Testing for the genomic footprint of conflict between life stages in an angiosperm

and a moss species

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Abstract

The maintenance of genetic variation by balancing selection is of considerable interest to

evolutionary biologists. An important but understudied potential driver of balancing selection is

antagonistic pleiotropy between diploid and haploid stages of the plant life cycle. Despite sharing

a common genome, sporophytes (2n) and gametophytes (n) may undergo differential or even

opposing selection. Theoretical work suggests the antagonistic pleiotropy between life stages can

generate balancing selection and maintain genetic variation. Despite the potential for far-reaching

consequences of gametophytic selection, empirical tests of its pleiotropic effects (neutral,

synergistic, or antagonistic) on sporophytes are generally lacking. Here, we examined the

population genomic signals of selection across life stages in the angiosperm *Rumex hastatulus*

and the moss *Ceratodon purpureus*. We compared gene expression among life stages and

between sexes, combined with neutral diversity statistics and the analysis of the distribution of

fitness effects. In contrast to what would be predicted under balancing selection due to

antagonistic pleiotropy, we found that unbiased genes between life stages are under stronger

purifying selection, likely explained by a predominance of synergistic pleiotropy between life

stages and strong purifying selection on broadly expressed genes. In addition, we found that 30%

of candidate genes under balancing selection in *R. hastatulus* are located within inversion

polymorphisms. Our findings provide novel insights into the genome-wide characteristics and

consequences of plant gametophytic selection.

Keywords: balancing selection, antagonistic pleiotropy, intralocus conflict, gene expression,

gametophytic selection

Significance

The genome-wide importance of balancing selection has been a key question in evolutionary

biology. Despite theoretical support for the potential of balancing selection generated by

antagonistic pleiotropy between plant life stages, no empirical studies have investigated this

problem. Our results revealed genome-wide patterns more consistent with synergistic pleiotropy,

with gametophyte-biased genes mostly likely to be under relaxed purifying selection in both

angiosperm and moss species we investigated. In addition, our study suggests gene expression

level and breadth has a greater effect on selection efficacy than differences between ploidy levels

in different life stages.

Introduction

Understanding how genetic variation is maintained for traits under selection remains a key open question in evolutionary biology. Balancing selection maintains genetic variation in a population when different alleles are favored in different contexts, e.g., antagonistic selection between sexes or life-history, hereafter life stages, (Kidwell et al. 1977; Immler et al. 2012). Despite persistent theoretical and empirical interests, we still lack empirical tests of the genome-wide importance of balancing selection. The increasing availability of genomic data helps us to understand how different forms of balancing selection shape genetic variation, and also to identify candidate regions under balancing selection (Bitarello et al. 2023). Here, we focus on testing for genome-wide signals of balancing selection generated by antagonistic pleiotropy between life stages using transcriptomic and population genomic data of the angiosperm *Rumex hastatulus* and the moss *Ceratodon purpureus*.

The alternation of life cycles between diploid and haploid phases in sexually reproducing eukaryotes allows for natural selection to occur during both phases (Mable and Otto 1998). In land plants, the haploid phase (gametophyte) and diploid phase (sporophyte) have varying degrees of complexity in the different major clades of land plants. For example, angiosperms generally have the most elaborated sporophytes and the most reduced gametophytes; their gametophytes (i.e., ovules and pollen) are smaller and ephemeral, have fewer cells, and are dependent on sporophytes. Contrastingly, the haploid gametophytes in bryophytes (mosses, liverworts, and hornworts) are multicellular and often perennial. Differential or even opposing selection can result from the differences in genetic, cellular and organismal levels between these two life stages, creating genomic conflicts (Qiu et al. 2012; reviewed in Immler 2019). How

selection on gametophytes (gametophytic selection) affect sporophytes is a key unresolved

question in plant evolutionary genetics.

The evolutionary consequences of gametophytic selection have been examined both theoretically

and empirically (Haldane 1932; Otto et al. 2015; Immler and Otto 2018; summarized in Immler

2019, Beaudry et al. 2020). The haploid nature of the gametophytic phase is predicted to increase

the efficacy of selection due to the lack of dominance (i.e., the masking hypothesis), allowing for

more efficient removal of recessive deleterious mutations (i.e., purging) and fixation of recessive

beneficial mutations (Crow and Kimura 1965; Kondrashov and Crow 1991; Gerstein and Otto

2009). Consistent with this prediction, some studies have found stronger purifying and positive

selection in gametophyte-specific than sporophyte-specific genes (Arunkumar et al. 2013;

Cervantes 2023; but see Harrison et al. 2017). Stronger selection on gametophyte-expressed

genes can also be driven by gametophytic competition (sexual selection) in angiosperms

(Arunkumar et al. 2013; Gossmann et al. 2014; Gutiérrez-Valencia et al. 2022). Additionally,

gene expression breadth can be a stronger predictor of selection efficacy than the masking effect

in angiosperms and bryophytes (Szövényi et al. 2013).

The common genome shared between life stages provides an opportunity to test how

gametophytic selection affects the evolution of the sporophyte stage due to the pleiotropy

between life stages. Plants share a substantial overlap in gene expression between life stages

based on both microarray and mRNA sequencing studies (summarized in Beaudry et al. 2020). In

ferns and bryophytes where the gametophytes are photosynthetic and exhibit indeterminate

growth, gene expression overlap may exceed that in angiosperms, e.g., estimates are ~60% in the

angiosperm *Arabidopsis thaliana* (Walbot and Evans 2003; Borg et al. 2009), 97.7% in the fern

Polypodium amorphum (Sigel et al. 2018), and 85% in the bryophyte *Physcomitrium patens*

(Ortiz-Ramírez et al. 2016). Previous studies on gametophytic selection have compared selective

forces in gametophyte -*vs.*- sporophyte-specific genes to test for the effects of haploid expression

on the distribution of selection coefficients (Arunkumar et al. 2013; Gossmann et al. 2014;

Harrison et al. 2019; Gutiérrez-Valencia et al. 2021; Cervantes et al. 2023). However, the

patterns of genetic diversity and selection pressures of genes with overlapping expression, which

are more likely to be pleiotropic due to the greater potential for competing selection pressures

across life stages (Immler et al. 2012; Peters and Weis 2018), have not yet been specifically

examined.

