Automatic Discovery of Optimal Discrete Character Models

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Abstract

Modeling discrete character evolution in a Markovian framework has become common practice in

phylogenetic comparative methods. The increasing size and complexity of these models reflects a trend

of analyses to include more taxa and more discrete characters. However, as complexity of the models

increase, so do the number of potential model structures and number of estimable parameters, making it

nearly impossible to consider all modeling options for a given dataset. To overcome this issue, I apply a

combination of regularization and simulated annealing to models of discrete character evolution. This

allows for the automatic searching and optimization across different model structures without user

specification. I test this framework under several simulation scenarios including hidden rates and

multiple discrete characters. The results indicate that regularized models significantly outperform

traditional approaches, yielding a far lower variance and a nearly tenfold reduction in the overall error

of parameter estimates in the most extreme scenarios. I illustrate the power of automatic model

selection by revisiting the ancestral state estimation of concealed ovulation and mating systems in Old

World monkeys. Using the dredge algorithm, I discover a previously unexamined model structure

which has both better statistical performance and a differing ancestral state reconstruction when

compared to default model sets. In general, these results highlight the dangers of an over-reliance on

default model sets. The combination of automatic model selection and regularization help overcome

problems of over-parameterization, and these results demonstrate that when inferences are drawn from

a larger model space, they can be both more statistically robust and biologically realistic.

Multistate discrete character models are now widely applied in phylogenetic comparative methods

(PCMs). These models, which were initially limited to relatively few characters and simple processes,

have been expanded in an attempt to incorporate various biological processes and sources of variation.

Complexity has been introduced to these models through correlated character evolution (Pagel 1994),

hidden rate variation (Beaulieu et al. 2013; Boyko and Beaulieu 2021), state-dependent speciation and

extinction (SSE) (Maddison et al. 2007; Beaulieu and O'Meara 2016), and the incorporation of contin-

uous character information (May and Moore 2020; Boyko et al. 2023).

This increasing complexity has also led to higher generality. For example, hidden Markov mod-

els (HMMs) which were initially introduced to model two binary characters (Beaulieu et al. 2013),

have been expanded to allow for any number of characters, observed states, and hidden states (Boyko

and Beaulieu 2021). However, while this additional flexibility allows for customization of the phyloge-

netic comparative method to the system at hand, the large model space can make it challenging for bi-

ologists to find the most appropriate model for their particular dataset. The typical approach, multi-

model inference framework (Burnham and Anderson 2002), has a biologist decide on a set of potential-

ly realistic models. The relative support for each model is then evaluated based on their fit to the data

using information criteria such as Akaike Information Criterion (AIC). This approach is powerful be-

cause it allows for model averaging, where inferences are made based on a weighted average of the

models included in the set (Burnham and Anderson 2002). However, the effectiveness of multi-model

inference relies heavily on the appropriateness of the model set. Several PCMs have been criticized for

high false positive rates due to the exclusion of the "correct" null hypothesis (Rabosky and Goldberg

2015). Although these criticisms have been addressed by introducing new model structures to serve as

better null hypotheses (Beaulieu and O'Meara 2016; Boyko and Beaulieu 2023), the problem, when

recast as a failure to include a complete model set (Boyko and Beaulieu 2023), suggests that there is

still a vast space of unexplored model structures.

Often the model set chosen for discrete character evolution are nested, with the difference being

which parameters are variable, and which are fixed. For example, for a two-character binary state

dataset, the difference between a correlated model (k=8) and the independent model (k=4) is whether

changes in the focal character depend on the state of the background character (Pagel and Meade

2006). In the independent model parameters representing this dependency process are fixed to be equal,

while in the correlated model they are freely estimated (Pagel and Meade 2006; Boyko and Beaulieu

2023). In this way, one can think of discrete models adding complexity as adding parameters to

increase "biological realism" and represent new processes or relationships between variables.

This increasing model complexity then leads to another, more technical, challenge. As the

number of estimable parameters grows it reaches a point where the number of parameters (k)

approaches the number of taxa (N). However, the rate at which the number of parameters increase often

surpasses the rate at which information can be gained through data (Felsenstein 2012). In the case of

correlated discrete character models, this is because each additional character requires considering its

relationship to all other traits. For instance, the most complex discrete model for a single binary

character has 2 parameters, while the most complex model with two binary characters has 12

parameters (k=8 if excluding dual transitions), and the most complex model with three binary

characters has 56 parameters (k=26 if excluding dual transitions; Fig. 1). In each instance we have

added a single character, but because we must consider the new character's relationship to all other

existing characters, the model complexity (as measured by the number of parameters) outpaces the

potential information gained from the new data. This is problematic because although Maximum

likelihood methods are consistent estimators when $N \gg k$, their performance deteriorates as models

become more parameter-rich, leading to unreliable and biased parameter estimates (Huelsenbeck et al.

2001). However, for complex models with a finite data, it is unlikely that all parameters will be

essential or necessary to best model the data (Lemey et al. 2009; Gelman et al. 2013).

Here, I address this challenge by introducing an approach for automatic model exploration

using regularization and simulated annealing. Regularization techniques, such as L1 (lasso) and L2

(ridge), constrain the magnitude of parameter estimates and encourage simpler, more generalizable

models (Hoerl and Kennard 1970; Tibshirani 1996). By incorporating regularization, we can balance

model complexity and goodness-of-fit, reducing the risk of overfitting and improving the stability of

parameter estimates. Additionally, to search the set of possible models, I implement a simulated

annealing routine that stochastically proposes and accepts model modifications according to a

temperature-controlled optimization schedule. This approach is most similar to Pagel and Meade

(2006), who introduced a reversible-jump Markov chain Monte Carlo (RJMCMC) to search across

models with different numbers of parameters. Their method explores the model space by proposing

merge/split and augment/reduce "moves", which effectively add or remove parameters during

sampling. Models are visited in proportion to their posterior probability and produce a distribution of

supported models rather than selecting a single best model. While RJMCMC provides a rigorous

approach to model selection, it is computationally intensive and currently limited in scope as existing

implementations only support two binary characters and do not accommodate hidden states. My

framework addresses these limitations by integrating simulated annealing and regularization within

corHMM (Beaulieu et al. 2022) implemented as the function corHMMDredge.

To test the dredge framework, I conduct an extensive set of simulations to explore the bias-

variance trade-off associated with regularization. Under regularization, it is expected that models will

have increased generality and lower variance. I test these expectations by examining regularized

model's accuracy of parameter estimates. Additionally, I perform a more detailed simulation test on a

subset of historically important discrete models, such as the hidden rates model (Beaulieu et al. 2013)

and correlated character evolution model (Pagel 1999; Pagel and Meade 2006; Boyko and Beaulieu

2021). Finally, as a case study, I reanalyze the dataset from Pagel and Meade (2006) testing whether

female Old World monkey estrus advertisement is associated with multimale mating systems. I use this

case study to demonstrate the dredge framework, guiding users through each step of the process

including cross-validation and uncertainty estimation.

Figure 1– Visualization of the exponential increase in model complexity as additional binary characters are introduced. (a) A single binary character with two states (k=2). (b) Two binary characters, each with two states (k=8, excluding dual transitions). (c) Three binary characters, each with two states (k=26, excluding dual transitions). Adding characters exponentially increases the number of parameters due to the need to account for relationships between all characters. The disallowance of simultaneous multistate transitions keep the models from being even more complex.

Methods

Regularization and Simulated Annealing

The likelihood of a discrete character model with its underlying framework as a continuous time

Markov chain (CTMC) is calculated as $L = P(D|Q, \Phi, \pi)$, which is the probability of observing the

data (D) given an instantaneous rate matrix (Q), root state frequency vector (π), and a phylogeny with a

fixed topology and set of branch lengths (Φ). The data consist of the observed character states (S) at the

tips of the phylogeny, while the rate matrix contains the rates of character state transitions (q_{ij}). The

likelihood function is then computed by integrating the product of transition probabilities along the

branches of the phylogeny (Felsenstein 1981, 2004; Pagel 1994; Lewis 2001). Though not

mathematically necessary, it may be useful to consider a mapping matrix (M; also referred to as an

index matrix (Revel 2024)) which gives the structure of the discrete model by indicating which

transition rates are estimated and/or fixed to be equal (FitzJohn, 2012; Beaulieu & O'Meara, 2016). For

example, if we consider a simple binary character with states 1 and 2, the instantaneous rate matrix is

given by $Q = \begin{bmatrix} -q_{12} & q_{12} \\ q_{21} & -q_{21} \end{bmatrix}$. However, the mapping matrix can specify several alternative model

structures. If $M_{er} = \begin{bmatrix} 0 & 1 \\ 1 & 0 \end{bmatrix}$, then $q_{12} = q_{21}$ and we have what is commonly known as the "equal rates"

model. An "all rates different" model can be indicated by $M_{ard} = \begin{bmatrix} 0 & 2 \\ 1 & 0 \end{bmatrix}$, where $q_{12} \neq q_{21}$ and a

"unidirectional" model can be constructed via $M_{dir} = \begin{bmatrix} 0 & 1 \\ 0 & 0 \end{bmatrix}$, where $q_{21} = 0$. In this specific example,

we have models of 1 ("equal rates" and "unidirectional") or 2 ("all rates different") estimable

parameters. To determine which model structure is optimal (best balances goodness-of-fit and

complexity) for the dataset, we can compute the maximum likelihood estimate (MLE) for each model

structure and compare them using a information criterion such as AIC (Akaike 1974; Burnham and

Anderson 2002).

