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The serotonin 1B receptor is required for some of the behavioral effects of psilocybin in
mice

Sixtine Fleury, Katherine M. Nautiyal*

Department of Psychological and Brain Sciences, Dartmouth College

*To whom correspondence should be addressed

6307 Moore Hall

Hanover, NH 03755

(603) 646-2778

Katherine.nautiyal@dartmouth,edu

Running title:

Abstract

Recent studies highlight the promising use of psychedelic therapies for psychiatric disorders, including depression. The persisting clinical effects of psychedelics such as psilocybin are commonly attributed to activation of the serotonin 2A receptor (5-HT2AR) based on its role in the acute hallucinatory effects. However, the active metabolite of psilocybin binds to many serotonin receptor subtypes, including the serotonin 1B receptor (5-HT1BR). Given the known role of 5-HT1BR in mediating depressive phenotypes and promoting neural plasticity, we hypothesized that it mediates the effects of psilocybin on neural activity and behavior. We first examined the acute neural response to psilocybin in mice lacking 5-HT1BR. We found that 5-HT1BR expression influenced brain-wide activity following psilocybin administration, measured by differences in the patterns of the immediate early gene c-Fos, across regions involved in emotional processing and cognitive function, including the amygdala and prefrontal cortex. Functionally, we demonstrated that 5-HT1BR mediates some of the acute and persisting behavioral effects of psilocybin. Although there was no effect of 5-HT1BR expression on the acute head twitch response, mice lacking 5-HT1BRs had attenuated hypolocomotion to psilocybin. We also measured the persisting effects of psilocybin on anhedonia and anxiety-like behavior using transgenic and pharmacological 5-HT1BR loss-of-function models and found that 5-HT1B is involved in mediating the decreased anhedonia and reduced anxiety-like behavior. Finally, using a network analysis, we identified neural circuits through which 5-H1BR may modulate the response to psilocybin. Overall, our research implicates the 5-HT1BR, a non-hallucinogenic serotonin receptor, as a mediator of the behavioral and neural effects of psilocybin in mice.

Introduction

In recent years, almost 10% of adults in the US meet the criteria for a mood disorder, and nearly 20% for an anxiety disorder (1). Despite the widespread use of selective serotonin reuptake inhibitors (SSRIs) as antidepressants and anxiolytics, these treatments require long-term administration and fail to provide relief for a significant proportion of patients (2). Developing novel therapeutic targets which include rapid-acting effective therapies is important for the treatment of depressive and anxiety disorders. Psychedelic compounds are emerging as potential therapeutic agents for treating psychiatric disorders. Growing evidence supports their efficacy in treating major depressive disorder (MDD) and other mood and anxiety disorders (3–8). Psilocybin is a serotonergic psychedelic found in *Psilocybe* mushrooms which is gaining traction as a promising therapeutic agent (9). Clinical studies report that one to two doses of psilocybin can produce antidepressant effects in MDD and treatment resistant depression patients that persist for weeks to months (5,7,10–14). However, the hallucinogenic properties of psilocybin limit its use for some patient populations and in everyday clinical practice. Uncovering the neural mechanisms responsible for the therapeutic benefits in preclinical studies is therefore crucial to developing other treatment strategies (15,16).

The persisting behavioral effects have been largely attributed to the activation of the 5-HT2A receptor, based on its well-documented role in mediating the acute hallucinogenic properties of serotonergic psychedelics (17–21). However, there is conflicting evidence emerging from preclinical studies on the necessity of 5-HT2AR in the behavioral effects, especially related to depressive behavior using pharmacological and genetic loss-of-function models (22–28). Some recent studies suggest that these behavioral and neural effects may be independent of 5-HT2AR activation (23,27). Psilocybin's capacity to drive neuroplastic changes has been demonstrated through structural changes in dendritic spines, including increased spine density and size in the prefrontal cortex and synaptic changes in the hippocampus (29) —all alterations which may remain intact with blockade of psilocybin binding to 5-HT2A receptors (23,30). Recent studies also implicate the neurotrophin receptor TrkB and brain-derived neurotrophic factor (BDNF) signaling in mediating psilocybin-induced synaptic plasticity, independent of 5-HT2ARs (31).

Psilocybin's active metabolite, psilocin, binds with high affinity to many serotonin receptor subtypes, not just 5-HT2A, and its polypharmacology is likely important for its clinical efficacy (32). One receptor besides 5-HT2A that binds psilocybin and may be important for the effects of psilocybin on behavior is the serotonin 1B receptor (5-HT1BR). As an inhibitory Gi/o-coupled G protein-coupled receptor (GPCR) that inhibits neurotransmitter release at axon terminals, it has been implicated in a number of plasticity mechanisms including long-term potentiation of cortical inputs to the hippocampus, and long-term depression of corticostriatal synapses (33,34). 5-HT1BRs have also been linked to increased anxiety and depressive behaviors (35–37). Finally, 5-HT1BR has been implicated in mediating the effects of other psychoactive substances including MDMA and Ketamine (38–42). These data suggest that 5-HT1B could be a critical mediator of psilocybin's persisting therapeutic effects.

Here, we hypothesize that psilocybin modulates depressive-like behaviors through 5-HT1B receptor activation. Using a genetic and pharmacological loss-of-function models,

we sought to investigate the role of 5-HT1B in neural and behavioral effects of psilocybin
in mice. We assessed the role of 5-HT1BR in modulating the acute drug effects of
psilocybin on brain-wide neural activity, head twitch, and locomotor behavior. We also
investigated the impact of 5-HT1BR on the long-term behavioral effects of psilocybin on
anxiety and anhedonia phenotypes, key features of depressive disorders. Our findings
suggest that the 5-HT1BR influences brain-wide neural changes following psilocybin
administration and is necessary for its enduring antidepressant-like effects in mice

