Institute of Plant and Animal Ecology UB RAS access to

their collections.

1253

Author Contributions

1255
1256 L.M.H. conceived and designed the project with guidance from S.G., M.H.S.S., and

1257 B.N.S. C.S.O., M.H.S.S., N.F.G.M. conducted laboratory work, H.F., G.S., B.H., F.H.,

1258 S.P.E.H., C.K., Ç.H.Ş., P.K., H.S., M.H.S.S., M.T.P.G. provided logistics, field work, wolf

1259 samples, and sequencing efforts, L.M.H. led data analysis with assistance from B.R.P.,

1260 A.N., X.S., N.F.G.M, L.M.H. wrote the manuscript with input from all co-authors.

1261

Data Availability

1263
1264 All raw reads have been submitted to the NCBI Project ID XXXX, with accession

1265 numbers XXXX-XXXX. The bioinformatic scripts and results from the phylogenetic

1266 analyses, D statistics, genetic load, and PSMC can be found on the github:

1267 https://github.com/hennelly/Asia_wide_wolf_genomics.

1268

1269

Figure Captions

1272

Figure 1. Population genetic structure genetic connectivity of wolves across Eurasia. (A) Sample location and individual admixture proportions estimated using genotype likelihoods of ~300,000 autosomal SNPs at K=6 for 98 wolves across Eurasia. The individual admixture proportions were estimated using a set of 101 gray wolves, 5 dogs, and 12 genomes from various basal canid species. **(B)** Principal component analysis using genotype likelihoods of ~10 million autosomal SNPs for 98 wolves across Eurasia. Colors indicate geographic region of each the individual wolf belongs to. **(C)** The estimated effective migration surface (EEMS) using ~8.7 million autosomal SNPs for 97 wolves across Eurasia. Colors of the EEMS correspond to lower-than-average (red) and higher-than-average (blue) effective migration as expressed as $\log_{10}(w)$. A lambda of 100 was used after performing cross-validation analyses.

Figure 2. Tibetan-like ancestry in wolves from central Asia and eastern Asia. **(A)** D statistic results to estimate derived allele sharing between selected wolf individuals in Asia and the Tibetan lineage. The only populations that show significant allele sharing with Tibetan wolves are wolves from Pakistan, Kyrgyzstan, Mongolia, and eastern China. **(B)** Autosomal phylogeny using 98 wolves from Eurasia, 3 North American gray wolves and 12 genomes from other canid species. The tree was inferred using ASTRAL and estimated using 1,000 regions in 20kb length that were randomly chosen across the dog reference genome. Normalized quartet score of selected nodes are shown. Colored circles indicate the individuals assigned to the Tibetan lineage (orange), Central Asian mountains (high altitude Pakistan, Tajikistan, Kyrgyzstan), North American wolves (gray), and Eastern Asia (eastern China, Mongolia). **(C)** A tree inferred using ASTRAL with only low recombination (<0.2cM/Mb) regions of the X chromosome. A total of 513K SNPs were used, which we split into regions of 10kb length to assess gene tree discordance using ASTRAL. Normalized quartet score of selected nodes are shown. Eastern Asian wolves form a well-supported clade that is basal within gray wolves, except Tibetan wolves. **(D)** The three topologies with the highest average topology weightings across the X chromosome when considering wolves from the central Asian mountains or eastern Asia as the focal population (X). The topology shaded in gray depicts a scenario where the focal wolf population is ancestral to other Holarctic wolves from Europe and Central Russia. Prevalence of 15 possible topologies in high, medium and low recombination regions on the X chromosome to assess whether wolves from **(E)** central Asian mountains and **(F)** eastern Asia show evidence of ancestry that is ancestral to northern Holarctic wolves. For both central Asian and eastern Asian wolves, three topologies (solid lines) show a negative relationship between recombination rate and topology weighting, in contrast to the 12 other topology (dotted lines). **(F)** Eastern Asian wolves have more thoroughly sorted windows when they form a lineage ancestral to other northern Holarctic populations (topology 10) in low recombination regions compared to wolves from the mountains of central Asian mountains.

