1 **Inferring demographic and selective histories from population genomic data using a two-**
2 **step approach in species with coding-sparse genomes: an application to human data**
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Abstract

The demographic history of a population, and the distribution of fitness effects (DFE) of

newly arising mutations in functional genomic regions, are fundamental factors dictating

both genetic variation and evolutionary trajectories. Although both demographic and DFE

inference has been performed extensively in humans, these approaches have generally

either been limited to simple demographic models involving a single population, or, where a

complex population history has been inferred, without accounting for the potentially

confounding effects of selection at linked sites. Taking advantage of the coding-sparse

nature of the genome, we propose a 2-step approach in which coalescent simulations are

first used to infer a complex multi-population demographic model, utilizing large non-

functional regions that are likely free from the effects of background selection. We then use

forward-in-time simulations to perform DFE inference in functional regions, conditional on

the complex demography inferred and utilizing expected background selection effects in the

estimation procedure. Throughout, recombination and mutation rate maps were used to

account for the underlying empirical rate heterogeneity across the human genome.

Importantly, within this framework it is possible to utilize and fit multiple aspects of the

data, and this inference scheme represents a generalized approach for such large-scale

inference in species with coding-sparse genomes.

Teaser text

The demographic history of a population, and the distribution of fitness effects (DFE) of

newly arising mutations in functional genomic regions, are fundamental factors dictating

both genetic variation and evolutionary trajectories. Here we describe a 2-step approach for

inferring complex models of demography and selection in coding-sparse genomes. We

demonstrate the utility of this approach by optimizing demographic and DFE parameters in

human populations.

Keywords

demography; distribution of fitness effects; background selection; selective sweeps; genome

scans; genetic hitchhiking

Introduction

Genetic variation is a fundamental concern of population genetics. Prior to the advent of next-generation sequencing, the dominant debate within the field was centered on whether levels of genetic variation were expected to be minimal or substantial (known as the *classical/balanced* debate; see Lewontin 1987; Crow 1987). Selection was assumed as the dominant process in both cases, be it purifying selection depressing levels of variation, or balancing selection maintaining polymorphism (Dobzhansky 1955). Despite molecular evidence confirming plentiful levels of genetic variation, Motoo Kimura's Neutral Theory of Molecular Evolution (Kimura 1968, 1983) instead posited that observed variation was largely a consequence of genetic drift; that is, of neutral alleles segregating in the process of drifting towards fixation or loss. This hypothesis – that neutral rather than selective processes can explain the majority of observed variation – has since been largely corroborated (as reviewed in Jensen et al. 2019).

However, quantifying the precise roles of selective and neutral processes in shaping observed levels of variation – and disentangling their individual effects - remains an ongoing challenge due to the similar manners in which multiple evolutionary processes affect patterns of variation. One notable example is the extent to which neutral population growth, background selection (BGS; Charlesworth et al. 1993), and recurrent selective sweeps (Maynard Smith and Haigh 1974) can all skew the site frequency spectrum (SFS, the distribution of allele frequencies) toward rare alleles (Kim 2006; Jensen et al. 2007; Nicolaisen and Desai 2012, 2013; Ewing and Jensen 2016; Johri et al. 2021; Soni et al. 2023; and see review of Charlesworth and Jensen 2021, 2024). The effects of these processes are

further modified by genomic heterogeneity in mutation and recombination rates in often

complex ways (Soni et al. 2024b). Therefore, if one wishes to quantify the strength and

frequency of rare and episodic processes such as positive selection, one must first construct

an evolutionarily appropriate baseline model that accounts for the effects of constantly

occurring processes including genetic drift as modulated by historical population size

changes, as well as the effects of purifying selection and BGS resulting from the removal of

deleterious mutations (Bank et al. 2014; Johri et al. 2022a), all whilst accounting for

underlying mutation and recombination rate variation. Failure to account for these

processes is likely to lead to misinference, particularly in light of the fact that many

commonly studied populations and species are thought to have experienced not only

population growth, but also recent and severe population bottlenecks [e.g. humans

(Gutenkunst et al. 2009; Gravel et al. 2011; Excoffier et al. 2013) and *Drosophila*

melanogaster (Li and Stephan 2006), as well as a variety of human pathogens (Irwin et al.

2016; Sackman et al. 2019; Jensen 2021; Morales-Arce et al. 2021)], a demographic history

that is itself often strongly confounded with selective sweeps (Barton 1998; Poh et al. 2014;

Matuszewski et al. 2018; Harris and Jensen 2020; Charlesworth and Jensen 2022; Jensen

2023).

