Female Moths Incorporate Plant Acoustic Emissions into Their Oviposition Decision-Making Process

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Abstract

Insects rely on plants' visual, chemical, tactile, and electrical cues when making various decisions. Recently it has been found that plants emit ultrasonic sounds, which are in the hearing range of many moths, especially under dehydration stress. In this study, we sought to determine whether insects also rely on plant acoustic signals when making decisions. We investigated whether female moths rely on ultrasonic clicks which are typically produced by dehydrated plants when deciding where to oviposit. In the absence of an actual plant, the moths indeed preferred to lay their eggs in proximity to acoustic signals which represent dehydrating plants. Tracking the moths' behavior prior to the decision showed that they examined both sides of the arena and gradually spent more time on the acoustic-playback side. Interestingly, when actual plants were added to the arena, the oviposition preference was reversed and the moths preferred silent plants, which is in accordance with their a-priori preference for hydrated plants. Deafening the moths eliminated their preference, confirming that the choice was based on hearing. Moreover, male moth signals did not affect female oviposition decision, suggesting that the response was specific to plant sound emissions. We reveal evidence for a first acoustic interaction between moths and plants, but as plants emit various sounds, our findings hint to the existence of more currently unknown insect-plant acoustic interactions.

Keywords

plant-insect interactions, ultrasound-hearing moths, plant dehydration ultrasonic clicks, plant bioacoustics.

Introduction

Plant-insect communication has been shown to rely on various modalities, including vision, olfaction, and mechanoreception (Boppré 1978; Kevan and Lane 1985; Gori 1989; Ne'eman 1995; Schiestl 2010; Brito *et al.* 2015; van Dam and Bouwmeester 2016). Plant-insect (airborne) acoustic communication, however, has never been demonstrated. It has long been known that plants vibrate at ultrasonic frequencies due to physiological processes such as cavitation, resulting from changes in their water pressure (Milburn and Johnson 1966; Tyree and Dixon 1983; Ponomarenko *et al.* 2014). Recently it has also been shown that these ultrasonic sounds produced by a drought-stressed or cut plant are airborne and are probably loud enough to be detected by ultrasound-hearing moths from a distance of a few meters (Khait *et al.* 2023). Moreover, it was shown that these sounds can serve as reliable cues for the condition of the plant, specifically indicating whether a plant is drought-stressed.

Ultrasonic hearing abilities and hearing organs located on different body parts have evolved multiple times independently in the Lepidoptera. Hearing sensitivity typically falls within the 20-60 kHz range in all groups of moths that have evolved ultrasonic hearing (Fenton and Fullard 1979; Hoy 1996; Conner 1999; Robert and Göpfert 2002; Moir *et al.* 2013; Göpfert and Hennig 2016). Two main hypotheses exist regarding the evolution of these hearing organs. The first suggests that they have evolved for sexual communication, i.e., to detect ultrasonic signals produced by male moths (Nakano *et al.* 2009). The second hypothesis suggests that they have evolved as an anti-predator mechanism to detect echolocation calls produced by bats (Conner 1999; Greenfield and Weber 2000; Nakano *et al.* 2014; but see Kawahara *et al.* 2019). Regardless of why it has evolved, ultrasonic hearing allows moths to detect various additional sounds (Spangler 1988), including plant dehydration sound clicks which have a wide spectrum that overlaps with moths' hearing range and peaks around 50kHz (Khait et al. 2023). We thus hypothesized that herbivor female moths with ultrasonic hearing might exploit ultrasonic plant emissions as cues to infer plant condition and employ this information for oviposition.

The selection of an oviposition site has a significant impact on the fitness of the hatching herbivor larvae and is thus one of the most critical decisions in the life of a female moth (Lhomme *et al.* 2018). Here we focused on the Egyptian cotton leafworm (*Spodoptera littoralis*) from the Noctuidae family, which is a generalist herbivore with ultrasonic-sensitive tympanic ears (Tougaard 1996, Skals *et al.* 2005, Anto *et al.* 2011).

The ears' sensitivity of many moths from the Noctuidae family have been fully
characterized and they typically show a wide range of sensitivy between ~20 - ~60 kHz
(Fullard 1998). The full audiogram of the Egyptian cotton leafworm moth has not been
documented, but (in accordance with the moths in the Noctuidae family) its hearing has
been shown to be most sensitive around 38kHz, a frequency which is part of the plant's
click spectrum (Tougaard 1998). Moreover, the spectra of the clicks of the males of this
species (Fig.1) which are clearly heard by the females broadly overlap with plant clicks.
We further demonstrated that the moth can hear echolocation calls which are in the
range between 40-80kHz, thus demonstrating sensitivity in the plant clicking range (see
Methods).

Much research has been conducted to characterize the females' oviposition choice in
this species with many factors suggested to be important for their decision-making
process. The females have been found to prefer certain species of host plants over others
(Salama *et al*. 1971; Sadek *et al*. 2010), to select plants based on their larval experience
(Proffit *et al.* 2015), and to choose plants devoid of parasitic larvae, possibly because
the presence of such larvae could promote the recruitment of secondary predators
(Sadek et al. 2010).

In this study, we investigated whether ultrasonic sounds typical of drought-stressed
plants influence oviposition decision making in the Egyptian cotton leafworm moths.
Based on their general behavioral preference for non-dry plants (as we validated, see
below), we hypothesized that the female moths would be affected by plant ultrasonic
signals when making oviposition decisions. Our results support this hypothesis,
providing the first evidence for the use of typical plant sounds by insects.

Results

In each of the following experiments, we placed between 10±3 (Mean±SD) fertile
female *S. littoralis* moths in the center of a $100 \times 50 \times 50$ cm^3 arena divided in the middle,
with two choices offered, one on either side of the arena (a two-alternative forced choice
paradigm, see Methods). To assess their choice, we compared the number of egg
clusters which the moths had laid on each side.

