1 **Elevational and Oceanic Barriers Shape the Distribution, Dispersal and Diversity of Aotearoa's**
2 **Kapokapowai (*Uropetala*) Dragonflies**
3

Authors: Ethan R. Tolman*[1,2], Chris D. Beatty*[2,3], Aaron Goodman*[2,4,], Priscilla Ramchand*[2,5], Tara
McAllister[6], Kelly Reyes[7], Akifa Zahara[7], Kassandra Taveras[7], Vincent Wade[7], John Abbott[8], Seth
Bybee[9], Paul B. Frandsen[10], Robert Guralnick[11], Manpreet K. Kohli[1,12], Jessica Ware[†2], Anton Suvorov[†1]
*Equal contributing authors to the manuscript
[†]Senior authors
[1]Department of Biological Sciences, Virginia Tech, Blacksburg, Virginia
[2]Division of Invertebrate Zoology, American Museum of Natural History, New York, New York
[3]Program for Conservation Genomics, Department of Biology, Stanford University, Stanford, California
[4]The Graduate Center, The City University of New York, New York, New York
[5] Brooklyn College, The City University of New York, Brooklyn, New York
[6]Te Wānanga o Aotearoa, New Zealand
[7]Science Research Mentorship Program, American Museum of Natural History, New York, New York
[8]Alabama Museum of Natural History and Department of Research and Collections, The University of
Alabama, Tuscaloosa, Alabama
[9]Department of Biology, Brigham Young University, Provo, Utah
[10]Department of Plant and Wildlife Sciences, Brigham Young University, Provo, Utah
[11]Florida Museum of Natural History, University of Florida, Gainesville, Florida
[12]Department of Natural Sciences, Baruch College, New York, New York

Author Contributions
Ethan R. Tolman: Data acquisition, data analysis, funding, writing, editing.
Chris D. Beatty: Data acquisition, data analysis, writing, editing.
Aaron Goodman: Data acquisition, data analysis, writing, editing.
Priscilla Ramchand: Data acquisition, data analysis, funding, writing, editing.
Tara McAllister[:]: Writing, editing.
John Abbott: Funding, writing, editing.
Kelly Reyes: Data analysis, writing, editing.
Akifa Zahara: Data analysis, writing, editing.
Kassandra Taveras: Data acquisition, data analysis, writing, editing.
Vincent Wade: Data acquisition, data analysis, writing, editing.
Paul B. Frandsen: Funding, writing, editing.
Robert Guralnick: Funding, writing, editing.
Seth Bybee: Funding, writing, editing.
Manpreet K. Kohli: Writing, editing, supervision.
Jessica Ware: Data acquisition, data analysis, funding, writing, editing, supervision.
Anton Suvorov: Data analysis, writing, editing, supervision.
40

Acknowledgements
We would like to thank Thomas Beatty, Kathleen Harding, and Mahealani Kaloi for their support of this
project. We would also like to acknowledge Dr. Milen Marinov for his generosity in providing Odonata,
as well as the Department of Entomology at the Smithsonian Institution's National Museum of Natural

History, Florida State Collection of Arthropods, and American Museum of Natural History for providing
specimens used in this analysis.
47

Funding
This work was funded by the Graduate Student Research Award from the Society of Systematic
Biologists, the Tow Undergraduate and Graduate International Research Stipends from The City
University of New York Brooklyn College, the Maxwell/Hanrahan Award from the American Museum of
Natural History, and by the National Science Foundation (NSF) Awards [Bybee: 2002432; Ware:
2002473; Guralnick: 2002457; Abbott: 2002489: Genealogy of Odonata (GEODE): Dispersal and color
as drivers of 300 million years of global dragonfly evolution].

55
56 **Data availability**
57 Filtered vcf files with and without *P. raptor*, and log and xml files from the BEAST2 runs have been
58 uploaded to figshare (10.6084/m9.figshare.27896910). The Raw reads have been uploaded to the
59 sequencing read archive (bioproject PRJNA1190089).
60
61 **Conflict of Interests Statement**
62 The authors declare no conflicts of interest.
63

Abstract
Mountains and islands are both model systems for studying the biogeography of diversification and
population fragmentation. Aotearoa is an excellent location to study both phenomena due to alpine
emergence and oceanic separation. While it would be expected that separation across Te Moana o
Raukawa and elevation gradients are major barriers to gene flow in aquatic insects, such hypotheses have
not been thoroughly explored in these taxa. Here, we show that mountains and oceanic separation
function as semi-permeable barriers for Kapokapowai dragonflies. We show that, although Te Moana o
Raukawa, is likely responsible for some of the genetic structure observed, speciation has not yet occurred
in populations separated by the strait. Although there is no evidence that they are an impervious barrier,
Kā Tiritiri-o-te-Moana do represent a major barrier to gene flow between named species. The distribution
of alpine Kapokapowai can also not be explained by the ancestral populations simply rising with the
uplifted tectonic plates. Although further research is needed, our findings suggest that the ancestral
Kapokapowai colonized alpine habitats after their formation, and then radiated back out to lowlands.
These findings suggest that aquatic insects could be an exciting new frontier in the study of the
biogeography of Aotearoa.