Constructing an evolutionarily appropriate baseline model for a given population will

therefore require inferring both a demographic history as well as the distribution of fitness

effects (DFE) of new mutations. However, because population history can confound DFE

inference, it is necessary to correct for the demographic history of the population in

question (Eyre-Walker and Keightley 2007; Boyko et al. 2008). The most commonly used

class of approaches are based on a framework in which demographic inference is performed

on putatively neutral sites, before utilizing that demographic history for DFE inference on

functional sites (Eyre-Walker and Keightley 2007; Boyko et al. 2008; Galtier 2016; Tataru and

Bataillon 2020; and see review of Johri et al. 2022b). Eyre-Walker and Keightley (2007)

obtained the first computationally inferred DFE estimates using this approach, and further

work incorporated a beneficial class of mutations into the inferred DFE (Boyko et al. 2008;

Eyre-Walker and Keightley 2009; Schneider et al. 2011; Galtier 2016).

Notably, this type of 2-step approach is often performed on functional regions under

the assumption that all sites are independent and unlinked, and that synonymous sites are

selectively neutral. However, these synonymous sites are likely experiencing BGS effects

(Charlesworth et al. 1993) due to linkage with directly selected and adjacent non-

synonymous sites, resulting in a skew in the SFS and thus mis-inference; in particular, these

BGS effects are often misinterpreted as population growth (Ewing and Jensen 2014; Johri et

al. 2021; and see review of Johri et al. 2022b). More generally speaking, there is indeed

substantial evidence that the effects of selection at linked sites may be widespread across

the genomes of many commonly studies species (see reviews of Cutter and Payseur 2013;

Charlesworth and Jensen 2021). Although recent work has shown that DFE inference is

relatively robust to the biasing effects of selection at linked sites (Kim et al. 2017; Huang et

al. 2021), that is not the case for demographic inference (Messer and Petrov 2013;

Nicolaisen and Desai 2013; Ewing and Jensen 2016; Schrider et al. 2016; Johri et al. 2021). It

is also noteworthy that these 2-step approaches are generally constrained to relatively

simple population histories utilizing a two-epoch model (Williamson et al. 2005; Keightley

and Eyre-Walker 2007; Kousanthanas and Keightley 2013).

The second class of methods involve using forward-in-time simulations (e.g., in SLiM;

Haller and Messer 2023) to jointly and simultaneously infer population history with the DFE

in an approximate Bayesian (ABC) framework (see Beaumont et al. 2002), as proposed by

Johri et al. (2020). Within this simultaneous inference scheme, it is neither necessary to

assume *a priori* the neutrality of synonymous sites, nor is it necessary to assume

independence amongst sites; as such, background selection can be directly modelled and

incorporated. While 2-step methods commonly infer a continuous distribution for the DFE,

this ABC framework infers a number of discrete DFE categories for various ranges of $2N_es$,

the population-scaled selection coefficient, where N_e is the effective population size and s is

the strength of selection acting on new mutations within the DFE category of interest. The

main drawback of such methods is that they are computationally expensive given the large

parameter space that must be explored when jointly inferring both demographic and DFE

parameters. As such, the inferred demographic models have thus far been limited to single-

step size changes in which the ancestral and current population sizes, as well as the timing

of size change, are inferred (Johri et al. 2020, 2023). Importantly however, in coding-dense

and/or non-recombining species in which sufficiently neutral, unlinked genomic regions may

not exist in the genome (thus precluding the needed neutral demographic inference

underlying 2-step approaches), this simultaneous inference framework remains the only

viable approach (e.g., Howell et al. 2023; Terbot et al. 2023a,b; Soni et al. 2024a).

It thus stands as an outstanding evolutionary inference question of how best to

accurately infer a necessarily complex and realistic demographic model, along with a

realistic DFE governing functional genomic regions, all whilst accounting for the variety of

discussed potential biases. Here we have investigated a modified 2-step approach applied to

human populations, in which the population history was inferred using non-functional

regions sufficiently distant from functional sites in order to avoid BGS effects, DFE inference

was then performed on exonic regions accounting for BGS effects and conditional on the

demographic history inferred in Step 1, and mutation and recombination rate maps were

utilized to account for the modulating effects of this underlying heterogeneity. By inferring

these parameters separately, a more biologically realistic population history was possible

accounting for the complexities of population size change, structure, and migration patterns

in these studied human populations, while the utilization of these distant non-functional

regions allowed for the reduction or elimination of the biasing effects of BGS on

demographic inference. Whilst a number of coalescent and diffusion approximation-based

approaches would be easily incorporated into our framework (e.g., Gutenkunst et al. 2009;

Excoffier et al. 2013; Jouganous et al. 2017; Wang et al. 2020), this approach – like the ABC

approach of Johri et al. (2020, 2022a) - has the benefit of utilizing various aspects of

population genomic data, including the SFS, associations between variants (linkage

disequilibrium, LD), and population differentiation.