Each treatment was repeated at least 9 times (i.e., with a new set of moths) but the
moths in each repetition were observed for several consecutive nights so that the
minimum number of egg-laying events was 17. Each night was considered an
independent observation because the moth could make a new decision regarding where

to lay her eggs. The treatment and the control sides were alternated between repetitions.

To ensure replicability, the main plant-acoustic treatments were run twice with a pause

of several months in between (see Table 1 in the Methods). In these experiments, we

used the number of egg clusters, rather than the total number of eggs, as the response

variable because each cluster represents a distinct oviposition decision. However, we

include a control experiment below where we evaluated the effect of the plant sounds

on egg number (and not cluster number).

To examine whether *S. littoralis* females prefer to lay their eggs on drying or fresh

tomato plants (without any playback sound, see Exp. 1 in the Methods), we first placed

them in an arena with one drying and one fresh plant. Female *S. littoralis* demonstrated

a strong preference to lay their eggs on fresh plants that were not drought-stressed (Fig.

1A, 2.2±2.7 vs. 0.9±1.1 egg clusters; Mean ± SE; clusters per night respectively, *p*

=0.004, Mixed effect generalized linear models – GLMM with the number of egg

clusters as the explained parameter, the treatment as a fixed effect and the number of

the arena and the repetition round and night as random effects, see statistics).

We next examined whether an ultrasonic acoustic stimulus affects moths' oviposition

decision making. To this end, we played drought-stressed sounds (recorded from a real

drying tomato plant) on one side of the arena and either placed nothing on the other

side or placed a decoy silent resistor to control for electric field sensing (Fig. 1A).see

Exp. 1 in the Methods). Because we aimed to examine the effect of sound only (without

other sensory cues such as visual or olfactory), in this condition, there was no plant in

the arena, and we placed a small mesh box wrapped with a paper towel in the center of

each side to encourage oviposition (the speaker was under the mesh so that the moth

could not sense the vibration directly, only through airborne sounds waves). Female

moths significantly preferred to lay their eggs on the side of the arena in which drying

plant sounds were played. The opposite choice to what was seen in the initial

experiment (Fig. 1B).

Notably, this experiment was repeated twice - six months apart - and the preference was

significant both times (Fig.1B1 1.1±0.8 vs 0.4±0.7 egg clusters per night for the

playback and the silent side respectively, Mean±SE *p*=0.0004, estimate=1*,* GLMM as

above, see Table 1 for the results of each session). The average number of egg clusters

(1.1 clusters per-night) in this condition was lower than in the baseline condition with

a plant (2.2 clusters), but this is reasonable when taking into account that there was no

plant in the arena. The playback rate was high with 60 drought clicks played per minute.
This is higher than the rate reported for a single young plant, but it is feasible when
considering a patch of adult plants as we have demonstrated experimentally (see
Methods). We repeated this experiment with a lower playback rate (of 30 per minute)
and got the same result – see below.

To make sure that the acoustic signals were the sole influential factor in the moths'
decision-making process, we deafened mated female moths and repeated the
experiment (drought-stressed sounds- no plant in the arena). We placed 9.3±1.8 female
moths in an arena and monitored their choice of oviposition sites. In accordance with
the acoustic hypothesis, the deafened moths did not show any preference in egg laying
(Fig.1B2, 0.70 ± 0.70 vs. 1.0 ± 1.09 egg clusters per night, $p = 0.55$, estimate = 0.12,
GLMM).

To examine the importance of plant sounds in oviposition decision making under
pseudo natural conditions, we placed two hydrated tomato plants - one on either side of
the arena - and added a speaker playing drought plant sounds on one side and on the
other side either a resistor (with the same impedance as the speaker) to control for
potential effects of the electric field, or nothing. Interestingly, females showed a
significant preference for the silent plant. In this case, the female preference was similar
to the initial experiment (without playback) in which the females avoided dehydrated
plants. The females laid 1.8±1.6 vs. 1.1±1.0 egg clusters per night on the silent and
playback sides, respectively. This treatment was also repeated twice over a 12-month
period (Fig. 1B3, estimate = -0.52, p=0.01, GLMM as above, see Table 1 for the results
of each repetition, note that the second repeat was only marginally significant).

To assess whether the moths' response was specific to plant sounds, we conducted
additional tests using male moth that were placed on one side of the arena (in a mesh-
box so females could not interact with males) and produced courtship clicks with a
spectral range similar to tomato clicks (as we validated, Methods). Females showed no
significant preference to lay their eggs near male moths (see Supplementary Fig. 2, Fig.
1C, p = 0.4, estimate = -0.25, GLMM as above). The egg-laying rate remained
consistent, with approximately one cluster per night on both sides of the arena.

Fig.1: **The setup and results.** In all experimental setup illustrations (A-C), the control is presented on
the left side, and the various treatments are on the right side. Because the number of egg clusters was low
(between 0-5 clusters) we present the Bayesian posterior (see Methods) for the probability to lay a cluster.
The posterior distribution is depicted by solid lines. The prior distribution (assuming random choice,
with a mean of 0.5 and an STD of 0.1) is represented by dashed lines. To create these plots, eggs laid on

the tested side (where the speaker was active, or hydrated plant in the initial experiment) are denoted as 1, while those on the opposite side are marked as 0. These plots thus demonstrate the probability of obtaining a 1 or 0 in each experiment.

