Spatial Proteomics by Trapped Ion Mobility supported MALDI MS/MS Imaging: A First Glance into Multiplexed and Spatial Peptide Identification

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Abstract

RATIONALE

In spatial proteomics, matrix-assisted laser desorption/ionization (MALDI) imaging enables rapid and cost-effective peptide measurement. Yet, in situ peptide identification remains challenging. Therefore, this study aims to integrate the trapped ion mobility spectrometry (TIMS)-based parallel accumulation-serial fragmentation (PASEF) into MALDI imaging of peptides to enable multiplexed MS/MS imaging.

METHODS

An initial MALDI TIMS MS1 survey measurement is performed, followed by a manual generation of a precursor list containing mass over charge values and ion mobility windows. Inside the dual TIMS system, submitted precursors are trapped, separately eluted by their ion mobility and analyzed in a quadrupole time-of-flight device, thereby

enabling multiplexed MALDI MS/MS imaging. Finally, precursors are identified by peptide to spectrum matching.

RESULTS

This study presents the first multiplexed MALDI TIMS MS/MS imaging (iprm-PASEF) of tryptic peptides. Its applicability is showcased on two histomorphologically distinct tissue specimens in a 4-plex and 5-plex setup. Precursors were successfully identified by the search engine MASCOT in one single MALDI imaging experiment for each respective tissue. Peptide identifications were corroborated by liquid-chromatography tandem mass spectrometry experiments and fragment colocalization analyses.

CONCLUSIONS

In the present study, we demonstrate the feasibility of TIMS-based MALDI MS/MS imaging for the multiplexed and rapid identification of tryptic peptides in a spatial manner. Hence, it represents a first step in the integration of MALDI imaging into the emerging field of spatial proteomics.

Abbreviations

ACN	Acetonitrile
AmBC	Ammonium bicarbonate
CHCA	Alpha-cyano-4-hydroxycinnamic acid matrix
DDA	Data-dependent acquisition
ESI	Electrospray ionization
FFPE	Formalin-fixed paraffin-embedded
IHC	Immunohistochemistry
iprm-PASEF	Imaging Parallel Reaction Monitoring, Parallel Accumulation SErial Fragmentation
ITO	Indium tin oxide
LC-MS/MS	Liquid-chromatography tandem mass spectrometry
m/z	Mass over charge
MALDI	Matrix-assisted laser desorption/ionization
MS	Mass spectrometry
prm	Parallel Reaction Monitoring
PASEF	Parallel Accumulation SErial Fragmentation
PDX	Patient-derived xenografts
PSM	Peptide to spectrum matching
Q-TOF	Quadrupole time-of-flight
TFA	Trifluoroacetic acid
TIMS	Trapped Ion Mobility Spectrometry
$1/K_0$	Inverse of reduced ion mobility

1 Introduction

Matrix-assisted laser desorption/ionization (MALDI) imaging is a powerful method to probe the spatial distribution of biomolecules including metabolites, lipids, peptides, and proteins. In the field of spatial proteomics, MALDI imaging stands out by enabling the antibody-independent, mass spectrometry-based, rapid, cost-effective, and spatial analysis of peptides and proteins[1-5]. Yet, in situ identification by mass spectrometry remains a major challenge in MALDI imaging since most measurements are limited to only MS1 spectra level, thus missing peptide fragment spectra (MS2), which are the backbone for peptide sequence information in mass spectrometry-based proteomics[6,7].

As a work-around for missing MS2 spectra, most MALDI imaging workflows rely on matching MALDI-based MS1 masses to peptide identifications of additional liquid-chromatography tandem mass spectrometry (LC-MS/MS) of the same (or an equivalent) sample. However, deep proteome coverage in LC-MS/MS often yields near-isobaric peptide sequences, which limits applicability of this strategy[6,8,9].

Direct in situ identification of peptide ions in MALDI imaging, i.e. by ion fragmentation, is considered to be optimal but is hampered by several hurdles[7,10,11]: First, the absence of chromatographic separation (as used for LC-MS/MS) leads to detection of numerous near-isobaric peptides resulting in co-fragmentation and chimeric MS2 spectra as well as limited enrichment of peptides and their ions. Second, most precursor selection systems for MALDI imaging have been typically confined to filtering for one precursor ion per set of laser shots, thus hindering multiplexing and resulting in a high sample consumption and long measurement times.

During recent years, quadrupole time-of-flight (Q-TOF) tandem mass spectrometry has been enhanced by preceding trapped ion mobility spectrometry (TIMS) for both electrospray ionization (ESI) and MALDI systems. A dual TIMS cell system sorts and traps precursor ions based on their collisional cross section (CCS), thereby allowing separation of isobaric and isomeric peptides. The TIMS cell measures the closely related ion mobility (K) value. Within this study we will report the inverse reduced ion mobility ($1/K_0$), which standardizes the ion mobility for temperature and pressure. TIMS-based synchronization of ion storage and release with subsequent, quadrupole-based ion selection and fragmentation steps leads to very efficient ion usage; a setup called parallel accumulation serial fragmentation (PASEF)[12]. It is already widely used in LC-MS/MS proteomics[13-18] and also applied to targeted proteomics as parallel reaction monitoring (prm)-PASEF[19]. More recently, novel acquisition software enhancements have made the prm-PASEF mode available to commercial MALDI imaging systems such as in the timsTOF fleX (Bruker Daltonics, Bremen, Germany), thereby enabling the multiplexed identification of peptide ions per set of laser shots. This software tool is called iprm-PASEF (Bruker Daltonics, Bremen, Germany) and its applicability has already been demonstrated for lipids and metabolites[8,20,21].

The present study is the first to optimize and apply TIMS-based acquisition modes including iprm-PASEF to MALDI imaging of tryptic peptides for improving peptide identifications.

2 Materials and Methods

2.1 Tissue Specimens and Tissue Processing

Mouse kidney tissues were obtained as surplus tissue from animals that had been sacrificed for unrelated experimental purposes in accordance with approved animal experimentation protocols. The mouse kidney samples represent leftover material that would have been discarded otherwise. Human patient-derived xenograft (PDX) tumor specimens were provided by Charles River Laboratories, Freiburg, Germany. This study was carried out in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the Society of Laboratory Animals (GV SOLAS). The PDX experiments were approved by the Committee on the Ethics of Animal Experiments of the regional council (Permit Numbers: I19-02). Bovine liver was obtained commercially from a local butcher. All tissues were formalin-fixed and embedded in paraffin (FFPE) according to standard protocols[22]. FFPE blocks were sliced into 2 µm thick sections and mounted onto IntelliSlides (Bruker Daltonics, Bremen, Germany) or indium tin oxide (ITO) slide (Bruker Daltonics, Bremen, Germany).

