Population genomics of the invasive Argentine ant (*Linepithema humile*) – adaptive evolution in introduced

supercolonies despite low genetic diversity

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Abstract

The Argentine ant (*Linepithema humile*), native to South America, has spread globally over the past 150

years, forming extremely large supercolonies in its introduced range. Despite the recent demographic

history, including founder effects, Argentine ants thrive in the introduced range and displace native ant

species. We adopted a comprehensive approach to investigate selection signals across the whole genome

of this highly invasive species, with a specific focus on introduced supercolonies. We also investigated

genome-wide diversity and divergence patterns and compared the results with earlier microsatellite

marker data studies. We used pooled whole-genome sequence data from 100 workers from the species'

native range and each of the three invasive supercolonies – European Main, Catalonia, and Chile. Invasive

supercolonies harboured low genetic diversity and were genetically highly differentiated. Despite this, we

could detect signs of positive selection in their genomes. Some of the selected genes showed parallel

adaptive evolution in invasive supercolonies. Furthermore, there were no signs that a social organisation

based on unrelated workers had led to impaired adaptability (genetic meltdown). We conclude that

introduced Argentine ant supercolonies evolve adaptively, indicating that founder effects, low genetic

diversity and supercolony social organisation do not always hamper species adaptability.

Keywords: pool-seq, demographic history, founder effect, genetic drift, selective pressure

    1. Introduction

Invasive species increasingly threaten biodiversity and ecosystems globally (Mollot et al., 2017; Seebens et

al., 2017). In the most harmful situations, they cause local extinctions, permanently changing the nature of

colonised areas (Holway et al., 2002; Mack et al., 2000). However, the factors contributing to the success of

invasive species are still poorly understood, as are their long-term evolutionary prospects in the new

ranges, which makes the possibility of controlling these species and mitigating their detrimental effects

challenging (Estoup et al., 2016; Pérez et al., 2006; Sherpa & Després, 2021).

Ants exhibit enormous species richness, inhabit diverse ecological niches, and play a crucial role in many

ecosystems (Hölldobler & Wilson, 1990; Moreau et al., 2006; Wilson & Hölldobler, 2005). Many ant species

are also among the most harmful invasive species (Holway et al., 2002; Lester & Gruber, 2016; McGlynn,

1999). One of these, the highly invasive Argentine ant (*Linepithema humile*), has spread, by accidental

human introduction, from its original habitat in South America to all continents except Antarctica during

the past 150 years (Suarez et al., 2001; Wetterer et al., 2006, 2009; Wetterer & Wetterer, 2006). Typical of

invasive ant species, the Argentine ant also forms large polygynous (multi-queen) and polydomous (multi-

nest) societies, known as supercolonies (Pedersen et al., 2006), in which territorial borders between nests

are absent over large areas, allowing individuals to move freely between them. In both the native and

introduced ranges, these supercolonies form closed breeding units where mating occurs within the nest,

and colonies spread by budding (Helanterä, 2022; Pedersen et al., 2006; Vogel et al., 2009). In the

introduced range, Argentine ants dominate competition between species, destroy local ecosystems, and

cause species extinctions (Giraud et al., 2002; Holway et al., 2002; Sanders et al., 2001, 2003). The

Argentine ants' supercolony size in the introduced range can be enormous, e.g., the European Main

supercolony spans over 6,000 kilometres from the Mediterranean coast in Italy to the Spanish Atlantic

coast (AntWeb, 2026; Giraud et al., 2002).

Previous research using microsatellite marker data indicated that introduced Argentine ant supercolonies

harbour low genetic diversity and are genetically highly differentiated from each other and from the native

range supercolonies (Blight et al., 2012; Brandt et al., 2009; Giraud et al., 2002; Jaquiéry et al., 2005; Tsutsui

et al., 2000; Vogel et al., 2010). These introduced supercolonies originate from a small number, in some

cases only a dozen, of reproducing individuals (Giraud et al., 2002; Vogel et al., 2010), indicating strong

founder effects, i.e., loss of genetic variation. Genetic drift (random fluctuations in allele frequencies, often

causing allele fixation or loss) is amplified in these small, newly established founder populations, leading to

even greater differentiation between populations (Keller & Passera, 1993; Nei et al., 1975). Low initial

diversity, combined with strong drift driven by small effective population sizes, should impede adaptive

evolution in the introduced Argentine ant supercolonies. Moreover, the general inefficiency of selection on

worker traits in supercolonies with low relatedness, due to multiple egg-laying queens, increases the

accumulation of deleterious mutations and decline in fitness (genetic meltdown) (Helanterä et al., 2009;

Linksvayer & Wade, 2009). However, the success and dominance of Argentine ants in the introduced range

(Vogel et al., 2010) suggest local adaptation to the new environments. This success in novel ranges despite

small effective population sizes and decreased selection efficacy raises an important question about the

effects of natural selection on the evolution of these ants.

Our earlier population genetic study, focusing on the evolution of immune genes in 18 Argentine ant

supercolonies worldwide, including the invasive supercolonies of this study, indicated that, despite recent

demographic history, positive selection has affected the evolution of immune genes in the introduced

range (Holmberg et al., 2024). In this study, we adopted a more comprehensive approach to investigate

signals of selection across the whole genome. We studied native-range supercolonies and three invasive

supercolonies from different parts of the world using pooled whole-genome sequence data from 100

workers from the native range and from each of the three invasive supercolonies. We aimed to investigate:

i) the overall genome-wide patterns of selection in invasive Argentine ants, i.e., whether signatures of

selection could be detected in their genomes, ii) signatures of parallel adaptive evolution among the three

invasive supercolonies, and iii) whether genome-wide data confirms the earlier findings, based on

microsatellite marker data, about demographic changes during introductions, supercolonial diversity and

divergence.

2. Materials and Methods

2.1. Sampling

We analysed four samples of Argentine ants. One of the samples represents native-range supercolonies, and three samples represent invasive supercolonies, one originating from South America and two from Europe (Table S1) (Wild, 2004). All samples include 100 diploid workers. The sample representing the native-range supercolonies was collected in Argentina, while the samples representing the three invasive supercolonies were collected in France, Spain and Chile. The three sampled invasive supercolonies arose from different primary introductions from the native range (Vogel et al., 2010). The two European samples were collected from the two known, separate supercolonies: the European Main supercolony (European Main; establishment recorded 1906) and the Catalonian supercolony (Catalonia; 1916), which live partly next to each other along the Mediterranean coastline (Giraud et al., 2002). The third invasive sample from the Chilean supercolony (Chile; 1965) was sampled from the only recognised supercolony in the country (Vogel et al., 2010). For each of the three samples representing the introduced ranges, ten individuals were collected from each of ten different nests within each supercolony (i.e., 100 individuals per supercolony). In the introduced ranges, ten nests were always close together at a single location. For the native-range sample, ten individuals were collected from ten nests each from a separate supercolony from four separate localities (i.e., a total of 100 individuals per ten supercolonies) to capture the total genetic diversity in the species' natural range (López et al., 2025) (Table S1).