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Materials and Methods
Animals. Mice were bred in the Dartmouth College vivarium and weaned at postnatal
day (PN) 21 into cages of 2–4 same-sex littermates. Cohorts of mice lacking 5-HT1BR
expression and littermate controls were generated by crossing female homozygous
floxed-tetO1B mice to male homozygous floxed-tetO1B mice with the βActin-tTS
transgene (tetO1B+/+ females crossed to tetO1B+/+::βActin-tTS + males), as previously
reported (43). N=27 (11 males, 16 females) were used in the c-Fos study. N=52 mice
(31 females, 21 males) were used to assess the acute behavioral effects of psilocybin.
N=155 (60 males, 95 females) were used to assess persisting behavioral effects of
psilocybin following chronic corticosterone administration. N=164 mice (61 females, 103
males) were used to assess persisting behavioral effects of psilocybin following chronic
behavioral despair. Mice were run in 3–4 experimental cohorts, and all treatment
conditions were represented within each cohort. N = 43 female mice were used for the
antagonist experiment. For the adult knockout experiment (N=30), temporal-specific
knockdown was achieved by withdrawal of doxycycline (Product # F5545, Bio-Serv,
Flemington, NJ) at PN60 from mice following administration to the dam throughout
gestation and nursing, and to the mice following weaning. As previously reported, this
results in normal levels of 5-HT1BR expression through development and knockout in
adulthood (43). Following PN120, mice were then administered chronic corticosterone
administration and persisting behavioral effects of psilocybin were examined. All mice
were maintained on a 12:12 light-dark cycle and on *ad libitum* chow and water except as
noted below for treatments and behavioral testing. Mice were between ages 3-9 months
old at time of testing. All procedures were approved by the Dartmouth College
Institutional Animal Care and Use Committee.

Drugs and treatments. Psilocybin (Cat # 14041, Cayman Chemical), or saline vehicle,
was injected intraperitoneally (i.p.) at the dose of 5mg/kg diluted in saline solution (0.9%
NaCl) at a volume of 10ml/kg. The dose of 5mg/kg was chosen based on our pilot
experiments in this strain of mice that induced post-acute behavioral changes. Although
the dose is on the higher side of the usual range of 1-5mg/kg commonly used in rodents,
is in line with other reports assessing behavioral effects of psilocybin in mice with doses
over 4mg/kg (44–46). For antagonist experiments, 0.9% saline vehicle, or the serotonin
5-HT1B/1D receptor antagonist GR127935 (CAT#508014; Sigma-Aldrich) was injected
i.p. at 10mg/kg (in 0.9% saline at 10ml/kg volume) 30 minutes prior to psilocybin or
saline vehicle administration, as previously used for pretreatment in behavioral
pharmacology studies to target 5-HT1B/D receptors (47–50).

Chronic corticosterone administration. Corticosterone (35 µg/mL; Cat # 2505, Sigma-
Aldrich) was dissolved in 0.45% beta-cyclodextrin (Cat # 4767, Sigma-Aldrich) in
drinking water, and delivered *ad libitum* in opaque bottles for 4 weeks. This paradigm
produces a corticosterone dosing of approximately 9.5 mg/kg/day (51). In non-stressed

mice, only 0.45% beta-cyclodextrin was dissolved in drinking water, and delivered *ad*
libitum in opaque bottles for 4 weeks.

Chronic behavioral despair model. As an alternative to chronic corticosterone
administration, a separate group of male mice were subjected to repeated stress
exposures via five forced swim sessions. Each mouse was placed in a 2-liter beaker
containing water (22-25ºC) for 10 minutes daily for five consecutive days. Mice remained
in their home cages between sessions. Mice were injected with psilocybin (1mg/kg, i.p)
or saline vehicle (10 ml/kg) 48 hours after the last swim session.

Behavioral tests.

Head Twitch Response and Locomotion. Head twitches and locomotion were analyzed
in a subset of mice after the administration of either vehicle or psilocybin (5mg/kg, i.p) in
both control and mice lacking 5-HT1BR. Immediately following injections, mice were
placed in an open field, and video recording was initiated with a camera positioned
directly above the arena for 1 hour. Three experimenters scored the first 15 minutes of
activity for head twitches while blinded to treatment, noting the total number of and time
stamp of each head twitch to compare across experimenters. Locomotion was scored
with EthoVision software.

Gustometer. A Davis Rig 16-bottle Gustometer (Med Associates MED-DAV-160 M) was
used to test the effects of psilocybin and 5-HT1BR expression on hedonic responding.
Mice were water-restricted for 5 days of initial training before being put on
corticosterone. On day 1 of training, mice were placed in the gustometer as a cage and
with free access to 5% sucrose for 15 minutes. On day 2, mice were placed individually
with free access to 5% sucrose for 15 minutes. On day 3, mice were placed individually
with access to 5% and 10% sucrose with tubes alternating for 30 minutes. Mice were
then food restricted for 2 days before being individually returned to the gustometer for 30
minutes with access to water, 2% sucrose, 4% sucrose, 6% sucrose, 8% sucrose, and
10% sucrose for a final training day using 6 bottle paradigm. Each concentration was
randomly presented for a maximum of 120 seconds at a time before switching to another
tube. Total licks at each tube were recorded using a capacitance-based system. Mice
that had less than 300 licks during the final training session were excluded from the
analysis. After the final training session, mice were put on corticosterone. The 6 bottle
paradigm was then repeated 24 hours after drug administration for the experimental
gustometer test. Each mouse was weighed before the 6 bottle paradigm and data were
normalized by weight. After the experimental test, mice were allowed food that day until
24 hours before the time of the novelty-suppressed feeding test (about ~1h30).

Novelty Suppressed Feeding (NSF). The NSF assay was conducted in a rodent
transport container (20' x 16') with the floor covered with approximately 2-4cm of corn
bedding to serve as an arena. Thirty minutes before testing, mice were placed
individually into holding cages. At the time of testing, a single pellet of food was placed
on a white platform (~4' diameter) in the center of the arena. The anxiogenic
environment was produced by placing a lamp with high luminosity (~120 lux) above the
center of the arena. Each mouse was placed in the same corner of the arena, facing the
wall, and a stopwatch was immediately started. The latency to eat, defined as the mouse
biting the pellet, was recorded over a session of 5 minutes. Immediately after a bite, the
pellet was removed from the arena. To control for hunger, mice were immediately placed

in their homecage after the testing session with a new food pellet, and the stopwatch
was started again to measure latency to eat within their homecage. Animals that did not
bite during the testing session were attributed the maximum value (300 seconds).
Animals were placed back in their homecage with food and water ad libitum once all
cage mates were tested. The NSF was performed 24 hours after the gustometer in the
corticosterone stress paradigm, and 48 hours after the EPM in the chronic behavioral
despair paradigm.