In theory, antagonistic pleiotropy between life stages, i.e., intralocus conflict when alleles of a

gene have opposite fitness effects between life stages, can cause balancing selection and maintain

genetic variation under some conditions (Immler et al. 2012; Peters and Weis 2018). Despite the

theoretical support, empirical evidence of antagonistic pleiotropy in angiosperms is limited to a

few studies suggesting sexual conflict across life stages, e.g., between male gametophyte and

female sporophyte during pollen competition (e.g., Travers and Mazer 2001; Lankinen and Kiboi

2007), and between female gametophyte and male sporophyte in a female meiotic drive system

(Fishman and Saunders 2008). In bryophytes where sporophytes are dependent only on female

gametophytes, parent offspring conflict may occur (Haig and Wilczek 2006; Johnson and Shaw

2016; Shortlidge et al. 2021), but the mechanism and genetic architecture underlying any such

conflict remain unclear. Testing the genome-wide prevalence of antagonistic pleiotropy between

life stages and its potential to maintain genetic variation is therefore key to understanding the

genetic architecture of conflict between plant life stages.

Given the potential for antagonistic pleiotropy between the life stages for genes with overlapping

expression, we predicted that if these effects are widespread, we should see population genetic

signals of balancing selection more commonly in genes with overlapping expression (Immler et

al. 2012; Peters and Weis 2018). An analogous finding is that genes with intermediate rather than

extreme sex bias between sexes show patterns of genetic diversity consistent with balancing

selection, as expected if extreme expression bias reflects resolved rather than ongoing conflict

(e.g., Cheng and Kirkpatrick 2016; Wright et al. 2018; Sayadi et al. 2019; summarized in Ruzika

et al. 2020). Similarly, in the case of intralocus conflict between life stages, we predict stronger

signals of balancing selection in genes with overlapping expression between life stages,

particularly those with relatively similar levels of expression between the two life stages (Immler

et al. 2012; Peters and Weis 2018), reflected by diversity statistics such as nucleotide diversity

(π) and Tajima's D. In addition, we also predicted that candidate genes under balancing selection

would be disproportionately biased in expression between life stages rather than being unbiased.

We examined gene expression and genetic diversity patterns in the gametophytic and sporophytic

life stages of two plant species: the angiosperm *Rumex hastatulus* (Polygonaceae) and the

bryophyte *Ceratodon purpureus* (Ditrichaceae). The annual *R. hastatulus* is dioecious, obligately

outcrossing, and wind-pollinated with an XY sex chromosome system (Smith 1964; Rifkin et al.

2021). The uniovulate flowers and the large quantity of pollen commonly produced in wind-

pollinated angiosperms suggests high potential for gametophytic selection during pollen

competition (Friedman and Barrett 2009; Field et al. 2012; Immler 2019). In the dioicous moss

C. purpureus, the female and male gametophytes possess the U or V sex chromosome,

respectively (Carey et al. 2021a), they produce gametes that fuse to form a diploid zygote (i.e.,

sporophyte, unsexed); the sporophyte matures and ultimately undergoes meiosis, but its entire

development occurs on the female gametophyte plant, providing multiple opportunities for

antagonistic pleiotropy (Carey et al. 2021b; McDaniel 2023). The contrasts in the length and

complexity of the gametophytic stage in these two species allow us to evaluate the evolutionary

consequences of biphasic plant life cycles in a broader context.

We first compared gene expression in gametophytic and sporophytic tissues of *R. hastatulus* and

C. purpureus, finding a larger overlap in gene expression and a smaller proportion of biased

genes between life stages in *C. purpureus* than *R. hastatulus*. We then examined the population

genetic signals of balancing selection due to antagonistic pleiotropy between life stages by

examining nucleotide diversity and Tajima's *D* across varying degrees of expression bias

between life stages. In contrast with our predictions for genome-wide patterns of antagonistic

pleiotropy, our results show elevated nucleotide diversity and Tajima's *D* in highly gametophyte-

biased genes in both *R. hastatulus* and *C. purpureus*. We found that expression levels and breadth

were a stronger predictor of selection efficacy than potential competing selective pressures

between life stages. Lastly, we identified in both species genomic regions containing hundreds of

candidate genes under balancing selection using a composite likelihood ratio test (Cheng and

DeGiorgio 2019, 2020, 2022), and examined the biological functions and differential gene

expression patterns of these candidate genes.

Results

Gene expression in gametophytes and sporophytes

We compared gene expression in gametophyte and sporophyte tissues of *R. hastatulus* and *C.*

purpureus. In *R. hastatulus*, we sequenced 75 mature pollen samples and 77 male leaf samples

representing the gametophyte and sporophyte stages, respectively (Table S1). In *C. purpureus*,

we compared expression between 21 sporophyte and 36 gametophyte samples (whole plant,

described in Carey et al. 2021a, Table S3, see Materials and Methods). In both species, there is a

large overlap in gene expression between the life stages (Table 1). In *R. hastatulus*, we identified

15,958 gametophyte-expressed genes, consisting of 52% of all annotated genes across the

genome, and 89% of genes that showed evidence of expression. In *C. purpureus*, we identified

24,097 gametophyte-expressed genes, consisting of 69% of all annotated genes and 93% of genes

with evidence of expression. The overlap in gene expression between life stages (89% in *R.*

hastatulus, and 93% in *C. purpureus*) sets up a strong potential for pleiotropic effects of

mutations between life stages.

We first identified genes that were differentially expressed (DE) between life stages and

exclusively expressed in one life stage (Table S4). There was a slightly higher percentage of

overlapping gene expression and a lower percentage of differentially expressed genes between

life stages in *C. purpureus* than *R. hastatulus* (Table 1). In *R. hastatulus*, we identified 5,635

gametophyte-biased and 8,653 sporophyte-biased genes; similarly in *C. purpureus*, there were

6,446 gametophyte-biased and 5,202 sporophyte-biased genes (padj < 0.1, FC > 2). Among

significant DE genes, we filtered for genes in the top 10% quantile of expression bias, allowing

us to compare genes with different tissue specificities. Our gene filtration process resulted in

similar numbers of gametophyte- and sporophyte-specific genes: 1,711 gametophyte-specific and

1,711 sporophyte-specific genes in *R. hastatulus*, and 1,886 gametophyte-specific and 1,867

sporophyte-specific genes in *C. purpureus*. Gene ontology (GO) enrichment analyses indicated

very different functional enrichment for biased and specific genes between life stages (Table S5,

S6), highlighting distinctive expression profiles across life stages in both species. For example, in

R. hastatulus, gametophyte-specific genes had protein phosphorylation functions (GO:0006468)

which are important during pollen-tube growth (Klodová and Fíla 2021) and sporophyte-specific

genes showed photosynthetic functions (GO:0015979).