2.2 Peptide Samples

For standard peptides and iprm-PASEF validation, an iRT synthetic peptide kit was used (Biognosys AG, Schlieren, Switzerland). For LC-ESI-TIMS-MS/MS optimization, Pierce HeLa Protein Digest Standard (Thermo Fisher Scientific, Waltham, Massachusetts, USA) was used. Further synthetic peptides were used for iprm-PASEF validation: Apelin-13 (Santa Cruz Biotechnology, Dallas, USA), Substance P (Merck, Darmstadt, Germany), and two further synthetic peptides (KLKESYCQRQGVPMN, KLKVIGQDSSEIHFKV) (GenScript, Piscataway Township, New Jersey, USA).

2.3 Sample Preparation for MALDI Imaging of Tryptic Peptides

For all MALDI imaging measurements, two adjacent tissue slides were prepared simultaneously as described previously[23]. Deparaffinization was carried out in a series of xylol and ethanol/water dilutions as described previously[22]. Tissue sections were washed twice with 10 mM ammonium bicarbonate (AmBC) for 1 min. Antigen retrieval was performed in a citric acid monohydrate buffer with pH 6.0 in a steamer for 1 h at approximately 100 °C. Slides were rinsed twice with 10 mM AmBC and dried for 10 minutes in a vacuum atmosphere at room temperature. Freshly prepared, N-tosyl-

L-phenylalanine chloromethyl ketone treated trypsin (Worthington, Lakewood, NJ, USA) (0.1 mg/mL in 10 % acetonitrile (ACN), 40 mM AmBC) was sprayed on the tissue slides using the M3+ sprayer (HTX Technologies, Chapel Hill, NC, USA) with the following parameters: Temperature 30 °C, nozzle velocity 750 mm/min, flow rate 30 µL/min, number of passes 8, track spacing 2 mm, pattern CC, drying time 0 s, and nitrogen gas pressure set to 10 psi. Afterwards, slides were incubated in a humidified chamber for 2 h at 50 °C. Alpha-cyano-4-hydroxycinnamic acid matrix (CHCA) (Sigma-Aldrich, Munich, Germany) (10 mg/mL, 70 % ACN, 0.1 % trifluoroacetic acid (TFA), 7 mM ammonium phosphate) was applied using the M3+ sprayer with the following parameters: Temperature 60 °C, nozzle velocity 1350 mm/min, flow rate 100 µL/min, number of passes 8, track spacing 3 mm, pattern CC, drying time 10 s, and nitrogen gas pressure set to 10 psi. Slides were either measured directly or stored in a vacuum desiccator for less than 24 h until final measurement. For validation purposes, two additional setups were carried out: First, FFPE bovine liver slides were prepared for MALDI imaging measurement in triplicates as described above. After tryptic digestion and before matrix application, iRT kit peptides were spotted twice in a dilution series (400 fmol, 200 fmol, 100 fmol, 50 fmol in 0.5 µL droplets). Second, iRT kit peptides, Apelin-13 , Substance P , and synthetic peptides (KLKESYCQRQGVPMN, KLKVIGQDSSEIHFKV) were mixed with CHCA matrix and spotted onto an empty ITO slide in triplicates to compare 1-plex and 5-plex iprm-PASEF measurements.

2.4 MALDI TIMS MS1 Imaging Acquisition and Data Processing

MALDI TIMS MS1 imaging datasets were acquired in the positive-ion reflector mode using a timsTOF fleX mass spectrometer (Bruker Daltonics, Bremen, Germany) at a pixel size of 50×50 µm, using 600 shots and a laser frequency of 10 kHz. The instrument is equipped with a Nd:YAG laser emitting at 355 nm. The *m*/*z* range was set to 800-2,000 *m*/*z*, the $1/K_0$ range to 1.2-2.1 V•s/cm^2, with a ramp time of 230 ms. Tuning parameters were optimized und set as follows: MALDI plate offset 80 V, deflection 1 delta 80 V, funnel 1 RF 250 Vpp, funnel 2 RF 500 Vpp, multiple RF 500 Vpp, collision energy 10 eV, collision RF 4000 Vpp, ion energy 5 eV, transfer time 185 ms, and pre pulse storage 20 µs. The TIMS cell pressure was decreased by adjusting the nitrogen flow until the calibration mass 1221 *m*/*z* (Hexakis(1H, 1H, 4H-hexafluorobutyloxy)phosphazine (Agilent Technologies, Santa Clara, USA)), aligned to 170 V, yielding a TIMS cell pressure of 2.0 mbar. $1/K_0$ and *m*/*z* calibration was performed over ESI with the ESI-low concentration tuning mix (Agilent Technologies, Santa Clara, USA). For validation purposes, MALDI TIMS off MS1 imaging measurements were carried out with TIMS turned off and all other parameters kept identical.

2.5 MALDI TIMS MS1 Imaging Data Analysis

MALDI TIMS MS1 imaging data was analyzed with the software SCiLS Lab (Bruker Daltonics, Bremen, Germany). The *m*/*z* variance was kept to 10 ppm for *m*/*z* and

0.01 V•s/cm^2 for 1/K_0. Root mean square normalization was always selected. Feature finding was carried out with T-ReX[3] (TIMS), spatial smoothing 2×2, 50 % coverage and 1 % relative intensity threshold. In cases where feature selection was slightly off, the peak maximum was manually selected in the two-dimensional *m*/*z* and 1/K_0 plot. Segmentation was performed only on the tissue region, denoising was set to none, with bisecting k-means and correlation distance selected. For the "Colocalization with Region" tool, the entire tissue region was selected and correlated with one of the segmented regions of interest determined by the spatial segmentation tool. For the "Colocalization with Feature" tool, the entire tissue region was selected and correlated with the feature of interest. Feature lists from feature finding as described above were used as input for all colocalization analyses.

2.6 Sample Preparation of Peptides Co-crystallized with MALDI Matrix for LC-ESI-TIMS-MS/MS

To recover the co-crystallized peptides after MALDI imaging acquisition, the matrix layer on the tissue slide was dissolved, collected with 70 % ACN, and vacuum-dried for ACN removal[8]. Peptides were cleaned up with PreOmics Cartridges (PreOmics, Planegg/Martinsried, Germany). The eluted peptides were vacuum-dried and dissolved in water and 0.1 % formic acid before loading on Evotips (Evosep, Odense, Denmark) for final desalting and LC-ESI-TIMS-MS/MS measurement.

