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Neural coding of choice and outcome are modulated by uncertainty in orbitofrontal but not secondary motor cortex

Juan Luis Romero-Sosa[1], Alex Yeghikian[1], Andrew M. Wikenheiser[1-3], Hugh T. Blair[1-3#] & Alicia Izquierdo[1-4#]

[1]Department of Psychology, University of California-Los Angeles, Los Angeles, California, USA.

[2]The Brain Research Institute, University of California-Los Angeles, Los Angeles, California, USA.

[3]Integrative Center for Learning and Memory, University of California-Los Angeles, Los Angeles, California, USA.

[4]Integrative Center for Addictions, University of California-Los Angeles, Los Angeles, California, USA.

Author contributions:

AI: Conceptualization, methodology, formal analysis, resources, writing-original draft, writing-review & editing, supervision, and funding. **JLRS**: Conceptualization, methodology, formal analysis, investigation, data curation, writing- original draft, writing- review & editing, software, and visualization. **AY:** Investigation, data curation, formal analysis, and visualization. **AMW:** Formal analysis and writing- review & editing. **HTB:** Formal analysis, resources, writing- review & editing, supervision, and funding.

Corresponding authors:

Juan Luis Romero-Sosa and Dr. Alicia Izquierdo

Abstract

Orbitofrontal cortex (OFC) and secondary motor cortex (M2) are both implicated in flexible reward learning but the conditions that differentially recruit these regions are not fully understood. We imaged calcium activity from single neurons in OFC or M2 during *de novo* learning of increasingly uncertain reward probability schedules. Predictions of choice were decoded from M2 neurons with high accuracy under all certainty conditions, but were more accurately decoded from OFC neurons under greater uncertainty. The number of choice- and reward-selective neurons was significantly higher in M2 than in OFC across schedules, but these proportions increased across levels of uncertainty only in OFC. Decoding accuracy of choice and outcome was predicted by behavioral strategies Win-Stay and Lose-Shift in OFC, but not M2. We also tested causal involvement of these regions with chemogenetic perturbation as rats learned increasing and then decreasing uncertainty schedules. Whereas inhibition of OFC neurons attenuated learning across all schedules, M2 neurons were found to support learning in the most certain reward schedule. Thus, OFC neurons preferentially encode choices and outcomes that foster a greater reliance on adaptive strategies under uncertainty. This reveals a novel functional heterogeneity within frontal cortex in support of flexible learning.

INTRODUCTION

Both the orbitofrontal cortex (OFC) and secondary motor cortex (M2) are involved in flexible reward learning and reward-motivated decision making. M2 receives dense inputs from diverse cortices, including sensorimotor, visual, retrosplenial, and orbital regions[1], and in turn, sends projections to striatum that fall along mediolateral and rostrocaudal gradients within M2[2], enabling precise control of actions. This anatomical connectivity positions M2 well to control action flexibly based on both sensory and motivation information. Functional studies have determined that M2 is essential for action updates [3] and that it enables the flexible assignment of either stimuli or motor antecedents to action plans[4-6]. Additionally, there has been discovery of different population dynamics in M2 depending on the strategy and stimulus modality that mice use in a flexible decision-making task[7]. Given its integration of sensory and motivation evidence, M2 has an essential role in higher-order movement control and orienting[8] but can also bias choice based on outcomes and choice history[9, 10]. Though a previous electrophysiological study indicated orbitofrontal cortex (OFC) is involved in updating outcome value from prior actions[11], authors found that M2 signaled choice earlier than any other area of rat frontal cortex, and remarkably also contained a value signal[12].

OFC also has a well-substantiated role in flexible decision making[13-16]. Several groups have reported choice-predictive neurons in OFC in equivalent or even larger numbers than value or outcome-encoding cells in this region[17-21]. One group found that over half of the neurons they recorded in OFC encoded both the direction of choice and the location of the goal, concluding that choice selective neurons in OFC represent spatial-motor information required to define specific goals and necessary actions to obtain them[21]. However, it is presently unclear if and how these representations change with uncertainty. By uncertainty we mean stochasticity, or moment-to-moment probabilistic outcomes[22] generating an 'expected uncertainty'[23-25], not uncertainty resulting from volatility, or the rate of change[22]. Indeed, a somewhat parallel literature suggests OFC may be critically involved in learning, especially under uncertain or probabilistic conditions[26-28], where it may set the expectation for the rest of learning[24, 25, 29, 30]. This is consistent with the idea that OFC stores long-term action values[24, 31], stimulus values[32], and causally contributes to an expected uncertainty or baseline risk in rats[33, 34]. This risk is signaled by single neurons in OFC in rats [35, 36] and in nonhuman primates[37, 38]. One study[39] reported that OFC neurons track expectations across a slow timescale (i.e., across-session probabilistic reversal learning) and are also recruited for trial-by-trial learning, or value updates, on a fast timescale (i.e., within a session). However, this study did not manipulate the reward uncertainty over time. Critically, aside from the

Hattori et al. (2023) study, recording and imaging studies in both rodent and primate species
typically assess the role of OFC in decisions in expert animals, not during the learning of risk or
expected uncertainty. Thus, it is unknown how choice and outcome cell selectivity may change
over the course of learning, especially with increasing uncertainty in OFC. All of the above
suggests that although M2 and OFC are involved in flexible learning and decision-making, their
involvement could dissociate over different timescales and increasingly uncertain conditions.

Using miniscopes, we compared populations of neurons and single cells in OFC and M2
over several sessions of learning in freely-behaving rats. Unlike the great number of studies
assessing these regions in well-trained animals, here we studied calcium dynamics during *de*
novo learning. We were primarily interested in determining if and when neural dynamics would
differentially predict choice and outcome in M2 vs. OFC, and which behavioral and task features
single cells were selective for in these regions across learning under uncertainty.

RESULTS

Learning under increasing uncertainty

Male and female Long–Evans rats first underwent stereotaxic surgery to infuse GCaMP6f
into either M2 or OFC (see Methods). In the same surgery, a GRIN lens was unilaterally implanted
over the site of the viral infusion (**Figure 1I-K**; photomicrographs of lens implant and calcium
indicator expression for all rats are shown in **Supplementary Figure 1**). Following surgical
recovery and baseplating, rats then underwent pretraining followed by six sessions of learning. In
a touchscreen-outfitted chamber, rats were required to initiate each trial by touching a center
stimulus and then make a choice to touch a stimulus on either the left or right side. If correct, rats
could retrieve a sucrose pellet reward from a food port located on the opposite side of the
touchscreen response zone, signaled by a light cue in the port. This visual cue appeared
concurrently with pellet delivery on rewarded, but not unrewarded, trials. The reward
contingencies reversed every 75 trials and increased in uncertainty on the last block of trials, with
reward schedules progressing from less to more reward uncertainty (100:0 to 70:30) (**Figure 1A-**
B). The probability of reward delivery depended on the rat's choice (left vs. right), with one side
delivering a higher probability of reward than the other. Each session consisted of 3 blocks of 75
trials (225 trials total); the response with the highest reward probability (left vs. right) was reversed
at the start of each block. Within each session, the probability ratio remained the same for the first
two trial blocks and then changed to a ratio with higher uncertainty for the last block of trials. As
shown in **Figure 1C**, sessions 1 and 2 used a ratio of 100:0 for blocks 1-2 and 90:10 for block 3

(Schedule 1), sessions 3 and 4 used a ratio of 90:10 for blocks 1-2 and 80:20 for block 3 (Schedule

2), and sessions 5 and 6 used a ratio of 80:20 for blocks 1-2 and 70:30 for block 3 (Schedule 3).

Figure 1D shows an example of a single rat's performance during a Schedule 2 session.

Results of an omnibus Generalized Linear Model (GLM) with a binomial distribution on

predictors of raw choices (0, 1) are found in **Table 1**. The main dependent variable was whether

the rats chose the higher reward probability location. Rats improved on this measure across trials

(GLM: $\beta_{trialnum}$ = 0.0559, p < 0.001) and blocks (β_{block} = 1.6671, p < 0.001), but not across

Figure 1. Probabilistic reversal learning task performance in freely-behaving, lens-implanted rats. (A) Schematic of trial timeline: Rats learn in an operant chamber outfitted with a touchscreen with three different interaction zones (left, center, and right) and a reward port on the opposite side of the chamber. A white circle is presented in the middle zone for 40 seconds, during which the rat is required to nosepoke to initiate a trial. Subsequently, two identical stimuli are presented on the left and right side of the screen for 60 seconds. The rat is required to nosepoke one of these stimuli, selecting either the left or right side. One of the sides has a higher reward probability than the other. The rat either gets rewarded 1 s after the nosepoke or undergoes a 5-second timeout. There is a 10 second Intertrial Interval (ITI) before a new trial begins. **(B)** A session consists of 3 blocks of 75 trials each, with two reversals, with the last block increasing in uncertainty. **(C)** Following surgery, recovery, handling, and pretraining to respond to touchscreen stimuli on either the left or the right, different schedules are introduced, with each schedule administered across two days each, and increasing in uncertainty. **(D)** Sample performance on Schedule 2 where the rat matches the value of the left side and adapts its behavior following a reversal. **(E)** There was a significant difference in accuracy, p(Better), by lens implant area (**Table 1**). **(F-H)** Performance measures by lens implant region to include latencies for Initiation **(F)**, Choice **(G)**, and Reward **(H)** for M2- and OFC-lens implanted rats across all the schedules. **(I)** Reconstructions of calcium indicator GCaMP6f and GRIN lens implant in all rats. **(J)** Photomicrograph of a lens in OFC. **(K)** Photomicrograph of a lens in M2.

schedules ($\beta_{schedule}$ = 0.1454, p = 0.58), **Figure 1E**. The M2 lens-implanted group exhibited
attenuated learning compared to the OFC lens-implanted group (β_{area} = 1.6671, p = 0.0416)
(**Table 1**).

Implanted animals also exhibited slightly attenuated learning compared to viral, non-
tethered controls (GLM: $\beta_{Implanted\_Lens}$ = -0.18, *p* < 0.01, **Supplementary Figure 2, Supplementary**
Table 1). For latency measures, non-tethered control animals initiated trials and collected rewards
more quickly but exhibited slower choice latencies (**Supplementary Tables 2-4**). Latencies to
initiate trials and collect reward did not differ in M2- vs. OFC-implanted animals, but choice
latencies were faster in the OFC-implanted animals than in the M2-implanted animals (GLM: β_{Area}
= -0.57, *p* < 0.01, **Figure 1F-H**, **Tables 2-4**).