This framework has been successfully applied for many years within PCMs (O'Meara 2012),

but growing model complexity (e.g. Fig. 1) is making it untenable for users to define all possible

relevant model structures or for method developers to construct a complete set of default models. As

such, regularization techniques may be a necessary step in discovering optimal model structures for

large and complex datasets. In corHMMDredge, I incorporate three regularization approaches, two of

which are analogous to *l1* and *l2* regularization (Hastie et al. 2015), and the third is based on the null

expectation that transition rates are all equal (*er*; (Zhou et al. 2024). Specifically, the regularized

likelihoods are defined as:

$L_{l1}(D|Q, \Phi, \pi; \lambda) = L(D|Q, \Phi, \pi) - \lambda \frac{1}{n}\sum_{i=1}^{n} Q_{ii},$ (1)

$L_{l2}(D|Q, \Phi, \pi; \lambda) = L(D|Q, \Phi, \pi) - \lambda \frac{1}{n}\sum_{i=1}^{n} Q_{ii}^{2},$ (2)

$L_{er}(D|Q, \Phi, \pi; \lambda) = L(D|Q, \Phi, \pi) - \lambda \frac{1}{k}\sum_{i \neq j} \theta_i - \theta_j,$ (3)

where $L(D|Q, \Phi, \pi)$ denotes the standard likelihood, $\frac{1}{n}\sum_{i=1}^{n} Q_{ii}$ is the elementwise mean of the diagonal

of the instantaneous rate matrix, $\frac{1}{n}\sum_{i=1}^{n} Q_{ii}^{2}$ is the squared elementwise mean of the diagonal of the

instantaneous rate matrix, $\frac{1}{k}\sum_{i \neq j} \theta_i - \theta_j$ is the mean pairwise distances between all freely estimated

parameters in their raw units, k is the number of freely estimate parameters, and λ is a hyper-parameter

that adjusts the severity of the penalty, ranging from 0 (no regularization) to 1 (full penalization). This

penalization scheme uses the mean rather than the sum of the parameter values because the number of

transition rates is a function of the number of possible discrete states and using the sum would cause

the penalization term to be a function of the number of states rather than the overall complexity of the

model (the number of parameters). It is important to note that λ cannot be jointly estimated alongside

the parameters via the maximized likelihood (Clavel et al. 2019). Instead, a form of cross-validation is

necessary to tune λ (see *Phylogenetically informed k-fold cross validation*). This scheme is similar to

Sanderson's penalized likelihood approach for estimating chronograms from phylogenetic trees

(Sanderson 2002). Under penalized likelihood, the penalty is based on differences of rates and is

essentially a regularization approach. Furthermore, the Sanderson method includes cross validation to

estimate the equivalent of the λ hyper-parameter (which he also labeled λ, the smoothing parameter).

The second component of the dredge algorithm involves a method to explore and assess

alternative model structures (Fig. 2). For this, I implemented a simulated annealing heuristic search

algorithm. Simulated annealing is a probabilistic optimization technique inspired by the process of

annealing in metallurgy, where a material is heated and then slowly cooled. The algorithm aims to find

a model that minimizes a given information criterion (e.g., AIC, BIC) and given enough time is

guaranteed to find the optimal solution for a discrete set (Kirkpatrick et al. 1983). This approach is

similar to Pagel and Meade (2006), who also use a Metropolis-Hastings sampling scheme to explore

alternative models in a Bayesian framework. While their method samples from the posterior

distribution, corHMMDredge optimizes each proposed model structure using a penalized likelihood. In

both approaches, poor-fitting models can still be temporarily accepted to escape local optima. The

search is performed independently for each number of hidden rate categories, from the initial category

up to max.rate.cat. Models are evaluated through an iterative process governed by a "temperature"

parameter, T, which starts high and is gradually lowered.

The search begins with an initial model structure (default is the model with the highest number

of free parameters) and its corresponding score. At each iteration, the algorithm proposes a move to a

new, neighboring model structure by making a small perturbation to the current mapping matrix. The

proposed model is then fit to the data, and its score is calculated. The decision to accept this new model

is probabilistic and depends on the change in the information criterion score ($\Delta E = E\_new - E\_current$)

and the current temperature, T. As in Pagel and Meade (2006), if $\Delta E \leq 0$, the proposed model is an

improvement (or equally good), and it is always accepted as the new current model. If $\Delta E > 0$, the

proposed model is worse than the current one, but following the Metropolis algorithm (Metropolis et al.

1953), it may still be accepted with a probability $P = \exp(-\Delta E / T)$. When the temperature T is high, the

acceptance probability is also high, allowing the search to freely explore the entire model space and

escape local minima. As the algorithm proceeds, T is gradually reduced according to an exponential

cooling schedule ($T\_i+1 = T\_i * \alpha$, where α is a cooling.rate typically close to 1.0 – default of 0.95). As

T approaches zero, the probability of accepting a worse model diminishes, causing the algorithm to

behave like a greedy hill-climbing search (Kirkpatrick et al. 1983; Russell and Norvig 2021).

To generate new candidate models, one of three stochastic move types is randomly selected at

each iteration: drop, merge, or free. A parameter drop simplifies the model. The algorithm identifies all

estimated rate parameters and stochastically selects one to remove, with the probability of selection

being inversely proportional to the parameter's estimated rate (i.e., smaller rates are more likely to be

dropped). Second, a parameter merge also simplifies the model by introducing a constraint. The

algorithm identifies pairs of parameters with similar estimated rates. A pair is randomly selected, with

the probability of selection being inversely proportional to the absolute difference between their rates

(i.e., more similar rates are more likely to be merged). The parameters are then constrained to be equal.

For hidden Markov models with multiple rate categories, this merging is restricted to occur only among

parameters within the same underlying rate class. Finally, a parameter free move increases model

complexity. It identifies transitions that are currently constrained (i.e., previously dropped or merged)

and randomly selects one to "free" by assigning it a new, unique parameter index. Note that these three

move types (drop, merge, free) are conceptually similar to the proposals used in reversible-jump

MCMC algorithms (Pagel and Meade 2006). Like Pagel and Meade (2006), corHMMDredge proposes

model simplifications or expansions through stochastic rules. However, unlike an RJ-MCMC,

corHMMdredge does not rely on priors and marginal likelihoods to navigate model space, instead the

dredge framework utilizes heuristics based on rate magnitudes to propose moves and evaluates model

quality using information criteria. This should make the method more scalable to high-dimensional

model spaces.

Finally, to prevent the search from becoming permanently trapped in a local minimum, a restart

mechanism is included. If the search fails to find a new best-fitting model or has not accepted a new

model for a predefined number of iterations (default of 20 attempts), the algorithm reverts to a

randomly selected, previously accepted (but not necessarily optimal) model from its search history.

The simulated annealing process for a given rate category terminates after a maximum number of

iterations or when the temperature becomes negligibly small. The algorithm maintains a record of all

unique models accepted during the search. Upon completion, any redundant models (i.e., models with

identical log-likelihoods and parameter counts but different rate structures) are pruned, and the

remaining unique models are ranked by their information criterion scores to provide a set of models for

further consideration.

a) current model

b) possible moves

Merge (1/3)   Drop (1/3)

Free (1/3)   Restart

c) evaluate and increment search

```
T = current_temp * cooling_rate
IF proposal_accepted:
    current_model = proposed_model
    reset_counter = 0
ELSEIF restart_needed:
    current_model = previous_model
ELSE:
    current_model = current_model
    reset_counter += 1
IF current_temp < threshold
    BREAK (search complete)
```

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Figure 2 – An overview of the simulated annealing algorithm. From a (a) current model, the algorithm
proposes a new model by stochastically selecting a move. (b) Possible moves include simplifying the
model by merging or dropping parameters or increasing complexity by freeing a constrained parameter.
A restart mechanism is also available to escape local optima. (c) The algorithm then decides whether to
accept the new model. Better models are always accepted, while worse models may be accepted with a
probability that decreases as the search "temperature" cools, allowing the search to converge on an
optimal solution.
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Phylogenetically informed k-fold cross validation

To optimize λ I choose to employ a phylogenetically informed k-fold cross-validation procedure

across the tips of the phylogeny. Cross-validation approaches, such as leave-one-out cross-validation

(Lartillot 2023), have been shown to be more adequate than alternatives for model selection. Here I

implement a k-fold method because it provides a computationally efficient and intuitive framework that

directly addresses phylogenetic non-independence through weighted sampling. K-fold cross-validation

is a common way to optimize λ when using regularization (Hastie et al. 2009). It involves dividing the

dataset into k separate subsets (or folds). The model is trained on k-1 of these folds with the remaining

fold used as the test set to evaluate the models' performance. This process is repeated k times, with

each of the k folds used exactly once as the test set. The results of these k tests are then averaged to

produce a single estimate. In the context of a discrete character model the metric chosen for evaluation

is reconstruction of tip state values. Each species within a k-fold is coded as having an unknown state

and a reconstruction based on the fitted model is then used for evaluation. The score for a fold is

determined by the average Jensen-Shannon divergence between the predicted and actual tip states.