2.2. DNA extraction and sequencing

Genomic DNA was extracted individually from each ant using the DNeasy Blood & Tissue Kit (QIAGEN). Equal amounts of DNA from ten individuals per nest were pooled and genome-amplified using REPLI-g Mini Kit (QIAGEN). Finally, all the genome-amplified sub-samples were pooled so that each final sample represented 100 individuals. The samples were sent to BGI Genomics for library preparation and paired-end sequencing using an Illumina Hiseq™ 2000 sequencer, with a read length of 90 base pairs (bp) and an

insert size of 200 bp. The clean reads provided by BGI Genomics were filtered to remove adapters,

contaminants, and low-quality sequences.

2.3. Data processing

The quality encoding of the sequenced raw reads was converted from Phred 64 to Phred 33 (also known as

Sanger), as it is more widely used and required in many software programs. The conversion was done using

the Seqtk tool (version 1.3) (*Seqtk*, 2018). After this, raw reads were filtered using trim-fastq.pl script from

the basic pipeline of PoPoolation (version 1.2.2) (Kofler et al., 2011). The reads were filtered using an

average minimum base quality of 20 and a minimum read length of 70 bp. The quality of the reads was

verified using FastQC (version 0.11.8) analyses after filtering (Andrews, 2010).

Filtered reads were aligned to the reference genome (Genome assembly Lhum_UNIL_v1.0, Pan et al., 2024)

using the BWA-MEM algorithm from the BWA software package (version 0.7.17) with default settings (Li &

Durbin, 2009). The resulting SAM files were sorted, quality-filtered and converted to more compressed

binary versions (BAM files) using the view command of SAMtools (version 1.10) (Li et al., 2009). Only

aligned reads with a mapping quality of at least 20 and a SAM flag "properly paired" were saved. Duplicates

were removed using Picard's MarkDuplicates tool (version 2.21.4) (*Picard Toolkit*, 2019). RepeatModeler

(version 2.0.7) was used to identify repeats, and RepeatMasker (version 4.1.9) was used to mask the

identified repeats in the reference genome (Smit & Hubley, 2015). Degeneracy of sites in coding regions

was annotated with degenotate (version 1.2.4) (Mirchandani et al., 2024).

2.4. Population genetic analyses

2.4.1. Genomic diversity

The genomic diversity of supercolonies was evaluated for the whole genome, as well as separately for four-

fold and zero-fold degenerate sites in coding regions. Two genetic diversity estimators, Tajima's π (π) and

Watterson's Θ (Θ_w), were computed using Grenedalf's (version 0.6.3) diversity command (Czech et al.,

2024). Aligned reads were subsampled to a maximum sequencing depth of 200 using the subsampling

method subscale. A minimum sequencing depth of four and a minor allele count of two were required per

sample. We also estimated microsatellite marker diversity, expected heterozygosity (H_{exp}) and allelic

richness (k') for five variable microsatellite loci obtained from Vogel et al. (2010), applying a similar sample

size and design as the genomic samples (see Text S2).

2.4.2. Genomic differentiation

To evaluate the genomic differentiation of supercolonies, we calculated Hudson's estimator of F_{ST} using

Grenedalf´s fst command (Czech et al., 2024). A minimum sequencing depth of four and a minor allele

count of two were required per sample. We also estimated microsatellite marker differentiation using

pairwise F_{ST} for five variable microsatellite loci obtained from Vogel et al. (2010), applying a sample size and

design similar to those of the genomic samples (see Text S2).

2.4.3. Population structure and adaptive evolution

We used the population genomics software BayPass (version 2.31), which utilises a Bayesian framework to

identify SNPs subjected to selection and associated with population-specific covariates (Gautier, 2015;

Olazcuaga et al., 2020). The different models of BayPass estimate and account for the neutral covariance

structure among population allele frequencies resulting from their shared history.

To evaluate population structure and identify SNPs under selection among invasive supercolonies, the

analysis was first run in core model mode with default settings. The differentiation outlier statistics, XtX,

consider all analysed populations simultaneously and identify SNPs that show unusually high differentiation

between populations relative to neutral expectations based on population structure and drift, and thus are

potential targets of selection in one or more populations. We analysed the three invasive supercolonies to

identify differentiated SNPs in at least one of them, making these SNPs candidates for selection. As this

analysis is based on differences in allele frequencies and includes solely invasive supercolonies, it only

identifies SNPs that have experienced different selective pressures in at least one of the studied

supercolonies, rather than SNPs that have a shared selective regime across all three invasive supercolonies

(these SNPs were identified with a different method, see below). XtX statistics were calibrated using a

simulated neutral dataset, i.e., a pseudo-observed data set (POD). For neutral data simulation, SNPs were thinned into 20 subsets, each comprising approximately 42K SNPs. These thinned SNP sets were used to estimate the covariance matrix, i.e., Ω of population allele frequencies resulting from their demographic history, which were then used to simulate PODs of 50K SNPs. PODs were then run in core model mode, and the 99th percentiles of XtX differentiation outlier statistics were calculated; the average of these percentiles serves as a threshold to identify outlier SNPs, i.e., candidates for selection.

Furthermore, a contrast model mode, C2 statistic, was run to estimate whether SNPs are associated with invasiveness. The C2 statistics contrast and compare allele frequencies between groups of populations defined by the binary covariable of interest, testing for a consistent allele frequency difference between groups. Hence, we contrasted the native-range sample with the group of three invasive supercolones. This analysis identifies SNPs that share similar allele frequencies across invasive supercolonies and simultaneously differ from those in natives. However, note that the association does not require complete similarity in allele frequencies across all invasive supercolonies, two might be enough. The C2 statistics were calibrated using PODs as described above to indicate whether outlier SNPs were associated with invasiveness. Both models accounted for population structure estimated as Ω of population allele frequencies.

Genes containing outlier SNPs with high XtX values were considered as potential targets of positive selection in one or more invasive supercolonies. Genes containing outlier SNPs with high C2 values were associated with invasiveness. We performed gene enrichment analysis using the genes identified as potential targets of positive selection as a test set and the annotated genes of the reference genome (Annotation GCF_040581485.1-RS_2024_08) as a reference set, using the ShinyGO tool (version 0.85) (Ge et al., 2020). Similarly, genes associated with invasiveness were analysed for enrichment. Moreover, genes identified by both the XtX and C2 statistics were analysed for enrichment.