Decoding choice from calcium activity

Binary support vector machine (SVM) classifiers were trained to predict *Chosen Side* (left
vs. right) from calcium traces aligned to the choice nosepoke (**Figure 2A-B)**. To determine the
most appropriate windows for statistical analysis, we generated peak-aligned heatmaps and
found cells that exhibited time-locked activity to either choice or reward cue **(Figure 2D-E)**.
Decoder training sets were balanced to include an equal number of rewarded/unrewarded and
left/right trials across all schedules (**Figure 2C**). In both M2 and OFC, choice decoding accuracy
ramped up from chance starting prior to trial initiation (suggesting that rats often anticipated which
side they would choose prior to trial initiation), and peaked ~500 ms after choice (**Figure 2F**). An
analysis of predictors of decoding accuracy in the bin just prior to choice (yellow shaded region in
Figure 2F) resulted in more accurate decoding in M2 than OFC (β_{Area} = -0.231, *p* = 0.0004), with
no effect of schedule ($\beta_{Schedule}$ = -0.0843, *p* = 0.19), but a significant Area x Schedule interaction
($\beta_{Area:Schedule}$ = 0.056, *p* = 0.0475), **Table 5**. *Chosen Side* was decoded from M2 with similarly high
accuracy under all probability schedules, whereas decoding was more accurate in OFC when the
schedule was more uncertain. Confirming this, post-hoc analyses on each schedule (**Table 6**)
revealed that decoding from M2 was more accurate than from OFC under conditions of higher
certainty (Schedule 1: β_{Area} = -0.174, *p* = 0.0027; Schedule 2: β_{Area} = -0.119, *p* = 0.0438), but under
the least certain schedule, OFC decoding accuracy increased to become equivalent to M2
decoding accuracy (Schedule 3: β_{Area} = -0.0621, *p* = 0.073). This certainty-dependent decoding
accuracy in OFC was not accounted for by experience (i.e., session) since experience was a
covariate in the GLM model and was not a significant predictor of choice accuracy ($\beta_{Session}$ =
0.0268, *p* = 0.234). Additionally, to check if the number of neurons used to train decoders affected
decoder accuracy, we entered this variable as a moderator in the GLM. We did not find any

significant effect or interaction with any other variable ($\beta_{NumCells}$ = -0.0008, p = 0.22, $\beta_{NumCells:Area}$ =

0.0008, p = 0.12, $\beta_{NumCells:schedule}$ = 0.0004, p = 0.17, $\beta_{NumCells:Area:schedule}$ = -0.0003, p = 0.19),

indicating that ensemble size did not significantly explain decoder accuracy results.

Decoding outcome from calcium activity

154 SVM classifiers were trained to predict *Trial Outcome* (rewarded vs. unrewarded) from

155 calcium traces aligned to the choice nosepoke (**Figure 2A-B**). At time points before the outcome

156 was known (that is, prior to 1 second after the choice nosepoke), the accuracy of outcome

Figure 2. OFC, but not M2, choice decoding accuracy increases with uncertainty. **(A)** Trial epochs selected for analysis of neural data: Initiation (left) and Choice (right). **(B)** Same epochs in A, shown in the trial timeline. **(C)** A flowchart of how the decoder was trained with balanced data and then tested on each trial. **(D)** Example of a cross-validated peak-aligned heatmap aligned at choice from an imaging session in M2 during schedule 2. **(E)** Same as D but in OFC. **(F)** Decoder accuracy from an SVM binary decoder trained using balanced neural data from either M2 or OFC. The solid line is the mean and the shaded region is the standard deviation. The decoder was trained to predict whether the choice would be left or right (*Chosen Side*). We found a main effect of Area and Area x Schedule interaction, with post-hoc analysis revealing better decoding in M2 than OFC in Schedule 1 and 2, but not Schedule 3. Decoder accuracy using shuffled neural data in both M2 and OFC is shown in black. **(G)** Similar to F but here the decoder was trained to predict whether the trial would be rewarded or not (*Trial Outcome*). We found main effects of Area and Schedule, but no significant interaction of Area x Schedule. *p< 0.05.

decoding was limited by the certainty of the outcome (which was highest under Schedule 1 and lowest under Schedule 3). In both M2 and OFC, decoding accuracy for *Trial Outcome* began ramping up from chance at trial initiation and then stalled at a plateau which persisted until the outcome was known, after which decoding accuracy began ramping upward again (**Figure 2G**). A GLM analysis of decoding accuracy in the bin just prior to the outcome (yellow shaded region in **Figure 2G**) resulted in, as expected, better decoding of outcome for certain than uncertain schedules (β_{Schedule} = -0.128, *p* = 0.0494). Decoding was also more accurate in M2 than OFC (β_{Area} = -0.138, *p* = 0.0248). There was similarly no effect of overall experience, or session number (**Table 7**). Here we also included the number of neurons as a moderator and again did not find a significant effect or interaction of ensemble size with any of the other predictors (β_{NumCells} = -0.00001, *p* = 0.98, $\beta_{\text{NumCells:Area}}$ = 0.0002, *p* = 0.59, $\beta_{\text{NumCells:schedule}}$ = 0.0002, *p* = 0.34, $\beta_{\text{NumCells:Area:schedule}}$ = -0.0002, *p* = 0.31). The same analysis was conducted on only co-registered cells (i.e., the decoder was trained on session 1 and tested on session 2, within the same schedule), **Supplementary Figure 6**. This revealed a similar pattern as **Figure 2F** and pointed to greater stability for Chosen Side decoding over Trial Outcome.

The ratio of choice, outcome, and reward-selective neurons increases with uncertainty in OFC, but not in M2

To link the decoder results to single cell selectivity, we removed an increasing number of cells with large beta coefficients (**Supplementary Figure 3**). As expected, decoder accuracy steadily dropped as more neurons with the largest beta coefficients were removed. We show heatmap plots of average trial activity for neurons that were significantly selective (p<0.05) for *Chosen Side*, *Trial Outcome*, and *Reward Retrieval* (**Figure 3A-C**), as well as trial-by-trial raster plots of calcium activity from individual example cells (**Figure 3G-I**).

M2 contained a higher ratio of neurons selective for *Chosen Side* than OFC throughout all schedules (GLM, β_{Area} = -0.238, *p* < 0.01). There was a significant interaction between area and schedule (GLM, $\beta_{\text{AreaxSchedule}}$ = 0.063, *p* = 0.007) (**Table 8**), reflecting the fact that the ratio of choice-selective neurons increased in OFC with uncertainty (GLM, β_{Schedule} = 0.06, *p* < 0.01, **Figure 3D**), but was not influenced by uncertainty in M2 (GLM, β_{Schedule} = 0.004, *p* = 0.78) (**Table 9**). Session (i.e., experience) was not a significant predictor of cell selectivity. Thus, the ratio of choice-selective neurons increases in OFC with uncertainty, but does not change with uncertainty in M2.

For the *Trial Outcome* epoch, we did not find a significant region difference in the ratio of outcome-selective cells, (GLM, β_{Area} = -0.10, p = 0.068). This ratio was also not modulated by experience alone as session was not a significant predictor. We did, however, find a significant main effect of schedule (GLM, $\beta_{Schedule}$ = -0.01, p = 0.044) and a significant interaction of area and schedule (GLM, $\beta_{AreaxSchedule}$ = 0.05, p = 0.03, **Table 10**), resulting from the fact that the ratio of outcome-predictive neurons in OFC increased with increasing reward uncertainty (GLM, $\beta_{Schedule}$ = 0.03, p = 0.009; **Figure 3E**), whereas the ratio of outcome-predictive neurons in M2 was similar regardless of uncertainty (GLM, $\beta_{Schedule}$ = -0.015, p = 0.44) (**Table 11**). It is somewhat paradoxical to observe that the proportion of neurons encoding trial outcome in M2 remained stable despite decreases in the certainty of reward. One explanation for this could be that under conditions of greater uncertainty, additional M2 neurons are recruited to encode choices.

Prior evidence suggests that value integration may occur at the time of *Reward Retrieval*[29, 40-42], so we performed an additional analysis to measure the proportion of neurons that were reward-selective during a 1-s time window centered on head entry into the reward port during rewarded trials versus a corresponding time window from unrewarded trials (see Methods). There was a larger proportion of reward-selective cells in M2 compared to OFC (GLM, β_{Area} = -0.27, p = 0.02) with a significant area by schedule interaction (GLM, $\beta_{AreaXSchedule}$ = 0.1, p = 0.048) (**Table 12**), resulting from the fact that OFC exhibited a higher proportion of reward-selective neurons for schedules with greater uncertainty (GLM, $\beta_{Schedule}$ = 0.09, p = 0.032, **Figure 3F**), whereas in M2 there was a similar proportion of reward-selective neurons across all schedules (GLM, $\beta_{Schedule}$ = -0.01, p = 0.68). M2 neurons thus appear to be more heavily recruited overall to encode reward as well as choices (described above) compared to OFC neurons. Finally, it is an increase in uncertainty and not experience (**Table 13**) that drives the increase in the ratio of reward-selective cells in OFC, since session was once again not a predictor of cell selectivity.

Figure 3. Decoder results are explained by changes in the ratio of selective cells across schedules. (A) Each row plots the difference in trial averaged activity of left trials minus right trials triggered by the time of choice. Cells plotted are all imaged neurons where we observed a significant effect of side, sorted from most negative beta coefficient to most positive one. The activity of the neurons was range-normalized. A value closer to 1 indicates more selectivity for the left side trials and a value closer to -1 indicates more selectivity for the right side. **(B,C)** Same as A but the difference is averaged activity of unrewarded trials minus rewarded trials, centered at time of reward cue (B) or reward retrieval (C). **(D)** Ratio of side selective neurons is overall larger in M2 but it increases with schedule only in OFC. **(E)** Ratio of outcome selective neurons increases with uncertainty only in OFC. **(F)** Ratio of reward selective neurons is overall larger in M2 but increases with schedule only in OFC. **(G)** Example of the z-score normalized activity of a significant side-selective M2 neuron on left and right trials. **(H)** Same as G but for a significant outcome-selective M2 cell. **(I)** Same as G but for a significant reward-selective OFC neuron. *p<0.05. ***p<0.001.

Behavioral strategies predict decoding accuracy in OFC, not in M2

To assess the degree to which different behavioral measures (**Figure 4A-F**) modulated decoding accuracy in OFC and M2, we analyzed the data using an omnibus mixed-effects GLM for *Chosen Side* (**Table 14**) and *Trial Outcome* (**Table 16**). As predictors, we included primary behavioral measures described above such as median initiation, choice, and reward latencies for the entire session (**Figure 4I-K**), as well as secondary measures such as Win-Stay (WS, proportion of trials when the rat repeated a response for which it had been rewarded on the previous trial) and Lose-Shift (LS, proportion of trials when the rat did not repeat a response for which it had been unrewarded on the previous trial), which we and others have previously used to assess adaptive strategies in both stimulus and action-based learning[24, 43]. As other predictors, we included a Flexibility Index (see Methods) that measured the animal's ability to adapt to trial block reversals, and a Perseveration Index, or the mean number of trials to shift choice behavior after being unrewarded, divided by the number of first unrewarded trials[44].

We found significant M2 vs. OFC area interaction effects for Win-Stay, Lose-Shift, and Perseveration Index in the bin just before the choice epoch (**Figure 4H**, **Table 14**). Post-hoc GLMs revealed that only initiation latencies (GLM: $\beta_{InitiationLatency}$ = -0.067, p = 0.0324) significantly predicted *Chosen Side* decoder accuracy for M2 (**Figure 4I**, **Table 15**). In contrast, WS (GLM: $\beta_{WinStay}$ = 0.843, p = 0.0008) and LS (GLM: $\beta_{LoseShift}$ = 1.466, p = 0.0240) positively predicted, whereas Perseveration Index (GLM: $\beta_{Perserveration\ Index}$ = -4.128, p < 0.0001) negatively predicted *Chosen Side* decoder accuracy in OFC (**Figure 4J**, **Table 15**). Similar post-hoc GLMs (**Figure 4K**, **Table 16**) found no significant behavioral predictors for *Trial Outcome* decoding accuracy in M2 (**Figure 4L, Table 17**), but again found that WS (GLM: $\beta_{WinStay}$ = 0.652, p = 0.0202) and LS (GLM: $\beta_{LoseShift}$ = 1.550, p = 0.0228) positively predicted and Perseveration Index (GLM: $\beta_{Perserveration\ Index}$ = -1.543, p = 0.0498) negatively predicted *Trial Outcome* decoding accuracy in OFC (**Figure 4M, Table 17**). These analyses indicate that complex, strategy-level behaviors contribute to decoding accuracy in OFC, but not M2.