Specifically, $score_k = \frac{1}{N_k} \sum_{i=1}^{N_k} D(L_i, \hat{L}_i)$ where N_k is the number of tips in the k-th fold, L_i are the

observed state likelihoods at the tips of the k-th fold (1 for the observed state and 0 for an unobserved

state), $\hat{L}_i$ are the predicted state likelihoods for the tips in the k-th fold, and D is the Jensen-Shannon

(JS) divergence (or any form of divergence more generally). The k scores are then averaged over all

folds to obtain a single score for the model. The JS divergence (available in corHMM as the function

js_divergence) is chosen because it is a symmetric and bounded measure of difference between

predicted and observed probability distributions while being robust to zero probabilities. An advantage

of using tip values to measure the model fit is that the phylogenetic structure of the data is unchanged

throughout the entire cross-validation procedure because no tips need to be explicitly dropped when

fitting the model; they only need to be set to an unknown value. The samples within a given k-fold are

chosen without replacement and with probabilities equal to $w = (I^T C^{-1})\vec{1}$ (Rohlf 2001) to ensure

phylogenetically even sampling between the k-folds, where w is roughly proportional to the

phylogenetic uniqueness of each species.

Simulation Study I: Bias and variance of parameter estimates

The expected behavior of a regularized model is that the introduction of a penalty term to the

likelihood will reduce sensitivity to the specific dataset, resulting in lower variance and greater

generalizability at the cost of a systematic error (bias). Though the behavior of regularization

techniques is well documented in generalized least squares regression (Hastie et al. 2009), it is less well

studied in the context of phylogenetic comparative methods (but see Khabbazian et al. 2016; Clavel et

al. 2019) and particularly understudied when applied to non-regression frameworks such as the Markov

models. In the context of discrete character models, the expected behavior is that rate estimates will

tend towards zero, resulting in a downward bias of parameter estimates. Biologically, this means that a

regularized model expects fewer changes in the character state through time and favors explanations of

homology over homoplasy. The benefit of a regularized model is the increased generalizability should

lead to a model which more accurately reflects the dynamics of unsampled taxa and leads to better

predictions of unknown character states (e.g., ancestral character estimation).

To test the bias-variance trade-off associated with the regularization framework introduced here,

I conduct a simulation study to evaluate the bias, variance, and overall error of parameter estimates for

regularized and unregularized models. I simulate data under three model structures which vary only in

the number of characters included. Specifically, I simulate data for a single character with binary states

(k=2), two characters with binary states (k=8), and three characters with binary states (k=24). It was not

required that all state combinations had to be observed. All models are simulated under an all rates

different (ARD) model with dual transitions disallowed. Parameters are sampled from a log-normal

distribution, where the logarithms of the parameters are normally distributed with a mean of 0 and

standard deviation of 0.25. Parameters are sampled 100 times for a given phylogenetic tree.

Phylogenetic trees of size 100, 250, and 500 taxa are simulated 10 times under a pure birth model with

a sampling fraction of 1 using the R-package TreeSim (Stadler 2019). Phylogenies are then rescaled to

a height of 1. These same 30 phylogenetic trees (10 each for the different number of taxa) are used for

the 3 different model structures. In total, each model structure is tested under 3000 simulated datasets

for a total of 9000 simulations.

Discrete character models are fit using the R-package corHMM (Beaulieu et al. 2022) with the

simulating model structure (i.e., ARD) specified. Models are fit with no penalty term (unregularized),

an *er* penalty term, an *l1* penalty term, and an *l2* penalty term with $\lambda = 1$. The maximum likelihood

estimates are then compared to the simulating values and their bias, variance, and Root Mean Squared

Error (RMSE) evaluated. Note that hidden Markov models are not included in Simulation Study I

because, for trees of this size, it would be difficult to simulate data which consistently had a strong

signal of rate heterogeneity (Boyko and Beaulieu 2021). This means that regularized models would

show an inflated negative bias as parameters in a second-rate class would be estimated as 0 when there

is weak evidence of hidden rates.

Simulation Study II: Automatic model selection

Apart from reducing the variance of discrete character models, a primary aim of the dredge

framework is to help users navigate large and complex model spaces. As such, in *Simulation Study II* I

simulate data under three historically significant and widely applied model structures and evaluate

whether the dredged model captures the most important features of the generating models. To conduct

this evaluation, I compare three model selection strategies. The first is a standard unregularized greedy

search, which acts as a baseline. The other two methods employ *l1* regularization to penalize model

complexity, one using a greedy search algorithm and the other using a more exhaustive simulated

annealing approach. Specifically, I simulate data under: (1) a correlated model, which captures the

influence of two or more traits on each other (Pagel 1994); (2) an ordered and unidirectional model of

trait change (e.g., Skinner et al. 2008); and (3) a hidden Markov model, where there is an unobserved

factor causing rate variation in an observed character (Felsenstein and Churchill 1996; Beaulieu et al.

2013; Boyko and Beaulieu 2021). Each of these models has a unique set of criteria for the "most

important features," depending on what is typically considered relevant in empirical hypotheses using

the particular model structure. Below I go into detail about which features are considered relevant for

this study in each case.

When the generating model is consistent with Pagel's (1994) correlated model, 100 datasets are

simulated under Q_{corr} with transition rates of $q_{(X_0 \to X_1)|Y_1} = 10$, $q_{(X_1 \to X_0)|Y_0} = 5$, $q_{(X_1 \to X_0)|Y_1} =$

$q_{(X_0 \to X_1)|Y_0} = 0.1$, and $q_{(Y_0 \to Y_1)|X_0} = q_{(Y_0 \to Y_1)|X_0} = q_{(Y_1 \to Y_0)|X_0} = q_{(Y_1 \to Y_0)|X_1} = 1$. A unique pure birth

phylogeny (birth rate = 1 events/MY) with 250 tips is simulated for each dataset and rescaled to have a

height of 1. To evaluate whether the dynamics of the correlated model are adequately captured, I

quantify how often does the best fitting dredge model find that: (1) $q_{(X_0 \to X_1)|Y_1} > q_{(X_0 \to X_1)|Y_0}$, (2)

$q_{(X_1 \to X_0)|Y_0} > q_{(X_1 \to X_0)|Y_1}$, (3) $q_{(Y_0 \to Y_1)|X_0} = q_{(Y_0 \to Y_1)|X_1}$, (4) $q_{(Y_1 \to Y_0)|X_0} = q_{(Y_1 \to Y_0)|X_1}$, and (5) all other tests are

passed. These comparisons are chosen because they are relevant to the interpretation of the correlated

model. Evidence for the correlated model comes from tests of whether changes in the focal character

depend on the state of the background character (Pagel and Meade 2006). As a hypothetical example,

consider the situation in which X is parity mode and Y is climatic preference in reptiles: X_0 is oviparity,

X_1 is viviparity, Y_0 is a warm climatic preference, and Y_1 is a cold climatic preference. An important

factor for the evolutionary transition from egg-laying (oviparity) to live-birth (viviparity) in reptiles is

the climate a lineage occupies, with squamate viviparity evolving frequently in colder climates, such as

those found at high altitudes and latitudes (Blackbum 1999). When a correlated model of discrete

character evolution is applied to this hypothetical example, the simulation parameters outlined above

indicate that the transition of oviparity to viviparity is faster when a lineage is in a cold environment

($q_{(X_0 \to X_1)|Y_1}$) than when the lineage is in a warm environment ($q_{(X_0 \to X_1)|Y_0}$) and that transitions from

viviparity to oviparity are faster when lineages are in a warm environment ($q_{(X_1 \to X_0)|Y_0}$) than when they

are in a cold environment ($q_{(X_1 \to X_0)|Y_1}$). If parity mode evolved independently of climatic environment,

then the expectations is that the transition rates between oviparity and viviparity are the same

regardless of which environment the lineages inhabit. If corHMMDredge consistently finds

$q_{(X_0 \to X_1)|Y_1} > q_{(X_0 \to X_1)|Y_0}$ and $q_{(X_1 \to X_0)|Y_0} > q_{(X_1 \to X_0)|Y_1}$, then it would seem to adequately capture

correlated trait dynamics. Additionally, the simulation includes a negative control. For this simulation,

transitions between climate do not dependent on the presence of viviparity or oviparity. This is captured

by tests (3) and (4) examine whether transitions from warm to cold climates ($q_{(Y_0 \to Y_1)|X_0} = q_{(Y_0 \to Y_1)|X_1}$)

and cold to warm climates ($q_{(Y_1 \to Y_0)|X_0} = q_{(Y_1 \to Y_0)|X_1}$) are the same regardless of the reproductive mode.

