Title: Species biology and demographic history determine species vulnerability to climate

change in tropical island endemic birds

Ratnesh Karjee[1,†], Vikram Iyer[1, †], Durbadal Chatterjee[1, †], Rajasri Ray[2], Kritika M. Garg[1,3],

Balaji Chattopadhyay[1,*]

[1]Department of Biology, Trivedi School of Biosciences, Ashoka University, Sonipat, Haryana,

India

[2]Centre for Studies in Ethnobiology, Biodiversity and sustainability (CEiBa), Malda, West

Bengal, India

[3]Centre for Interdisciplinary Archaeological Research, Ashoka University, Sonipat, Haryana,

India

[†]These authors contributed equally to this work

*Corresponding author: Balaji Chattopadhyay

Email address: balaji.chattopadhyay@ashoka.edu.in

ABSTRACT

Climate change and associated habitat fluctuations can expedite the diversification of insular

lineages or lead to isolation and extinction. Tropical island birds are a model system to assess

responses to climate change owing to their insular nature and ability to diversify rapidly. While

there is some understanding of the diversification of tropical island avian clades, we are yet to

understand the vulnerability of these species to climate change. Long-term species genetic

diversity and demography are critical predictors of this. Therefore, we investigated the sensitivity

of tropical island endemic birds to climate change and analysed how species traits determine

these responses by comparing species traits with demographic histories and paleohabitat

fluctuations during the Last Glacial Period (LGP). From publicly available whole genome and

paleoclimatic datasets, we reconstructed tropical island endemics' past demographic histories

(effective population size (Ne)) using Pairwise Sequential Markovian Coalescent (PSMC) (n =

22) and suitable habitats (n = 28) during the LGP and the Holocene. We observed that most

species experienced an increase in suitable habitat between the Last Interglacial and Last Glacial

Maximum. However, a concomitant increase in Ne was only observed in the hyper-diverse

passerine clade, attesting to their ability to rapidly diversify. Overall, diet specialists and large-

bodied species showed a decrease in Ne during the LGP. Our results indicate that species traits

dictate tropical island endemics' demographic responses to climate, and a plastic response to

habitat availability could be a consequence of clades' abilities to rapidly occupy new niches and

diversify. Further, our analyses revealed that most species entered the Holocene with low genetic

diversity. Given that tropical island endemics have small geographic ranges and are groups

vulnerable to climate change, special efforts are necessary to conserve them. We recommend that

conservation management policies add components like historical demography and species traits

while assessing extinction threats for island populations.

KEYWORDS: Climate change, Effective population size, Genetic diversity, Tropical island

endemic birds, Demographic history, Species Distribution Modelling, PSMC

INTRODUCTION

Tropical islands of the Indo-Australian Archipelago (IAA), the Indo-Pacific, and the Caribbean and Atlantic have had complex geological pasts that have affected their species distribution patterns. While several Pacific islands are true oceanic islands arising *de novo* from the seafloor, many islands of the Caribbean and the IAA were once connected with each other or to the continental landmasses during periods of global sea-level fall in the Quaternary (Lohman et al., 2011; Voris, 2000). These land bridges and the additional habitat they offered during periods of sea-level fall facilitated on one hand colonisations among islands and between islands and the mainland (Andersen et al., 2015; Cros et al., 2020; Irestedt et al., 2013; Moyle et al., 2009; Ng et al., 2017; Pujolar et al., 2022) and on the other, range expansions within islands in response to fluctuations in suitable habitat space. The habitat type and quality of these intermittently exposed land bridges varied, ranging from open savannah biomes to lowland forests (Cannon et al., 2009; Lohman et al., 2011). In the case of stratified elevational gradients, with tropical upland montane forests expanding onto lower slopes during periods of global cooling, upland montane species could disperse across otherwise persistent dual barriers of land and sea (Cannon et al., 2009).

Tropical islands' geologic pasts make them an important natural system to study species responses to changing habitat at both ecological and evolutionary timescales. Birds are an ideal clade for this because they are taxonomically well-characterised, are ecologically well-studied, and charismatic. Avian lineages have arisen on tropical islands during the Pleistocene (Andersen et al., 2015; Garg et al., 2018; Irestedt et al., 2013; Moyle et al., 2009), concomitant with complex, clade-wise colonisation and recolonisation trajectories (Filardi and Moyle, 2005; Jønsson et al., 2008, 2011b, 2014) including colonisations of the mainland (Jønsson et al., 2011a)

and repeated, independent colonisation events (Cibois et al., 2011). In birds, these responses to

changing habitat are governed by species traits in several continental taxa and non single-island

endemics (Brüniche-Olsen et al., 2021, 2019), but responses in single-island endemics are poorly

understood.

With such complex biogeographic pasts, tropical, single-island endemics represent either refugial

populations, or *in-situ* diversifications as explained by taxon cycles (Ricklefs, 1970; Ricklefs and

Cox, 1978, 1972). Being confined to single islands, these species are exceptionally vulnerable to

ongoing anthropogenic climate change which is unprecedented and unlike past Pleistocene and

early Holocene climatic changes (Crowley, 1990). This is exacerbated because we are currently

experiencing a period of relatively high sea levels with reduced habitat availability for species

resulting in population bottlenecks across taxa (Hewitt, 2000; Willis et al., 2004) including birds

(Nadachowska-Brzyska et al., 2015; Smith et al., 2021). Information on tropical, single-island

endemics' demographic responses to past climate change can inform conservation efforts, owing

to the genomic signatures that predispose a species to extinction (Mays et al., 2018; Spielman et

al., 2004).

Pairwise Sequential Markovian Coalescent (PSMC) is a powerful method to reconstruct the

effective population size (Ne) for a species using a single diploid genome (Li and Durbin, 2011)

and has been used to infer demographic history across taxa (Chattopadhyay et al., 2019; Kim et

al., 2016; Kozma et al., 2016; Murray et al., 2017; Nadachowska-Brzyska et al., 2015). Using

this along with paleo-ecological niche modelling allows us to directly correlate the demographic

history of a species with its distributional range at different time points (Chattopadhyay et al.,

2019). We perform these analyses on a global panel of tropical single-island endemics to

understand the effects of past climatic changes on species with assumed low genetic diversity

owing to their status as refugial species or newly arisen lineages.

RESULTS

Bird species panel

Our final species panel comprises 30 tropical single-island endemic bird species. Out of these,

PSMC analyses were possible for 22 species and Ecological Niche Modelling (ENM) was

possible for 28 species. Both PSMC and ENM analyses could be done successfully for 20

species (Table 1). These included seven Papuan species, five Philippine species, three Caribbean

species, and three species from the Bismarck Archipelago. Of these 20 species, 13 species are

passerines, with two from the white-eye family (Zosteropidae). Of the non-passerines, two

species are parrots (family Psittacidae). All other families are represented by single species.

Paleo-habitat reconstruction

Among the 19 climatic variables used, we observed precipitation of the warmest quarter (BIO18)

to contribute the most for each island endemic bird species except for the Caribbean species

Amazona guildingii. 24 out of the 28 bird species experienced an increase in suitable habitat

from the Last Interglacial (LIG) to the Last Glacial Maximum (LGM) (Supplementary table S4).

