Title:

IntelliProfiler: a novel analytic tool for behavior dynamics of multiple animals in a

home cage

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Abstract

There is a growing demand for sophisticated behavioral analysis systems that minimize

bias in multi-animal and semi-natural environments. We present "IntelliProfiler," an

advanced system using radio frequency identification (RFID) technology to enable fully

automated behavior analyses of multiple mice within a home cage. IntelliProfiler

continuously monitors up to 16 mice, capturing both locomotor activity and social

dynamics over extended periods. Our findings revealed that male mice maintain broader

social distances than females, with group size playing a key role in shaping male social

network dynamics. Furthermore, aging in males significantly impacts both locomotor

activity and social interaction in a group size-dependent manner. With its ability to

provide in-depth analysis of individual and group behaviors, IntelliProfiler offers a

novel approach for exploring complex social interactions and group dynamics,

advancing the field of behavioral science.

Introduction

Behavioral analyses using model animals are crucial for understanding neural functions

and pathological states. Due to its ease of handling and suitability for genetic

manipulations, the laboratory mouse is widely used as a model animal to study neural

bases of various conditions, including psychiatric, neurodevelopmental, and

neurodegenerative disorders. Traditional methods for analyzing neural functions in mice

involve assessing behaviors, such as locomotion, exploration, social interaction, and

learning/memory through specific tests including the open field test, the elevated plus

maze, the social interaction test, and the Morris water maze[1–4]. However, these

traditional approaches come with several limitations.

A major limitation of traditional behavioral analyses is the low throughput of data

acquisition, as most analyses are manually conducted over limited periods under

artificial conditions. Typically, behavior is recorded for a single mouse in a specific

experimental setup. For instance, the open field test measures general locomotor activity

and anxiety-related behavior by placing a single mouse in an open arena and tracking its

movements and time spent in different zones[1]. Similarly, the three-chamber social

interaction test assesses sociability by comparing the time a mouse interacts with a

novel mouse versus an empty chamber, offering insights into social preferences[5]. A

more significant concern, however, is that interactions between experimenters and

animals often introduce biases that influence results[6–8]. Handling by experimenters can

elevate heart rate, body temperature, spontaneous activity, and anxiety, potentially

impacting memory and learning abilities[7,9]. Moreover, variations in behavioral data

have been linked to the researcher's gender and environmental conditions[8,10]. Therefore,

there is a growing need for automated, unbiased systems to analyze behavior in natural

(home-caged) conditions to ensure more robust and reliable data.

Recent trends in behavioral analyses have shifted towards studying group dynamics in

semi-natural environments. One prominent approach is the use of video tracking

systems with computer visions, such as DeepLabCut (DLC)[11–13]. DLC leverages deep

learning to achieve markerless pose estimation, offering high precision tracking with

minimal training data[14]. Despite its accuracy, video tracking systems face limitations

when tracking larger groups. Identifying individual animals becomes technically

challenging due to issues like visual cue swapping or obscuring during group behaviors,

such as huddling or fighting.

Another approach involves tracking systems using contactless tags, such as radio

frequency identification (RFID) chips. IntelliCage, a pioneering RFID system, utilizes

RFID readers placed at the corners of a cage to automatically monitor the frequency and

timing of animals entering chambers[15]. IntelliCage has been widely used to investigate

spontaneous behavior and learning in group-housed mice[16,17]. Another RFID-based

system, ECO-HAB, consists of four open fields connected by tubes, with RFID readers

positioned at the junctions, allowing the study of free movement and social interactions

among mice by tracking their movement between fields[18,19]. More recently, RFID floor

arrays have been developed to track multiple animals with higher time resolution[20].

While these RFID systems provide valuable insights into group behavior, they often

lack the spatial or temporal resolution required for more detailed analyses.

In this study, we developed an innovative RFID-based behavioral analysis system,

IntelliProfiler, to automatically monitor multiple mice in a home-cage environment.

IntelliProfiler excels in its ability, accommodating groups ranging from four to 16 mice,

while offering superior spatial and temporal resolution compared to existing systems. It

enables long-term, continuous monitoring of both activity and sociability with minimal

researcher intervention. Using IntelliProfiler, we conducted 72-hour continuous

behavioral analyses on large groups of male and female wild-type mice and compared

behavioral patterns between sexes, as well as between young and aged male mice, to

investigate the effects of aging on social dynamics. The insights gained through

IntelliProfiler provide a deeper understanding of group behavior dynamics, advancing

the precision and reliability of behavior research.

Results

Long-term activity measurement of group-housed mice

First, we set up our IntelliProfiler system using the antenna board comprising 96

antenna tiles arranged in a 12 x 8 grid (Fig. 1a). A large plastic cage (58 cm x 38 cm x

21.5 cm = width × depth × height) with a cage lid with food pellets, and three water

bottles for long-term housing was placed on the antenna board (Fig. 1a, b). The

IntelliProfiler software (see Methods) offers multiple functions: (1) data collection and

preprocessing, (2) calculation of travel distance, relative distance between two mice,

and spatial interaction proportion, (3) network analysis, (4) principal component

analysis (PCA), and (5) figure visualization. Data including time, antenna ID, and

transponder ID, were exported from the antenna board via Universal Serial Bus (USB)

using TeraTerm software (Fig. 1c). In the data collection and preprocessing step, raw

data were converted into a per-second format with time, antenna ID, transponder ID,

and X-Y coordinates using initial function of IntelliProfiler software we developed (Fig.

1d, see Data collection and preprocessing in Method section). Further calculations,

analyses, and figure visualization were conducted using the converted data generated in

the data collection and preprocessing step (see Travel distance for each mouse, Relative

distance between two mice, Spatial interaction analysis, Close contact: defining social

proximity, Network Analysis, Principal component analysis, Figure visualization in

Method section). We conducted continuous 72-hour behavioral analyses on 8-week old

male and female mice with group sizes of four, eight, and 16 mice (Fig. 1e). Due to the

dislodging of one RFID tag in the group of 16 male mice, data from 15 male mice were

used for analysis, following the methodology of a previous article[21].