Elevated Plus Maze (EPM). Mice were placed in the center facing a closed arm and
videotaped while allowed to explore the maze undisturbed for 6 minutes. The maze has
four arms (35cm long x 5cm wide) that were 60cm above the ground. The two opposite
closed arms had 20cm high walls, while the two opposing open arms had a 1cm lip.
Behavior was scored using EthoVision software for the number of entries and time spent
in each arm. Mice were excluded from the analysis as outliers if the number of open arm
entries was greater than or less than two standard deviations from the mean. The EPM
was performed 24 hours after the NSF in the corticosterone stress paradigm, and 24
hours after the gustometer in the chronic behavioral despair paradigm.

c-Fos Whole Brain Analysis

Brain samples. A set of male and female tTS- (N=14) and tTS+ (N=14) mice not
maintained on doxycycline was used to perform a whole brain c-Fos analysis following
vehicle or psilocybin (5mg/kg, i.p) administration. Brain-wide c-Fos expression was
induced during the acute phase of psilocybin administration. Mice were injected with
either vehicle or psilocybin (5mg/kg), and then placed back in their home cage for 2h,
before perfused with 0.1 M PBS followed by 4% formaldehyde. Brains were extracted
and postfixed for 24 hours in 4% formaldehyde before being cryoprotected in 30%
sucrose. Brains were then sectioned in a coronal plane with a thickness of 40um on a
cryostat (Leica Biosystems) and stored in 0.1 M PBS at 4ºC in 24 well plates. About 20
to 30 sections per brain were selected for staining. One brain in the saline-WT group
was eliminated due to technical issues with tissue sections prior to processing.

Tissue processing. Sections were first washed three times in PBS-T (0.1%) for 10
minutes. Sections were then blocked for 1h in 2% Normal Donkey Serum (#NC9624464,
Fisher) in PBS-Triton (0.1%). Sections were then incubated in primary antibody (rb anti-
cfos Rabbit Recombinant Monoclonal c-Fos antibody, ab214672) at 1:1000 in blocking
solution overnight at 4ºC. The following day, sections were washed 3 times again in
PBS-T (0.1%). Sections were then incubated in secondary antibody Alexa Fluor 488
anti-rabbit (ab150073, Abcam) at 1:250 and DAPI at dilution 1:10,000 in PBT (0.15) for 2
hours away from light. Sections were then washed again three times and mounted.
Once dry, the slides were cover slipped with ProLong Gold. Whole sections were then
imaged on Keyence BZ-X800 microscope with a 10x objective. Fluorophores were
imaged with DAPI and GFP filter cubes.

Segmentation of fluorescent labels and quantification. Cells expressing a c-Fos label
were segmented using the machine learning-based pixel and object classification
program, *Ilastik*, as previously described in Berg et al., 2019. *Ilastik* was also used to
generate binary images of the segmented c-Fos labels. Registration and quantification of

histological images on a brain-wide scale were performed using the open-source atlas
registration tool FASTMAP, as previously described in (52). Regions that were not
adequately aligned or identified on stained sections were left out of the overall analysis.

Statistical Analyses

All behavioral analyses were performed using GraphPad Prism (v.10.2.2) or RStudio
(v.2023.06.1+524). The threshold of statistical significance was set at 0.05. Data were
analyzed using one-, two-, and three- ANOVAs, linear mixed models, generalized linear
models, and t-tests using GraphPad Prism and RStudio, with Tukey's correction for
multiple comparisons when appropriate. Area under the curve for locomotion was
calculated using the trapz function in RStudio. Functional connectivity analyses were
performed on a collection of 30 regions based on our interest and ability to reliably and
consistently delineate these regions using a DAPI stained reference image. A psilocybin
index was generated by subtracting average total c-FOS counts. The density of regional
c-Fos expression was cross-correlated within each group to generate pairwise
correlation matrices, and difference matrices were obtained by subtracting coefficients
within each element. To quantify psilocybin-induced changes in network synchrony
within each genotype, we extracted the upper triangle to obtain pairwise correlation
changes. We then performed a one-sample-t-test on each vector to determine whether
the mean change in correlation significantly differed from zero within each genotype. The
similarity and strength of matrices were assessed using nonparametric permutation
testing Monte Carlo and Spearman correlation to generate rho and p values. For
network analysis, we performed a fisher r-to-z transformation on correlation and selected
significant edges at 95%, 90%, 85%, and 80% confidence intervals.
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276 **Results**
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Psilocybin-induced changes in neural activity require 5-HT1BR expression

To investigate how psilocybin changes brain-wide neural activity, and whether 5-HT1BR
expression influences those changes, we performed a whole-brain c-Fos analysis on
male and female mice lacking the 5-HT1BR and their littermate controls. Brains were
collected 2h following a single injection of psilocybin (5mg/kg) or saline vehicle, and we
found that the number of c-Fos+ cells was relatively consistent across animals within
brain regions (Fig 1A). Overall, there was a significant effect of psilocybin on neural
activity measured by c-Fos staining (main effect of psilocybin: $F_{(1,23)}=5.708$, p=0.026).
Interestingly, the neural activity was significantly influenced by 5-HT1BR expression
(genotype by drug interaction: $F_{(1, 23)}=5.986$, p=0.023), though the pattern varied by
brain region (Fig S1; genotype by drug by region interaction: $F_{(28, 628)} = 2.352$,
p=0.0001). Some brain regions showed significant effects of psilocybin regardless of 5-
HT1BR expression, such as the somatosensory cortex and the claustrum (CLA) which
both express high levels of 5-HT2ARs (Fig 1B, main effect of drug on CLA: $F_{(1, 22)}=21.74$, p=0.0001). There were also select brain regions, such as the basolateral
amygdala (BLA) which showed differential baseline c-Fos activity based on genotype
(main effect of genotype: $F_{(1, 22)} = 4.832$, p = 0.039) which potentially drove the
differential effect of psilocybin between genotypes (genotype by drug interaction: $F_{(1, 22)} = 11.13$, p = 0.003). Most interestingly, quite a few other brain regions showed an
effect of 5-HT1BR expression on the c-Fos response to psilocybin including prefrontal
cortical areas and the central amygdala (CEA) which showed increased c-Fos in control,

but not 5-HT1BR KO mice (drug by genotype interaction for CEA: $F(1, 23)=8.084$,
p=0.009; controls: p=0.0002, 5-HT1BR KOs: p=0.669). The globus pallidus showed a
similar pattern with increased c-Fos expression in controls, but not 5-HT1BR KO mice
(drug by genotype interaction for GP: $F(1, 22)=4.954$, p=0.037; controls: p=0.003; 5-
HT1BR KO: p=0.829). Importantly, these regions express high levels of 5-HT1BR protein
(Fig S2; (53,54), and may be key sites for psilocybin acting via these receptors to
influence neural activity. Overall, psilocybin differentially influenced neural activity
throughout the brain leading to both increases and decreases depending on the brain
area, and these changes in neural activity were differentially influenced by 5-HT1BR
expression (Fig 1C). This points to a role for the 5-HT1BR to mediate the behavioral
effects of psilocybin in rodents.