2.7 LC-ESI-TIMS-MS/MS Acquisition Focused on Singly Charged Peptides

LC-ESI-TIMS-MS/MS measurements of matrix peptides including 10 fmol iRT Kit were conducted on a timsTOF fleX mass spectrometer equipped with a CaptiveSpray ion source (Bruker Daltonics, Bremen, Germany). Chromatographic separation was performed with the Evosep One HPLC system (Evosep, Odense, Denmark), using the 30 samples per day method (44 min gradient, 500 nL/min) and the EV1137 performance 15 cm reversed phase column (Evosep, Odense, Denmark) with buffer A consisting of 0.1 % aqueous formic acid and buffer B of 0.1 % formic acid in ACN. Samples were measured in positive data-dependent acquisition (DDA)-PASEF mode. To increase measurement of 1+ charged peptides, adjustment of the TIMS cell pressure and ion mobility calibration were performed as described in MALDI TIMS MS1 acquisition. The mass range was set to 100–1,700 *m/z* and 1/K_0 range to 0.7–2.1 V•s/cm^2. Collision-induced dissociation energy was set to 20–75 eV, ion accumulation time to 166 ms with a 100 % duty cycle. The PASEF precursor polygon region was adjusted manually to include singly charged peptide traces. Tuning parameters were optimized und set as follows: Deflection 1 delta 80V, funnel 1 RF 250 Vpp, funnel 2 RF 500 Vpp, multiple RF 500 Vpp, collision energy 10 eV, collision RF 2,500 Vpp, ion energy 5 eV, transfer time 70 ms, and pre pulse storage 15 µs. The capillary voltage was set to 1,600 V and the drying gas flow rate was 3 L/min with 180 °C drying temperature.

2.8 LC-ESI-TIMS-MS/MS Data Analysis

The FragPipe pipeline (v22.0) was employed for the analysis of the DDA-PASEF datasets[24-26]. Peptide identification of MS2 spectra was obtained using MSFragger with raw (.d) files as input. Protein databases, either human (UP000005640, downloaded August 13, 2024) or mouse (UP000000589, downloaded June 10, 2024), were both prepared with common contaminants, and internal standards (iRT Kit peptides). Reversed sequences were added as decoys. Default settings for label free quantification were applied with proline +15.999 (hydroxylation) set as a variable modification in MSFragger.

2.9 iprm-PASEF Acquisition and Data Processing

The iprm-PASEF datasets were acquired using a prototype version of the iprm-PASEF tool provided by Bruker Daltonics GmbH, Bremen, Germany. Measurements were performed in MS/MS and positive ion reflector mode on the timsTOF fleX at a pixel size of 100×100 µm, using 2,000 shots and a laser frequency of 5 kHz. The *m*/*z* range was set to 100–2,000 *m*/*z*, the $1/K_0$ range was set to 1.2–2.1 V•s/cm^2 with a ramp time of 230 ms. Tuning parameters were optimized and set as follows: Collision RF 750 Vpp, ion energy 5 eV, transfer time 55 ms, pre pulse storage 8 µs. Adjustment of the TIMS cell pressure and $1/K_0$ calibration were performed as described in MALDI TIMS MS1 imaging acquisition. The iprm-PASEF methods were set up with up to 5 precursor ions with a predefined $1/K_0$ and *m*/*z* range. Collision energies were adjusted depending on the precursor mass from 45–140 eV. For validation purposes, MALDI TIMS off MS/MS imaging measurements were performed with only TIMS turned off and all other parameters kept identical. In the DataAnalysis software (Bruker Daltonics, Bremen, Germany), MS2 spectra of each individual precursor were averaged over all pixels, exported from the iprm-PASEF datasets as individual ".mgf" files and submitted to the "MASCOT MS/MS Ions Search" (Matrix Science, London, UK) against the Swiss-Prot (2024/04) database including contaminants filtered by the respective taxonomy (mus musculus, homo sapiens)[27]. Enzymatic digestion was set to "Trypsin" with two missed cleavages allowed. The precursor tolerance was set to 100 ppm, fragment ion tolerances to 0.3 Da, and target FDR to 1 %. Histidine, tryptophane, proline and methionine oxidation as variable modification were included. MS2 spectra and assigned theoretical b- and y-ions were visualized using the pyteomics package in python[28]. Theoretical b- and y-ions were matched with a tolerance of 0.3 Da. Ion images of peptide fragments were plotted in SCiLS Lab. For colocalization analysis of fragments, features were picked manually in SCiLS Lab and the "Colocalization to Feature" tool was exploited.

3 Results

3.1 Overview of Ion Mobility-Enabled, Multiplexed and MS/MS-based MALDI Imaging of Peptides

Spatial proteomics using MALDI imaging benefits from the direct, in situ MS/MS based identification of peptide ions. TIMS-enabled MALDI imaging supports this approach by ion mobility-based precursor separation and entrapment according to distinct ion mobility features, thereby allowing multiplexing per set of laser shots. For measurement in iprm-PASEF, precursor ions of interest are selected within predefined ranges of their relevant molecular properties, namely *m*/*z* and $1/K_0$. The spatial proteomics approach performed in this study consists of four steps: (A) An initial MALDI TIMS MS1 survey measurement (50×50 µm), (B) manual generation of a precursor list, (C) iprm-PASEF based MALDI imaging for the creation of MS2 spectra of those precursors (100×100 µm), and (D) a peptide to spectrum matching (PSM) and potential corroboration. We used 50 µm resolution in the MS1 scan for refined spatial analysis, while iprm-PASEF at 100 µm for identification focused on a faster acquisition and higher spectral quality.

The initial MALDI TIMS MS1 measurement (Figure 1A) yields an overview of locally abundant MS1 precursor ions including *m*/*z* and $1/K_0$ values from which a feature list (typically 2,000-8,000 features) is derived. However, multiplexed precursor ion trapping in the first TIMS chamber for iprm-PASEF relies on non-overlapping $1/K_0$ windows. In addition, those windows need to be sufficiently wide to allow for efficient isolation. These requirements typically limit the number of precursor ions to 25 entries with distinct ion mobility windows. Therefore, a sophisticated precursor selection is necessary.

As iprm-PASEF is a targeted workflow, the precursor definition relies on manual selection (Figure 1B). Strategies for automated, “on the fly” precursor extraction for MALDI imaging (similar to DDA in LC-ESI-TIMS-MS/MS) are highly anticipated but, to our knowledge, presently not yet available. Exemplary approaches for manual precursor selection include exploratory statistical analyses of the MS1 imaging data, filtering the MS1 data for the most intense signals, a literature-based search for specific peptides/proteins of interest, an in-silico digestion of specific proteins of interest, and LC-ESI-TIMS-MS/MS-based libraries generated from measurements of the same or comparable tissue specimen. For demonstration purposes, the precursors selected in this study were compiled by exploratory statistical analysis combined with a literature search. This resulted in a customized precursor list containing *m*/*z* values and $1/K_0$ windows.