Keywords
Anisoptera, Epiprocta, biogeography, population genomics, Petaluridae, Islands, Odonata

Table 1: Reference table of colonial and Māori names of localities and taxa.

Geographic localities

Māori name	Colonial name
Aotearoa	New Zealand
Te Moana o Raukawa	Cook Strait
Te Ika-a-Māui	North Island
Te Waipounamu	South Island
Kā Tiritiri-o-te-Moana	Southern Alps

Population Names

Māori names used in this work	Scientific name and locality
Kapokapowai North (KN)	*Uropetala carovei*, Te Ika-a-Māui
Kapokapowai South (KS)	*Uropetala carovei*, Te Waipounamu (below 4000 ft)
Kapokapowai Montane (KM)	*Uropetala chiltoni*, Kā Tiritiri-o-te-Moana

Introduction

Islands are widely recognized for their unique role in shaping biodiversity (Barreto et al., 2021; Conway & Olsen, 2019; Gleditsch et al., 2022; Wilson, 2009). The Theory of Island Biogeography by MacArthur and Wilson (1963) summarized several of these phenomena, and this theory was later extended to mountaintop habitats (Wyckhuys et al., 2022). The islands of Aotearoa (New Zealand; see table 1 for nomenclature) have often been considered as a natural laboratory to study both island and mountain biogeography (Shepherd et al., 2024; Thomas et al., 2023; Wallis & Buckley, 2024; Waters & Craw, 2006). This is due to the opening and closing of the Te Moana o Raukawa, referred to by some post-European arrival as the Cook Strait, which separates Te Ika-a-Māui and Te Waipounamu (also known as the North and South Islands respectively) and last opened 500,000 years ago (Lewis et al., 1994), and the uplift of Kā Tiritiri-o-te-Moana (the Southern Alps), a relatively young mountain range on Te Waipounamu (South Island) (Table 1).

Despite its young age, Te Moana o Raukawa (the Cook Strait) has been proposed as a driver of allopatric speciation and population divergence in several taxa including amphipods (Stevens & Hogg, 2004), the Weka (terrestrial birds) and their ectoparasites (Trewick et al., 2017), and skink lizards (O'Neill et al., 2008). The uplift of the Kā Tiritiri-o-te-Moana mountain range, known colonially as the Southern Alps, likely opened niches and acted as a driver of diversification in flora (Thomas et al., 2023)

and fauna (Buckley et al., 2024) of Te Waipounamu. Most lineages of alpine flora arrived in the later Tertiary, and the environmental upheaval in the Pliocene and Pleistocene drove diversity in these lineages (Thomas et al., 2023). Most alpine invertebrate fauna are relatively young, with ages corresponding to the increased uplift of the Kā Tiritiri-o-te-Moana range 5 million years ago (Ma), and the development of a permanent snow line just over 1 Ma (Buckley et al., 2024). Similar to the flora, it is broadly understood that most invertebrate fauna evolved from lineages already on the island as niches opened with the increased elevation (Buckley et al., 2024), although it has been hypothesized that invertebrates rode tectonic lift to higher elevation (Heads, 2017).

Although aquatic insects have been used to better understand the past climate and geology of Aotearoa, they have been relatively underrepresented in biogeographic and broader biodiversity work (Leschen et al., 2023) and knowledge about their biogeography remains limited for many taxa. One such group of aquatic macroinvertebrates are the dragonflies first referred to as Kapokapowai (among other names) by Māori (Grant, 2014), and later, named *Uropetala* by colonial European scientists (Selys 1858).

According to Māori cosmology, dragonflies and other insects are considered the children of Tāne Māhuta (the god of forests, birds and insects). Dragonflies appear in whakaairo (traditional carvings which often adorn the walls of Māori meeting houses) and waiata (songs) of Māori (Indigenous Peoples of Aotearoa New Zealand). Stories about dragonflies also feature in pūrakau (legends) belonging to specific iwi (tribes) and hapū (subtribes). All insects, including dragonflies, were created before humans in Māori genealogical histories and are therefore tuakana (roughly translated as an elder family member) and are respected accordingly. Kapokapowai or Kapowai are names commonly-used for the Petaluridae of Aotearoa, which loosely translates to water snatchers, though there are other recorded names including Tītīwaiora, Kihihiwaru and Uruururoroa (Grant, 2014). As this work was conducted with the express permission of the iwi who own the lands of Aotearoa, we refer to these dragonflies by the Māori name, Kapokapowai, in our work here. We also note that the name Kapokapowai was used to describe these dragonflies in the literature, three years prior to the first use of *Uropetala* by Selys (Grant, 2014; Taylor, 1855, 1870), and it has continued to be used to refer to dragonflies in this genus in scientific literature (Durrant, 2024; Fong, 2024; E. R. Tolman et al., 2024). However, we are not making any change to the binary taxonomic nomenclature of this genus here.