5-HT1BR may influence some of the acute behavioral effects of psilocybin

To investigate the functional effect of 5-HT1BR activation on the acute behavioral
response to psilocybin, we measured the head twitch and locomotor response to
psilocybin in male and female control and 5-HT1B KO mice. Mice treated with psilocybin
demonstrated significantly more head twitch responses than mice treated with saline
(Fig 2A; main effect of psilocybin: $F (1, 28) = 312.3$). However, there was no significant
difference in the number of head twitches between genotypes (main effect of genotype:
$F (1, 28) = 0.4982$), indicating that psilocybin robustly induces head twitches in mice
independent of 5-HT1BR. This is consistent with previous research implicating 5-HT2AR
activation in this acute response to psilocybin (55,56), and demonstration of normal
levels of 5-HT2AR expression in mice lacking 5-HT1BRs (57). We also analyzed the
locomotor response to psilocybin in mice lacking 5-HT1BR expression, and found that
unlike the head twitch response, 5-HT1BR may be involved in the acute hypolocomotor
effects (Fig 2B). Specifically, psilocybin significantly decreased locomotion in the 30min
following administration ($F(1, 48)=8.18$, p<0.0063), without effects on velocity (Fig S3A).
This effect seemed attenuated in mice lacking 5-HT1BR, however this interaction didn't
reach statistical significance (genotype by psilocybin interaction $F(5, 240)=2.00$,
p=0.0791). Using AUC values for planned comparisons within genotype (Fig 2C), there
were significant reductions in locomotor activity following psilocybin in control ($t_{28}=2.92$,
p=0.0068), but not in 5-HT1BR KO mice ($t_{20}=1.26$, p=0.16). Overall, these data show
that 5-HT1BR could partially mediate some acute behavioral responses to psilocybin,
though not the classic head twitch response.

5-HT1BR is required for antidepressant-like effects of psilocybin in female mice
treated with corticosterone

Given that there are long lasting behavioral effects of a single administration of
psilocybin in humans, we next wanted to examine the role of the 5-HT1BR in the
persisting behavioral effects in mice. We used two paradigms to model stress-induced
anhedonia and anxiety-like behaviors. First, we administered chronic corticosterone in
the drinking water for four weeks to induce a stress-phenotype to measure
antidepressant-like effects of psilocybin (Fig 3A), which did not influence the acute
responses to psilocybin (Fig S3B-C). Interestingly, we tested mice lacking 5-HT1BR
expression and littermate controls in a series of behavioral tests that evaluated
anhedonia and anxiety-like behaviors, performed 24-72h after a single i.p. injection of
either vehicle or 5mg/kg psilocybin. In female mice, psilocybin significantly increased
hedonic responding in the gustometer as measured by increased licking for sucrose (Fig

3B; treatment by sucrose concentration: $F_{(2,384)} = 5.71$, p = 0.004). There was also a
significant effect of genotype on licking across sucrose concentrations ($F_{(1, 384)} = 9.12$,
p = 0.003) which is consistent with the baseline differences between genotypes that we
have previously reported (58). Though the interaction of genotype and treatment did not
reach statistical significance ($F_{(2, 384)}=2.53$, p=0.081), because there were baseline
differences, we compared the effect of psilocybin within each genotype. While control
mice showed a significant effect of treatment on sucrose consumption (Fig 3B; $F_{(2, 42)}$
=6.09, p=0.005), there was no effect in female mice lacking 5-HT1BR ($F_{(2, 30)} = 0.73$,
p=0.489). Looking specifically at consumption of the highest concentration of sucrose in
control mice (Fig 3C; main effect of treatment $F_{(2, 72)}=5.66$, p=0.005), psilocybin
increases consumption compared to saline treated mice (p=0.012), restoring it back to
non-cort baseline levels (p=0.266). In 5-HT1BR KO mice, although cort decreased
sucrose consumption (no cort/saline vs cort/saline: p=0.030), psilocybin did not
significantly increase the consumption (cort/saline vs cort/psilocybin: p=0.549). Overall,
these data suggest that psilocybin may influence anhedonia in mice through actions at
the 5-HT1BR.

Expression of 5-HT1BR also significantly modulated the anxiolytic response to psilocybin
in female mice. First, we measured anxiety-like behavior in the conflict-based novelty-
suppressed feeding (NSF) assay 48h following psilocybin administration. Interestingly,
there was an anxiolytic response to psilocybin that was significantly blunted in the
absence of 5-HT1BR expression (genotype by drug interaction: $F_{(2,72)} = 2.99$, p =
0.049). Specifically, control mice injected with psilocybin showed reduced latency to eat
compared to saline-treated mice (Fig. 3D, $\chi^2 = 9.62$, p = 0.002), while we saw no
significant effect in the 5-HT1BR KO mice (Fig. 3D, $\chi^2 = 1.85$, p = 0.174). There was also
a main effect of genotype independent of treatment ($F_{(1,72)}=7.29$, p=0.009), suggesting
that there are again baseline effects of brain-wide, life-long absence of 5-HT1BR
expression on anxiety-like behavior. There were no significant effects of genotype or
drug treatment in the home cage condition suggesting that the effects of psilocybin were
on anxiety behavior rather than feeding behavior (Fig S4A). We also tested mice in the
elevated plus maze (EPM), as a second measure of anxiety-like behavior, 72h following
psilocybin administration. We again found that psilocybin-induced decreases in anxiety
behavior that were dependent on 5-HT1BR expression (Fig. 3E; main effect of drug by
genotype ($F_{(1, 51)} = 4.080$, p = 0.0487). Psilocybin administration significantly
decreased anxiety in female control mice, as measured by increased entries in open
arms (p=0.01), but not in mice lacking 5-HT1BR (p = 0.732). This was also somewhat
reflected in the time spent in open arms of the EPM (Fig S4B), and was not due to any
persisting changes in overall locomotion (Fig S4C). Overall, convergent evidence from
two conflict-based assays shows that psilocybin's persisting anxiolytic response requires
5-HT1BR. Overall, these behavioral results indicate that in female mice, psilocybin
produces an antidepressant-like phenotype in the days following administration which
we measured as decreased anhedonia in the gustometer and decreased anxiety-like
behaviors in the NSF and EPM. Interestingly, these behavioral effects were attenuated
in the absence of 5-HT1BR expression, suggesting that the antidepressant-like effects of
psilocybin are dependent on 5-HT1BR following chronic corticosterone administration in
female mice.