11 out of 13 Papuan species showed an increase in habitat availability from the LIG to the LGM,

with *Cicinnurus regius* and *Pseudorectus ferrugineus* as exceptions. All Philippine species

showed an increase in habitat from the LIG to the LGM as well (Figure 1, Supplementary

information S2). Two out of the three Caribbean species experienced a decline in suitable habitat

during this period. Of the remaining species, all showed an increase in available habitat. Upland

Papuan species remained confined to upland montane regions at all the time points for which we

reconstructed available habitats, with *Rhagologus leucostigma* as a notable exception

(Supplementary information S2). Papuan lowland species also remained largely confined to

regions which were not newly exposed land bridges . From the LGM to the present day, habitat

decreased for 19 out of the 28 bird species, and two species experienced no change

(Supplementary table S3). For the present study, we primarily concentrated on habitat fluctuation

between the LIG and the LGM as this is the period for which we could obtain comparative

evidence of fluctuations of both habitat as well as genetic diversity.

Strong Quaternary fluctuations in genetic diversity

PSMC analyses could be successfully performed for 22 species (Table 1). Results indicate large

genetic diversity variations in almost all bird species, corresponding to Quaternary climate shifts

(Figure 2). The demographic history of these species reconstructed using Pairwise Sequentially

Markovian Coalescent (PSMC) analyses extends back over a million years (Supplementary

information S1). Reconstructed Ne values were generally concordant across the three PSMC

settings used, except for five species where Ne values had large differences across PSMC

settings for Ne values at the LIG and LGM. However, in these species as well, trends of Ne

increase or decrease from the LIG to LGM were robust across all three PSMC models

considered. *Centropus unirufus, Dicaeum eximium, Irena cyanogastra, Sterrhoptilus*

dennistouni, Zosterops hypoxanthus were the five species which had large differences

(Supplementary information S1). We found no significant difference between the different sets

of PSMC settings used (Kruskal-Wallis test, χ^2 = 0.217, DF = 2, p = 0.897).

The 14 passerine species we analysed showed overall lower Ne values in the LGM as compared

to the LIG (Figure 1). We obtained similar results for non-passerines as well, with *Rhynochetos*

jubatus as a notable exception using the –p "4 + 30 * 2 + 4 + 6 + 10" setting, displaying a large

peak followed by a crash in Ne (Supplementary information S1). This is likely a PSMC artefact,

because plots using other parameter settings did not show a peak (Supplementary information

S1). Along with overall lower values, we also see a smaller range of Ne values in the LGM as

compared to the LIG (Figure 1).

Correlation between historical fluctuations in Ne and distribution

Habitat change was poorly associated with change in Ne for the 20 species for which both PSMC

and ENM analyses were possible (Cramer's V = 0.15). However, passerine species only showed

a strong association (Cramer's V = 0.96), while non-passerines showed a weak negative

association (Cramer's V = –0.15).

Bayesian multivariate regression models revealed that an overall fluctuation in Ne was

associated with species biology and habitat change during the Last Glacial Period (LGP) even

after controlling for geographical island group (Figure 3). Most confidence intervals and beta

coefficients did not overlap with zero (Figure 3) suggesting significant relationships. We found a

positive association between change in habitat area (LIG to LGM) and Ne (β = 9.45, 95% CI:

[1.95, 21.1]). This means that species with increasing habitat area in the LGP also showed an

increase in Ne. Change in Ne and body mass showed a significant negative association ($\beta =$

-8.63, 95% CI: [16.27, –2.61]). That is, large-bodied species showed decreases in Ne during the

LGP. Diet specialists like frugivores ($\beta = -11.89$, 95% CI: [–22.65, –2.99]) and invertivores ($\beta =$

-11.7, 95% CI: [–22.66, –2.94]) also showed significant negative associations with population

change. However, omnivores ($\beta = -1.39$, 95% CI: [–13.38, 11.5]) did not show any association

with population change. Whether or not a species was a passerine was an important predictor of

Ne only in combination with the change in habitat from LIG to LGM ($\beta = 7.97$, 95% CI: [1.82,

16.91], suggesting that passerines respond positively to habitat change, as suggested by Cramer's

V as well. The interaction between habitat change and body mass ($\beta = 10.05$, 95% CI: [–0.3,

24.41]) suggests that there is no impact of habitat change in larger species. Finally, the random

intercept for Country (sd (Intercept)) showed a marginal positive influence ($\beta = 0.85$, 95% CI:

[0.04, 2.24]) (Figure 3, Supplementary table S6).

DISCUSSION

Island birds are vulnerable to the effects of climate change

Our results on the fluctuations in paleohabitat and genetic diversity (Ne) during the LGP

highlight the vulnerability of island birds to climatic fluctuations. Climate fluctuations change

habitat availability and, in turn, genetic diversity, making these species particularly sensitive to a

changing environment. In response to landmass expansion, several species in our panel across

islands showed an increase in their genetic diversity during the LGP (Figure 1, Supplementary

information S1), although the overall relationship is weak (Cramer's V = 0.15). This could be

because large amounts of intermittently exposed landbridge habitat remain uninhabited by the

species in our panel (Supplementary figure S2). For example, we know that large amounts of

exposed landbridge habitat available to our panel's Papuan species ranged from lowland

evergreen to seasonal rainforest, with an expansion of upland habitats of evergreen to seasonal

rainforest. Transitional hill forest formed an ecotone between these two (Cannon et al., 2009).

However, most of our species inhabit dense forest (Supplementary table S7), which is unlikely to

comprise newly exposed landbridge habitat.

Our results also reveal that both passerine and non-passerine island endemics have entered the

Holocene with low genetic diversity (Figure 1). A loss of genetic diversity predisposes species to

extinction (Mays et al., 2018; Spielman et al., 2004) with a potentially causal relationship for

birds (Evans and Sheldon, 2008). The current climate crisis has greatly imperilled biodiversity

with the documented loss of many vertebrate species (Steadman and Martin, 2003; Tan et al.,

2023; Willis et al., 2004). Fossil evidence has revealed the extinction of several species in the

Caribbean since the LGM (Morgan and Woods, 1986; Orihuela et al., 2020; Steadman et al.,

1984), and many of the species in our panel are Southeast Asian where rapid habitat loss is

ongoing (Sodhi et al., 2004). Climate warming, sea level rise, and changes in vegetation are all

associated with this loss, and flightless birds and endemics are particularly prone to extinction

(Fromm and Meiri, 2021).

Habitat loss, rise in sea level, and warming temperatures can rapidly accelerate species

extinction, particularly in the tropics (Şekercioğlu et al., 2012). Effective population size, and in

extension genetic diversity are associated with species survival and extinction risk (Frankham,

2005). Coalescent effective population size as used in PSMC is an important predictor of species

vulnerability to extinction (Brüniche-Olsen et al., 2021; Wilder et al., 2023). Thus, an evolution-

informed understanding of species vulnerability to climate change and their associations with

paleohabitats can be an important tool to predict species vulnerability to the current climate crisis

and future extinction risk (Brüniche-Olsen et al., 2021; Chattopadhyay et al., 2019; Gabrielli et

al., 2024; Germain et al., 2023).