3. Results

3.1. Properties of the data

Sequencing reads covered 92.78%, 93.11%, 82.71%, and 92.38% of the reference genome in native-range

sample, Main, Catalonia and Chile, while the mean sequencing depths were 87, 98, 61, and 78, respectively.

Repetitive regions, which comprised 31% of the reference genome, were removed from all downstream

analyses.

3.2. Introduced supercolonies are expanding and harbour low genetic diversity

We evaluated the genetic diversity of supercolonies with two genetic diversity estimators, Tajima's pairwise

nucleotide diversity (π) and Watterson's theta (Θ_w), describing the population mutation rate. To ensure

comparability, we accepted only sites with data from all supercolonies for diversity calculations. These sites

contained 502,659 whole-genome SNPs, of which 59,575 were 4-fold SNPs, and 122,678 were 0-fold SNPs.

SNPs were identified simultaneously across the entire data set.

According to π, the native-range supercolonies were more genetically diverse than the invasive

supercolonies (p-values < 0.05 in all pairwise comparisons, see details in Table S3; Table 1). Both expected

heterozygosity (H_{exp}) and allelic richness (k') estimates derived from microsatellite marker diversity yielded

similar results to π – the sample of native-range supercolonies harbours higher diversity than the invasive

supercolonies (Table S4, Figure S5). Instead, Θ_w indicated that Catalonia had the highest genetic diversity

(p-values < 0.05 in all pairwise comparisons, see details in Table S3; Table 1). The Catalonia supercolony

harboured an excess of rare alleles, as evidenced also by the high peaks on the left side of the folded site

frequency spectrum (Figure 1). For each supercolony, Θ_w was always higher than π, especially pronounced

in the invasive supercolonies, indicating expanding population sizes (p-values < 0.05 in all pairwise

comparisons, see details in Table S3; Table 1). Moreover, four-fold sites had consistently higher diversity

than zero-fold sites, as expected (p-values < 0.05 in all pairwise comparisons, see details in Table S3; Table

1).

3.3. Introduced supercolonies are genetically different from each other

We used pairwise F_{ST} values to evaluate genetic differentiation between supercolonies. The results

indicated that supercolonies were genetically highly differentiated from each other (Table 2). The pairwise

F_{ST} values were consistently highest between Catalonia and the other supercolonies, indicating that

Catalonia was the most genetically differentiated supercolony (Table 2). The microsatellite marker data also

revealed genetic differences between supercolonies (Table S6, Figure S7). The marker data indicated that

invasive supercolonies were more differentiated from each other than from the native-range supercolonies

(Table S6, Figure S7) – a pattern that we did not observe in the genomic data.

3.3. Argentine ants can adapt to new environments

The core model and the XtX statistics of the BayPass software were used to identify candidate genes for

positive selection among invasive supercolonies. 1220 genes (see Table S8) contained one or more SNPs

with an XtX value greater than the calibrated threshold (6.3825), indicating them as outlier loci in the

genome scan for adaptive differentiation, and thus local adaptation in one or more of the invasive

supercolonies. The XtX outlier SNPs were distributed across the genome (Figure S9). Enriched molecular

function and biological process GO terms among the candidate genes were mainly related to nervous

system development, ion transport, signalling-related processes, and transcriptional regulation (Table S10).

Contrast analysis based on C2 statistics of the BayPass software were used to identify genes associated with

invasiveness. 285 genes (see Table S11) contained one or more SNPs with a C2 value greater than the

calibrated threshold (4.7973), indicating them as outlier loci in the genome scan for association with

invasiveness. Thus, these genes showed signs of parallel adaptive evolution across invasive supercolonies.

These C2 outlier SNPs were also distributed across the genome (Figure S12). Enriched molecular function

and biological process GO terms among genes associated with invasiveness were mainly related to ion

transport, cellular signalling and communication, and gene regulation and transcriptional processes (Table

S13). Notably, 97 genes were identified by both XtX and C2 analyses. These genes showed both high

differentiation between invasive supercolonies and an association with invasiveness, indicating parallel

adaptive evolution at each gene in two out of the three invasive supercolonies studied. One molecular

function GO term, GO:0030125 Semaphorin receptor binding, was enriched among these genes (Table S14).

The remaining 188 genes associated with invasiveness (high C2) were those that showed parallel adaptive

evolution across all three invasive supercolonies (as not identified with the XtX).

Both models estimated and accounted for the covariance structure among population allele frequencies

resulting from their shared history. The SVD-based visualisation of the Ω matrix summarises the genetic

structure among the native-range sample and the three invasive supercolonies (Figure 2). PC1 explains

approximately 50% of the total genetic variation and distinguishes native-range supercolonies from invasive

ones. PC2 explains approximately 35% of the variation and distinguishes the Catalonia supercolony from

the other invasive supercolonies (Figure 2).

4. Discussion

This study aimed to shed light on the highly invasive Argentine ant's recent evolutionary history and

genome dynamics, specifically focusing on the introduced supercolonies. We searched for answers to the

following questions: i) can signs of selection be detected in the genomes of the highly invasive Argentine

ant?, ii) are there signatures of parallel adaptive evolution among invasive supercolonies?, iii) are we able

to verify, using genomic data, the earlier population genetic findings on diversity and divergence based on

microsatellite marker data?

Despite the recent demographic history, including strong founder effects, we detected signs of selection in

the invasive supercolonies, indicating adaptability. Numerous genes were identified as candidates for

positive selection among invasive supercolonies. This finding demonstrates local adaptation in introduced

ranges with novel selective pressures. Due to the global nature of the analysis considering allele frequency

differences, it cannot be said whether these genes were positively selected in one or more of the

supercolonies, or, if selected in more than one, whether the selected allele is the same or different. But it is

evident that positive selection shapes the evolution of invasive Argentine ant supercolonies, and that the

identified genes play a key role in local adaptation. Moreover, some genes were associated with

invasiveness, indicating parallel adaptive evolution in invasive supercolonies. Thus, selective regimes are

probably not entirely different between invasive supercolonies, and some selective pressures are shared

across invasive ranges. Whilst the Mediterranean-like climate is shared across the native and invasive

ranges of the Argentine ant (Roura-Pascual et al., 2011), other biotic and abiotic factors, such as pathogens

and competitors, likely differ between regions, leading to distinct local selective pressures and local

adaptation (Holmberg et al., 2024; Lester & Gruber, 2016).