Interestingly, in male mice, we were unable to find any behavioral effect of psilocybin
following chronic corticosterone treatment, which is consistent with previous findings
(27). Specifically, psilocybin had no effect on hedonic responding in the gustometer as

measured by licking for various sucrose concentrations (Fig 3F; $F_{(2, 230.4)}$=2.14,
p=0.12). Similar to the females, we again saw an effect of genotype on licking over the
sucrose concentrations in males ($F_{(1, 294)}$=11.36 , p<0.001), as previously reported
(58), however there was no significant effect of treatment ($F_{2, 294}$=2.37, p=0.1).
Analyzed within each genotype, there was no effect of psilocybin in either 5-HT1BR KOs
or controls ($F_{(2,23)}$=0.60, p=0.55 for controls, $F_{(2, 31)}$=1.1, p=0.34 for KOs). Similar
patterns were seen when looking at consumption of the highest concentration of sucrose
(Fig 3G). There was a significant genotype effect, but no effect of treatment ($F_{(1, 54)}$=5.3, p=0.028 for genotype, $F_{(2, 54)}$=1.3, p=0.28 for treatment).
54)=5.3, p=0.028 for genotype, $F_{(2, 54)}$=1.3, p=0.28 for treatment).

We saw a similar trend for males in two tests of anxiety-like behavior – there were
effects of cort treatment and of genotype, but no significant effects of psilocybin on these
measures. In the NSF test (Figs 3H and S4D) 5-HT1BR KOs showed reduced anxiety-
like behavior ($F_{(1, 55)}$=18.12, p<0.0001), but again there were no effects of psilocybin,
despite cort treatment increasing anxiety-like behavior (main effect of treatment:
$F_{(2,55)}$=5.92, p=0.005, veh/saline vs cort/saline: χ_1 = 6.72, p=0.006 for controls, χ_1 =
11.13, p=0.001 for KOs; cort/saline vs cort/psilocybin: χ_1 = 0.50, p=0.48 for controls, χ_1 =
1.66, 0.24 for KOs). In the EPM (Figs 3I, 4SE-F), there were no effects of genotype on
the number of entries into the open arms in males ($F_{(1,54)}$=2.22, p=0.14). There was a
significant effect of treatment ($F_{(2, 54)}$=12.64, p<0.0001), though this effect was driven
by the effect of cort (veh/saline vs cort/saline: p=0.0007), rather than psilocybin
(cort/saline vs cort/psilocybin: p=0.57). Overall, although genotype and cort had some
behavioral effects in males, psilocybin consistently did not influence anhedonia and
anxiety-like behavior.

Acknowledging potential compensatory effects of the absence of 5-HT1BR throughout
development, we used two approaches to address this alternative interpretation of the
psilocybin effect in females. First, we used an adult-knockdown approach, in which mice
were reared with normal expression of 5-HT1BRs through PN60 via administration of
doxycycline to block tTS-mediated knockdown (as previously reported and validated
(58)). Following withdrawal of doxycycline as adults, mice lacking 5-HT1BR expression
were tested for the persisting behavioral effects of psilocybin during chronic
corticosterone administration in the elevated plus maze (Fig 4A). Though limited in
number, a planned comparison shows no significant effect of psilocybin in adult
knockdown mice (t_{14}=0.26, p=0.80). Though the interpretation of this result is limited by
an effect in controls that was only trending toward significance (t12=1.461, p=0.11), a
lack of effect in the adult knockdown suggests that any developmental compensation
was likely not responsible for the lack of effects seen in the whole-brain whole-life
knockouts. This effect was mirrored in the time spent in the open arms (Fig S5A), and
was not due to differences in locomotion (Fig S5B). Additionally, to investigate if the
behavioral effects in the constitutive 5-HT1BR KO were due to activation of the 5-HT1BR
during the acute phase of psilocybin, and to attempt to circumvent genotype differences
seen in some of the behavioral tests, we performed another experiment in which we
pharmacologically blocked 5-HT1BR with pre-treatment with a 5-HT1B/1D receptor
antagonist, GR127935, during psilocybin administration in control mice. Psilocybin again
decreased anxiety-like behavior as measured by increased open-arm entries as a
function of 5-HT1BR activation (Fig. 4B, pretreatment x psilocybin interaction: $F_{(1, 48)}$=4.30, p=0.044). Specifically, in the absence of any pretreatment, psilocybin
48)=4.30, p=0.044). Specifically, in the absence of any pretreatment, psilocybin
increased open arm entries (p=0.0084), however when pre-treated with GR127935, the
effect of psilocybin was blocked (p=0.89). Measures of open arm duration or locomotion

were not significantly influenced by treatment (Fig S5C-D). Overall, these results
suggests that blockade of 5-HT1BR during the acute psilocybin response may be
sufficient to block the post-acute anxiolytic effects of psilocybin seen in the EPM.

5-HT1BR mediates the anxiolytic effects of psilocybin in a behavioral despair
model in both female and male mice

We used a second stress paradigm to confirm the results seen in the female mice in the
chronic cort model, and because the chronic cort model was not effective to measure the
persisting behavioral effect of psilocybin in male mice. Using a chronic behavioral
despair paradigm implemented through repeated forced swim stress, we examined the
effect of the absence of 5-HT1BR on the behavioral effects of psilocybin in mice (Fig
5A). Interestingly, in this paradigm we were able to elicit an effect of psilocybin in both
sexes. There were significant effects of treatment on consumption across the sucrose
concentrations (Fig 5B; treatment x concentration: $F_{(2, 538)}=3.63$, $p=0.027$). Given that
we saw a baseline genotype effect again (genotype x concentration: $F_{(1, 538)}=10.29$,
$p=0.001$; genotype x treatment x concentration $F_{(2, 538)}=2.41$, $p=0.09$), we analyzed
the treatment effects within each genotype separately. In control mice, there was a
significant effect of treatment on sucrose consumption (treatment x concentration: $F_{(10, 330)}=2.87$, $p=0.002$), which was largely driven by a increase in consumption of 10%
330)=2.87, p=0.002), which was largely driven by a increase in consumption of 10%
sucrose following psilocybin compared to saline treatment (Fig 5C; $p=0.009$). Though
there were no significant sex differences ($F_{(1,488)}=0.86$, $p=0.35$) and both sexes
showed similar trends, the effects of psilocybin in females may be greater (Figs S6A-D).
On the other hand, there was no effect of treatment in 5-HT1BR KO mice ($F_{(10, 190)}=0.75$, $p=0.68$), suggesting that stress and/or psilocybin don't significantly influence
190)=0.75, p=0.68), suggesting that stress and/or psilocybin don't significantly influence
hedonic responding in the absence of 5-HT1BR expression in mice.