Table 1. Summary of samples used and gene expression analyses in *R. hastatulus* and *C.*

purpureus. P-adj: Benjamini-Hochberg adjusted *p*-value with a false discovery rate of 0.1, FC:

fold change. All percentages are based on comparison to the number of expressed genes.

	Genome size	Sample size of RNA samples	Number of annotated genes	Number of expressed genes	Number of gametophyte-expressed genes	Number of biased genes (padj < 0.1, FC > 2)
Rumex hastatulus	1.7 Gb (2n)	75 (n), 77 (2n)	30,641 (2n)	17,984	15,958 (89%)	5,635 (n, 31%) 8,653 (2n, 48%)
Ceratodon purpureus	358 Mb (n)	36 (n), 21 (2n)	34,392 (2n)	25,880	24,097 (93%)	6,446 (n, 25%) 5,202 (2n, 20%)

Nucleotide diversity and expression bias between life stages

Population genomic analyses can be used to provide an indirect test for pleiotropic effects across

life stages. We calculated nucleotide diversity at 4-fold (π_s) and 0-fold (π_n) sites for each gene

and looked at the linear relation between π_s or π_n and the direction and degree of expression bias

between life stages (log2FoldChange) across genes. The linear regression of π_s or π_n on the

direction and degree of expression bias between life stages (log2FoldChange) showed a

significant and positive relation, but with near-zero slopes in both species (Figure S1, S2). We

next divided the genes into eight bins based on the direction and degree of expression bias

between life stages and calculated the weighted mean of π_s and π_n for each bin, weighted by the

number of sites per gene. In both species, highly gametophyte-biased genes exhibited the highest

levels of π_s, π_n and π_n/π_s ratio (Figure 1, Table S7). This result contrasts with our expectation that

genes with overlapping and intermediate bias in expression should show the highest diversity

under a model of widespread antagonistic pleiotropy. In both species, π_s and π_n increase with the

degree of expression bias between life stages in both directions (less so in *R. hastatulus*), but

values of π_s and π_n were greatest for the highly gametophytic-biased genes (Figure 1). In *R.*

hastatulus, the increase in π_n with the degree of expression bias was less symmetrical than in π_s,

with sporophyte-biased genes having lower π_n than genes with overlapping expression (Figure

1a, 1b). Our results are robust to our choice of weighting π_s and π_n by the number of sites per

gene, as when we estimated the mean and standard errors without weighting by the number of

sites, the patterns remain the same (Figure S3).

There are two possible explanations for the patterns of elevated π_s and π_n in the most

gametophyte-biased genes, a greater prevalence of balancing selection under the assumption of

neutrality of synonymous mutations or relaxed purifying selection on this set of genes. If there is

relaxed purifying selection on gametophytic-biased genes, this could drive higher π_n and higher

π_n/π_s. The observed π_n/π_s ratios showed a similar trend as π_s and π_n, especially in *C. purpureus*,

with highly gametophyte-biased genes having the highest values (Figure 1c, 1f), suggesting

either balancing selection, or a reduction in the strength of purifying selection on the most

gametophyte-biased genes.

Figure 1. Effect of expression bias between life stages on nucleotide diversity in *R. hastatulus* (a-c) and *C. purpureus* (d-f). x-axis: quantiles of log2FoldChange between gametophyte and sporophyte expression. Number of genes in each bin: 1,009-1,847 for *R. hastatulus*; 1,943-2,564 for *C. purpureus*. The weighted mean and 95% confidence intervals are based on 1000 bootstraps of the original dataset. Note that the scales on the y-axis are different between species.

To further distinguish between the two alternative explanations for increased nucleotide diversity in gametophyte biased genes, we examined diversity patterns at genes specifically expressed in one of the two life stages, since these genes should experience the least pleiotropy across stages with reduced expression breadth. We compared π_s, π_n and π_n/π_s ratio for gametophyte-specific, sporophyte-specific and unbiased genes (Figure 2, S4; Table S8). The results are mostly consistent with Figure 1, with gametophyte-specific genes having a higher level of π_s and π_n than

sporophyte-specific and unbiased genes in both species (Figure 2a, 2b, 2d, 2e), although the π_n/π_s

ratios are not elevated above unbiased genes in *R. hastatulus* like *C. purpureus* (Figure 2c, 2f).

Figure 2. Weighted mean nucleotide diversity of gametophyte-specific, sporophyte-specific,

unbiased genes in *R. hastatulus* (a-c) and *C. purpureus* (d-f). Number of genes in each category:

1149 (s), 920 (g), 523 (unbiased) for *R. hastatulus*; 1,232 (s), 1,217 (g), 3,309 (unbiased) for *C.*

purpureus. The weighted mean and 95% confidence intervals are based on 1000 bootstraps of the

original dataset. Note that the scales on the y-axis are different between species.

One possible additional factor that could contribute to differences in the strength of purifying

selection is if these genes differ in expression level. Gene expression has been shown to be a

strong predictor of the strength of purifying selection genome-wide in many species (e.g., Urrutia

and Hurst 2003; Slotte et al. 2011; Zhang and Yang 2015). We used this relation to further

examine the possible effects of relaxed purifying selection on π_s, and divided the genes into ten

bins of expression level (baseMean across all samples) to examine how expression level affects

π_s, π_n and π_n/π_s in gametophyte- and sporophyte-biased genes. As expression level increases, both

π_n and π_s decrease, with a bigger decrease in π_n than π_s (Figure 3). Gametophyte-biased genes

have higher π_s, π_n and π_n/π_s than sporophyte-biased genes, regardless of expression level in both

species, except for the bins with overlapping confidence intervals in π_s of *R. hastatulus* (Figure

3). Our results suggest that expression level has a strong effect on patterns of diversity. Overall,

contrary to our expectations, genes with overlapping expression levels do not show signals of

elevated diversity due to balancing selection. Instead, heterogeneity in diversity likely reflects

differences in the strength of purifying selection.

Figure 3. Effect of expression level on weighted mean nucleotide diversity in *R. hastatulus* (a-c)

and *C. purpureus* (d-f). Number of genes in each bin: 686-1,464 for *R. hastatulus*; 1,034-2,328

for *C. purpureus*. The weighted mean and 95% confidence intervals are based on 500 bootstraps

of the original dataset. qt1: lowest expression, qt10: highest expression. Note that the scales on

the y-axis are different between species.