In the males, cumulative travel distances for every 2 hours exhibited the highest travel

distance values in the four-mouse group than the eight- and 15-mouse groups during

most of the time (Fig. 2a). The male mice exhibited the highest travel distance

immediately after entry, dropping by less than half within 2 hours (Fig. 2a). Their travel

distance increased again during the transition from light to dark period (Fig. 2a). In

contrast, females showed less active and stable travel distances across almost all time

points (Fig. 2b).

To understand the general trends of locomotor activity in the male and female mice, we

analyzed cumulative travel distances in 12-hour intervals (Fig. 2c, d). The males

showed the highest travel distance just after their entry and significantly higher travel

distance in the group of four mice than in the groups of eight and 15 mice (Fig. 2c).

Small but significant differences were observed between the eight- and 15-mouse

groups in the light period on day 1 and the dark period on day 3 (Fig. 2c). On the other

hand, females showed similar travel distances through the period among groups except

for day 1 (Fig. 2d).

To examine sex differences in locomotor activity within the same group size, we

compared 2-hour intervals activities in the groups of four, eight, and 15/16 mice (Fig.

2e-g). In the four-mouse group, male mice showed higher travel distances throughout

the period than females, with significantly higher travel distances observed at six time

points in the dark periods (Fig. 2e). However, no periodic trend was observed in eight-

and 15/16-mouse groups (Fig. 2f, g). In the cumulative travel distances in 12-hour

intervals (Fig. 2h-j), males showed higher travel distances throughout the period in the

four-mouse group than female mice except for the light period on day 3 (Fig. 2h). On

the other hand, no noticeable sex difference was observed in the groups of eight and

15/16 mice (Fig. 2i, j).

The time course of relative locomotor activity for each mouse was visualized as a

heatmap (Fig. 2k-n). Male mice showed a marked increase in activity immediately after

the introduction and at the onset of the dark period (Fig. 2k). A pronounced contrast of

locomotor activity was observed between light and dark periods over the course of 3

days (Fig. 2m). In contrast, female mice displayed relatively more variable activity

patterns over time (Fig. 2l), with some showing elevated activity even during the light

period (Fig. 2n). These findings suggest that male mice, particularly in smaller group

sizes, exhibit higher locomotor activity, pointing to potential sex-specific behavioral

responses to group size.

Social interactions between two individuals of group-housed mice

To assess social interactions among group-housed mice, we tracked changes in the

distance between two individual mice. The spatial relationships were classified into four

categories based on proximities: "Same" - the other mouse occupied the same grid as

the focal mouse; "Close" - the other mouse was in the adjacent grids; "Intermediate" -

the other mouse was located beyond the surrounding grids; and "Away" - the other

mouse was farther than "Intermediate" (Fig. 3a). The transition in social distance ratios

were recorded in 2-hour intervals for groups of four, eight, and 15 male mice (Fig. 3b-d)

and for groups of four, eight, and 16 female mice (Fig. 3e-g).

The proportion of "Same" increased during the light period and decreased during the

dark period in the four-mouse group, but this pattern was not observed in the other

groups (Fig. 3b-g). A similar trend was found for the "Close" category, where its

proportion rose in the light period and declined in the dark period across groups of four,

eight, and 15 males, as well as four, eight, and 16 females (Fig. 3b-g). The proportion of

"Intermediate" was relatively low in the four-mouse group but increased with group

size, without showing a consistent periodic trend observed (Fig. 3b-g). In contrast, the

proportion of "Away" decreased during the light period and increased in the dark period

across all groups (Fig. 3b-g).

The average proportion of each social distance category during the light and dark

periods was compared across groups (Fig. 3b-g). The proportion of "Same" decreased as

group size increased (Fig. 3b-g). During the light period, the percentage of "Close" was

higher than during the dark period across all groups, though there was no clear trend

concerning group size (Fig. 3b-g). The proportion of "Intermediate" increased with

group size during the light period, and was higher in the groups of eight and 15/16 mice

than the four-mouse group during the dark period, but similar between the eight-mouse

and 15/16-mouse groups (Fig. 3b-g). In contrast, the proportion of "Away" was highest

in the 15-male group during both periods, although no clear pattern emerged for the

female groups. In summary, the "Same" category was more prevalent in the four-male

and four- or eight-female groups during the light period, but no consistent periodic trend

was found in the larger groups. Meanwhile, the proportion of "Away" was consistently

lower across all groups during the light periods (Fig. 3b-g). These analyses revealed

qualitatively distinct patterns of social distance in the eight- and 15/16-mouse groups

compared to the four-mouse group.

Significant differences in the proportions of each category between the light and dark

periods were observed in the groups of four males, as well as in the groups of four,

eight, and 16 females (Fig. 3b, e-g). However, no significant differences were found in

the groups of eight and 15 males (Fig. 3c, d). Regarding group size, significant

differences were noted in the male groups during both the light and dark periods, and in

the female groups during the light period (Fig. 3b-g), but no significant differences were

observed in the female groups during the dark period (Fig. 3e-g).

We then introduced the Close Contact Ratio (CCR), a metric to quantify social

proximity to analyze social interactions. This ratio reflects the proportion of time two

mice are either in the "Same" or "Close" (Fig. 4a). When CCR data were divided into 2-

hour intervals for both male and female groups (Fig. 4b, c), males in the four-mouse

group exhibited the highest CCR compared to the eight- and 15-mouse groups

throughout the entire period (Fig. 4b). In male mice, the CCR was initially low

immediately after entry, gradually increasing until 18:00, followed by a gradual decline

until 20:00, with a similar pattern recurring from day 2 onward (Fig. 4b). In contrast, the

females in the 16-mouse group showed a significantly lower CCR compared to the four-

and eight-mouse groups (Fig. 4c), although no consistent time-dependent pattern was

observed. We also analyzed the average CCR over 12-hour intervals (Fig. 4d, e). In

male mice, the CCR decreased as the group size increased from four to eight to 15 mice

during the light period (Fig. 4d). The 15-mouse group had the lowest CCR, while no

significant difference was found between the four- and eight-mouse groups during the

dark period (Fig. 4d). Similarly, female mice in the 16-mouse group tended to have the

lowest CCR during the light period compared to the four- and eight-mouse groups,

though no consistent differences were observed between the latter two groups (Fig. 4e).