A) Drought-stressed vs. thriving plant (no playback). B1) Silence vs. drought-stressed plant playback (without a plant). B2) Deaf females in a setup with silence vs. drought-stressed plant playback (without a plant). B3) Silent plant vs. playback of drought-stressed plant. C) A box with male moth's vs an empty box. Tomato and male clicks are presented (time signal and spectrum) in panels B and C. The horizontal black bar depicts 0.1ms.

To control for a few of the assumptions made above, we conducted another experiment testing the main effect – of plant sounds on oviposition (See sound gradient experiment in the Methods). Namely, in this experiment we tested a single moth each time, with a lower biological-feasible plant click rate and with a long sound gradient. To this end, we placed a single female moth in a 150 cm long arena. On one side of the arena (location -75), a speaker played sounds recorded from a drought-stressed tomato plant (at a rate of 30 clicks per minute). On the other side of the arena (location +75), there was a silent resistor. A feeder with 60% sugar solution was positioned at the center (location 0, Fig. 2A). We then measured the distance from the center and the number of eggs for each cluster. The results, for both egg and cluster numbers, revealed a clear bimodal distribution with peaks near the feeder and the speaker. Hence, most clusters were laid very close to the feed or the speaker while no eggs were laid near the resistor the closest egg was 21cm away (Fig. 2B, both egg and cluster number distributions were significantly different from the expected H0 distribution which was estimated using permutation, K-S test, $p = 2.2 \times 10^{-16}$ for the clusters, $p = 3.9 \times 10^{-14}$ for the eggs, and see Methods and supplementary Fig. 3). This was thus a third independent validation that females prefer to lay eggs near plant playback and that this behavior is seen both when quantifying the individual egg or the cluster level

A Playback of drought- stressed plant vs. silence

Eggs

Drought sounds

Inverted sugar

Silence

-75   0   75
Distane (cm)

150 cm

B

Fig.2: Females lay eggs near acoustic playback. A) The long arena creates an acoustic gradient,

allowing us to investigate whether female moths prefer to lay their eggs in specific locations based on

the sound environment. Additionally, there is sugar water in the center of the arena, which serves as the

adult moth's food. B) Egg count density (solid line) and cluster density (dashed line). Both figures

display a bimodal distribution, with one peak near the speaker (-75) and another near the feeder (0).

The points under the graph depict laid clusters, illustrating the relationship between the number of eggs

per cluster and their spatial distribution within the arena.

To gain further insight into the moths' decision-making process, we repeated

experiment 2 (Fig. 1B) where drought-stressed sounds were played on one side of an

arena *without a plant* in three additional repetitions (with a total of N=13 females) while

videoing and tracking the entire behavior. In these repetitions, eggs were laid only on

the playback side of the arena. The continuous tracking showed that most moths (8 of

the 13) visited both sides of the arena, crossing sides 4.2 ±5.7 times (Mean±SD) on

average during the night (Fig.3 A). Moreover, over time, there was a significant increase in the female moths' tendency to spend more time in the playback side (Logistic GLMM, $p<0.004$, Fig.3 B.).

Fig.3: Females' movement and decision making. A) The continuous location over time in the arena (top-view) of 4 individual moths during one trial of the drought sounds vs. silent treatment. Time is represented by color in minutes, with a red triangle indicating the playback side and red X's marking the locations where eggs were laid. Note that we cannot be sure which of the individuals laid the eggs. B) The proportion of time moths spent in the playback side (in bins of 30 minutes) increased over time.

The moths' preference for the silent plants was not as strong as in the case of a dehydrating vs a fresh plant (Fig.1A). One possible explanation is that in the case of a real dehydrating plant, there are also olfactory cues that can assist the moth. To examine

the potential role of additional (olfactory) cues in the moths' decision, we recorded the

moths' antennal physiological response to the volatiles secreted from drying and

hydrated plants using Electroantennograms. Applying a Machine-Learning classifier,

we were able to discriminate between the response of the antenna to drying and

hydrated plants with high precision (an accuracy of 90±14%, see Supplementary Fig.2).

Discussion

We reveal first evidence for the use of acoustic information and specifically of sounds

typically emitted by plants in insect decision making. Despite decades of research on

plant vibrations, it has only recently been shown that these vibrations can be detected

remotely by organisms with ultrasonic hearing ability (Khait et al. 2023). Our current

results suggest that *Spodoptera littoralis* females detect and respond to ultrasonic clicks

which are typically emitted by drought-stressed tomato plants and adjust their choice

of oviposition accordingly. This finding opens a whole new range of possibilities for

animal-plant acoustic interactions.

Moreover, the presence of clicking male moths had no significant effect on the females'

oviposition preference. This suggests that female moths can distinguish between sounds

and specifically respond to plant-like sounds. Although the moth's hearing system

might be too simple to distinguish among the spectral properties of the different sounds,

i.e., male clicks vs. plant sounds (Nakano *et al.* 2013), the temporal patterns of the

sequences emitted in these varied situations are very different. While male moths emit

bursts of several clicks (Suppl. Fig.2), plants emit sporadic clicks with no clear temporal

order (as used in our playback). Playback of additional sound signals are needed to

examine moth specificity.

Although females responded in both treatments when ultrasonic drought-stressed

signals were played, they exhibited opposite preferences depending on the presence of

a plant. When there was no plant in the arena, the moths showed a strong preference to

the playback side, while when plants were present in the arena, the moths switched

preference to lay their eggs on the silent side. This latter choice was in accordance with

their preference to lay eggs on thriving vs. dry plants while the first choice (without a

plant) was somewhat surprising.

