Accuracy in bird predation assessment: Camera traps testing the efficacy of plasticine caterpillars as prey models

Accuracy in bird predation assessed by camera traps

Laura Schillé[1,¤], Nattan Plat[1,¤], Luc Barbaro[2], Hervé Jactel[1], Frédéric Raspail[1], Jean-Baptiste Rivoal[1], Bastien Castagneyrol[1,#], Anna Mrazova[1,3,#]

Affiliations
[1]BIOGECO, INRAE, University Bordeaux, Cestas, France
[2]DYNAFOR, University of Toulouse, INRAE, Toulouse, France
[3]Institute of Entomology, Biology Centre of the Czech Academy of Sciences, Ceske Budejovice, Czech Republic
[¤]Laura Schillé and Nattan Plat share the first co-authorship.
[#]Anna Mrazova and Bastien Castagneyrol share the last co-authorship.

Contributions

A.M., L.S., and N.P. developed the original idea. A.M., B.C., L.S., and N.P. developed the

methodology. A.M., B.C., H.J., JB.R., L.B., L.S., and N.P. conducted data collection. F.R.

implemented the computer code. A.M. and L.S. did the formal analysis and wrote the

original draft, and all co-authors provided critical reviews of the manuscript.

Abstract

Sampling methods that are both scientifically rigorous and ethical are cornerstones of any

experimental biological research. Since its introduction 30 years ago, the method of using

plasticine prey to quantify predation risk has become increasingly popular in the field of

biology. However, recent studies have questioned the accuracy of the method, arguing that

inaccuracy in inferring evidence of predator bite marks and the artificiality of models may

bias the results. Yet, bias *per se* might not be a methodological issue as soon as its statistical

distribution in the samples is even, quantifiable, and thus correctable in quantitative

analyses. In this study, we focus on avian predation of lepidopteran larvae models, which

represent one of the most widespread predator-prey interactions in various ecosystems

worldwide. We compared bird predation on plasticine caterpillar models to that on dead

caterpillars of similar size and color, using camera traps and observer posterior assessment

to identify biases and quantify actual predation events. Although camera trap analyses

revealed that birds respond similarly to plasticine models and dead caterpillars, suggesting

no advantage in using dead prey for assessing avian predation, the results of posterior

identification of predation marks showed contradictory results, making the interpretation

of the results ambiguous. Observer inconsistencies in detecting predation clues on models

highlight potential biases, with camera trap data showing these clues do not reliably

indicate actual bird predation. Given the uncertainties and shortcomings of technological

character revealed, the development of adequate monitoring methods and further research

is essential to refine the assessment approach and better understand the biases in using

plasticine models.

Keywords: artificial prey; bird predation; camera trap; ecological methods; fake prey;

plasticine caterpillar models; real caterpillars

Introduction

The functional role of insectivorous birds in terrestrial ecosystems [1,2] has been the focus

of many studies aimed at understanding the predation patterns, feeding behavior, predator

control of prey populations and evolutionary ecology [3–6]. Traditional methodological

approaches to these research topics include direct field observations [7], experimental

manipulation in the laboratory [8,9] or in the field [10], and gut or fecal DNA analyses

(11,12). Each of these methods has its own set of limitations regarding standardization,

logistics, and ethical considerations such as not sparing living prey or handling captive

birds. To address these challenges, the use of artificial prey, particularly those made of

plasticine, introduced over 30 years ago by Brodie III (13), has become a valuable tool in

bird predation research (14). The method is primarily appreciated for its ability to preserve

the bite marks of predators, the identification and quantification of which has been used to

infer attacks and predation risk from specific predators.

While plasticine caterpillar models are widely employed as simple, standardized prey

models to investigate predation activity in diverse ecosystems (15–17), recent studies have

raised concerns regarding their limitations and potential inaccuracies (14,18–20). The main

concerns relate to the accuracy of inferences of predator identity and activity from the

posterior assessment of putative predation marks on prey models.

Inaccuracy of predator identity - Observers may struggle to attribute a considerable

portion of attack marks to a specific predator type, e.g., bird, reptile, or arthropod (18,19).

For example, Valdés-Correcher et al. (21) found that up to 24% of predation mark

identifications were inaccurate when assessed by untrained scientists using photos of

attacked plasticine caterpillar models. This was observed in comparison to evaluations

conducted by an expert who had access to the actual plasticine caterpillar models. This

result shows that the identification of predator marks often relies on subjective human

judgment, which can lead to observer bias. This variability is accentuated by the absence of

a standardized training system for observers, thus giving them more certainty in

recognizing such marks (18,22; but see Low et al. (23), for examples of predator-type

classified bite mark images), leading to potential discrepancies in interpretations among

researchers.

Inaccuracy in determining real predation events among all identified marks - Even

though marks were attributed to birds, questions would remain about what one can infer

from them. Incidental contact or non-predatory interactions (e.g., by bird claws, attraction

of non-predatory animals, or curiosity) with the plasticine caterpillar models can result in

marks that are difficult to distinguish from genuine predation attempts, potentially leading

to an overestimation of predation rates (24). The misinterpretation of marks can lead to

distorted results on predation pressure and misleading conclusions about predator-prey

interactions, especially in areas where the attack rate is low (25).

Furthermore, the artificial appearance and texture of plasticine may not adequately

simulate the characteristics of real prey, including smell, possibly affecting the behavior of

insectivorous birds and thus the validity of predation assessment (19). The literature

contains several studies that compared the attack rates on plasticine larvae to those of real

larvae, with the latter being quantified based on missing or partially consumed prey items

(19,20,24). As opposed to plasticine caterpillar models, the attack rate on living and dead

prey has shown comparable results (19), thus justifying not using living prey for ethical

reasons (not to glue or pin them alive). By simultaneously studying predation on dead

caterpillars versus plasticine models, it becomes possible to compare the response of

predators to both types of prey within the same experimental design with the use of passive

monitoring methods. This comparative approach makes it possible to test the assumptions

on which the plasticine caterpillar model method is biased and to correct it in relation to

observable predation patterns on real dead prey. Quantifying this bias is critical to

determining whether plasticine models can reliably mimic real prey, and thus whether the

data they provide can reflect an accurate depiction of predation pressure.