No clear trend was observed during the dark period in female mice (Fig. 4e).

We also examined the sex differences in CCR using 2-hour intervals in four-, eight-,

and 15/16-mouse groups (Fig. 4f-h). In the four-mouse group, females showed a

significantly higher CCR during the initial 4 hours (Fig. 4f). However, CCR fluctuated

between males and females throughout the remaining time points, suggesting that

consistent sex differences in social proximity might not be present in this group (Fig.

4f). In contrast, in the eight- and 15/16-mouse groups, females consistently exhibited

higher CCR than males at most time points, indicating more pronounced sex differences

in social proximity (Fig. 4g, h). To assess overall trend, we analyzed the CCR in 12-

hour intervals (Fig. 4i-k). In the four-mouse group, no clear sex differences in CCR

were observed, except during the light period on day 2, when females had a higher CCR

(Fig. 4i). However, in the eight-mouse group, females had a higher CCR than males

across most periods, except during the dark period on day 3 (Fig. 4j). Similarly, in the

16-mouse group, females consistently exhibited a higher CCR than males across all the

periods (Fig. 4k).

The time course of CCR, visualized as a heatmap for individual mice (Fig. 4l-o),

revealed a strong periodic trend in the males, with CCRs peaking during the light period

and decreasing during the dark period (Fig. 4l). In contrast, the female groups generally

displayed higher CCRs for longer durations compared to males (Fig. 4l, m). The

periodic patterns of CCRs were less pronounced in the 15/16 mouse groups, though the

trend remained more apparent in males than females (Fig. 4l-o). These findings suggest

that sex-specific social dynamics become more prominent in larger groups.

Aging effect

Since aging has been reported to affect many behavioral aspects, including locomotor

activity and social interaction[22,23], we compared young (8-week-old) and aged (53-

week-old) male mice in groups of four and eight (Fig. 1e). Activity levels of the mice

were analyzed using methods as those in Fig. 2. Both young and aged males in the four-

mouse group generally had longer travel distances compared to those in the eight-mouse

group at most time points (Fig. 5a). Regarding the group effect, aged males in the four-

mouse group showed a longer travel distance immediately after their introduction and

after the next transition from the dark and light period than those in the eight-mouse

group (Fig. 5b). Regarding the age effect, there was no significant difference in travel

distance between the young and aged males (Fig. 5c). In contrast, the aged male mice in

the eight-mouse group had consistently longer travel distances at almost all time points

compared to young male mice (Fig. 5d). The differences between young and aged mice

in the eight-mouse group was also significant than other comparison between aged mice

in the four- and eight-mouse groups as well as between young and aged mice in the

four-mouse group (Fig. 5e-g).

In the heatmap analysis, activity was also prominent immediately after the introduction

and at the onset of the dark period in all groups, although aged mice in the eight-mouse

group showed more activity throughout the period (Fig. 5h, i). In the heatmap of

cumulative activity over 12-hour intervals, aged males exhibited more diverse and

desynchronized activity, particularly in the eight-mouse group (Fig. 5j, k). Therefore,

the spontaneous activity was different between young and aged mice.

Next, we examined the changes in spatial relationships among young and aged male

mice in the four- and eight-mouse groups (Fig. 6a), similar to those in Fig. 3. Regarding

group effects, aged mice in the four-mouse group exhibited lower percentage of "Same"

category during the light and dark periods compared to young mice in the four-mouse

group, while aged mice in the eight-mouse group showed lower percentage of "Away"

in both periods compared to younger counterparts (Fig. 6b-e). In contrast, aged mice in

both the four- and eight-mouse groups consistently exhibited higher percentage of

"Intermediate" during both periods compared to young mice (Fig. 6b-e). These analyses

revealed distinct social proximity patterns in aged male mice compared to young male

mice, especially in the increased proportion of "Intermediate" in the aged groups.

We then analyzed CCRs in 2-hour intervals for both young and aged male mice in the

four- and eight-mouse groups (Fig. 7a), using methods similar to those in Fig. 4. Young

males in the four-mouse group exhibited a higher CCR than those in the eight-mouse

group at most time points (Fig. 7b). In contrast, aged males in the four-mouse group

displayed a higher CCR from 18:00-20:00 on day 2 compared to those in the eight-

mouse group (Fig. 7c). Regarding the age effect, aged mice in the four-mouse group

consistently showed a lower CCR during the light period at nearly all time points

compared to young mice in the four-mouse group (Fig. 7d). On the other hand, aged

mice in the eight-mouse group displayed non-consistent periodic trends, with lower

CCR amplitude of the light-dark cycle, compared to young mice in the eight-mouse

group (Fig. 7e). The CCR differences between young and aged mice in the four-mouse

group during the light period was also consistently more significant than those in the

comparisons between aged mice in the four- and eight-mouse groups, as well as

between young and aged mice in the eight-mouse group (Fig. 7f-h). In the heatmap

analysis, the young mice exhibited robust periodic trends, with elevated CCR values

during the light period and decreased values during the dark period, particularly in the

four-mouse group (Fig. 7i). In contrast, the aged mice in both the four- and eight-mouse

groups showed more stable, intermediate CCR values compared to young mice (Fig. 7j).

In the heatmap of cumulative activity over 12-hour intervals, aged males demonstrated

more ambiguous cyclic CCR patterns, particularly in the eight-mouse group (Fig. 7k, l).

These findings suggest that aging in male mice altered not only locomotor activity but

also the temporal dynamics of social behaviors.