A strong passerine association might be driven by their rapid diversification

Passerines are a hyperdiverse clade representing over 60% of extant avian diversity. The most

recent, fossil-calibrated passerine phylogeny (Oliveros et al., 2019) shows that passerines began

diversifying in the Middle (47 mya) to Late Eocene (38–39 mya) with increasing diversification

rates suggested as we move towards the present. Crown passerines originated in the Australo-

Pacific region (Oliveros et al., 2019), and this places the many Papuan and Australian species in

our panel close to the passerine diversification centre which is known to have rapid

diversification rates (McCullough et al., 2022). Because bird lineages are known to have arisen

on tropical islands in the Pleistocene (Andersen et al., 2015; Irestedt et al., 2013; Moyle et al.,

2009), it is likely that the Southeast Asian species in our panel represent newly arisen lineages

over refugia, possibly belonging to currently rapidly-diversifying clades. This is supported by the

fact that several of these species belong to small, often oligotypic families such as Oreoicidae

(*Aleadryas rufinucha*), Ptilonorhynchidae (*Amblyornis subalaris*), Cnemophilidae (*Cnemophilus*

loriae), Eulacestomatidae (*Eulacestoma nigropectus*), Ifritidae (*Ifrita kowaldi*),

Machaerirhynchidae (*Machaerirhynchus nigripectus*), and Rhagologidae (*Rhagologus*

leucostigma). Many of these species belong to poorly-studied tropical genera for which species-

level divergence times are unavailable. The strong passerine association between change in Ne

and available habitat area (Cramer's V = 0.96; figure 3, β = 9.45) reveals that passerines are

strongly plastic to the environment, most possibly as a consequence of their ability to rapidly

diversify.

Non-passerines representing a paraphyletic outgroup do not show this trend (Cramer's V = –

0.15). The seven non-passerine species for which both ENM and PSMC analyses were possible

belong to six different families (Table 1), representing a diverse sample across the avian

phylogeny and precluding non-passerine phylogenetic insight. Notably, we confirm (Cramer's V

= 0.15) that habitat area is positively correlated with genomic diversity in a clade non-specific

manner for birds in general (Brüniche-Olsen et al., 2021, 2019).

Species traits influenced historic fluctuations in Ne during the LGP

The interplay of species traits and habitat availability determine how Ne values fluctuate with

time in non-endemic birds (Brüniche-Olsen et al., 2021), and our results confirm this for tropical

island endemics as well. Habitat change in the LGP was positively associated with Ne

fluctuations (Figure 3, β = 9.45), that is, species which showed an increase in habitat in the LGP

also showed a concurrent increase in Ne. While diet generalists like omnivores showed no

significant association with Ne fluctuations, specialists like frugivores (Figure 3, β = –11.89) and

invertivores (Figure 3, β = –11.7) showed significant negative associations with fluctuation in

Ne, implying that specialist species tended to show decreases in Ne in the LGP. Previous work

has shown that large-bodied species have lower values of standing genetic diversity at a given

time-point (Brüniche-Olsen et al., 2021, 2019; Eo et al., 2011). We find large-bodied species to

show a negative association with Ne fluctuations (Figure 3, β = −8.63) as well. This implies that

Ne for large-bodied species have decreased in the LGP. Traits like body size and diet determine

extinction risk (Ripple et al., 2017; Willis et al., 2004), and our results add to the understanding

of how traits modulate Ne changes through time.

Assessing the relative contribution of species traits and habitat availability in determining Ne

requires their quantitative measurements. Our methods generated values for available habitat at

various time-points (Supplementary table S3). However, Ne values can so far only be

reconstructed using coalescent methods like PSMC. These values depend upon the parameter

settings used. While we find these to be generally concordant across settings (Kruskal-Wallis, p

= 0.897, Supplementary table S1), the precise, estimated values vary. We therefore chose to

cautiously measure only the direction of change (increase or decrease) and not use numerical

values.

Caveats of PSMC analyses

Coalescent methods for inferring demographic history like PSMC can trace a local population's

demographic history rather than the entire species' history (Gattepaille et al., 2013; Heller et al.,

2013; Ptak and Przeworski, 2002). Population structure is thus a confounding factor. However,

this is unlikely to be a problem given that all our species are single-island endemics. Large peaks

followed by apparent collapses in Ne have also recently been shown to be an artefact of PSMC

analyses (Hilgers et al., 2024). We try three different parameter combinations, and look for

consensus amongst them in order to try and mitigate this. Because reconstructed Ne values are

inferred based on these parameter settings, we do not use them directly for analyses, instead

looking at the direction of population size change (increase/decrease/no change). Lastly, we

emphasise that Ne is a scaled value of the real population size (Wang et al., 2016), and is

measured to be up to one-tenth of the real population size in wild populations (Frankham, 1995).

We emphasise caution in interpreting Ne results as a proxy for real population size.

METHODS

Species selection

We queried Avibase (Lepage et al., 2014) for all species endemic to tropical islands and selected

all species for which assembled whole genome sequences were available on GenBank using

short-read data from the Illumina platform. We only selected species that had not been analysed

using similar methods already. We excluded genomes assembled from museum specimens. This

resulted in a panel of 30 species (Table 1).

Pairwise Sequential Markovian Coalescent (PSMC) analyses

For each assembled diploid genome, we identified the contigs corresponding to the sex

chromosomes using Satsuma ver. 2.0 (Grabherr et al., 2010) by aligning contigs to a *Gallus*

gallus (chicken; BioSample: SAMN02981218) genome as a reference. All contigs mapping to

the sex chromosomes were removed. Next, we obtained all available Illumina short reads for

each species from the Sequence Read Archive (SRA) database of the NCBI, and checked them

for errors using FastQC (Andrews, 2010). We used Trimmomatic (Bolger et al., 2014) for

preprocessing and trimming the reads. The cleaned reads were aligned to the autosomes using

BWA-MEM ver. 0.7.17 (Li, 2013). Next, we used SAMtools ver. 1.10 (Danecek et al., 2021) to

merge all the resulting files to generate a single bam file for each species. We further sorted and

removed duplicates using SAMtools and estimated the depth of coverage for each species. We

excluded species with a depth of coverage < 18x. This is a sufficient value of coverage for

PSMC analyses (Li and Durbin, 2011). Next, we implemented the SAMtools mpileup-bcftools

pipeline to identify SNPs. The minimum and maximum depths for calling SNPs was set to one-

third and double the mean depth respectively following Nadachowska-Brzyska et al.

(Nadachowska-Brzyska et al., 2015).

We used three sets of parameters for PSMC analyses and looked for consensus trends amongst

them because PSMC analyses can result in spurious peaks of effective population size (Hilgers et

al., 2024). We used the following parameter sets for PSMC analyses: –t5 –b –r1 –p "4 + 30 * 2

+ 4 + 6 + 10", –t5 –b –r1 –p "2 + 2 + 30 * 2 + 4 + 6 + 10", and –t5 –b –r1 –p "1 + 1 + 1 + 1 + 30

* 2 + 4 + 6 + 10". We performed 30 iterations for parameter optimisation and ran 100 bootstrap

replicates to judge the uncertainty in our estimates. Bootstrapping was done by randomly

sampling chromosome segments with replacement and running PSMC on them. To plot the

results from the PSMC analyses, we used previously estimated values of mutation rates and

generation times for passerines (1.4 x 10e–9 years/site and 2 years respectively; (Ellegren et al.,

2012; Nadachowska-Brzyska et al., 2016) and non-passerines (1.91 x 10e–9 years/site

and 1 year respectively) (Nam et al., 2010) (Supplementary material S1, supplementary table

S1).

