Research Article

Tumoral hypoxic extracellular vesicles create a protective microenvironment for triple-negative breast cancer

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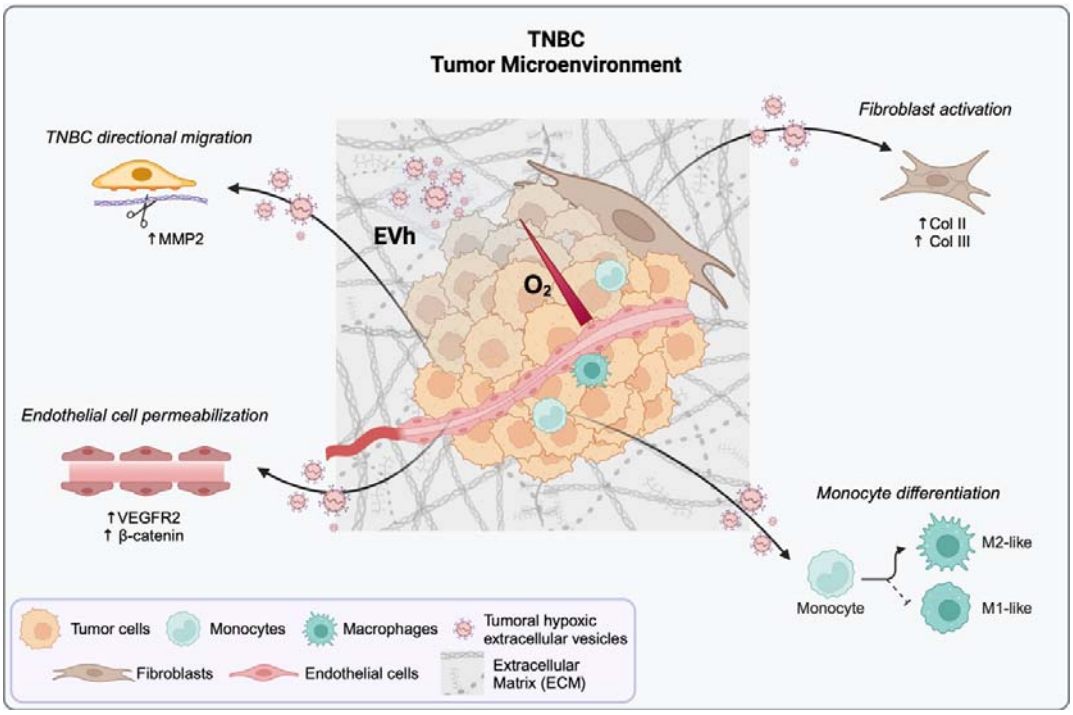
Graphical Abstract



The function of hypoxic extracellular vesicles in triple-negative breast cancer tumor microenvironment

Abstract

The highly metastatic triple-negative breast cancer (TNBC) relies on its tumor microenvironment (TME) to maintain phenotypic heterogeneity and progression. Extracellular vesicles from hypoxic TNBC were previously shown to aid tumoral invasion but their function in the tumor microenvironment is still unclear. We used a novel method to investigate the TME *in vitro* called multicellular circulating co-culture, to characterize how TNBC-derived hypoxic EVs (EVh) interfere with tumoral and endothelial cells, fibroblasts, monocytes and macrophages. EVh promoted monocyte differentiation to M2-like macrophages and inhibited macrophage-derived phagocytosis in endothelial and tumoral cells. The protection of endothelial, tumoral and stromal cellular integrity by EVh increased

pro-tumoral and pro-angiogenic signaling, collagen matrix synthesis and a potential differentiation to cancer-associated fibroblasts. Our findings highlighted the critical role of EVh in protecting tumor cells, indicating its cooperation towards a protective TME, which was demonstrated by the multicellular circulating co-culture and conventional co-culture protocols, leading to an adequate system with potential for investigating other tumor-related processes, including circulating tumor cells and metastasis.

Keywords: phagocytosis; extracellular vesicles; hypoxia; tumor microenvironment.

Statement of Significance

We are evaluating the cellular communication and interaction in the tumor microenvironment through a novel co-culture model while offering insights into the function of hypoxic extracellular vesicles in triple-negative breast cancer.

Introduction

Triple-negative breast cancer (TNBC) is mainly characterized by the lack of expression of receptors for estrogen, progesterone, and the human epidermal growth factor (HER2) (1). Comprised of a heterogeneous mass with a high risk of metastasis, TNBC affects over 15% of all breast tumors and presents poor overall survival (2). Its development is plagued with hypoxia, a condition in which oxygen distribution is reduced due to the increased cellular activity within the tumoral tissue, the inefficient tumoral vasculature, and the constant competition for nutrients within the mass (3,4). Hypoxia and tumorigenesis favor the secretion of extracellular vesicles (EV), nanoparticles (30-300 nm) released by all known cells with a major function in intercellular communication (5,6). Tumor-derived hypoxic EV (EVh) cargo has been previously described to favor pro-invasive pathways (7), but their direct action in the TNBC tumor microenvironment (TME) is still unknown.

The TME is a myriad of cellular and acellular components that evolve as the tumor phenotypic heterogeneity grows according to the stages of cancer progression (8–10). In the pre-invasive TME, autocrine signaling by epithelial (EGF), hepatocyte (HGF), and transforming growth factors (TGF-β) favors cell proliferation, migration, and plasticity (11). The extracellular matrix (ECM) is modified mainly by fibroblasts called, in this context, cancer-associated fibroblasts (CAF) via protein synthesis and remodeling enzyme expression, which alters the protein content, activity and crosslinking to favor collagen synthesis and deposition (12,13). Pro-inflammatory mediators induce monocyte differentiation into macrophages and their subsequent polarization into tumor-associated macrophages (TAM) (13). The increased cell activity of the tumor stimulates the release of reactive oxygen species, which favors DNA damage and the secretion of pro-angiogenic stimuli including VEGF and fibroblast growth factor (FGF-2) (11). Once the invasive profile is achieved, cell motility is favored by the polarization of the cytoskeleton, the interplay between adhesion molecules, and ECM remodeling by matrix metalloproteases (MMPs) (14). These changes in cell behavior result from the secretion of TGF-β, interleukins (IL-6 and IL-8), and VEGF by CAFs, which favors the epithelial-mesenchymal transition, survival, and maintenance of immune control (11).

The collective complexity of the TME renders its study challenging, particularly when using *in vitro* techniques such as 2D cell culture. Due to the intrinsic method limitations, including the restriction in the number of simultaneous cell cultures and the low correspondence with the physiology of clinical TME, there has been a shift towards 3D cell culture models, which add a novel layer of complexity to *in vitro* studies. However, one pitfall of these models is the difficulty in observing individual cellular responses from the mass (15). In this work, we describe a novel method for studying the TME *in vitro* called the multicellular circulating co-culture (MC-CC) to determine how TNBC-derived hypoxic EVs influence individual

immortalized cellular models in a TME setting. Using this approach, we demonstrated for the first time that EVh protects cells from macrophage-derived phagocytosis. This pro-tumorigenic function of EVh confirms previous suppositions on the tumoral function of EVs and supports the development of novel EV-based strategies to overcome resistance to anticancer therapies.