Kapokapowai represent one of only five genera of Petaluridae, a family that comprises 11 currently described species (E. R. Tolman et al., 2024; Ware et al., 2014). Kapokapowai includes two currently described species, *Uropetala carovei* (Selys, 1843) which is thought to inhabit lower elevations of both Te Ika-a-Māui and Te Waipounamu (North and South Islands respectively), and *Uropetala chiltoni* (Tillyard, 1921), which inhabits higher elevations of Te Waipounamu (South Island). We refer to the populations by their Māori name in this work, referring to *U. chiltoni* as Kapokapowai Montaine (KM), and *U. carovei* from either island as Kapokapowai North (KN) and Kapokapowai South (KS)(Table 1), representing the three putative species in the Kapokapowai species complex.

Debate about whether this complex should appropriately be considered to have two species has been ongoing for decades (Andress, 1998; L. S. Wolfe, 1952; Winstanley & Rowe, 1980). According to Tillyard's original description, KM is characterized by large pale blotches on its labrum, which are not present in KN, and by black femur leg segments, which are brown in KN. However, variation in the splotches of the labrum have been used to suggest that hybrid populations between KM and KS can be found at the north end of Te Waipounamu (South Island) (Winstanley & Rowe, 1980). Behavioral work suggests that female KM lay all their eggs in one plant, and in repeating this egg laying process females tend to badly damage their wings; such damage has not been seen in KN or KS (W. J. Winstanley, 1981).

This behavioral work should be replicated to confirm that this is a species–wide behavior. It has also been
suggested that KN and KS may be different species, as Kapokapowai North on Te Ika-a-Māui may have
broader superior appendages and larger imagos than their counterparts on Te Waipounamu (L. S. Wolfe,
1952). Two different karyotypes have been identified for this genus, but it is unclear if this is due to
laboratory error (Kuznetsova & Golub, 2020); no other attempts have yet been made to determine the
number of species in this genus using molecular data. Based on known morphological variation and
behavioral data (L. S. Wolfe, 1952; Winstanley & Rowe, 1980),, the dragonflies known as Kapokapowai
are hypothesized here to comprise between one and three species, the diversification of which was shaped
by both Te Moana o Raukawa (Cook Strait) and Kā Tiritiri-o-te-Moana (Southern Alps).
Here, we use population and functional genomics, alongside scanning electron microscopy and
environmental niche models to determine the number of species and the population dynamics within
Kapokapowai and to study how Kā Tiritiri-o-te-Moana and Te Moana o Raukawa may have driven
speciation in this genus.
164
Materials and Methods
Population genomics
To determine the population structure of Kapokapowai using a sampling of individuals, we sequenced 31
individuals, comprising 16 KM and eight KN from Te Ika-a-Māui, as well as eight KS from Te
Waipounamu, and seven *Phenes raptor* (Supplementary table 1), using a 20kb anchored-hybrid
enrichment dataset (E. R. Tolman et al., 2024). We also utilized reads previously generated for this
dataset for two KM from Te Waipounamu, and one KN from Te Ika-a-Māui (E. R. Tolman et al., 2024).
We mapped these reads to the soft-masked assembly of an individual KS (E. R. Tolman et al., 2023) with
bwa2 (0.7.17-r1188) (H. Li & Durbin, 2009), and calculated mapping statistics, average depth and
converted the output to a sorted bam file with samtools (1.16.1) (Danecek et al., 2021). We called variants
with bcftools (1.15.1) (Danecek et al., 2021), using the options: bcftools mpileup -a AD,DP,SP -Ou -f
reference.fasta. Input_bam | bcftools call -f GQ,GP -mO z -o output. We then filtered the resulting vcf file
for a minimum quality score of 30, and a minimum and maximum depth of 10 and 350 respectively to
capture the region targeted with hybrid probes and the adjacent flanking region that is also sequenced and
often more variable.
We then performed a PCA analysis and calculated the F_{ST} between and within each of the three
putative species using vcftools (0.1.16) (Purcell et al., 2007). To visualize admixture among populations
we generated admixture models assuming 1-7 ancestral populations with admixture v1.3.0 (Skotte et al.,
2013) and evaluated the models with cross-validation error. The best fitting admixture-model and the
PCA plots were generated in R using mapmixture (1.1.0) (Jenkins, 2024).
To test for isolation by distance, we used genetic differences calculated with VCFtools (Danecek
et al., 2021), and performed a Mantel test using the vegan package in R v2024.04.2+764 (Oksanen et al.,
2024), using 1000 permutations to determine the significance of the correlation. We also tested for
introgression between KM and KN and KS from either island with an ABBA-BABA test from Dsuite
v0.5 r57, which is optimized to work on lineages in shorter evolutionary time scales (Malinsky et al.,
2021). In this test, seven sequenced individuals of *P. raptor* (the sister genus to *Uropetala* (E. R. Tolman
et al. 2024)) were used as the outgroup taxa.
We used the speedemon v.1.1.1 (Douglas & Bouckaert, 2022) pipeline on 395 biallelic SNPs
from our dataset, using snapper v1.1.4 (Stoltz et al., 2021) and BICEPS v1.1.2 (Bouckaert, 2022) in
BEAST2 v2.7.7 with a chain length of 10,000,000 to identify species delimitations using our dataset. In