Characterization of group behaviors

To investigate the behavioral characteristics of each group, we conducted principal

component analysis (PCA) using four key parameters. First, we defined normalized

activity as the activity at time t, divided by the maximum activity value recorded over

Days 1-3, and multiplied by 100 to standardize the data. This allowed us to calculate the

normalized activity in both the light (AL) and dark (AD) periods by averaging activity

values at each time point over three days. We selected four parameters for PCA:

sociability, measured as CCR values during the light (SL) and dark (SD) periods

(parameters 1 and 2, respectively), and normalized activity during the light (AL) and

dark (AD) periods (parameters 3 and 4, respectively). Correlations among these

parameters are shown in Supplementary Fig. 1, 2, while the PCA results with estimated

clusters are displayed in Fig. 8. The number of clusters was determined using the Elbow

method[24] and further assessed through Silhouette plots[25] (Supplementary Fig. 3).

The PCA revealed distinct sex-biased groupings. Notably, the male mice in the 15-

mouse group formed a well-defined separated cluster, whereas the male and female

mice in the four-mouse groups and female mice in the eight-mouse group clustered

together (Fig. 8a). Interestingly, female mice in the 16-mouse group split into two

clusters, with one cluster aligning with the male mice in the eight-mouse group (Fig.

8b). This suggests that while smaller groups tend to blend, significant sex differences

emerge in larger groups. In the feature space, AL and AD vectors pointed in the same

direction, indicating similar patterns of activity, while SL and SD vectors diverged,

reflecting different sociability dynamics between light and dark periods (Fig. 8c).

In terms of aging effects, young and aged mice in the four-mouse groups were largely

positioned in the positive X and Y directions, whereas those in the eight-mouse groups

clustered near the origin (0,0) or in the negative X direction (Fig. 8d). K-means

clustering effectively separated the four- and eight-mouse groups (Fig. 8e), and each

feature vector had a distinct direction (Fig. 8f), indicating that aging influenced both

activity and sociability in distinct ways, aligning with our earlier observations (Fig. 5-

7).

Network analysis of social interactions

We explored how group size influences social interaction networks (Fig. 9).

Qualitatively, there was no noticeable difference in network structure between male and

female mice in the four- and eight-mouse groups (Fig. 9). However, in the 15/16-mouse

groups, male mice exhibited increased social proximity and developed more expanded

networks, influenced by light-dark circadian rhythms. In contrast, female mice

maintained relatively uniform network structures (Fig. 9). When comparing young and

aged male mice in the four-mouse groups, no significant differences in network

structure were observed (Fig. 10). In the eight-mouse groups, however, young male

mice displayed increased social proximity and formed diverse networks that fluctuated

with light-dark cycles, while aged male mice maintained more stable, uniform networks

with only slight variation corresponding to circadian rhythms (Fig. 10*).* These findings

suggest that male and female mice create distinct social environments, and that aging

may impact the complexity and heterogeneity of the networks in larger groups.

Analysis of individual social interactions

Lastly, we focused on individual differences in social interactions through a pairwise

analysis of CCR. In the heatmap presentations (Supplementary Fig. 6), no obvious

individual differences were observed between sexes in the four- and eight-mouse

groups. However, in the 15/16-mouse groups, two males (mouse ID #1 and #2)

exhibited lower CCR scores compared to the others, while all female mice showed

similar CCR patterns. Additionally, male mice tended to display higher CCR during the

light period, which decreased during the dark period, whereas female mice maintained

relatively stable CCR levels with less fluctuation across the light and dark periods,

indicating a sex-dependent response to light. Regarding the aging effect, aged males

showed lower CCR scores and more ambiguous, less defined time-dependent patterns

compared to young males (Supplementary Fig. 7), consistent with mean values

presented in Fig. 7. These findings suggest that individuality may develop progressively

with aging in males, potentially contributing to the observed social dynamics.

Discussion

In this study, we developed a novel RFID-based behavioral analysis system,

IntelliProfiler, which allows for continuous, unbiased monitoring of up to 16 mice

within the home-cage environment. This system has provided clear insights into the

effects of group size, sex, and aging on both locomotor activity and social interactions,

offering a more comprehensive understanding of the semi-natural behaviors of group-

housed mice than using traditional behavioral methods can achieve.

One of our key findings was the impact of group size on behavior. IntelliProfiler

revealed significant differences in activity levels and social proximity, particularly

among males, where larger groups reduced activity. Traditional behavior paradigms,

such as the open-field test, are typically designed for individual animals[26,27], and

therefore fail to capture the complexities of behavior in group settings. These

conventional paradigms may overlook critical dynamics unique to multi-animal

environments. A recent article in *Nature Digest* emphasized the importance of studying

animals under 'natural conditions' to gain deeper insights into their behavior[28].

IntelliProfiler enables us to monitor authentic locomotor activity under semi-natural

conditions, offering a more comprehensive understanding of how group size shapes

individual activity and social dynamics.

In this study, we introduced CCR as a novel social parameter to measure social

proximity, representing a significant advancement in behavioral analysis (Fig. 4).

Traditional social interaction tests, such as the three-chamber test, offer limited insight

into short-term interactions, but the CCR provides a more detailed, long-term view of

group dynamics. Notably, we found that females tended to form stable clusters, while

males exhibited more fluid, group size-dependent social patterns (Fig. 4, 8, 9). This

underscores IntelliProfiler's ability to detect subtle, yet important sex-specific social

behaviors, often overlooked by conventional methods.

Aging also influenced locomotor activity and social interactions, with aged males

displaying less distinct and more variable CCR patterns compared to young males. This

finding aligns with previous research on aging and social behavior,[23,29] but also suggests

that IntelliProfiler offers a more nuanced understanding of how aging affects social

dynamics, especially in larger groups (Fig. 7).

IntelliProfiler further provides an inexpensive, high-throughput platform for social

network analysis, surpassing the limitations of human observation. Although sexual

interaction between males and females is beyond our current purpose, our system would

be applicable to analyze sexual or courtship behaviors in large groups. While video

tracking systems have been useful for analyzing small groups, IntelliProfiler extends

this capability to larger groups, making it a crucial tool for studying population-level

behaviors.