We detected positive selection in several immune genes (emp, imd, hop, bsk, Trax, Toll-6, and Sp1) in the

whole-genome scanning. We could not find overlap between the positively selected immune genes

identified here and those reported in the earlier Argentine ant immune gene study, even though the

studied supercolonies are the same (worker specimens differ) (Holmberg et al., 2024). This might be due to

differences in sampling strategies and the methods used. However, together, these findings indicate that

immune genes play important roles in how invasive Argentine ants adapt to new ranges.

Wagner et al. (2023) reported that Argentine ants were excellent olfactory learners. They proposed that

efficient learning combined with other features of Argentine ants, such as rapid discovery of food sources

and other resources, and rapid recruitment, might provide a competitive advantage over local species in

introduced ranges (Wagner et al., 2023). Intriguingly, the positively selected genes include olfactory

learning (trp, mura, mnb, Mob2, cher, PKA-R1, Fas3, Fas1) and memory genes (eag, aPKC, dnc).

The enriched GO terms among the positively selected genes included a range of molecular functions. Some

of these overlap with GO terms for genes that have evolved under positive selection across the ant

phylogeny over a long evolutionary time (Barkdull & Moreau, 2023). These GO terms were GO:0015103

Inorganic anion transmembrane transporter activity, GO:0022836 Gated channel activity, GO:0022857

Transmembrane transporter activity, GO:0004930 G protein-coupled receptor activity, GO:0004672 Protein

kinase activity, GO:0005515 Protein binding, and GO:0008061 Chitin binding.

The general observation of low diversity and high divergence in the invasive supercolonies based on our

genome-wide data largely confirms the results of earlier marker data studies of Argentine ants (Blight et al.,

2012; Brandt et al., 2009; Giraud et al., 2002; Jaquiéry et al., 2005; Tsutsui et al., 2000; Vogel et al., 2010).

However, there were some discrepancies between the results of genomic data and the microsatellite loci

obtained from Vogel et al. (2010). Catalonia appeared to be even more differentiated from other

supercolonies based on the genomic data than the marker data suggested, with an excess of rare alleles

(high Θ_w). Similarly, in another invasive ant species, *Cardiocondyla obscurior,* genome-wide evidence

showed reduced genetic diversity in invasive populations (Errbii et al., 2021). Moreover, marker data have

demonstrated reduced genetic diversity in several other invasive ant populations of *Nylanderia fulva,*

Pheidole megacephala, Solenopsis invicta, *Solenopsis geminata*, and *Wasmannia auropunctata* (Fournier et

al., 2005, 2009, 2012; Ross et al., 1993, 1996; Wauters et al., 2018). Together, these results indicate that

introduced populations of invasive ant species typically harbour less genetic diversity than native

populations. This is in line with the typical supercolonial lifestyle of the invasive ants, i.e., the formation of

closed breeding units and the fact that populations in the introduced ranges usually comprise single

supercolonies.

Our results give no indication that the demographic and ecological changes that accompany an invasion,

namely the expansion of supercolonies in the absence of competition that drastically reduces the

relatedness between sterile workers and reproductive queens (Giraud et al., 2002; Helanterä, 2022;

Helanterä et al., 2009; Pedersen et al., 2006), would have amplified the predicted accumulation of harmful

mutations or hampered positive selection on worker traits (Linksvayer & Wade, 2009) in the invasive

supercolonies. This could be due to, firstly, the relatively short time span – less than 100 generations –

since introductions, or secondly, the rarity of genes only expressed in workers, where such mutation

accumulation is predicted and shown to occur (Linksvayer & Wade, 2009; Warner et al., 2017). The rarity of

such genes could mask any genome-wide effect, and testing this hypothesis in invasive supercolonies

requires high-quality gene expression data.

The high divergence between the invasive Argentine ant supercolonies included in this study may be

explained by the fact that they have originated from different primary introductions from the native range

without secondary reinforcements (Giraud et al., 2002; Vogel et al., 2010). Moreover, founder effect likely

initiated the first shift toward genetic differentiation within the introduced supercolonies, as the effective

queen number establishing the European Main as well as the Catalonian supercolonies has been estimated

to be between 6 to 13 individuals (Giraud et al., 2002; Vogel et al., 2010), while in the native range

supercolonies, it has been estimated to be extremely high, reaching infinity (Pedersen et al., 2006).

Subsequent gene flow between invasive supercolonies has been prevented by two factors: i) Argentine ant

workers are highly aggressive toward males originating from other colonies, and ii) there is considerable

geographic distance between the studied supercolonies, except the European Main and Catalonian

supercolonies, which live partly next to each other (Blight et al., 2012; Giraud et al., 2002; Jaquiéry et al.,

2005; Vogel et al., 2010). Although the European Main and Catalonia supercolonies live close to each other,

there is no gene flow between them (Blight et al., 2012; Giraud et al., 2002; Jaquiéry et al., 2005). In

addition, amplified genetic drift in small founder populations might have further increased fluctuations in

allele frequencies, contributing to diversification within expanding supercolonies (Nei et al., 1975; Vogel et

al., 2010). Especially in Catalonia, the excess of rare alleles might indicate both substantial expansion after

population foundation and strong purifying selection. Other social insects, such as the yellow-legged hornet

Vespa velutina and German wasp *Vespula germanica,* have also successfully invaded regions outside their

native range despite low genetic diversity (Arca et al., 2015; Brenton-Rule et al., 2018; Eloff et al., 2020;

Takeuchi et al., 2017). In the yellow-legged hornet, successful invasion possibly originated from as few

individuals as just one (one queen mated with multiple males) (Arca et al., 2015; Takeuchi et al., 2017). In

addition, other non-social insects, such as the African fig fly *Zaprionus indianus* (Comeault et al., 2020), and

other species, such as the invasive wart comb jelly *Mnemiopsis leidyi* (Jaspers et al., 2021) have also proven

that invasion success does not require high genomic diversity. Thus, the general assumption of invasive

populations' lower adaptive potential is incorrect, as the founder effect and low diversity do not always

prevent invaders' success.

The success of a wide range of invasive species populations with low adaptive potential due to low genetic

diversity has been explained by factors such as gene flow, stress-induced accelerated mutation rate or TE-

activation, epigenetic changes and the release from natural pathogens and enemies (Errbii et al., 2021;

Manfredini et al., 2019; Marin et al., 2019; North et al., 2021; Schrader et al., 2014). Despite the low genetic

diversity, the invasive Argentine ants continue to evolve adaptively. Pre-adaptation in the native range has

recently received more attention across species (Davidson et al., 2011; Elst et al., 2016; Estoup et al., 2016;

Guo et al., 2014; Hufbauer et al., 2012), and we suggest that this might also play a role in the initial invasive

success of the Argentine ant. The unstable environment in the native range of the Argentine ant,

characterised by frequent flooding (LeBrun et al., 2007), may enhance the species' adaptive potential.