the analysis we designated KN, KS and KM as three separate populations, and used the mean pairwise
percent genetic divergence between individuals of KM (~.54), as calculated with from the VCF file using
the pandas (McKinney, 2010) library for data manipulations and numpy (Harris et al., 2020) for
numerical computations, as the prior mean on theta, and used an alpha of 2.0, and beta of 0.26942 to
achieve this mean in the Gamma distribution of the theta prior, and followed the developer's suggestions
for all other parameters in the BEAST2 run (Douglas & Bouckaert, 2022). We independently ran the
analysis two times to check for convergence, and tested for adequate mixing of and model convergence
using Effective Sample Size (ESS), both in TRACER v1.7.2 (Rambaut et al., 2018).
203

Morphological analysis
To identify potential morphological synapomorphies between the putative species of Kapokapowai we
evaluated the male secondary genitalia. We specifically looked at the vesica spermalis, or sperm pump.
This structure is located on the ventral side of the second abdominal segment in male Odonata, to which
sperm is transferred prior to copulation, and may provide informative morphological characters for
species determination (Kennedy, 1922; May, 1997; Ware, 2008). The vesica spermalis of Odonata is
composed of four segments that are variable between species in their relative size and shape. In
Petaluridae, a barb on the second segment near the junction with the third segment and the horns on the
lateral side of the third segment have been used to support intrageneric difference (E. Tolman, 2024). We
imaged the vesica spermalis from two males from each putative species to try and identify more definitive
diagnostic features for species delimitation. We followed the protocol for scanning electron microscopy
outlined by Tolman et al. (2024). Briefly, penes were mounted laterally and dorsally on aluminum stubs
and coated with gold palladium using a Cressington 108E sputter coater and specimens were then imaged
using an S-7400 Hitachi scanning electron microscope.
218

Environmental niche modeling
Occurrence Records
We acquired occurrence records of adult KN, KS, and KM from the Global Biodiversity Information
Facility (GBIF)(available in supplementary data). We selected occurrences from GBIF possessing
preserved museum samples and research grade observations, which include verified latitude and longitude
coordinates, a photograph of the sighting, observation date, and at least ⅔ agreement on species
identification by the community. We filtered occurrences by removing sightings from erroneous localities
(middle of the ocean, locations of large museums). We divided occurrences for KN and KS between Te
Ika-a-Māui and Te Waipounamu (North and South Islands) to be used as individual models
(Supplementary Table 3).
229

Environmental Data
We acquired averaged sets of environmental predictor variables for modeling consisting of purely
bioclimatic variables. All analyses were conducted using the statistical programming language R v. 4.1.2.
We acquired environmental rasters at 2.5 arc-second resolution (~5km at the equator) from the CHELSA
climate database v2.1 (Karger et al., 2017, 2023) . We downloaded the 'Anthropocene' bioclimatic
dataset (1979 – 2013) consisting of 19 bioclimatic variables which follow Worldclim v2 (Fick &
Hijmans, 2017). We omitted bioclimatic variables 8 and 9 after visual inspection of interpolation
discontinuities (Booth, 2022). Although such spatial artifacts were minor within our initial study extent,