$$Q_{corr} = \begin{array}{c|cccc} & X_0Y_0 & X_1Y_0 & X_0Y_1 & X_1Y_1 \\ \hline X_0Y_0 & - & q_{(X_0 \to X_1)|Y_0} & q_{(Y_0 \to Y_1)|X_0} & - \\ X_1Y_0 & q_{(X_1 \to X_0)|Y_0} & - & - & q_{(Y_0 \to Y_1)|X_1} \\ X_0Y_1 & q_{(Y_1 \to Y_0)|X_0} & - & - & q_{(X_0 \to X_1)|Y_1} \\ X_1Y_1 & - & q_{(Y_1 \to Y_0)|X_1} & q_{(X_1 \to X_0)|Y_1} & - \end{array} = \begin{array}{c|cccc} & X_0Y_0 & X_1Y_0 & X_0Y_1 & X_1Y_1 \\ \hline X_0Y_0 & - & 0.1 & 1.0 & - \\ X_1Y_0 & 5.0 & - & - & 1.0 \\ X_0Y_1 & 1.0 & - & - & 10.0 \\ X_1Y_1 & - & 1.0 & 0.1 & - \end{array}$$

To simulate under an ordered trait evolution model, I construct a matrix Q_{ord} which allows for

transitions from state 1 to state 2 to state 3, where reversions are possible only in state 2. All transition

rates are set to 1, such that $q_{1\to2} = q_{2\to1} = q_{2\to3} = 1$ and a unique pure birth tree with 250 taxa is

simulated with birth rate 1 for 100 datasets and trees are rescaled to a height of 1. To evaluate whether

the dynamics of a ordered model are actually captured by the dredge framework, I conduct 4 tests: (1)

$q_{1\to3} = 0$, (2) $q_{2\to1} > 0 \wedge q_{2\to3} > 0$, (3) $q_{3\to1} = q_{3\to2} = 0$, and (4) all other tests are passed. The first 3

tests examine whether the correct structure of the ordered model is discovered by corHMMDredge. A

biological example of this type of model would be transitions between outcrossing and obligate selfing

in angiosperms, through an intermediate facultative selfing state and the irreversibility of obligate

selfing state. If this were the case, X_1 would represent out-crossing, X_2 would represent facultative

selfing, and X_3 would represent obligate selfing. The first test asks whether corHMMDredge has

accurately discovered that it is impossible to directly transition from outcrossing to obligate selfing.

The second test naturally follows the first examining whether transitions from the intermediate

facultative selfing state are allowed to transition to both obligate selfing and outcrossing. The third tests

asks whether the obligate selfing is correctly inferred to be a sink state, incapable of reverting back to

either facultative selfing or outcrossing. This ordered model is somewhat unique among the historical

models I am testing in that it is often clearly linked to apriori biological expectations related to

developmental biology. And in those cases, it may not be appropriate to apply a dredge framework as

the model set will be well defined. However, these sorts of ordered dynamics are possible in settings

where developmental information is not readily available, and the goal of these simulations is to

determine if the corHMMDredge framework can detect ordered models in general.

$$Q_{ord} = \begin{array}{c} \\ X_1 \\ X_2 \\ X_3 \end{array} \begin{array}{ccc} X_1 & X_2 & X_3 \\ - & q_{1\to2} & - \\ q_{2\to1} & - & q_{2\to3} \\ - & - & - \end{array} = \begin{array}{c} \\ X_1 \\ X_2 \\ X_3 \end{array} \begin{array}{ccc} X_1 & X_2 & X_3 \\ - & 1.0 & - \\ 1.0 & - & 1.0 \\ - & - & - \end{array}$$

The final historically significant model being tested is a hidden Markov model where

unobserved rate variation is introduced via a hidden factor. The matrix, Q_{HMM}, describes a simple

HMM for a single character, X, with two states, 0 and 1. The structure of an HMM is not unlike a

correlated model, except that in the correlated case, both characters are observed and for an HMM we

are trying to detect the hidden character based on the rate differences in the observed character (Boyko

and Beaulieu 2021). For simulations, transition rates are set to be $q_{(X_0\to X_1)|A} = q_{(X_1\to X_0)|A} = 10$,

$q_{(X_0\to X_1)|B} = q_{(X_1\to X_0)|B} = 0.01$, and $q_{(A\to B)|X_0} = q_{(A\to B)|X_1} = q_{(B\to A)|X_0} = q_{(B\to A)|X_1} = 1$ and simulated on

100 unique pure birth tree with 250 taxa. To evaluate whether the dynamics of a hidden Markov model

are captured by the corHMMDredge framework, I conduct 2 tests: (1) $n\_rate\_class > 1$ (2)

$q_{(X_0 \leftrightarrow X_1)|A} / q_{(X_0 \leftrightarrow X_1)|B} \geq 100$, (3) $q_{(X_0 \leftrightarrow X_1)|A} \vee q_{(X_0 \leftrightarrow X_1)|B} > q_{(A \leftrightarrow B)}$. The first test asks whether

there is support for hidden rates at all. The second test, conditioned on the first, asks whether the

difference between the fastest rate class 100 times is at least greater than the slower rate class. I note

that the simulating difference of 1000 times is a large difference (and arguably unrealistic) difference in

rate classes but is chosen to ensure that simulating data had a signal of rate heterogeneity. Test 3

examines whether the fastest rates belong to transitions between observed states, rather than between

hidden states.

$$Q_{HMM} = \begin{array}{c|cccc} & X_{0A} & X_{1A} & X_{0B} & X_{1B} \\ \hline X_{0A} & - & q_{(X_0 \to X_1)|A} & q_{(A \to B)|X_0} & - \\ X_{1A} & q_{(X_1 \to X_0)|A} & - & - & q_{(A \to B)|X_1} \\ X_{0B} & q_{(B \to A)|X_0} & - & - & q_{(X_0 \to X_1)|B} \\ X_{1B} & - & q_{(B \to A)|X_1} & q_{(X_1 \to X_0)|B} & - \end{array} = \begin{array}{c|cccc} & X_{0A} & X_{1A} & X_{0B} & X_{1B} \\ \hline X_{0A} & - & 10.0 & 1.0 & - \\ X_{1A} & 10.0 & - & - & 1.0 \\ X_{0B} & 1.0 & - & - & 0.01 \\ X_{1B} & - & 1.0 & 0.01 & - \end{array}$$

Empirical example: concealed ovulation and mating systems in Old World monkeys

Concealed ovulation has been considered an adaptation in primates as either a means of

promoting parental care by increasing confidence in paternity or improving male behavior towards

their potential offspring by confusing paternity issues (Sillen-Tullberg and Moller 1993). Addressing

whether concealed ovulation increases confidence or confusion in paternity issues is difficult to study

at a phylogenetic scale, so previous examinations of this question have focused on the context in which

the trait has evolved. An initial analysis by Sillen-Tullberg and Moller (1993) found that the common

ancestor of all anthropoid primates showed signs of ovulation (or estrus advertisement) and had a

multimale mating system. Concealed ovulation then evolved from this state 8 to 11 times in a

nonmonogamous context and at most once in a monogamous context. In contrast, in their examination

of this same question, Pagel and Meade (2006) found the common ancestor of all Old World primates

to have concealed ovulation and a monogamous mating system. This discrepancy was explained by the

difference in methodology. Pagel and Meade (2006) had used rate estimates from Markov models and

estimating the ancestral state of the Old World primate common ancestor is crucial for the subsequent

interpretation as it will imply whether estrus advertisement or concealed ovulation is the derived state

in primates, though an updated dataset will likely be necessary before we can trust the robustness of

any biological interpretations from these analyses.

To examine the evolution of concealed ovulation and mating systems, I apply corHMMDredge

to the dataset of Pagel and Meade (2006). The dredge models use *l1* regularization and I conduct

phylogenetic cross-validation to determine the optimal λ value. For cross-validation I examine 5 folds

for λ values of 0, 0.25, 0.5, 0.75, and 1 with Jensen-Shannon divergence used as a metric of accuracy.

All model fits undergo 10 random restarts to increase the chances of finding the global optimum for the

rate estimates. The best dredge model is used to estimate ancestral states based on the regularized

maximum likelihood estimate. The root prior is set to "maddfitz" (FitzJohn et al. 2009) in the corHMM

R-package. I examine the profile likelihoods of the rate estimates to generate 95% confidence intervals

around the MLE independently for each parameter. To do this, I implement a new profile likelihood

function within corHMM and examine 200 fixed values for each parameter spanning 100 orders of

magnitude around the MLE. Profile likelihood operates by examining what the maximum likelihood

would be given a fixed value and with the remaining parameters freely estimated.

Results

Simulation Study I: regularized models substantially reduce variance

The introduction of *l1* and *l2* regularization led to a significant reduction in total variance across

all tested models for the raw parameter estimates (Table 1). Furthermore, the absolute average bias

($\frac{1}{n}\sum_{i=1}^{n}|\hat{\theta} - \theta|$) for phylogenies with 100 taxa and 250 taxa was found to be lower for *l1* and *l2*

regularized models, contrary to the expected increase in negative bias (Table 1). This trend reversed for

l2 regularized models fit to datasets with trees of 500 taxa, but *l1* regularization consistently showed

the lowest bias across all tested tree sizes. More surprisingly, the bias of *l1* regularized models was

slightly positive on average, contrary to the expected downward bias, although a negative bias is found

if one uses the median difference instead of the arithmetic mean. Finally, when considering overall

error (RMSE), *l1* and *l2* regularized models showed substantial improvements over unregularized (*l0*)

and *er* regularized models (Table 1). This improvement in RMSE highlights the effectiveness of *l1* and

l2 regularization in achieving better generalizability and prediction accuracy, despite the potential

trade-off with increased bias. It is important to note that several of these findings go against

expectation. Particularly, the absolute bias of an unregularized model is not expected to be greater than

that of regularized models. There are two possible explanations for this which we can examine. First, it

could be that several of the parameter estimates lack information to be well estimated causing them to

reach an upper or lower bound during the maximum likelihood search. Indeed, as one removes outlier

transition rate estimates the overall variance of the unregularized model estimates decreases

substantially (Table 1). The second explanation is that parameter estimates should be evaluated on a log

scale because fold changes are more meaningful for rate estimates than the raw units.