Studies comparing predation on both real caterpillars and plasticine models have shown

discrepancies, revealing that plasticine models may not be a sufficiently accurate method

for assessing predation (19), particularly across large biogeographical ranges (24), varying

thermal regimes (26), different seasons of the bird life cycle (20), and for different taxa

(19). There appears to be a gap in the literature regarding more detailed analyses that

elucidate the origins of the marks found on plasticine models. Moreover, missing plasticine

models are usually excluded from the statistical analyses (16,27). Yet, at least part of those

missing models can be the result of a predation attempt that was not recorded. On the

contrary, studies working with real prey have often assumed that a missing prey was

equivalent to a depredated prey (19,20,24), an assumption that may not always hold true.

Without direct observation, the reasons behind the disappearance of the prey remain

speculative.

To address these methodological uncertainties, and following recommendations from

Bateman et al. (18) and Rößler et al. (22), we employed camera traps. As they have been

already used to study bird frugivory (28,29), we suggest their application also to

experimentally record predation on plasticine models (hereafter "models") paired with

dead caterpillars (hereafter "corpses") on the experimental trees. We hypothesized that the

integration of camera traps could visually confirm predation attempts, providing a direct

and unbiased record of predator interactions with the prey. This visual evidence could

enable the differentiation between actual predation marks and those resulting from

incidental contact or environmental factors (e.g., rubbing by leaves or branches), therefore

increasing the accuracy of predation rate assessment. Ultimately, we believe that the results

of this study will help to refine the plasticine caterpillar model method as a tool for

ecological research, ensuring that it produces scientifically relevant results.

We used camera traps to quantify predation events on both prey types and possible sources

of bias in inferences made from plasticine models. We first assessed bias in the detection of

predation clues on plasticine models due to variation among observers (H1). We further

hypothesize that observer detection of predation clues leads to an overestimation of

predation on plasticine models by attributing unrelated or uncertain marks to actual

predation attempts (H2). Taking into account the innate ability of insectivorous birds to

recognize suitable prey and the differences in the surface texture of real vs. plasticine

caterpillars, we hypothesize that there will be evidence of more predation on corpses than

on models, both assessed using camera traps (H3). Finally, we hypothesize that observer

detection of predation would lead to an overestimation of the predation on corpses by

accounting for missing individuals as a predation event, lacking direct evidence for why the

prey disappeared (H4). If we obtain comparable results of the bias between observers and

cameras for both models and corpses, and if the test of H3 provides evidence of more

predation on corpses than on models, we could conclude that the use of corpses with

human post-detection would be a more accurate method than the use of plasticine models

with human post-detection.

Material and Methods

Study area

We carried out this experiment in a pine and oak forest stand of 5ha near Bordeaux, France (44°440N, 00°460W). The climate is oceanic, with 12.8°C average temperature and 870mm annual rainfall. We synchronized the start of the experiment as closely as possible with the breeding period of local insectivorous birds, specifically, when the chicks were in the nest and fed by the parents (starting generally mid-April for resident breeders or early migrants). We choose this timing to quantify bird predation by experienced adults rather than later exploratory predation attempts by young, inexperienced birds. The main insectivorous bird species in the study area were Great Tit (*Parus major*), Eurasian Blue Tit (*Cyanistes caeruleus*), Eurasian Blackcap (*Sylvia Atricapilla*), Common Redstart (*Phoenicurus phoenicurus*), Eurasian Blackbird (*Turdus merula*), Western Bonelli's Warbler (*Phylloscopus bonelli*), Common Chiffchaff (*Phylloscopus collybita*), Short-toed Treecreeper (*Certhia brachydactyla*), Eurasian Nuthatch (*Sitta europaea*), European Robin (*Erithacus rubecula*), Eurasian Wren (*Troglodytes troglodytes*), and Great Spotted Woodpecker (*Dendrocopos major*) (30).

These bird species have home ranges that vary between 0.3 ha for the Blue Tit (31) and 7.9 ha for the Common Chiffchaff (32), the latter value being extreme as compared to most of the insectivorous species (33,34). It is therefore likely that multiple individuals of the same insectivorous bird species defended separate territories in the study area. Together with the fact that we distributed the experiment across the whole forest stand (see below) and that oak trees host a large density of herbivorous insects (N. Plat, A. Mrazova, and L. Schillé

personal observation), we doubt that birds learned to avoid corpses and models during the

experiment.

Study design

First, we haphazardly selected 12 pedunculate oaks (*Quercus robur*, the dominant

broadleaved species of the area) located at least 10m apart from each other. On one branch

accessible from the ground of each of these trees, we glued five plasticine models, and on

another branch as far as possible from the first one, we glued another set of five corpses of

Operophtera brumata larvae, previously killed by freezing at -80°C. We use this level of prey

density because (i) the natural density of caterpillars was high at this time of the year, and

(ii) to optimize the use of camera traps by ensuring an adequate number of statistical

replicates. The five caterpillars on each branch were spaced at least 5 cm apart to remain in

the focus of camera traps.

We used the lepidopteran species *Operophtera brumata* for experimentation, a widespread

and often abundant oak-defoliating moth. The caterpillars, whether models or corpses,

were directly glued to the branches with two glue drops. We used corpses for ethical

reasons, to avoid gluing living caterpillars.

We modeled artificial caterpillars with green plasticine (Staedtler brand, model 8421-5),

forming 1.5 × 0.3 cm cylindrical shapes and mimicking Lepidopteran larvae. They had the

same color as most plasticine models commonly used in studies of predation interactions in

ecology (15–17,23,35) and the same size as *Operophtera brumata* dead larvae used as

corpses.