we observed major breaks of climate smoothing when we extrapolated our model to a larger spatial extent
within the last glacial maximum and interglacial periods.
240
Model Construction
We spatially thinned occurrences to the resolution of our environmental variables to reduce the effects of
sampling bias and artificial clustering (Veloz, 2009). We thinned occurrences by 5km to match the spatial
resolution of our environmental data. We chose study extents for occurrences of KN and KS found in Te
Ika-a-Māui and Te Waipounamu, defined as polygons around all localities between both islands
respectively; we chose the same study extent for occurrences of KM found in Te Ika-a-Māui. We chose
these study extents to include potential suitable habitat, while excluding large areas outside the species'
dispersal limitations (Peterson & Lieberman, 2012). Within this extent, we randomly sampled 50,000
background points for modeling and extracted their environmental values. We used these values to
calculate correlations between variables using the 'vifcor' and 'vifstep' functions in the usdm package
(Naimi, 2023) and filtered out variables with correlation coefficients higher than 0.7 and a Variance
Inflation Factor (VIF) threshold of 10, both standard cutoffs in such analyses (Goodman et al., 2024).
To model the distribution of putative Kapokapowai species, we used the presence-background
algorithm Maxent v3.4.4 (Phillips et al., 2017), using the R Package ENMeval 2.0.0 (Kass et al., 2021)
for model building, parameterization, evaluation with different complexity settings, and reporting of
results. We partitioned our data using the 'checkerboard2' strategy. To prevent our model from
extrapolating beyond the bounds of training data, we clamped our models by omitting ranges of
environmental data which fall outside of the training data. All final models were fitted to the full dataset.
We tuned model complexity to find optimal settings using different feature classes, including linear (L),
quadratic (Q), and hinge (H) as well as regularization multipliers 1 through 5 (higher numbers penalize
complexity more) (Radosavljevic & Anderson, 2014; Warren & Seifert, 2011).
We assessed our model using averages of threshold-dependent (omission rate) and
threshold-independent (Area Under the Receiver Operating Characteristic or AUC) discrimination
performance metrics calculated on withheld validation data (Warren & Seifert, 2011). The 10th-percentile
omission rate sets a threshold as the lowest suitability value for occurrences after removing the lowest
10% suitability values (Kass et al., 2021; Radosavljevic & Anderson, 2014). Validation AUC is a
measure of discrimination accuracy that can be used to make relative comparisons between ENMs with
different settings fit on the same data (Lobo et al., 2008; Radosavljevic & Anderson, 2014). Finally, we
checked the results against the Akaike Information Criterion with correction for small sample sizes
(AICc; calculated on the full dataset) (Warren & Seifert, 2011). To investigate model behavior, we
examined marginal response to suitability. Marginal response curves show the modeled relationship of
each variable individually with the occurrence data when all other variables are held constant (Phillips et
al., 2017).
We made habitat suitability predictions for KM, and KN and KS from both islands using our
'Anthropocene' environmental predictor variables. We projected our modern-day models to a new study
extent to encapsulate the potential habitat of Kapokapowai within the historic past, defined as a polygon
around both Islands of Aotearoa. We generated thresholdless predictions by converting raw predictions to
a scale of 0-1 to approximate probability of occurrence using the 'cloglog' transformation (Phillips et al.
2017). We also generated a threshold prediction, calculated from the 10-percentile omission rate from our
model evaluation. To estimate habitat suitability of Kapokapowai within the geologic past, we generated
predictions using environmental predictor variables from the PaleoClim dataset (Brown et al., 2018). We

generated predictions within three distinct time periods: Mid-Holocene (8.3 – 4.2ka), Last Glacial
Maximum (21ka), and the Last Interglacial (130ka).
284
Ordination Analysis
To determine niche differentiation between putative species, we compared niche overlap in occurrences
of KN and KS from Te Ika-a-Māui and Te Waipounamu using an ordination framework by first reducing
dimensionality within the datasets via a Principal Component analysis (PCA). Using the 'espace_pca'
function in the *Wallace* v2.0.5 and *ade4* package v1.7 in R (Dray & Dufour, 2007; Kass et al., 2018)),
we generated a Principle Components Analysis (PCA) using environmental variables from Kapokapowai
occurrences and plotted with correlation loadings to infer the degree of influence particular environmental
variables possess in the distribution of background points within niche space; we chose bioclimatic
variables which were uncorrelated and shared among both *Kapokapowai* maxent models. Using the
'espace_occDens' function in the package *ecospat* v3.2 (Di Cola et al., 2017, p. 2) an occurrence density
grid was estimated for both the environmental values at each occurrence point and background extent
points using a kernel density estimation approach. Niche overlap between occurrence density grids of
environmental values at each occurrence point and background points were compared using Schoener's D
(Schoener, 1968) 'espace_nicheOv' function. Finally, using the 'ecospat.plot.overlap.test' function, we
conducted a niche similarity test in which Kapokapowai niches from both islands are randomly shifted in
the background extent and permuted 1000 times, and the niche overlap between the two are recalculated.
A p-value <0.05 indicates that the niches from both time periods are significantly similar with each other.
302
Results
Population genomics
After filtering for mapping quality and depth, we retained 365 SNPs for the population structure analysis.
The weighted F_{ST} values were estimated at 0.23 between KM and KS, 0.29 between KM and KN and
0.11 between KN and KS (Table 2). We then separated the data into subsets as follows by their
distributions along PC1 (Fig 1A): KM (all individuals), KM (negative PC axes), KN (Te Ika-a-Māui) and
KS (Te Waipounamu). The F_{ST} values differed when only a subset of KM was used (Supplementary
Table 3). The centroid of KM was distal from the centroids of KN and KS across PC1, while the centroid
of KN and KS differed along PC2 (Fig. 1A). Despite the differing centroids across PC1 and PC2, there
was overlap in the space occupied between all three putative species (Fig. 1A). Because of the low cross
validation-error (Supplementary table 4), and the three clusters identified in the PCA analysis, we
assumed three ancestral populations in our main admixture analysis (Fig. 1B), but also provide
assumptions of K=2, and K=4 (Supplementary fig. 1). Ancestral population 1 was only present in KM,
while ancestral populations 2 and 3 were present in all three putative species (Fig. 1B). There was no
evidence that geographic separation was correlated with genetic distance in this dataset (pearson
correlation coefficient r=0.02462, p=0.322), however, the ABBA-BABA test showed moderate evidence
of gene flow from KM to KS (p=.03, f4-ratio=.45, D-statistic=.11). All parameters in the speedemon
analysis showed adequate mixing (ESS>400), and model convergence (see supplementary data). The
species were recovered as currently described with very high (99%) posterior support.
322
323
324
325