Species and climate records

We performed reconstructions of paleohabitats through ecological niche models and

reconstructed species distributions from four time periods: the Last Interglacial (LIG, approx.

110,000–130,000 years ago), the Last Glacial Maximum (LGM, approx. 20,000 years ago), the

Mid-Holocene (MDH, approx. 6000 years ago) and Current (CUR, present day).

We used R (ver. 4.2.1; R Core Team, 2021) for all paleo-habitat modelling analyses. We

accessed the location records of 20 bird species from the Global Biodiversity Information

Facility (GBIF) in September, 2023 using the "rbgif" package (ver. 3.7.9) (Chamberlain and

Boettiger, 2017) in R. We discarded duplicates and retained only human observed records using

the "tidyverse" (v-2.0.0) package (Wickham and RStudio, 2023) followed by the

'CoordinateCleaner' package (v-3.0.1) (Zizka et al., 2023). We used the "spThin" package

(Aiello-Lammens et al., 2014) to account for spatial autocorrelation and finally discarded

spurious distribution records like records overlaying water bodies, buildings, roadways, and

railways using QGIS (v-3.34+; https://www.qgis.org/). GBIF dataset keys used to access all the

data points are in Supplementary Table S8.

We used 19 climate variables (labelled BIO1 – BIO19; Supplementary table S2) for the four

different time periods considered. We downloaded CHELSA (https://chelsa-climate.org/; Karger

et al., 2021) climate predictors at a resolution of 2.5 arc-minutes (~5 km) for all regions except

for *Amazona guildingii* from Saint Vincent where we used a resolution of 30 arc-seconds

(~1km). For the latter, we used WORLDCLIM (https://www.worldclim.org/; Fick and Hijmans,

2017) for data for the LIG and MDH while CHELSA was used for the LGM and CUR.

We followed Chattopadhyay et al., (2019) and used a global dataset approach to account for

idiosyncratic biases due to smaller datasets to extract the location points of each endemic-island

group against each climatic variable. We tested for multicollinearity using variance inflation

factor (VIF) using the 'usdm' R package (Naimi, 2014) and considered variables for further

analyses if their VIF value was ≤ 5 (Shrestha, 2020).

Habitat Suitability Modelling and Area Calculation

We used R to reconstruct paleo-climatic suitable habitat for 28 endemic island birds through

ecological niche models for the four time-periods considered. For this, we accessed the Global

Biodiversity Information Facility (GBIF) to extract our species' occurrence records. We could not

perform ENM analyses for two species due to a paucity in the number of GBIF occurrence points

for them (Table 1).

Data partitioning and model evaluation

For each species, we used available occurrence records from the present day to generate pseudo-

absence data points. We generated 500–10,000 pseudo-absence/background points for each

species depending upon the area of the island (Supplementary table S3), except for *Amazona*

guildingii endemic to Saint Vincent where we generated 40 points. Saint Vincent's small land

area could not accommodate more points than this. We used a subset of bioclimatic variables to

generate a weighted average ensemble species distribution model (eSDM) for each island group.

eSDMs were implemented in the R package "sdm" (v-1.2.37; Naimi and Araujo, 2016) applying

the 'MaxEnt', 'GLM', and 'BRT' algorithms. eSDMs account for the limitations of different

models by generating a weighted average of multiple models (Araújo and New, 2007; Dormann

et al., 2018; Naimi and Araujo, 2016). We used k-fold cross-validation (CV) with replication for

each method for training and test datasets across each endemic island group (Supplementary

table S3). Model accuracy was measured using the Area Under the Curve (AUC) and True Skill

Statistic (TSS) metrics. We first performed the above analyses for the present-day (CUR)

distribution using the weighted-average of AUC and TSS. AUC and TSS values greater than 0.9

and 0.75 respectively have been shown to be indicators of superior model performance (Ahmad

et al., 2019) and we chose a similar threshold (AUC ≥ 0.9 & TSS ≥ 0.8) to get the best ensemble

model for most of the species except for 10 species for which data quality was poor

(Supplementary table S3). Models for the other time points: the LIG, LGM, and MDH, were

generated using the eSDM model generated based on CUR data.

Suitable area analysis

All spatial analyses were carried out using the R package 'terra' (ver. 1.7.13) (Hijmans et al.,

2024). Equal area projections were used to calculate the absolute suitable area of each species

across the four time-periods. To account for a spherical Earth and the limitations of landmass

depiction of the original eSDM raster (World Geodetic System 1984) we reprojected it into a

pseudocylindrical projection (Eckert IV). Further, we used the average quantile threshold pixels

to measure the availability of absolute suitable areas for the LIG, LGM, MDH, and CUR periods

(Supplementary table S4).

Statistical Analyses

We checked for associations between Ne and habitat change during the Last Glacial Period (from

LIG to LGM) (LGP) using Cramer's V implemented in the"polycor" package in R (Fox, 2022).

Cramver's V varies between -1 to 1, with values closer to 0 suggesting no-correlation. This was

done for the entire dataset, and then for passerines and non-passerines separately.

We further explored the effects of species biology, habitat, and phylogenetic constraint on the

fluctuation in Ne during the LGP using Bayesian Multilevel Models (MLMs), using the "brms"

package in R (Bürkner, 2018). Our fixed effect predictors were habitat change from the LIG to

LGM, diet (invertivore, frugivore, or omnivore), body mass, and clade identity (passerine and

non-passerine). The latter allows us to check if belonging to the rapidly diversifying passerine

clade results in a signal. We also included country (the island/archipelago a species is endemic

to) as a random effect variable for the analysis to account for country-specific effects. For the

response variable i.e., the change in Ne, a Bernoulli distribution with a logit link was used

because it is a binary response variable.

We ran a series of models (n=12) to find the best-fit model (brms11, Supplementary table S6)

based on leave-one-out cross-validation. We generated a total of 16,000 posterior samples by

using the No-U-Turn Sampler (Hamiltonian Monte Carlo) algorithm with 4 chains with 5000

iterations each (with 1000 warm-up iterations; supplementary information S3). Further, we

estimated the posterior parameters with 95% confidence intervals to find negative or positive

associations between the Ne and its predictors. In addition, we assessed the posterior

convergence of the sample through the R-hat statistic ($R^\wedge = 1$) where values close to 1 suggest an

ideal convergence. We also reported the effective sample size for bulk and tail distributions

(Supplementary table S5).

ACKNOWLEDGEMENTS

BC acknowledges the support from the Trivedi School of Biosciences. KMG acknowledges the

support from the DBT-Ramalingaswami Fellowship (No. BT/HRD/35/02/2006). VI is supported

by DBT grant (BT/PR42830/BRB/10/1995/2021) to KMG.

DATA AVAILABILITY

The raw data used for this study are available on NCBI and details of the accession IDs are

provided in Table S1. All codes used in this study are provided on the Zenodo URL

https://doi.org/10.5281/zenodo.14603966 .