In the EPM, there was a significant effect of genotype on the anxiolytic effect of
psilocybin (Fig 5D; interaction of genotype and treatment: $F_{(2, 155)}=4.22$, $p=0.016$). In
control mice, stress decreased entries into the open arms ($p<0.0005$), and psilocybin
rescued this effect ($p=0.03$). Interestingly, in the mice lacking 5-HT1BR expression,
psilocybin had no effect on entries into the open arms ($p=0.99$), though stress also had
no effect in this strain ($p=0.84$). This pattern was replicated in the duration of time spent
in open arms of the EPM (Fig S6E-G). There were also no effects on locomotion
generally in the EPM (Fig S6H-J), and no significant effects of sex on the EPM behavior
(genotype x treatment x sex: $F_{(2, 155)}=0.146$, $p=0.86$).

In a second measure of anxiety behavior in the NSF assay, psilocybin also decreased
anxiety-like behavior as measured by latency to eat (Fig 5E, $X_5=31.40$, $p<0.0001$). There
were again some baseline differences between genotypes, but importantly within control
mice, psilocybin decreased the latency to eat (stress/saline vs stress/psilocybin:
$X_1=6.41$, $p=0.011$), but had no significant effect in 5-HT1BR KO mice ($X_1=1.70$, $p=0.19$).
There were again no effects of sex on the treatment effects (sex by treatment: $F_{(2, 104)}=0.14$, 0.87). There were no effects of genotype or psilocybin on latency to eat in
104)=0.14, 0.87). There were no effects of genotype or psilocybin on latency to eat in
the homecage suggesting that hunger was not a cause of the differences in behavior
(Fig S6K-M).

Neural circuits impacted by psilocybin are influenced by 5-HT1BR expression

To identify the neural circuits through which 5-HT1BR may mediate the effects of psilocybin on behavior, we performed a network analysis on neural activity following psilocybin to probe functional connectivity. Returning to the c-Fos data (shown in Fig 1), we generated pairwise correlation matrices of brain region activity as measured by number of c-Fos+ cells to determine how the activity of brain regions vary together. A difference matrix for each genotype, representing the change in correlated activity following psilocybin administration, illustrates how the functional connectivity throughout the brain is changed by psilocybin and differs based on 5-HT1BR expression (Fig. 6A). Comparing these two correlation matrices reveals statistically distinct patterns between control and 5-HT1BR KO mice (Fig 6A; rho = -0.01, p = 0.80). Control mice showed more desynchrony following psilocybin compared to saline (t = -5.336 with p<0.0001). Interestingly, this is reversed in mice lacking 5-HT1BR (t=+5.083 p < 0.0001 for KOs), suggesting that neural activity is more positively correlated across regions following psilocybin treatment. We then examined which psilocybin-responsive networks were most influenced by 5-HT1BR expression and found significant effects of 5-HT1BR on correlated activity between the central amygdala and the anterior cingulate cortex (z=2.00, p=0.05). Widening the confidence interval, we found suggestive differences in the correlated activity of the central amygdala with the claustrum, amongst amygdalar regions, and between the globus pallidum and lateral amygdala and anterior cingulate cortex (Fig 6B). Overall, this network analysis shows that 5-HT1BR expression significantly influences neural circuits that are relevant to the effects of psilocybin on cognition and mood. Furthermore, it points to potential neural circuits through which 5-HT1BR may influence some of the persisting effects of psilocybin.

Discussion

Our results highlight a critical role for 5-HT1BR in the response to psilocybin in mice. We found that the absence of this non-hallucinogenic receptor impacts the neural response to psilocybin, as well as the acute and persisting behavioral effects. Using c-Fos as a marker of neural activity, we find increased activity in many regions throughout the brain including in sensory and prefrontal cortices, claustrum, amygdala, hypothalamus, and basal ganglia, which is consistent with previous reports (22,59–61). Interestingly, we saw that mice lacking expression of 5-HT1BR have notable differences in the pattern of neural activity following psilocybin administration, including in prefrontal, hippocampal, and amygdalar areas, all brain regions that express high levels of 5-HT1BR. Functionally, a lack of 5-HT1BR had no effect on the head twitch response, though it did attenuate the acute hypolocomotor effects of psilocybin. Importantly, there were also effects of 5-HT1BR on the persisting behavioral effects of psilocybin seen by the amelioration of stress-induced anhedonia and anxiety-like behaviors. Overall, this shows that 5-HT1BR expression is involved in mediating some the neural and behavioral effects of psilocybin..

Though we find that 5-HT1BR is required for the persisting effects of psilocybin, our data do not address its sufficiency. The profound neural and behavioral effects of psilocybin are likely supported by its polypharmacology at multiple serotonin receptors (32). Although there is some evidence questioning the necessity of the 5-HT2AR in the behavioral effects of psilocybin in mice (23,26), there are mixed findings on this topic

with other reports showing that 5-HT2AR expression is necessary for the anti-
depressant effects of psilocybin (22). Tangential to this debate, our results show that
acute activation of the 5-HT2A receptor by psilocybin is insufficient to rescue stress-
induced behaviors in the absence of the 5-HT1BR.

There are many lines of evidence linking the 5-HT1BR to antidepressant function as well
as to the behavioral and neural response to hallucinogens (62,63). As an inhibitory $G_{i/o}$-
coupled receptor, its activation reduces calcium influx in axon terminals of serotonin
(autoreceptors) and non-serotonin (heteroreceptors) neurons decreasing
neurotransmitter release (64–66). Past research shows that 5-HT1BR agonists promote
antidepressant-like effects and can increase the rewarding effects of cocaine and social
interaction (67–70). 5-HT1BR also interacts with p11 to regulate depression states
(71,72). Additionally, 5-HT1BR is necessary for the behavioral and hippocampal
plasticity effects of fluoxetine (33). In addition to mediating depressive phenotypes, there
is also evidence implicating the 5-HT1BR in antidepressant response to ketamine
(39,42), and in other behavioral effects of MDMA (38,40,41).