Balancing selection, which maintains alternative alleles in fluctuating conditions, could contribute to their

continued and long-term success by enabling rapid adaptation to new habitats following introductions as

indicated here. The exact roles of all selected genes must be carefully resolved among the complex

biological processes and pathways in upcoming genomic surveys, such as comparative population genomic

studies between native and introduced supercolonies using both high-quality genomic and transcriptomic

data at the individual level. Our study provides an excellent groundwork and frame for future studies of the

Argentine ant in the field of invasion genomics.

Tables and Figures

Table 1. The average per-site diversity estimators, π and Θ_W, for whole-genome, four-fold sites, and zero-fold
sites.

	Sample	π	Θ_W
whole-genome	Native	0.00094	0.00203
	Main	0.00048	0.00200
	Catalonia	0.00058	0.00593
	Chile	0.00056	0.00210
4-fold	Native	0.00220	0.00521
	Main	0.00120	0.00486
	Catalonia	0.00124	0.01219
	Chile	0.00127	0.00544
0-fold	Native	0.00049	0.00237
	Main	0.00031	0.00221
	Catalonia	0.00043	0.00550
	Chile	0.00029	0.00246

Figure 1. Folded site frequency spectra for Native, Main, Catalonia, and Chile.

363   Table 2. Genetic differences measured as pairwise F_{ST} values, genome-wide average.

F_{ST}	Native	Main	Catalonia	Chile
Native	0	0.2812	0.4434	0.2670
Main		0	0.5034	0.2507
Catalonia			0	0.5148
Chile				0

Figure 2. The SVD-based visualisation of the covariance matrix, i.e., Ω of population allele frequencies.

Author contribution

JSP, HH, and LV designed the research. JP did the lab work. IP analysed the data and wrote the manuscript,

and all authors contributed to editing and revising.

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References

Andrews, S. (2010). *FastQC A Quality Control tool for High Throughput Sequence Data*.

https://www.bioinformatics.babraham.ac.uk/projects/fastqc/

AntWeb (Version 8.114.). (2026, March 12). AntWeb; California Academy of Science, online at

https://www.antweb.org.

Arca, M., Mougel, F., Guillemaud, T., Dupas, S., Rome, Q., Perrard, A., Muller, F., Fossoud, A., Capdevielle-

Dulac, C., Torres-Leguizamon, M., Chen, X. X., Tan, J. L., Jung, C., Villemant, C., Arnold, G., Silvain, J.-F.,

Dupas, S., Fossoud, Á. A., Capdevielle Dulac, Á. C., … Villemant, Á. C. (2015). Reconstructing the

invasion and the demographic history of the yellow-legged hornet, Vespa velutina, in Europe.

Biological Invasions, *17*(8), 2357–2371. https://doi.org/10.1007/S10530-015-0880-9

Barkdull, M., & Moreau, C. S. (2023). Worker Reproduction and Caste Polymorphism Impact Genome

Evolution and Social Genes Across the Ants. *Genome Biology and Evolution*, *15*(6).

https://doi.org/10.1093/gbe/evad095

Blight, O., Berville, L., Vogel, V., Hefetz, A., Renucci, M., Orgeas, J., Provost, E., & Keller, L. (2012). Variation

in the level of aggression, chemical and genetic distance among three supercolonies of the Argentine

ant in Europe. *Molecular Ecology*, *21*(16), 4106–4121. https://doi.org/10.1111/J.1365-

294X.2012.05668.X

Brandt, M., Wilgenburg, E. Van, & Tsutsui, N. D. (2009). Global-scale analyses of chemical ecology and

population genetics in the invasive Argentine ant. *Molecular Ecology*, *18*(5), 997–1005.

https://doi.org/10.1111/J.1365-294X.2008.04056.X

Brenton-Rule, E. C., Dobelmann, J., Baty, J. W., Brown, R. L., Dvorak, L., Grangier, J., Masciocchi, M.,

McGrannachan, C., Shortall, C. R., Schmack, J., van Zyl, C., Veldtman, R., & Lester, P. J. (2018). The

origins of global invasions of the German wasp (Vespula germanica) and its infection with four honey

bee viruses. *Biological Invasions*, *20*(12), 3445–3460. https://doi.org/10.1007/S10530-018-1786-

0/TABLES/3

Comeault, A. A., Wang, J., Tittes, S., Isbell, K., Ingley, S., Hurlbert, A. H., Matute, D. R., & Parsch, J. (2020).

Genetic Diversity and Thermal Performance in Invasive and Native Populations of African Fig Flies.

Molecular Biology and Evolution, *37*(7), 1906. https://doi.org/10.1093/MOLBEV/MSAA050

Czech, L., Spence, J. P., Es Exp Osito-Alonso, M., Associate, E.-A.), & Schwartz, R. (2024). grenedalf:

population genetic statistics for the next generation of pool sequencing. *Bioinformatics*, *40*(8). https://doi.org/10.1093/BIOINFORMATICS/BTAE508

Davidson, A. M., Jennions, M., & Nicotra, A. B. (2011). Do invasive species show higher phenotypic plasticity than native species and, if so, is it adaptive? A meta-analysis. *Ecology Letters*, *14*(4), 419–431. https://doi.org/10.1111/J.1461-0248.2011.01596.X

Eloff, J., Veldtman, R., Bulgarella, M., & Lester, P. J. (2020). Population genetics of the invasive wasp Vespula germanica in South Africa. *Insectes Sociaux*, *67*(2), 229–238. https://doi.org/10.1007/S00040-020-00752-X

Elst, E. M., Acharya, K. P., Dar, P. A., Reshi, Z. A., Tufto, J., Nijs, I., & Graae, B. J. (2016). Pre-adaptation or genetic shift after introduction in the invasive species Impatiens glandulifera? *Acta Oecologica*, *70*, 60–66. https://doi.org/10.1016/J.ACTAO.2015.12.002

Errbii, M., Keilwagen, J., Hoff, K. J., Steffen, R., Altmüller, J., Oettler, J., & Schrader, L. (2021). Transposable elements and introgression introduce genetic variation in the invasive ant Cardiocondyla obscurior. *Molecular Ecology*, *30*(23), 6211–6228. https://doi.org/10.1111/MEC.16099