Tajima's D and expression bias between life stages

Tajima's D is a metric summarizing information of the distribution of allele frequencies in a

population (Tajima 1989) and is influenced by both selection and demographic history. Positive

values are associated with balancing selection or population contraction and negative values

indicate purifying selection or population expansion. We further tested the effect of expression

bias between life stages on Tajima's D at synonymous (D_s) and nonsynonymous (D_n) sites

(Figure 4). In both species, D_n was smaller than D_s, consistent with purifying selection on

nonsynonymous sites (Figure 4a v.s. 4b, 4c v.s. 4d). In *R. hastatulus*, both D_n and D_s were

negative, D_s was higher in gametophyte-biased genes than unbiased and sporophyte-biased genes,

as expected if they had a higher proportion of genes under balancing selection (Figure 4a). The

negative values in D_n and D_s in *R. hastatulus* may possibly be attributed to population expansion,

but we currently have no evidence on demographic history. In *C. purpureus*, the patterns of D_n

were more similar to π_s and π_n than D_s, both D_s and D_n were higher in highly gametophyte-biased

genes than unbiased genes (Figure 4c, 4d). The higher and more positive Tajima's D values in *C.*

purpureus compared to *R. hastatulus* might be due to spatially variable selection on

gametophytes in *C. purpureus* with a global sampling of populations from different habitats. We

also compared D_s and D_n in gametophyte- and sporophyte-biased genes across expression level

quantiles. In *R. hastatulus*, D_s and D_n remained roughly constant across expression level

quantiles, whereas in *C. purpureus*, D_s and D_n decreased with higher expression (Figure S5).

Gametophyte- and sporophyte-biased genes had similar levels of D_s and D_n across expression

levels in both species. Combined with patterns of π (Figure 1-3) our results are consistent with

the hypothesis that gametophyte-biased genes are under relaxed purifying selection.

Figure 4. Effect of expression bias between life stages on Tajima's D in *R. hastatulus* (a, b) and

C. purpureus (c, d). x-axis: quantiles of log2FoldChange between gametophyte and sporophyte

expression. Number of genes in each bin: 1,009-1,734 (D_s), 1,197-1,966 (D_n) for *R. hastatulus*;

2,138-2,534 (D_s), 2,097-2,579 (D_n) for *C. purpureus*. Error bars represent mean ± SEM across

genes in each bin. Note that the scales on the y-axis are different between species.

Distribution of fitness effects

Tajima's D values reflect a summary of the site frequency spectrum rather than an explicit

quantification of the strength of purifying selection on nonsynonymous sites. To address this, we

estimated the distribution of fitness effect (DFE) to further understand the strength of purifying

selection across life stages while controlling for expression level (Supplementary Methods). We

first compared the DFE of gametophyte-specific, sporophyte-specific and unbiased genes (Figure

S6, Table S9), then we binned the gene sets into four expression level quantiles (Figure S7, Table

S10). In both species, higher expression level was associated with lower proportions of

effectively neutral mutations ($0 < N_e s < 1$), consistent with an increasing level of constraint due

to higher expression level (Figure S7).

In *C. purpureus*, unbiased genes had a lower proportion of effectively neutral mutations and a

higher proportion of strongly deleterious mutations than gametophyte- or sporophyte-specific

genes. This finding is consistent with unbiased genes experiencing higher levels of constraint

whereas stage-specific genes are more likely under relaxed purifying selection (Figure S6b).

Similarly, in the highest expression level quantile (qt4), unbiased genes had a slightly lower

proportion of effectively neutral mutations than gametophyte- or sporophyte-specific genes.

However, in the other expression level quantiles the same pattern was not evident: unbiased

genes had a higher proportion of effectively neutral mutations in the first quantile, and lower

proportion of strongly deleterious mutations in the second quantile (Figure S7b). In *R. hastatulus*,

gametophyte-specific and unbiased genes had similar proportions of effectively neutral and

strongly deleterious mutations and sporophyte-specific genes had the lowest proportion of

effectively neutral mutations and the highest proportion of strongly deleterious mutations (Figure

S6a), consistent with Figure 2c. Across expression level quantiles, the confidence intervals

mostly overlapped between gene sets and mutation categories in *R. hastatulus* (Figure S7a).

Based on the haploid purging hypothesis, we had predicted that selection efficacy would be

stronger in haploid than diploid phases. We therefore expected gametophyte-specific genes to

have higher proportions of slightly ($1 < N_e s < 10$) or strongly ($N_e s > 10$) deleterious mutations, or

a lower proportion of effectively neutral mutations ($0 < N_e s < 1$). In contrast to previous studies

(Arunkumar et al. 2013; Gossmann et al. 2014; Harrison et al. 2019; Gutiérrez-Valencia et al.

2021; Cervantes et al. 2023), our results in both species did not support the haploid purging

hypothesis. In *R. hastatulus*, gametophyte-specific genes had a higher proportion of strongly

deleterious mutations than sporophyte-specific genes, in *C. purpureus* the proportions were

similar with overlapping confidence intervals (Figure S6). In the low to intermediate expression

quantiles (qt1 to qt3), sporophyte-specific genes showed a slightly higher proportion of strongly

deleterious mutations ($N_e s > 10$) than gametophyte-specific genes in both species (Figure S7). In

the highest expression quantile (qt4), gametophyte-specific genes had slightly higher proportions

of strongly deleterious mutation ($N_e s > 10$) than sporophyte-specific genes in both species, but

the differences were not significant with overlapping confidence intervals (*R. hastatulus*: 67.19 -

70.21% in n, 65.38 - 68.66% in 2n; *C. purpureus*: 77.09 - 80.36% in n, 74.22 - 78.78 % in 2n,

Figure S7, Table S10).