Corpses darken when exposed in the field; after 48 hours, their color differs from that of

live individuals. We cannot exclude that their smell change too. For these reasons, we

exposed fresh models and corpses on a new set of 12 trees every two days. After 16 days,

this procedure resulted in a total of 480 model- and 480 corpse- caterpillars installed on 96

oaks.

Predation analysis

We took plasticine models collected in the field to the laboratory in plastic vials. Note that

none of the plasticine models disappeared during the experiment. All models were

examined by three experienced observers who identified marks of attempted predation by

birds using a magnifying glass. The models were considered 'attacked' if they exhibited at

least one attack mark on the plasticine surface. Observers carefully reread the bite mark

guide published by Low et al. (23) beforehand. These observations were used to evaluate

consistency among the observers. In cases of doubt regarding whether a model was

attacked or not, the observers discussed and reached a consensus. Hereafter, we refer to

the presence of marks reported by human observers' consensus as 'predation clues'.

Predation clues on corpses were directly examined in the field. A missing corpse or one

with a missing part was considered attacked, as is generally the case in the literature

(19,20,24).

Image analysis

Interactions between predators and corpses as well as models were recorded using camera

traps (Brand Ltl Acorn model Ltl-5210M). We installed two camera traps on the trunk of

each tree facing the two branches holding models and corpses, respectively. The camera

was positioned about 1m from the caterpillars to make it easier to focus (without blurring)

(Fig 1). The camera traps were active 24 hours a day for the 14 consecutive days of the

experiment. They were set to the highest sensitivity and automatically took bursts of three

photos when the passive infrared (PIR) sensors detected motion with a 0.5s trigger speed.

At each field visit, we changed the SD cards and checked the batteries to replace them if

necessary.

We analyzed the images taken by camera traps using the application EcoAssist (36)

supporting the MegaDetector learning model (Machine Learning for Wildlife Image

Classification, (37)). This learning model automatically detects the presence of animals in

images. We used the most conservative detection confidence threshold of 0.01 for the

model in the EcoAssist application, as this threshold previously tested showed a 100%

success rate (i.e., all birds present in the images were detected). Following this detection

process, we carried out a visual examination by an observer of the images. This examination

concerned both corpses and plasticine models.

Our camera traps were insufficient to monitor all predation events due to the birds' small

size and stealthy behavior. Therefore, we defined 'evidence of predatory bird presence' at

the branch level as the presence of at least one image showing a foraging bird on the branch

(hereafter 'evidence of bird presence'). These images could show individuals in various

contexts: birds looking at the camera (Fig 2d and 2e), flying over the target branch (Fig 2a),

engaging in foraging activity on the target branch but with natural prey on the bill (Fig 2c

and 2e), foraging on experimental prey (Fig 2f and 2h), standing on the experimental

branch (Fig 2b and 2g), or feeding during early morning (Fig 2e).

Subsequently, we statistically compared this evidence of bird presence to the predation clue

determined by human observers on the same branch. A branch was categorized as having a

predation clue if at least one of its five caterpillars showed a predation clue.

All observed evidence of predator presence, regardless of whether the predator was a bird

or not, was also carefully recorded for further discussion. It should be noted, however, that

arthropod predators were unlikely to be detected by camera traps. We identified all species

associated with the evidence of bird presence.

Fig 1. Photographs of the experimental design in the field.

Red arrows point toward the camera traps. Blue and yellow arrows point toward five

plasticine models (a) and three corpses (b), respectively.

Fig 2. Camera trap photographs of birds taken in various contexts.

Birds exhibit different behaviors (perched on a branch, actively foraging, flying, etc., see main text). The blue squares correspond to MegaDetector detections with the associated confidence level. The bird species included are: the Great Tit (Parus major, Figs 2a and 2f), the Long-tailed Tit (Aegithalos caudatus, Figs 2b and 2g), the European Robin (Erithacus rubecula, Fig 2c), the Eurasian Jay (Garrulus glandarius, Fig 2d), the European Pied Flycatcher (Ficedula hypoleuca, Fig 2e), and the Common Chiffchaff (Phylloscopus collybita, Fig 2h)

Statistical analysis

All analyses were performed in R (38). Fig 3 summarizes all the hypotheses tested using the experimental design.

Evaluation of the degree of consistency between observers in the posterior detection of bird predation clues (H1)

We relied on the intra-class correlation coefficient (ICC) to assess the degree of consistency between observers in detecting predation clues on plasticine models. The ICC quantifies the degree of agreement between groups or between observers and is bounded between 0 and 1, where 1 indicates perfect intra-group agreement and 0 indicates no agreement beyond what would be expected by chance. According to Koo and Li (39), an ICC value below 0.50 is considered poor, between 0.50 and 0.75 moderate, between 0.75 and 0.90 good, and above 0.90 excellent. We calculated the ICC using the '*icc*' function of the *irr* package (40), with a two-way random effect model because each plasticine model was evaluated by the same set of three observers.

 Comparison between evidence of bird presence assessed by camera traps and predation clues

detected by human observers branches with models and corpses (H2 and H4)

We used two generalized linear mixed models (GLMM, *lme4* package; (41)) with a binomial

error distribution to evaluate the bias between observer detection of predation clues and

evidence of bird presence assessed by camera traps on (i) branches with models (H2) and

on (ii) branches with corpses (H4). In both cases, the detection method (predation clues or

evidence of bird presence assessed by a camera) was set as a fixed effect, and tree identity

nested in temporal permutation was considered a random factor. The model equation was:

264
$$Y_{ij} = \beta_0 + \beta_1 \times Detection_{Clues,ij} + \gamma_j + \epsilon_{ij}$$

where Y_{ij} is the response variable (one data point per detection method and branch). Y_{ij}

could be zero (no predation clue on any model (H2) or corpse (H4) or no evidence of bird

presence) or one (at least one model (H2) or corpse (H4) with predation clue or evidence of

bird presence). β_0 is the model intercept (i.e., $Detection_{EvidenceBird,ij}$), β_1 the coefficient of

the fixed effect of the predation clues detected by the observer (i.e., $Detection_{Clues,ij}$), γ_j the

random intercept for tree identity nested in temporal permutation, and ϵ_{ij} the residuals.