One explanation for this reversal in preference might be the multi-modal moth decision-

making process. When drought-stressed signals alone (without a plant) were presented

to the female moths, they might have become the only reliable signals for the presence

of a plant in the arena, which can explain their strong preference for this side. In
contrast, when we integrated thriving plants into the arena, the moths' decision making
became multi-factorial. Namely, on both sides of the arena, there were visual, texture,
and olfactory cues of thriving plants, while the treatment side also exhibited an acoustic
signal of a stressed plant. In this setup, the females' oviposition preference was reversed
to the side without the acoustic signal. This might suggest that the acoustic signal
interpretation is content dependent, i.e., that the playback of stress sounds in a multi-
factorial setup became a reliable signal of the physiological state of the plant. Therefore,
the females reverted back to their original preference to oviposition on thriving plants.
Supporting this hypothesis, the probability of laying eggs at all was significantly higher
when a plant was present than in the absence of a plant. Specifically, eggs were laid on
68% vs. 54% of the nights with and without plants respectively (p=0.009; Binomial test
comparing experiments two and three). The number of egg clusters was also higher
when a plant was present (see Fig.1). We conclude that the moths were more reluctant
to lay their eggs when no plant was present.
The preference for the silent plant vs. a plant with stress acoustic playback was not as
clear as the preference for the thriving hydrated plants (compare Fig.1A and Fig.1B3).
There are several potential explanations for this difference. First, moths probably rely
on various cues, including olfaction, to detect a drying plant. Although the playback
allowed us to isolate the specific effect of the acoustic cue, and we tried to select equal
plants, we could not control for other cues provided by the plant, and we may have
provided the animal with a partial (and likely even contradictory) set of cues. For
instance, the plants might have secreted drought-related volatiles and (although
watered) might have occasionally emitted sounds spontaneously, reducing the effect of
our playback. Indeed, a physiological measurement of plant volatiles suggested that
drying plants can be (at least partially) distinguished by the moths (Fig.S3).
We further investigated the behavioral mechanism of the female moths as they explored
the arena. We quantified the moths' movement during the decision process in the
experimental setup with drought-stressed acoustic signals played on one side, and with
an equal-impedance resistor on the other side. Our findings indicated that their decision
process typically included crossing over between the two sides of the arena and
spending an increasing amount of time on the (drought-stressed) playback side. This
suggests that females explore the available space and ultimately decide based on
comparing the two.

Various plant species emit airborne ultrasonic clicks when they are drought-stressed, which can serve as reliable cues for the physiological condition of the plant (Khait et al. 2023). Our findings demonstrate that moths with auditory abilities use these clicks when choosing a site for oviposition. We hypothesize that some other species of insects might also exploit these acoustic cues to their advantage in different contexts. Pollinating insects, for example, might use drought-related sounds when choosing where to forage. Some insects might even be able to distinguish between clicks produced by different plants or under different conditions, such as drying plants vs. plants under a pathogen attack.

Plant clicks are ultrasonic and thus very different from most other outdoors sounds (such as wind sounds, as we also show in Khait et al. 2023). Moreover, because the clicks are ultrasonic and not very intense, they can only be picked up by the moths from a short distance (~1.5m) which allows the moths to localize them in space.

The sounds emitted by drought-stressed plants are probably a cue rather than a signal, i.e., they did not evolve to convey information to insects. The interaction that we have demonstrated in this study therefore cannot be considered "communication" according to the conservative definition of the term, which relies on signals that have evolved to convey a specific message (Searcy and Nowicki 2005; Skyrms 2010). However, it is possible that some plants have evolved an ability to amplify their emissions or modify their spectral content to facilitate desirable interactions with animals and perhaps even with other plants (Veits et al. 2019). One exciting possibility would be that plants signal an insect attack by amplifying click intensity to recruit potential predators of the attacking insects, such as predatory insects, rodents, or bats. Such amplification could be achieved by various morphological modifications. Insects, on the other hand, might have evolved behavioral strategies to move near plants and pick up these weak acoustic signals. In conclusion, our study shows that moths are able to detect and respond to acoustic signals emitted by plants. This discovery suggests the existence of a third type of acoustic signal that moths utilize, in addition to those produced by bat echolocation and moth courtship clicks, raising new questions about the evolution of moth hearing. We predict that future studies will uncover more examples of acoustic communication between plants and animals.

Methods

Experimental setup –We collected pupae of *Spodoptera littoralis* that were reared under controlled breeding conditions (reared on castor bean leaves, 25 ± 1 °C, 40% relative humidity with a 12–12 h light–dark cycle). Newly-emerged female and male moths were closed together until egg-laying was detected (approximately two days). Then we transferred the females to an experimental arena. Each arena was 100 x 50 x 50 cm3 in size, divided in the middle by a plastic partition half the height of the arena (Fig. 1A). On the partition, we placed a closed test tube with cotton wool containing 60% inverted sugar solution for ad libitum feeding throughout the experiment. Experiment 1 (see below) was performed in a greenhouse (2.5 x 4.5 x 3.5 m^3) to simulate optimal conditions for plant development. The experiments involving acoustic signals (see below experiments 2,3,4,5 and 6) were performed in an acoustically shielded room (2.5×4×2.5 m^3) to prevent acoustic interference. Each of the following treatments was performed simultaneously in up to four arenas. Moths could choose between the treatments presented on each side of the arena (see below) and oviposition was monitored daily for three days by counting the number of egg clusters. We then repeated the experiments under the same conditions until acquiring at least nine nights with egg-laying observations (eggs were not always laid, which is not surprising given the artificial conditions in the acoustic room used for these experiments). We refer to the cluster and not to the individual egg as the moth's decision unit, because each cluster requires a decision about the location of oviposition, whereas the number of eggs could be affected by the general condition of the female or by external interference. Indeed, there was much variation in the number of eggs per cluster - 68 ±134 eggs (mean ± SE). However, to determine whether counting eggs would have altered our results, we conducted an experiment comparing cluster counts to individual egg counts (experiment 6). For experiments with actual plants, a young tomato plant (*Solanum lycopersicum*) in a small pot was used in all experiments. All the treatments are illustrated in Fig.1A-C. The number of repetitions of each treatment is noted in Table 1 and data is presented in supplementary Table 1.