As human populations have naturally been highly studied, with numerous published

demographic models, we here provide an optimized and well-fitting 4-population

demographic model for the Out-of-Africa (OOA) expansion. Conditional on this model, we

additionally optimized a DFE using genic regions, fitting both levels and patterns of

polymorphism and divergence, and finding consistency with the recent DFE estimates of

Johri et al. (2023). Finally, we have evaluated the degree to which positively selected

mutations may be identifiable within the context of this fit model. This work thus provides a

valuable and improved framework for evolutionary inference in coding-sparse genomes,

and for the construction of evolutionary baseline models in such species.

Methods and Materials

Data

This study was based on the GRCh37 human reference genome, with SNP data and

accessibility masks obtained from 1000 genomes variant call format and bed files,

respectively (The 1000 Genomes Project Consortium 2015). The data was split into

continental populations, informed by levels of admixture, as determined by The 1000

Genomes Project Consortium (2015). The total number of samples from each of the four

considered populations were: African – 99; European – 502; East Asian – 104; South Asian –

489. We obtained recombination and mutation rate maps from Halldorsson et al. (2019) and

Froncioli et al. (2015), respectively, gene annotations from NCBI (Sayers et al. 2022),

ancestral sequences from the six-way EPO alignments available from Ensembl (Flicek et al.

2014; Cunningham et al. 2022), and we identified conserved elements via the 100-way

PhastCons score (Siepel et al. 2005; Pollard et al. 2010). See Supplementary Table S1 for

links to all downloaded data.

Selecting non-functional regions for demographic inference

For demographic inference we identified non-functional regions of the human

genome that were at a distance of at least 10kb from the nearest functional region (as per

the NCBI GFF file [Sayers et al. 2022]). We then masked these regions using both strict

accessibility masking (The 1000 Genomes Project Consortium 2015) and conserved element

masks (i.e., with a phastCons score > 0 [Siepel et al. 2005; Pollard et al. 2010], in order to

remove sites potentially experiencing purifying selection and generating background

selection effects (e.g., binding sites (Simkin et al. 2014))). Across each region, we calculated

mean recombination and mutation rates, with any regions lacking this information being

removed. Finally, we set a minimum length threshold of 15kb to ensure that regions were

long enough to reliably calculate summary statistics. Following these steps, we were left

with a total of 146 non-functional regions. Finally, we used the B maps of McVicker et al.

(2009) to compare the distribution of B values (i.e., the estimated reduction in diversity

attributed to BGS by McVicker et al. (2009)) to the distribution of our non-functional

regions. For this analysis we lifted over B map coordinates from the hg18 human genome

assembly to the GRCh37 assembly using the UCSC liftover tool (Karolchik et al. 2003).

Supplementary Figure S1 provides plots of this comparison, as well as the distributions of

region lengths, SNPs, and mutation and recombination rates for our set of curated non-

functional regions.

Selecting exons for DFE inference

We used the set of exons curated by Johri et al. (2023), although our focus was on

the exonic regions only, as opposed to the exons and the neighbouring intergenic regions.

Because we used different recombination and mutation rate maps (as described in the data

section above), we recalculated mean rates across the 465 exonic regions, removing regions

for which we did not have rate information, leaving a total of 397 exonic regions.

Calculating empirical summary statistics

We calculated summary statistics for each population sampled using the python

library for libsequence, Pylibseq v0.2.3 (Thornton 2003), except for F_{ST} which was estimated

using scikit-allel (Miles et al. 2024). The number of segregating sites and F_{ST} were calculated

per site, whilst Tajima's D (Tajima 1989) and mean r^2 were calculated over 10kb windows for

each non-functional region, and per region for each exon.

Exonic divergence was calculated based on the number of fixed differences between

the reference and ancestral sequences, with polymorphic sites masked, relative to total

region size.

Calculating summary statistics from simulated data

We calculated summary statistics from simulated data in a manner that replicated

the empirical data, using the same software as described above. Thus, sites that had been

masked in the empirical data were also masked in the simulated data prior to calculating

summary statistics.

Exonic divergence was calculated as the number of fixations post-burn-in from

forward-in-time simulations (see the 'Simulating human population history with selection

using SLiM' section).

We calculated the mean and standard deviation for each region across its respective

100 replicates. For plotting purposes, we plotted the mean of all regions as the data point,

and the mean of the standard deviations across all regions as the confidence intervals.