The iprm-PASEF proof-of-concept measurement was performed at 100×100 µm spatial resolution, resulting in measurement times less than 100 min for 1 cm^2 area (2,000 number of laser shots with 5 kHz frequency). In our preliminary experience, the

reduction to 5 kHz enables prolonged accumulation times that benefit spectra intensity. The entire tissue section was scanned in iprm-PASEF mode resulting in spatially resolved information of fragment ions. For subsequent PSM of individual MS2 spectra, we resorted to the online version of the well-established MASCOT software. For this purpose, ion mobility filtered MS2 spectra were averaged and exported to the ".mgf" format as implemented in the DataAnalysis software (Figure 1C).

We further aimed to demonstrate additional analyses to validate peptide identification by iprm-PASEF (Figure 1D): For a visual and spatial-resolved corroboration of the peptide identification, the mean MS2 spectra were plotted and correctly detected b- and y-ions of the identified peptides were visualized. The ion images of the fragments were examined example-wise to show similar spatial distribution as expected and statistically analyzed by the "Colocalization to Feature" tool in SCiLS Lab. We further aimed to support the iprm-PASEF peptide identifications with PSM of LC-ESI-MS/MS (incl. TIMS) data. This step can be pursued with additional sample material or with matrix extracted peptides that have remained following initial MALDI imaging[8]. If the same peptide is identified by LC-ESI-TIMS-MS/MS, precursor m/z and $1/K_0$ values can be compared with MALDI imaging data to validate their identities.

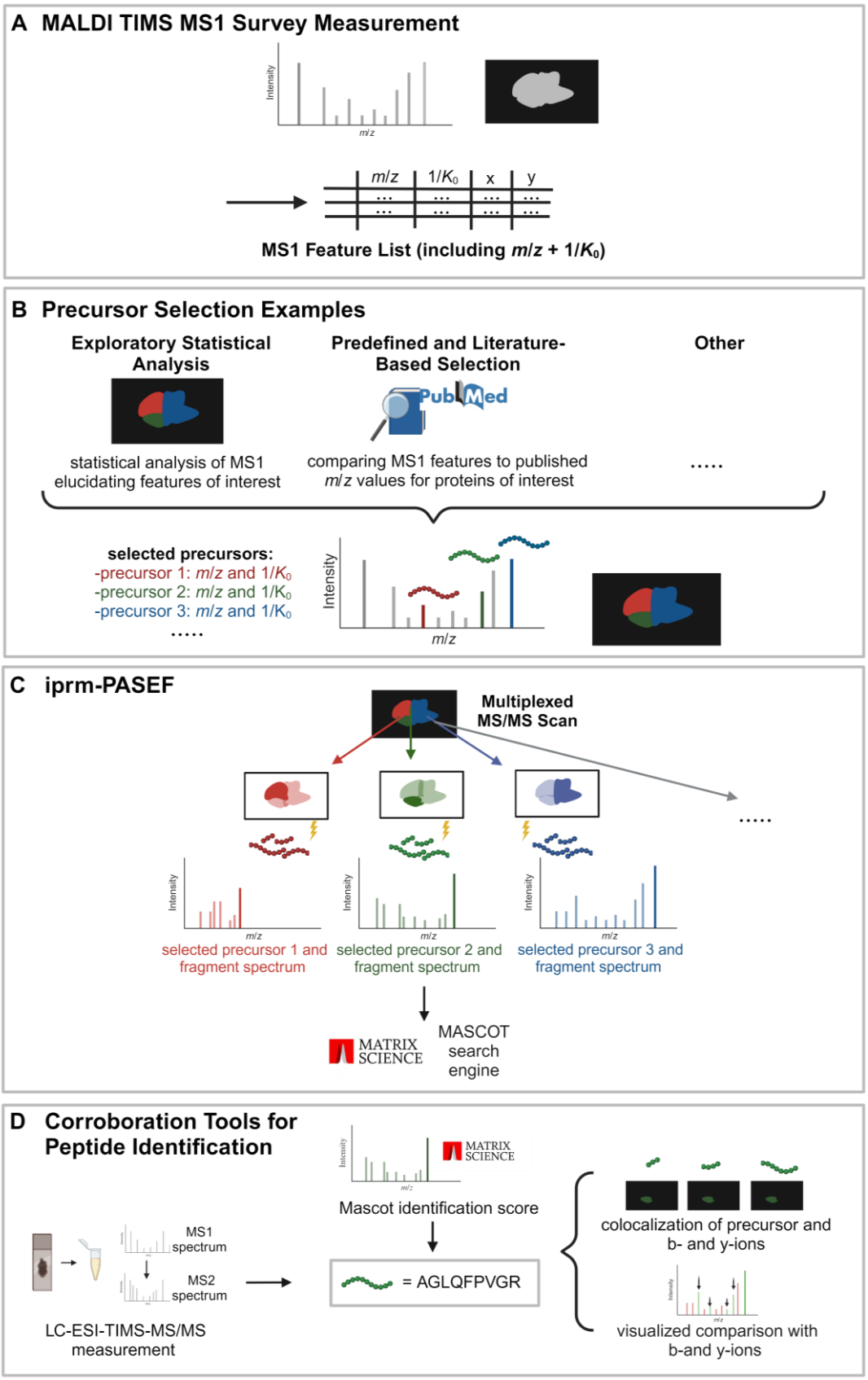

Figure 1: Overview of the iprm-PASEF Workflow used in this study. (A) The MALDI TIMS MS1 survey imaging scan provides a feature list. **(B)** Precursors are selected from the feature list. Two selection options are visualized in this figure. **(C)** With the selected precursors and their distinct *m/z* and $1/K_0$ values, a multiplexed MALDI TIMS MS/MS imaging (iprm-PASEF) scan is performed. The resulting MS2 spectra are identified using the MASCOT search engine. **(D)** Overview of the corroboration tools used in this study. (Created in BioRender. Schilling, O. (2024) https://BioRender.com/g48n665)

3.2 Dual TIMS Does Not Impede MS1 Sensitivity in MALDI Imaging

As outlined, usage of TIMS for selection and entrapment of precursor ions is fundamental for iprm-PASEF. We aimed to probe whether the dual TIMS step prior to Q-TOF measurement impacts MS1 sensitivity as compared to a Q-TOF measurement on its own. To this end, we adjusted TIMS parameters to accommodate for peptidic precursors: The ramp time was increased to 230 ms for sufficient separation and the TIMS cell nitrogen gas flow was lowered to 2.0 mbar to optimize accumulation.

To evaluate the effect of upstream dual TIMS usage on MS1 sensitivity, the iRT peptide kit was spotted in a dilution series on trypsin-digested bovine liver tissue using standard settings for MS1 MALDI imaging. Without dual TIMS (hence Q-TOF on its own), the lowest detectable spot was at 50 fmol per peptide. Inclusion of the dual TIMS step for MALDI TIMS MS1 yielded a very comparable result (Figure 2A). For all iRT ion images see Supplementary Figure 1. We conclude that inclusion of the dual TIMS step for MALDI MS1 does not impede MS1 sensitivity.