Despite its advantages, IntelliProfiler has a few limitations. First, its spatial resolution is

lower than video-based tracking systems like DLC[11,30], as the current grid size is 5 cm x

5 cm, allowing two mice located in parallel. Additionally, determining body orientation

is challenging when the animal remains within the same grid. Second, the system has

difficulty distinguishing detailed social contacts, such as sniffing, mounting, or

huddling. Lastly, the current RFID system requires surgery for transponder

implantation, and it is not yet possible to eliminate the invasive effects due to the size of

commercially available transponders. These challenges should be addressed in future

developments.

Conclusion

The IntelliProfiler makes a significant advancement in animal behavior research,

enabling continuous, detailed, and unbiased monitoring of individual movements and

interactions within dynamic and complex social environments. Our findings highlight

the critical roles of group size, sex, and aging in shaping behavior. This system provides

a powerful tool for studying behavior in more naturalistic settings, offering new avenues

for large-scale behavioral research and deeper insights into social dynamics.

Methods

Animals

C57BL/6J mice at 7- or 52-week old were purchased from CLEA Japan, Inc., and

maintained in the animal facility at Tohoku University Graduate School of Medicine.

An RFID tag (7 mm x 1.25 mm, Phenovance LCC) was subcutaneously implanted into

the abdomen of each mouse under anesthesia using 2.0% isoflurane (MSD Japan). The

mice were recovered for 1 week before starting behavioral recording. The recordings

started at 8- or 53-week old. All experimental procedures were approved by the Ethics

Committee for Animal Experiments at the Tohoku University Graduate School of

Medicine (approval number: 2021MdA-020-13).

Hardware

The antenaboard, comprising 96 antenna tiles arranged in 12x8 grids (5 cm by 5 cm

each) (Fig. 1a), was purchased from Phenovance LCC, Japan. The home cage used for

the study was 2000P cage (58 cm x 38 cm x 21.5 cm = width × depth × height,

Tecniplast, Italy), equipped with a cage lid with food pellets, and three water bottles for

long-term housing (Fig. 1a, b). Both the antenaboard and home cage were placed inside

a soundproof box (Phenovance LCC). The light-dark cycle in the soundproof box and

testing room was 12:12 hours (08:00-20:00, light; 20:00-08:00, dark). Mice were

introduced at 8:00 in the light period on day 1.

Software

R language (Version 4.2.2) was mainly used for preprocessing, data analysis, and

visualization[31]. Python (Version 3.9.7), Prism 9, and Microsoft Excel (Version 16.86)

were also used for preprocessing, data analysis, and visualization. R and Python

packages required for the scripts are listed in Supplementary Table 1.

Data collection and preprocessing

Data including time, antenna ID, and transponder ID, were exported from the

antenabord via Universal Serial Bus (USB) using TeraTerm software (Fig. 1c). The raw

data were converted into a format per second with time, antenna ID, transponder ID,

and X-Y coordinates (Fig. 1d). Missing values were interpolated by the immediately

preceding value.

Travel distance for each mouse

The Euclidean distances of mouse movement were calculated from consecutive RFID

positions at time t and t+1. This was pivotal in understanding the spatial dynamics of

the subjects. Graphical representations of tracked movements, including 2D positional

plots and time series plots of X and Y coordinates, were generated to visualize

trajectories and movement patterns over time.

Relative distance between two mice

The distances between different pairs of mice were calculated from the Euclidean

distance between RFID positions. The total travel distance for each mouse was also

calculated and plotted over the experimental period, providing a quantitative measure of

activity levels.

Spatial interaction analysis

Spatial relationships of paired mice on the XY plane of the IntelliProfiler were

classified, based on the distance L between the paired mice, into four categories (Fig.

3a). "Same" indicates that two mice were in the same grid ($L=0$ (cm)), suggesting

immediate proximity. "Close" represents two mice were in adjacent grids ($L=5$ or $5\sqrt{2}$

(cm)), demonstrating close but distinct positions. "Intermediate" ($L=10$, $5\sqrt{5}$ or $10\sqrt{2}$

(cm)) places one mouse on the outer perimeter of "Close," while "Away" ($L > 10\sqrt{2}$

(cm)) signifies that one mouse was outside of the range of "Intermediate," indicating

highly separation.

Close contact: defining social proximity

To examine social interactions within a group of mice, we defined the Close Contact

Ratio (CCR) as the cumulative percentage of time a pair of mice spent in the same grid

(defined as "Same") or adjacent grids (defined as "Close"). This measure serves as a

quantitative score of sociality between two mice, indicating their tendency to remain in

immediate or proximity. Utilizing Excel and Prism 9 (GraphPad Software), we

calculated the proportionate shifts of social distances ("Same," "Close," "Intermediate,"

and "Away") and CCR every 2 hours and in light and dark periods.

Network Analysis

Using Cytoscape 3.9.1[32], we conducted network analyses for four-, eight-, and 15/16-

mouse groups. We imported values ten times the CCR as edge betweenness into

Cytoscape 3.9.1. The network layout was generated using the Edge-weighted Spring

Embedded Layout, facilitating the visual interpretation of social interactions among the

mice based on their proximity metrics[32,33].

Principal component analysis

Principal component analysis (PCA) was conducted using Python 3 (Version 3.9.7) on

four datasets derived from long-term behavior mice in large groups under light and dark

periods, focusing on relative activity levels and CCR. Three distinct types of PCA plots

were generated: 1) PCA contribution vectors displayed on the PCA plot; 2) K-means

clustering analysis, with the optimal number of clusters determined using the elbow

method[24] and silhouette method[25] applied to the PCA plot; and 3) PCA plots color-

coded by gender and genotype to distinguish different groups.

Figure visualization

Graphical representations were primarily created using Prism 9 (GraphPad Software).

Time series line graphs and heat maps of travel distances or cumulative CCRs as well as

transitions in the proportionate of social distances ("Same," "Close," "Intermediate,"

and "Away") were visualized. Pie charts illustrating the distribution across social

distances were also developed. The PCA plots were designed in Python 3, and the social

network graphs for the groups were constructed using Cytoscape version 3.9.1.