Genome-wide scan for balancing selection

These overall patterns of diversity do not show a genome-wide signal of an enrichment of

balancing selection on genes with overlapping expression between the life stages; however, it is

possible that individual genes may still be subject to balancing selection. We therefore took an

alternative approach by conducting a genome-wide scan for signals of balancing selection and

explored whether they were enriched for genes that might be subject to antagonistic pleiotropy

between life stages. We applied a composite likelihood ratio test to scan the genome for

candidate genes under balancing selection using values of polymorphism and divergence (Cheng

and DeGiorgio 2019, 2020, 2022). In *R. hastatulus,* a total number of 4,945,561 sites were tested

in windows based on genetic positions (Figure 4). We found 79,431 sites with strong signals of

balancing selection across all chromosomes (CLR > 9.5), located at 449 genes, 405 of which had

evidence of expression. The candidate genes were only located in regions with considerable

recombination, based on our genetic maps (Figure 4). However, among the genes found in

regions with evidence of balancing selection, 138 of them were located within inversion

polymorphisms identified from two phased whole genome assemblies of *R. hastatulus* on

chromosomes A2 (63), A3 (74), and A4 (1) (Sacchi et al. 2024; Sacchi et al, in prep), consisting

of more than 30% of all candidate genes; whereas these inversions contained 16.7% of genes

across the genome. Because inversion heterozygotes are likely to experience reduced

recombination, these candidate genes may be subject to non-independent signals of balanced

polymorphism. In *C. purpureus*, the scans were performed in physical position windows as site-

specific recombination rates were unavailable (Figure S8). We tested 7,743,902 sites and

identified 339,375 sites under balancing selection (CLR > 956.2), located at 963 genes, 807 of

which had evidence of expression in this study. In *R. hastatulus*, candidate genes under balancing

selection had significantly higher π_s, and significantly lower π_n/π_s than genes that were not under

balancing selection (Welch Two Sample *t*-test, *p*-value = 0.0001511, 0.0003204, respectively). In

C. purpureus, there was no significant differences in π_s or π_n/π_s between genes under balancing

selection and gene that were not (Welch Two Sample *t*-test, *p*-value = 0.6029; Two Sample *t*-test,

p-value = 0.5956, respectively).

We performed contingency tests on the number of genes in different expression bias categories

(gametophyte-biased, sporophyte-biased, unbiased) for genes under balancing selection and

across the whole genome (Tables S11). In both species, we found no enrichment of any category

of expression bias under long-term balancing selection compared to the whole genome (Fisher's

exact test, p > 0.05). We examined the gene functions for all gametophyte-biased genes in

regions under balancing selection (Table S12, S13). We performed GO enrichment after

excluding the genes found within inversion polymorphisms in *R. hastatulus*, since the

suppression of recombination between inversion types can influence our ability to find functional

enrichments of causal loci. The GO enrichment results after excluding inversions were not

compelling (Table S12). In *R. hastatulus*, two genes under balancing selection had probable

disease resistance functions based on orthologs to *Arabidopsis thaliana* (At4g19060,

At4g14610). In *C. purpureus*, two genes under balancing selection were expressed in both life

stages with high gametophyte bias and have sexual reproduction functions (log2FoldChange > 3,

GO:0019953). These genes were also in the top 10% quantile of π_s.

Figure 5. Genome-wide scan of balancing selection in *R. hastatulus*. y-axis: genetic positions of

sites being tested (top), composite likelihood ratio (bottom). $\hat{a}$: estimated dispersion parameter, a

positive $\log_{10}\hat{a}$ value suggests balancing selection.

Discussion

We combined population genomic and differential gene expression analyses to examine the population genetic signals of balancing selection due to potential conflict between life stages in *R. hastatulus* and *C. purpureus*. First, we showed that there is larger overlapping gene expression and less differential expression in the moss *C. purpureus* than in the angiosperm *R. hastatulus,* based on our sampling of tissues. If most of the signature of balancing selection is attributable to antagonistic pleiotropy between life stages, we would expect a signal of greater balancing selection on genes with overlapping expression between life stages. In contrast, we found elevated π_s, π_n and D_n in highly gametophyte-biased genes in both species. Combined with the observations that π_s, π_n and π_n/π_s all decreased as gene expression level increased, the elevated diversity suggests relaxed purifying selection in highly gametophyte-biased genes rather than balancing selection. In addition, based on DFE comparisons we found no evidence that gametophyte-specific genes undergo stronger purifying selection than sporophyte-specific genes. Lastly, we performed genome-wide scans and identified hundreds of candidate genes under balancing selection in both species; however, we did not observe an enrichment of gametophyte-biased genes in candidates. Below we discuss the implications of our results for the evolutionary consequences of gametophytic selection in species in which the gametophytic life stage differs in duration and the challenges of using genome-wide population genomics to test models of conflict.

Relaxed purifying selection rather than balancing selection

Balancing selection generated by opposing selective pressures between sexes (sexual antagonism) has been tested in a series of empirical studies using genetic diversity and sex-biased expression (Cheng and Kirkpatrick 2016; Kasimatis et al. 2017; Mank 2017; Wright et al. 2018;

Sayadi et al. 2019). These studies have shown elevated diversity in weakly sex-biased genes

consistent with the prediction of intralocus sexual conflict. This is because highly sex-biased

genes likely reflect resolved conflict with little potential for ongoing antagonism, whereas fully

unbiased genes may be enriched in genes with highly concordant effects in males and females.

Similarly, we predicted higher genetic diversity with lower or intermediate expression bias due to

intralocus conflict between life stages. In contrast, we found higher π_s, π_n and D_n in highly

gametophyte-biased genes in both species (Figure 1), which may be explained by balancing

selection or relaxed purifying selection on nonsynonymous and synonymous sites in highly

gametophyte-biased genes. As highly expressed genes are under stronger selective constraint

than lowly expressed genes (e.g., Urrutia and Hurst 2003; Slotte et al. 2011; Zhang and Yang

2015), an observation of high π_s in gametophyte-biased genes with high expression level would

suggest balancing selection. However, π_s decreases with higher expression level in both

gametophyte- and sporophyte-biased genes in both species (Figure 3), and thus we conclude that

relaxed purifying selection on both nonsynonymous and synonymous sites is the more plausible

explanation. Overall, our results are not consistent with our predictions for widespread intralocus

conflict between the life stages. However, other processes are likely to contribute to the patterns

of elevated genetic variation but were not tested in this study; below we discuss the alternative

processes and caveats based on our dataset.

Two processes could lead to higher π_s in genes subject to relaxed purifying selection: weaker

background selection, or less direct selection on synonymous sites themselves, or both.

Background selection reduces neutral nucleotide diversity due to selection on linked deleterious

sites; the reduction in diversity is larger when the linked region contains more constrained sites

(Elyashiv et al. 2016). Highly gametophyte-biased genes could be subject to weaker background

selection, resulting in elevated π_s and π_n. Synonymous mutations are generally assumed to be

neutral as they do not affect amino acid sequences of the encoded protein (King and Jukes 1969).