Figure 4. Chosen Side and Trial Outcome decoding accuracy is predicted by behavioral strategy only in OFC. **(A-G)** Behavioral measurements that were entered as regressors in omnibus Generalized Linear Models (GLM). Latency measures **(A-C)** were calculated as medians of all trials for each session. WinStay and LoseShift **(D-E)** were calculated as the proportion of total trials that were WinStay and LoseShift at the session level. Perserveration Index and Flexibility Index **(F-G)** were calculated using formulas described in the Methods. **(H)** Beta coefficients for regressors where significant area (M2 vs. OFC) interactions were found in predicting decoder accuracy of *Chosen Side*. Modeling was performed only on the accuracy from the bin right before choice (highlighted in yellow, Figure 2) **(I)** Post-Hoc GLM performed on sessions from rats with lenses in M2. **(J)** Post-Hoc GLM performed on sessions from rats with lenses in OFC. Strategy indices were significant predictors of *Chosen Side* decoding accuracy in OFC, but not in M2. **(K)** Beta coefficients for regressors where significant area (M2 vs. OFC) interactions were found in predicting *Trial Outcome*. Similarly, to H, analyses were only performed on the bin before reward cue (highlighted in yellow, Figure 2). **(L)** Post-Hoc GLM performed on sessions from rats with lenses in M2. **(M)** Post-Hoc GLM performed on sessions from rats with lenses in OFC. Strategy indices were significant predictors of *Trial Outcome* decoding accuracy in OFC, but not in M2.

OFC, but not M2, neurons preferentially support adaptive learning under uncertainty

Male and female Long-Evans rats underwent stereotaxic surgery to infuse hM4Di DREADDs in either OFC or M2 (see Methods). After following the same pretraining scheme as rats with lens implants, this cohort performed schedules first in order of increasing (i.e., ascending) uncertainty- identical to the cohort with lens implants- and then in order of decreasing uncertainty, which rats with lens implants did not experience. Each schedule was administered

Figure 5. OFC, but not M2, neurons support learning under all uncertainty levels. (A) Following surgery, recovery, handling, and pretraining rats were introduced to different schedules, first in increasing order of uncertainty followed by decreasing (reverse) order. Each schedule was administered across two days, the first under CNO inhibition and the second following an injection of vehicle (VEH). **(B)** Win-stay behavior was reduced across all schedules following OFC inhibition, but not M2. **(C)** OFC inhibition resulted in attenuated accuracy throughout learning. In contrast, M2 inhibition impaired only Schedule 1 performance (CNO: β_{Drug} = -0.2018, p < 0.0001). **(D)** Reconstructions of DREADD expression across all rats. **(E)** Representative photomicrographs of hM4Di-mCherry DREADD expression in M2 and OFC. **(F)** OFC, but not M2, inhibition slowed choice latencies. *p<0.05 ***p<0.001.

twice- with CNO inhibition on the first session and an injection of vehicle (VEH) on the second

session- before the rat advanced to the next schedule (**Figure 5A**).

As in **Figure 1E**, accuracy was measured as the raw choice (0,1) of the higher reward
probability option (see Methods). Using omnibus GLMs with binomial distribution (**Tables 18 and
19**), we assessed if there were interactions between schedule (1, 2, 3), order (ascending or
descending uncertainty), and drug (CNO, VEH). OFC inhibition impaired accuracy across all
schedules, as the effect of drug did not interact with schedule or order (CNO: β_{Drug} = -0.2831, p <
0.02), **Table 18**. In contrast, while there was also an effect of inhibition of M2 on accuracy (CNO:
β_{Drug} = -0.4698, p < 0.0001), there was a drug by schedule interaction (**Table 19**), which revealed
its effect was in Schedule 1 (CNO: β_{Drug} = -0.2018, p < 0.0001) (**Table 20**). Due to an interaction
of drug by order, we conducted a post-hoc analysis which revealed that decreased performance
following M2 inhibition was in the ascending (CNO: β_{Drug} = -0.0950, p = 0.0176) but not descending
order (**Table 21**). Consistent with prior findings, OFC neurons support first reversal experience[24,
31, 45] as well as learning under all subsequent uncertainty levels. In contrast, the largest effect size
for M2 neuron involvement during learning occurred in Schedule 1.

We also examined the effect of inhibition on adaptive strategies (**Tables 22–25**). OFC
inhibition reduced the Win-Stay strategy across all schedules (CNO: β_{Drug_CNO} = -0.020, p < 0.01),
with no effect on Lose-Shift. M2 inhibition had no significant effect in the use of either strategy
(**Figure 5B**). Finally, we analyzed initiation, choice, and reward latencies following inhibition.
While M2 inhibition had no effect on choice latencies (**Table 26**), OFC inhibition significantly
slowed choice latencies (**Table 27**) (**Figure 5F**). All analyses included only animals with
confirmed, bilateral expression (**Figure 5D–E**).

DISCUSSION

We imaged single-cell activity in M2 and OFC while freely-moving rats underwent learning.
Using a binary support vector machine decoder, we found evidence of functional heterogeneity in
frontal cortex at both the population- and single-cell level. Specifically, M2 consistently contained
more information about choice on individual trials than OFC, yet decoder accuracy increased with
uncertainty, not just experience, only in OFC, not M2. We found that decoder accuracy was
predicted by adaptive strategies in OFC, but not M2. We also determined that M2 and OFC
neurons exhibited different neural trajectories in learning trial types: these changed dynamically
in more uncertain schedules in OFC, but not M2 (**Supplementary Figure 4**). Our single cell
analyses revealed a recruitment of an increasing ratio of choice-, outcome- and reward- selective

neurons in OFC with uncertainty, not M2. Finally, chemogenetic inactivation of each of these regions confirmed functional roles for OFC, but not M2, as preferentially involved in learning under uncertainty. Collectively, these results suggest that neurons in OFC are more engaged in conditions with greater task demands that require adaptive strategies, such as increased uncertainty during learning.

There are several novel and confirmatory findings here. First, studies of neural activity in the frontal cortex in mice[46], rats[47, 48], and especially non-human primate subjects [49] typically involve extensive training prior to surgery to ensure that the animal is a task expert prior to any recording or imaging. In contrast, in the present experiment, rats learned the task *de novo* while we imaged single cells from these regions, which allowed us to determine that the dynamics in OFC changed as the uncertainty increased. Quite differently, M2 signaling of choice was robust and stable throughout the experiment. With the exception of two single-unit electrophysiological studies[11, 12], these two brain regions have not been systematically compared for their signaling in a reward-motivated task until now, and importantly, the previous studies administered a dynamic foraging task where subjects were not required to learn any task structure. We used a similar task to that in a recent report by Hattori and colleagues[39], but those authors also did not vary the uncertainty in the reward environment, as we did here. Consistent with their findings, we confirm that OFC plasticity is required for 'across session' learning (what authors call "meta-learning"), and here we extend this to conditions of increasing uncertainty encoded in OFC, but not in M2, neurons.

Studies following manipulations in rat OFC in flexible reward learning have included targeting the entire ventral surface [50, 51] or comparisons of medial vs. lateral OFC[27, 28]. Similar to a recent study using identical targeting[24], here we examined the role of ventrolateral OFC, a subregion not as often probed as medial and more lateral OFC[52], and found evidence consistent with its encoding of expected uncertainty[25]. Specifically, several studies using different methods [32-34, 53] suggest OFC must be 'online' to establish a baseline risk associated with probabilistic outcomes from which to gauge meaningful changes (i.e., reversals or block switches). OFC-inhibited animals also do not employ effective Win-Stay strategies[24], consistent with the GLM modeling results of the imaging experiment and the causal chemogenetic perturbation result here. A limitation in the present experimental design is that we inactivated OFC or M2 neurons on the first experience with each schedule (i.e., CNO was administered on the first and VEH was administered on the second of two sessions of each schedule). While there is converging evidence that OFC is most involved in first reversals[24, 31, 45] we were mainly interested in the pattern across schedules, and across M2 vs. OFC, not within a schedule. Indeed, we found the pattern

of learning attenuations was different across schedules, and the two regions supported different facets of learning.

We also find evidence that OFC neurons learn task structure details that are ultimately useful in predicting the consequences of decisions in uncertain and/or novel conditions[54]. In support of this, we found that OFC neurons do not encode choices as well as M2 neurons during more certain schedules, evidenced in both decoder and stability analyses where M2 decoder accuracy is greater than that of OFC in the more certain schedules. Additionally, we observed an increase in the proportion of neurons that signaled reward with increasing uncertainty in OFC, not in M2. These findings are consistent with the idea that OFC has a pivotal role in either using or creating a task map[55, 56]: just as a spatial map helps one move better through space, a task map would help with adaptive behavioral strategies. We show evidence of this with decoder performance in OFC positively predicted by adaptive strategies WinStay, LoseShift, and negatively correlated with an inflexibility measure like Perseveration Index.

By comparison to OFC, M2 is often overlooked in learning studies due to its presumed stronger role in motor skill and action execution in both rodents and non-human primates[57-59]. There is evidence that M2 encodes task-relevant features, such as whether a rat will choose the left or right side in a bandit task [12] and a risky or safe option[60]. Thus, our results provide additional evidence that M2 encodes important task parameters such as choice and trial outcome. Indeed, M2 is relatively unchanging: its encoding of choice remained robust and dynamics in this region contained a "full picture" of task-relevant features even from the initial phases of learning. In support of this, we found that M2 contained information related to blocks throughout all different epochs of a trial, more so than OFC **(Supplementary Figure 5)**. Additionally, chemogenetic inactivation of M2 preferentially impacts learning in the most certain condition (Schedule 1). Given that M2 biases decision-making processes[5, 10], a read-out of "intention" could be a useful brain-machine interface application, given its stable coding.

These findings demonstrate distinct roles for M2 and OFC in processing task-specific information during learning under uncertainty, with M2 neurons maintaining robust, stable activity throughout learning, and OFC neurons exhibiting significant adaptations in response to uncertainty. This suggests a mechanism by which OFC neurons make adjustments to dynamic environments, while M2 provides a more consistent response control mechanism. Interestingly, unlike the present findings that suggest a role for rodent OFC in both outcome and reward probability encoding, primate OFC is not as important for encoding of reward "availability, as indexed by its probability"[61]. Instead, OFC is more specialized in primates for value and

349   "desirability," not probability. This may be an important primate-rodent difference to consider when

350   interpreting experimental findings in frontal cortex across species in the future.

352   **Acknowledgements**

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METHODS

Subjects

363     A total of N=32 Long-Evans rats was used. Subjects for single-cell calcium imaging were

n=7 adult Long-Evans rats: n=4 (3 males, 1 female) with lenses in OFC and n=3 (2 females, 1

male) with lenses in M2. Subjects for chemogenetic manipulation were N=25 adult Long-Evans

rats. Of these, seven animals were excluded from analyses: one due to a failure to complete

pretraining and six due to weak or unilateral viral expression. Thus, the total for the chemogenetic

experiment was N=18: OFC: n=9, 5 females; M2: n=9, 5 females. Rats were obtained from a

colony of GAD-Cre transgenic rats (RRRC#751; Rat Resource and Research Center, MO) / TH-

Cre (RRRC#659; Rat Resource and Research Center, MO), genotyped as wildtype. Rats were

pair-housed when they arrived at our housing facilities and then single-housed after undergoing

surgery. Rats were housed in plastic transparent tubs filled with bedding media made of wood

shavings in a room with a reverse 12-hour light cycle. Pretraining and testing occurred during their

dark period, seven days per week until all experimental sessions were completed. Motivation was

achieved through a progressive food restriction until the weight was kept above 85% of their initial

free-feeding weight. Rats had free access to water at all times. Before training, rats were handled

for 10-20 min for a minimum of 5 consecutive days. Subsequently, rats were introduced to a three-

finger hold that enabled intraperitoneal injections or the attachment of a miniscope after

baseplating. Experiments were conducted with the approval of, and following the guidelines
established by, the Chancellor's Animal Research Committee at UCLA.