Simulating human population history using Msprime

234   Step 1 in our 2-step inference framework was the inference of population history.

We simulated human demography using the coalescent simulator Msprime (Baumdicker et

al. 2022) for each of our 146 non-functional regions, with region specific mutation and

recombination rates. Our demographic model was comprised of 5 populations (four

sampled populations: African, European, South Asian and East Asian; as well as the

unsampled ancestral Eurasian population) and 25 parameters. Parameter ranges were taken

from the human demographic inference literature, with midpoints of all ranges used as the

initial starting parameterization. A generation time of 26.9 years was used to appropriately

scale simulations (Wang et al. 2023). For details of the demographic model see Figure 1 and

Table S2. 100 replicates were simulated for each of the 146 non-functional regions, with a

single mutation and recombination rate per region, calculated as the average across the

region from the Francioli et al. (2015) mutation rate map and the Halldorsson et al. (2019)

recombination rate map (see Supplementary Figure S1 for distributions of region lengths,

mutation rates and recombination rates across curated regions). Parameters were

optimized to the data using F_{ST}, the number of segregating sites, Tajima's D (Tajima 1989)

and mean r^2, across all four populations. Demographic inference plots (e.g., Figure 1) were

produced using Demes software (Gower et al. 2022).

Simulating human population history with selection using SLiM

253   For Step 2, we simulated the inferred population history from Step 1 using the

forward-in-time simulator SLiM (v4.0.1 [Haller and Messer 2023]), for our 397 exonic

regions, with region specific mutation and recombination rates. We simulated to the

human-chimpanzee split (12mya; Moorjani et al. 2016). Thus, the simulations considered 12

million years (446,100 generations) before starting the 10N generation burn-in period.

Exonic mutations were drawn from a DFE comprised of 4 fixed classes (following Johri *et al.*

2020), with frequencies denoted by f_i: f_0 with $0 \leq 2N_{AFRancestral}\, s < 1$ (i.e., effectively neutral

mutations), f_1 with $1 \leq 2N_{AFRancestral}\, s < 10$ (i.e., weakly deleterious mutations), f_2 with $10 \leq$

$2N_{AFRancestral}\, s < 100$ (i.e., moderately deleterious mutations), and f_3 with $100 \leq 2N_{AFRancestral}\, s$

(i.e., strongly deleterious mutations), where $2N_{AFRancestral}$ is the initial African population size

and s is the reduction in fitness of the mutant homozygote relative to the wild-type. We

initially simulated the DFE from Johri et al. (2023) - comprised of neutral and deleterious

mutations - which fit the empirical data well.

Simulating selective sweeps

<u>Recurrent</u>

We simulated recurrent selective sweeps by adding a beneficial DFE category for our

397 exonic regions. We simulated three different beneficial rates (0.1%, 1%, and 10% of new

mutations), with the effectively neutral DFE category (f_0) reduced to account for the

addition of the beneficial category. Three different beneficial classes were separately

simulated: $1 \leq 2N_{AFRancestral}\, s_b < 10$; $10 \leq 2N_{AFRancestral}\, s_b < 100$ and $100 \leq 2N_{AFRancestral}\, s_b <$

1,000, where s_b is the increase in mutant homozygote fitness relative to the wild-type.

<u>Individual</u>

To simulate a single hard selective sweep, we ran our inferred demographic model

with the inferred DFE, with three different scenarios for introducing a beneficial mutation:

model 1) the beneficial mutation was introduced into the African population immediately

after burn-in; model 2) the beneficial mutation was introduced into the ancestral Eurasian

population immediately after splitting from the African population; and model 3) the

beneficial mutation was introduced into the European population immediately after

splitting from the Eurasian population. In model 1, simulations were terminated and

restarted if the beneficial mutation did not fix in all 4 sampled populations. In model 2,

simulations were terminated and restarted if the hard sweep did not fix in the European,

East Asian and South Asian populations. Finally, in model 3 simulations were terminated and

restarted if the hard sweep did not fix in the European population. For each scenario, two

different strengths of selection were simulated: $2N_e s_b$ = 1,000 and 10,000, where N_e is the

ancestral African population size ($N_{AFRancestral}$) and s_b is the beneficial selection coefficient.

For these simulations, we utilized the chromosomal structure of Soni and Jensen

(2024), with functional regions comprised of 9 exons (each of size 1,317bp) and 8 introns

(each of size 1,520bp), separated by intergenic regions (each of size 4,322bp) [The 1000

Genomes Project Consortium 2015]. The number of exons and introns per functional region

were taken from Sakharkar et al. (2004). The chromosomal region contained 7 functional

regions in total, resulting in a total simulated region length of 198,345bp.

Variable mutation and recombination rates were drawn from a uniform distribution

such that the mean recombination rate across the simulated region for each replicate was

equal to the Kong et al. (2010) mean, and the mean mutation rate across the simulated

region for each replicate was equal to the Kessler et al. (2020) mean.

Sweep inference with SweepFinder2

We performed selective sweep inference by running SweepFinder2 (DeGiorgio et al.

2016) on each simulated replicate of each exonic region from our hard sweep simulations.