For example, DFE analyses often use synonymous (4-fold) sites as a neutral control to compare

the site frequency spectra with nonsynonymous (0-fold) selected sites (Keightley and Eyre-

Walker 2007). However, studies in fruit flies (*Drosophila melanogaster*) and yeast

(*Saccharomyces cerevisiae*) found that 22% and three quarters of examined synonymous

mutations are strongly deleterious, respectively (Lawrie et al. 2013; Shen et al. 2022; but see

Kruglyak et al. 2023). One explanation for selection on synonymous mutations is due to codon

usage bias, where synonymous codons that lead to more efficient translation are favored (Plotkin

and Kudla 2011; Hunt et al. 2014). Codon usage bias is also found to affect transcription and

gene expression levels (Zhou et al. 2016). A study in the liverwort *Marchantia polymorpha*

found relaxed purifying selection in sporophyte-biased genes and evidence for lower codon usage

bias in these genes (Shen et al. 2024). In our case, both synonymous and nonsynonymous sites

may be under constraint, and the level of constraint may be lower in highly gametophyte-biased

genes, resulting in elevated π_s and π_n.

Expression level and expression breadth across multiple tissues may have a stronger effect on

selection efficacy than the antagonistic pleiotropy across life stages. Many studies have found

expression level as a strong factor in predicting selection efficacy (e.g., Urrutia and Hurst 2003;

Slotte et al. 2011; Zhang and Yang 2015). Consistent with these findings, our results show that π_n

is lower than π_s across expression levels (Figure 3) and there is a decrease in π_n/π_s (Figure 3) and

the proportions of effectively neutral mutations (Figure S7) with higher expression level. If

gametophyte- and sporophyte-biased genes are likely to have more specialized functions than

unbiased genes, the reduced expression breadth in highly gametophyte-biased genes could

potentially explain why they are under relaxed purifying selection than unbiased genes. In

contrast with antagonistic pleiotropy, the prevalence of synergistic pleiotropy between life stages

could explain why we observe signals of stronger purifying selection on unbiased genes (e.g.,

deleterious mutations in unbiased genes are under purifying selection in both life stages), and this

may predominate in affecting patterns of genome-wide diversity over signals of antagonistic

pleiotropy elevating neutral diversity in these genes.

The limited sampling of tissues especially in the angiosperm *R. hastatulus* makes it challenging

to accurately account for expression breadth and to differentiate alternative processes that might

contribute to the patterns we observed. In *R. hastatulus*, we only used one tissue per life stage

(pollen for gametophyte and leaf for sporophyte), female gametophytic tissue (i.e., ovule) and

other sporophytic tissues (e.g, roots, stem, etc.) were not sampled. As pollen is a highly

specialized tissue, pollen-biased genes will likely have a smaller expression breadth than

unbiased or leaf-biased genes (e.g., Honys and Twell 2003). The lack of female gametophytic

tissue (i.e., ovules) in *R. hastatulus* makes it difficult to capture the potential sexual conflict

within the gametophytic stage (i.e., between pollen and ovule) and conflict between sporophytes

and female gametophytes; as a result, other potential forms of pleiotropy may have been missed.

Sperm competition can also contribute to the maintenance of genetic variation (Clark 2002;

Dapper and Wade 2016). The sperm and vegetative cells within pollen tissue might experience

different strengths of selection which could explain the patterns we see in the DFE results of *R.*

hastatulus (Arunkumar et al. 2013; Gossmann et al. 2014; Gutiérrez-Valencia et al. 2022). In *C.*

purpureus, we generated expression data from gametophytic and sporophytic tissues that include

many cell types. Why we observed more relaxed purifying selection in highly gametophyte- than

sporophyte-biased or unbiased genes in *C. purpureus* is less clear. The increase in π_s and π_n with

stronger expression bias between the life stages is more symmetrical in *C. purpureus* than *R.*

hastatulus. This result suggests there is a smaller difference in expression breadth between

gametophytes and sporophytes with multiple cell types in *C. purpureus* than pollen and leaf

tissues in *R. hastatulus.* Future studies will benefit from a wider and more complete sampling of

tissues from different plant species.

It is noteworthy that we found a common pattern of elevated genetic diversity in highly

gametophyte-biased genes in our two study species with contrasting predominance of the

gametophytic stage, despite the differences in data collection methods and genome properties.

The source population for nucleotide polymorphism data and genome properties, such as

recombination landscapes, differ in our two study species. The whole-genome resequencing data

of *C. purpureus* are generated from isolates collected from five locations across the Northern

Hemisphere (Carey et al. 2021c). The whole-genome sequencing data of R. *hastatulus* were

obtained from a single population in Texas, US (Pickup and Barrett 2013). The large non-

recombining regions in *R. hastatulus* affects gene density and, as a result, patterns of genetic

diversity across the genome (Rifkin et al. 2022). In contrast, bryophyte species have a higher

average recombination rate than vascular plants, the distribution of gene density is more uniform

across the genome (Gaut et al. 2007; Bowman et al. 2017; Lang et al. 2018; Healey et al. 2023).

As the genomic data from *C. purpureus* consist of populations from ecologically different

habitats, spatially variable selection could potentially maintain different locally adapted alleles

especially in the long-lived gametophytes, which would increase nucleotide diversity and

Tajima's *D* in gametophyte-biased than sporophyte-biased genes. Because both species show

similar patterns of both π_s and π_n despite differences, relaxed purifying selection in highly

gametophyte-biased genes is likely to be the most plausible explanation for both species.

Genome-wide scan for balancing selection

The maintenance of genetic variation is a key question in evolutionary biology, but determining

the genomic prevalence of balancing selection remains an unresolved issue despite intensive

effort. Standard methods of detecting balancing selection often use summary statistics like π and

Tajima's D, which could lack power compared to composite likelihood ratio (CLR) based

methods (summarized in Bitarello et al. 2023). We identified hundreds of genes in regions under

balancing selection across the genome in *R. hastatulus* (449) and *C. purpureus* (964) using a

CLR-based method called B statistics (Cheng and DeGiorgio 2019, 2020, 2022), within which

138 genes were located within inversion polymorphisms previously identified in *R. hastatulus*.

Note that because recombination should be suppressed in inversion heterozygotes, many of these

138 genes may be showing the signal of balancing selection due to linkage disequilibrium, rather

than being direct targets of balancing selection. Even though we did not find a genome-wide

pattern for balancing selection due to intralocus conflict between life stages in both species, we

performed functional enrichment of candidate genes under balancing selection to identify genes

subject to intralocus conflict between life stages. Similar approaches will be useful in sexual

conflict studies (Ruzicka et al. 2019), e.g., to test whether the identified sexually antagonistic

SNPs are under balancing selection. We identified two genes expressed in both life stages of *C.*

purpureus that show gametophyte bias and signals of balancing selection; studies of such genes

could shed light on the nature of any intergenerational antagonistic pleiotropy. Although we did

not see an enrichment of gametophyte-biased genes under balancing selection compared to the

whole genome, or more compelling functional enrichment, it is noteworthy that focal genes

under balancing selection and nearby unselected genes are indistinguishable due to linkage,

which affect our functional enrichment results. Challenges also remain with CLR-based methods

in interpreting results when a genetic map is unavailable, as information of recombination rate is

important in determining the window sizes of different detection methods (Bitarello et al. 2023).