Behavioral Pretraining

Our operant chambers were outfitted with a touchscreen on one end, divided into three
zones (left, center, and right), and a reward port opposing the touchscreen (Models 80604 and
80604A, Lafayette Instrument, Inc., Lafayette, IN). Ten sucrose pellets were placed in the rat's
homecage one day before training to acclimate them to the pellets. The rats were placed in the
operant chambers on the first training day, and 5 pellets were dispensed. The rats remained in
the chambers for 20 min, where they could explore, eat the pellets, and become habituated to the
testing environment. The criterion for the rat to move on to the next phase is eating all 5 pellets
within the allotted time. The following day, they begin Initial Touch Center (ITC), which was the
first time the rat had learned to interact with the touchscreen. A trial commenced with the display
of a white rectangle in the center zone to indicate the rat is required to interact with it (e.g., nose
poke or paw touch). Once the rat interacted with the center zone, a pellet was immediately
delivered to the reward port. The rat only had 40 s to interact with the center zone before the
rectangle disappeared, and the trial was counted as an initiation omission. If the rat successfully
interacted with the center zone and collected the sucrose pellet, the trial was considered a
committed trial. For the rat to move on from ITC, it had to complete 60 committed trials within 45
min. The final phase of pretraining was Immediate Reward (IR). In this phase, the rat was required
to initiate the trial by touching the center zone when the white rectangle is presented, and then it
was required to touch a stimulus presented pseudo randomly either on the left or right zone of the
touchscreen. The rat had 60 s to interact with this stimulus to obtain a reward. Similar to the ITC
criterion, for the rat to complete IR, it had to complete 60 committed trials within 45 min. After
completing IR, rats were considered ready for surgery.

Miniscope Assembly and Imaging

We purchased assembled Miniscope V4.4 kits from *Open Ephys*. The miniscope was
placed in a 3D-printed, custom-designed, and constructed helmet (a link to the 3D design would
be excellent here). The 3D-printed helmet enabled us to utilize a more resilient SMA connector
for rat movement. Once the miniscope was fitted inside the helmet, we ensured LED power, the
electrowetting lens, and the CMOS were working correctly. The tether used to connect the
miniscope to the operating system had a female SMA connector on one end and a male HDMI
connector on the opposite end. A motorized commutator from Doric Lenses (Part Number:

AERJ_24_HDMI+4) allowed the tether to rotate with the rat. The commutator transferred the

signal to the miniscope DAQ v3.3, also obtained from *Open Ephys*. The DAQ sent the signal over

USB to a recording computer. Additionally, the DAQ produced a TTL pulse every time the

miniscope recorded a frame of data. These TTL pulses were recorded in parallel with TTL pulses

sent by the operant chamber whenever certain behaviors within a trial were performed (i.e., trial

initiation, reward port entry) and were used to time lock neural data to specific epochs within trials

and within single sessions. These TTL pulses were recorded using a CerePlex Direct data

acquisition system and later used to sync the neural data. The USB signal from the miniscope

was recorded onto a Framework Laptop 13 (11th Gen Intel) using the Miniscope DAQ Software

(build v1_11). All imaging was conducted at a rate of 20 frames-per-second (fps).

GCaMP and Lens Implant Surgeries

Rats were anesthetized with isoflurane (5% induction, 2% maintenance in 2.5 L/min O_2)

and placed in a stereotaxic apparatus (Kopf Instruments, Tujunga, CA). After clearing the skull,

we leveled the skull by ensuring the difference in DV around bregma was not greater than 0.05

mm. An etchant was applied to the skull for 1 min to ensure the adhesive would adhere. After

cleaning the etchant, we partially drilled seven tiny holes that would be used for seven anchor

screws (Fine Science Tools #19010-00). Two anchor screws were placed in the very front, two

directly posterior from bregma, two anterior to lambda, and one contralateral to where the GRIN

lens would be placed. Subsequently, a 1.2 mm diameter craniotomy was drilled above the brain

regions of interest. For OFC placement, the coordinates were AP: +3.7mm, ML: +2.5mm, and for

M2 placement, the coordinates were AP: +2.0mm, ML: +0.7mm. The craniotomy was cleaned to

ensure that the superior sagittal sinus was intact, and then the dura was carefully removed. A

1 L Hamilton syringe and pump delivered GCaMP6f (AVV9-CamKII-GCaMP6f-WPRE-SV40,

Addgene, # 100834) through cannulae and injectors in either OFC or M2. For OFC surgeries, we

unilaterally infused virus into two different sites (AP: +3.7mm, ML: +2.5mm, DV: −4.2mm

and −4.6mm). For M2, we also infused unilaterally into two different sites (AP: +2.0mm, ML:

+0.7mm, DV: −1.4mm and −0.9mm). We dialed down 0.05mm more ventrally in either region to

create a pocket for infusions. We then injected 0.5 L of GCaMP6f per site, thus totaling 1 L of

virus per surgery. The syringe was placed in a Hamilton pump with a speed of 0.1 L per minute.

Following each infusion, 5 min elapsed before the cannula was moved. Lens implantation

occurred immediately after viral infusions. A 23-gauge needle was attached to the stereotaxic

instrument, zeroed at bregma, and then lowered to make clear a path for the lens. Next, the lens

was lowered to the halfway point between the DV coordinates as above using a vacuum. After

the GRIN (Gradient Index) lens had been placed in its location, we used viscous Loctite super glue to glue the sides of the lens to the top of a dry skull, then added a layer of Metabond bone cement that coats the lens, anchor screws, and the most superficial layer of the skull. After allowing 5-10 min for the Metabond to dry, we poured opaque dental dement on the Metabond to fully cover the anchor screws and cover the sides of (but not obscure) the GRIN lens. Before the dental cement thoroughly dried, we affixed a custom 3D-printed lens cover. After drying the dental cement, we applied a layer of topical antibiotic to the edge of the dental cement head cap. Animals were administered carprofen (5mg/kg s.c.) and saline (1cc) prior to completion of surgery. Post-operative care consisted of carprofen (5mg/kg s.c.) for five days following surgery.

We waited 3 to 4 weeks post-surgery to allow for GCaMP expression before attaching the baseplate. We anesthetized the rat for baseplating, removed the lens protector, and cleaned the top of the dental cement head cap. We attached a baseplate to a fully assembled miniscope using a custom 3D holder. We lowered the miniscope and the baseplate until we could focus entirely on the top of the implanted GRIN lens, searching for visible expression in the area of interest, then fixing the baseplate using dental cement. GCaMP expression and lens implant reconstructions are shown for OFC (**Figure 1D, left**) and for M2 (**Figure 1D, right**), along with representative photomicrographs (**Figure 1E-F**). All GCaMP and lens placements for all animals are shown in **Supplementary Figure 1**.

Viral Surgeries

Craniotomies were performed similarly to the GCaMP surgeries. To express inhibitory hM4Di DREADDs (Designer Receptors Exclusively Activated by Designer Drug) in cortical excitatory neurons, a CaMKIIα promoter drove the hM4Di-mCherry sequence (Addgene viral prep # 50477-AAV8, Watertown, MA). This construct was infused bilaterally into either OFC or M2 at the same coordinates as previously described. 0.5 L was infused per site, thus totaling 1 L per hemisphere and 2 L per surgery. The scalp was carefully closed using self-dissolving sutures to ensure a fast and less invasive recovery.

Experimental testing began a minimum of three weeks after surgery to allow for receptor expression. Reconstructions of DREADD expression are shown for OFC (**Figure 5D, left**) and for M2 (**Figure 5D, right**), along with representative photomicrographs (**Figure 5C**).

Behavioral Testing

After the baseplating surgery, rats were again put on food restriction before behavioral testing: they were eased into a restricted diet over three days to no more than 85% of their free-feeding weight. Additionally, rats were once again habituated to a gentle hold and plugging the miniscope into the baseplate. Rats were given a second round of pretraining as an acclimating imaging session using a miniscope. We ensured rats performed many trials in the IR phase (> 200 trials in 45 mins) before moving the rat onto the main behavioral task.

Learning of Probabilistic Reversals. Similar to pretraining, a trial began when a circle was displayed in the center zone of the touchscreen. The rat must interact with it by nose-poking or touching it with its paw within 40 s; otherwise, it is scored as an initiation omission. After touching the circle, two identical stimuli were presented on both the right and left touchscreen zones. The rat had to interact with either the left or the right zones. The rat had 60 s to make a choice otherwise, it was scored as a choice omission. One of the zones was associated with a higher probability of reward, while the other was associated with a lower probability of reward. If the rat received a sucrose pellet reward, it was dispensed in the reward port on the opposite side of the chamber, and the subsequent trial began after a 10-s inter-trial interval (ITI). If the rat did not receive a reward, a 5-s time-out was imposed in addition to the 10-s ITI. If the rat initiated a trial and made a choice within the allotted time, the trial was scored as a committed trial regardless of whether it was rewarded. Food motivation was assessed purely by the number of committed trials, not accuracy. In each session, there were three blocks of 75 committed trials each.

From one block to the next, the location of the highest-valued side reversed. The probability of each option itself varied by block according to schedule, with Schedule 1 including deterministic blocks (100:0, 0:100, 90:10) and Schedules 2 (90:10, 10:90, 80:20) and 3 (20:80, 80:20, 70:30) being entirely probabilistic. There was an increase in uncertainty over testing. There were two primary reasons for this design: 1) reversals (i.e., block design) keep the animals engaged and less likely to fall into a motor habit of always responding left, for example, and 2) we enhance the chance of including the same cell population during learning at one level of uncertainty (e.g., 100-0) with another (90-10), if these blocks occur in the same session. Task structure is visualized in **Figure 1A**. Violin plots for all initiation, choice, and reward latencies are shown by schedule (**Figure 1I-K**), and the median per session collapsed across all schedules for each of these groups.

Chemogenetic inhibition. In the chemogenetic cohort (n=18), inhibition of OFC or M2 was achieved with systemic administration of clozapine-N-oxide, CNO (6 mg/kg, i.p., in 95% saline, 5% DMSO; HelloBio, Princeton, NJ) 30 min prior to behavioral testing. To control for the

nonspecific effects of injection and handling, vehicle solution, VEH (95% saline, 5% DMSO), was

alternatively administered. The effects of CNO were assessed in a within-subjects design where

rats completed each schedule twice, first under CNO administration and second with an injection

of VEH. To investigate learning in changing environments while controlling for the effect of task

experience in navigating the task, rats first completed the schedules in increasing order of

uncertainty, then repeated these schedules in decreasing order **(Figure 5A).**

Histology

Following completion of behavioral testing, all rats were euthanized with an intraperitoneal

dose of sodium pentobarbital and sodium phenytoin (Euthasol® Euthanasia Solution) and

transcardially perfused with PBS and a 4% formalin solution. Brains were extracted, placed in 4%

formalin solution for at least 24 h, and subsequently left in a 30% sucrose solution for at least 24

h. Forty μm coronal sections were mounted onto slides and coverslipped with DAPI to visualize

GCaMP or hM4Di-mCherry expression. Imaging was performed using a LSM 900 confocal

microscope (Zeiss) and BZ-X710 florescence microscope (Keyence).

Post-processing calcium imaging pipeline

Miniscope recording videos were processed using custom Python code and the Python

version of CaImAn[62]. We concatenated videos and cropped them to focus the field of view (FOV)

with the neural activity. We then passed it through the motion correction algorithm in CaImAn,

with the central parameter *gSif_filt* set between 6 and 12 for our data. All of the motion correction

for our data was conducted with the piece-wise rigid motion correction Boolean setting set to true.