Estoup, A., Ravigné, V., Hufbauer, R., Vitalis, R., Gautier, M., & Facon, B. (2016). Is There a Genetic Paradox of Biological Invasion? *Annual Review of Ecology, Evolution, and Systematics*, *47*, 51–72. https://doi.org/10.1146/ANNUREV-ECOLSYS-121415-032116

Fournier, D., De Biseau, J. C., & Aron, S. (2009). Genetics, behaviour and chemical recognition of the invading ant Pheidole megacephala. *Molecular Ecology*, *18*(2), 186–199. https://doi.org/10.1111/J.1365-294X.2008.04018.X

Fournier, D., Foucaud, J., Loiseau, A., Cros-Arteil, S., Jourdan, H., Orivel, J., Le Breton, J., Chazeau, J., Dejean, A., Keller, L., & Estoup, A. (2005). Characterization and PCR multiplexing of polymorphic microsatellite loci for the invasive ant Wasmannia auropunctata. *Molecular Ecology Notes*, *5*(2), 239–242. https://doi.org/10.1111/J.1471-8286.2005.00889.X

Fournier, D., Tindo, M., Kenne, M., Mbenoun Masse, P. S., van Bossche, V., de Coninck, E., & Aron, S.

(2012). Genetic Structure, Nestmate Recognition and Behaviour of Two Cryptic Species of the Invasive

Big-Headed Ant Pheidole megacephala. *PLOS ONE*, *7*(2), e31480.

https://doi.org/10.1371/JOURNAL.PONE.0031480

Gautier, M. (2015). Genome-wide scan for adaptive divergence and association with population-specific

covariates. *Genetics*, *201*(4), 1555–1579. https://doi.org/10.1534/GENETICS.115.181453/-/DC1

Ge, S. X., Jung, D., Jung, D., & Yao, R. (2020). ShinyGO: a graphical gene-set enrichment tool for animals and

plants. *Bioinformatics*, *36*(8), 2628–2629. https://doi.org/10.1093/BIOINFORMATICS/BTZ931

Giraud, T., Pedersen, J. S., & Keller, L. (2002). Evolution of supercolonies: The Argentine ants of southern

Europe. *Proceedings of the National Academy of Sciences of the United States of America*, *99*(9),

6075–6079. https://doi.org/10.1073/pnas.092694199

Guo, W. Y., Lambertini, C., Nguyen, L. X., Li, X. Z., & Brix, H. (2014). Preadaptation and post-introduction

evolution facilitate the invasion of Phragmites australis in North America. *Ecology and Evolution*,

4(24), 4567. https://doi.org/10.1002/ECE3.1286

Helanterä, H. (2022). Supercolonies of ants (Hymenoptera: Formicidae): ecological patterns, behavioural

processes and their implications for social evolution. *Myrmecoigal News*, *32*, 1–22.

Helanterä, H., Strassmann, J. E., Carrillo, J., & Queller, D. C. (2009). Unicolonial ants: where do they come

from, what are they and where are they going? *Trends in Ecology & Evolution*, *24*(6), 341–349.

https://doi.org/10.1016/J.TREE.2009.01.013

Hölldobler, B., & Wilson, E. O. (1990). *The Ants*. Harvard University Press.

Holmberg, I., Tolonen, L., Paviala, J., Pedersen, J. S., Helanterä, H., & Viljakainen, L. (2024). Positive

selection has shaped the evolution of Argentine ant immune genes both in native and introduced

supercolonies. *Journal of Evolutionary Biology*, *37*(2), 131–140. https://doi.org/10.1093/JEB/VOAD014

Holway, D. A., Lach, L., Suarez, A. V., Tsutsui, N. D., & Case, T. J. (2002). The Causes and Consequences of

Ant Invasions. *Annual Review of Ecology and Systematics*, *33*, 181–233.

https://doi.org/10.1146/ANNUREV.ECOLSYS.33.010802.150444

Hufbauer, R. A., Facon, B., Ravigné, V., Turgeon, J., Foucaud, J., Lee, C. E., Rey, O., & Estoup, A. (2012).

Anthropogenically induced adaptation to invade (AIAI): contemporary adaptation to human-altered

habitats within the native range can promote invasions. *Evolutionary Applications*, *5*(1), 89–101.

https://doi.org/10.1111/J.1752-4571.2011.00211.X

Jaquiéry, J., Vogel, V., & Keller, L. (2005). Multilevel genetic analyses of two European supercolonies of the

Argentine ant, Linepithema humile. *Molecular Ecology*, *14*(2), 589–598.

https://doi.org/10.1111/J.1365-294X.2005.02433.X

Jaspers, C., Ehrlich, M., Pujolar, J. M., Künzel, S., Bayer, T., Limborg, M. T., Lombard, F., Browne, W. E.,

Stefanova, K., & Reusch, T. B. H. (2021). Invasion genomics uncover contrasting scenarios of genetic

diversity in a widespread marine invader. *Proceedings of the National Academy of Sciences*, *118*(51),

e2116211118. https://doi.org/10.1073/PNAS.2116211118

Keller, L., & Passera, L. (1993). Incest avoidance, fluctuating asymmetry, and the consequences of

inbreeding in Iridomyrmex humilis, an ant with multiple queen colonies. *Behavioral Ecology and*

Sociobiology, *33*, 191–199. https://doi.org/10.1007/BF00216600

Kofler, R., Orozco-terWengel, P., De Maio, N., Pandey, R. V., Nolte, V., Futschik, A., Kosiol, C., & Schlötterer,

C. (2011). PoPoolation: A Toolbox for Population Genetic Analysis of Next Generation Sequencing Data

from Pooled Individuals. *PLoS ONE*, *6*(1), e15925. https://doi.org/10.1371/journal.pone.0015925

LeBrun, E. G., Tillberg, C. V., Suarez, A. V., Folgarait, P. J., Smith, C. R., & Holway, D. A. (2007). AN

EXPERIMENTAL STUDY OF COMPETITION BETWEEN FIRE ANTS AND ARGENTINE ANTS IN THEIR

NATIVE RANGE. *Ecology*, *88*(1), 63–75. https://doi.org/https://doi.org/10.1890/0012-

9658(2007)88[63:AESOCB]2.0.CO

Lester, P. J., & Gruber, M. A. M. (2016). Booms, busts and population collapses in invasive ants. *Biological*

Invasions, *18*, 3091–3101. https://doi.org/https://doi.org/10.1007/s10530-016-1214-2

Li, H., & Durbin, R. (2009). Fast and accurate short read alignment with Burrows-Wheeler transform.