Statistical analysis

Statistical analyses were performed with Prism version 9 (GraphPad). Two-way

ANOVA followed by Holm-Sidak's multiple comparisons test was applied to assess

differences between the two groups, which has been presented in part in a previous

article[34]. Values in the bar or line graphs were presented as mean $\pm$ SD, and $p* < 0.05$,

$p** < 0.01$, $p*** < 0.001$ indicated statistically significant differences. The p-values, F

values and degrees of freedom for all results, are provided in Supplementary Table 2.

All experiments were performed in a single repetition.

Data availability

The behavioral data generated in this study are provided in Supplementary Information.

Code availability

All code and scripts used for data analysis in this study are available on GitHub. To

access the public repository, please use the following access token along with the

repository URL:

- Repository URL: https://github.com/ShoheiOchi/IntelliProfiler

References

1.   Hall, C. S. Emotional behavior in the rat. I. Defecation and urination as measures

529   of individual differences in emotionality. *J Comp Psychol* **18**, 385–403 (1934).

530   2.   Pellow, S., Chopin, P., File, S. E. & Briley, M. Validation of open:closed arm

531   entries in an elevated plus-maze as a measure of anxiety in the rat. *J Neurosci*

532   *Methods* **14**, 149–167 (1985).

533   3.   File, S. E. & Hyde, J. R. Can social interaction be used to measure anxiety? *Br. J.*

534   *Pharmac* **62**, 19–24 (1978).

535   4.   Morris, R. Developments of a water-maze procedure for studying spatial learning

536   in the rat. *J Neurosci Methods.* **11**, 47–60 (1984).

537   5.   Moy, S. S. *et al.* Sociability and preference for social novelty in five inbred

538   strains: an approach to assess autistic-like behavior in mice. *Genes Brain Behav*

539   **3**, 287–302 (2004).

540   6.   Balcombe, J. P., Barnard, N. D. & Sandusky, C. Laboratory routines cause

541   animal stress. *Contemp Top Lab Anim Sci* **43**, 42–51 (2004).

542   7.   Gouveia, K. & Hurst, J. L. Optimising reliability of mouse performance in

543   behavioural testing: The major role of non-aversive handling. *Sci Rep* **7:44999**,

544   (2017).

545   8.   Georgiou, P. *et al.* Experimenters' sex modulates mouse behaviors and neural

546   responses to ketamine via corticotropin releasing factor. *Nat Neurosci* **25**, 1191–

547   1200 (2022).

548   9.   Hurst, J. L. & West, R. S. Taming anxiety in laboratory mice. *Nat Methods* **7**,

549   825–826 (2010).

550   10.   Sorge, R. E. *et al.* Olfactory exposure to males, including men, causes stress and

551   related analgesia in rodents. *Nat Methods* **11**, 629–632 (2014).

552   11.   Mathis, A. *et al.* DeepLabCut: markerless pose estimation of user-defined body

553   parts with deep learning. *Nat Neurosci* **21**, 1281–1289 (2018).

554   12.   de Chaumont, F. *et al.* Real-time analysis of the behaviour of groups of mice via

555   a depth-sensing camera and machine learning. *Nat Biomed Eng* **3**, 930–942

556   (2019).

557   13.   Hakataya, S., Katsu, N., Okanoya, K. & Toya, G. An exploratory study of

558   behavioral traits and the establishment of social relationships in female

559   laboratory rats. *PLoS One* **18**, (2023).

560   14.   Ye, S. *et al.* SuperAnimal pretrained pose estimation models for behavioral

561   analysis. *Nat Commun* **15**, (2024).

562   15.   Lipp, H.-P. High-throughput and automated behavioural screening of normal and

563   genetically modified mice. *Business Briefing: Future Drug Discovery* 1–5

564   (2005).

565   16.   Endo, T. *et al.* Automated test of behavioral flexibility in mice using a behavioral

566   sequencing task in IntelliCage. *Behavioural Brain Research* **221**, 172–181

567   (2011).

568   17.   Kiryk, A. *et al.* IntelliCage as a tool for measuring mouse behavior – 20 years

569   perspective. *Behavioural Brain Research* **388**, (2020).

570   18.   Chen, X. *et al.* Modelling collective behavior in groups of mice housed under

571   semi-naturalistic conditions. *bioRxiv* (2024) doi:10.1101/2023.07.26.550619.

572   19.   Winiarski, M. *et al.* Social deficits in BTBR T+ Itpr3tf/J mice vary with

573   ecological validity of the test. *Genes Brain Behav* **21**, (2022).

574   20.   Lipp, H. P. *et al.* IntelliCage: the development and perspectives of a mouse- and

575   user-friendly automated behavioral test system. *Front Behav Neurosci* **17**,

576   (2024).

577 21. Smutek, M. *et al.* A model of alcohol drinking under an intermittent access

578   schedule using group-housed mice. *PLoS One* **9**, (2014).

579 22. Yanai, S. & Endo, S. Functional Aging in Male C57BL/6J Mice Across the Life-

580   Span: A Systematic Behavioral Analysis of Motor, Emotional, and Memory

581   Function to Define an Aging Phenotype. *Front Aging Neurosci* **13**, (2021).

582 23. Shoji, H., Takao, K., Hattori, S. & Miyakawa, T. Age-related changes in

583   behavior in C57BL/6J mice from young adulthood to middle age. *Mol Brain* **9**,

584   (2016).

585 24. Thorndike, R. L. Who belongs in the family? *Psychometrika* **18**, 267–276 (1953).

586 25. Rousseeuw, P. J. Silhouettes: A graphical aid to the interpretation and validation

587   of cluster analysis. *J Comput Appl Math* **20**, 53–65 (1987).

588 26. Yoshizaki, K. *et al.* Paternal aging affects behavior in Pax6 mutant mice: A

589   gene/environment interaction in understanding neurodevelopmental disorders.

590   *PLoS One* **11**, (2016).