1. Drought-stressed vs. well-hydrated plants: We placed a single-stem tomato plant, 10 cm high, on either side of the arena. The plant on one side was drought-stressed (three days without watering), and the other was thriving and well-hydrated. Moths could lay eggs on either plant (Fig.1A).

2. <u>Playback of a drought-stressed plant vs. silence (without plants):</u> Each side contained an oviposition box (10 x 15 x 5 cm^3 made of 0.5 x 0.5 cm^2 mesh), covered with a paper towel. A speaker playing sounds recorded from a drought-stressed tomato plant (Khait et al. 2018) was placed under one of the two oviposition boxes (on one side of the arena). The speaker played drought sounds at the same intensity measured for real plants at a rate of 1 Hz, with a stochastic 10% error in the intervals between clicks (see below for details on assessing intensity and playback rate). The oviposition box on the other side either had a resistor similar to the speaker in shape and identical in impedance to control for potential effects of the electric field created by the speaker, or no resistor. The experiment was performed twice to strengthen the confidence in its results: the first trial was performed during August and September 2021 and the second during February to May 2022 (A pool of both trials and controls - with and without resistors - is presented in Fig.1B2).

3. Deaf females in a setup with silence vs. drought-stressed plant playback (without a plant): we deafened mated females by puncturing their tympanic membrane and placed them in an arena to assess their response to drought-stressed sounds, compared to a silent control (as described in experiment 2). Deafening surgical procedure: We performed a surgical procedure on female moths to deafen them. The procedure involved puncturing the tympanic membrane located at the thoraco-abdominal juncture using an entomological needle #2. The female moths recovered from the procedure within 2 minutes and were able to fly normally. We tested a sample of these females in a standard rearing box and found that they were able to lay eggs normally. To confirm that the surgery had successfully deafened the females, we conducted an inspection by playing a bat playback (the same as described below). We deafened a group of 20 moths and compared their reactions to a control group of 25 non-deafened moths. During the experiment, the moths were released in a dark acoustically isolated room (5.5 × 4.5 × 2.5 m3) with acoustic foam on the walls and ceiling and a single light source (12W mercury vapor bulb peaked at 1,650 lux), and while they were in flight around the light source, we emitted the sound. In the control group, 5 moths exhibited a response (such as falling or a significant change in direction), upon hearing the sound (scored by a naïve viewer who did not know whether the moths were treated). In contrast, none of the deafened

399   moths displayed any reaction to the clicking stimulus (Q=4.5, p=0.03, Chi-
400   square test).

4. Well-hydrated plants with and without playback of drought-stressed plant
    sound: There was an oviposition box on each side of the arena. One side played
    drought-stressed sounds while the other remained silent, with either a resistor
    or no sound (same as experiment number 2). Additionally, a thriving, healthy
    tomato plant was placed on each oviposition box. This experiment was
    performed twice, 12 months apart, to strengthen the confidence of its results (a
    pool of both trials and controls [with and without resistors] is presented in
    Fig.1B3). To determine the specificity of the response to plant sounds, two
    additional controls were performed:

5. Male moths: Five males were enclosed under the oviposition box with sugar
    water to maintain them. The control box had only sugar water without any
    moths (Fig.1C). We validated that males in this condition produced clicks by
    recording the sounds emitted by the five male moths enclosed overnight in an
    acoustically isolated container which showed that the males frequently click.
    The test was repeated five times and clicks were always emitted by the males
    (Supplementary Fig.1).

Playback:

Drought sounds were recorded using an Hm16 Avisoft microphone and an HM116
Avisoft A/D from a distance of 10 cm in an isolated container with walls covered with
acoustic foam (Khait et al. 2018). These recordings revealed emission intensities of at
least 60 dB SPL (Re 20μPa) at a distance of 10 cm. The sounds were played using a
Vifa speaker connected to an Avisoft D/A converter (Player 116).

We ensured that playback sound intensity was similar to that measured in real plants
on the playback side of the arena (i.e., ~60dB SPL at a distance of 10 cm) and that
sound level on the control side was below the detection range of our system, that is,
below 30dB SPL at 10 cm. We performed four calibration measurements using a
calibrated GRAS 40DP microphone during the period of the experiments to validate
that sound levels had not changed over time. Using the GRAS calibrated microphone,
we also validated that the average sound intensity of the male moth sequences was the
same as that of the playback plant sounds.

Validating the playback rate: The drying plant sounds in the box arenas (experiments
2-4, Fig.1B) were played back at a rate of 1 Hz with up to 10% error in the intervals
(caused by the computer controlling the system). This frequency is substantially higher
than that found for a single young tomato plant (Khait *et al.* 2023). However, the rate
that we played (60 Hz) is ecologically relevant when considering a patch of tomato (or
other) plants. To validate this, we aggregated 45 tomato seedlings in a planting tray (30
x 30 cm^2) and placed the tray in an empty greenhouse. The plants were not watered for
three days, and we recorded sound continuously for 50 hours (using the same Hm116
microphone setup noted above). When placing the microphone ~20cm above the tray –
as a flying moth would do, we measured a maximum click rate of 0.33 Hz (i.e., 20 per
minute). This is three-fold slower than the rate we used, but very similar to the rate that
we used in the gradient experiment (see below). Moreover, when taking into account
the moth's detection range for this emission intensity which is likely ~1.5 meters at
least (Khait *et al.* 2023), a female moth could be exposed to a rate over three-fold higher
(i.e., higher than 1 Hz) in a patch of drying plants (which would contain more than 100
seedlings in a typical bush of agricultural or wild hosts typical of this species). Notably,
nearly every plant that we examined was found to emit similar ultrasonic clicks when
dehydrating (Khait *et al.* 2023), so this behavior could be relevant to other plants many
of which grow as dense bushes.