Comparison between the evidence of bird presence on corpses versus models both assessed by

camera traps (H3)

We used another GLMM with a binomial distribution to assess the predation bias on models

compared to corpses. Fixed effect was the type of prey (model or corpse) and random

intercepts were tree identity nested in temporal permutation. The model equation was:

276
$$Y_{ij} = \beta_0 + \beta_1 \times PreyType_{Model,ij} + \gamma_j + \epsilon_{ij}$$

where Y_{ij} is the binomial response variable (presence or absence of evidence of bird

presence at the branch level). β_0 is the model intercept (i.e., $PreyType_{Corpse,ij}$), β_1 the

coefficient of the fixed effect of the plasticine models (i.e., $PreyType_{Model,ij}$), γ_j the random

intercept for tree identity nested in temporal permutation, and ϵ_{ij} the residuals.

For both types of GLMMs, we used the '*nlminbwrap*' optimizer, which facilitated model

convergence and provided results comparable to the default optimizer of the *lme4* package.

It should be noted that since the two GLMMs we used are binomial with a *logit* link function,

an inverse transformation of the coefficients must be applied to estimate it as a percentage

of predation.

Fig 3. The experimental design testing the four hypotheses.

H1: Evaluation of the degree of consistency between observers in the posterior detection of

bird predation clues; H2: Comparison between evidence of bird presence assessed by camera

 traps and predation clues detected by human observers branches with models; H3:

 Comparison between the evidence of bird presence on corpses versus models both assessed by

 camera traps; H4: Comparison between evidence of bird presence assessed by camera traps

 and predation clues detected by human observers branches with corpses.

Results

Consistency among observers in the detection of bird predation clues (H1)

Observer A detected ten predation clues on 480 plasticine models, whereas Observer B and Observer C detected only three and four predation clues, respectively. At first, the three of them only agreed on two caterpillars (Fig 4a). The intra-class correlation coefficient (ICC) score was 0.47 (CI 95%: 0.41 - 0.52) indicating a poor agreement between the three observers in their assessments of predation clues on the 480 plasticine models ($F_{(479, 960)}$ = 3.61, P < 0.001). The three observers therefore re-assessed predation clues to reach a consensus on the attacked vs non-attacked status. The final, consensual status resulting in seven caterpillars with bird predation clues was used in further analyses.

Evidence of bird presence on branches with corpses or models assessed by camera traps (H3)

We obtained 148,157 images from the cameras. The MegaDetector algorithm detected animal presence in 37,495 images (using the lowest detection threshold of 0.01 confidence). After visual evaluation, we ultimately retained only 146 photos showing foraging birds on or around 30 target branches (out of 192). They corresponded to 42 bird individuals belonging to 10 different species (*Aegithalos caudatus*, *Anthus trivialis*,

Phylloscopus collybita, *Erithacus rubecula*, *Garrulus glandarius*, *Ficedula hypoleuca*, *Cyanistes*

caerulus, *Parus major*, *Fringilla coelebs*, and *Turdus philomelos*). We also detected other

potential predators on certain branches, such as rodents of the genera *Mus* or *Eliomys* and a

Red Squirrel (*Sciurus vulgaris*) (see Figs S1m and z, respectively).

Evidence of bird presence, defined as the presence on images of a foraging bird on or flying

toward a branch, did not differ significantly between branches with plasticine models

vs. branches with corpses ($\beta_{1,Corpse} \pm$ SE = -0.5 $\pm$ 0.4, z = -1.3, P = 0.20, Fig 4b). A

substantial portion of the variance in bird presence assessment was attributed to random

effect (marginal R^2 = 0.019, conditional R^2 = 0.19)

Evidence of bird presence vs. predation clues on branches with models (H2) and corpses (H4)

We found predation clues on 7 plasticine models (7 branches, Fig 5) and 181 corpses (64

branches). Predation clues on branches with plasticine models were associated with the

presence of a bird on the same branches in the case of three branches. We observed the

presence of birds on 15 branches with no predation clues on models. Predation clues on

models were observed on four branches without the presence of birds being detected by

camera traps (Fig 4a). We therefore detected significantly more evidence of bird presence

than predation clues on branches with plasticine models ($\beta_{1,Evidence} \pm$ SE: 1.3 $\pm$ 0.5, z = 2.4,

P < 0.05, Fig 4c). A substantial portion of the variance in predation assessment on branches

with plasticine models was attributed to random effect (marginal R^2 = 0.08, conditional R^2 =

0.34).

Predation clues on branches with corpses were associated with evidence of bird presence

on the same branches in 8 cases. In contrast, we observed evidence of bird presence

without predation clues on corpses on 4 branches, and predation clues on corpses without

the presence of bird being detected on 56 branches (Fig 4a.). There was therefore

significantly less evidence of bird presence than predation clues on branches with corpses

($\beta_{1,Evidence} \pm$ SE: -2.7 $\pm$ 0.4, z = -6.8, P < 0.001, Fig 4d.). The random effect accounted for

only a small fraction of the variance (marginal R^2 = 0.35, conditional R^2 = 0.38).

Fig 4. Main results of the study.

Euler diagram of consistency between the three observers (a, upper panel), and Euler diagrams of the number of branches with predation clues and with evidence of bird presence for models and corpses (a, lower right and lower left panels, respectively). Results of the generalized mixed-effects models with binomial distribution for the three hypotheses tested: H2 (c), H3 (b), and H4 (d).

Fig 5. Photographs with mark details.
Seven caterpillars that observers considered to have bird predation clues.

Discussion

Lack of consistency in observations of predation clues on plasticine caterpillar models

Our study confirms the lack of consistency between observers' evaluation of predation

marks on plasticine models. Despite all trained observers re-reading the attack mark guide

(23), they did not detect the same number of predation clues. Similar results, even among

experienced scientists, were reported by Valdés-Correcher et al. (21) and Castagneyrol et al.