591 27. Crawley, J. N. Exploratory behavior models of anxiety in mice. *Neurosci*

592   *Biobehav Rev* **9**, 37–44 (1985).

593 28. Smith, K. Lab mice go wild: making experiments more natural in order to decode

594   the brain. *Nature* **618**, 448–450 (2023).

29.   Shoji, H. & Miyakawa, T. Age-related behavioral changes from young to old age

in male mice of a C57BL/6J strain maintained under a genetic stability program.

Neuropsychopharmacol Rep **39**, 100–118 (2019).

30.   Lauer, J. *et al.* Multi-animal pose estimation, identification and tracking with

DeepLabCut. *Nat Methods* **19**, 496–504 (2022).

31.   R Core Team. R: A Language and Environment for Statistical Computing. R

Foundation for Statistical Computing, Vienna. (2021).

32.   Shannon, P. *et al.* Cytoscape: A software Environment for integrated models of

biomolecular interaction networks. *Genome Res* **13**, 2498–2504 (2003).

33.   Otasek, D., Morris, J. H., Bouças, J., Pico, A. R. & Demchak, B. Cytoscape

Automation: Empowering workflow-based network analysis. *Genome Biol* **20**,

(2019).

34.   Endo, N. *et al.* The effects of maternal separation on behaviours under social-

housing environments in adult male C57BL/6 mice. *Sci Rep* **11**, (2021).

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Contributions

Conceptualization, S.O., H.I. and N.O.; behavior analysis, S.O.; data process, S.O. and

H.I.; code preparation, S.O. and H.I.; figure preparation, S.O.; writing—review and

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version of the manuscript.

Competing interests

629 The authors declare that this study was conducted in the absence of any commercial or

630 financial relationships that could be construed as a potential conflict of interest.

Figures

Fig. 1: IntelliProfiler automated behavioral measurement system.

a Schematic of the IntelliProfiler. The system tracks the location of mice implanted with

RFID tags via detectors placed in a 96-grid array at the base, with each grid measuring 5

cm x 5 cm. Location information is converted into XY coordinates (X=12, Y=8), based

on antenna IDs (1-96). **b** Photograph of the IntelliProfiler. **c** Original data, capturing

information on Time, Antenna ID, and Transponder ID. **d** Processed data converted for

the IntelliProfiler. The data is refined to includ Time, Antenna ID, Transponder ID, and

XY coordinates. Decimal information is removed from the time data. Iif multiple

individuals are detected on the same antenna ID simultaneously, their information is

separated into different rows. **e** Behaior analysis was conducted over a continuous 72-

hour period for male and female mice in groups of four-, eight- and 15/16. Additionally,

young (8-week old) and aged (53-week old) male mice were analyzed in groups of four

and eight.

Fig. 2: Activity analysis of group-housed mice using the IntelliProfiler.

a-j Cumulative travel distances measured in 2-hour intervals (**a, b, e-g**) and over 12-

hour intervals (light-dark cycles) (**c, d, h-j**). **a-d** Analysis of locomotor activity based on

group size, comparing groups of four, eight, and 15 males (**a, c**) and four, eight, and 16

female mice (**b, d**). **e-j** Analysis of sex differences in locomotor activity, comparing

groups of four (**e, h**), eight (**f, i**) and 15-16 (**g, j**) male and female mice. **k-n** Heatmaps

showing relative locomotor activity over 2-hour intervals (**k, l**) and 12-hour intervals

(**m, n**) for groups of four, eight, and 15-16 mice. Values in the bar or line graphs are

presented as mean ± SD, with box plots displaying min/max whiskers. Statistical

analysis was conducted using Holm-Sidak method for multiple comparison.

Significance levels are indicated as *$p < 0.05$, **$p < 0.01$, and ***$p < 0.001$. $p < 0.001$

(**c**), $p = 0.011$ (**d**), $p < 0.001$ (**e**), $p = 0.894$ (**f**), $p = 0.009$ (**g**), $p < 0.001$ (**h**), $p = 0.920$ (**i**),

$p = 0.139$ (**j**).

Fig. 3: The spatial relationships among group-housed mice using the

IntelliProfiler.

a Definition of the spatial relationships between two individual mice. When another

mouse is in the same grid as the focal mouse (both depicted in yellow), the relationship

is defined as "Same." If the other mouse is in the surrounding grids (depicted in green),

it is defined as "Close." If the other mouse is outside the surrounding grids (depicted in

light blue), it is defined as "Intermediate." If the other mouse is beyond the

"Intermediate" zone (depicted in blue), the relationship is categorized as "Away." **b-g**

The 2-hour bin transition of proportions based on "Same," "Close," "Intermediate," and

"Away" for groups of four male mice (**b**), eight male mice (**c**), 15 male mice (**d**), four

female mice (**e**), eight female mice (**f**), and 16 female mice (**g**). The average proportion

of distance categories during the light period and the dark period is shown for each

group. Chi-square p-values comparing the proportions between light and dark periods: p

< 0.001 (**b**, four males), $p = 0.256$ (**c**, eight males), $p = 0.114$ (**d**, 15 males), $p = 0.005$

(**e**, four females), $p = 0.049$ (**f,** eight females), $p = 0.007$ (**g**, 16 females). The p-values of

chi-square for the group size, $p < 0.001$ (**b-d**, males in the light phase), $p = 0.002$ (**b-d**,

males in the dark phase), $p < 0.001$ (**e-g**, females in the light phase), $p = 0.092$ (**e-g**,

females in the dark phase).