326   Figure 1: Population Genomics of Kapokapowai

327
328   Fig. 1: (A) PCA of Kapokapowai, colored by putative species: Kapokapowai Montane (Blue), KN
329   (Green) and KS (Teal). (B) Admixture analysis of Kapokapowai assuming three ancestral populations,
330   with the ancestral populations 1-3 colored as blue, green, and teal respectively per each sampled
331   individual. (C) Adult Female Kapokapowai North from Te Ika-a-Māui. (D) Nymphs of Kapokapowai
332   Montane.
333
334

Table 2: F_{st} values between putative species of Kapokapowai			
Putative species 1	Putative species 2	Mean F_{ST}	Weighted F_{ST}
Kapokapowai Montane	Kapokapowai South	0.20	0.23
Kapokapowai Montane	Kapokapowai North	0.27	0.29
Kapokapowai North	Kapokapowai South	0.05	0.11

335
336
337 *Morphological analysis*
338
339 Figure 2: Male Secondary Genitalia of the Three Hypothesized Kapokapowai Species

340
341 Fig. 2: SEM images of male penes from six sampled Kapokapowai (two from each putative species).
342 Divergent features include (1) curves at the end of the horns only found in Kapokapowai Montane, (2) the
343 barb near the joint of the second and third segments, and (3) a knob found near the barb from one
344 individual from Kapokapowai North.
345
346 The "horns" of KM (see figure 2, feature 1) are down curved at the tips in a way not observed in any of
347 the three KN or KS individuals (Fig. 3). The angle and curvature of the barb (figure 2, feature 2), and the
348 presence of a "knob" near it (figure 2, feature 3) differs between individuals, but does not appear to be
349 fixed between any populations. One individual of KN had a knob near the barb.
350
Ecological Niche Modeling
Optimal model settings varied considerably for Kapokapowai based on 10-percentile omission rate and
mean validation AUC metrics. Based on the results of the collinearity analysis, we used the following six
predictor variables to build all models: Isothermality (bio03), temperature seasonality (bio04), maximum
temperature of the warmest month (bio05), precipitation seasonality (bio15), precipitation of the warmest
quarter (bio18), and precipitation of the coldest quarter (bio19). Models varied in complexity with
expressing either hinge-loss, or linear-quadratic feature classes with regularization multipliers of 3-4

(higher complexity penalty). Mean validation AUC was adequate for KS and KM (0.75 and 0.86
respectively), while 10% omission rate was low relative to its expected value of 0.1 (0.11 and 0.09
respectively) (Supplementary Table 2). Furthermore, optimal models for KS and KM possessed low
amounts of non-zero lambdas (coefficients) and AICc values suggesting low amounts of model
overinflation (Supplementary Table 2). Optimal models for KN performed poorly, possessing an
unacceptably low AUC value (0.44) and high 10% omission rate (0.16) (Supplementary Table 2).
364   Optimal models retained 4 – 6 predictor variables after regularization, and relied mainly on
contributions from Isothermality (bio03), temperature seasonality (bio04), maximum temperature of the
warmest month (bio05), and precipitation of the wettest and driest quarters (bio18 and bio19 respectively)
(See Supplemental Fig. 1). Response curves of climate variables revealed differing relationships with
suitability among Kapokapowai models (Supplementary Fig. 1). Response curves for KN expressed
jagged optimal ranges, with habitat suitability not following a smooth or continuous gadient. Temperature
Seasonality (bio03) and precipitation of the coldest quarter expressed positive linear/hinge relationships
with suitability among all three Kapokapowai models, while precipitation of the wettest quarter (bio18)
expressed negative linear/hinge relationships with suitability for KN and KS; precipitation of the wettest
quarter possessed no relationship in KM. Isothermality (bio03), and maximum temperature of the
warmest month (bio05) expressed opposite relationships with suitability within Kapokapowai.
Isothermality (bio03) expressed positive linear or linear/quadratic relationships with suitability within
KM and KN, but a negative linear relationship with KS. Maximum temperature of the warmest month
(bio05) expressed negative linear or quadratic/linear relationships with suitability within KM and KS but
a positive linear relationship within Kapokaowai South (Supplementary Fig. 1).
379   Predictions using our 'Anthropocene' environmental variables for KS within Te Waipounamu
revealed areas of high suitability along the coast (Westland, Nelson and Marlborough provinces) (Fig
3D). Suitability was highest for KM across the Southern Alps (interior Nelson and Marlborough
provinces) (Fig. 3A). Predictions for Kapokaowai North within Te Ika-a-Māui revealed areas of high
suitability within the interior (Wellington and Hawke's Bay, along the Tararura, Ruahine, and
Kaimanawa Mountain Ranges), as well as the Northern portion of the island within Auckland (Fig 3G).
Threshold predictions reflect our 'cloglog' predictions (See Supplementary Fig. 2).
386   Predictions generated from our paleoclimate variables demonstrated variability in suitability over
time for Kapokapowai compared to our modern-day 'Anthropocene' variables. For KM, suitability
expands within the mid-Holocene (8.3 – 4.2ka), encompassing most of Te Waipounamu except for the
coastal margins (Fig 3H.). Projection to the Te Ika-a-Māui within the mid-Holocene indicates suitable
habitat within the island's center (Te Ika-a-Māui Volcanic Plateau, Tararua Range, and Raukumara
Range) (Fig 3H.). Within the LGM, suitability contracts within Te Waipounamu, with suitable habitat
only being located within a small portion of the Kā Tiritiri-o-te-Moana (Southern Alps) and Kaikōura
Mountain Ranges (Fig 3I.). Within Te Ika-a-Māui, suitability expands, encompassing most of the western
and southern mountain ranges. Although a land bridge connected both Islands during the LGM (Te
Moana o Raukawa), suitability was low (Fig 3I.) while our threshold predictions suggest suitable habitat
across the strait (See Supplemental Figure 3).
397   For KS within Te Waipounamu, suitability contracts within the mid-Holocene, being restricted to
a small portion of the coast (northern Westland) (Fig 3E.). When projected to Te Ika-a-Māui, suitability is
high within the Northern Cape and southwestern cape (Taranaki Province), and Auckland (Fig 3E).
Within the LGM, suitability expands encompassing the northern coastal regions of Te Waipounamu
(Westland and Nelson), as well as the Northern Cape, Taranaki Province, and Auckland within Te Ika-a-