We performed the non-rigid algorithm if this algorithm did not fix the motion artifacts. Once the

motion correction was completed, the motion-corrected videos were passed on to the CNMF-E

portion of CaImAn[62]. The main parameters we used for the CNMF-E were as follows: the half size

of a neuron = 8; the half size of the neuron's bounding box = 16; for every session, we chose a

value to separate the first and most significant population of minimum peak-to-noise (PNR) ratio

resulting in use of a range between 7-16; for the minimum correlation value we found values

between 0.75 and 0.85 worked best. Every other setting was left to CaImAn default. After CNMF-

E, we performed a component analysis. We set the Boolean parameter *use_cnn* to true so that

CaImAn would use a trained convolution-neural network (CNN) model to assign a value between

0-1 that determined how close the components matched the appearance of soma. The algorithm

used three different values to determine if a component should be as accepted as a cell: Spatial

footprint consistency (*rval*), Signal-to-Noise ratio (*snr*), and CNN threshold value (*cnn_thr*), with

the minimum values for each of these set to 2.5, 0.85, and 0.85, respectively. We then visually-

inspected random components. Since, according to CaImAn documentation, it is not advised to

calculate deltaF/F when using 1P data (due to the inability to confidently calculate baseline with

background activity) we used the function *estimates.detrend_df_f* with the flag *detrend_only* set

to true to calculate fluorescence where baseline has been subtracted but has not been normalized

by it. Additional settings used were: quantileMin = 8, frames_window = 250, use_residuals = True.

Finally, we calculated the deconvolved traces of the accepted components using the CaImAn

function *estimates.deconvolve*. This removed any change in the fluorescence not due to the

dynamics of the neurons. After the detrended and deconvolved values were calculated, all of the

outputs were saved into a .mat file (MATLAB, MathWorks Inc., Natick, MA), for all further

processing. In MATLAB, we combined the output from CaImAn with TTL pulses to align activity

of neurons around specific epochs of behavior.

Some of the videos had banding artefact caused by a known issue with the USB controller

of the UCLA miniscope DAQ. When banding occurred, we first saved the frames in which this

occurred and then filled the frames with the last reliable frame before the banding. Following

processing with CaImAn, we replaced the data from those frames with NaNs.

Data Analysis

All data analyses were conducted using custom-written code in MATLAB.

Learning and Performance. The probability of choosing the better option was the primary

measure of interest, and secondarily Initiation Latency, Choice Latency, and Reward Latency

were each analyzed with a series of mixed-effects General Linear Models (GLMs) (*fitglme*

function; Statistics and Machine Learning Toolbox) first with omnibus analyses that included lens

group (M2-implanted, OFC-implanted). Follow-up GLMs were conducted pending significant

interactions in the omnibus models. The coding of variables for GLM were: 0=females, 1=males,

0=OFC-implanted, and 1=M2-implanted). Statistical significance for GLM analyses was noted as

p-values of less than 0.05. The variable of interest p(Better) was calculated starting with a boolean

vector where a 1 corresponded to a choice for the better option: 1 with the higher probability of

reward and a 0 for the worse option. This vector of raw choices (0, 1) was then analyzed using a

GLM with a binomial distribution. The Win-Stay and Lose-Shift values presented in **Figure 5D**

and Tables 22-25 were calculated similarly to p(Better), but for the session, not trial-by-trial. A

boolean vector was generated for the entire session: 1 if the trial was Win-Stay / Lose-Shift and

0 if it was not. These boolean vectors were then inputted into *movmean* to generate a proportion

of trials within this window that were Win-Stay or Lose-Shift. The following are definitions of

predictors: *TrialNum*: The number of trials within an entire session (from 1 to 255); *Session*:

Experimental day (from 1-6 in the calcium imaging and 1-10 in the DREADDs experiment); *Block*:

The number of blocks within a single session (from 1-3); *Area*: The brain area for calcium imaging

(1=M2, 0=OFC); *Schedule*: The uncertainty schedule (from 1-3). The remaining predictors are

defined in the Results section.

Cross Validated Peak-Aligned Heatmap. A window of detrended fluorescence 1 s before

and 3 s after choice was obtained for every trial in a session. This resulted in a matrix of cell-by-

frame-by-trial averaged across all trials for a resulting cell-by-frame matrix. This was normalized

using the normalize function on Matlab with the 'range' parameter. The max in a cell's activity was

1, and the minimum was 0 to ensure fair comparison across all cells in that session. The odd trials

of neurons were then peak aligned and the resulting indices were used to plot the even trials

using *imagesc*. Vertical dashed bars show the moment of choice and the moment of reward cue.

Decoder Analysis. A 4-s window of detrended fluorescence centered around Initiation was

obtained for every trial in a session. Separately, another 4-s window (1-s before and 3-s after) of

detrended fluorescence around choice was obtained for every trial in a session. This resulted in

a cell-by-frame-by-trial matrix for initiation and a separate one for choice. We temporally

downsampled so that activity in bin 1 was the average of frames 1-6, activity in bin 2 was the

average of frames 3-9, and so on. Every trial was also labeled as either a left or right choice,

whether rewarded or unrewarded (win or lose). One trial was left out for testing the decoder, and

the rest of the trials were then balanced so that there would be the same number of left-rewarded,

left-unrewarded, right-rewarded, and right-unrewarded conditions across all sessions in training

of the decoder. The balanced trials were randomly selected for every test trial to sample the entire

dataset of trials per session. Once the balanced subset of trials was selected, the neural data for

that session and the labels were inputted into the Matlab function *fitcsvm* with the

"KernelFunction" set to 'linear.' The decoder was then tested on the neural data for the test trial,

and the result was tallied as a 1 if the label matched the test label and a 0 if it was mismatched.

This was conducted for every single trial in a session. Additionally, we reran the decoder on all

trials per session, but with the trial type labels randomly shifted. This data is shown as "Shuffled"

in **Figure 2F-G**. Statistical comparisons and post-hocs were performed using Matlab's *fitglme*

function, and the formulas are included in the corresponding tables. Accuracy measures from the

bin (average of 5 frames) before choice were utilized for the *Chosen Side* GLMs and accuracy

measures before the reward cue were used for the *Trial Outcome* GLM.

604   *Single Cell Selectivity GLM*. Activity for every neuron was obtained on the same bins used

605 for the GLMs for decoder accuracy. In addition to *Chosen Side* and *Trial Outcome* activity, we

606 used the averaged activity 5 frames before and after *Reward Retrieval* for single cell selectivity

607 analysis. Statistics were conducted using Matlab's *fitglme* and the formulas for γ = neural activity

608 were γ ~ [1 + side] for Chosen Side and γ ~ [1 + rewarded + side] for both Trial Outcome and

609 Reward Retrieval. We added side as a covariate to remove the influence of side selective neurons

610 appearing as Trial Outcome or Reward Retrieval selective neurons. After a GLM was fitted for

611 every cell per session, the ratio of selective cells (ones with a p-value lower than 0.05) was

612 obtained. We then fit GLM models to the ratios across both areas and all three schedules. The

613 Beta coefficients used for **Supplementary Figure 3** were optained from the same GLMs used to

614 obtain these ratios. They are also from the same time bins as **Figure 2F-G** and the Reward

615 Retrieval timepoint described above.

616   *Behavioral measurements*. Latencies were recorded from ABET in Lafayette Instrument

617 chambers. Initiation latency measured the time from the presentation of the white circle on the

618 center touch zone until a rat interacts with the middle zone. Choice latency measured the time

619 from the presentation of both stimuli to the time the rat interacts with one of the stimuli to make a

620 choice. Reward latency measures the time from the dispensed pellet until the rat enters the reward

621 port to retrieve the pellet. WinStay was calculated as the proportion of all trials when the rat

622 received a reward on the previous trial and chose the same side for the following trial. LoseShift

623 was calculated as the proportion of all trials when the rat did not receive a reward and chose a

624 different side for the subsequent trial. The Perseveration Index was calculated as the average

625 number of trials to change choice after being unrewarded. Flexibility Index was calculated as

626 follows:

627 $FlexIndex = mean[(mean(C_{Better}\text{Block1}, C_{Better}\text{Block2}), (mean(C_{Better}\text{Block2}, C_{Better}\text{Block3})]$

628 where C_{Better} is equal to 1 if the animal chose the better option and 0 otherwise, with the better

629 option defined according to the reward schedule for each block. Statistical comparisons and post-

630 hocs were performed using Matlab's *fitglme* function, and the formulas are included in the

631 corresponding tables. Accuracy measures from the bin before choice were utilized for the *Chosen*

632 *Side* GLMs and accuracy measures from the bin before the reward cue were used for the *Trial*

633 *Outcome* GLM.

<div style="text-align:center">**REFERENCES**</div>

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771 **TABLE 1**

Across all Blocks in all Sessions for Rats with Lens Implants						
γ = Probability Better Option						
Formula	γ~[1 + Session + Schedule *Block * TrialNum * Area + (1 + Session + Block * TrialNum|RatID)]					
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
Intercept	1.4085	0.6099	2.3094	9433	0.0209	0.2129	2.6040
TrialNum	0.0559	0.0088	6.3735	9433	**<0.0001**	0.0387	0.0731
Schedule	0.1454	0.2598	0.5597	9433	0.5757	-0.3638	0.6546
Session	0.0578	0.0557	1.0376	9433	0.2995	-0.0514	0.1669
Area	1.6671	0.8182	2.0376	9433	**0.0416**	0.0633	3.2709
Block	-2.7860	0.4844	-5.7517	9433	**<0.0001**	-3.7355	-1.8365
TrialNum:Schedule	-0.0133	0.0031	-4.3179	9433	**<0.0001**	-0.0193	-0.0073
TrialNum:Area	0.0226	0.0118	1.9126	9433	0.0558	-0.0006	0.0458
Schedule:Area	-0.4912	0.3328	-1.4760	9433	0.1400	-1.1436	0.1612
TrialNum:Block	-0.0052	0.0031	-1.6777	9433	0.0934	-0.0113	0.0009
Schedule:Block	0.3826	0.2039	1.8761	9433	0.0607	-0.0171	0.7823
Area:Block	-2.0621	0.6598	-3.1253	9433	**0.0018**	-3.3555	-0.7687
TrialNum:Schedule:Area	-0.0078	0.0042	-1.8756	9433	0.0607	-0.0159	0.0004
TrialNum:Schedule:Block	0.0018	0.0011	1.6260	9433	0.1040	-0.0004	0.0039
TrialNum:Area:Block	0.0013	0.0042	0.3071	9433	0.7588	-0.0069	0.0094
Schedule:Area:Block	0.6375	0.2757	2.3122	9433	**0.0208**	0.0970	1.1780
TrialNum:Schedule:Area:Block	-0.0001	0.0015	-0.0730	9433	0.9418	-0.0030	0.0028

772   Alpha Level 0.05.

774 **TABLE 2**

Across all Blocks in all Sessions Comparing Lens Rats

γ = Initiation Latency

Formula	γ~[1+Session*Block*TrialNum+Area+(1+Session*Block*TrialNum|RatID)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	5.7688	1.5593	3.6996	9441	0.0002	2.7123	8.8254
TrialNum	-0.0450	0.0147	-3.0611	9441	**0.0022**	-0.0737	-0.0162
Session	-0.2227	0.2056	-1.0833	9441	0.2787	-0.6257	0.1803
Area	-0.4731	0.3832	-1.2344	9441	0.2171	-1.2242	0.2781
Block	0.0716	0.8186	0.0874	9441	0.9303	-1.5330	1.6762
TrialNum:Session	0.0053	0.0027	2.0071	9441	**0.0448**	0.0001	0.0105
TrialNum:Block	0.0114	0.0048	2.3661	9441	**0.0180**	0.0020	0.0209
Session:Block	-0.1536	0.1565	-0.9820	9441	0.3261	-0.4603	0.1530
TrialNum:Session:Block	-0.0010	0.0008	-1.2089	9441	0.2267	-0.0026	0.0006