(42), using a more robust experimental design. In our case, the lack of consistency among

observers may have been aggravated by the small size of plasticine models mimicking

Operophtera brumata caterpillars (1.5 × 0.3 cm). These models were smaller than the

artificial prey commonly used in the scientific literature (3 cm on average across the five

following studies (16,17,19,23,24)), resulting in a perception of predation marks that

differs from what we typically observed on larger models.

To our knowledge, most studies using predation clues on plasticine models rely on a single

observer. Information about whether multiple observers could be involved is also rarely

provided (e.g., when the primary observer consults one or more colleagues for marks on

models that are difficult to assess). Some studies have employed two independent

observers evaluating the models by consensus (43,44), which could be a good method to

avoid observer bias. It could be also advisable to develop an online standardized training

system for identifying bird bill marks on plasticine models, as it is done for insect herbivory

(e.g., ZAX herbivory trainer: https://zaxherbivorytrainer.com/(45)). This could

complement the comprehensive guide developed by Low et al. (23) and be tailored to

different biomes, considering the substantial variations in predator and prey sizes and

behaviors.

Bird presence assessed by camera traps does not differ between branches with plasticine models

and corpses

The main result of our study revealed that the evidence of bird presence assessed by

camera traps does not significantly differ between experimental branches. Considering that

camera traps caught flying birds and birds foraging under poor light conditions (S1 Figure),

we assume that the cameras do not miss birds when they are present. Such an outcome

would suggest that plasticine models can be used to estimate bird predation, as they offer

the additional advantage of overcoming the ethical issues associated with the use of real

prey, whether dead or alive. This result differs from our findings regarding the number of

predation clues assessed by observers and contradicts other studies that find more bird

attacks on real prey compared to plasticine ones (19,20).

Specifically, we observed a ratio of 7 predation clues on plasticine models to 181 on

corpses. Similar to the findings by Zvereva & Kozlov (20), we believe that the corpses were

primarily removed and attacked by arthropods as we found ants crawling around 24

corpses out of 480, but other predators such as spiders or other arthropods could also be

involved. These findings are also in accordance with Nimalrathna et al. (24) who found that

the attack rate by vertebrate predators was lower on dead prey than on live prey and

plasticine models, while invertebrates predated most on dead prey followed by live prey

and plasticine models. Considering our and Nimalrathna et al.'s (24) results, we conclude

that our observations of predation on corpses are driven by arthropod consumers.

Although tempting to conclude that birds do not distinguish between real and plasticine

caterpillars, and thus claim this method reliable, it must be noted that we cannot rely on the

(i) posterior evaluation of clues on corpses; and (ii) the evidence of bird presence assessed

by camera traps as evidence of predation. Therefore, the evaluation of the plasticine model

method is mostly limited by available technology. To our knowledge, there is currently no

reliable, affordable technique for accurately identifying birds preying on insects. Camera

traps available on the market have motion detection speeds that are too slow for birds or

do not allow for fast sequences of pictures or continuous recording. Cameras that do offer

continuous recording have other limitations in data storage, power supply, and encoding

time. There is thus a need for advanced cameras, that allow us to observe fast behavioral

phenomena of relatively small animals, such as foraging birds. These cameras could be

used in various fields such as trophic interaction ecology, conservation ecology, and

behavioral ecology. Although some initiatives offering prototypes exist (e.g., (46)), the costs

are currently still too high to be affordable for scientific purposes.

The most limiting trade-off is between the camera performance and energy consumption.

For scientific research, we need at least 24 hours of endurance, which is currently not

possible for continuous recording. The cameras could therefore operate by capturing a

sequence of pictures at intervals of at least 0.5 seconds continuously to maximize the

chance of capturing the moment of predation (86,400 pictures/day). At the same time, the

batteries must be removable for quick changes in the field. Additionally, the cameras should

be affordable, relatively small, light, and waterproof, and finally have programmed a short

encoding time.

Predation clues assessed by observers do not correlate with bird presence assessed by camera

traps

Predation clues assessed by observers on plasticine models or corpses did not reflect bird

foraging activity as revealed by camera traps. Another study comparing camera traps and

posterior detection to assess predation on artificial snakes also revealed that predatory

birds were underrepresented in the videos triggered after motion detection compared to

the number of predation marks on artificial replicates (47). Moreover, O'Brien & Kinnaird

(48) demonstrated in 2008 that camera traps were more suitable for larger birds such as

pheasants than for functionally insectivorous birds targeted in our study. We acknowledge

that modern camera trap technology has significantly improved since 2008, though, to our

knowledge, however, no recent studies specifically address this in the context of bird

predation.

Contrary to our hypothesis, we found more evidence of bird presence than predation clues

on plasticine models suggesting that fake prey underestimates the foraging activity of avian

insectivores. Considering the particularly high density of live prey during this time of year

(L.S., N.P., A.M. pers. observation), an explanation could be that experienced birds prefer

live caterpillars found on the same branch over the models (see also (20)). It is, however,

important to consider the low numbers of both observations (clues vs. evidence) (Fig 4c)

implying a weak indicative value.

The glue used to attach the prey sometimes caused such small models to harden, potentially

preventing the later detection of predation clues. Finally, we could not exclude curiosity-

driven behaviors from some individuals, notably Eurasian jays, who seemed to gaze intently

at the camera rather than at the available prey. While we anticipated this curiosity

behavior, we expected it to manifest more in predation clues on the models (e.g., such as

claw marks). We carefully re-checked models on which no predation clues were initially

identified, whereas we obtained evidence of birds foraging on the corresponding branches.

The three observers confirmed no predation clue was visible on these models.

We observed more predation clues (181) than presence evidence on corpses (12). As

described above, we believe that the effect size is most likely driven by arthropod

consumers. This assumption, however, cannot be currently tested as we are not able to

distinguish between arthropod and bird clues on corpses and at the same time cannot rely

on the evidence of bird presence to be evidence of a genuine predation event. Additionally,

we note that photographs from the camera traps revealed potentially predatory rodents on

one of the branches with corpses.