Māui (Fig 3F.). Suitability across the Te Moana o Raukawa is intermediate, with a few pockets of high
suitability (Fig 3F.) while our threshold predictions suggest suitable habitat across the strait (See
Supplemental Figure. 3).
For KN within Te Ika-a-Māui, suitability contracts within the mid-Holocene, with only a few
pockets of high suitability within the southern coasts (Tararua Ranges), and Auckland (Fig. 3B.). When
projected to Te Waipounamu, suitability is highest along the coastal ranges of southern Westland and
Otago. Within the LGM, suitability expands to include most of the southern Mountain Ranges within Te
Ika-a-Māui, as well as Kaimanawa Mountain Ranges within the middle of the Island (Fig 3C). Within Te
Waipounamu, suitability slightly expands southward into the coastal ranges of Otago (Fig 3D). Suitability
within the Te Moana o Raukawa remains intermediate with a small pocket of high suitability off the coast
of Taranaki (Fig 3D.) while our threshold predictions suggest suitable habitat across the strait (See
Supplemental Figure 3).
414
Ordination Analysis
Environmental variables for KN and KSwithin Te Ika-a-Māui and Te Waipounamu expressed an
intermediate degree of overlap in ordination analysis, but were statistically more similar than random
(Fig. 4). Within our PCA analysis, the first principal component explained ~34% of the variance with
PC2 explaining ~32%. Isothermality (bio03), temperature seasonality (bio04), and precipitation of the
coldest quarter (bio19) loaded strongly on PC1, while precipitation of the warmest and coldest quarters
(bio18 and bio19) loaded strongly on PC2 (Table 3). Environmental niche overlap for Kapokapowai was
intermediate between Te Ika-a-Māui and Te Waipounamu (Schoener's D = 0.24), with niche similarity
tests showing observed overlaps being higher than 95% of simulated overlaps (p = 0.01), supporting niche
similarity between both islands (Fig. 4).
425
426

Species	PC1	PC2	Overlap (%)	Unique North Island (%)	Unique South Island (%)	Schoener's D	Similarity test (p)
Kapokapowai (North and South)	34.9	32.6	93	6.9	13.2	24.7	0.01

Environmental Variable	PC1	PC2	PC3
bio 3	-0.51	-0.27	-0.38
bio 4	-0.51	-0.17	-0.48
bio 5	0.11	0.47	-0.54
bio 15	0.24	0.49	-0.37
bio 18	0.38	-0.56	-0.20
bio 19	0.51	-0.35	-0.40

428 **Table 3:** PCA loadings, importance of variables, percent overlap, percent of unique niche overlap from
429 both islands, environmental similarity (Schoener's D), and results of similarity tests from our EcoSpat
430 analyses.

448 Figure 3: Niche Models of Kapokapowai Populations

Figure 3: Maxent predictions for Kapokapowai projected to the modern day using our 'Anthropocene
variables', mid-Holocene (Northgrippian: 4.2 – 8.326ka) generated from the PaleoClim dataset (Brown et
al. 2018), and Last Glacial Maximum (LGM)(21ka) and Last Interglacial (LIG)(130ka) generated from
the CHELSA database (Karger et al. 2017). Predictions were derived from the optimal model using the
criterion of mean valuation AUC (AUC$_{val}$) values being the highest, and 10% omission rate being the

455 lowest. Predictions were transformed using the 'cloglog' function in Maxent v3.4.4 (Phillips et al., 2017),
456 in which raw values are converted to a range of 0 - 1 to approximate a probability of occurrence. Brighter
457 colors (yellow, green, blue) indicate areas of higher suitability (higher probability of occurrence), while
458 darker colors (violet) indicate areas of lower suitability (lower probability of occurrence).
459
460 Figure 4: Niche Overlap of Kapokapowai North and Kapokapowai South
461