When bias and variance are examined on log-transformed parameter estimates, I find that

variance is slightly higher for regularized models than the unregularized models. Additionally, all

models demonstrate a negative bias suggesting that transition rates are generally underestimated. Both bias and variance decrease as sample size increases. Once again, these results are counterintuitive, not only because the regularized models have a larger variance estimate, but because unregularized models are showing a consistent negative bias. Because I expect that this is due to outlier estimates I applied an aggressive filtering strategy, removing parameter estimates that 5 orders of magnitude above or below the true generating parameter values (Table 1; for original units the filtering strategy is to remove differences in transition rates of greater than 10 units). Unsurprisingly, without outliers the bias, variance, and RMSE all decrease substantially. More importantly, when examining log-transformed parameter estimates, I find that regularized models have a negative bias with an absolute bias that is greater than the unregularized model and that regularized models have a lower variance than the regularized model matching expectations(Table 1).

Table 1 - Bias, variance, and overall error (RMSE) for regularized and unregularized models across phylogenies of varying sizes. Results are summarized across all generating models and presented in both original units and log-transformed units. Values in brackets are the bias, variance, or RMSE after filtering outliers. For original units, outliers are defined as having a transition rate estimate that is 10 transitions per unit time greater than the generating value. For log units, outliers are defined as being 5 orders of magnitude greater or lower than the generating value.

Reg. type	Ntax a	Original units			Log units		
		Bias	Variance	RMSE	Bias	Variance	RMSE
l0	100	1.65 (0.34)	87.83 (3.92)	9.45 (1.92)	-1.77 (0.09)	15.29 (0.85)	4.22 (0.68)
l0	250	0.48 (0.23)	15.91 (2.48)	3.92 (1.43)	-1.08 (0.02)	9.19 (0.57)	3.11 (0.42)
l0	500	0.16 (0.13)	2.69 (1.52)	1.45 (0.99)	-0.57 (0.002)	4.95 (0.47)	2.17 (0.29)
er	100	1.68 (0 0.34)	90.66 (3.94)	9.6 (1.92)	-1.77 (0.09)	15.32 (0.83)	4.22 (0.67)
er	250	0.47 (0.22)	15.47 (2.5)	3.87 (1.43)	-1.1 (0.02)	9.42 (0.57)	3.16 (0.42)
er	500	0.16 (0.13)	2.71 (1.52)	1.46 (0.99)	-0.57 (0.003)	4.86 (0.47)	2.15 (0.29)
l1	100	-0.05 (-0.05)	2.08 (2.03)	1.4 (1.38)	-2.51 (-0.18)	17.46 (0.65)	4.82 (0.64)
l1	250	0.04 (0.04)	1.71 (1.66)	1.18 (1.16)	-1.31 (-0.09)	9.93 (0.51)	3.32 (0.41)
l1	500	0.04 (0.04)	1.21 (1.18)	0.86 (0.84)	-0.65 (-0.04)	5.19 (0.43)	2.25 (0.28)
l2	100	-0.5 (-0.5)	0.46 (0.46)	0.98 (0.98)	-2.81 (-0.38)	16.43 (0.42)	4.89 (0.7)
l2	250	-0.33 (-0.33)	0.5 (0.5)	0.81 (0.81)	-1.61 (-0.21)	10.04 (0.34)	3.48 (0.43)
l2	500	-0.21 (-0.21)	0.49 (0.49)	0.64 (0.64)	-0.86 (-0.12)	5.57 (0.34)	2.41 (0.32)

Simulation Study II: performance for model selection

Both regularized and unregularized methods preformed equally well when it came to automatically inferring the generating model structure (Table 2). The most substantial differences are seen when comparing the simulated annealing routine versus a greedy hill climbing approach (9 out of the 12 tests simulated annealing preforms as well or better). For the correlated model, while all methods accurately recovered the primary dependencies, the simulated annealing search was roughly twice as successful at passing all focal tests simultaneously (10.1% vs 5.1-6.1%). Its advantage was more pronounced for the ordered model, where simulated annealing was substantially more effective at identifying both forbidden transitions and irreversible sink states, leading to an overall success rate of 33% compared to ~20% for the greedy searches. Finally, for hidden states model, simulated annealing was better at detecting the large magnitude difference between the fast and slow rate classes, succeeding in 75.8% of simulations compared to 65.3% for the *l1*-regularized greedy search and only 25.3% for the unregularized greedy search. This indicates that while greedy searches are generally proficient, the more exhaustive search of simulated annealing can help uncover additional complex model structures, particularly those involving rate heterogeneity.

Table 2: Results from structure tests comparing a greedy hill climbing search to simulated annealing (SA). Right hand values indicate the proportion of time each framework passed the row's Focal Test. All methods performed reasonably in identifying the generating model structure.

Model	Focal Test	Greedy (l0)	Greedy (l1)	SA (l1)
dependent	$q_{(X_0 \to X_1)\|Y_1} > q_{(X_0 \to X_1)\|Y_0}$	0.98	**0.99**	0.97
dependent	$q_{(X_1 \to X_0)\|Y_0} > q_{(X_1 \to X_0)\|Y_1}$	0.899	0.899	**0.909**
dependent	$q_{(Y_0 \to Y_1)\|X_0} = q_{(Y_0 \to Y_1)\|X_1}$	**0.303**	0.293	**0.303**
dependent	$q_{(Y_1 \to Y_0)\|X_0} = q_{(Y_1 \to Y_0)\|X_1}$	0.162	0.253	**0.293**
dependent	All dependent model tests passed	0.051	0.061	**0.101**
ordered	$q_{1 \to 3} = 0$	0.66	0.54	**0.71**
ordered	$q_{2 \to 1} > 0 \wedge q_{2 \to 3} > 0$	**0.53**	0.48	0.51
ordered	$q_{3 \to 1} = q_{3 \to 2} = 0$	0.44	0.68	**0.72**
ordered	All ordered model tests passed	0.21	0.2	**0.33**
hidden states	$n\_rate\_class > 1$	0.726	0.863	**0.905**
hidden states	$q_{(X_0 \leftrightarrow X_1)\|A} / q_{(X_0 \leftrightarrow X_1)\|B} \geq 100$	0.253	0.653	**0.758**
hidden states	$q_{(X_0 \leftrightarrow X_1)\|A} \vee q_{(X_0 \leftrightarrow X_1)\|B} > q_{(A \leftrightarrow B)}$	0.705	**0.8**	0.789

Canalization of mating systems through time

The ancestral state reconstruction at the root under the dredge model had considerable uncertainty regarding the evolutionary origins of mating system traits. The most likely estimated ancestral state was concealed ovulation with a monogamous mating system (state 00) having a marginal probability of 61%, while state 01 (estrus display and monogamous mating system) had a marginal probability of

38% (Fig. 3a). The result of simulated annealing identified a two-parameter model incorporating

hidden rates as the optimal framework for describing trait evolution (Fig. 3b). The parameter estimates

revealed distinct transition rate classes: states 00, 01, and 11 shared a common transition rate of 0.038

transitions/MY (95% CI: 0.017, 0.093), while transitions from 00R1 to 00R2 and 11R1 to 11R2

occurred at a rate of 0.029 (95% CI: 0.00099, 0.12). A particularly notable feature of the optimal model

is the presence of two sink states within the R2 rate classes. Once a lineage transitions to rate class two

for either concealed ovulation with a monogamous mating system with (00R2) or estrus display with

multimale mating (11R2), the model predicts no further evolutionary transitions, effectively trapping

these lineages in their current state. This is similar to the covarion model, where molecular

evolutionary processes can cease entirely in certain lineages (Tuffley and Steel 1998). However, this

model differs in that lineages entering these sink states are not expected to resume evolutionary

transitions, suggesting a form of evolutionary canalization rather than temporary stasis. Based on state

reconstructions at the tips for the unknown hidden states, there are some clade-specific patterns of

evolutionary canalization. Clade 1 (*Hylobates*) appears to have entered this canalized state, remaining

permanently as monogamous mating with concealed ovulation (00), while Clade 2 (*Macaca*) is

trending towards a canalization of multimale mating and estrus display (11). Additionally, the common

ancestor of humans and their closest relatives is estimated to be at an intermediate state (01), which

subsequently diverged to state 00R1 in humans and state 11R1 in both *Pan paniscus* and *Pan*

troglodytes (Clade 3).