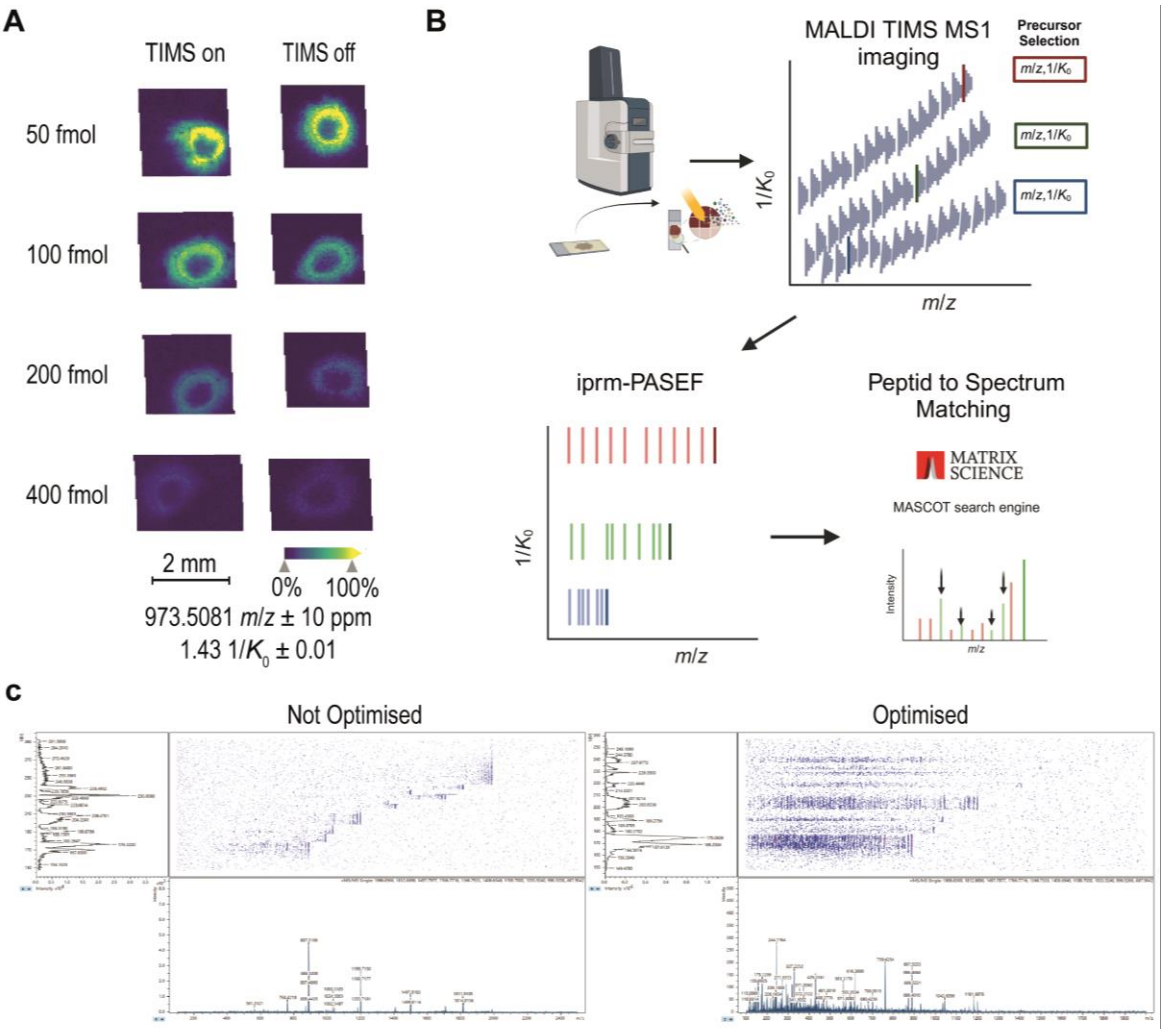

Figure 2: Evaluation of TIMS-supported MALDI MS1 imaging of tryptic peptides. (A) Synthetic tryptic peptides (iRT kit) were spotted on a bovine slice tissue section, CHCA-matrix was applied and MALDI MS1 imaging measurement was carried out either in TIMS on or TIMS off mode. The data was visualized and analyzed using the SCiLS Lab software. **(B)** Overview of the iprm-PASEF methodology. After a MALDI TIMS MS1 imaging scan, specific precursors are selected for a MALDI TIMS MS/MS imaging (iprm-PASEF) scan. MS2 spectra are analyzed using MASCOT (Created in BioRender. Schilling, O. (2024) https://BioRender.com/j85k835) **(C)** Comparison of the mobility heatmap between default and optimized method, visualized in timsControl (Bruker Daltonics, Bremen, Germany).

3.3 iprm-PASEF of Tryptic Peptides

iprm-PASEF is an implementation of multiplexed MALDI TIMS MS/MS imaging to generate spatially resolved MS2 spectra (Figure 2B). When establishing the TIMS-enabled, multiplexed MS/MS data acquisition of tryptic peptides, we noticed the need to optimize the acquisition method to enable the detection of low *m/z* fragment signals with additional sensitivity for high molecular weight precursor masses (Figure 2C). Compared to the default peptide imaging method provided by Bruker Daltonics, we adjusted relevant tuning parameters as follows: “Collision RF” from 4,000 Vpp to 750 Vpp, “Transfer Time” from 185 ms to 55 ms and “Pre Pulse Storage” from 20 µs to 8 µs. The *m*/*z* range was set to 100-2,000 *m*/*z*. Collision energies were increased dependent on *m*/*z* from 45 to 140 eV.

We further evaluated the impact of dual TIMS on MS2 spectra quality. Equally sized areas of tryptic digested bovine liver tissue slice were imaged in MS/MS mode in triplicates. Three precursors were selected for the MS/MS measurement (Supplementary Table 1). Apart from TIMS turned on or off, all other parameters were kept identical. Since independent measurements are required for every single precursor with TIMS turned off, the measurement area was tripled and measurement time took longer compared to iprm-PASEF. All MS2 spectra per precursor were averaged, exported in the ”.mgf” format, and submitted to MASCOT for PSM. In iprm-PASEF, all three precursors were matched to peptide sequences with MASCOT scores >35. Without the TIMS dimension, only one precursor was matched to a peptide sequence. However, for TIMS off methods, individual, precursor-specific refinement of acquisition parameters may yet improve spectral quality and was not pursued in the present study. Overall, iprm-PASEF allows for the simultaneous and multiplexed acquisition of MS2 spectra over a wide precursor *m*/*z* range (Figure 3).