Figure 3. Persistence of Indian-like ancestry in wolves of southwest Asia. **(A)**
Declines in autosomal ancestry proportion at K=6 assigned to the Indian lineage
(purple), the Tibetan lineage (orange) and African wolf (*Canis lupaster*) (blue) for
selected wolves as geographic distance increases from the distributional extent of these
three taxa. Assigned Indian ancestry does not decline with geographic distance for
wolves in southwest Asia (purple triangles). **(B)** The model with the highest likelihood
inferred using fastsimcoal2 corresponding to an introgression event between the Indian
lineage and wolves of southwest Asia. **(C)** The topology with the highest average
topology weight when testing the relationship between 4 wolf populations and the
African wolf, resulting in 105 possible topologies inferred using 100-SNP windows with
Twisst. **(D)** A maximum likelihood phylogeny using autosomal genomic regions in which
the topology weight of topology where Indian and southwest Asian wolves are sister
(topology 73) was above 0.3 (~99.99th percentile; ~40kb length total). The tree was
inferred using IQ-tree where the best model was TVM+F+R5 through ModelFinder and
1000 bootstraps were used. Local posterior probability is labeled at selected nodes. **(E)**
A maximum likelihood phylogeny that excludes wolves from Pakistan and Ladakh,
which was inferred using autosomal genomic regions where the topology weight of
topology 73 was above 0.3 (~99.99th percentile; ~40kb length total).

Figure 4. Recent and long-term demographic history of wolves in Eurasia. (A)
Estimates of genome-wide heterozygosity inferred using ANGSD for 99 wolves from
Eurasia**. (B)** Runs of homozygosity (ROH) for 43 wolves showing the sum of the length
of ROH in four size ranges: 0.1-1Mb (Short ROH), 1-5Mb (Short-medium ROH), and 5-
10Mb (medium-long ROH), and 10-100Mb (long ROH). Individuals are arranged from
top to bottom by the sum length of ROH with wolves most representing the Indian and
Tibetan lineages bolded. **(C)** Pairwise sequentially Markovian coalescent (PSMC)
analysis of the demographic histories of 33 wolves representing multiple populations in
Eurasia and North America. Thicker lines indicate population belonging to the Indian
and Tibetan lineages, and a single individual from Shanxi, China. We assumed a
mutation rate of 4.5 x 10^{-9} and a generation time of 4.4 years.

Figure 5. Genetic load in selected wolf individuals in Asia and North America. **(A)**
Total genetic load, **(B)** masked load, and **(C)** realized load for 13 wolf individuals wihtin
five populations. Wolves found in historically smaller populations (Mexican, Indian, and
Tibetan) have a lower total load, lower masked load, and higher realized load. The total
derived allele count of each wolf within the five populations, where generally Mexican,
Indian, and Tibetan wolves show overall less derived alleles. The derived allele count of
(E) Loss of function (LOF) mutations classified as high impact, **(F)** missense mutation
classified as moderate impact, and **(G)** synonymous mutations classifed as low impact
per 100,000 derived alleles found in non-coding regions, assumed to be neutral. When
taking into account differences in overall number of derived count, Mexican, Indian, and
Tibetan wolves show reduced number of missense and synonymous mutations, but not
for loss of function mutation, compared to central Asian and southwest Asian wolves.

1362

1363
1364

1365 **Figure 1. Population genetic structure genetic connectivity of wolves across**
1366 **Eurasia.**

Figure 2. Tibetan-like ancestry in wolves from central Asia and eastern Asia.

1406

1407
1408
1409 **Figure 3. Persistence of Indian-like ancestry in wolves of southwest Asia.**

1424
1425
1426
1427

1428
1429
1430

Figure 4. Recent and long-term demographic history of wolves in Eurasia.

1431

1454 **Figure 5. Genetic load in selected wolf individuals in Asia and North America.**