Fig. 4: Close contact between two individuals among group-housed mice analyzed

using the IntelliProfiler.

a Definition of close contact, where two mice either in the "Same" grid (depicted in

yellow) or in adjacent grids, classified as "Close" (depicted in green). **b-k** Average

Close Contact Ratio (CCR) measured in in 2-hour intervals (**b, c, f-h**) and 12-hour

intervals (**d, e, i-k**). **b-e** Analysis of CCR based on group size, comparing groups of

four, eight, and 15 male mice (**b, d**) and four, eight, and 16 female mice (**c, e**). **f-k**

Analysis of sex differences in CCR, comparing groups of four (**f, i**), eight (**g, j**) and

15/16 (**h, k**) male and female mice. **l-o** Heatmaps showing the time course of CCR over

2-hour intervals (**l, m**) and 12-hour intervals (**n, o**) for groups of four, eight, and 15-16

mice. Values in the bar or line graphs are presented as mean $\pm$ SD, with box plots

showing min/max whiskers. Statistical analysis was conducted using Holm-Sidak

method for multiple comparisons. Significance levels are indicated as *$p < 0.05$, **$p <$

0.01, and ***$p < 0.001$. $p < 0.001$ (**b**), $p < 0.001$ (**c**), $p = 0.882$ (**f**), $p < 0.001$ (**g**), $p <$

0.001 (**h**), $p = 0.873$ (**i**), $p < 0.001$ (**j**), $p < 0.001$ (**k**).

Fig. 5: Activity analysis of group-housed aged male mice using the IntelliProfiler.

a-g Cumulative travel distances measured in 2-hour intervals (**a-d**) and over 12-hour

intervals (**e-g**). **b, e** Analysis of locomotor activity based on group size, comparing

groups of four and eight aged mice. **c, d, f, g** Analysis of aging effects in locomotor

activity, comparing groups of four young and aged male mice (**c, f**), groups of eight

young and aged male mice (**d, g**). **h-k** Heatmaps showing relative locomotor activity

over 2-hour intervals (**h, i**) and 12-hour intervals (**j, k**) for groups of four and eight

mice. Values in the bar or line graphs are presented as mean ± SD, with box plots

displaying min/max whiskers. Statistical analysis was conducted using Holm-Sidak

method for multiple comparisons. Significance levels are indicated as *$p < 0.05$, **$p <$

0.01, and ***$p < 0.001$. $p = 0.338$ (**c**), $p < 0.001$ (**d**), $p = 0.585$ (**f**), $p < 0.001$ (**g**).

Fig. 6: The spatial relationships among group-housed aged male mice using the

IntelliProfiler.

a Definition of the spatial relationships between two individual mice. When another

mouse is in the same grid as the focal mouse (both depicted in yellow), the relationship

is defined as "Same." If the other mouse is in the surrounding grids (depicted in green),

it is defined as "Close." If the other mouse is outside the surrounding grids (depicted in

light blue), it is defined as "Intermediate." If the other mouse is beyond the

"Intermediate" zone (depicted in blue), the relationship is categorized as "Away." **b-e**

The 2-hour bin transition of proportions based on "Same," "Close," "Intermediate," and

"Away" for groups of four young male mice (**b**), eight young male mice (**c**), four aged

male mice (**d**), eight aged male mice (**e**). **b-e** The average proportion of distance

categories during the light period and the dark period is shown for each group. Chi-

square p-values comparing the proportions between light and dark periods, $p < 0.001$ (**b**,

four young males), $p = 0.256$ (**c**, eight young males), $p = 0.836$ (**d**, four aged males), $p =$

0.439 (**e**, eight aged males). The p-values of chi-square for the group size, $p < 0.001$ (**b**,

c, young males in the light phase), $p = 0.020$ (**b, c**, young males in the dark phase), $p =$

0.115 (**d, e**, aged males in the light phase), $p = 0.477$ (**d, e**, aged males in the dark

phase).

Fig. 7: Close contact between two individuals among group-housed aged male mice

analyzed using the IntelliProfiler.

a Definition of close contact, where two mice either in either "Same" (depicted in

yellow) or in adjacent grids, classified as "Close" (depicted in green). **b-h** Average

Close Contact Ratio (CCR) measured in 2-hour intervals (**b-e**) and 12-hour intervals (**f-**

h). **c, f** Analysis of CCR based on group size, comparing groups of four and eight young

and aged males. **d, e, g, h** Analysis of CCR based on aging effects, comparing groups of

four young and aged male mice (**d, g**), groups of eight young and aged male mice (**e, h**).

i-l Heatmaps showing the time course of CCR over 2-hour intervals (**i, j**) and 12-hour

intervals (**k, l**) for groups of four and eight mice. Values in the bar or line graphs are

presented as mean $\pm$ SD, with box plot showing min/max whiskers. Statistical analysis

was conducted using Holm-Sidak method for multiple comparisons. Significance levels

are indicated as *$p < 0.05$, **$p < 0.01$, and ***$p < 0.001$. $p < 0.001$ (**c**), $p = 0.338$ (**d**), p

$= 0.072$ (**e**), $p < 0.001$ (**f**), $p = 0.007$ (**g**), $p = 0.109$ (**h**).

Male - Female

Aged - Young

750   **Fig. 8: Principal component analysis of group behavior using the IntelliProfiler.**

751   **a-f** Principal component analysis (PCA) of group behavior in four, eight, and 15/16

752   male and female mice (**a-c**) and in young and aged mice in groups of four and eight (**d-**

753   **f**). **b, e** K-means clustering in PCA plots. **c, f** Analysis of feature vectors, focusing on

754   Close Contact Ratio (CCR) and relative activity during the light and dark periods.

Fig. 9: Network analysis of social interaction in the male and female mice using the

IntelliProfiler.

Network analysis of social interactions during the light period on day 1 and the dark

period on day 3 in groups of four, eight, and 15/16 male and female mice. Nodes

represents individual mice, and edges reflects the relative social contacts between two

individuals. The network graph was generated using Cytoscape. Supplementary Fig. 4

provides additional network diagrams, where unique color code for each individual

highlight the individual-specific social dynamics within the network.

Fig. 10: Network analysis of group interaction in young and aged male mice using

the IntelliProfiler.

Network analysis of social interactions during the light period on day 1 and the dark

period on day 3 in groups of four and eight young and aged males. Each circle

represents an individual mouse, and the proximity of the lines indicates the relative

social distance between two individuals. The network graph was generated using

Cytoscape. Supplementary Fig. 5 provides further details, offering a clear understanding

of individual-specific social dynamics across the networks.