REFERENCES

Ahmad, R., Khuroo, A.A., Charles, B., Hamid, M., Rashid, I., Aravind, N.A., 2019. Global

distribution modelling, invasion risk assessment and niche dynamics of Leucanthemum vulgare

(Ox-eye Daisy) under climate change. Sci. Rep. 9, 11395. https://doi.org/10.1038/s41598-019-

47859-1

Aiello-Lammens, M.E., Boria, R.A., Radosavljevic, A., Vilela, B., Anderson, R.P., 2014.

spThin: Functions for Spatial Thinning of Species Occurrence Records for Use in Ecological

Models. https://doi.org/10.32614/CRAN.package.spThin

Andersen, M.J., Shult, H.T., Cibois, A., Thibault, J.-C., Filardi, C.E., Moyle, R.G., 2015. Rapid

diversification and secondary sympatry in Australo-Pacific kingfishers (Aves: Alcedinidae:

Todiramphus). R. Soc. Open Sci. 2, 140375. https://doi.org/10.1098/rsos.140375

Andrews, S., 2010. FastQC: A Quality Control Tool for High Throughput Sequence Data.

Araújo, M.B., New, M., 2007. Ensemble forecasting of species distributions. Trends Ecol. Evol.

22, 42–47. https://doi.org/10.1016/j.tree.2006.09.010

Bolger, A.M., Lohse, M., Usadel, B., 2014. Trimmomatic: a flexible trimmer for Illumina

sequence data. Bioinformatics 30, 2114–2120. https://doi.org/10.1093/bioinformatics/btu170

Brüniche-Olsen, A., Kellner, K.F., Belant, J.L., DeWoody, J.A., 2021. Life-history traits and

habitat availability shape genomic diversity in birds: implications for conservation. Proc. R. Soc.

B Biol. Sci. 288, 20211441. https://doi.org/10.1098/rspb.2021.1441

Brüniche-Olsen, A., Kellner, K.F., DeWoody, J.A., 2019. Island area, body size and

demographic history shape genomic diversity in Darwin's finches and related tanagers. Mol.

Ecol. 28, 4914–4925. https://doi.org/10.1111/mec.15266

Bürkner, P.-C., 2018. Advanced Bayesian Multilevel Modeling with the R Package brms. R J.

10, 395–411.

Cannon, C.H., Morley, R.J., Bush, A.B.G., 2009. The current refugial rainforests of Sundaland

are unrepresentative of their biogeographic past and highly vulnerable to disturbance. Proc. Natl.

Acad. Sci. 106, 11188–11193. https://doi.org/10.1073/pnas.0809865106

Chamberlain, S., Boettiger, C., 2017. R Python, and Ruby clients for GBIF species occurrence

data. PeerJ Prepr.

Chattopadhyay, B., Garg, K.M., Ray, R., Rheindt, F.E., 2019. Fluctuating fortunes: genomes and

habitat reconstructions reveal global climate-mediated changes in bats' genetic diversity. Proc.

R. Soc. B Biol. Sci. 286, 20190304. https://doi.org/10.1098/rspb.2019.0304

Cibois, A., Beadell, J.S., Graves, G.R., Pasquet, E., Slikas, B., Sonsthagen, S.A., Thibault, J.-C.,

Fleischer, R.C., 2011. Charting the course of reed-warblers across the Pacific islands. J.

Biogeogr. 38, 1963–1975. https://doi.org/10.1111/j.1365-2699.2011.02542.x

Cros, E., Chattopadhyay, B., Garg, K.M., Ng, N.S.R., Tomassi, S., Benedick, S., Edwards, D.P.,

Rheindt, F.E., 2020. Quaternary land bridges have not been universal conduits of gene flow.

Mol. Ecol. 29, 2692–2706. https://doi.org/10.1111/mec.15509

Crowley, T.J., 1990. Are There Any Satisfactory Geologic Analogs for a Future Greenhouse

Warming?

Danecek, P., Bonfield, J.K., Liddle, J., Marshall, J., Ohan, V., Pollard, M.O., Whitwham, A.,

Keane, T., McCarthy, S.A., Davies, R.M., Li, H., 2021. Twelve years of SAMtools and

BCFtools. GigaScience 10, giab008. https://doi.org/10.1093/gigascience/giab008

Dormann, C.F., Calabrese, J.M., Guillera-Arroita, G., Matechou, E., Bahn, V., Bartoń, K., Beale,

C.M., Ciuti, S., Elith, J., Gerstner, K., Guelat, J., Keil, P., Lahoz-Monfort, J.J., Pollock, L.J.,

Reineking, B., Roberts, D.R., Schröder, B., Thuiller, W., Warton, D.I., Wintle, B.A., Wood,

S.N., Wüest, R.O., Hartig, F., 2018. Model averaging in ecology: a review of Bayesian,

information-theoretic, and tactical approaches for predictive inference. Ecol. Monogr. 88, 485–

504. https://doi.org/10.1002/ecm.1309

Ellegren, H., Smeds, L., Burri, R., Olason, P.I., Backström, N., Kawakami, T., Künstner, A.,

Mäkinen, H., Nadachowska-Brzyska, K., Qvarnström, A., Uebbing, S., Wolf, J.B.W., 2012. The

genomic landscape of species divergence in Ficedula flycatchers. Nature 491, 756–760.

https://doi.org/10.1038/nature11584

Eo, S.H., Doyle, J.M., DeWoody, J.A., 2011. Genetic diversity in birds is associated with body

mass and habitat type. J. Zool. 283, 220–226. https://doi.org/10.1111/j.1469-7998.2010.00773.x

Evans, S.R., Sheldon, B.C., 2008. Interspecific Patterns of Genetic Diversity in Birds:

Correlations with Extinction Risk. Conserv. Biol. 22, 1016–1025. https://doi.org/10.1111/j.1523-

1739.2008.00972.x

Fick, S.E., Hijmans, R.J., 2017. WorldClim 2: new 1-km spatial resolution climate surfaces for

global land areas. Int. J. Climatol. 37, 4302–4315. https://doi.org/10.1002/joc.5086

Filardi, C.E., Moyle, R.G., 2005. Single origin of a pan-Pacific bird group and upstream

colonization of Australasia. Nature 438, 216–219. https://doi.org/10.1038/nature04057

Fox, J., 2022. polycor: Polychoric and Polyserial Correlations.

Frankham, R., 2005. Genetics and extinction. Biol. Conserv. 126, 131–140.

https://doi.org/10.1016/j.biocon.2005.05.002

Frankham, R., 1995. Effective population size/adult population size ratios in wildlife: a review.

Genet. Res. 66, 95–107. https://doi.org/10.1017/S0016672300034455

Fromm, A., Meiri, S., 2021. Big, flightless, insular and dead: Characterising the extinct birds of

the Quaternary. J. Biogeogr. 48, 2350–2359. https://doi.org/10.1111/jbi.14206

Gabrielli, M., Leroy, T., Salmona, J., Nabholz, B., Milá, B., Thébaud, C., 2024. Demographic

responses of oceanic island birds to local and regional ecological disruptions revealed by whole-

genome sequencing. Mol. Ecol. 33, e17243. https://doi.org/10.1111/mec.17243

Garg, K.M., Chattopadhyay, B., Wilton, P.R., Malia Prawiradilaga, D., Rheindt, F.E., 2018.

Pleistocene land bridges act as semipermeable agents of avian gene flow in Wallacea. Mol.