Next, we assessed whether multiplexing in iprm-PASEF affects MS2 spectrum quality. Five synthetic peptides (see Supplementary Figure 2 for sequences) were mixed with CHCA matrix and spotted on an ITO slide (see section 2.2). Then, iprm-PASEF measurements were performed; either using multiplexing of all five precursors or individually aiming for one precursor after another. Visual inspection of the resulting MS2 spectra suggests highly similar MS2 spectra from both measurement approaches (Supplementary Figure 2).

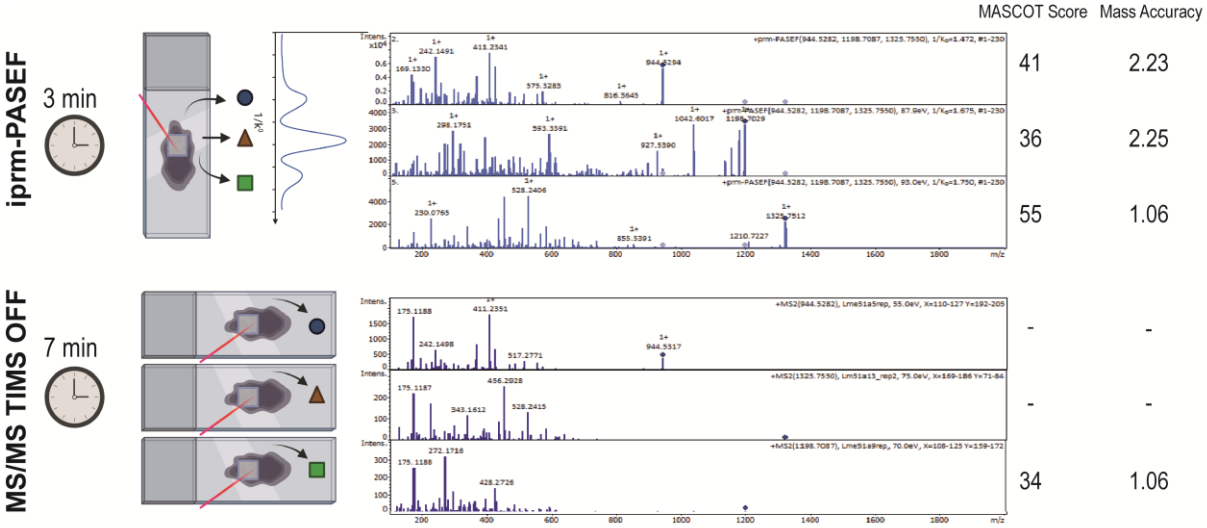

Figure 3: Validation of the multiplexed MALDI MS/MS imaging using the novel iprm-PASEF Software. Comparison between iprm-PASEF (TIMS on) and MALDI MS/MS imaging TIMS off measurements. Three precursors are targeted in both modes on a prepared bovine liver tissue slice. In iprm-PASEF, all three precursors can be measured simultaneously whereas in MALDI MS/MS imaging in TIMS off mode only one precursor can be targeted at a time. Mass accuracy shown in ppm (Created in BioRender. Schilling, O. (2024) https://BioRender.com/j51r217, modified)

3.4 Optimization of LC-ESI-TIMS-MS/MS Measurement to Focus on Singly Charged Peptides

LC-ESI-TIMS-MS/MS based peptide identifications may serve to corroborate MALDI TIMS MS/MS based peptide identifications. MALDI ionization produces mostly singly charged ions in the typical mass range of tryptic peptide, whereas ESI usually yields tryptic peptide ions of 1+ to 4+. For improved comparability of MALDI and ESI data, we sought to focus on singly charged ions in LC-ESI-TIMS-MS/MS, especially since $1/K_0$ values are charge state dependent[29-31]. Therefore, we optimized the tuning parameters in LC-ESI-TIMS-MS/MS measurements to increase the detection of 1+ charged peptides. An overview of an optimization experiment testing different settings can be found in Supplementary Table 2. Key adjustments include the TIMS cell pressure set similar to the MALDI imaging acquisition, increasing the upper $1/K_0$ limit and redefining the PASEF polygon precursor region to include singly charged precursor ions.

Next, we assessed whether $1/K_0$ values for the same peptide sequence are comparable between LC-ESI-TIMS-MS/MS and MALDI imaging. To this end, iRT peptides (Biognosys AG, Schlieren, Switzerland) were measured in both modes, (LC-ESI-TIMS-MS/MS and MALDI TIMS MS1 imaging). The *m/z* and $1/K_0$ values were highly comparable between those two measurements with an *m/z* variance of <5 ppm and a $1/K_0$ variance of <0.01 V•s/cm^2 (Table 1).

Table 1: *m/z* and $1/K_0$ values of iRT peptides obtained from MALDI imaging spots on bovine liver. In LC-ESI-TIMS-MS/MS, 20 fmol of iRT Kit was measured together with 100 ng HeLa digest.

Peptide	*m/z* LC-ESI-TIMS-MS/MS	*m/z* MALDI	*m/z* error (ppm)	$1/K_0$ LC-ESI-TIMS-MS/MS (V•s/cm^2)	$1/K_0$ MALDI (V•s/cm^2)	$1/K_0$ error (V•s/cm^2)
LGGNEQVTR	973.5061	973.5055	-0.616	1.443	1.434	0.009
GTFIIDPGGVIR	1,244.6997	1,244.6981	-1.285	1.739	1.745	0.006
TPVISGGPYEYR	1,338.6688	1,338.6705	1.270	1.755	1.751	0.004
TPVITGAPYEYR	1,366.7000	1,366.6939	-4.463	1.777	1.772	0.005
DGLDAASYYAPVR	1,397.6695	1,397.6699	0.286	1.791	1.789	0.002

3.5 Exemplary Application to FFPE Tissue Specimens

To further elaborate the novel, MS/MS-enabled, MALDI-based spatial proteomics technique, two samples were selected as histomorphologically distinct tissue specimens. An FFPE mouse kidney sample was analyzed in a 5-plex setting and breast cancer tissue from murine patient-derived xenografts (PDX tumor) was measured in a 4-plex setup, demonstrating the applicability of the workflow to different tissue species.

3.5.1 MALDI-TIMS-MS1 Survey Measurement

MS1 peptide imaging took 138 minutes generating 107 minutes for 22,977 pixels (mouse kidney) and 30,245 pixels (PDX tumor) with a spatial resolution of 50 µm, resulting in a measurement speed of approximately 3.5 pixels per second. Initial data assessment including feature finding, data visualization and normalization was carried out in SCiLS Lab. Feature finding resulted in 7,536 (PDX tumor) and 6,119 features (mouse kidney), respectively.

