Characterisation of the genomic sequence of an iflavirus, a protoambidensovirus, and of three microviruses detected in mosquitoes (*Aedes albopictus* and *Culex quinquefasciatus*)

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Running title

insect-viruses and phages identified in mosquitoes

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Abstract

We report the complete CDS of five viruses: an iflavirus, a protoambidensovirus and three microviruses), which were detected by viromics surveillance of mosquitoes (*Aedes albopictus* and *Culex quinquefasciatus*) from the Réunion Island. We detected the protoambidensovirus, which belongs to a clade previously reported only in *C. pipiens*, in *A. albopictus*.

Announcement

Our knowledge of mosquito viruses remains limited, although they can modulate the transmission of arboviruses **(1)**. We processed 13 *Aedes albopictus* and 9 *Culex quinquefasciatus* larvae samples collected in La Réunion Island in 2019. Morphological characteristics were used for species identification, and larvae were pooled by location. Samples weighted between 0.5g and 1.49g, and stored at -80°C. Viromes were obtained as described in **(2)**. We followed manufacturers' instructions and default parameters except where otherwise noted. Samples were homogenised by a bead-beater, filtered through a 0.45 μm filter and centrifuged at 140,000 g for 2.5 hours. The pellets were resuspended, digested by DNaseI and RNaseA incubation at 37°C for 1.5 h. RNA and DNA were extracted using a NucleoSpin kit. Reverse transcription was performed by SuperScript III, cDNAs were purified by a QIAquick PCR Purification Kit and dsDNA synthetised by Klenow DNA polymerase I. DNA was amplified

by random PCR, and PCR products were purified using a NucleoSpin gel and PCR clean-up kit. Libraries

were prepared using a NEBNext Ultra DNA PCR free with Ilumina adapter kit without fragmentation

step, and sequenced on an Illumina HiSeq3000 in 2x150bp paired-end mode.

Adaptors were removed and reads were filtered for quality (q30 quality and read length >45 nt) using

cutadapt 2.19 **(3)**. 10,150,317 reads were assembled into contigs by MEGAHIT 1.2.9 (4). Taxonomic

assignment was achieved using DIAMOND 0.9.30 with against the NCBI nr protein database **(5)**. Genome

coverage was assessed by mapping using Bowtie2 3.5.1 (end–to-end very-sensitive) **(6)**. Open Reading

Frames (ORFs) were identified using ORF finder (length cutoff >400 nt) on Geneious prime 2024.0.5 **(7)**,

and were annotated by blastn query-centered alignment against RefSeq viral database on October 2024.

Genome circularisation was performed using Simple-Circularise (overlap >20nt).

We reconstructed the complete circular genomes of three bacteriophages species from the *Microviridae*

family (*Gokushovirinae* subfamily) (PQ041302, PQ041303 and PQ041304). They share 54.5% to 75.0% of

amino acid identity in major capsid and replication-associated proteins with their closest relatives, which

were isolated from water environments (**Table 1**). There is currently no species demarcation criteria

based on genome similarity defined by the ICTV for the *Microviridae* family **(8)**. Their high abundance in

Aedes samples suggests than they may infect its microbiota, although their presence could also be due

to environmental or diet contamination.

We report a complete CDS of a lineage of *XiangYun picorna-like virus 4* species (*Iflaviridae*, *Iflavirus*)

(**Table 1**, PQ041301). It shares a whole polyprotein pairwise identity of 98.5% with a lineage discovered

in *Culex theileri* from Yunnan, China (OL700176) **(9)**. This lineage clustered in a monophyletic clade

containing only viral taxa isolated from dipteran species (mosquitoes and true flies), indicating that it is

likely specific of dipterans (**10.6084/m9.figshare.28565888**). Further work is needed to determine the

prevalence of XiangYun picorna-like virus 4 in natural populations of mosquitoes and its impact on

mosquito fitness.

We report the complete CDS of a distant lineage of the mosquito infecting *Dipteran*

protoambidensovirus 1 species (*Parvoviridae*, *Densovirinae*, *Protoambidensovirus*, **Figure 1**, PQ041300).