Phylogenet. Evol. 125, 196–203. https://doi.org/10.1016/j.ympev.2018.03.032

Gattepaille, L.M., Jakobsson, M., Blum, M.G., 2013. Inferring population size changes with

sequence and SNP data: lessons from human bottlenecks. Heredity 110, 409–419.

https://doi.org/10.1038/hdy.2012.120

Germain, R.R., Feng, S., Chen, G., Graves, G.R., Tobias, J.A., Rahbek, C., Lei, F., Fjeldså, J.,

Hosner, P.A., Gilbert, M.T.P., Zhang, G., Nogués-Bravo, D., 2023. Species-specific traits

mediate avian demographic responses under past climate change. Nat. Ecol. Evol. 7, 862–872.

https://doi.org/10.1038/s41559-023-02055-3

Grabherr, M.G., Russell, P., Meyer, M., Mauceli, E., Alföldi, J., Di Palma, F., Lindblad-Toh, K.,

2010. Genome-wide synteny through highly sensitive sequence alignment: Satsuma.

Bioinformatics 26, 1145–1151. https://doi.org/10.1093/bioinformatics/btq102

Heller, R., Chikhi, L., Siegismund, H.R., 2013. The Confounding Effect of Population Structure

on Bayesian Skyline Plot Inferences of Demographic History. PLOS ONE 8, e62992.

https://doi.org/10.1371/journal.pone.0062992

Hewitt, G., 2000. The genetic legacy of the Quaternary ice ages. Nature 405, 907–913.

https://doi.org/10.1038/35016000

Hijmans, R.J., Bivand, R., Dyba, K., Pebesma, E., Sumner, M.D., 2024. terra: Spatial Data

Analysis.

Hilgers, L., Liu, S., Jensen, A., Brown, T., Cousins, T., Schweiger, R., Guschanski, K., Hiller,

M., 2024. Avoidable false PSMC population size peaks occur across numerous studies.

https://doi.org/10.1101/2024.06.17.599025

Irestedt, M., Fabre, P.-H., Batalha-Filho, H., Jønsson, K.A., Roselaar, C.S., Sangster, G.,

Ericson, P.G.P., 2013. The spatio-temporal colonization and diversification across the Indo-

Pacific by a 'great speciator' (Aves, Erythropitta erythrogaster). Proc. R. Soc. B Biol. Sci. 280,

20130309. https://doi.org/10.1098/rspb.2013.0309

Jønsson, K.A., Fabre, P.-H., Ricklefs, R.E., Fjeldså, J., 2011a. Major global radiation of corvoid

birds originated in the proto-Papuan archipelago. Proc. Natl. Acad. Sci. 108, 2328–2333.

https://doi.org/10.1073/pnas.1018956108

Jønsson, K.A., Irestedt, M., Bowie, R.C.K., Christidis, L., Fjeldså, J., 2011b. Systematics and

biogeography of Indo-Pacific ground-doves. Mol. Phylogenet. Evol. 59, 538–543.

https://doi.org/10.1016/j.ympev.2011.01.007

Jønsson, K.A., Irestedt, M., Christidis, L., Clegg, S.M., Holt, B.G., Fjeldså, J., 2014. Evidence of

taxon cycles in an Indo-Pacific passerine bird radiation (Aves: Pachycephala). Proc. R. Soc. B

Biol. Sci. 281, 20131727. https://doi.org/10.1098/rspb.2013.1727

Jønsson, K.A., Irestedt, M., Fuchs, J., Ericson, P.G.P., Christidis, L., Bowie, R.C.K., Norman,

J.A., Pasquet, E., Fjeldså, J., 2008. Explosive avian radiations and multi-directional dispersal

across Wallacea: Evidence from the Campephagidae and other Crown Corvida (Aves). Mol.

Phylogenet. Evol. 47, 221–236. https://doi.org/10.1016/j.ympev.2008.01.017

Karger, D.N., Wilson, A.M., Mahony, C., Zimmermann, N.E., Jetz, W., 2021. Global daily 1 km

land surface precipitation based on cloud cover-informed downscaling. Sci. Data 8, 307.

https://doi.org/10.1038/s41597-021-01084-6

Kim, Soonok, Cho, Y.S., Kim, H.-M., Chung, O., Kim, H., Jho, S., Seomun, H., Kim, J., Bang,

W.Y., Kim, C., An, J., Bae, C.H., Bhak, Y., Jeon, S., Yoon, H., Kim, Y., Jun, J., Lee, HyeJin,

Cho, S., Uphyrkina, O., Kostyria, A., Goodrich, J., Miquelle, D., Roelke, M., Lewis, J.,

Yurchenko, A., Bankevich, A., Cho, J., Lee, S., Edwards, J.S., Weber, J.A., Cook, J., Kim,

Sangsoo, Lee, Hang, Manica, A., Lee, I., O'Brien, S.J., Bhak, J., Yeo, J.-H., 2016. Comparison

of carnivore, omnivore, and herbivore mammalian genomes with a new leopard assembly.

Genome Biol. 17, 211. https://doi.org/10.1186/s13059-016-1071-4

Kozma, R., Melsted, P., Magnússon, K.P., Höglund, J., 2016. Looking into the past – the

reaction of three grouse species to climate change over the last million years using whole

genome sequences. Mol. Ecol. 25, 570–580. https://doi.org/10.1111/mec.13496

Lepage, D., Vaidya, G., Guralnick, R., 2014. Avibase – a database system for managing and

organizing taxonomic concepts. ZooKeys 420, 117–135.

https://doi.org/10.3897/zookeys.420.7089

Li, H., 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM.

https://doi.org/10.48550/arXiv.1303.3997

Li, H., Durbin, R., 2011. Inference of human population history from individual whole-genome

sequences. Nature 475, 493–496. https://doi.org/10.1038/nature10231

Lohman, D.J., Bruyn, M. de, Page, T., Rintelen, K. von, Hall, R., Ng, P.K.L., Shih, H.-T.,

Carvalho, G.R., Rintelen, T. von, 2011. Biogeography of the Indo-Australian Archipelago.

Annu. Rev. Ecol. Evol. Syst. 42, 205–226. https://doi.org/10.1146/annurev-ecolsys-102710-

145001

Mays, H.L., Hung, C.-M., Shaner, P.-J., Denvir, J., Justice, M., Yang, S.-F., Roth, T.L., Oehler,

D.A., Fan, J., Rekulapally, S., Primerano, D.A., 2018. Genomic Analysis of Demographic

History and Ecological Niche Modeling in the Endangered Sumatran Rhinoceros *Dicerorhinus*

sumatrensis. Curr. Biol. 28, 70-76.e4. https://doi.org/10.1016/j.cub.2017.11.021

McCullough, J.M., Oliveros, C.H., Benz, B.W., Zenil-Ferguson, R., Cracraft, J., Moyle, R.G.,

Andersen, M.J., 2022. Wallacean and Melanesian Islands Promote Higher Rates of

Diversification within the Global Passerine Radiation Corvides. Syst. Biol. 71, 1423–1439.

https://doi.org/10.1093/sysbio/syac044

Morgan, G.S., Woods, C.A., 1986. Extinction and the zoogeography of West Indian land

mammals. Biol. J. Linn. Soc. 28, 167–203. https://doi.org/10.1111/j.1095-8312.1986.tb01753.x

Moyle, R.G., Filardi, C.E., Smith, C.E., Diamond, J., 2009. Explosive Pleistocene diversification

and hemispheric expansion of a "great speciator." Proc. Natl. Acad. Sci. 106, 1863–1868.