Materials and Methods

Cell culture

Human cells from triple-negative mammary adenocarcinoma MDA-MB-231 (ATCC® CRM-HTB-26™) were grown between passages 36 and 45 in Leibovitz L-15 10% FBS media at 37 ºC. Dermal fibroblasts HDFa (ATCC® PCS-201-012™) and umbilical cord endothelial cells HUVEC (ATCC® CRL-1730™) were maintained between passages 6 and 15 in DMEM 10% FBS 1% pen/strep at 37 ºC, 5% CO_2. Monocytes THP-1 (ATCC® TIB-202™) were kept under suspension in RPMI 1640 10% FBS 1% pen/strep at 37 ºC, 5% CO_2. Cells were grown under sterile conditions for no longer than 4 weeks after thawing and were frequently tested for *Mycoplasma* using the MycoAlert® Assay kit (Lonza).

EVh separation and characterization

EVs from MDA-MB-231 cells grown in hypoxia (1% O_2) in EV-depleted media were separated and characterized as described previously (7), following the MISEV2023 guidelines (16). EVh samples were characterized by nanoparticle tracking analysis (NTA; Nanosight NS300, Malvern), transmission electron microscopy (TEM; FEI TECNAI G2 F20 HRTEM), protein quantification and western blotting detection of ALIX, calnexin, CD63 and flotillin-1. Detailed methods are described in Suppl. 1A-D.

Monocyte differentiation in macrophages

Using OptiMEM 1% pen/strep (Gibco), THP-1 cells were seeded ($6x10^5$ cells/ml) in 6-well plates and treated with vehicle control (PBS), EVh (10^9 particles/ml) or phorbol-12-myristate-13-acetate (PMA, 100 nM) (17). Cell differentiation was evaluated with time-lapse light microscopy (Lux2, CytoSMART) with hourly snaps taken for 72h at 37 ºC, 5% CO_2 (18). Cell tracking was assessed using the TrackMate plugin from FIJI (18,19). Cell supernatant was collected and evaluated for IL-6, IL-10, TNF-α, and IL-1β with ELISA.

Time-lapse cell motility assay

In a 24-well dish, MDA-MB-231 ($5x10^4$ cells/ml), HUVEC ($5x10^4$ cells/ml) or HDFa (10^4 cells/ml) were seeded in complete growth media for adhesion over 24h at 37 ºC, 5% CO_2, followed by media replacement with OptiMEM 1% pen/strep. THP-1 cells were diluted in OptiMEM to achieve a 10^5 cells/ml concentration. An untreated group was maintained parallel to an EVh-treated group (10^9 particles/ml) for 24h at 37 ºC, 5% CO_2, under time-lapse light microscopy (Lux2, CytoSMART). Snaps were taken every 30 minutes and single-cell tracking was processed using the *TrackMate* plugin in FIJI (18,19).

Fluorescent gelatin invasion assay

Black 96-well plates coated with fluorescein-conjugated gelatin (0.2 mg/ml, G13187, Molecular Probes) were used to seed MDA-MB-231, HUVEC, HDFa or THP-1 cells in growth media. Plates were incubated overnight at 37 ºC, 5% CO_2. Media was substituted to OptiMEM 1% pen/strep containing EVh (10^9 particles/ml) or PBS for 24h at 37 ºC, 5% CO_2. Cell viability, morphology, and gelatin degradation were assessed following the full protocol (20).

Cell viability assay

Cells seeded for invasion assays were mixed with a 10% resazurin solution (0.1 ml/ml, #199303, Sigma-Aldrich) in OptiMEM and incubated for 4h at 37 ºC, 5% CO_2, alongside a

blank no-cell control and a 100%-reduced control. The supernatant was transferred to a new plate and analyzed by fluorescence (λ_{Exc} 555 nm, λ_{Em} 585 nm; SpectraMax i3, Molecular Devices).

Gelatin invasion in direct co-culture

MDA-MB-231, HUVEC, HDFa, or THP-cells were stained either with CellTracker™ Red CMTPX (5 µM, C34552, Invitrogen) or CellTrace™ CSFE (5 µM, C34554, Invitrogen) in OptiMEM. Two cell lineages were simultaneously seeded in a gelatin-coated 96-well black plate, treated with EVh (10^9 particles/ml) or PBS, and incubated at 37 ºC, 5% CO_2 for 24h. Cell viability, morphology, and gelatin degradation were determined following the full protocol (21).

Cell invasion in transwell co-culture

MDA-MB-231 cells previously attached to gelatin-coated coverslips were assembled in Boyden chambers with inserts containing a monolayer of HUVEC or HDFa cells. The system was maintained in OptiMEM for 24h at 37 ºC, 5% CO_2. Cells and conditioned media were processed following the full protocol (22).

Multicellular circulating co-culture (MC-CC)

MDA-MB-231, HUVEC, and HDFa cells were seeded to gelatin, Matrigel, and FN-coated coverslips, respectively, overnight at 37 ºC, 5% CO_2 and assembled in sterile QV500 (*Kirkstall*) culture chambers with 1 ml of OptiMEM 1% pen/strep. THP-1 cells were pre-stained with CellTracker™ Red CMTPX (5 µM, C34552, Invitrogen) and transferred to the reservoir containing EVh (10^9 particles/ml) or vehicle (PBS). The tubing system was closed and connected to the MDA-MB-231, HUVEC, and HDFa chambers, propelled by a peristaltic pump set to a 50 µl/s flow rate. The circuit was incubated at 37 ºC, 5% CO_2 for 24h. Coverslips and the conditioned media were processed following the full protocol (23).

Individual static and circulating culture controls

Cells seeded in ECM-coated coverslips (23) were assembled individually in a sterile QV500 (*Kirkstall*) culture chamber with 1 ml of OptiMEM 1% pen/strep and connected with the tubing system to a reservoir containing 13 ml of media. Following the previous method, the system was maintained with a peristaltic pump, parallel to a static individual control. Coverslips were removed from the compartments, counterstained, and imaged at 40x magnification (ImageXpress Micro XRS, Molecular Devices). Cell circularity index and gelatin degradation were quantified using FIJI (7,18).

Enzyme-Linked Immunosorbent Assay - ELISA

High-affinity microplates were sensitized with monoclonal anti-cytokine antibodies (IL-6: 555220; IL-10: 555157; TNF-α: 555212; IL-1β: 557953, OptEIA, BD Biosciences) for 16-18h. Plates were washed thrice (0.05% Tween 20-PBS), incubated in 4% FBS-PBS for 1 hour at RT, and washed again. Both samples and the standard curve of recombinant cytokines were added to the wells for 2h, RT, and excess was washed off. Anti-cytokine antibodies conjugated with peroxidase were incubated for 1.5h, RT, followed by five washes with 0.05% Tween 20. A developing solution (3,3',5,5'-tetramethylbenzidine, TMB) was added to wells and the reaction was stopped with sulfuric acid. Plates were evaluated by spectrophotometry ($O.D._{450nm}$; SpectraMax i3, Molecular Devices). Sample concentrations were calculated from the titration curve of the cytokine standards, and final concentrations were expressed in pg/ml.