Studies on balancing selection are important in helping the understanding of how polymorphisms

in structural variants such as inversions are maintained. Chromosomal inversions play a key role

in evolutionary processes such as local adaptation and speciation (Dobzhansky 1970; Kirkpatrick

and Barton 2006; Huang and Rieseberg 2020). Inversion polymorphisms in a population can be

identified through genomic studies (e.g., Sacchi et al. 2024). Studies in humans found an

enrichment of balancing selection signatures in inversion polymorphisms (Giner-Delgado et al.

2019). Similarly, our results show over 30% of candidate genes for balancing selection are

located within inversion polymorphisms in *R. hastatulus*. The potential predominance of

balancing selection signatures within inversion polymorphisms suggests a role of balancing

selection in maintaining inversion polymorphisms in populations (Dobzhansky 1970; Kirkpatrick

and Barton 2006). Future studies using genomic data from more taxa including trans-specific

polymorphisms will provide more evidence on the biological functions and evolutionary roles of

balancing selection.

Gene expression profiles in species with biphasic life cycles

The differentially expressed genes between life stages and their functional enrichment suggest

distinctive ecological and reproductive roles for gametophytes and sporophytes. There is a

smaller overlap in gene expression (87% vs 93%) and a larger proportion of differentially

expressed genes (79% vs 45%) between life stages in *R. hastatulus* than in *C. purpureus*. The

smaller overlap and greater differentiation in gene expression in the angiosperm *R. hastatulus*

may reflect the reduction of the gametophytic stage during land plant evolution, as gametophytes

become short-lived, more specialized and much more reduced in angiosperms than bryophytes.

Similar results were found when comparing the moss *Funaria hygrometrica* and the angiosperm

Arabidopsis thaliana (Szövényi et al. 2011). Similarly, the overlap in gene expression is larger in

the fern *Polypodium amorphum* (97.7%) of which the gametophytes are usually photosynthetic

and sometimes independent when mature compared to angiosperms (Sigel et al. 2018).

Furthermore, the proportions of sporophyte- and gametophyte-biased genes are more similar in

C. purpureus than in *R. hastatulus*, with slightly more biased genes towards the more dominant

life stage in each species respectively (Table 1). However, the RNA sequencing technologies,

sample sizes and the nature of gene annotations are different between these studies. Therefore,

comparative analyses using a wider sampling of species from all major land plant lineages, e.g.,

gymnosperms (Cervantes 2023), and directly comparable methods would be useful to test how

the varying complexity and length of life stages affects the proportion of overlapping and

differential gene expression between life stages. Overall, our results suggest the two plant life

stages undergo more concordant than discordant selection on gene expression despite differences

in their ploidy, genetics, development and morphology. This result is consistent with previous

plant-breeding studies showing selection on pollen has synergistic pleiotropic effects on the

fitness of sporophytic offspring, e.g., cold treatment in pollen increases cold tolerance in the

offspring in maize (reviewed in Hormaza and Herrero 1992). Future studies on the

developmental and evolutionary genetics of gametophytes and sporophytes in more plant species

will benefit both biotechnology and the understanding of land plant evolution.

Methods

Plant materials

For *R. hastatulus*, we collected seeds from a population in Rosebud, Texas, US (Pickup and

Barrett 2013) and planted seedlings in a glasshouse at Earth Science Center, University of

Toronto. We collected pollen grains from the male F_1 plants for RNA isolation based on a

modified protocol from Lu (2011). We collected leaf tissues from male and female F_1 plants for

DNA and RNA isolation. We used the standard protocols of Qiagen Plant Mini kit and Sigma

Aldrich Spectrum Plant Total RNA Kits for DNA and RNA isolation, respectively. All libraries

were prepared and sequenced at the Genome Quebec Innovation Centre in Montréal, Canada.

The DNA samples were sequenced at a depth of 10-15X. The median read numbers for DNA and

RNA sequences are 72,907,022 (range: 64,585,743 - 88,888,320) and 23,412,838 (range:

18,618,763 - 58,151,564), respectively (Table S1, S2).

For *C. purpureus*, gametophyte DNA sequences including 18 samples at 6 locations are available

from Carey et al. (2021c). All RNA samples were taken from juvenile (protonema) and mature

(gametophore) tissues at the gametophyte stage. Gametophyte RNA sequences from 3 male-

female sibling pairs at 2 developmental stages (each with 3 biological replicates) are available

from Carey et al. (2021a). We generated sporophyte tissues for RNA sequencing using the same

methods and at the same time with Carey et al. (2021a). The sporophyte RNA sequences

included 26 samples from 6 crosses, the number of replicates per cross is between 2 and 10

(Table S3). We removed one sporophyte RNA sample from our analyses because it had low

coverage. The median read number for the sporophyte RNA sequences is 52,038,814 (range:

33,030,143 - 64,278,575) (Table S3).

RNA Seq data and expression analysis

For *R. hastatulus*, we used the genome assembly and genome annotation from Rifkin et al.

(2022) and assessed the quality of RNA reads using FastQC (Andrews 2010). In *C. purpureus*,

we used a hybrid genome consisting of the male R40 genome assembly (autosomes, the V sex

chromosome) and the U sex chromosome from the female GG1 genome assembly (Carey et al.

2021a). We trimmed adaptor sequences and filtered out low-quality reads using trimmomatic for

all RNA samples (Bolger et al. 2014), and trimmed polyG sequences using fastp for the

sporophyte RNA samples (Chen 2023). We used FastQC to assess the quality of RNA reads

before and after filtering (Andrews 2010). In *C. purpureus*, we excluded four sporophyte samples

for being an outlier in PCA from the majority of sporophyte samples, our final sample size for

sporophyte RNA samples is 21.