Overall, the results discussed here provide only a limited window to the interpretation of

whether the use of plasticine models should be regarded as a reliable method. Therefore,

we encourage future protocols to explore methods that involve exposing live prey on

branches while non-invasively preventing their escape (e.g., applying glue to branch ends)

in combination with cameras providing unquestionable evidence of bird predation.

Perspectives

Our study still has not answered the most important question, namely whether plasticine

models accurately assess actual predation by birds or not. However, it paves the way for the

use of new technologies, such as passive monitoring methods like video surveillance, to

obtain real evidence of predation. We, therefore, advocate for their development and use,

which will likely refine the widely used plasticine model method in the ecology of trophic

interactions.

Taking into account all results and technical difficulties of this study as well as the results of

other studies, we summarized the advantages and disadvantages of each method for

quantifying bird predation, along with future recommendations in Table 1.

Table 1. *Advantages, disadvantages, comments, and recommendations for the three methods of assessing bird predation:*

plasticine models, real prey, and cameras. Note that cameras must be used in combination with one of the other two methods. If a

cell in the third column spans two rows in the first column labeled 'Methods,' it applies to both methods. Literature references

supporting our arguments are marked in blue.

Methods	Advantages	Disadvantages	Comments/ recommendations
Plasticine models	- Low-cost - Easy method - Standardized - Ethical - Posterior detection possible	- Artificiality: immobile, too bright color resulting in unnatural contrast with branch; non-natural smell; often oversized regarding the size of the species naturally occurring in the habitat - Inconsistency of human detection	- Most models used are too large compared to native European caterpillars *((14); see S2 Figure)*. Therefore, we recommend adjusting the models' size according to the species likely to be present at study sites. - The vivid green color is not always the most suitable choice for plasticine. We recommend adjusting the color of larvae in a given geography and habitat *(e.g., (49))*. - We recommend several independent observers *(e.g., (43,44))* or an online training system *(on the model of (45))* to mitigate the observer detection bias.
			- It is necessary to consider a realistic density of models concerning the actual abundance of prey in the study area at any given time *(14)*. - Consider exposing the prey (models or real caterpillars) according to the natural behavior of the species they represent (e.g., petioles of

			the leaves, inner space of the rolled leaves, tree trunks, etc.) *(e.g., (15,17,27))*.
			- Consider the appropriate attachment method regarding the size of the prey, (e.g., the glue may not be the best solution in the case of small prey and could be substituted with pins or double-sided adhesive tape) *(e.g., (15,17,27,50))*.
Real prey	- Really reflects predation	- Expensive and time-consuming rearing - Unethical	
			- Use live prey to better reflect predation. Develop protocols to utilize live prey without attachment for ethical reasons while preventing their escape from branches *(e.g., (19,20,51))*. - Combination of real prey and cameras.
Camera traps	- Theoretically allows obtaining evidence of predation by birds *(but this study showed that current camera traps do not yet allow this)* - Identification of bird species - Detection of other vertebrate predators - Knowing the exact time of predation	- Expensive *(52)* Currently: - Passive infrared sensors are not suitable for small birds *(48)* - Too slow motion detection speeds for photos and even slower for videos - Only a few photos in burst mode after motion detection decrease the chance of capturing the predation event - Too short video times - Non-existing long-term continuous recording - Low data storage - Short-term power supply	- We strongly encourage programmers and electronicians to use low-cost microcontrollers coupled with small cameras and high-capacity batteries to obtain real evidence of not only bird predation but generally animal behavior under natural conditions. Such a monitoring method could allow us to conduct reproducible experiments on a large geographic scale, in different habitats, and with multiple target larva species to model an overall bias of the plasticine model method.

		- Long encoding time - High power supply *(52)* - Need for artificial intelligence to process thousands of photos *(46)*	

Conclusion

Our study sheds new light on the evaluation of the plasticine model method for

characterizing bird predation. It confirms the existence of an inter-observer consistency

bias in the posterior evaluation of predation marks on these models. It also suggests that

predation clues do not accurately reflect the presence of predatory birds as assessed by

camera traps, neither on the branches bearing the plasticine models nor on those bearing

dead natural caterpillars. As we found no differences in the behavioral responses of birds to

artificial prey compared with dead natural ones, we would conclude that using caterpillar

corpses did not present particular advantages over using plasticine models for assessing

avian predation on folivores. However, taking into account predation clues analyzed by

observers showing contradictory results indicates a non-accurate monitoring approach for

assessing the reliability of this method. Existing technologies do not yet allow this method

to be improved and the use of plasticine to assess bird predation cannot be claimed or

refused using currently available techniques. Our study suggests it is necessary to be able to

evaluate the inherent biases of using plasticine models in field conditions. To this end, we

advocate for the development of more precise monitoring devices and more global

initiatives building upon our study, as soon as such a reliable monitoring method can be

offered.

Acknowledgments

This study was permitted by the financial support of the BNP Paribas Foundation through its Climate & Biodiversity Initiative for the 'Treebodyguards' citizen science project. A.M. is financially supported by the Czech Science Foundation (CSF) within the Postdoctoral Individual Fellowship - Outgoing Programme (no. 23-07045O). We thank Alba Lazaro-Gonzalez, Heidy Schimann, Inge Van Halder, Tom Barlier, Thomas Ribot, and Thibaud Coupart for their help in the field.

Data availability plan

The R Markdown format will be used to ensure that the statistical analyses are fully reproducible, with the code and data made publicly available (https://recherche.data.gouv.fr/) upon acceptance of the study, following the FAIR principles for scientific data management.

References

1.   Díaz-Siefer P, Olmos-Moya N, Fontúrbel FE, Lavandero B, Pozo RA, Celis-Diez JL. Bird-mediated effects of pest control services on crop productivity: a global synthesis. J Pest Sci. mars 2022;95(2):567-76.

2.   Remmel T, Davison J, Tammaru T. Quantifying predation on folivorous insect larvae: the perspective of life-history evolution. Biol J Linn Soc. sept 2011;104(1):1-18.