Figure 3 – Results of the empirical dredge analysis of primate mating systems and ovulation. (a) Ancestral state reconstruction on the phylogeny under the best-fitting model. Node and tip colors correspond to the combined character states and hidden rate classes (R1, R2). Numbered clades are referenced in the text. (b) The structure of the final dredged model, a two-parameter hidden Markov model. Colored states and solid arrows represent the inferred model, while grayed-out elements show the full potential model space. Note the "sink" states (00R2 and 11R2) which have no outgoing transitions. (c) The simulated annealing trace plot, showing the AICc score over iterations for models with one and two rate classes. The bar at the bottom indicates the type of move proposed at each step.

The dredge analysis was conducted using a temperature of 4 and cooling rate of 0.9 (Fig. 3c).

However, the initial run's final model included a transition involving an unobserved state (10),

suggesting that the simulated annealing routine had not reached the global optimum and the

temperature parameter maybe have been too low for the model's complexity. To address this limitation,

a second dredge run was initiated using the first run's final model as a starting point. This iterative

approach demonstrates the utility of human-guided model refinement in corHMMDredge, allowing

researchers to apply biological intuition to improve model realism. Finally, cross-validation analysis

supported the robustness of the final model, with a lambda value of 1 yielding the highest validation

scores (0.2234; Table 3). While our cross-validation was performed on the final dredged model, I note

that this approach may introduce a bias, as the model search itself was conducted under a fixed lambda.

Ideally, cross-validation would be integrated into the entire simulated annealing routine. However,

given the significant computational cost of this procedure, I recommend most users perform cross-

validation as a final, confirmatory step. For studies where computational resources are not a limiting

factor, running the entire dredge search across a range of lambda values and comparing the cross-
validation scores of each final model would provide the most rigorous validation.

Table 3 - Results of the 5-fold cross-validation used to determine the optimal l1 regularization penalty (lambda) for the final dredged model. Each value represents the validation score for a given fold and lambda setting. The optimal value was determined to be lambda = 1.0, as it produced the best average score across all five folds.

Lambda	0	0.25	0.5	0.75	1
Fold 1	0.2836	0.2259	0.2256	0.2254	0.2252
Fold 2	0.2722	0.2180	0.2171	0.2163	0.2156
Fold 3	0.2282	0.2276	0.2271	0.2266	0.2262
Fold 4	0.2626	0.2617	0.2608	0.2600	0.2592
Fold 5	0.1918	0.1915	0.1913	0.1912	0.1911
Average Score	0.2477	0.2249	0.2244	0.2239	0.2235

Discussion

Uncertainty in ancestral estimation and the challenges of model interpretation

The dredge algorithm was able to find a unique, non-standard, model structure in the empirical

case study (Fig. 3b). The dredge model estimated the ancestral state at the root to be a monogamous

mating system with concealed ovulation (marginal probability of 61%), though it was fairly uncertain.

Sillen-Tullberg and Moller (1991) found that the root state was likely to be estrus advertisement with

multimale mating system and therefore discussed the evolution of concealed ovulation in this context.

In contrast, and as in this study, Pagel and Meade (2006) had found that the best supported root state

was concealed ovulation with monogamous mating. In the absence of direct observation, it is nearly

impossible to say which of these outcomes is more correct. However, revisiting the hypotheses of

Sillen-Tullberg and Moller (1991) under the dredged model results do provide some interesting

alternative interpretations for the evolution of estrus advertisement and mating system.

Sillen-Tullberg and Moller (1991) had considered concealed ovulation to be the derived state

and found that it had almost always evolved in non-monogamous lineages. This provided evidence that

concealed ovulation was more likely to be a prerequisite for the evolution of monogamy, rather than a

consequence of it (as had been hypothesized by (Burley 1979). However, these modeling results

suggest an entirely different pathway with estrus display evolving only after multimale mating has

established itself. This finding somewhat contradicts the hypothesis that signs of ovulation will

disappear in non-monogamous mating systems because paternity confusion will be beneficial (Hrdy

1979). That is one possible outcome and is inferred to occur at a rate of 0.038 transitions/MY (11 →

01), but from the concealed ovulation and multimale state, a lineage is just as likely to regain overt

ovulation signs as it was to have lost them in the first place (0.038 transitions/ MY; 01 → 11). This is

because under the dredged model structure, concealed ovulation with multimale mating, is an

intermediate state. Furthermore, a particularly novel insight from the dredged model is the inference of

evolutionary canalization. The model identifies sink states where, after a lineage transitions into a

second rate-class for either monogamy and concealed ovulation (00R2) or multimale mating with

estrus display (11R2), it becomes evolutionarily trapped. Though it is not possible to know the

mechanism from comparative data, this model could explain the relative stability of mating systems in

certain clades like gibbons.

There are two important caveats when interpreting these results. First, this biological

interpretation is contingent on the single best-fitting model. While this model had the lowest AIC score,

several others had values well within the recommended threshold of ΔAIC < 2 (Burnham and Anderson

2002), implying that alternative evolutionary scenarios also have substantial support. As discussed in

the section Choosing the "best" model, model averaging to account for this uncertainty is non-trivial

when dealing with hidden states. Second, any comparative analysis is fundamentally dependent on the

input data. This study re-analyzes the Pagel and Meade (2006) dataset, which is now nearly two

decades old. Revisions to the phylogeny or character state assignments in that time could significantly

alter the outcome. Therefore, while the dredged model provides a compelling alternative hypothesis for

the evolution of mating systems and ovulation, its conclusions should be viewed as provisional and

should be reexamined with contemporary data.

The cost of regularization and eliminating very high transition rates

The dredge algorithm, and regularization more generally, will result in a downward bias

proportional to the magnitude of the transition rate. It is worth considering what, if any, is the cost of

this downward bias. In empirical settings, it is not uncommon to have transition rates approaching and

being estimated at an upper bound. When interpreted as 100s of state transitions occurring over short

time intervals, this may be cause for concern because it often seems biologically implausible. However,

high transition rates do not necessarily predict that many transitions of that type have occurred. Often,

high transition rates may correspond to only a few inferred transitions on a tree (though they will have

occurred very rapidly). The two inferences, extremely high transition rates and relatively few

transitions, may seem to be contradictory, but they are perfectly compatible since a lineage must first be

in the initial state to transition, and the initial state need not be common. Instead, a more serious cause

for concern is the amount of information available to infer the transition rate if that transition only

occurs rarely throughout the clade's history. This was the case for the Old World monkey dataset when

examining an all-rates-different model. Several parameter estimates seemed to have very little

information and fell along a likelihood ridge. The combination of regularization and model structure

searching removes this ridge but also has the benefit of estimating rates which better match intuition

that a transition which occurs rarely throughout the clade has a corresponding transition rate that is also

low. Ultimately, a regularization scheme moves towards a more parsimonious view of character

evolution, with rate estimates tending to be biased towards explanations of homology over homoplasy.

The value of multiple characters

Part of the value of a dredge approach lies in its ability to identify important relationships

between characters by eliminating or equating parameters that lack significant support. This is

particularly useful for phenotypic complexes with multiple interacting characters (e.g., pollinator

syndromes). However, a dataset containing many characters also introduces a large state space with

numerous testable model structures. One solution to address this issue is to independently model each

character and then compile the results into a cohesive view. This approach is computationally tractable

and can yield well-behaved parameter estimates. But modeling each character independently ignores

correlated character evolution and dependent relationships can influence both biological inferences and

ancestral state estimation (Boyko and Beaulieu 2021). When the evolution of one character affects the

rate of change in another, independent modeling may lead to uncertainty in certain parts of the

phylogeny. By accounting for correlated character evolution, shared information between characters

can improve inference (Boyko and Beaulieu 2021). Of course, correlation between discrete characters

is a hypothesis that should first be tested. These tests should account for character-independent rate

variation (Boyko and Beaulieu 2023), which will introduce even more parameters than standard

independent models (Pagel 1994). Searching a state space that includes multiple characters and hidden

rate classes can be a daunting task even for experienced comparative biologists and it can be tempting

to rely on the most used default model sets. However, there is greater flexibility in potential dependent

relationships beyond typical fixed model sets. Correlation tests have traditionally compared

independent and correlated models, but not all characters need to show dependent relationships for

there to be evidence of correlation (see also Pagel and Meade 2006). A dredge framework is valuable

here, as it searches potential model structures and identifies only the necessary dependent relationships.

Choosing the "best" model

Model averaging is central to both Bayesian frameworks (e.g., Pagel and Meade 2006) and

frequentist multi-model inference (MMI; Burnham and Anderson 2002; Jhwueng et al. 2014) and

improves predictive power by integrating results over a set of plausible models. In contrast, model

selection aims to identify a single best-fitting model or a small set of top models, often guided by an

information criterion that balances model fit and complexity. While model averaging is powerful, its direct application following the use corHMMDredge faces some technical challenges, leading to a methodology centered on model selection and examination of the top-ranked models. The most critical challenge is the label switching problem inherent to hidden Markov models (Stephens 2000). For example, one model might assign high transition rates to hidden rate class 1, while another equally likely model assigns them to rate class 2. Averaging parameters across these models without resolving this arbitrary class assignment would produce meaningless estimates and incorrect conclusions. This issue can be mitigated in some cases, such as in diversification models, by focusing on tip rates rather than internal parameters (Title and Rabosky 2019; Vasconcelos et al. 2022).