In keeping with our previous work showing that the 5-HT1BR has a known role in
mediating reward and anxiety behavior (36,43,58), we do find baseline differences in
some of the behaviors we tested in 5-HT1BR KO mice. Additionally, there were also
behavioral assays in which mice lacking 5-HT1BR seemed to show more resilience to
chronic corticosterone treatment, particularly in the gustometer measure of anhedonia.
These differences may arise from compensatory changes due to the absence of 5-
HT1BR throughout development. Alternatively, these baseline differences could be a
reflection of the important role of 5-HT1BR in mediating these phenotypes in adulthood.
However, the baseline differences leave open the limitation of potential behavioral and
mechanistic ceiling effects which could have obscured the effects of psilocybin in this
strain. This would be possible either through maxing out the range of the behavioral
assays and/or through saturation of the 5-HT1BR-dependent mechanism through which
psilocybin has its effects. We first addressed these limitations by using at least one
behavioral assay with no baseline differences, namely the elevated plus maze.
Additionally, we tested mice with adult knockdown of the receptor to eliminate any
developmental compensation. Finally, we used a pharmacological blockade of 5-HT1BR
by administering an antagonist prior to psilocybin administration. This not only allowed
us to circumvent any baseline differences, but also helped identify that 5-HT1BR is
involved during the acute drug effect.

Consistent with past research, we find some sex differences in our behavioral effects of
psilocybin. We found that the post-acute behavioral effects of psilocybin on anhedonia
and anxiety-like behavior were influenced by an interaction of stressor-type and sex.
Specifically, psilocybin was able to rescue the anhedonia and anxiety-like behavior in
females exposed to chronic corticosterone treatment, however, this effect was not seen
in males. This sex difference may have emerged from a differential response to the
stress paradigm rather than a differential response to psilocybin, particularly due to a
more robust effect of the corticosterone treatment on the behavior of males. This is
consistent with past work showing that there are sex-specific effects of chronic
corticosterone administration in the drinking water (73,74). On the other hand, using a
behavioral despair model via repeated forced swim stress, psilocybin was able to rescue
stress-induced behavioral phenotypes in both male and female mice. This suggests that
different stress modalities may induce sex-dependent alterations by engaging different

neural circuits that are amenable to varied pharmacological interventions. For example,
recent work shows that psilocybin differentially influences hypothalamic reactivity to
aversive stimuli and reduces ethanol consumption specifically in male, but not female
rodents (59,75). On the other hand, females display more head twitch behavior to the
psychedelic 1-(2,5-dimethoxy-4-iodophenyl)-2-aminopropane (DOI) and less of an
augmentation of pre-pulse inhibition in males, in a strain dependent manner (76,77).
Ultimately, there appears to be a substantial impact of sex on the effect of psilocybin on
behavior, which are dependent on strain and paradigm, reinforcing the importance of
examining sex differences in future studies.

While the 5-HT2AR plays a central role in psilocybin's acute psychedelic effects, its
therapeutic potential in the context of stress-induced depression may be limited without
the support of other serotonin signaling pathways, including those acting via 5-HT1B.
The absence of 5-HT1BR signaling may prevent psilocybin from achieving its full
therapeutic potential as the feedback inhibition of serotonin release is disrupted, leading
to overactivation of serotonergic circuits, and limiting the ability to modulate neural
mechanisms effectively. This could result in a mechanistic ceiling of 5-HT2A, essentially
maxing out the 5-HT2A-dependent effects on anhedonia and anxiety-like behaviors. This
hypothesis arises in part from studies showing increased 5-HT2AR expression following
chronic exposure to stress (78–80). However, neither stress nor the knock-out model
seem to have affected acute 5-HT2AR activation in our study, as measured by head
twitch responding, and past studies showing no effect of 5-HT1BR KO on 5-HT2AR
binding (57). Additionally, although our hypothesis of 5-HT1BR involvement is via direct
binding of psilocin to 5-HT1BRs on neural circuits implicated in anxiety and depressive
behaviors, it is also possible that these effects may be indirect. For example, 5-HT1BR
could influence the hypothermic response to psilocybin which could then influence the
antidepressant-like behavioral response (81–83). Overall, in combination with other
work, our results highlight that psilocybin's persisting effects on behavior may depend on
an optimal balance of serotonin tone acting across different serotonin receptor subtypes
potentially across a number of physiological systems.

Our studies begin to point to the anterior cingulate cortex, the claustrum, and the
amygdala as potential targets of psilocybin which are modulated by 5-HT1BR
expression. Some preclinical studies have already highlighted the amygdala, especially
the basolateral and central nucleus of the amygdala, as important loci for the therapeutic
effects of psilocybin (84–87). Given the role of the amygdala in mediating the effects of
psilocybin on threat responding (88), it is possible that 5-HT1BR expression on
amygdalar terminals underlies our reported behavioral effects. We also found that the 5-
HT1BR-effects on functional connectivity seemed to involve the amygdala as a node
given its high number of connections with other brain regions that were modulated by 5-
HT1BR expression. Our current studies are focusing on region-specific knockdown of 5-
HT1BRs in the amygdala to test their functional role in modulating the effects of
psilocybin on emotional reactivity. Projection specific studies will also investigate if
prefrontal-amygdala networks are involved, and how psilocybin may influence these
connections via 5-HT1BR since serotonin is known to gate cortical glutamatergic inputs
into the BLA via 5-HT1B receptors (89). Understanding how psilocybin affects
interregional connectivity within these networks via its actions on 5-HT1BR activation will
help identify the mechanisms through which psilocybin exerts its antidepressant effects.

Overall, our work highlights the complexity of the serotoninergic targets involved in the
therapeutic efficacy of psilocybin. Specifically, these studies add to our understanding of
the importance of non-5-HT2AR targets by implicating 5-HT1BRs in the antidepressant-
like effects of psilocybin in mice. These data contribute to the broader understanding of
the polypharmacology of psilocybin and highlight the need for multi-receptor approaches
to better understand psilocybin's mechanisms of action. Additionally, our data
demonstrate a dissociation of the acute head twitch response from the longer-term
effects of psilocybin, suggesting that different mechanisms of actions may underlie the
acute psychedelic effect of psilocybin and the persisting behavioral effects. We suggest
that psilocybin's effects are not solely driven by 5-HT2AR activation, and that activation
of other serotonin receptors are critical for its therapeutic potential. These results also
raise the potential for development of effective non-hallucinogenic pharmacotherapies
targeting 5-HT1BR for the treatment of depressive disorders.