3.   Ford AT, Goheen JR. Trophic Cascades by Large Carnivores: A Case for Strong Inference and Mechanism. Trends Ecol Evol. déc 2015;30(12):725-35.

4.   Nyffeler M, Şekercioğlu ÇH, Whelan CJ. Insectivorous birds consume an estimated
400–500 million tons of prey annually. Sci Nat. août 2018;105(7-8):47.

5.   Sherry TW, Kent CM, Sánchez NV, Şekercioğlu ÇH. Insectivorous birds in the
Neotropics: Ecological radiations, specialization, and coexistence in species-rich
communities. The Auk. 24 déc 2020;137(4):ukaa049.

6.   Thiollay JM. Comparative Foraging Success of Insectivorous Birds in Tropical and
Temperate Forests: Ecological Implications. Oikos. juill 1988;53(1):17.

7.   Collins CT, Watson A. Field Observations of Bird Predation on Neotropical Moths.
Biotropica. mars 1983;15(1):53.

8.   Mäntylä E, Kleier S, Lindstedt C, Kipper S, Hilker M. Insectivorous Birds Are Attracted
by Plant Traits Induced by Insect Egg Deposition. J Chem Ecol. déc 2018;44(12):1127-38.

9.   Poloni R, Dhennin M, Mappes J, Joron M, Nokelainen O. Exploring polymorphism in a
palatable prey: predation risk and frequency dependence in relation to distinct levels of
conspicuousness. Evol Lett. 11 janv 2024;qrad071.

10.   van Bael SA, Philpott SM, Greenberg R, Bichier P, Barber NA, Mooney KA, et al. Birds
as predators in tropical agroforestry systems. Ecology. avr 2008;89(4):928-34.

11.   Michalski M, Nadolski J, Marciniak B, Loga B, Bańbura J. Faecal Analysis as a Method
of Nestling Diet Determination in Insectivorous Birds: A Case Study in Blue Tits *Cyanistes*
caeruleus and Great Tits *Parus major*. Acta Ornithol. déc 2011;46(2):164-72.

12.   Sam K, Koane B, Jeppy S, Sykorova J, Novotny V. Diet of land birds along an
elevational gradient in Papua New Guinea. Sci Rep. 9 mars 2017;7(1):44018.

13.   Brodie III ED. Differential avoidance of coral snake banded patterns by free-ranging
avian predators in Costa Rica. Evolution. 1993;47(1):227-35.

14.   Lövei GL, Ferrante M. A review of the sentinel prey method as a way of quantifying
invertebrate predation under field conditions: Measuring predation pressure by sentinel
prey. Insect Sci. août 2017;24(4):528-42.

15.   Roslin T, Hardwick B, Novotny V, Petry WK, Andrew NR, Asmus A, et al. Higher
predation risk for insect prey at low latitudes and elevations. Science. 19 mai
2017;356(6339):742-4.

16.   Schillé L, Valdés-Correcher E, Archaux F, Bălăcenoiu F, Bjørn MC, Bogdziewicz M, et
al. Decomposing drivers in avian insectivory: Large-scale effects of climate, habitat and bird
diversity. J Biogeogr. 4 févr 2024;jbi.14808.

17.   Valdés-Correcher E, Moreira X, Augusto L, Barbaro L, Bouget C, Bouriaud O, et al.
Search for top-down and bottom-up drivers of latitudinal trends in insect herbivory in oak
trees in Europe. Keith S, éditeur. Glob Ecol Biogeogr. mars 2021;30(3):651-65.

18.   Bateman PW, Fleming PA, Wolfe AK. A different kind of ecological modelling: the use
of clay model organisms to explore predator–prey interactions in vertebrates. J Zool. avr
2017;301(4):251-62.

19.   Rodriguez-Campbell A, Rahn O, Chiuffo MC, Hargreaves AL. Clay larvae do not
accurately measure biogeographic patterns in predation. J Biogeogr. 2023;

20.   Zvereva EL, Kozlov MV. Predation risk estimated on live and artificial insect prey
follows different patterns. Ecology. mars 2023;104(3):e3943.

21.   Valdés-Correcher E, Mäntylä E, Barbaro L, Damestoy T, Sam K, Castagneyrol B.
Following the track: accuracy and reproducibility of predation assessment on artificial
caterpillars. Entomol Exp Appl. 2022;170(10):914-21.

22.   Rößler DC, Pröhl H, Lötters S. The future of clay model studies. BMC Zool. déc
2018;3(1):6.

23.   Low PA, Sam K, McArthur C, Posa MRC, Hochuli DF. Determining predator identity
from attack marks left in model caterpillars: guidelines for best practice. Entomol Exp Appl.
août 2014;152(2):120-6.

24.   Nimalrathna TS, Solina ID, Mon AM, Pomoim N, Bhadra S, Zvereva EL, et al.
Estimating predation pressure in ecological studies: controlling bias imposed by using
sentinel plasticine prey. Entomol Exp Appl. janv 2023;171(1):56-67.

25.   Valdés-Correcher E, van Halder I, Barbaro L, Castagneyrol B, Hampe A. Insect
herbivory and avian insectivory in novel native oak forests: Divergent effects of stand size
and connectivity. For Ecol Manag. août 2019;445:146-53.

26.   Muchula K, Xie G, Gurr GM. Ambient temperature affects the utility of plasticine
caterpillar models as a tool to measure activity of predators across latitudinal and
elevational gradients. Biol Control. févr 2019;129:12-7.

27.   Mrazova A, Houska Tahadlová M, Řehová V, Sam K. The specificity of induced
chemical defence of two oak species affects differently arthropod herbivores and arthropod
and bird predation. Arthropod-Plant Interact. avr 2023;17(2):141-55.

28.   Campos RC, Steiner J, Zillikens A. Bird and mammal frugivores of *Euterpe edulis* at
Santa Catarina island monitored by camera traps. Stud Neotropical Fauna Environ. août
2012;47(2):105-10.