Allele frequency files were generated for each replicate, following Huber et al.'s (2016)

recommendation of including only polymorphic and substitution data. Inference was

performed at each SNP via a grid file, following Nielsen et al. (2005). The background SFS

was taken from the sweep-free simulations inferred in this study. The following command

line was used for inference:

SweepFinder2 -lru GridFile FreqFile SpectFile RecFile OutFile

Sweep inference with H12

We ran the H12 method of Garud et al. (2015) on each simulated replicate of each

exonic region from our hard sweep simulations, using a custom python script. H12 was

estimated over 1kb, 2kb, 5kb, 10kb, 20kb, and 40kb windows at each SNP, with the SNP at

the center of each window.

For both SweepFinder2 and H12 inference, we calculated true- and false-positive

rates based on the inference values at each site, generating ROC curves from this

information.

Results and Discussion

Our implemented 2-step approach to demographic and DFE inference involves inferring population history using non-functional regions that are at a sufficient distance from functional sites so as to reasonably ensure that they are not experiencing purifying or background selection effects. DFE inference is then performed on exonic regions in Step 2, conditional on the demographic history inferred in Step 1 and incorporating expected background selection effects. We have applied this approach to human population genomic data from the 1000 genomes project (The 1000 Genomes Project Consortium 2015), in order to better characterize the evolutionary parameters governing recent human history.

Step 1: Demographic inference on non-functional regions

In order to avoid the biasing effects of purifying selection and BGS, we performed demographic inference on our curated set of 146 non-functional regions, with mean recombination and mutation rates calculated for each region from the rate maps of Halldorsson et al. (2019) and Francioli et al. (2015), respectively. For details of the data curation steps, please see the Methods section. While one would typically begin with an evaluation of numerous demographic models and topologies in less well-characterized species (see Beaumont et al. 2002; Johri et al. 2020), given the considerable literature on human demographic history (e.g., Gutenkunst et al. 2009; Gravel et al. 2011; Schiffels and Durbin 2014; Terhorst et al. 2017; Hu et al. 2023), and inferred levels of admixture in The 1000 Genomes dataset (The 1000 Genomes Project Consortium 2015), we began with a model of the Out-Of-Africa (OOA) colonization in which the ancestral Eurasian population splits from the African population, followed by the European, South Asian and East Asian

populations dispersing from the ancestral Eurasian population, along with the Bantu

expansion in the African population. Thus, our demographic model was comprised of 5

populations (African, ancestral Eurasian, European, South Asian and East Asian, of which all

but the ancestral Eurasian population were sampled) and 25 parameters that capture

population sizes, bottleneck severities, growth rates, timings of each event, and migration

rates between populations. Parameter ranges were drawn from the extensive literature on

human population history (Mellars 2006; Gutenkunst et al. 2009; Gravel et al. 2011;

Tennessen et al. 2012; Terhorst et al. 2017). Figure 1a provides the parameter ranges for

our model, and see Methods section for further details.

359   We simulated 100 replicates for each of our 146 non-functional regions using the

coalescent simulator MSprime (Baumdicker et al. 2022) with region-specific mutation and

recombination rates, initially starting with midpoint values for each of our parameters (see

Figure 1a). For each replicate we estimated four summary statistics for each population (or

pairs of populations): the number of segregating sites, Tajima's D (Tajima 1989), mean r^2,

and F_{ST}, giving us a total of 18 summary statistics. Fitting these four statistics enabled us to

account for multiple aspects of the data including levels of diversity, the SFS, LD and

population structure. Figure 1a provides the optimized fit of each parameter within the

context of previously published parameter ranges, and Figure 1b the total inferred

demographic model. As shown in Figure 2, the summary statistics resulting from this

demographic model well fit observed empirical data.

371   It is notable that the African population in our model is larger than the African

populations in the Gutenkunst et al. (2009) and Gravel et al. (2011) best-fitting models.

There are two likely contributing factors. Firstly, these previous studies fit the model to the

SFS, whereas we have here fit multiple diverse summaries of the data. Secondly, these

previous studies modeled the African population with a fixed size that undergoes a single

instantaneous expansion. Here we modelled the recent Bantu expansion, and thus our final

African population size was notably larger, though our final African population size of 87,594

falls within the range of previous estimates (Schiffels and Durbin 2014; Terhorst et al. 2017;

Johri et al. 2023). Finally, it is worth noting that numerous other coalescent and diffusion

approximation-based approaches have been used to infer the OOA model of human

population history (Gutenkunst et al. 2009; Gravel et al. 2011; Excoffier et al. 2013;

Jouganous et al. 2017; Wang et al. 2020). These studies have masked genic regions to avoid

the biasing effects of selection. However, BGS can still affect demographic inference if not

accounted for; nonetheless, our parameter estimates fall within previously inferred ranges,

confirming the modest nature of BGS effects in humans (Johri et al. 2021; Buffalo and Kern

2024).