https://doi.org/10.1073/pnas.0809861105

Murray, G.G.R., Soares, A.E.R., Novak, B.J., Schaefer, N.K., Cahill, J.A., Baker, A.J.,

Demboski, J.R., Doll, A., Da Fonseca, R.R., Fulton, T.L., Gilbert, M.T.P., Heintzman, P.D.,

Letts, B., McIntosh, G., O'Connell, B.L., Peck, M., Pipes, M.-L., Rice, E.S., Santos, K.M.,

Sohrweide, A.G., Vohr, S.H., Corbett-Detig, R.B., Green, R.E., Shapiro, B., 2017. Natural

selection shaped the rise and fall of passenger pigeon genomic diversity. Science 358, 951–954.

https://doi.org/10.1126/science.aao0960

Nadachowska-Brzyska, K., Burri, R., Smeds, L., Ellegren, H., 2016. PSMC analysis of effective

population sizes in molecular ecology and its application to black-and-white Ficedula

flycatchers. Mol. Ecol. 25, 1058–1072. https://doi.org/10.1111/mec.13540

Nadachowska-Brzyska, K., Li, C., Smeds, L., Zhang, G., Ellegren, H., 2015. Temporal

Dynamics of Avian Populations during Pleistocene Revealed by Whole-Genome Sequences.

Curr. Biol. 25, 1375–1380. https://doi.org/10.1016/j.cub.2015.03.047

Naimi B, Hamm Na, Groen TA, Skidmore AK, Toxopeus AG (2014). Where is positional

uncertainty

a problem for species distribution modelling. *Ecography*, 37, 191-203.

doi:10.1111/j.1600-0587.2013.00205.x

Naimi, B., Araujo, M.B., 2016. sdm: a reproducible and extensible R platform for species

distribution modelling. Ecography 39, 368–375. https://doi.org/10.1111/ecog.01881

Nam, K., Mugal, C., Nabholz, B., Schielzeth, H., Wolf, J.B., Backström, N., Künstner, A.,

Balakrishnan, C.N., Heger, A., Ponting, C.P., Clayton, D.F., Ellegren, H., 2010. Molecular

evolution of genes in avian genomes. Genome Biol. 11, R68. https://doi.org/10.1186/gb-2010-

11-6-r68

Ng, N.S.R., Wilton, P.R., Prawiradilaga, D.M., Tay, Y.C., Indrawan, M., Garg, K.M., Rheindt,

F.E., 2017. The effects of Pleistocene climate change on biotic differentiation in a montane

songbird clade from Wallacea. Mol. Phylogenet. Evol. 114, 353–366.

https://doi.org/10.1016/j.ympev.2017.05.007

Oliveros, C.H., Field, D.J., Ksepka, D.T., Barker, F.K., Aleixo, A., Andersen, M.J., Alström, P.,

Benz, B.W., Braun, E.L., Braun, M.J., Bravo, G.A., Brumfield, R.T., Chesser, R.T., Claramunt,

S., Cracraft, J., Cuervo, A.M., Derryberry, E.P., Glenn, T.C., Harvey, M.G., Hosner, P.A.,

Joseph, L., Kimball, R.T., Mack, A.L., Miskelly, C.M., Peterson, A.T., Robbins, M.B., Sheldon,

F.H., Silveira, L.F., Smith, B.T., White, N.D., Moyle, R.G., Faircloth, B.C., 2019. Earth history

and the passerine superradiation. Proc. Natl. Acad. Sci. 116, 7916–7925.

https://doi.org/10.1073/pnas.1813206116

Orihuela, J., Viñola, L.W., Jiménez Vázquez, O., Mychajliw, A.M., Hernández de Lara, O.,

Lorenzo, L., Soto-Centeno, J.A., 2020. Assessing the role of humans in Greater Antillean land

vertebrate extinctions: New insights from Cuba. Quat. Sci. Rev. 249, 106597.

https://doi.org/10.1016/j.quascirev.2020.106597

Ptak, S.E., Przeworski, M., 2002. Evidence for population growth in humans is confounded by

fine-scale population structure. Trends Genet. 18, 559–563. https://doi.org/10.1016/S0168-

9525(02)02781-6

Pujolar, J.M., Blom, M.P.K., Reeve, A.H., Kennedy, J.D., Marki, P.Z., Korneliussen, T.S.,

Freeman, B.G., Sam, K., Linck, E., Haryoko, T., Iova, B., Koane, B., Maiah, G., Paul, L.,

Irestedt, M., Jønsson, K.A., 2022. The formation of avian montane diversity across barriers and

along elevational gradients. Nat. Commun. 13, 268. https://doi.org/10.1038/s41467-021-27858-5

R Core Team, 2021. R: A Language and Environment for Statistical Computing. R Foundation

for Statistical Computing, Vienna, Austria.

Ricklefs, R.E., 1970. Stage of Taxon Cycle and Distribution of Birds on Jamaica, Greater

Antilles. Evolution 24, 475–477. https://doi.org/10.2307/2406820

Ricklefs, R.E., Cox, G.W., 1978. Stage of Taxon Cycle, Habitat Distribution, and Population

Density in the Avifauna of the West Indies. Am. Nat. 112, 875–895.

https://doi.org/10.1086/283329

Ricklefs, R.E., Cox, G.W., 1972. Taxon Cycles in the West Indian Avifauna. Am. Nat. 106, 195–

219. https://doi.org/10.1086/282762

Ripple, W.J., Wolf, C., Newsome, T.M., Hoffmann, M., Wirsing, A.J., McCauley, D.J., 2017.

Extinction risk is most acute for the world's largest and smallest vertebrates. Proc. Natl. Acad.

Sci. 114, 10678–10683. https://doi.org/10.1073/pnas.1702078114

Şekercioğlu, Ç.H., Primack, R.B., Wormworth, J., 2012. The effects of climate change on

tropical birds. Biol. Conserv. 148, 1–18. https://doi.org/10.1016/j.biocon.2011.10.019

Smith, B.T., Gehara, M., Harvey, M.G., 2021. The demography of extinction in eastern North

American birds. Proc. R. Soc. B Biol. Sci. 288, 20201945.

https://doi.org/10.1098/rspb.2020.1945

Sodhi, N.S., Koh, L.P., Brook, B.W., Ng, P.K.L., 2004. Southeast Asian biodiversity: an

impending disaster. Trends Ecol. Evol. 19, 654–660. https://doi.org/10.1016/j.tree.2004.09.006

Spielman, D., Brook, B.W., Frankham, R., 2004. Most species are not driven to extinction before

genetic factors impact them. Proc. Natl. Acad. Sci. 101, 15261–15264.

https://doi.org/10.1073/pnas.0403809101

Shrestha, N., 2020. Detecting Multicollinearity in Regression Analysis. American Journal of

Applied Mathematics and Statistics 8, 39–42. https://doi.org/10.12691/ajams-8-2-1

Steadman, D.W., Martin, P.S., 2003. The late Quaternary extinction and future resurrection of

birds on Pacific islands. Earth-Sci. Rev. 61, 133–147. https://doi.org/10.1016/S0012-

8252(02)00116-2

Steadman, D.W., Pregill, G.K., Olson, S.L., 1984. Fossil vertebrates from Antigua, Lesser

Antilles: Evidence for late Holocene human-caused extinctions in the West Indies. Proc. Natl.