29.   Zhu C, Li W, Gregory T, Wang D, Ren P, Zeng D, et al. Arboreal camera trapping: a
reliable tool to monitor plant-frugivore interactions in the trees on large scales. Remote
Sens Ecol Conserv. 2022;8(1):92-104.

30.   Schille L, Valdés-Correcher E, Archaux F, Bălăcenoiu F, Bjørn MC, Bogdziewicz M, et
al. Data and codes for the article « Decomposing drivers in avian insectivory: large-scale
effects of climate, habitat and bird diversity » [Internet]. Recherche Data Gouv; 2024 [cité 2

févr 2024]. Disponible sur:
https://entrepot.recherche.data.gouv.fr/citation?persistentId=doi:10.57745/0E0JEA

31.   Naef-Daenzer B. Radiotracking of great and blue tits: new tools to assess
territoriality, home-range use and resource distribution. 1994;

32.   Lerche-Jørgensen M, Mallord JW, Willemoes M, Orsman CJ, Roberts JT, Skeen RQ, et
al. Spatial behavior and habitat use in widely separated breeding and wintering
distributions across three species of long-distance migrant *Phylloscopus* warblers. Ecol Evol.
juin 2019;9(11):6492-500.

33.   Cornell Laboratory of Ornithology. Birds of the World. In: Billerman SM, Keeney BK,
Rodewald PG, Schulenberg TS, éditeurs. Cornell Laboratory of Ornithology, Ithaca, NY, USA
[Internet]. 2022. Disponible sur: https://birdsoftheworld.org/bow/home

34.   Naguib M, Titulaer M, Waas JR, Van Oers K, Sprau P, Snijders L. Prior territorial
responses and home range size predict territory defense in radio-tagged great tits. Behav
Ecol Sociobiol. mars 2022;76(3):35.

35.   Sam K, Koane B, Novotny V. Herbivore damage increases avian and ant predation of
caterpillars on trees along a complete elevational forest gradient in Papua New Guinea.
Ecography. 2015;38(3):293-300.

36.   van Lunteren P. EcoAssist: A no-code platform to train and deploy custom YOLOv5
object detection models. J Open Source Softw. 2023;8(88):5581.

37.   Beery S, Morris D, Yang S. Efficient Pipeline for Camera Trap Image Review
[Internet]. arXiv; 2019 [cité 16 mars 2024]. Disponible sur:
http://arxiv.org/abs/1907.06772

38.   R Core Team. R: A Language and environment for statistical computing. [Internet]. R
Foundation for Statistical Computing, Vienna, Austria; 2021. Disponible sur:
https://www.r-project.org/

39.   Koo TK, Li MY. A Guideline of Selecting and Reporting Intraclass Correlation
Coefficients for Reliability Research. J Chiropr Med. juin 2016;15(2):155-63.

40.   Gamer M, Lemon J, Singh IFP. irr: Various Coefficients of Interrater Reliability and
Agreement [Internet]. 2019 [cité 17 mars 2024]. Disponible sur: https://cran.r-
project.org/web/packages/irr/index.html

41.   Bates D, Mächler M, Bolker B, Walker S. Fitting Linear Mixed-Effects Models Using
lme4. J Stat Softw. 2015;67(1):5-8.

42.   Castagneyrol B, Valdés-Correcher E, Bourdin A, Barbaro L, Bouriaud O, Branco M, et
al. Can School Children Support Ecological Research? Lessons from the *Oak Bodyguard*
Citizen Science Project. Citiz Sci Theory Pract. 18 mars 2020;5(1):10.

43.   Czarnecki C, Manderino R, Parry D. Reduced avian predation on an ultraviolet-
fluorescing caterpillar model. Can Entomol. 2022;154(1):e10.

44.   Franco JC, Branco M, Conde S, Garcia A, Fernandes MR, Lima Santos J, et al. Ecological
Infrastructures May Enhance Lepidopterans Predation in Irrigated Mediterranean
Farmland, Depending on Their Typology and the Predator Guild. Sustainability. 25 mars
2022;14(7):3874.

45.   Xirocostas ZA, Debono SA, Slavich E, Moles AT. The ZAX Herbivory Trainer—Free
software for training researchers to visually estimate leaf damage. Methods Ecol Evol.
2022;13(3):596-602.

46.   Darras KFA, Balle M, Xu W, Yan Y, Zakka VG, Toledo-Hernández M, et al. Eyes on
nature: Embedded vision cameras for multidisciplinary biodiversity monitoring [Internet].
2023 [cité 19 juin 2024]. Disponible sur:
http://biorxiv.org/lookup/doi/10.1101/2023.07.26.550656

47.   Akcali CK, Adán Pérez-Mendoza H, Salazar-Valenzuela D, Kikuchi DW, Guayasamin
JM, Pfennig DW. Evaluating the utility of camera traps in field studies of predation. PeerJ. 25
févr 2019;7:e6487.

48.   O'Brien TG, Kinnaird MF. A picture is worth a thousand words: the application of
camera trapping to the study of birds. Bird Conserv Int. sept 2008;18(S1):S144-62.

49.   Zvereva EL, Castagneyrol B, Cornelissen T, Forsman A, Hernández-Agüero JA,
Klemola T, et al. Opposite latitudinal patterns for bird and arthropod predation revealed in
experiments with differently colored artificial prey. Ecol Evol. déc 2019;9(24):14273-85.

50.   Richards LA, Coley PD. Seasonal and habitat differences affect the impact of food and
predation on herbivores: a comparison between gaps and understory of a tropical forest.
Oikos. janv 2007;116(1):31-40.

51.   Sam K, Jorge LR, Koane B, Amick PK, Sivault E. Vertebrates, but not ants, protect
rainforest from herbivorous insects across elevations in Papua New Guinea. J Biogeogr. 19
juill 2023;jbi.14686.

52.   Glover-Kapfer P, Soto-Navarro CA, Wearn OR. Camera-trapping version 3.0: current
constraints and future priorities for development. Remote Sens Ecol Conserv.
2019;5(3):209-23.