775   Alpha Level 0.05

776 **TABLE 3**

Across all Blocks in all Sessions Comparing Lens Rats

γ = Choice Latency

Formula	γ~[1+Session*Block*TrialNum+Area+(1+Session*Block*TrialNum|RatID)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	2.0249	0.6468	3.1304	9441	0.0018	0.7569	3.2928
TrialNum	-0.0085	0.0065	-1.3061	9441	0.1916	-0.0213	0.0043
Session	0.0307	0.2050	0.1498	9441	0.8810	-0.3712	0.4326
Area	-0.5705	0.1600	-3.5666	9441	**0.0004**	-0.8840	-0.2569
Block	0.6944	0.4989	1.3920	9441	0.1640	-0.2835	1.6724
TrialNum:Session	-0.0006	0.0022	-0.2781	9441	0.7809	-0.0048	0.0036
TrialNum:Block	-0.0004	0.0032	-0.1219	9441	0.9030	-0.0066	0.0058
Session:Block	-0.1717	0.1449	-1.1843	9441	0.2363	-0.4558	0.1125
TrialNum:Session:Block	0.0008	0.0007	1.0717	9441	0.2839	-0.0006	0.0022

777   Alpha Level 0.05.

778 **TABLE 4**

Across all Blocks in all Sessions Comparing Lens Rats						
γ = Reward Latency						
Formula	γ~[1+Session*Block*TrialNum+Area+(1+Session*Block*TrialNum|RatID)]					
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	1.5710	0.4575	3.4336	5809	0.0006	0.6740	2.4679
TrialNum	-0.0024	0.0032	-0.7300	5809	0.4654	-0.0087	0.0040
Session	-0.0524	0.0695	-0.7539	5809	0.4509	-0.1887	0.0839
Area	-0.3749	0.2199	-1.7047	5809	0.0883	-0.8060	0.0562
Block	0.2466	0.2591	0.9518	5809	0.3413	-0.2613	0.7545
TrialNum:Session	0.0002	0.0005	0.4166	5809	0.6770	-0.0008	0.0012
TrialNum:Block	-0.0002	0.0017	-0.0989	5809	0.9212	-0.0036	0.0032
Session:Block	-0.0540	0.0441	-1.2253	5809	0.2205	-0.1405	0.0324
TrialNum:Session:Block	0.0001	0.0003	0.2610	5809	0.7941	-0.0005	0.0007

779   Alpha Level 0.05

781 **TABLE 5**

Comparing Chosen Side Decoder Accuracy Across all Schedules in M2 and OFC γ = Decoder Accuracy before Choice							
Formula	γ~[1+Area*Schedule+Session+(1|RatID:Session)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	1.0210	0.0983	10.389	37	<0.0001	0.8219	1.2202
Area	-0.2312	0.0593	-3.9013	37	**0.0004**	-0.3513	-0.1111
Session	0.0268	0.0222	1.2109	37	0.2336	-0.0181	0.0718
Schedule	-0.0843	0.0633	-1.3307	37	0.1914	-0.2126	0.0440
Area:Schedule	0.0562	0.0274	2.0500	37	**0.0475**	0.0007	0.1118

782   Alpha Level 0.05**.**

784 **TABLES 6 (Table 5 Post-Hoc)**

Comparing Chosen Side Decoder Accuracy Across Schedules 1 ONLY in M2 and OFC γ = Decoder Accuracy before Choice in Schedule 1							
Formula	γ~[1+Area+(1|RatID:Session)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.9752	0.0661	14.755	12	<0.0001	0.8312	1.1192
Area	-0.1746	0.0401	-4.3533	12	**0.0009**	-0.2620	-0.0872

Comparing Chosen Side Decoder Accuracy Across Schedules 2 ONLY in M2 and OFC γ = Decoder Accuracy before Choice in Schedule 2							
Formula	γ~[1+Area+(1|RatID:Session)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.9503	0.0747	12.7138	12	<0.0001	0.787	1.113
Area	-0.1195	0.0454	-2.6330	12	**0.0219**	-0.218	-0.021

Comparing Chosen Side Decoder Accuracy Across Schedules 3 ONLY in M2 and OFC γ = Decoder Accuracy before Choice in Schedule 3							
Formula	γ~[1+Area+(1|RatID:Session)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.9140	0.0522	17.5113	12	<0.0001	0.800	1.028
Area	-0.0621	0.0317	-1.9613	12	0.0735	-0.131	0.007

785   Alpha Level 0.05.

788 **TABLE 7**

Comparing Trial Outcome Decoder Accuracy Across all Schedules in M2 and OFC						
γ = **Decoder Accuracy before Reward Cue**						
Formula	γ~[1+Area*Schedule+Session+(1|RatID:Session)]					
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.8561	0.0976	8.7707	37	<0.0001	0.6583	1.0539
Area	-0.1378	0.0589	-2.3405	37	**0.0248**	-0.2571	-0.0185
Session	0.0300	0.0220	1.3612	37	0.1817	-0.0146	0.0746
Schedule	-0.1278	0.0629	-2.0321	37	**0.0494**	-0.2552	-0.0004
Area:Schedule	0.0482	0.0273	1.7686	37	0.0852	-0.0070	0.1034

789   Alpha Level 0.05.

792 **TABLE 8**

Comparing the Ratio of Side Selective Neurons Across all schedules in M2 and OFC							
γ = Ratio of Side Selective Neurons							
Formula	γ~[1+Area*Schedule+Session+(1|RatID)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.5512	0.0824	6.6895	37	<0.0001	0.3843	0.7182
Session	0.0208	0.0179	1.1569	37	0.2547	-0.0156	0.0571
Schedule	-0.0999	0.0512	-1.9501	37	0.0588	-0.2037	0.0039
Area	-0.2375	0.0497	-4.7775	37	**<0.0001**	-0.3383	-0.1368
Area:Schedule	0.0630	0.0222	2.8364	37	**0.0074**	0.0180	0.1079

793 Alpha Level 0.05

795 **TABLE 9 (Table 8 Post-Hoc)**

Comparing the Ratio of Side Selective Neurons Across all Schedules in M2 ONLY							
γ = Ratio of Side Selective Neurons in M2							
Formula	γ~[1+Schedule+(1|RatID)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.3033	0.0407	7.4487	16	<0.0001	0.2170	0.3896
Schedule	0.0046	0.0166	0.2758	16	0.7862	-0.0305	0.0397
Comparing the Ratio of Side Selective Neurons Across all Schedules in OFC ONLY							
γ = Ratio of Side Selective Neurons in OFC							
Formula	γ~[1+Schedule+(1|RatID)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.0658	0.0310	2.1207	22	<0.0001	0.0015	0.1301
Schedule	0.0675	0.0144	4.7031	22	**0.0001**	0.0378	0.0973

796 Alpha Level 0.05.

800 **TABLE 10**

Comparing the Ratio of Outcome Selective Neurons Across all schedules in M2 and OFC							
γ = Ratio of Outcome Selective Neurons							
Formula	γ~[1+Area*Schedule+Session+(1|RatID)]						
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.3351	0.0897	3.7352	37	0.0006	0.1533	0.5168
Session	0.0181	0.0161	1.1285	37	0.2664	-0.0144	0.0507
Schedule	-0.0959	0.0459	-2.0875	37	**0.0438**	-0.1889	-0.0028
Area	-0.1020	0.0542	-1.8805	37	0.0679	-0.2119	0.0079
Area:Schedule	0.0447	0.0199	2.2443	37	**0.0309**	0.0043	0.0850

801   Alpha Level 0.05

803 **TABLE 11 (Table 10 Post-Hoc)**

Comparing the Ratio of Outcome Selective Neurons Across all Schedules in M2 ONLY							
γ = Ratio of Outcome Selective Neurons in M2							
Formula	γ~[1+Schedule+(1|RatID)]						
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.2240	0.0415	5.4034	16	<0.0001	0.1361	0.3119
Schedule	-0.0149	0.0188	-0.7920	16	0.4399	-0.0548	0.0250
Comparing the Ratio of Outcome Selective Neurons Across all Schedules in OFC ONLY							
γ = Ratio of Outcome Selective Neurons in OFC							
Formula	γ~[1+Schedule+(1|RatID)]						
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.1220	0.0358	3.4079	22	0.0025	0.0478	0.1963
Schedule	0.0297	0.0104	2.8584	22	**0.0091**	0.0082	0.0513

804   Alpha Level 0.05.

808 **TABLE 12**

Comparing the Ratio of Reward Selective Neurons Across all schedules in M2 and OFC							
γ = Ratio of Reward Selective Neurons							
Formula	γ~[1+Area*Schedule+Session+(1|RatID)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.7543	0.1819	4.1471	37	0.0002	0.3857	1.1228
Session	0.0080	0.0360	0.2223	37	0.8253	-0.0649	0.0809
Schedule	-0.1124	0.1028	-1.0940	37	0.2810	-0.3207	0.0958
Area	-0.2430	0.1099	-2.2120	37	**0.0332**	-0.4656	-0.0204
Area:Schedule	0.0909	0.0445	2.0408	37	**0.0484**	0.0007	0.1811

809   Alpha Level 0.05

811 **TABLE 13 (Table 12 Post-Hoc)**

Comparing the Ratio of Reward Selective Neurons Across all Schedules in M2 ONLY							
γ = Ratio of Reward Selective Neurons in M2							
Formula	γ~[1+Schedule+(1|RatID)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.5073	0.0390	12.9905	16	<0.0001	0.4245	0.5900
Schedule	-0.0056	0.0133	-0.4179	16	0.6816	-0.0337	0.0226
Comparing the Ratio of Reward Selective Neurons Across all Schedules in OFC ONLY							
γ = Ratio of Reward Selective Neurons in OFC							
Formula	γ~[1+Schedule+(1|RatID)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.2643	0.0906	2.9182	22	0.0080	0.0765	0.4521
Schedule	0.0853	0.0373	2.2886	22	**0.0321**	0.0080	0.1626

812   Alpha Level 0.05

814 **TABLE 14**

Decoder Accuracy modeled by Behavioral Measurements. γ = Chosen Side Decoder Accuracy							
Formula	γ~[1+Session+FlexibilityIndex+PerserverationIndex+InitiationLatency+Choice Latency+RewardLatency+WinStay+LoseShift+Area*(Session+FlexibilityIndex +PerserverationIndex+InitiationLatency+ChoiceLatency+RewardLatency+Win Stay+LoseShift)+(1|RatID:Session)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	1.2994	0.4269	3.0436	25	0.0054	0.4201	2.1787
Initiation Latency	-0.0739	0.0653	-1.1326	25	0.2681	-0.2083	0.0605
Choice Latency	0.0264	0.1127	0.2344	25	0.8166	-0.2057	0.2586
Reward Latency	-0.0873	0.0783	-1.1152	25	0.2754	-0.2484	0.0739
WinStay	-1.5011	0.8803	-1.7052	25	0.1006	-3.3140	0.3119
Area	-0.5611	0.2587	-2.1691	25	**0.0398**	-1.0939	-0.0283
LoseShift	-2.2113	1.4948	-1.4793	25	0.1515	-5.2899	0.8673
Flexibility Index	1.1537	0.8096	1.4250	25	0.1665	-0.5137	2.8211
Perseveration Index	3.9990	1.6714	2.3926	25	**0.0246**	0.5566	7.4413
Session	0.0035	0.0099	0.3565	25	0.7244	-0.0168	0.0238
Area:InitiationLatency	0.0608	0.0407	1.4962	25	0.1471	-0.0229	0.1446
Area:ChoiceLatency	-0.0599	0.0841	-0.7127	25	0.4826	-0.2331	0.1132
Area:RewardLatency	0.0242	0.0438	0.5542	25	0.5844	-0.0659	0.1144
Area:Win Stay	1.3807	0.5670	2.4352	25	**0.0224**	0.2130	2.5485
Area:Lose Shift	2.1181	0.9107	2.3258	25	**0.0284**	0.2425	3.9937
Area:Flexibility Index	-0.7755	0.5260	-1.4743	25	0.1529	-1.8589	0.3078
Area:Perseveration Index	-3.9170	1.0290	-3.8066	25	**0.0008**	-6.0363	-1.7978