Table 1. Summary of experimental conditions including the number of repetitions, i.e. the

number of times that new moths were placed in the arenas and the number of observations (each

repetition was observed for approximately three consecutive nights). The total number of egg

clusters and the P-values for each experiment are reported. Experiments that were replicated

twice appear in two separate lines denoted for combined statistics and by #1 or #2. Experiments

and observations that did not produce any egg-laying were excluded from the data set and that

is why the number of observations is often the same as the number of repetitions.

Experiment	#Repetitions	#Observations	#Egg clusters	Mean± SE clusters on the side of the treatment	Mean ± SE clusters on the side of the Control	P-Value	Estimates (# of Egg clusters)
Drought-stressed plants vs. well-hydrated plants	17	17	53	0.88±1.11	2.23±2.68	0.005	0.93
Playback of a drought-stressed plant vs silence, combined trials (playback: 60 per minute)	38	45	67	1.08±0.82	0.4±0.65	0.000	1.00
Playback of a drought-stressed plant vs silence 1# (playback: 60 per minute)	11	17	24	1.11±0.69	0.29±0.58	0.012	1.34
Playback of a drought-stressed plant vs silence2# (playback: 60 per minute)	27	28	43	1.07±0.89	0.46±0.69	0.015	0.84
Deafened moths -Playback of a drought-stressed plant vs silence	23	23	39	0.7 ± 0.7	1 ± 1.09	0.550	0.12
Well-hydrated plants and playback of a drought-stressed plant, combined trials (playback: 60 per minute)	29	39	110	1.05±0.99	1.76±1.64	0.010	-0.52
Well-hydrated plants and playback of a drought-stressed plant 1# (playback: 60 per minute)	9	19	44	0.78±0.91	1.52±1.38	0.05	-0.66
Well-hydrated plants and playback of a drought-stressed plant 2# (playback: 60 per minute)	20	20	66	1.3±1.03	2±1.86	0.10	-0.43
Males vs no-males	19	29	48	0.72±0.92	0.93±1.33	0.39	-0.25

459 <u>Sound gradient experiment</u>

We used elongated arenas ($150 \times 20 \times 5$ cm^3, Fig.2A). In the center of the arena (location 0), we placed a closed test tube with cotton wool containing a 60% sugar solution for ad libitum feeding. To facilitate accurate measurement of egg distances from the speaker (location -75), we printed a ruler and placed it along the bottom of the arena. Each moth was placed at the center of the arena at the beginning of the experiment. On the next morning, we recorded the locations of the egg clusters and counted the number of eggs in each cluster using a stereoscopic microscope, or a magnifying glass if the eggs were not laid on the ruler. Each female remained in the arena during the days starting three days after emerging from the pupa and mating, and until it died. After each night, we switched the locations of the speaker and the resistor within the arena. We measured a 30 dB SPL difference in intensity between the side of the speaker and the side of the resistor. The clicks were emitted at a frequency of 0.5 clicks per second (30 per minute).

Tracking the females' decision-making process

In order to investigate how moths survey the experimental arena and subsequently engage in a decision-making process, we conducted two additional trials in which we continuously recorded the movement of the moths throughout the night. In each trial, we placed four female moths on a platform in the middle of the arena, in which a speaker played drought-stressed plant sounds on one side, while on the other, control side we placed a silent resistor (as in treatment 3 above). We exchanged sides between trials and tracked the moths for six hours using an IR camera (Reolink RLC-511-5MP camera) placed above the arena. We then documented the position of each moth at 12 seconds intervals using the DLTdv 8 software (Hedrick 2008). Each individual was recognized according to its proximity to the last tracking point in order to reconstruct its full movement. We quantified how many times each individual crossed the center of the arena (the platform in the center was divided in the middle), and the proportion of time it spent in each side.

Statistics

Mixed effect generalized linear models (GLMM) were used (in MATLAB) to examine the females' choice of oviposition. Random effects were set as intercepts. The number of clusters was set as the explained variable. The treatment, i.e., playback or control, and the number of female moths in the arena, was set as a fixed effect. The number of the arena, the month in which the experiment was performed, the number of repetitions,

and the night of the repetition were considered as random effects. Because we were analyzing counts (number of clusters), the model was run using a Poisson distribution. In the experiments in which we ran two repetitions of the same experiment, we added the session as another fixed parameter and we also ran the stats separately for each session.

To deepen our understanding of the trends observed in the experiments, we implemented Bayesian model fittings for each choice-based experiment. In this analysis, "oviposition choices" were considered as distinct decisions. A value of 1 was assigned when the egg cluster was located on the side with the active speaker (or on the hydrated plant in the initial experiment) and a value of 0 was assigned for oviposition on the opposite side. We employed a Gaussian model, incorporating the number of females in each experiment as a random effect, with a prior mean of 0.5 and a standard deviation of 0.1. For each experiment, we sampled our data 16,000 times to calculate the posterior distribution from these samples. We used a Binomial GLMM to determine the effect of the treatment on the moths' decision making. To achieve this, the proportion of time spent in each side of the arena was set as an explained variable, the playing side as a fixed effect, with the trial and the individual moth as random effects. To study the effect of time on the movement of the moths, we used Logistic GLMM in which the accumulated amount of time spent on the sound-playing side was set as an explained variable, the time as a fixed effect, and the trial and the individual moths were set as random effects.