Bioinformatics, *25*(14), 1754–1760. https://doi.org/10.1093/bioinformatics/btp324

Li, H., Handsaker, B., Wysoker, A., Fennell, T., Ruan, J., Homer, N., Marth, G., Abecasis, G., & Durbin, R.

(2009). The Sequence Alignment/Map format and SAMtools. *Bioinformatics*, *25*(16), 2078–2079.

https://doi.org/10.1093/bioinformatics/btp352

Linksvayer, T. A., & Wade, M. J. (2009). GENES WITH SOCIAL EFFECTS ARE EXPECTED TO HARBOR MORE

SEQUENCE VARIATION WITHIN AND BETWEEN SPECIES. *Evolution*, *63*(7), 1685–1696.

https://doi.org/10.1111/J.1558-5646.2009.00670.X

López, M. E., Ozerov, M., Pukk, L., Noreikiene, K., Gross, R., & Vasemägi, A. (2025). Dynamic Outlier Slicing

Allows Broader Exploration of Adaptive Divergence: A Comparison of Individual Genome and Pool-Seq

Data Linked to Humic Adaptation in Perch. *Molecular Ecology*, *34*(4), e17659.

https://doi.org/10.1111/MEC.17659;PAGEGROUP:STRING:PUBLICATION

Mack, R. N., Simberloff, D., Lonsdale, W. M., Evans, H., Clout, M., & Bazzaz, F. A. (2000). Biotic Invasions:

Causes, Epidemology, Global Consequences, and Control. *Ecological Applications*, *10*(3), 689–710.

Manfredini, F., Arbetman, M., & Toth, A. L. (2019). A Potential Role for Phenotypic Plasticity in Invasions

and Declines of Social Insects. *Frontiers in Ecology and Evolution*, *7*, 375.

https://doi.org/10.3389/FEVO.2019.00375/BIBTEX

Marin, P., Genitoni, J., Barloy, D., Maury, S., Gibert, P., Ghalambor, C. K., & Vieira, C. (2019). Biological

invasion: The influence of the hidden side of the (epi)genome. *Functional Ecology*, *34*(2), 1–16.

https://doi.org/10.1111/1365-2435.13317/SUPPINFO

McGlynn, T. P. (1999). The worldwide transfer of ants: geographical distribution and ecological invasions.

Journal of Biogeography, *26*(3), 535–548. https://doi.org/10.1046/J.1365-2699.1999.00310.X

Mirchandani, C. D., Shultz, A. J., Thomas, G. W. C., Smith, S. J., Baylis, M., Arnold, B., Corbett-Detig, R.,

Enbody, E., & Sackton, T. B. (2024). A Fast, Reproducible, High-throughput Variant Calling Workflow

for Population Genomics. *Molecular Biology and Evolution*, *41*(1).

https://doi.org/10.1093/MOLBEV/MSAD270

Mollot, G., Pantel, J. H., & Romanuk, T. N. (2017). The Effects of Invasive Species on the Decline in Species

Richness: A Global Meta-Analysis. *Advances in Ecological Research*, *56*, 61–83.

https://doi.org/10.1016/BS.AECR.2016.10.002

Moreau, C. S., Bell, C. D., Vila, R., Archibald, S. B., & Pierce, N. E. (2006). Phylogeny of the ants:

Diversification in the Age of Angiosperms. *Science*, *312*(5770), 101–104.

https://doi.org/10.1126/SCIENCE.1124891/SUPPL_FILE/MOREAU_SOM.PDF

Nei, M., Maruyama, T., & Chakraborty, R. (1975). The Bottleneck Effect and Genetic Variability in

Populations. *Evolution*, *29*(1), 10. https://doi.org/10.2307/2407137

North, H. L., McGaughran, A., & Jiggins, C. D. (2021). Insights into invasive species from whole-genome

resequencing. *Molecular Ecology*, *30*(23), 6289–6308. https://doi.org/10.1111/MEC.15999

Olazcuaga, L., Loiseau, A., Parrinello, H., Paris, M., Fraimout, A., Guedot, C., Diepenbrock, L. M., Kenis, M.,

Zhang, J., Chen, X., Borowiec, N., Facon, B., Vogt, H., Price, D. K., Vogel, H., Prud'Homme, B., Estoup,

A., & Gautier, M. (2020). A Whole-Genome Scan for Association with Invasion Success in the Fruit Fly

Drosophila suzukii Using Contrasts of Allele Frequencies Corrected for Population Structure. *Molecular*

Biology and Evolution, *37*(8), 2369–2385. https://doi.org/10.1093/MOLBEV/MSAA098

Pan, Q., Darras, H., & Keller, L. (2024). LncRNA gene ANTSR coordinates complementary sex determination

in the Argentine ant. *Science Advances*, *10*(22), 31.

https://doi.org/10.1126/SCIADV.ADP1532/SUPPL_FILE/SCIADV.ADP1532_SUPPLEMENTARY_FILES_S1

_TO_S9.ZIP

Pedersen, J. S., Krieger, M. J. B., Vogel, V., Giraud, T., & Keller, L. (2006). NATIVE SUPERCOLONIES OF

UNRELATED INDIVIDUALS IN THE INVASIVE ARGENTINE ANT. *Evolution*, *60*(4), 782–791.

https://doi.org/10.1111/j.0014-3820.2006.tb01156.x

Pérez, J. E., Nirchio, M., Alfonsi, C., & Muñoz, C. (2006). The Biology of Invasions: The Genetic Adaptation

Paradox. *Biological Invasions*, *8*, 1115–1121. https://doi.org/10.1007/S10530-005-8281-0

Picard Toolkit. (2019). Broad Institute, GitHub Repository. Http://Broadinstitute.Github.Io/Picard/; Broad

Institute. https://broadinstitute.github.io/picard/

Ross, K. G., Vargo, E. L., & Keller, L. (1996). Social evolution in a new environment: the case of introduced

fire ants. *Proceedings of the National Academy of Sciences of the United States of America*, *93*(7),

3021–3025. https://doi.org/10.1073/PNAS.93.7.3021

Ross, K. G., Vargo, E. L., Keller, L., & Trager, J. C. (1993). Effect of a founder event on variation in the genetic

sex-determining system of the fire ant Solenopsis invicta. *Genetics*, *135*(3), 843–854.

https://doi.org/10.1093/GENETICS/135.3.843

Roura-Pascual, N., Hui, C., Ikeda, T., Leday, G., Richardson, D. M., Carpintero, S., Espadaler, X., Gómez, C.,

Guénard, B., Hartley, S., Krushelnycky, P., Lester, P. J., McGeoch, M. A., Menke, S. B., Pedersen, J. S.,

Pitt, J. P. W., Reyes, J., Sanders, N. J., Suarez, A. V., … Worner, S. P. (2011). Relative roles of climatic

suitability and anthropogenic influence in determining the pattern of spread in a global invader.