Statistical analysis

Cellular assays were repeated on three distinct occasions in technical triplicates. MC-CC was repeated four times with both groups running simultaneously. Each sample was applied twice on zymograms, and average band densitometry was used for statistics. Datasets were checked

for outliers using ROUT's test and distribution by Shapiro-Wilk ($n < 9$) or D'Agostino-Pearson omnibus K2 ($n \geq 9$). Parametric data was evaluated using unpaired t-tests (2 groups) or ANOVA one-way with Tukey's multiple comparison test (3+ groups). Non-parametric data was analyzed using the Mann-Whitney test (2 groups) or Kruskal-Wallis analysis of variance with Dunn's multiple comparison test (3+ groups). Values of $p < 0.05$ were considered statistically relevant (GraphPad Prism (v. 9.3)).

Results

EVh characterization

The isolation of EVh samples followed a previous method, where samples were thoroughly characterized (7). Incorporating the recommendations outlined in the MISEV2023 guidelines (16), EVh samples were separated from 2.2×10^7 MDA-MB-231 cells grown in hypoxia in serum-free media, leading to a population with modal size of 130.2 nm (± 78.7) and concentration of 3.69×10^{11} particles/ml (± 8.74×10^{10}) (Fig. 1A). Samples contained 236.6 µg/ml of protein and yielded 167,346.94 particles per cell or 1.07×10^{-5} µg protein per cell. Known EV biomarkers CD63 and flotillin-1 were abundant in EV samples, whereas Alix was faintly identified, and calnexin was absent (Fig. 1B). Visualization by TEM confirmed EVh abundance and typical morphology (Fig. 1C).

EVh mediates THP-1 differentiation into M2-like macrophages

The adherent phenotype of THP-1 cells was detected in both EVh- and PMA-treated groups after 16-24 hours of treatment, while untreated cells remained in suspension for up to 72 h (Fig. 2A and Suppl. 2A,B). The distinct cell morphologies found in EVh- and PMA- treated groups suggest alternative differentiation mechanisms (Fig. 2B). Gelatin invasion decreased upon EVh treatment (Fig. 2C,D). However, cell viability and circularity remained unchanged (Fig. 2E,F). The tracking of cell movement showed the undirected motility in untreated and

PMA-differentiated cells, whereas a directional migration of EVh-differentiated cells was observed with higher mean speed and average distance traveled (Fig. 2G-I). Inflammatory cytokine profiling of THP-1 conditioned media detected TNF-α and IL-1β only in the PMA group (Fig. 2J,K), a non-significant trend of IL-6 reduction in the EVh group (Fig. 2L), and continuity of IL-10 expression in all three groups (Fig. 2M).

Tumoral reprogramming of endothelial cells in vitro

EVh treatment in HUVEC promoted gelatin invasion and elongated cell morphology, consistent with the invasive phenotype (Fig. 3A-C). Cell viability was unaffected (Fig. Suppl. 3A). Unlike THP-1, there was non-directional motility in EVh-exposed HUVEC (Fig. 3D). Although the average speed of movement was similar between the groups, a higher distance traveled by cells was seen in EVh groups (Fig. 3E,F).

The pro-tumoral effect of EVh in endothelial cells was further investigated using indirect and direct co-culture assays with MDA-MB-231 cells. In indirect co-culture, where tumor cells were separated from endothelial cells by an insert (Fig. 3G), HUVEC exhibited typical morphology and significant intercellular interaction, while the tumoral cells showed multiple cytoskeleton extensions, which gave them a downy appearance (Fig. 3H). In EVh-treated conditions, the cytoskeleton retracted as endothelial cells visibly lost intercellular interactions, and tumor cells showed a more elongated phenotype, without F-actin extensions (Fig. 3I). Tumoral gelatin invasion was detected in both co-culture groups, likely due to MMP-2 activation (Fig. 3J-K; Suppl. Fig. 3B). Both MMP-2 activity and gelatin degraded area were slightly higher in EVh-treated groups (Fig. 3L). Adding to previous results (7), we found EVh directs movement in MDA-MB-231, which facilitates tumoral invasion in gelatin matrices (Fig. Suppl. 3C-E).

In the direct co-culture approach, where both cells were seeded in the same well simultaneously (Fig. 3M), the proximity between cells led to a stress response in which the endothelial cytoskeleton turned to a star-shape, while tumoral cells became more cohesive (Fig. 3N). Once exposed to EVh, cells recapitulated their original morphological characteristics (Fig. 3N; Suppl. Fig. 3F,G). These changes were reflected in the circularity index, suggesting the importance of EVh in the regulation of intercellular interactions on the TME (Fig. 3O). Like the indirect co-culture model, EVh-treated cells also increased gelatin invasion (Fig. 3P-Q). In this scenario, cell viability remained unchanged (Fig. 3R).

Fibroblast subtle response to tumor cells and EVh

HDFa promoted visible gelatinase activity, especially after EVh administration (Fig. 4A). Fibroblasts showed a migratory phenotype and increased gelatin degradation, coupled with a significant rise in cell viability after EVh exposure (Fig. 4B-D). HDFa movement was polarized and resulted from reduced track speed and increased total distance traveled (Fig. 4E-G).

To evaluate the responses of HDFa in different models of tumoral association, then an indirect co-culture assay was assembled between fibroblasts and gelatin-seeded MDA-MB-231. There was a subtle fibroblast response to tumoral signaling, regardless of EVh exposure (Fig. 4H-I). Cell area was reduced, making them broader than the individual cell controls, and tumoral gelatin degradation remained unchanged (Fig. 4J-K). The cell circularity index supported these observations. Zymograms suggest MMP-2 activity is reduced by EVh exposure (Fig. 4L). EVh also favors anti-invasive behaviors in a direct co-culture model. Both cells show a visibly distinct phenotype (Fig. 4M, Suppl. Fig. 3H), the overall cell morphology is more circular (Fig. 4N), gelatin degradation is reduced after EVh administration (Fig..4O) and cell viability remains unchanged (Suppl. Fig. 3I).

EVh inhibits phagoptosis via macrophages in the TME

A multicellular circulating co-culture (MC-CC) was developed to recreate the cellular complexity of a TNBC tumor *in vitro* for evaluating EVh response in a TME-like setting (Fig. 5A). In the system, each TME cell type was seeded in a compartment for individual assessment of targets, while maintaining communication by a constant flow of 50 µl/s, set by previous optimization steps containing isolated cells (Fig. Suppl. 4A). A closed system comprised of MDA-MB-231, HUVEC, HDFa and THP-1 cells circulated for 24 hours showed high levels of phagoptosis (i.e., death by macrophage phagocytosis) (24), especially in the TNBC and endothelial cell chambers (Fig. 5B). In these chambers, integer nuclei and cytoplasm were seldomly found. EVh exposure led to more frequently sighted cells with integer nuclei and cytoplasm throughout chambers, suggesting phagoptosis inhibition. Cell morphological plasticity was confirmed by the circularity indexes of MDA-MB-231 and HUVEC (Fig. Suppl. 4B-C). The media flow reduced gelatin invasion in TNBC cells compared to individual static controls. EVh treatment partially recovered the cell invasive ability (Fig. 5C). The conditioned media was tested for gelatinases, and MMP-2 activity remained unchanged (Fig. 5D, Suppl. 4D). Cytokine IL-1β was present in similar levels in both groups, but IL-6 was more abundant in the EVh-treated system (Fig. 5E,F).