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Figure 1: Psilocybin-induced changes in neural activity are dependent on 5-HT1BR
(A) Heatmap shows log-transformed counts of c-Fos+ cells for individual mice (columns) across
brain regions (rows) for control and 5-HT1BR knockout (KO) mice treated with saline or
psilocybin. White squares indicate missing data points. **(B)** Group means of c-Fos+ cells in six
representative brain regions following psilocybin or saline treatment. **(C)** Percent change in c-
Fos+ cells following psilocybin administration is shown in three diagrams of coronal sections of
the brain (at bregma +1.94mm, +0.98mm, and -2.18mm) in control (left hemisphere of each
diagram) and 5-HT1BR knockout (right hemisphere of each diagram) mice. Psilocybin leads to
both increases (shown in red) and decreases (blue) in c-Fos+ cell counts, with notable
differences between genotypes, calculated as relative changes in c-Fos expression compared to
saline-injected controls, with increases colored as red if they were greater than 200%. Data are
presented as mean ± SEM. *p < 0.05, **p < 0.01, ***p < 0.001. ILA: infralimbic cortex, PL:
prelimbic cortex; ORB: orbitofrontal cortex, ACA: anterior cingulate cortex, CLA: claustrum, ISO:
isocortex, COA: cortical amygdala, BLA: basolateral amygdala, BMA: basomedial amygdala,
CEA: central amygdala, LA: lateral amygdala, MEA: medial amygdala, PA: posterior amygdala
nucleus, BST: bed nucleus of the stria terminalis, LS: lateral septum, TH: thalamus, HY:
hypothalamus, ACB: nucleus accumbens, CP: caudate putamen, STR: dorsal striatum, GP:
globus pallidus, PAL: pallidum, OT: olfactory tubercle, OLF: olfactory areas, PIR: piriform cortex,
EP: endopiriform nucleus, ENT: entorhinal area, dHIP: dorsal hippocampus, vHIP: ventral
hippocampus, MB: midbrain

Figure 2: 5-HT1BR may be involved in some of the acute effects of psilocybin

(A) Head twitch response is shown for 5-HT1BR KO mice and littermate controls over the 15-
minute period following administration of saline or psilocybin. Psilocybin treatment significantly
increased the number of head twitches in both controls and KO mice compared to saline-treated
controls with no significant difference between genotypes. (B) Locomotor activity is shown for 5-
HT1BR KO mice and littermate controls mice over the 30 minutes following administration of
saline or psilocybin. (C) Area under the curve of the locomotor activity over the 30 min following
treatment summarizes the locomotor effects of psilocybin. Individual animals are shown as circles
(saline) or squares (psilocybin) and colored by sex (pink for females, blue for males). Group data
is represented as mean ± SEM. *, $p < 0.05$; ****$p < 0.0001$.

Figure 3: 5-HT1BR is required for behavioral effects of psilocybin in female mice.

(A) Experimental timeline is shown for studies testing the post-acute behavioral effects of psilocybin. **(B)** Gustometer data shows the number of licks in females across 6 concentrations of sucrose in female control and 5-HT1BR KO mice, following chronic cort with saline or psilocybin treatment, and for the no stress control condition. **(C)** Inset shows the number of licks at the 10% sucrose concentration in females. **(D)** Kaplan-Meier survival curves represent the cumulative

proportion of females eating over time in the NSF for controls (shades of black) and 5-HT1BR
KOs (shades of pink). Inset shows individual data points and average group latency to eat. **(E)**
Open arm entries are shown for females in the EPM for controls and KOs over the three
treatment groups. **(F)** Gustometer data shows the number of licks in males across 6
concentrations of sucrose in female control and 5-HT1BR KO mice, following chronic cort with
saline or psilocybin treatment, and for the no stress control condition. **(G)** Inset shows the number
of licks at the 10% sucrose concentration in males. **(H)** Kaplan-Meier survival curves represent
the cumulative proportion of males eating over time in the NSF for controls (shades of black) and
5-HT1BR KOs (shades of pink). Inset shows individual data points and average group latency to
eat. **(I)** Open arm entries are shown for males in the EPM for controls and KOs over the three
treatment groups. Data points on bar graphs represent individual mice and group averages are
shown as mean ± SEM. *p<0.05, **p<0.005, ***p<0.001

Figure 4: (A) There was no significant effect of psilocybin in mice with adult knockout of 5-
HT1BR expression, however the effect of psilocybin in control mice did not reach statistical
significance in this cohort either. **(B)** Psilocybin significantly increased entries into the open arms
of the EPM in saline-treated controls mice, suggesting reduced anxiety. However, pre-treatment
with the 5-HT1BR antagonist GR127935 blocked this effect, indicating the importance of 5-
HT1BR activation during the acute phase of psilocybin for its anxiolytic effects. #p=0.1; *p < 0.05

Figure 5. The effects of Psilocybin on behavior in a behavioral despair model are mediated by 5-HT1BR expression in both male and female mice. (A) Experimental timeline is shown for testing the post-acute behavioral effects of psilocybin following forced swim stress exposure. **(B)** Gustometer data shows the number of licks across 6 concentrations of sucrose in control and 5-HT1BR KO mice, following forced swim stress with saline or psilocybin treatment, and for the no stress control condition. **(C)** The number of licks at the 10% sucrose concentration is shown for control and 5-HT1BR KO mice in the three treatment conditions. **(D)** Open arm entries are shown in the EPM for both sexes in control and 5-HT1BR KO mice in three treatment groups. **(E)** Kaplan-Meier survival curves represent the cumulative proportion of mice eating over time in the NSF for controls (shades of black) and 5-HT1BR KOs (shades of yellow). Inset shows individual data points and average group latency to eat. Data points on bar graphs represent individual mice, with females colored pink and males colored blue. Group averages are shown as mean ± SEM. *p<0.05, **p<0.005, ***p<0.001

A. CTRL (lower triangle) and KO (upper triangle)

B.

Figure 6: 5-HT1BR expression influences select neural circuits influenced by psilocybin

(A) The difference correlation matrix shows psilocybin-induced alterations in functional connectivity between control (lower triangle) and 5-HT1BR KO mice (upper triangle). The matrix represents the pairwise correlation differences across 30 regions, illustrating desynchrony patterns between the two genotypes, with control mice exhibiting more desynchrony following psilocybin than 5-HT1BR KO mice do. (B) Circular network plot highlighting psilocybin-responsive connections most influenced by 5-HT1BR expression. Significant changes in correlated activity were observed between the CEA and the ACA (p = 0.05). Expanding the confidence interval to 90%, 85%, and 80% revealed additional differences in connectivity in CEA-CLA, ACA-GP, CEA-COA, CEA-BLA, CLA-ENT, GP-LA, PA-CEA, PA-BLA. Line width corresponds to the confidence interval, with the thickest edge representing 95% CI.

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