However, a further complication with model averaging is the potential for pseudo-replication of model structures. The model space for more complex models (e.g., those with two rate classes or correlated character evolution) is inherently larger than that for simpler models (e.g., with one rate class or independent evolution). As a result, the total number of models available for averaging may disproportionately represent more complex models, even when there is roughly equal support for simpler alternatives. This can distort model averaging, as the AIC weights may reflect the sheer number of models rather than the actual support for distinct, biologically meaningful models. The most extreme example of this would be if identical models are refit, effectively double counting them. However, this problem can also occur if models are nearly identical, such as when two parameters are equated (e.g., k=7 vs. k=8). These models might still receive significant support despite being only minor variations of the most complex model. Handling this model structure pseudo-replication becomes particularly important for automated model selection.

This was not a problem when Burnham and Anderson (2002) originally proposed multi-model inference because they emphasized fitting a set of biologically plausible (and distinct) models (Burnham and Anderson 2002). Each model was justified based on specific hypotheses and therefore tested the contribution of different factors. For exploratory analyses on large model spaces, the rationale behind model averaging becomes less clear. In a dredge run, one is not testing specific hypotheses but rather comparing parameter estimates across many models. The final model of a dredge run does not necessarily reflect the combination of several well-supported biological models but is the byproduct of optimizing the bias-variance trade-off through AIC and encouraging sparse parametrizations through regularization. This is valuable from an inference perspective since the more complete a model set is, the less likely it is for unanticipated biases to creep into the results (Maddison and FitzJohn 2015; Rabosky and Goldberg 2015).

Nonetheless, the dredge approach and MMI need not be mutually exclusive. In cases where a

set of biologically plausible hypotheses are known beforehand, a MMI approach remains an excellent

option. And so long as procedures are clearly documented, there are advantages to combining the

approaches by considering which model structures are biologically reasonable and determining what

the optimal model structure could be for a given dataset. Furthermore, the most robust biological

inferences can then be drawn not by relying on a single "best" model, but by examining the top models

in the corHMMDredge set (e.g., those within ΔAIC < 2). This will allow researchers to assess whether

key parameter estimates and biological interpretations are stable across a range of well-supported

model structures. And because these models are ultimately phenomenological, the biological insights

we draw from the best-supported model, and how these insights shape our understanding of

evolutionary processes, are ultimately derived from subsequent interpretations and considerations in

concert with other pieces of evidence (Gardner and Organ 2021). This perspective leaves room for

quantifying the patterns in a general way via comparative modeling results, without necessarily forcing

too much biological meaning on any particular parameter estimate.

Conclusion

Determining which models to test in an empirical setting is an incredibly valuable process in

which biologists take hypotheses and formally structure them so that they can be compared. This

process can help clarify thinking around biologically relevant interactions between variables and will

lead to more robust inferences. However, if only a subset of model structures are considered (e.g., only

examining all-rates-different and equal-rates), there is a risk of overlooking plausible and important

model structures. This issue is exacerbated by the fact that the knowledge necessary to manipulate

PCMs is often difficult to acquire and may be hidden in highly technical texts, making it challenging

for biologists to explore and customize model sets effectively (Cooper et al. 2016). It is evident that a

comprehensive model set is important for trustworthy inferences in comparative biology (FitzJohn et

al. 2009; Rabosky and Goldberg 2015; Beaulieu and O'Meara 2016; Boyko and Beaulieu 2023), but

the growing complexity of discrete character models makes it challenging for users to determine which

models are potentially realistic and important to consider. Here I have shown how the corHMMDredge

framework may help alleviate this burden. This framework will enable biologists to focus more on

model interpretations rather than model construction and may even lead to the discovery of model

structures which imply unique hypotheses that would have not been considered otherwise.

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Data Availability

All the original data and scripts necessary to reproduce the analyses reported in this study can be

accessed through the Dryad link:

http://datadryad.org/share/HmmoM5OiQz2KUQr230La7ntdprGKF3kUR5rU0N-CZjs

Supplementary Material

Supplementary files including pseudocode for simulated annealing search and the relative model

support for the Old World monkey analysis is available at SYSBIO online.

References

Akaike H. 1974. A new look at the statistical model identification. IEEE Transactions on Automatic Control. 19:716–723.

Beaulieu J., O'Meara B., Oliver J., Boyko J. 2022. corHMM: Hidden Markov Models of Character Evolution. .

Beaulieu J.M., O'Meara B.C. 2016. Detecting Hidden Diversification Shifts in Models of Trait-Dependent Speciation and Extinction. Systematic Biology. 65:583–601.

Beaulieu J.M., O'Meara B.C., Donoghue M.J. 2013. Identifying Hidden Rate Changes in the Evolution of a Binary Morphological Character: The Evolution of Plant Habit in Campanulid Angiosperms. Systematic Biology. 62:725–737.

Blackbum D.G. 1999. Viviparity and Oviparity: Evolution and Reproductive Strategies. .

Boyko J.D., Beaulieu J.M. 2021. Generalized hidden Markov models for phylogenetic comparative datasets. Methods in Ecology and Evolution. 12:468–478.

Boyko J.D., Beaulieu J.M. 2023. Reducing the Biases in False Correlations Between Discrete Characters. Systematic Biology. 72:476–488.

Boyko J.D., O'Meara B.C., Beaulieu J.M. 2023. A novel method for jointly modeling the evolution of discrete and continuous traits. Evolution. 77:836–851.

Burley N. 1979. The Evolution of Concealed Ovulation. The American Naturalist. 114:835–858.

Burnham K.P., Anderson D.R. 2002. Model Selection and Multimodel Inference. New York, NY: Springer New York.

Clavel J., Aristide L., Morlon H. 2019. A Penalized Likelihood Framework for High-Dimensional Phylogenetic Comparative Methods and an Application to New-World Monkeys Brain Evolution. Systematic Biology. 68:93–116.

Cooper N., Thomas G.H., FitzJohn R.G. 2016. Shedding light on the 'dark side' of phylogenetic comparative methods. Methods in Ecology and Evolution. 7:693–699.

Felsenstein J. 1981. Evolutionary trees from DNA sequences: A maximum likelihood approach. J Mol Evol. 17:368–376.

Felsenstein J. 2004. Inferring phylogenies. Inferring phylogenies. p. 664–664.

Felsenstein J. 2012. A Comparative Method for Both Discrete and Continuous Characters Using the Threshold Model. The American Naturalist. 179:145–156.

Felsenstein J., Churchill G.A. 1996. A Hidden Markov Model approach to variation among sites in rate of evolution. Molecular Biology and Evolution. 13:93–104.

FitzJohn R.G., Maddison W.P., Otto S.P. 2009. Estimating Trait-Dependent Speciation and Extinction Rates from Incompletely Resolved Phylogenies. Systematic Biology. 58:595–611.

Gardner J.D., Organ C.L. 2021. Evolutionary sample size and consilience in phylogenetic comparative analysis. Systematic Biology. 70:1061–1075.

Gelman A., Carlin J.B., Stern H.S., Dunson D.B., Vehtari A., Rubin D.B. 2013. Bayesian Data Analysis. Boca Raton London New York: Chapman and Hall/CRC.

Hastie T., Tibshirani R., Friedman J. 2009. The Elements of Statistical Learning: Data Mining, Inference, and Prediction, Second Edition. New York, NY: Springer.

Hastie T.H., Tibshirani R., Wainwright M. 2015. Statistical Learning with Sparsity The Lasso and Generalizations. .

Hoerl A.E., Kennard R.W. 1970. Ridge Regression: Biased Estimation for Nonorthogonal Problems. Technometrics. 12:55–67.

Hrdy S.B. 1979. Infanticide among animals: A review, classification, and examination of the implications for the reproductive strategies of females. Ethology and Sociobiology. 1:13–40.

Huelsenbeck J.P., Ronquist F., Nielsen R., Bollback J.P. 2001. Bayesian Inference of Phylogeny and Its Impact on Evolutionary Biology. Science. 294:2310–2314.

Jhwueng D.-C., Huzurbazar S., O'Meara B.C., Liu L. 2014. Investigating the performance of AIC in selecting phylogenetic models. Statistical Applications in Genetics and Molecular Biology. 13:459–475.

Khabbazian M., Kriebel R., Rohe K., Ané C. 2016. Fast and accurate detection of evolutionary shifts in Ornstein–Uhlenbeck models. Methods in Ecology and Evolution. 7:811–824.

Kirkpatrick S., Gelatt C.D., Vecchi M.P. 1983. Optimization by Simulated Annealing. Science. 220:671–680.

Lartillot N. 2023. Identifying the Best Approximating Model in Bayesian Phylogenetics: Bayes Factors, Cross-Validation or wAIC? Systematic Biology. 72:616–638.

Lemey P., Salemi M., Vandamme A.-M. 2009. The Phylogenetic Handbook: A Practical Approach to Phylogenetic Analysis and Hypothesis Testing. Cambridge: Cambridge University Press.

Lewis P.O. 2001. A Likelihood Approach to Estimating Phylogeny from Discrete Morphological Character Data. Systematic Biology. 50:913–925.

Maddison W.P., FitzJohn R.G. 2015. The unsolved challenge to phylogenetic correlation tests for categorical characters. Systematic biology. 64:127–136.