Pro-tumoral components were quantified based on immunofluorescence images. In endothelial cells, the expression of β-catenin and VEGFR2 expression, which are involved in intercellular junctions and the angiogenic cascade, respectively, was higher in EVh groups (Fig. 5G,H). In fibroblasts, where cell integrity was best maintained among all cell types, subtle morphological differences were found (Fig. Suppl. 4E). Regarding ECM deposition, their expression of collagen type II and III was increased by EVh treatment, located mostly around the cellular body (Fig. 5I,J). Due to technical restraints, collagen type I was not investigated.

Interestingly, most macrophages in the MC-CC were identified within the MDA-MB-231 chamber (Fig. 6A). THP-1 was less frequently sighted in EVh-treated groups than in the controls, reinforcing our hypothesis that EVh protects the TME from phagoptosis. A direct co-culture with each cell line was executed to check individual responses to THP-1 with EVh treatment, leading to similar results. We found high levels of phagoptosis in both MDA-MB-231 and HUVEC wells, and cell debris were also found in fibroblasts (Fig. 6B). EVh treatment partially inhibited macrophage activity and recapitulated the original phenotype in all cell types. A corona of THP-1 cells attached to the fibroblasts was observed, suggesting that both cells may exchange information. In this case, fibroblasts became broader, branched, and more polarized, suggesting their activation (Fig. 6C). Cell viability was unchanged by EVh in all three settings (Fig. Suppl. 4F), but treatment increased gelatin invasion in tumor and endothelial cells upon direct culture with THP-1 and decreased in fibroblasts (Fig. 6D).

Discussion

The role of EVs in tumor progression and development is well described, and as new facets of tumorigenesis are revealed, their importance to cancer continuity becomes apparent. Tumor-associated EVs have been implicated in promoting angiogenesis (25), immune modulation (26), tumoral invasion (7,27), migration (28) and organotropic metastasis (29). The establishment of hypoxia in tumor progression promotes EV secretion in breast cancer, with distinct composition and characteristics (5–7). After describing the pro-invasive potential of hypoxic EV in comparison to normoxic EVs from TNBC cell line MDA-MB-231 (7), we show in this work that EVh differentiated THP-1 cells in M2-like macrophages, downregulating phagoptosis in tumor microenvironment models *in vitro*. Moreover, under the MC-CC model, EVh protected endothelial and tumoral cellular integrity, promoted pro-tumoral responses in endothelial cells and induced collagen synthesis in fibroblasts.

Monocyte recruitment and activation are crucial steps of the tumor-induced immune cascade, often triggered by the macrophage colony-stimulating factor (M-CSF) and other environmental cues (30). The differentiation of monocytes to macrophages *in vitro* relies on PMA to generate non-polarized cells, which may be induced to a pro-inflammatory M1 phenotype or an anti-inflammatory M2 profile, each with a distinct cytokine secretion pattern (17,31). We found that PMA-induced cells were enriched with pro-inflammatory components IL-1β, IL-6 and TNF-□, suggesting an M1-like profile, and the subtle reduction in IL-6 and sustained expression of anti-inflammatory IL-10 suggested that EVh promoted an M2-like phenotype (31–34).

The macrophage profiles were consistently maintained during co-culture with tumoral and endothelial cells, where phagoptosis was abundant in the control groups but avoided with EVh treatment. Cell death by phagocytosis results from the imbalance between the eat me and don't eat me signals from the target cell and their recognition by receptors in the phagocytic cell (24). In our models, EVh likely modulated don't eat me signals, such as CD47 and MHC-I, to improve cell survival, corroborating previous findings in which hypoxia inhibited macrophage phagocytosis in a HIF-1□-dependent manner (35,36). Further studies are required to confirm this hypothesis.

Studying the TME *in vitro* is challenging due to methodological, practical, and technological limitations. The recent development of techniques such as multicellular co-cultures, microfluidics and 3D culture has improved the complexity with which we can investigate basic mechanisms (37–39). A significant limitation of multicellular 3D co-cultures is the difficulty of evaluating responses in individual cells from the cultured mass, which often relies upon more refined and less accessible techniques (15). To circumvent this issue, we described a novel method to study the TME *in vitro* where cells are cultured *in tandem* in a commercially available system, henceforth named a multicellular circulating co-culture (MC-

CC). Results obtained from the MC-CC model were complementary to other co-culture techniques, including direct and indirect co-culture, but we have not investigated whether it is a better alternative to 3D culture. This method was previously described for studying the blood-brain barrier but has several limitations, including the number of available co-culture chambers, flow shear force, and lack of direct contact between chambers which reduces our ability to evaluate certain cellular responses (40). We acknowledge that different experimental designs, including other cell types, order of chambers, incubation conditions, flow rate, and media composition, may interfere with the results and have not been investigated in the present study.

Our experimental design permitted the evaluation of cell morphology alongside cytokine and enzyme secreted in the circulating medium across all MC-CC chambers. EVh cause stromal and tumor cells to favor a more pro-tumoral phenotype, likely due to the increased levels of IL-6, a cytokine whose overexpression is closely related to the development of various tumors, regulation of the acute phase of inflammation, and modulation of T and B lymphocytes (41). Furthermore, the high levels of IL-6 and IL-1β favor the inflammatory phenotype of cancer-associated fibroblasts, which is consistent with our evaluation of fibroblast response (42). There was a non-statistical tendency for an increase in IL-1β levels in EVh-treated cells, suggesting fibroblast activation, but additional testing to detect differences in □-smooth muscle actin (□-SMA) is required to confirm this supposition.

EVh in endothelial cells under MC-CC increased the expression of markers for cell-cell adhesion and proangiogenicity, including VEGFR2 and β-catenin (43). This may facilitate tumor development and metastatic spread by increasing the connections between endothelial and tumoral cells and encouraging angiogenesis (43,44). Intercellular junctions are crucial for endothelial integrity and depend on the function of multiple proteins, especially VE-cadherin (45). With nearly exclusive expression in vascular endothelium, VE-cadherin is

intracellularly linked to β-catenin, which directly connects to the cytoskeleton (46,47). In the indirect co-culture assay, we could not identify which adhesive molecules were influenced by EVh; however, multicellular co-culture under flow suggested an increase in β-catenin expression in HUVEC under EVh action. These conflicting results hint the tumor microenvironment is essential for maintaining endothelial intercellular adhesions but require additional investigation. Furthermore, we suggest that EVh disrupts endothelial cell adhesion to aid EVh-mediated tumor cell invasion.

Fibroblasts are structural support cells that can associate with tumors, transforming into cancer-associated fibroblasts (CAFs) through prolonged tumor-associated inflammation (TAI) signaling (42). TAI is sustained by continuous tumor mass growth and hypoxia, which have been described as determinants of the inflammatory phenotype of fibroblasts in pancreatic cancer (48). EVh increased fibroblast activity, modified their morphology and, in MC-CC, promoted the synthesis of type II and type III collagen, Both collagens are fibrillar and associated with increased ECM stiffness, a crucial factor for metastasis (49). Hypoxic extracellular vesicles also play a significant role in modulating fibroblast activity, leading to increased production of collagen types II and III. This process of extracellular matrix remodeling through MMP production and excessive collagen formation has been shown to promote tumor growth, progression, and resistance to treatments such as radiotherapy (43,44,50,51). In this setting, we show that hypoxic extracellular vesicles from triple-negative breast cancer impact fibroblast activity.