In both species, we mapped the RNA reads to the genome assembly using STAR in two pass

mode (Dobin et al. 2013, v2.7.6a). We sorted the alignment SAM files and added read groups

using PicardTools (http://broadinstitute.github.io/picard/). We generated raw read counts for each

gene using featureCount (Liao et al. 2014) and then normalized the raw read counts by library

depth using DESeq2 (Love et al. 2014). In the expression analysis of *C. purpureus*, we included

both male and female gametophyte samples for autosomal genes, and only included female or

male gametophyte samples for U and V genes, respectively. A gene was identified as expressed if

the mean normalized read count for either life stages was > 5; we only kept expressed genes in

the subsequent analyses. We calculated expression ratios between life stages for each gene by

dividing the mean normalized read counts at each life stage. A gene was considered tissue-

specific if: a) it was in the 10% or 90% quantiles of expression ratio among all significantly

differentially expressed genes (adjusted p-value < 0.1), and b) it had an absolute value of

log2FoldChange > 2. We performed GO enrichment for biased and specific genes using topGO,

with expressed genes from each species serving as the gene universe for comparison. (Alexa and

Rahnenfuhrer 2023). We divided the genes into four bins based on quantiles of log2FoldChange

for gametophyte-biased (log2FoldChange > 0) and sporophyte-biased (log2FoldChange > 0)

genes separately. We defined unbiased genes as the middle two bins with lowest absolute value

of log2FoldChange as well as an adjusted p >= 0.05. We divided the expressed genes into four or

ten quantiles of expression levels and used the same cutoff for later analyses.

DNA Seq data and variant calling

For *R. hastatulus*, we assessed the quality of DNA raw reads using FastQC (Andrews 2010) and

mapped the reads to the genome assembly using bwa-mem2 (Vasimuddin et al. 2019). We sorted

the BAM files using SAMtools (Danecek et al. 2021), added read groups, and removed PCR

duplicates using PicardTools (http://broadinstitute.github.io/picard/). We called SNPs jointly on

all samples using freebayes (Garrison and Marth 2012) with invariant sites saved in the output (-

-report-monomorphic). The variant and invariant sites were filtered separately using VCFtools

(Danecek et al. 2011), and concatenated together using BCFtools (Danecek et al. 2021). For both

variant and invariant sites we removed sites with a proportion of missing data > 0.8 or with a

mean read depth per site <10 or >40. For variant sites, we only kept bi-allelic SNPs with

genotype quality > 30.

For *C. purpureus*, we removed adaptor sequences using trimmomatic (LEADING:3

TRAILING:3 SLIDINGWINDOW:10:30 MINLEN:40) (Bolger et al. 2014), and performed

quality control using FastQC before and after trimming (Andrews 2010). We masked the V sex

chromosome when mapping female samples and masked the U sex chromosome when mapping

male samples using BEDTools maskfasta (Quinlan and Hall 2010). The read mapping and BAM

file processing methods are the same as described above in *R. hastatulus*. We called SNPs on all

samples jointly using BCFtools mpileup with a ploidy of 1 and the -B option (Danecek et al.

2021). We only kept the VCF for autosomal genes for the subsequent analysis, as the U and V

sex chromosomes have low recombination (Carey et al. 2021a, 2021c). We performed VCF

filtering using BCFtools (Danecek et al. 2021). We removed sites with a combined read depth <

5 and filtered variant and invariant sites separately. For variant sites, we kept bi-allelic SNPs and

removed sites with low quality or mapping score (QUAL<30 && MQ<30). Finally, we removed

sites with a proportion of missing data > 0.8 for both invariant and variant sites.

Diversity statistics

In both species we identified 0-fold and 4-fold degenerate sites in the genome using the script

codingSiteTypes.py(https://github.com/simonhmartin/genomics_general/blob/master/codingSite

Types.py, accessed in 2020). We intersected the list of 0-fold and 4-fold sites with the filtered

VCFs to generate 0-fold and 4-fold VCFs for population genetic analyses. The numbers of sites

(variant and invariant) in each VCF are 11,432,804 (0-fold) and 2,891,775 (4-fold) in *R.*

hastatulus; 17,394,329 (0-fold) and 4,759,819 (4-fold) in *C. purpureus*. We used pixy to

calculate the 0-fold and 4-fold average per site nucleotide diversity for each gene across the

genome (Korunes and Samuk 2021, v1.2.6.beta1). Only genes with a minimum of 50 sites for

both 4-fold and 0-fold nucleotide diversity were kept in the subsequent analyses.

We used the script popgenWindows.py

(https://github.com/simonhmartin/genomics_general/blob/master/popgenWindows.py, accessed

in 2020) to calculate Tajima's *D* for each gene based on the variant-only VCFs. We calculated

the weighted average nucleotide diversity for each bin based on the number of sites for each gene

(see supplementary methods). We bootstrapped the genes in each bin 1000 times to generate the

95% confidence intervals. We calculated the weighted average nucleotide diversity (mean and

95% CI) in the same way for gametophyte-specific, sporophyte-specific and unbiased genes. We

removed zeros for π_s and π_n to calculate the π_n/π_s ratio for each gene.

All DNA samples in *R. hastatulus* are female, this reduces the bias in nucleotide diversity

estimates due to the divergence between the X and Y sex chromosomes in males. In *C.*

purpureus, nucleotide diversity was estimated from 16 DNA samples (8 males and 8 females)

from 5 locations, see sample information from (2021a).

Balancing selection scan

We used BalLeRMix+ to perform genomic scans for balancing selection with B2 statistics and

default options (Cheng and DeGiorgio 2022). In *R. hastatulus*, the scans were performed on each

chromosome using the genetic positions. In *C. purpureus*, the scans were performed in 1 Mb

sliding windows based on the physical positions on each chromosome, using a default

recombination rate of 1 cM/Mb. The threshold for signature of balancing selection was set as the

5% quantile in the non-zero values of composite likelihood ratio. We calculated the maximum

CLR per gene. We identified the genes that had at least one site with strong signal of balancing

selection and also with evidence of expression in this study. We performed GO enrichment for

candidate balancing selection genes using topGO, with expressed genes as the gene universe for

comparison (Alexa and Rahnenfuhrer 2023). For candidate balancing selection genes with

gametophyte bias, we used gametophyte-biased genes as the comparison to test the functional

enrichment for balancing selection.

Data availability

Raw reads of *R. hastatulus* DNA and RNA Seq are available in NCBI SRA database under

BioProject PRJNA744278 (to be released on Aug 1, 2025 or upon publication). Raw reads of *C.*

purpureus sporophyte RNA Seq are available through JGI genome portal, see project IDs in

supplementary Table S3. Scripts used in this study are available at

https://github.com/imengyuan/lifestage_conflict (private repository, to be made public upon

review/publication).

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