To compare the distribution of eggs in the elongated arena to a random distribution, we generated an H0 distribution by randomly shuffling the locations of the speaker and resistor for each laid egg. This distribution was then compared to our actual egg count distribution using the Kolmogorov-Smirnov test (Supplementary Fig. 3).

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References

Amichai E, Yovel Y (2017) Bats pre-adapt sensory acquisition according to target

distance prior to takeoff even in the presence of closer background objects. Scientific

Reports 7:. https://doi.org/10.1038/s41598-017-00543-8

Amichai E, Yovel Y (2021) Echolocating bats rely on an innate speed-of-sound

reference. Proceedings of the National Academy of Sciences 118:.

https://doi.org/10.1073/PNAS.2024352118

Anton, S., Evengaard, K., Barrozo, R. B., Anderson, P., & Skals, N. (2011). Brief

predator sound exposure elicits behavioral and neuronal long-term sensitization in the

olfactory system of an insect. *Proceedings of the National Academy of Sciences of the*

United States of America, 108(8), 3401–3405.

https://doi.org/10.1073/pnas.1008840108

Boppré M (1978) Chemical communication, plant relationships, and mimicry in the

evolution of danaid butterflies. *Entomologia Experimentalis et Applicata* 24:264–277

Brito VLG, Weynans K, Sazima M, Lunau K (2015) Trees as huge flowers and flowers

as oversized floral guides: The role of floral color change and retention of old flowers

in Tibouchina pulchra. *Frontiers in Plant Science* 6:1–10.

https://doi.org/10.3389/fpls.2015.00362

Conner WE (1999) "Un chant d'appel amoureux": Acoustic communication in moths.

Journal of Experimental Biology 202:1711–1723.

https://doi.org/10.1242/jeb.202.13.1711

Fenton MB, Fullard JH (1979) The influence of moth hearing on bat echolocation

strategies. *Journal of Comparative Physiology* A .132:77–86.

https://doi.org/10.1007/BF00617734

Fullard, J. H. (1998). The sensory coevolution of moths and bats. In *Comparative*

Hearing: Insects pp. 279-326. New York: Springer-Verlag

Göpfert MC, Hennig RM (2016) Hearing in Insects. *Annual Review of Entomology*

61:257–276. https://doi.org/10.1146/annurev-ento-010715-023631

Gori DF (1989) Floral Color Change in Lupinus argenteus (Fabaceae): Why Should

Plants Advertise the Location of Unrewarding Flowers to Pollinators? *Evolution*

43:870

Greenfield MD, Weber T (2000) Evolution of ultrasonic signaling in wax moths:

Discrimination of ultrasonic mating calls from bat echolocation signals and the

559 exploitation of an antipredator receiver bias by sexual advertisement. *Ethology*

560 *Ecology and Evolution* 12:259–279. https://doi.org/10.1080/08927014.2000.9522800

561 Hedrick TL. (2008), Software techniques for two- and three-dimensional kinematic

562 measurements of biological and biomimetic systems, *Bioinspiration & Biomimetics,*

563 3(3):034001

564 Hoy RR (1996) Tympanal Hearing in Insects. *Annual Review of Entomology* 41:433–

565 450. https://doi.org/10.1146/annurev.ento.41.1.433

566 Kawahara AY, Plotkin D, Espeland M, Meusemann K, Toussaint EFA, DonathA,

567 Gimnich F, Frandsen PB, Zwick A, dos Reis M, Barber JR, Peters RS, Liu S, Zhou X,

568 Mayer C, Podsiadlowski L, Storer C, Yack JE, Misof B, Breinholt JW (2019),

569 Phylogenomics Reveals the Evolutionary Timing and Pattern of Butterflies and

570 Moths, *Proceedings of the National Academy of Sciences of the United States of*

571 *America* 116(45):22657–63. https://pnas.org/doi/full/10.1073/pnas.1907847116

572 Kevan P. G., and Lane, M. A., (1985). Flower petal microtexture is a tactile cue for bees.

573 *Proceedings of the National Academy of Sciences*, *82*(14), 4750–4752.

574 https://doi.org/10.1073/pnas.82.14.4750

575 Khait, I., Lewin-Epstein, O.,Sharon, R., Saban, K., Goldstein, R., Anikster, Y., Zeron,

576 Y., Agassy, C., Nizan, S., Sharabi, G., Perelman, R., Boonman, A., Sade, N., Yovel,

577 Y., & Hadany, L. (2023). Sounds emitted by plants under stress are airborne and

578 informative. *Cell*, *186*(7), 1328-1336.e10. https://doi.org/10.1016/j.cell.2023.03.009

579 Lhomme P, Carrasco D., Larsson M., Hansson B., Anderson P. (2018) A context-

580 dependent induction of natal habitat preference in a generalist herbivorous insect.

581 *Behavioral Ecology:*29:360–367, https://doi.org/10.1093/beheco/arx173

582 Milburn JA, Johnson RPC (1966) The conduction of sap. Planta 69:43–52

583 Moir HM, Jackson JC, Windmill JFC (2013) Extremely high frequency sensitivity in a

584 "simple" ear. *Biology Letters* 9:. https://doi.org/10.1098/rsbl.2013.0241

585 Nakano R Takanashi T Fujii T Skals N Surlykke A Ishikawa Y. (2009) Moths are not

586 silent, but whisper ultrasonic courtship songs. *Journal of Experimental Biology*.

587 212:4072–4078.

588 Nakano R, Takanashi T, Surlykke A, Skals N, Ishikawa Y. (2013) Evolution of deceptive

589 and true courtship songs in moths. *Scientific Reports*.3:2003. doi: 10.1038/srep02003.

590Nakano R, Takanashi T, Surlykke A (2014) Moth hearing and sound communication.