In summary, by optimizing within previously published parameter ranges, we have

identified a neutral demographic model that well explains multiple facets of the genomic

data in distant non-coding regions.

Step 2: DFE inference on functional regions

Given the strong fit of the neutral demographic model to the intergenic data, we

next moved to Step 2: inference of the DFE using functional regions. We utilized the curated

set of functional regions from Johri et al. (2023). After obtaining region-specific mutation

and recombination rates we were left with a total of 397 functional regions. Unlike Johri et

al. (2023) who simulated exons and their neighboring regions, we focused on the exons only

(given that the model fit was consistent across both exons and adjacent regions in their

study). First, we simulated our 397 functional regions under the demographic model

inferred in Step 1, using the forward-in-time simulator SLiM (v4.0.1 [Haller and Messer

2023]). For the purpose of DFE inference, we simulated to the human-chimpanzee split time

(12mya [Moorjani et al. 2016]) to allow us to compare empirical and simulated divergence,

which is expected to be shaped by selection at functional sites. When simulating these

functional regions under selective neutrality, we found that the fit to the empirical data was

poorer than for the non-functional regions (Supplementary Figure S2); an expected result

given the action of selection in these exonic regions. Next, we simulated under the Johri et

al. (2023) DFE using our fit demographic model, and found a good fit of the simulated

summary statistics to the empirical data (Figure 3). These results are encouraging given the

differing approaches taken between the two studies: we here took the 2-step approach as

described, whilst Johri et al. utilized a simultaneous inference scheme. Importantly

however, both studies accounted for expected BGS effects, a relative rarity in DFE inference.

Though the inclusion of population history, purifying and background selection

effects, and mutation and recombination rate heterogeneity were alone sufficient to explain

empirically observed data patterns, that does not necessarily imply the absence of beneficial

mutations; rather, it suggests that this additional parameter is not needed in order to fit

observed patterns of variation. While this observation is itself meaningful, it indeed raises

the question of what rate of beneficial mutation may be consistent with the data but simply

unidentifiable. In order to investigate this question, we added a beneficial DFE category to

the Johri et al. (2023) DFE, in an attempt to understand what rate of input of beneficial

mutations may be compatible with the observed levels of variation, the SFS, LD, divergence

and F_{ST}. Initially, we considered three beneficial DFE proportions, f_{bo} = [0.1%, 1%, or 10% of

newly arising mutations], with $1 \leq 2N_{AFRancestral} s_b < 10$ (i.e., weakly beneficial mutations).

Under this model, we correspondingly reduced f_0 - the proportion of effectively neutral

mutations – in order to account for the addition of this beneficial DFE class. Supplementary

Figures S3-S5 provide the fit of the summary statistics from these simulations to the

observed data. At f_{b0} = 0.1% or 1%, all summary statistics remain reasonably well fit - in

other words, they are not significantly modified from the expectations in the absence of

positive selection. However, divergence was notably increased relative to that observed at

f_{b0} = 10%, due to the greater rate of beneficial fixation.

Given that this beneficial mutation rate of 10% appears inconsistent with empirical

divergence, we next examined f_{b0} = 0.1% and 1% only, whilst increasing the population-

scaled strength of selection to $10 \leq 2N_{AFRancestral} s_b < 100$ (i.e., moderately beneficial

mutations). Supplementary Figures S6 and S7 provide the fit of summary statistics from

these simulations to the observed data. With this increased strength of selection, the

modelled divergence only fit the empirical data at the lowest beneficial frequency, f_{B0} =

0.1%. Finally, we increased the population-scaled strength of selection further to $100 \leq$

$2N_{AFRancestral} s_b < 1000$ (i.e., strongly beneficial mutations), at f_{B0} = 0.1%. Even at this low

frequency, the resulting divergence was too high relative to the empirical data (see

Supplementary Figure S8 for all summary statistics). It is notable that regardless of beneficial

mutation frequency or strength of selection, the other summary statistics fit the data well -

this owes to the relative waiting time between selective sweeps under these models; that is,

selective sweeps are too old on average to strongly impact patterns of polymorphism

(Jensen 2009), while being frequent enough to modify divergence over the 12mya time-

scale.

Taken together, these results suggest that whilst the addition of a beneficial DFE

class is not necessary to explain the patterns observed in the human population genomic

data here considered, a modest input of weakly beneficial mutations and/or a low input of

moderately beneficial mutations would remain consistent with the observed data.