In summary, we found that tumor-derived hypoxic EVs show a distinct immune-modulating function, allowing the differentiation of M2-like macrophages in a TNBC in the TME *in vitro* and inhibiting endothelial and tumoral phagoptosis. EVh administration protected the endothelial and tumoral cellular integrity from phagoptosis. It promoted pro-tumoral and pro-angiogenic responses in endothelial cells, inducing collagen synthesis in fibroblasts and their

potential differentiation to CAFs. In addition, our work describes a novel method for studying the TME *in vitro* using a commercial system with great potential for investigating other tumor-related processes, including circulating tumor cells and metastasis.

Author's contribution

BCP: Conceptualization; Data curation; Formal Analysis; Funding acquisition; Investigation; Methodology; Project administration; Software; Writing – original draft. PHTB: Formal Analysis; Investigation; Visualization; AMM: Formal Analysis; Investigation; Writing – review & editing; CAC: Formal Analysis; Investigation; Methodology; Writing – review & editing; GG: Investigation; Visualization; LTG: Methodology; Visualization; MMG: Investigation; Writing – review & editing; ADZ: Investigation; Visualization; AMF: Conceptualization; Resources; Writing – review & editing; MRC: Funding acquisition; Resources; Software; Writing – review & editing; WFA: Conceptualization; Resources; Supervision; Writing – review & editing; HSSA: Conceptualization; Funding acquisition; Resources; Supervision; Writing – review & editing.

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Data availability

The data generated in this study are available within the article and its supplementary data files. Raw files are available upon reasonable request to the corresponding author. Detailed protocols have been submitted to the “protocols.io” platform (20–23). We have submitted all relevant data of our EV experiments to the EV-TRACK knowledgebase (EV-TRACK ID: EV240149) (52). This study does not contain artificial intelligence-generated data.

Ethics statement

The authors state that all research conducted in this study adhered to the highest ethical standards, ensuring integrity in research practices and compliance with institutional and legal requirements.

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Figures and Legends

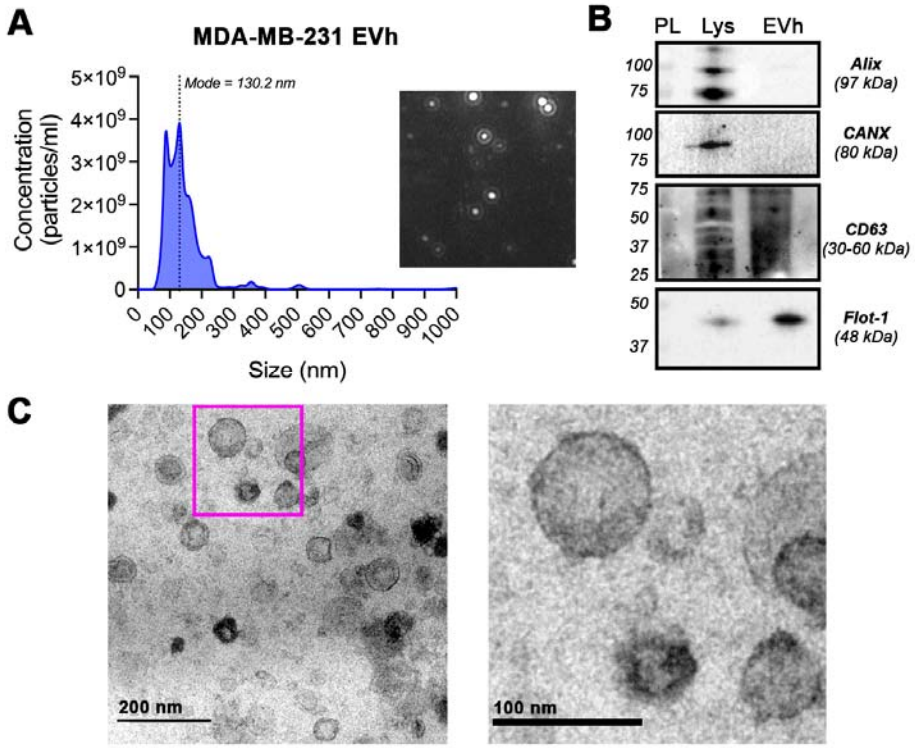

Figure 1. EVh characterization. (A) Size dispersion and EV abundance by nanoparticle tracking analysis of MDA-MB-231-derived EVh. (B) Detection of ALIX, calnexin (CANX), CD63 and flotillin-1. (C) Wide-field and close-up TEM images of EVh. Scale bars: 200 nm and 100 nm.