591 J*ournal of Comparative Physiology A:* 201:111–121

Ne'eman G (1995) Pollination Ecology and the Significance of Floral Color Change in

Lupinus Pilosus L. (Fabaceae). *Israel Journal of Plant Sciences*.

https://doi.org/10.1080/07929978.1995.10676599

Ponomarenko A., Vincent O., Pietriga A., Cochard H., Badel É. and Marmottant P.

(2014) Ultrasonic emissions reveal individual cavitation bubbles in water-stressed

wood. *Journal of Royal Society Interface*: 11. https://doi.org/10.1098/rsif.2014.0480

Proffit M, Khallaf MA, Carrasco D, Larsson MC, Anderson P (2015) 'Do you remember

the first time?' host plant preference in a moth is modulated by experiences during

larval feeding and adult mating. *Ecology Letters* 18:365–374

Robert D, Göpfert MC (2002) Novel schemes for hearing and orientation in insects.

Current opinion in neurobiology 12:715–20

Sadek MM, Hansson BS, Anderson P (2010) Does risk of egg parasitism affect choice of

oviposition sites by a moth? A field and laboratory study. *Basic and Applied Ecology*

11:135–143. https://doi.org/10.1016/j.baae.2009.09.003

Salama HS, Dimetry NZ, Salem SA (1971) On the Host Preference and Biology of the

Cotton Leaf. *Zeitschrift für Angewandte Entomologie* 67:261–266

Schiestl FP (2010) The evolution of floral scent and insect chemical communication.

Ecology Letters 13:643–656. https://doi.org/10.1111/j.1461-0248.2010.01451.x

Searcy, W. A. & Nowicki, S. (2005). The Evolution of Animal Communication:

Reliability and Deception in Signalling Systems. Princeton University Press,

Princeton.

Shvil, N., Golan, A., Yovel, Y., Ayali, A., & Maoz M., B. (2023). The Locust antenna as

an odor discriminator. *Biosensors and Bioelectronics*, *221*.

https://doi.org/10.1016/j.bios.2022.114919

Skals, N., Anderson, P., Kanneworff, M., Löfstedt, C., & Surlykke, A. (2005). Herodours

make him deaf: Crossmodal modulation of olfaction and hearing in a male moth.

Journal of Experimental Biology, *208*(4), 595–601. https://doi.org/10.1242/jeb.01400

Skyrms B. (2010) Signals: Evolution, Learning, and Information. Oxford University

Press

Spangler HG (1988) Moth hearing, defense, and communication. *Annual Review of*

Entomology Vol 33 59–81. ttps://doi.org/10.1146/annurev.en.33.010188.000423

Tougaard, J. (1996). Energy detection and temporal integration in the noctuid A1

auditory receptor. *Journal of Comparative Physiology A:* 178.

Tougaard J. (1998). Detection of short pure-tone stimuli in the noctuid ear: what are

temporal integration and integration time all about? *Journal of Comparative*

Physiology A: 183, 563-572.

Tyree MT, Dixon MA (1983) Cavitation Events in *Thuja occidentalis L*.? . *Plant*

Physiology 72:1094–1099. https://doi.org/10.1104/pp.72.4.1094

van Dam NM, Bouwmeester HJ (2016) Metabolomics in the Rhizosphere: Tapping into

Belowground Chemical Communication. *Trends in Plant Science* 21:256–265.

https://doi.org/10.1016/j.tplants.2016.01.008

Veits, M., Khait, I., Obolski, U., Zinger, E., Boonman, A., Goldshtein, A., Saban, K.,

Seltzer, R., Ben-Dor, U., Estlein, P., Kabat, A., Peretz, D., Ratzersdorfer, I., Krylov,

S., Chamovitz, D., Sapir, Y., Yovel, Y., & Hadany, L. (2019). Flowers respond to

pollinator sound within minutes by increasing nectar sugar concentration. *Ecology*

Letters, 22(9), 1483–1492. https://doi.org/10.1111/ele.13331

Supplementary Fig 1. Male Egyptian cotton leaf moths (*Spodoptera littoralis*) courtship sequences recorded when we placed males in the arena (spectrogram presented).

Supplementary Figure 2. Electro-antennograms (EAG) were used to record the electrophysiological responses of (n=9) female moth antennae (removed from the body) while exposing them to drying or hydrated tomato plant odors (n=3 in each group). To present the odors, we utilized six tomato plants of identical age, of which three were deprived of irrigation for 72 hours before the experiment. To introduce the odors to the antennae, we fashioned a tube from a leaf of each plant, through which air, carrying the scent, was blown towards the antennae using a pump. The full recording information can be found here (Shvil *et al.* 2023). The recordings (n=30) are presented in the left panel (Mean±SE) with dry in red and hydrated in blue. We applied a Support Vector Machine (SVM) to examine the separation between the dry and dehydrated plant recordings using 80% (24 signals) of the available data for training and a 5-fold cross validation. This procedure resulted in average accuracy of 90±14% when tested on the remaining test signals (Right panel shows a confusion matrix of the test signals of all 5 cross validations).

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Supplementary Fig 3. On the left, comparison between the egg count results (solid
line) in the elongated arena and the pseudo-random distribution (dashed line) (K-S test,
D = 0.3, p = 2.2 × 10⁻¹⁶). On The right, comparison between the clusters count results
(solid line) pseudo-random distribution (dashed line) (K-S test, D = 0.21, p = 3.9 ×
10⁻¹⁴).The speaker was placed on location -75, a feeder was placed on the center
(location 0) and a resistor was placed on location 75.
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669
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A

Playback of drought- stressed plant vs. silence

Eggs

Drought sounds

Inverted sugar

Silence

-75   0   75
Distane (cm)

150 cm

B

Density / Egg count (#) vs Distance (cm)

A

B