It shares 89.0% genome-wide nucleotide identity to its closest relative, Culex pipiens densovirus

(FJ810126.1) (**Table 1**) **(10)**. This lineage belongs to the CpDV-3 clade which was previously only

represented by sequences collected from Beijing (China) samples in 2003 **(11)**. Reported CpDV host

range currently only includes *Culex pipiens* **(12,13)**, its broadening to *A. albopictus* warrants further

confirmation.

Data availability

The genomic sequences of the five full-length viral genomes or CDS have been deposited at GenBank

under the accession numbers PQ041300 to PQ041304. High-throughput sequencing reads were

deposited in SRA under the accession no. SRR29133481 to SRR29133504 under PRJNA1114772

BioProject.

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Figure legend

Table 1 Information on the five viruses reconstructed from mosquito viromic data

Figure 1 Maximum likelihood phylogenetic tree based on the NS1 nucleotide sequences of 52
Dipteran protoambidensovirus 1 (common virus name: Culex pipiens densovirus (CpDV))
sequences. The alignment of 892 nucleotides in length was produced by MAFFT v7.450
using the G-INS-i algorithm. The tree is mid-point rooted. Bootstrap values (100
replicates) are indicated at each node. Scale bar corresponds to nucleotide substitutions
per site. The CpDV sequence obtained from our sample is coloured in red.

Table 1 Information on the five viruses reconstructed from mosquito viromic data

Virus	GenBank accession number	Genome		Coverage			Putative proteins			Closest identified relatives			
		size (nt)	%GC	average	number of reads	sample	name	size (nt)	size (aa)	virus name	accession number	pairwise identity (aa)	source
Culex quinquefasciatus associated densovirus, isolate 2019_VP12-D87	PQ041300	5 459	36,7	12 521	501 573	Cxqfts virome sample 3 (D87) SRR29133503				Culex pipiens densovirus	FJ810126		Culex pipiens
							NS1	687	229			94,7	
							NS2	372	214			92,2	
							NS3	642	124			42,4	
							VP	2 208	736			90,5	
Culex quinquefasciatus associated iflavirus, isolate 2019_VP12-D85	PQ041301	10 074	36,9	2 777	194 020	Cxqfts virome sample 1 (D85) SRR29133486	polyprotein	9 231	3 077	XiangYun picorna-like virus 4	OL700176	98,5	Culex theileri
Aedes albopictus associated microvirus 1, isolate 2019_VP12-D83	PQ041302	4 365	43,9	243	8 198	Aedes virome sample 11 (D83) SRR29133492	major capsid protein	1 650	550	Microvirus sp. isolate BS1_474	MT309944	75,0	wastewater
							DNA pilot protein	678	226	Microviridae sp. isolate CD-SXD-WGA-1-k141_518849	OR998997	43,3	bacteria
							internal scaffolding protein	429	143	Microvirus sp. isolate BS1_462	MT309947	54,2	wastewater
							replication initiator protein	1 023	341	Arizlama microvirus	MW697615	54,9	fresh water
Aedes albopictus associated microvirus 2, isolate 2019_VP12-D75	PQ041303	4 387	46,3	62	2 200	Aedes virome sample 3 (D75) SRR29133501	major capsid protein	1 635	545	Microviridae sp. isolate SD_MC_72	MH572427	67,3	Ciona robusta
							DNA pilot protein	801	267	Microvirus sp. isolate LSGA_102419	OR349779	23,4	mollusc
							internal scaffolding protein	474	158	Microviridae sp. isolate ctda862	MH616902	65,8	fish
							replication initiator protein	747	249	Microvirus sp. isolate BS1_438	MT309954	74,7	wastewater
Aedes albopictus associated microvirus 3, isolate 2019_VP12-D82	PQ041304	4 813	50,5	197	6 618	Aedes virome sample 10 (D82) SRR29133485	major capsid protein	1 653	551	Microviridae sp. isolate ctcc708	MH617066	73,4	fish
							DNA pilot protein	795	265	Microviridae sp. isolate ctji855	MH617641	37,6	fish
							internal scaffolding protein	444	148	Microviridae sp. isolate ctcc708	MH617066	61,9	fish
							replication initiator protein	783	261	Microvirus sp. isolate 1712115_622	MT310309	68,6	wastewater