Evaluating power to detect selective sweeps within this human baseline model

Recurrent sweep models, such as the one studied above, involve a scenario in which

beneficial mutations occur randomly across a chromosome according to a time-

homogenous Poisson process at a per-generation rate (Kaplan et al. 1989; Wiehe and

Stephan 1993; Stephan 1995; Pavlidis et al. 2010; Soni et al. 2023). Although this is a more

realistic model of positive selection, in that the beneficial mutations underlying selective

sweeps naturally occur at a per-generation rate - meaning that they are naturally associated

with an average time since fixation - the more commonly studied model involves a single

selective sweep in which fixation occurred immediately prior to sampling. As such, these

models consider a best-case scenario for sweep detection, both in that sweeps are as recent

as possible thus maximizing detectable polymorphism-based patterns (see review of Nielsen

2005), but also because it avoids the possibility of interference between positively selected

mutations (i.e., Hill and Robertson 1966).

Furthermore, these models are often simulated on otherwise neutral backgrounds,

which is additionally unrealistic in the sense that beneficial mutations occur in functional

regions, which will be dominated by newly arising deleterious mutations. Thus, as a step

towards biological reality, we here have modelled single selective sweeps within the context

of our evolutionary baseline model, using our inferred demographic history, DFE, as well as

mutation and recombination rate maps, thereby accounting for constantly-operating

evolutionary processes in order to characterize the power to identify an episodic selective

sweep (as described by Johri et al. 2022a).

Under this model, we simulated a large genomic region comprised of functional and

non-functional regions in which a single hard selective sweep occurred in a functional

element (see Methods section for more details about simulated chromosomal structure, as

well as parameterizations). Sweep inference was conducted using two methods: the

composite-likelihood ratio (CLR) SFS-based method, SweepFinder2 (DeGiorgio et al. 2016),

and a haplotype-based approach, H12. Three different sweep models were simulated: 1) a

beneficial mutation introduced into the ancestral African population immediately after

simulation burn-in, with the fixed beneficial present in the sampled African, European, East

Asian and South Asian populations; 2) a beneficial mutation introduced into the ancestral

Eurasian population immediately after splitting from the ancestral African population, with

the fixed beneficial present in the sampled European, East Asian and South Asian

populations; and 3) a beneficial mutation introduced into the European population

immediately after splitting from the Eurasian population, with the fixed beneficial present in

the sampled European population. Figure 4 presents ROC plots, plotting the false positive

rate (FPR) against the true positive rate (TPR) for inference on each model across 100

replicates with SweepFinder2 (with inference performed at each SNP) and H12 (with

inference performed across 1kb windows, centered on each SNP; see Supplementary

Figures S9-13 for additional window sizes).

At the lowest strength of selection ($2N_es_b$ = 100), no beneficial mutations reached

fixation by the sampling time (i.e., the present day) across the replicates. As such, Figure 4

presents ROC plots for $2N_es_b$ values of 1,000 and 10,000 only. Although SweepFinder2

showed greater inference power than H12, there was limited power to detect selective

sweeps for both approaches. While potentially appearing counter-intuitive, in some

circumstances $2N_es_b$ = 1,000 had greater power than $2N_es_b$ = 10,000, as the fixations of the

former were more recent given the longer sojourn time, and thus experienced less post-

fixation decay in patterns of polymorphism (Kim and Stephan 2000; Soni et al. 2023). These

results thus suggest that detectable selective sweeps would necessarily be the result of

positive selection that was strong and recent enough to leave a detectable signature,

consistent with previous work (Przeworski 2002; Kim and Stephan 2002; Jensen et al. 2007;

Crisci et al. 2013). Moreover, the modest power under our baseline model is likely explained

by the severe bottlenecks and expansions characterizing these populations, as the

fundamental difficulty in distinguishing between population bottlenecks and selective

sweeps has been previously demonstrated (Barton 1998; Jensen et al. 2005). These results

suggest that caution is needed when performing genomic scans for selection in humans due

to their complex recent demographic history, and likely supports previous assertions that

strong selective sweeps have been rare in recent human history (Hernandez et al. 2011).

Conclusions

In this study we have demonstrated the viability of a 2-step approach for inferring

population history along with the DFE in coding-sparse genomes, such as that characterizing

humans. This condition, together with being a recombining genome, is important for the

existence and availability of non-functional regions sufficiently distant from functional sites

so as to be free from the effects of purifying and background selection, as such regions are

necessary for the accurate inference of population history. By contrast, organisms with

genomes that are either coding dense or experience limited recombination may not have

such regions in sufficient number, in which case demographic inference must be performed

within the context of background selection effects. As these background selection effects

will be dictated partially by the DFE in functional regions, this genomic architecture requires

the joint and simultaneous inference of demographic and selective parameters - a situation

that spans organisms ranging from *Drosophila* to many viruses (see review of Johri et al.