Acad. Sci. 81, 4448–4451. https://doi.org/10.1073/pnas.81.14.4448

Tan, H.Z., Jansen, J.J., Allport, G.A., Garg, K.M., Chattopadhyay, B., Irestedt, M., Pang, S.E.,

Chilton, G., Gwee, C.Y., Rheindt, F.E., 2023. Megafaunal extinctions, not climate change, may

explain Holocene genetic diversity declines in Numenius shorebirds. eLife 12, e85422.

https://doi.org/10.7554/eLife.85422

Voris, H.K., 2000. Maps of Pleistocene sea levels in Southeast Asia: shorelines, river systems

and time durations. J. Biogeogr. 27, 1153–1167. https://doi.org/10.1046/j.1365-

2699.2000.00489.x

Wang, J., Santiago, E., Caballero, A., 2016. Prediction and estimation of effective population

size. Heredity 117, 193–206. https://doi.org/10.1038/hdy.2016.43

Wickham, H., RStudio, 2023. tidyverse: Easily Install and Load the "Tidyverse."

Wilder, A.P., Supple, M.A., Subramanian, A., Mudide, A., Swofford, R., Serres-Armero, A.,

Steiner, C., Koepfli, K.-P., Genereux, D.P., Karlsson, E.K., Lindblad-Toh, K., Marques-Bonet,

T., Munoz Fuentes, V., Foley, K., Meyer, W.K., Zoonomia Consortium, Ryder, O.A., Shapiro,

B., 2023. The contribution of historical processes to contemporary extinction risk in placental

mammals. Science 380, eabn5856. https://doi.org/10.1126/science.abn5856

Willis, K.J., Bennett, K.D., Walker, D., Hewitt, G.M., 2004. Genetic consequences of climatic

oscillations in the Quaternary. Philos. Trans. R. Soc. Lond. B. Biol. Sci. 359, 183–195.

https://doi.org/10.1098/rstb.2003.1388

Zizka, A., Silvestro, D., Andermann, T., Azevedo, J., Ritter, C.D., Edler, D., Farooq, H.,

Herdean, A., Ariza, M., Scharn, R., Svanteson, S., Wengstrom, N., Zizka, V., Antonelli, A., sp,

B.V. (Bruno updated the package to remove dependencies on, raster, rgdal, maptools,

packages), and rgeos, ropensci, I.S. (Irene reviewed the package for, see

https://github.com/ropensci/onboarding/issues/210), ropensci, F.R.-S. (Francisco reviewed the

package for, see <https://github.com/ropensci/onboarding/issues/210>), 2023.

CoordinateCleaner: Automated Cleaning of Occurrence Records from Biological Collections.

     FIGURES

Figure 1: Pairwise Sequential Markovian Coalescent (PSMC) plots using the settings –p "2 + 2

+ 30 * 2 + 4 + 6 + 10" displaying reconstructed effective population size values with time for

passerines (above) and non-passerines (below) based on whether habitat availability increased

(left) or decreased (right) during the Last Glacial Period (LGP). Colours indicate the

archipelago/island the bird belongs to, and the line style indicates the dietary habit of the bird

species. Bold lines indicate large (> 50 g body mass) bird species. The grey bands indicate the

approximate durations of the Last Interglacial (LIG) and the Last Glacial Maximum (LGM).

Black arrows indicate if habitat availability increased or decreased during the LGP. A mutation

rate of 1.4 x 10e–9 years/site and a generation time of 2 years for passerines, and a mutation rate

of 1.91 x 10e–9 years/site and a generation time of 1 year for non-passerines were used to

785   generate plots. *Zosterops hypoxanthus* is not displayed because its Ne values far exceed those of

786   the other species.

Figure 2: Example Ecological Niche Modelling plots for species from Papua (*Cnemophilus loriae)*, Australia (*Alectura lathami*), the Philippines (*Irena cyanogastra*), and the Caribbean (Puerto Rico, *Todus mexicanus*). LIG = Last Interglacial. LGM = Last Glacial Maximum. MDH = Mid Holocene. CUR = Current. The continuous heatmap represents the probability of occurrence of the species and red points are known occurrences from GBIF. For all the plots see Figure S2.

Figure 3: Results of the best Bayesian multivariate regression model performed. The

response variable represents the changes in effective population size (increased (=1) or

decreased (=0)) during the Last Glacial Period. Circles represent mean values and lines

represent 95% confidence intervals. Model parameters and coefficients are provided in table

S6. ΔHabitat = the change in suitable habitat from the Last Interglacial to the Last Glacial

Maximum.

TABLES

Table 1: Details of the taxa included in this study. PSMC = Pairwise Sequential Markovian

Coalescent, ENM= Ecological Niche Modelling, y = yes, n = no.

Species	PSMC	ENM	Endemic Region	Passerine	Family
Actenoides hombroni	y	y	Philippines	n	Alcedinidae
Aleadryas rufinucha	y	y	Papua New Guinea	y	Oreoicidae
Alectura lathami	y	y	Australia	n	Megapodiidae
Amazona guildingii	y	y	St Vincent	n	Psittacidae
Amazona vittata	y	y	Puerto Rico	n	Psittacidae
Amblyornis subalaris	y	n	Papua New Guinea	y	Ptilonorhynchidae
Cacatua alba	y	n	Indonesia	n	Cacatuidae
Centropus unirufus	y	y	Philippines	n	Cuculidae
Chaetorhynchus papuensis	n	y	Papua New Guinea	y	Rhipiduridae
Cicinnurus regius	y	y	Papua New Guinea	y	Paradisaeidae
Cnemophilus loriae	y	y	Papua New	y	Cnemophilidae

			Guinea		
Dicaeum eximium	y	y	Bismarck Archipelago	y	Dicaeidae
Diphyllodes magnificus	y	y	Papua New Guinea	y	Paradisaeidae
Eulacestoma nigropectus	n	y	Papua New Guinea	y	Eulacestomatidae
Ifrita kowaldii	n	y	Papua New Guinea	y	Ifritidae
Irena cyanogastra	y	y	Philippines	y	Irenidae
Machaerirhynchus nigripectus	y	y	Papua New Guinea	y	Machaerirhynchidae
Myiagra hebetior	y	y	Bismarck Archipelago	y	Monarchidae
Oreocharis arfaki	n	y	Papua New Guinea	y	Paramythiidae
Pachycephala philippinensis	y	y	Philippines	y	Pachycephalidae
Paradisaea raggiana	y	y	Papua New Guinea	y	Paradisaeidae
Paradisaea rubra	n	y	Indonesia	y	Paradisaeidae

Parotia lawesii	n	y	Papua New Guinea	y	Paradisaeidae
Pseudorectes ferrugineus	n	y	Papua New Guinea	y	Pachycephalidae
Ptilorrhoa leucosticta	n	y	Papua New Guinea	y	Cinclosomatidae
Rhagologus leucostigma	y	y	Papua New Guinea	y	Rhagologidae
Rhynochetos jubatus	y	y	New Caledonia	n	Rhynochetidae
Sterrhoptilus dennistouni	y	y	Philippines	y	Zosteropidae
Todus mexicanus	y	y	Puerto Rico	n	Todidae
Zosterops hypoxanthus	y	y	Bismarck Archipelago	y	Zosteropidae