3.5.2 Precursor Selection

Segmentation and colocalization tools in SCiLS Lab were exploited to reveal region-specific features. Spatial segmentation was carried out to segregate two distinct tissue subregions (Figure 4A, red and green). The “Colocalization to Region” tool was employed, resulting in colocalization scores indicating correlation of features to the respective subregion. Additionally, a “Colocalization to Feature” was carried out for 836.4425 *m/z* on the mouse kidney to find features with similar spatial distribution (Supplementary Table 3). Results were further combined with a literature search to

reveal putative peptide identifications by *m/z* value. MS1 ion images of all selected precursors are shown in Figure 4B.

3.5.3 iprm-PASEF

For iprm-PASEF of the mouse kidney, the set of five precursors chosen by exploratory statistical analysis was expanded with two randomly chosen, additional precursors due to $1/K_0$ window availability (Supplementary Table 4). The iprm-PASEF measurement of the PDX tumor employed a set of four predefined precursors (Supplementary Table 4).

The iprm-PASEF measurements generated 5,666 pixels in 47 minutes (mouse kidney) and 10,406 pixels in 81 minutes (PDX tumor) with a spatial resolution of 100 µm, resulting in a measurement speed of approximately 2 pixels per second. Mean MS2 spectra for each precursor were separated by their $1/K_0$ traces and submitted to the search engine MASCOT, allowing for the assignment of peptide identities to the measured spectra. All precursors determined by MS1 statistical analysis were successfully identified by MASCOT (Table 2, Figure 4B). In contrast, the identity of the randomly selected precursors for the mouse kidney remained elusive. Mass accuracies were lower than 20 ppm for all precursors. The MASCOT scores ranged from 23.8 to 89.9 and were comparable with scores obtained in other published in situ MALDI MS/MS studies[32-35].

Presently, all acquired MS2 spectra per precursor over the entire tissue region are being averaged to yield a single MS2 spectrum, which we employed for the PSM step. We consider the possibility that quality filter steps prior to MS2 spectra averaging (e.g. signal-to-noise thresholds, subregion selection) may further improve final MS/MS data quality. However, this aspect was not further investigated in the present proof-of-concept study.

3.5.4 Corroboration Tools for Peptide Identification

To corroborate the peptide identifications obtained by MASCOT, we performed a number of additional analyses (Figure 1C):

Since the online MASCOT tool only provides an identification score, the pyteomics package was employed to examine theoretical b- and y-ions as corroboration. All detected b- and y-ions and the mean MS2 spectra can be found in the supplementary information (Supplementary Figure 3, Supplementary Table 5) and in Table 2. For instance, all possible y-ions and 9.1 % of possible b-ions of the respective peptide DNIQGITKPAIR (1,325.7528 *m*/*z*, PDX tumor) and for peptide GPAGPQGPR (836.4425 *m*/*z*, mouse kidney) all possible y-ions and 50 % of b-ions were detected. Additionally, fragment colocalization was investigated to include the acquired spatial distribution. For demonstration purposes, the colocalization analysis was only performed on one precursor for each tissue, 1,325.7528 *m*/*z* for the PDX tumor and

836.4425 *m*/*z* for the mouse kidney. In SCiLS Lab, the “Colocalization to Feature” tool was used with the respective precursor *m*/*z* as feature. Colocalization scores were calculated for high quality ion images of the fragment ions and reached from 0.50 to 0.90 (PDX tumor) and 0.40 to 0.56 (mouse kidney) (Supplementary Table 6). Peptides ending on a C-terminal arginine are usually well detected in MALDI-TOF devices, explaining the high amount of identified y-ions[36]. The MS2 spectra and ion images of the fragment signals including the percentage of detected b- and y-ions are visualized for the two exemplary precursors in Figure 4C.

To demonstrate further corroboration by LC-ESI-TIMS-MS/MS, we extracted the residual matrix peptides after the MALDI imaging measurement and submitted them to LC-ESI-TIMS-MS/MS analysis[8]. By focusing on singly charged peptides, we detected seven out of nine precursors in 1+ charge, allowing for the comparison of ion mobility in addition to the *m*/*z* value. One precursor (1,198.7003 *m*/*z*) was only detected on 2+ charge and one precursor at 836.4425 *m*/*z* failed to be detected in LC-ESI-TIMS-MS/MS. For the mouse kidney, *m*/*z* variance was lower than 10 ppm and ion mobility variance lower than 0.04 V•s/cm^2. For the PDX tumor, the variance for *m*/*z* was lower than 15 ppm and for ion mobility lower than 0.01 V•s/cm^2 (Table 2).

Using these two test FFPE tissues, we demonstrated that iprm-PASEF enables multiplexed in situ MS/MS, making it a valuable tool in the field of spatial proteomics.