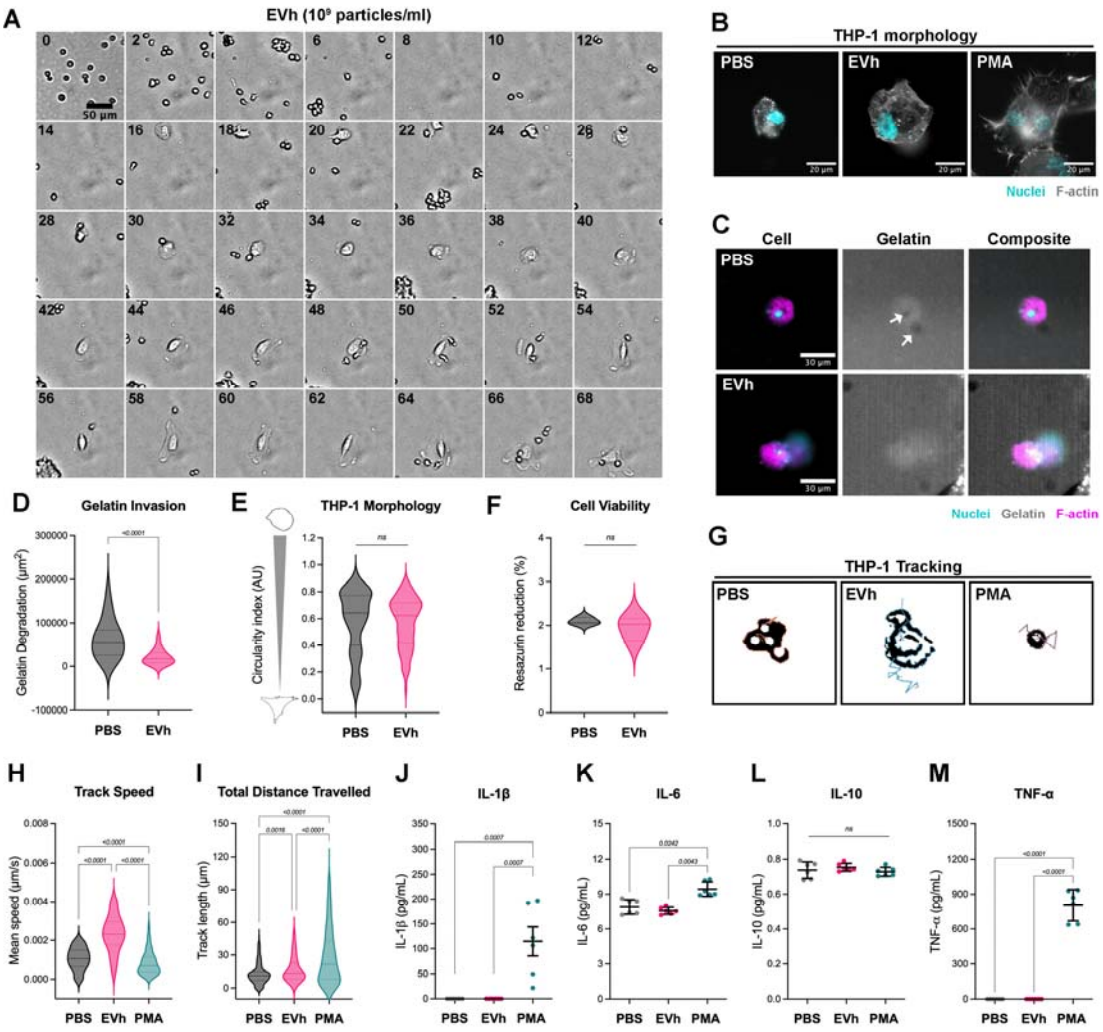

Figure 2. EVh-mediated macrophage differentiation. **A -** Timelapse tracking of THP-1 cells treated with EVh. Snaps from the same site were compiled in 2-hour increments over 3 days. Image brightness and contrast were adjusted for better visualization (scale bar: 50 µm). **B –** Representative images of THP-1 treated with vehicle (PBS), EVh or PMA after 72h differentiation (nuclei in cyan and F-actin in gray; scale bar: 20 µm). **C –** Representative images of THP-1 cells (nuclei in cyan, F-actin in magenta) treated with PBS or EVh on a gelatin coating (gray), with degradation spots identified by white arrows (scale bar: 30 µm). **D –** Quantification of degraded gelatin area (µm^2). **E -** Cell circularity index for quantification of THP-1 morphology. **F -** Cell viability of cells under invasion assay. **G –** Tracking of THP-1 trajectory over 24h. **H –** Average track speed of cells in 24 h (µm/s). **I -** Total distance traveled by cells in 24 h (µm). **J-M -** Quantification of cytokines TNF-α, IL-1β, IL-6, and IL-10, respectively. Assays were repeated over three different occasions with technical duplicates or triplicates. *p* values indicated above comparative bars with statistical significance.

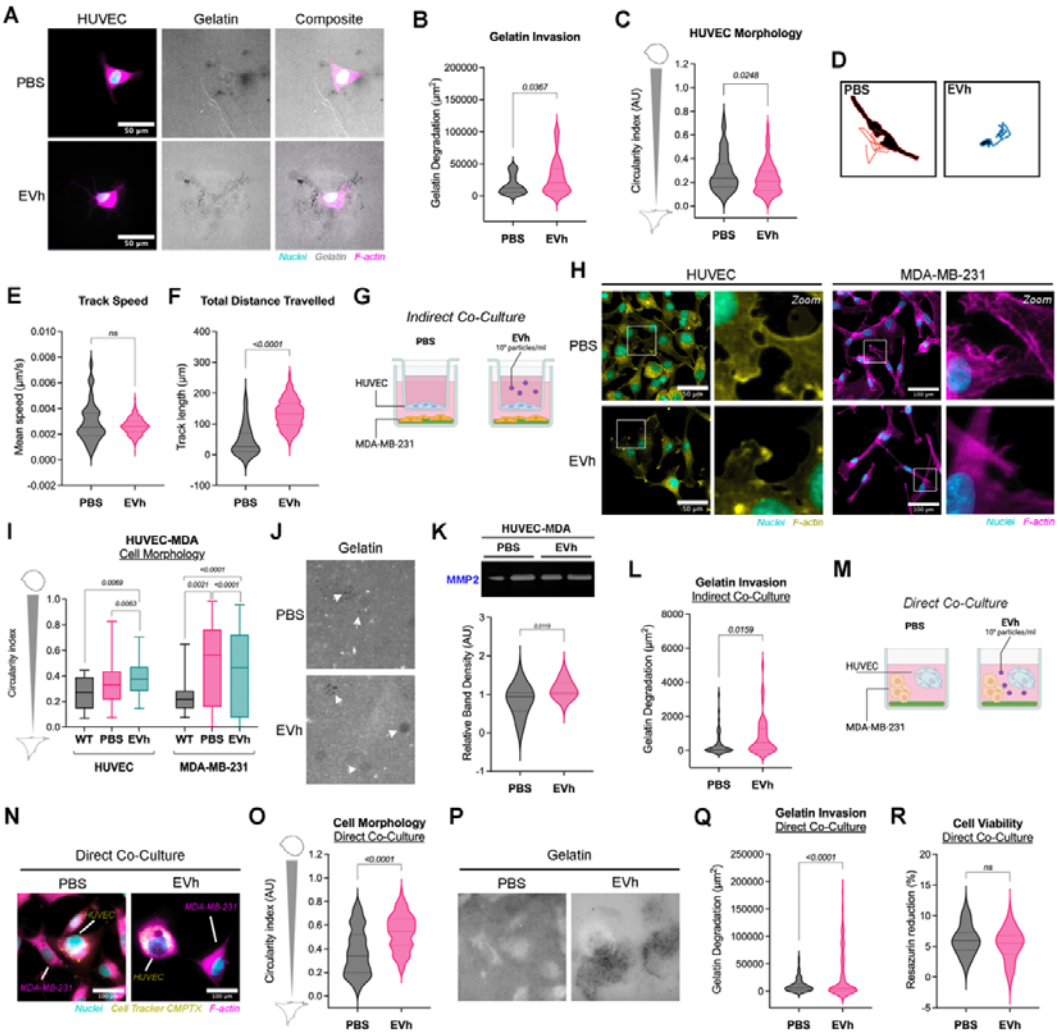

Figure 3. Endothelial cells respond to tumoral signaling. **A** - Representative images of HUVEC (nuclei in cyan, F-actin in magenta) treated with PBS or EVh on a gelatin coating (gray), with degradation spots in black (scale bar: 50 µm). **B –** Quantification of degraded gelatin area (µm^2) in individual HUVEC cultures. **C –** Cell circularity index for quantification of HUVEC morphology in individual culture. **D** - Tracking of HUVEC trajectory over 24h. **E –** Average track speed of cells in 24 h (µm/s). **F** - Total distance traveled by cells in 24 h (µm). **G –** Experimental design of the indirect co-culture assay, created in BioRender.com. **H –** Representative images of HUVEC (nuclei in cyan, F-actin in yellow; scale bar: 50 µm) in indirect co-culture with MDA-MB-231 (nuclei in cyan, F-actin in magenta; scale bar: 100 µm). **I –** Cell circularity index for quantification of HUVEC and MDA-MB-231 morphology in direct co-culture. **J –** Representative images of gelatin matrix (gray) with degradation spots identified by white arrows. **K** - MMP-2 detected by gelatin zymography and densitometry analysis to identify gelatinase activity in indirect co-culture. **L –** Quantification of degraded gelatin area (µm^2) in indirect co-culture. **M** - Experimental design of the direct co-culture assay, created in BioRender.com. **N –** Representative images of HUVEC (cytoplasm in yellow) in direct co-culture with MDA-MB-231 (nuclei in cyan, F-actin in magenta) seeded atop a gelatin coating (gray) (scale bar: 100 µm). **O** - Cell circularity index for quantification of combined cell morphology in direct co-culture. **P** - Quantification of degraded gelatin area (µm^2) in direct co-culture. **Q –** Cell viability of HUVEC and MDA-MB-231 direct co-

culture. Assays were repeated on three different occasions with technical triplicates. *p* values indicated above comparative bars with statistical significance.