Proceedings of the National Academy of Sciences of the United States of America, *108*(1), 220–225.

https://doi.org/10.1073/PNAS.1011723108/SUPPL_FILE/PNAS.201011723SI.PDF

Sanders, N. J., Barton, K. E., & Gordon, D. M. (2001). Long-term dynamics of the distribution of the invasive

Argentine ant, Linepithema humile, and native ant taxa in northern California. *Oecologia*, *127*(1), 123–

130. https://doi.org/10.1007/S004420000572

Sanders, N. J., Gotelli, N. J., Heller, N. E., & Gordon, D. M. (2003). Community disassembly by an invasive

species. *Proceedings of the National Academy of Sciences of the United States of America*, *100*(5),

2477. https://doi.org/10.1073/PNAS.0437913100

Schrader, L., Kim, J. W., Ence, D., Zimin, A., Klein, A., Wyschetzki, K., Weichselgartner, T., Kemena, C., Stökl,

J., Schultner, E., Wurm, Y., Smith, C. D., Yandell, M., Heinze, J., Gadau, J., & Oettler, J. (2014).

Transposable element islands facilitate adaptation to novel environments in an invasive species.

Nature Communications, *5*(1), 1–10. https://doi.org/10.1038/ncomms6495

Seebens, H., Blackburn, T. M., Dyer, E. E., Genovesi, P., Hulme, P. E., Jeschke, J. M., Pagad, S., Pyšek, P.,

Winter, M., Arianoutsou, M., Bacher, S., Blasius, B., Brundu, G., Capinha, C., Celesti-Grapow, L.,

Dawson, W., Dullinger, S., Fuentes, N., Jäger, H., … Essl, F. (2017). No saturation in the accumulation of

alien species worldwide. *Nature Communications*, *8*(1), 1–9. https://doi.org/10.1038/ncomms14435

Seqtk. (2018). Toolkit for Processing Sequences in FASTA/Q Formats. https://github.com/lh3/seqtk

Sherpa, S., & Després, L. (2021). The evolutionary dynamics of biological invasions: A multi-approach

perspective. *Evolutionary Applications*, *14*, 1463–1484. https://doi.org/10.1111/EVA.13215

Smit, A., & Hubley, R. (2015). *RepeatModeler* (Open-1.0).

Suarez, A. V., Holway, D. A., & Case, T. J. (2001). Patterns of spread in biological invasions dominated by

long-distance jump dispersal: Insights from Argentine ants. *Proceedings of the National Academy of*

Sciences, *98*(3), 1095–1100. https://doi.org/10.1073/PNAS.98.3.1095

Takeuchi, T., Takahashi, R., Kiyoshi, T., Nakamura, M., Minoshima, Y. N., & Takahashi, J. (2017). The origin

and genetic diversity of the yellow-legged hornet, Vespa velutina introduced in Japan. *Insectes*

Sociaux, *64*(3), 313–320. https://doi.org/10.1007/S00040-017-0545-Z

Tsutsui, N. D., Suarez, A. V., Holway, D. A., & Case, T. J. (2000). Reduced genetic variation and the success of

an invasive species. *Proceedings of the National Academy of Sciences of the United States of America*,

97(11), 5948–5953. https://doi.org/10.1073/pnas.100110397

Vogel, V., Pedersen, J. S., D'Ettorre, P., Lehmann, L., & Keller, L. (2009). Dynamics and genetic structure of

argentine ant supercolonies in their native range. *Evolution*, *63*(6), 1627–1639.

https://doi.org/10.1111/j.1558-5646.2009.00628.x

Vogel, V., Pedersen, J. S., Giraud, T., Krieger, M. J. B., & Keller, L. (2010). The worldwide expansion of the

Argentine ant. *Diversity and Distributions*, *16*(1), 170–186.

https://doi.org/https://doi.org/10.1111/j.1472-4642.2009.00630.x

Wagner, T., Galante, H., Josens, R., & Czaczkes, T. J. (2023). Systematic examination of learning in the

invasive ant Linepithema humile reveals fast learning and long-lasting memory. *Animal Behaviour*,

203, 41–52. https://doi.org/10.1016/j.anbehav.2023.06.012

Warner, M. R., Mikheyev, A. S., & Linksvayer, T. A. (2017). Genomic Signature of Kin Selection in an Ant with

Obligately Sterile Workers. *Molecular Biology and Evolution*, *34*(7), 1780–1787.

https://doi.org/10.1093/MOLBEV/MSX123

Wauters, N., Dekoninck, W., & Fournier, D. (2018). Introduction history and genetic diversity of the invasive

ant Solenopsis geminata in the Galápagos Islands. *Biological Invasions*, *20*(11), 3207–3226.

https://doi.org/10.1007/S10530-018-1769-1

Wetterer, J. K., Espadaler, X., Wetterer, A. L., Aguin-Pombo, D., & Franquinho-aguiar, A. M. (2006). Long-

term impact of exotic ants on the native ants of Madeira. *Ecological Entomology*, *31*(4), 358–368.

https://doi.org/10.1111/J.1365-2311.2006.00790.X

Wetterer, J. K., & Wetterer, A. L. (2006). A Disjunct Argentine Ant Metacolony in Macaronesia and

Southwestern Europe. *Biological Invasions*, *8*, 1123–1129. https://doi.org/10.1007/S10530-005-8641-

9

Wetterer, J. K., Wild, A. L., Suarez, A. V., Roura-Pascual Núria, & Espadaler, X. (2009). Worldwide spread of

the Argentine ant, Linepithema humile (Hymenoptera: Formicidae). *Myrmecological News*, *12*, 187–

194.

Wild, A. L. (2004). Taxonomy and Distribution of the Argentine Ant, Linepithema humile (Hymenoptera:

Formicidae). *Annals of the Entomological Society of America*, *97*(6), 1204–1215.

Wilson, E. O., & Hölldobler, B. (2005). The rise of the ants: A phylogenetic and ecological explanation.

Proceedings of the National Academy of Sciences, *102*(21), 7411–7414.

https://doi.org/10.1073/PNAS.0502264102

Data Accessibility Statement

Raw sequence reads are deposited in the SRA (BioProject PRJNA846165).

Benefit-Sharing Statement

Benefits from this research accrue from sharing our data and results on public databases as described

above.