815   Alpha Level 0.05

824 **TABLE 15 (Table 14 Post-Hoc)**

Post-Hoc Statistics on Area Interaction with M2 Lens Rats only							
γ = **Chosen Side Decoder Accuracy in M2 Rats**							
Formula	γ~[1+PerseverationIndex+WinStay+LoseShift+(1|RatID:Session)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.9846	0.1674	5.8818	14	<0.0001	0.6255	1.3436
Win Stay	-0.0131	0.1558	-0.0839	14	0.9343	-0.3472	0.3210
Lose Shift	-0.6778	0.5746	-1.1796	14	0.2578	-1.9102	0.5546
Perseveration Index	-1.0741	0.7841	-1.3699	14	0.1923	-2.7559	0.6076

Post-Hoc Statistics on Area Interaction with OFC Lens Rats only							
γ = **Chosen Side Decoder Accuracy in OFC Rats**							
Formula	γ~[1+PerseverationIndex+WinStay+LoseShift+(1|RatID:Session)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.2041	0.1828	1.1167	20	0.2774	-0.1771	0.5853
Win Stay	0.8435	0.2010	4.1975	20	**0.0004**	0.4243	1.2627
Lose Shift	1.4664	0.5304	2.7648	20	**0.0120**	0.3600	2.5729
Perseveration Index	-4.1286	0.6312	-6.5405	20	**<0.0001**	-5.4453	-2.8118

825   Alpha Level 0.05

827 **TABLE 16**

Decoder Accuracy modeled by Behavioral Measurements.						
γ = **Trial Outcome Decoder Accuracy**						

Formula	γ~[1+Session+FlexibilityIndex+PerserverationIndex+InitiationLatency+ChoiceLatency+RewardLatency+WinStay+LoseShift+Area*(Session+FlexibilityIndex+PerserverationIndex+InitiationLatency+ChoiceLatency+RewardLatency+WinStay+LoseShift)+(1|RatID:Session)]						
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
Intercept	1.4433	0.4581	3.1505	25	0.0042	0.4998	2.3868
Initiation Latency	-0.1490	0.0700	-2.1278	25	**0.0434**	-0.2932	-0.0048
Choice Latency	0.0677	0.1210	0.5595	25	0.5808	-0.1814	0.3168
Reward Latency	0.1892	0.0840	2.2530	25	**0.0333**	0.0162	0.3621
WinStay	-2.0423	0.9446	-2.1622	25	**0.0404**	-3.9877	-0.0970
Area	-0.7024	0.2776	-2.5304	25	**0.0181**	-1.2741	-0.1307
LoseShift	-2.7002	1.6040	-1.6835	25	0.1047	-6.0037	0.6032
Flexibility Index	0.8666	0.8687	0.9976	25	0.3281	-0.9226	2.6557
Perseveration Index	3.7763	1.7935	2.1056	25	**0.0455**	0.0826	7.4700
Session	0.0087	0.0106	0.8186	25	0.4207	-0.0131	0.0305
Area:InitiationLatency	0.0946	0.0436	2.1676	25	**0.0399**	0.0047	0.1845
Area:ChoiceLatency	-0.0641	0.0902	-0.7109	25	0.4837	-0.2499	0.1217
Area:RewardLatency	-0.0971	0.0469	-2.0688	25	**0.0491**	-0.1938	-0.0004
Area:WinStay	1.5174	0.6084	2.4940	25	**0.0196**	0.2643	2.7704
Area:LoseShift	2.3028	0.9772	2.3565	25	**0.0266**	0.2902	4.3154
Area:Flexibility Index	-0.6076	0.5644	-1.0765	25	0.2920	-1.7701	0.5549
Area:Perseveration Index	-2.4325	1.1041	-2.2031	25	**0.0370**	-4.7065	-0.1585

828   Alpha Level 0.05

 TABLE 17 (Table 16 Post-Hoc)

Post-Hoc Statistics on Area Interaction with M2 Lens Rats only						
γ = **Trial Outcome Decoder Accuracy in M2 Rats**						
Formula	γ~[1+PerseverationIndex+InitiationLatency+RewardLatency+Win Stay+LoseShift+(1|RatID:Session)]					
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.8398	0.1998	4.2042	12	0.0012	0.4046	1.2750
Initiation Latency	-0.0678	0.0281	-2.4183	12	**0.0324**	-0.1290	-0.0067
Reward Latency	0.0835	0.0398	2.0961	12	0.0579	-0.0033	0.1703
Win Stay	-0.2921	0.2255	-1.2951	12	0.2197	-0.7834	0.1993
Lose Shift	-0.1878	0.6896	-0.2723	12	0.7901	-1.6903	1.3148
Perseveration Index	0.9463	0.8409	1.1254	12	0.2824	-0.8858	2.7785

Post-Hoc Statistics on Area Interaction with OFC Lens Rats only						
γ = **Trial Outcome Decoder Accuracy in OFC Rats**						
Formula	γ~[1+PerseverationIndex+InitiationLatency+RewardLatency+Win Stay+LoseShift+(1|RatID:Session)]					
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
(Intercept)	0.0749	0.1868	0.4007	18	0.6934	-0.3176	0.4674
Initiation Latency	0.0193	0.0151	1.2789	18	0.2172	-0.0124	0.0509
Reward Latency	-0.0069	0.0236	-0.2919	18	0.7737	-0.0564	0.0426
Win Stay	0.6517	0.2268	2.8730	18	**0.0101**	0.1751	1.1283
Lose Shift	1.5504	0.5505	2.8161	18	**0.0114**	0.3937	2.7070
Perseveration Index	-1.5430	0.6310	-2.4452	18	**0.0249**	-2.8687	-0.2172

838   Alpha Level 0.05

 TABLE 18

Across all Blocks in all Sessions for Rats with DREADDs in OFC γ = **Probability Better Option**							
Formula	γ ~ [1 + Drug*Schedule*Order + TrialNum*Block + Sex + (1 + TrialNum*Block | RatID)]						
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
Intercept	3.4514	0.3179	10.8558	20238	<0.0001	2.8282	4.0746
TrialNum	0.0364	0.0031	11.8630	20238	**<0.0001**	0.0303	0.0424
Schedule	-0.1897	0.0377	-5.0383	20238	**<0.0001**	-0.2635	-0.1159
Order	1.2519	0.1697	7.3779	20238	**<0.0001**	0.9193	1.5845
Sex	-0.0729	0.0726	-1.0052	20238	0.3148	-0.2152	0.0693
Drug	-0.2831	0.1204	2.3502	20238	**0.0188**	0.0470	0.5192
Block	-3.0267	0.2796	-10.823	20238	**<0.0001**	-3.5748	-2.4786
Schedule:Order	-0.4388	0.0967	-4.5357	20238	**<0.0001**	-0.6284	-0.2492
Schedule:Drug	-0.0222	0.0545	-0.4071	20238	0.6839	-0.1291	0.0847
Order:Drug	-0.3706	0.2443	-1.5169	20238	0.1293	-0.8495	0.1083
TrialNum:Block	0.0002	0.0009	0.1891	20238	0.8500	-0.0015	0.0019
Schedule:Order :Drug	0.1861	0.1397	1.3321	20238	0.1829	-0.0877	0.4599

841   Alpha Level 0.05

849 **TABLE 19**

<table>
<tr><td colspan="8" align="center">Across all Blocks in all Sessions for Rats with DREADDs in M2
γ = **Probability Better Option**</td></tr>
<tr><td>Formula</td><td colspan="7">γ ~ [1 + Drug*Schedule*Order + TrialNum*Block + Sex +
(1 + TrialNum*Block | RatID)]</td></tr>
<tr><td>Coefficients</td><td>β</td><td>SE</td><td>tStat</td><td>DF</td><td>P</td><td>CIL</td><td>CIU</td></tr>
<tr><td>Intercept</td><td>2.2371</td><td>0.3472</td><td>6.4436</td><td>20238</td><td><0.0001</td><td>1.5566</td><td>2.9175</td></tr>
<tr><td>**TrialNum**</td><td>0.0200</td><td>0.0047</td><td>4.2983</td><td>20238</td><td>**<0.0001**</td><td>0.0109</td><td>0.0292</td></tr>
<tr><td>**Schedule**</td><td>0.0702</td><td>0.0346</td><td>2.0301</td><td>20238</td><td>**0.0424**</td><td>0.0024</td><td>0.1381</td></tr>
<tr><td>**Order**</td><td>0.7681</td><td>0.1371</td><td>5.6007</td><td>20238</td><td>**0.0000**</td><td>0.4993</td><td>1.0368</td></tr>
<tr><td>**Sex**</td><td>0.2504</td><td>0.1193</td><td>2.0995</td><td>20238</td><td>**0.0358**</td><td>0.0166</td><td>0.4842</td></tr>
<tr><td>**Drug**</td><td>-0.4698</td><td>0.1066</td><td>4.4067</td><td>20238</td><td>**<0.0001**</td><td>0.2609</td><td>0.6788</td></tr>
<tr><td>**Block**</td><td>-2.4059</td><td>0.2808</td><td>-8.5687</td><td>20238</td><td>**<0.0001**</td><td>-2.9563</td><td>-1.8556</td></tr>
<tr><td>**Schedule:Order**</td><td>-0.3030</td><td>0.0799</td><td>-3.7922</td><td>20238</td><td>**0.0001**</td><td>-0.4596</td><td>-0.1464</td></tr>
<tr><td>**Schedule:Drug**</td><td>-0.1866</td><td>0.0492</td><td>-3.7900</td><td>20238</td><td>**0.0002**</td><td>-0.2832</td><td>-0.0901</td></tr>
<tr><td>**Order:Drug**</td><td>-0.4036</td><td>0.1956</td><td>-2.0637</td><td>20238</td><td>**0.0391**</td><td>-0.7869</td><td>-0.0203</td></tr>
<tr><td>**TrialNum:Block**</td><td>0.0027</td><td>0.0012</td><td>2.2015</td><td>20238</td><td>**0.0277**</td><td>0.0003</td><td>0.0052</td></tr>
<tr><td>Schedule:Order :Drug</td><td>0.1914</td><td>0.1137</td><td>1.6829</td><td>20238</td><td>0.0924</td><td>-0.0315</td><td>0.4142</td></tr>
</table>

850 Alpha Level 0.05

859 **TABLE 20 (Table 19 Post-Hoc)**

Post-Hoc Statistics on Schedule by Drug Interaction with Schedule 1 collapsed, M2 Rats γ = **Probability Better Option**							
Formula	γ ~ [1+Drug+Sex+Block*TrialNum+(1+Block*TrialNum|Rat)]						
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
(Intercept)	2.3493	0.3596	6.5333	8094	<0.0001	1.6444	3.0542
TrialNum	0.0212	0.0050	4.2491	8094	**<0.0001**	0.0114	0.0310
Sex	0.4271	0.1365	3.1299	8094	**0.0018**	0.1596	0.6947
Drug	-0.2018	0.0505	3.9978	8094	**0.0001**	0.1028	0.3007
Block	-2.4932	0.2741	-9.0943	8094	**<0.0001**	-3.0306	-1.9558
TrialNum:Block	0.0030	0.0015	1.9742	8094	**0.0484**	0.0000	0.0061