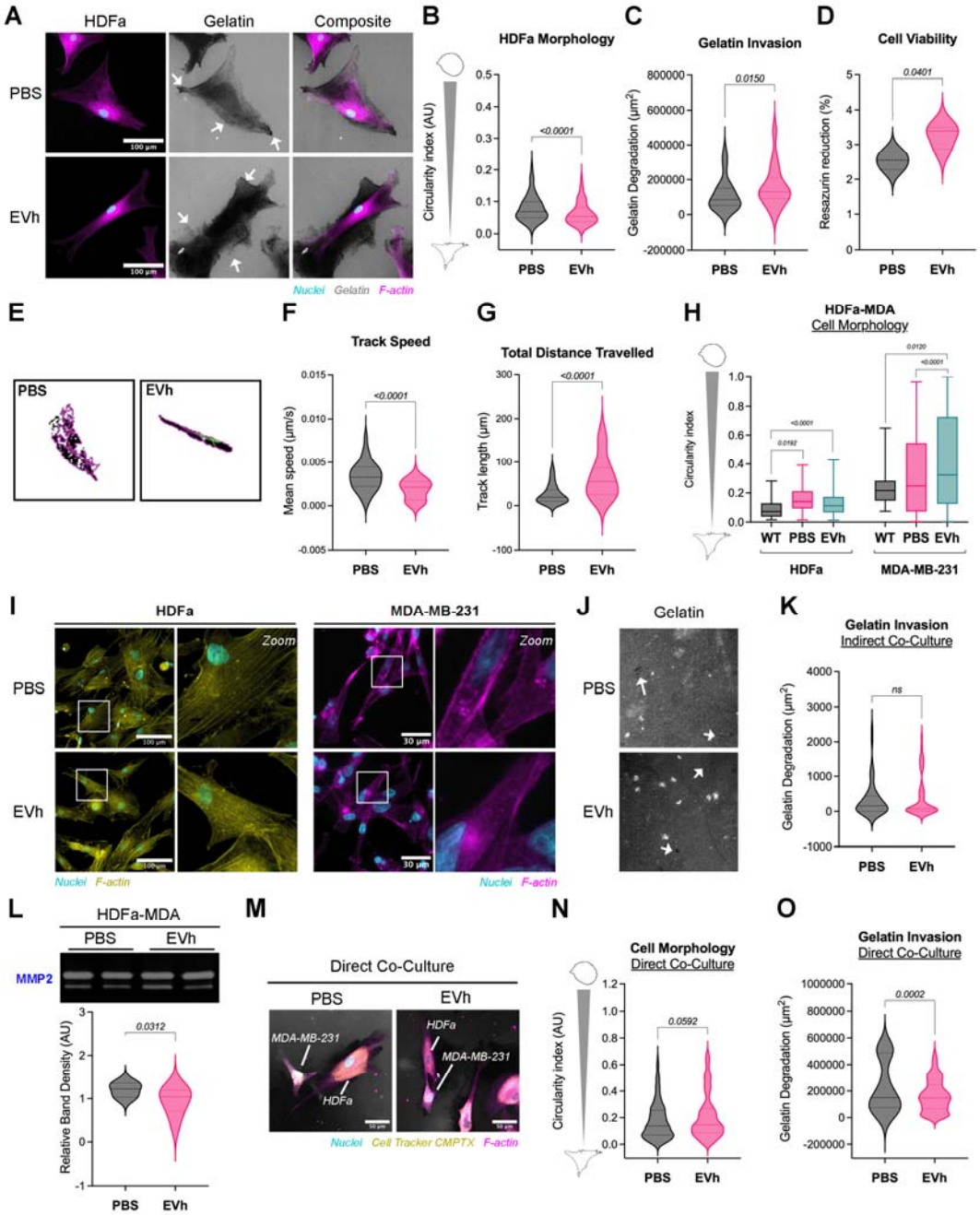

Figure 4. Fibroblast response to tumoral signaling. **A -** Representative images of HDFa (nuclei in cyan, F-actin in magenta) treated with PBS or EVh on a gelatin coating (gray), with degradation spots identified by white arrows (scale bar: 100 µm). **B –** Cell circularity index for quantification of HDFa morphology in individual culture. **C -** Quantification of degraded gelatin area (µm^2) in individual HUVEC cultures. **D –** HDFa cell viability in individual culture. **E -** Tracking of HDFa trajectory over 24h. **F –** Average track speed of cells in 24 h (µm/s). **G -** Total distance traveled by cells in 24 h (µm). **H –** Representative images of HDFa (nuclei in cyan, F-actin in yellow; scale bar: 100 µm) in indirect co-culture with MDA-MB-231 (nuclei in cyan, F-actin in

magenta; scale bar: 30 µm). **I –** Cell circularity index for quantification of HDFa and MDA-MB-231 morphology in indirect co-culture. **J -** Representative images of gelatin matrix (gray) with degradation spots identified by white arrows. **K -** Quantification of degraded gelatin area (μm^2) in indirect co-culture. **L –** MMP-2 detected by gelatin zymography and densitometry analysis to identify gelatinase activity in indirect co-culture. **M –** Representative images of MDA-MB-231 (cytoplasm in yellow) in direct co-culture with HDFa (nuclei in cyan, F-actin in magenta) seeded atop a gelatin coating (gray) (scale bar: 100 µm). **N -** Cell circularity index for quantification of combined cell morphology in direct co-culture. **O –** Quantification of degraded gelatin area (μm^2) in direct co-culture. Assays were repeated on three different occasions with technical triplicates. *p* values indicated above comparative bars with statistical significance.