Maddison W.P., Midford P.E., Otto S.P. 2007. Estimating a Binary Character's Effect on Speciation and Extinction. Systematic Biology. 56:701–710.

May M.R., Moore B.R. 2020. A Bayesian Approach for Inferring the Impact of a Discrete Character on Rates of Continuous-Character Evolution in the Presence of Background-Rate Variation. Systematic Biology. 69:530–544.

Metropolis N., Rosenbluth A.W., Rosenbluth M.N., Teller A.H., Teller E. 1953. Equation of State
Calculations by Fast Computing Machines. The Journal of Chemical Physics. 21:1087–1092.

O'Meara B.C. 2012. Evolutionary Inferences from Phylogenies: A Review of Methods. Annual Review
of Ecology, Evolution, and Systematics. 43:267–285.

Pagel M. 1994. Detecting correlated evolution on phylogenies: a general method for the comparative
analysis of discrete characters. Proceedings of the Royal Society of London. Series B:
Biological Sciences. 255:37–45.

Pagel M. 1999. The Maximum Likelihood Approach to Reconstructing Ancestral Character States of
Discrete Characters on Phylogenies. Systematic Biology. 48:612–622.

Pagel M., Meade A. 2006. Bayesian Analysis of Correlated Evolution of Discrete Characters by
Reversible⬚Jump Markov Chain Monte Carlo. The American Naturalist. 167:808–825.

Rabosky D.L., Goldberg E.E. 2015. Model Inadequacy and Mistaken Inferences of Trait-Dependent
Speciation. Systematic Biology. 64:340–355.

Rohlf F.J. 2001. Comparative Methods for the Analysis of Continuous Variables: Geometric
Interpretations. Evolution. 55:2143–2160.

Russell S., Norvig P. 2021. Artificial Intelligence: A Modern Approach. Hoboken, NJ: Pearson.

Sanderson M.J. 2002. Estimating Absolute Rates of Molecular Evolution and Divergence Times: A
Penalized Likelihood Approach. Molecular Biology and Evolution. 19:101–109.

Sillen-Tullberg B., Moller A.P. 1993. The Relationship between Concealed Ovulation and Mating
Systems in Anthropoid Primates: A Phylogenetic Analysis. The American Naturalist. 141:1–25.

Skinner A., Lee M.S., Hutchinson M.N. 2008. Rapid and repeated limb loss in a clade of scincid
lizards. BMC Evolutionary Biology. 8:310.

Stadler T. 2019. TreeSim: Simulating Phylogenetic Trees. .

Stephens M. 2000. Dealing With Label Switching in Mixture Models. Journal of the Royal Statistical
Society Series B: Statistical Methodology. 62:795–809.

Tibshirani R. 1996. Regression shrinkage and selection via the lasso. Journal of the Royal Statistical
Society Series B: Statistical Methodology. 58:267–288.

Title P.O., Rabosky D.L. 2019. Tip rates, phylogenies and diversification: What are we estimating, and
how good are the estimates? Methods in Ecology and Evolution. 10:821–834.

Tuffley C., Steel M. 1998. Modeling the covarion hypothesis of nucleotide substitution. Mathematical
Biosciences. 147:63–91.

Vasconcelos T., O'Meara B.C., Beaulieu J.M. 2022. A flexible method for estimating tip diversification
rates across a range of speciation and extinction scenarios. Evolution. 76:1420–1433.

Zhou Y., Gao M., Chen Y., Shi X. 2024. Adaptive Penalized Likelihood method for Markov Chains. .

Pseudocode for simulated annealing search

```
758    Pseudocode for simulated annealing search

759    FUNCTION SimulatedAnnealingSearch(data, phylogeny, initial_model_structure,
760    rate_cat, params):
761      // SA Initialization
762      FIT initial_model_structure to get initial_fit and initial_score
763      SET current_model = initial_fit
764      SET current_score = initial_score
765      SET best_model = current_model
766      SET best_score = current_score
767      CREATE list of visited_models, add initial_fit
768      SET current_temp = params.initial_temp
769      SET steps_since_accept_or_best = 0
770
771      // Main SA iterative loop
772      FOR iteration = 1 TO params.max_iterations:
773        // A. Check for restart condition
774        IF steps_since_accept_or_best >= params.restart_interval:
775          current_model = RANDOMLY_SELECT a model from visited_models
776          current_score = score of current_model
777          RESET steps_since_accept_or_best = 0
778          CONTINUE to next iteration
779        END IF
780
781        // B. Propose a new model structure
782        move_type = RANDOMLY_SELECT from {"drop", "merge", "free"}
783        IF move_type == "drop":
784          Calculate drop probabilities P(drop Qi) for all parameters in current_model
785          SELECT parameter to drop based on these probabilities
786          CREATE proposed_structure by removing the parameter
787        ELSE IF move_type == "merge":
788          Calculate merge probabilities P(merge Qi, Qj) for all pairs in current_model
789          SELECT pair to merge based on these probabilities
790          CREATE proposed_structure by merging the pair
791        ELSE IF move_type == "free":
792          Identify constrained transitions
793          RANDOMLY_SELECT one transition to free
794          CREATE proposed_structure by freeing the transition
795        END IF
796
797        // C. Evaluate the proposed model
```

```
798        proposed_fit = FIT model with proposed_structure
799        IF fit failed, CONTINUE to next iteration
800        proposed_score = GET information criterion from proposed_fit
801        delta_score = proposed_score - current_score
802
803        // D. Metropolis acceptance criterion
804        acceptance_prob = 1.0 IF delta_score <= 0 ELSE exp(-delta_score / current_temp)
805        IF random number [0,1] < acceptance_prob:
806          // Accept the new model
807          current_model = proposed_fit
808          current_score = proposed_score
809          ADD current_model to visited_models
810          IF current_score < best_score:
811            best_model = current_model
812            best_score = current_score
813            RESET steps_since_accept_or_best = 0
814          ELSE:
815            INCREMENT steps_since_accept_or_best
816          END IF
817        ELSE:
818          // Reject the new model
819          INCREMENT steps_since_accept_or_best
820        END IF
821
822        // E. Cool the temperature
823        current_temp = current_temp * params.cooling_rate
824        IF current_temp < a minimum threshold, BREAK loop
825      END FOR
826
827      RETURN visited_models
828    END FUNCTION
829
830
```

Table S1 – Relative model support for the Old World monkey analysis. This table includes the number of parameters (np), the number of rate categories (nRateCat), the log likelihood value for a given model (lnLik), the Akaike Information Criterion value (AIC), the difference from the models AIC value to the best AIC value (dAIC) and the AIC weight. These are the unique models which were accepted during the simulated annealing routine. Many more models were attempted but not accepted during the search.

np	nRateCat	lnLik	AIC	dAIC	AICwt
4	1	-42.756941	93.5138814	5.59242874	0.0084073
7	1	-42.823622	99.6472448	11.7257921	0.00039157
3	1	-42.791402	91.5828036	3.66135092	0.02207928
2	1	-43.055279	90.1105582	2.18910557	0.04609766
3	1	-43.570587	93.1411748	5.21972214	0.01012952
2	1	-43.231785	90.4635702	2.54211759	0.03863877
24	2	-40.360785	128.721571	40.800118	1.90E-10
23	2	-40.505853	127.011707	39.090254	4.47E-10
22	2	-41.620411	127.240823	39.31937	3.99E-10
6	2	-40.839915	93.6798291	5.75837645	0.00773787
17	2	-40.840757	115.681515	27.7600622	1.29E-07
14	2	-40.83998	109.67996	21.758507	2.60E-06
4	2	-41.181067	90.3621331	2.4406804	0.04064902
3	2	-41.2575	88.514999	0.59354638	0.10236468
12	2	-40.673954	105.347907	17.4264544	2.26E-05
12	2	-40.673961	105.347921	17.4264684	2.26E-05
6	2	-40.504566	93.0091319	5.08767929	0.01082085
8	2	-40.948669	97.8973384	9.97588573	0.00093929
6	2	-40.948663	93.8973261	5.97587344	0.00694053
5	2	-41.189251	92.3785021	4.45704939	0.01483205
3	2	-41.533733	89.0674668	1.1460141	0.07765748
11	2	-40.839923	103.679846	15.7583936	5.21E-05
9	2	-40.839917	99.6798338	11.7583811	0.00038525
7	2	-40.839914	95.679829	7.75837634	0.0028466
4	2	-42.052089	92.1041774	4.18272478	0.01701257
3	2	-42.192072	90.3841433	2.46269067	0.04020412
3	2	-41.935946	89.8718921	1.9504394	0.05194031
5	2	-40.839919	91.6798375	3.7583848	0.02103363
3	2	-41.175615	88.3512292	0.42977656	0.11109955
3	2	-42.56508	91.130161	3.20870832	0.02768694
2	2	-43.114338	90.2286759	2.30722325	0.04345402
3	2	-41.300881	88.6017622	0.68030953	0.09801888
3	2	-42.779014	91.5580279	3.63657528	0.02235449
3	2	-43.381054	92.7621085	4.8406558	0.0122434
4	2	-41.620225	91.240449	3.31899638	0.0262015
2	**2**	**-41.960726**	**87.9214527**	**0**	**0.13773271**