2022b). However, given the multiple jointly inferred parameters, the demographic histories

under these joint inference schemes have been highly simplified in current

implementations. Thus, this 2-step approach has a distinct advantage for coding-sparse

genomes, in that previously developed and sophisticated neutral demographic inference

approaches may be leveraged in Step 1 - such as that employed here estimating a 25-

parameter human demographic model consisting of multiple population size changes, split

times, and migration rates - allowing DFE inference to be focused upon in Step 2 conditional

on that inferred history.

It is additionally important to consider the extent to which a consideration of these

BGS effects matters for human demographic inference. Indeed, given the coding-sparseness

of the genome, these effects are expected *a priori* to be limited, and that is fully consistent

with the observation that our optimized demographic parameter values fall within

previously published parameter ranges. However, apart from accounting for the effects of

selection at linked sites, this approach also utilizes patterns of variation in addition to the

site frequency spectrum (e.g., linkage disequilibrium and population-differentiation), which

provide a further valuable 'sanity check' on estimated models. This combination of factors

has resulted in incrementally improved - but indeed improved - parameter estimates for the

populations studied, as assessed by the fit between the estimated model and the empirical

data. Thus, this proof-of-principle approach applied here to publicly-available human data

will likely provide a highly relevant and informative inference framework for the analysis of

future genomic resources in comparatively poorly-studied species with a similar genomic

architecture (e.g., non-human primates).

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Figure 1: Demographic model representing the Out-of-Africa expansion. a) Parameter ranges for all 25 parameters (represented by the blue bars on the plots). Orange dots indicate the best fitting parameter values identified. b) Plot of demographic model with the best fitting parameter values. Population key: AFRancestral = initial ancestral African population; AFR = African population; EURASI = unsampled Eurasian population; EUR = European population; EAS = East Asian population; SAS = South Asian population. Parameter key: τ = time of splits between specified populations (with τ_{BANTU} representing the time of start of the Bantu expansion in the African population); r = growth parameter; N = population size; B = bottleneck severity; m = migration rate. Demographic model graphic generated using Demes software (Gower et al. 2022).

Figure 2: Summary statistics calculated from putatively neutral non-functional regions from population samples for empirical (blue) data, compared to simulated (red) data under the best-fitting demographic model. Means and standard deviations were calculated for 100 replicates. Data points represent the mean across regions, while bars represent the mean of the standard deviations across all regions.

Figure 3: a) to e) Summary statistics calculated from functional regions from population samples for empirical (blue) data, compared to simulated (red) data under the best-fitting neutral demographic model with the addition of purifying and background selection modelled using

the Johri et al. (2023) DFE (shown in panel f). Following this DFE, exonic mutations were drawn from a DFE comprised of 4 fixed classes with frequencies denoted by f_i: f_0 with $0 \leq 2N_{AFRancestral} \, s < 1$ (i.e., effectively neutral mutations), f_1 with $1 \leq 2N_{AFRancestral} \, s < 10$ (i.e., weakly deleterious mutations), f_2 with $10 \leq 2N_{AFRancestral} \, s < 100$ (i.e., moderately deleterious mutations), and f_3 with $100 \leq 2N_{AFRancestral} \, s$ (i.e., strongly deleterious mutations), where $N_{AFRancestral}$ was the initial population size and s the reduction in fitness of the mutant homozygote relative to wild-type. Means and standard deviations were calculated for 100 replicates. Data points represent the mean across regions, while bars represent the mean of the standard deviations across all regions.

Figure 4: ROC curves, providing the change in true-positive rate (TPR) with false-positive rate (FPR), for sweep inference with SweepFinder2 (SF2) and the H12 statistic under the demographic model inferred in this study (see Figure 1) together with the Johri et al. (2023) DFE for functional regions, and variable mutation and recombination rates (see Methods section). Here, a single beneficial mutation was introduced into the population at three different time points and in three different populations: Model 1: the beneficial mutation was introduced into the ancestral African population immediately after the burn-in period, the beneficial fixation is present in all populations, and sweep inference was conducted on all sampled populations; Model 2: the beneficial mutation was introduced into the ancestral Eurasian population immediately upon splitting from the ancestral African population, the beneficial fixation is present in all non-African populations, and sweep inference was conducted on the European, East Asian and South Asian populations; Model 3: the beneficial mutation was introduced into the European population immediately upon splitting from the Eurasian population, the beneficial fixation is present in the European population, and sweep inference was conducted on this population only. For each model, two different strengths of selection were modelled: $2N_e s_b$ = 1,000 and $2N_e s_b$ = 10,000, where N_e is the size of the ancestral African population and s_b is the selection coefficient of the beneficial mutation. Inference with SweepFinder2 was performed on each SNP and substitution, whilst H12 inference was performed on each SNP over a 1kb window, with the SNP at the center of the window.