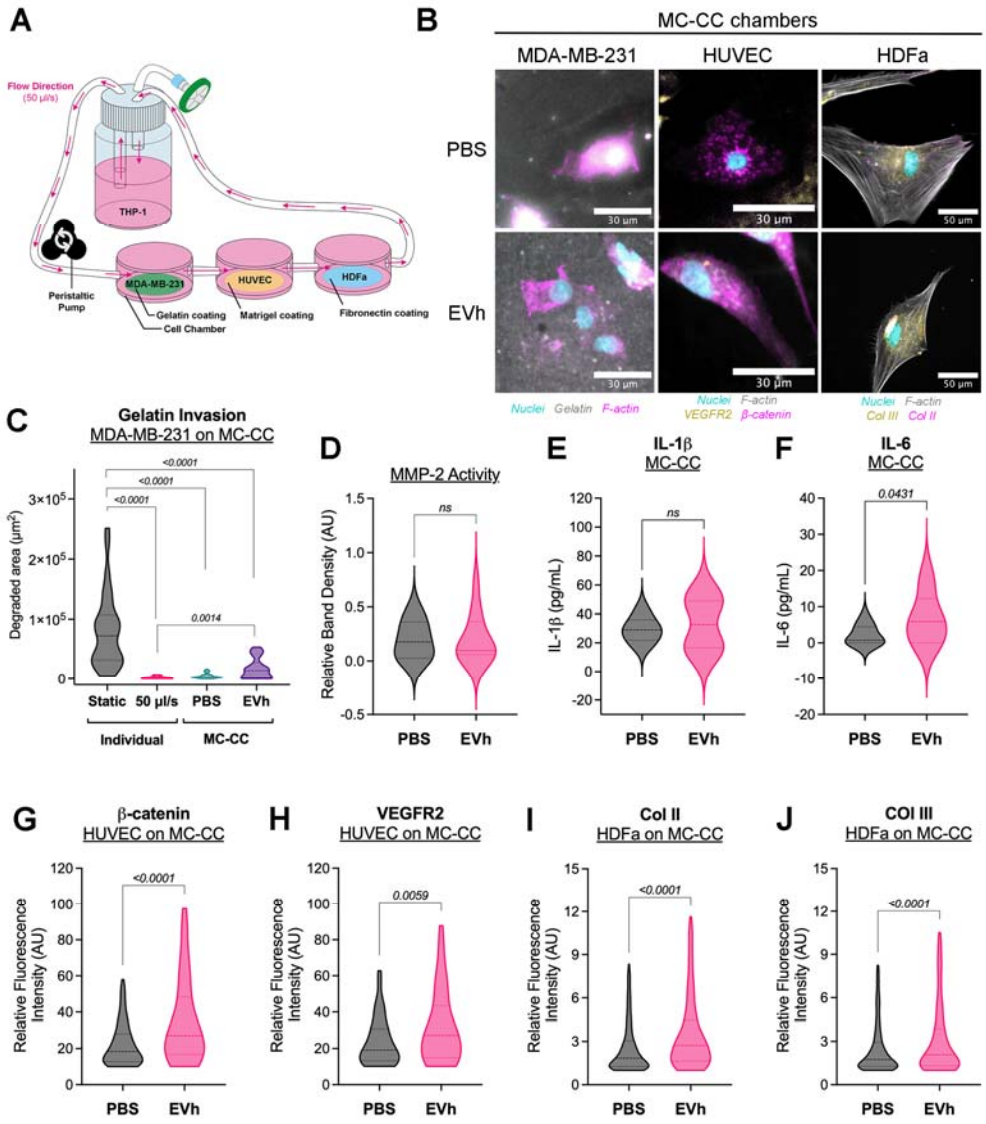

Figure 5. EVh inhibits phagoptosis in the TME. **A –** Experimental design of the multicellular circulating co-culture assay (MC-CC) using the QV500 system (Kirstall). **B -** Representative images of MC-CC chambers containing MDA-MB-231, HUVEC or HDFa treated with PBS or EVh. (scale bars: 30 µm and 50 µm). **C -** Quantification of degraded gelatin area (µm^2) in the MC-CC MDA-MB-231 chamber. **D –** Band densitometry of MMP-2 detected by gelatin zymography from the MC-CC conditioned media. **E,F –** Cytokines IL-1β and IL-6 probed from the MC-CC conditioned media by ELISA. **G,H –** Relative fluorescence intensity of β-catenin and VEGFR2 in HUVEC, respectively. **I,J –** Relative fluorescence intensity of collagen II and III in HDFa, respectively. Assays were repeated on four different occasions with a single replicate. *p* values indicated above comparative bars with statistical significance.

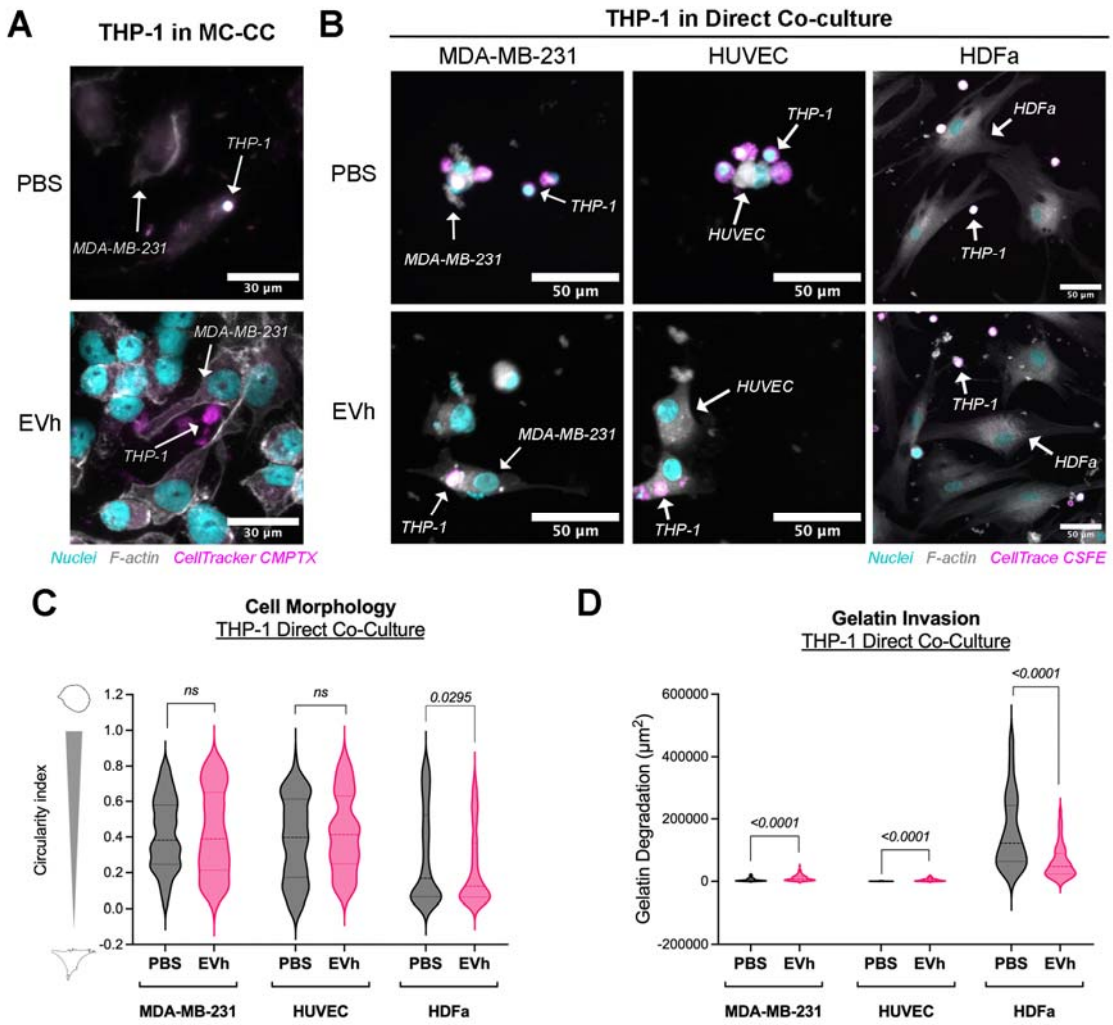

Figure 6. Macrophage remodeling by EVh in TME models *in vitro* A – Representative images of THP-1 (magenta, stained with CellTracker CMPTX) attachment to MDA-MB-231 MC-CC chambers (nuclei in cyan, F-actin in gray; scale bar: 30 µm). **B -** Representative images of THP-1 cells (magenta, stained with CellTrace CSFE) in direct co-culture with MDA-MB-231, HUVEC or HDFa (nuclei in cyan, F-actin in gray; scale bars: 50 µm). **C -** Cell circularity index for quantification of combined cell morphology in THP-1 direct co-culture with MDA-MB-231, HUVEC and HDFa. **D –** Quantification of degraded gelatin area (μm^2) in THP-1 direct co-culture with MDA-MB-231, HUVEC and HDFa. Assays were repeated on three different occasions with technical triplicates. *p* values indicated above comparative bars with statistical significance.