Post-Hoc Statistics on Schedule by Drug Interaction with Schedule 2 collapsed, M2 Rats γ = **Probability Better Option**							
Formula	γ ~ [1+Drug+Sex+Block*TrialNum+(1+Block*TrialNum|Rat)]						
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
(Intercept)	2.4211	0.3567	6.7876	8094	<0.0001	1.7219	3.1203
TrialNum	0.0264	0.0064	4.1355	8094	**<0.0001**	0.0139	0.0390
Sex	0.2689	0.1034	2.6021	8094	**0.0093**	0.0663	0.4715
Drug	-0.0476	0.0507	0.9380	8094	0.3483	-0.0518	0.1470
Block	-2.4086	0.3349	-7.1911	8094	**<0.0001**	-3.0651	-1.7520
TrialNum:Block	0.0008	0.0016	0.4725	8094	0.6366	-0.0024	0.0040

Post-Hoc Statistics on Schedule by Drug Interaction with Schedule 3 collapsed, M2 Rats γ = **Probability Better Option**							
Formula	γ ~ [1+Drug+Sex+Block*TrialNum+(1+Block*TrialNum|Rat)]						
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
(Intercept)	2.6601	0.5251	5.0659	4044	<0.0001	1.6306	3.6896
TrialNum	0.0071	0.0056	1.2606	4044	0.2075	-0.0039	0.0180
Sex	0.4522	0.1762	2.5666	4044	**0.0103**	0.1068	0.7975
Drug	0.0491	0.0684	-0.7179	4044	0.4728	-0.1832	0.0850
Block	-2.3052	0.3974	-5.8000	4044	**<0.0001**	-3.0844	-1.5260
TrialNum:Block	0.0057	0.0021	2.7799	4044	**0.0055**	0.0017	0.0098

860   Alpha Level 0.05

863 **TABLE 21 (Table 19 Post-Hoc)**

Post-Hoc Statistics on Order by Drug Interaction with Ascending collapsed, M2 Rats
γ = **Probability Better Option**

Formula	γ ~ [1+Drug+Sex+Block*TrialNum+(1+Block*TrialNum|Rat)]						
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
(Intercept)	2.6389	0.3007	8.7758	12144	<0.0001	2.0495	3.2284
TrialNum	0.0180	0.0049	3.6885	12144	**0.0002**	0.0084	0.0276
Sex	0.0654	0.1015	0.6447	12144	0.5192	-0.1335	0.2644
Drug	-0.0950	0.0400	2.3744	12144	**0.0176**	0.0166	0.1735
Block	-2.5241	0.2593	-9.7345	12144	**<0.0001**	-3.0324	-2.0158
TrialNum:Block	0.0037	0.0013	2.7929	12144	**0.0052**	0.0011	0.0063

Post-Hoc Statistics on Order by Drug Interaction with Descending collapsed, M2 Rats
γ = **Probability Better Option**

Formula	γ ~ [1+Drug+Sex+Block*TrialNum+(1+Block*TrialNum|Rat)]						
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
(Intercept)	2.2890	0.4609	4.9665	8094	<0.0001	1.3856	3.1925
TrialNum	0.0243	0.0051	4.7432	8094	**<0.0001**	0.0143	0.0344
Sex	0.5643	0.1814	3.1111	8094	**0.0019**	0.2088	0.9199
Drug	-0.0757	0.0520	1.4559	8094	0.1455	-0.0262	0.1777
Block	-2.2855	0.3692	-6.1910	8094	**<0.0001**	-3.0091	-1.5618
TrialNum:Block	0.0012	0.0014	0.8352	8094	0.4036	-0.0016	0.0039

864   Alpha Level: 0.05

866 **TABLE 22**

<table>
<tr><td colspan="8" align="center">Across all Sessions for Rats with DREADDs in OFC
γ = **Win-Stay**</td></tr>
<tr><td>Formula</td><td colspan="7">γ ~ [1 + Drug*Schedule*Order + Sex + (1 | Rat)](1 | RatID)]</td></tr>
<tr><td>Coefficients</td><td>β</td><td>SE</td><td>tStat</td><td>DF</td><td>P</td><td>CIL</td><td>CIU</td></tr>
<tr><td>Intercept</td><td>0.6894</td><td>0.0293</td><td>23.5256</td><td>81</td><td><0.0001</td><td>0.6311</td><td>0.7477</td></tr>
<tr><td>**Schedule**</td><td>-0.0548</td><td>0.0112</td><td>-4.8942</td><td>81</td><td>**<0.0001**</td><td>-0.0771</td><td>-0.0325</td></tr>
<tr><td>**Order**</td><td>0.2558</td><td>0.0429</td><td>5.9638</td><td>81</td><td>**<0.0001**</td><td>0.1705</td><td>0.3411</td></tr>
<tr><td>Sex</td><td>-0.0329</td><td>0.0232</td><td>-1.4209</td><td>81</td><td>0.1592</td><td>-0.0791</td><td>0.0132</td></tr>
<tr><td>**Drug**</td><td>-0.0782</td><td>0.0342</td><td>2.2852</td><td>81</td><td>**0.0249**</td><td>0.0101</td><td>0.1463</td></tr>
<tr><td>**Schedule:Order**</td><td>-0.0923</td><td>0.0250</td><td>-3.6874</td><td>81</td><td>**0.0004**</td><td>-0.1422</td><td>-0.0425</td></tr>
<tr><td>Schedule:Drug</td><td>-0.0138</td><td>0.0158</td><td>-0.8730</td><td>81</td><td>0.3853</td><td>-0.0453</td><td>0.0177</td></tr>
<tr><td>Order:Drug</td><td>-0.0826</td><td>0.0607</td><td>-1.3623</td><td>81</td><td>0.1769</td><td>-0.2033</td><td>0.0381</td></tr>
<tr><td>Schedule:Order:Drug</td><td>0.0341</td><td>0.0354</td><td>0.9621</td><td>81</td><td>0.3389</td><td>-0.0364</td><td>0.1045</td></tr>
</table>

867 Alpha Level 0.05

869 **TABLE 23**

Across all Sessions for Rats with DREADDs in M2 γ = **Win-Stay**							
Formula	γ ~ [1 + Drug*Schedule*Order + Sex + (1 | Rat)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
Intercept	0.5246	0.0557	9.4118	81	<0.0001	0.4137	0.6355
Schedule	-0.0015	0.0190	-0.0781	81	0.9379	-0.0392	0.0363
Order	0.1949	0.0726	2.6828	81	**0.0088**	0.0504	0.3394
Sex	-0.1342	0.0520	-2.5838	81	**0.0116**	-0.2376	-0.0309
Drug	-0.0863	0.0580	1.4884	81	0.1405	-0.0291	0.2016
Schedule:Order	-0.0790	0.0424	-1.8627	81	0.0661	-0.1634	0.0054
Schedule:Drug	-0.0328	0.0268	-1.2241	81	0.2245	-0.0862	0.0205
Order:Drug	-0.0443	0.1027	-0.4310	81	0.6676	-0.2487	0.1601
Schedule:Order:Drug	0.0185	0.0600	0.3087	81	0.7583	-0.1008	0.1379

870   Alpha Level 0.05

879 **TABLE 24**

	Across all Sessions for Rats with DREADDs in OFC γ = **Lose-Shift**						
Formula	γ ~ [1 + Drug*Schedule*Order + Sex + (1 | Rat)]						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
Intercept	0.1143	0.0161	7.0819	81	<0.0001	0.0822	0.1464
Schedule	0.0272	0.0056	4.8427	81	**<0.0001**	0.0160	0.0383
Order	-0.0825	0.0215	-3.8395	81	**0.0002**	-0.1252	-0.0397
Sex	0.0014	0.0147	0.0968	81	0.9231	-0.0278	0.0306
Drug	0.0184	0.0171	-1.0760	81	0.2851	-0.0525	0.0157
Schedule:Order	0.0291	0.0125	2.3232	81	**0.0227**	0.0042	0.0541
Schedule:Drug	-0.0010	0.0079	-0.1245	81	0.9012	-0.0168	0.0148
Order:Drug	0.0189	0.0304	0.6232	81	0.5349	-0.0415	0.0794
Schedule:Order:Drug	-0.0074	0.0177	-0.4177	81	0.6773	-0.0427	0.0279

880   Alpha Level 0.05

882 **TABLE 25**

Across all Sessions for Rats with DREADDs in M2 γ = **Lose-Shift**						
Formula	$\gamma \sim [\ 1 + \text{Drug*Schedule*Order} + \text{Sex} + (1\ |\ \text{Rat})\ ]$					
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
Intercept	0.1584	0.0225	7.0291	81	<0.0001	0.1135	0.2032
Schedule	0.0057	0.0080	0.7105	81	0.4794	-0.0102	0.0216
Order	-0.0514	0.0306	-1.6777	81	0.0973	-0.1123	0.0095
Sex	0.0349	0.0200	1.7466	81	0.0845	-0.0049	0.0747
Drug	-0.0026	0.0244	-0.1079	81	0.9144	-0.0512	0.0460
Schedule:Order	0.0225	0.0179	1.2571	81	0.2123	-0.0131	0.0580
Schedule:Drug	-0.0025	0.0113	-0.2184	81	0.8276	-0.0250	0.0200
Order:Drug	-0.0122	0.0433	-0.2814	81	0.7791	-0.0983	0.0740
Schedule:Order:Drug	-0.0514	0.0306	-1.6777	81	0.0973	-0.1123	0.0095

883   Alpha Level 0.05

887 **TABLE 26**

	Across all Sessions for Rats with DREADDs in M2 γ = **Choice Latency**						
Formula	$\gamma \sim [\ 1 + Drug*Schedule*Order + Sex + (1\ |\ Rat)\ ]$						
Coefficients	**β**	SE	tStat	DF	P	CIL	CIU
Intercept	0.5028	0.2232	2.2530	81	0.0270	0.0588	0.9468
Schedule	0.1222	0.0970	1.2599	81	0.2113	-0.0708	0.3151
Order	0.0091	0.3713	0.0246	81	0.9804	-0.7297	0.7480
Sex	0.3677	0.1185	3.1020	81	**0.0026**	0.1319	0.6036
Drug	-0.1114	0.2962	-0.3760	81	0.7079	-0.7008	0.4780
Schedule:Order	0.0183	0.2168	0.0843	81	0.9330	-0.4131	0.4497
Schedule:Drug	0.0592	0.1371	0.4315	81	0.6673	-0.2137	0.3320
Order:Drug	0.1751	0.5251	0.3335	81	0.7396	-0.8697	1.2200
Schedule:Order:Drug	-0.0578	0.3066	-0.1886	81	0.8509	-0.6679	0.5522

888   Alpha Level 0.05

891 **TABLE 27**

	Across all Sessions for Rats with DREADDs in OFC γ = **Choice Latency**						
Formula	$\gamma \sim [\ 1 + Drug*Schedule*Order + Sex + (1\ |\ Rat)\ ]$						
Coefficients	β	SE	tStat	DF	P	CIL	CIU
Intercept	1.2705	0.2036	6.2401	81	<0.0001	0.8654	1.6756
Schedule	-0.0897	0.0616	-1.4576	81	0.1488	-0.2122	0.0327
Order	-0.6209	0.2357	-2.6341	81	**0.0101**	-1.0900	-0.1519
Sex	0.0135	0.2101	0.0643	81	0.9489	-0.4046	0.4316
Drug	-0.3819	0.1880	-2.0306	81	**0.0456**	-0.7560	-0.0077
Schedule:Order	0.1862	0.1376	1.3526	81	0.1799	-0.0877	0.4600
Schedule:Drug	0.1301	0.0870	1.4940	81	0.1390	-0.0431	0.3033
Order:Drug	0.4106	0.3334	1.2317	81	0.2216	-0.2527	1.0739
Schedule:Order:Drug	-0.2023	0.1946	-1.0392	81	0.3018	-0.5896	0.1850

892   Alpha Level 0.05