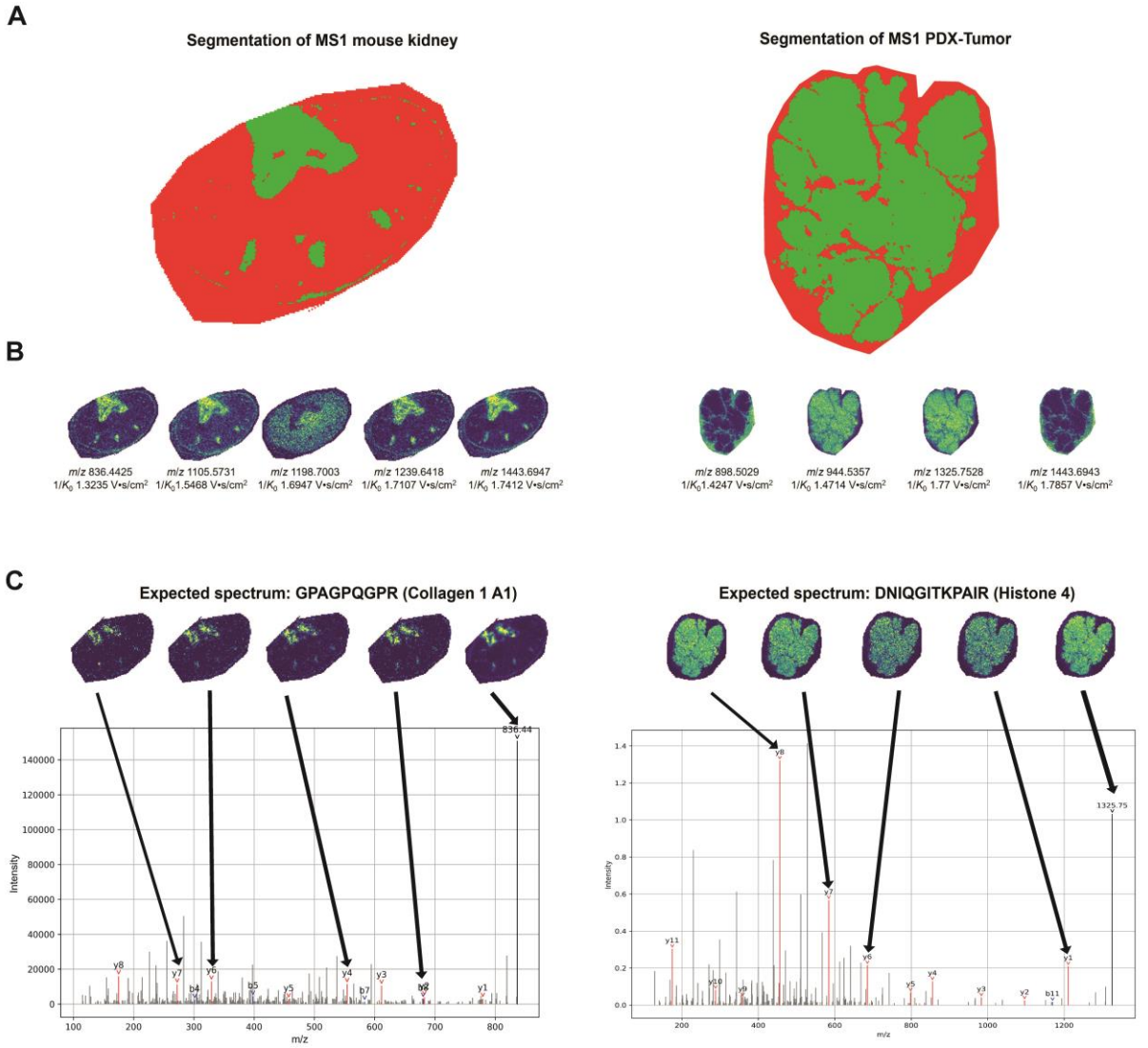

Figure 4: Selection and multiplexed identification of precursors in one single iprm-PASEF measurement **(A)** Segmentation of MS1 datasets conducted in SCiLS Lab are shown up to two subregions. **(B)** MS1 ion images of precursors selected for iprm-PASEF. **(C)** MS2 spectra acquired in iprm-PASEF mode with a spatial resolution of 100 µm are shown for one selected precursor of each test tissue. B-ions are highlighted in red and y-ions in blue. Ion images of respective fragments are shown on top of each spectrum.

Table 2: Overview of peptide identity corroborations by iprm-PASEF and LC-ESI-TIMS-MS/MS measurements of two test tissues.

Protein name	Peptide Sequence	*m/z* MALDI	MASCOT score (higher better)	MASCOT mass accuracy (ppm)	*m/z* LC-ESI-TIMS-MS/MS	*m/z* difference (ppm)	Charge stage LC-ESI-TIMS-MS/MS	$1/K_0$ MALDI (V•s/cm^2)	$1/K_0$ LC-ESI-TIMS-MS/MS (V•s/cm^2)	$1/K_0$ difference (V•s/cm^2)	Number of fragment ions detected	Measured Tissue
Collagen 1A1	GVVGLP[16]GQR	898.5029	2.8	38.6	898.5035	-0.67	1	1.4247	1.4167	0.008	3/8 (b) 7/8 (y)	PDX tumor
Histone H2A type 1-B/E	AGLQFPVGR	944.5373	2.4	34.27	944.5239	14.19	1	1.4714	1.4676	0.0038	1/8 (b) 8/8 (y)	PDX tumor
Histone 4	DNIQGITKPAIR	1,325.7528	2.3	89.94	1,325.7487	3.09	1	1.77	1.7707	-0.0007	1/11 (b) 11/11 (y)	PDX tumor
Collagen 1 A1	GSAGPPGATGFP[16]GAAGR	1,443.6980	19.1	43.3	1,443.6866	7.90	1	1.7822	1.7857	-0.0035	5/16 (b) 12/16 (y)	PDX tumor
Collagen 1 A1	GPAGPQGPR	836.4425	6.1	52.67	-	-	-	1.3255	-	-	4/8 (b) 8/8 (y)	Kidney
Collagen 1 A1	GVQGPP[16]GPAGPR	1,105.5731	2.4	39.07	1,105.5647	7.60	1	1.5468	1.5854	-0.0386	7/11 (b) 10/11 (y)	Kidney
Actin A	AVFPSIVGRPR	1,198.7003	3.0	28.92	1,198.7029	-2.17	2	1.6947	0.9633		4/10 (b) 8/10 (y)	Kidney
Collagen 1 A2	GIP[16]GPAGAAGATGAR	1,239.6418	1.6	28.8	1,239.6357	4.92	1	1.7107	1.7408	-0.0301	7/14 (b) 12/14 (y)	Kidney
Collagen 1 A1	GAAGPP[16]GATGFP[16]GAAGR	1,443.6943	1.9	23.84	1,443.6807	9.42	1	1.7412	1.758	-0.0168	8/16 (b) 12/16 (y)	Kidney

4 Discussion

In this study, we present the first performance of iprm-PASEF on tryptic peptides as an initial step in improving MALDI-based spatial proteomics. This method enables us to tackle the issue of efficient spatial peptide identification as a key challenge in MALDI imaging of peptides. We demonstrate the ability to identify peptidic precursor ions on two different FFPE tissue samples in a spatially resolved manner using iprm-PASEF. Peptide identification was corroborated using the MASCOT PSM search of MS2 spectra and additional LC-ESI-TIMS-MS/MS measurements.

Notably, the presented technique is highly adaptable and can be customized to suit specific experimental needs. For instance, MS1 imaging and iprm-PASEF can be conducted on the same slide with a laser offset, reducing sample usage and simplifying co-registration. Choosing a laser offset can also allow for multiple iprm-PASEF runs with different precursor lists on the same slide, thereby increasing the number of targeted precursors. Furthermore, our data analysis pipeline relied on four different software programs (DataAnalysis, SCiLS Lab, MASCOT, Python/pyteomics), requiring multiple import and export steps. Manual adjustments were necessary due to e.g. limited performance of the SCiLS Lab feature detection tool for ion mobility dimensional data. This highlights significant potential for further optimization and automation of the data analysis workflow.

In conclusion, this study integrates TIMS and PASEF into MALDI imaging of tryptic peptides as a novel approach in the field. These methods have already significantly advanced LC-ESI-TIMS-MS/MS shotgun proteomics, therefore providing great potential to improve peptide MALDI imaging as well. Our results demonstrate the feasibility of this approach for multiplexed peptide identification, making tryptic peptide MALDI imaging a promising method for spatial proteomics. Looking forward, further applications may be possible, such as extending isolation windows to allow a data-independent-acquisition-PASEF-like methodology, thereby allowing accumulation of specific ions. Therefore, TIMS technology offers great potential to enhance the sensitivity for low abundant peptides, which will be explored in the future. Overall, the presented workflow for the spatial identification of tryptic peptides by MALDI imaging provides a powerful tool with possible applications for future clinical research and diagnostics.

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