462
Figure 4: Ordination plot (PCA), niche overlap, correlation circle, and niche similarity tests showing
niche differences between Kapokapowai North and South from Te Ika-a-Māui (North) and Te
Waipounamu (South). Ordination plot represents principal component points of occurrence and
background environmental values using type 2 scaling (Mahalanobis distance). PCA points for Te
Waipounamu are in blue, while Te Ika-a-Māui is red. Solid contour lines in the niche overlap illustrate
full range (100%) of climate space ('fundamental niche') while dashed lines indicate 50% confidence
intervals. Contour lines of Te Waipounamu are in orange, and Te Ika-a-Māui is blue. Shading shows the

density of species occurrences per grid cell of kernel density analysis ('realized niche'), and violet pixels
show niche stability (climate conditions occupied in both time periods). Orange shading indicates climate
conditions only occupied by Te Waipounamu, while blue indicates climate conditions only occupied by
Te Ika-a-Māui. Correlation circle indicates climactic variable loadings on the PCA space. Length and
direction of arrows indicates influence and distribution of variables within PCA space. Bioclimatic
variables which follow Worldclim v2 (Fick and Hijmans 2017). Histograms represent observed (red bar)
and randomly simulated overlaps of niche similarity. The observed overlap is higher than 95% ($p < 0.05$)
for both analyses supporting the niche similarity across both islands.
478
Discussion
Biogeography
The genetic differentiation between KM and KS is much higher than the differentiation between KN and
KS (Table 2). Although Te Moana o Raukawa (Cook's Strait) is currently a barrier to the dispersal of
Kapokapowai, our species distribution models suggest that this has not always been the case (Fig. 4,
supplementary Fig. 2). Perhaps Te Moana o Raukawa has not been a barrier to the dispersal of
Kapokapowai for a sufficient period of time for a strong genetic structure to evolve (Fig. 3). On the other
hand, the peaks of Kā Tiritiri-o-te-Moana have not contained suitable habitat for KN and KS in any of
the tested time periods, and have likely served as a longer barrier to the dispersal of these populations,
shaping the genetic structure currently observed in Kapokapowai. The physical boundary of the
mountains is further compounded by the fact that the alpine habitat has shortened the flight season of KM,
compared to KN and KS (L. S. Wolfe, 1952). Despite this barrier, both the PCA (Fig. 1A) and admixture
(Fig. 1B) analyses suggest there is some level of interbreeding and movement between populations, and
neither barrier seems to be entirely restricting gene flow, as evidenced by the Fst values $<.3$, and the F4
ratios (Table 2).
494
Patterns of genetic diversity in Kapokapowai
Two main hypotheses have been posited to explain how alpine species evolved in Aotearoa: either the
species arrived by passively being carried with tectonic uplift, (Heads, 2017)), or they actively colonized
the mountains after their formation (Buckley et al., 2024). Our data offer support for the former in
Kapokapowai. Both PCA (fig. 1A) and admixture (fig. 1B) analyses showed KM to have much higher
genetic diversity than either KN or KS. This suggests that KM is closest to the ancestral state of all three
populations, and has thus retained the highest amount of genetic diversity, while KN and KS lost diversity
due to the founder effect, similar to many other systems (Anopheles gambiae 1000 Genomes Consortium
et al., 2017; Auton et al., 2015; B. Li et al., 2024). A historical demographic model, generated from the
reference genome of one individual of KS from Te Waipounamu using the Pairwise Sequential
Coalescent (Liu & Hansen, 2017) shows a severe bottleneck in effective population size *after* alpine uplift
(E. R. Tolman et al., 2023). Taken together with the population genomics presented here, a plausible
explanation is that all sampled Kapokapowai are descendents of a population that colonized alpine
habitats, while KN and KS descend from populations that later moved into lowland habitats. Such a
model would reconcile the bottleneck of ancestral Kapokapowai observed in the PSMC, and patterns of
genetic diversity as demonstrated by the PCA (Fig. 1A) and admixture (Fig. 1B) analyses, which show
that KM maintains the highest genetic diversity, which would have been lost in KN and KS as the
ancestral populations dispersed to lowlands. This hypothesis is not without precedent, as plant lineages
have also been hypothesized to have colonized mountains, and then recolonized the lowlands of Aoteora

(Thomas et al., 2023). A demographic model for KM from a reference genome, and revisiting the
specimens sampled here with genome-wide resequencing data could be used to generate a demographic
model for all populations of Kapokapowai, and a historical demographic model of dispersal and gene
flow between populations using the site-frequency-spectrum (Noskova et al., 2023), both of which could
be used to robustly test this hypothesis.
519
Species delimitations
The genetic structure of the three putative Kapokapowai species demonstrate that incipient speciation
(Fig. 1A,B) is underway in this genus, thus current species delimitations are difficult to make. Given the
genetic differentiation between KM and KN and KS(F_{ST} >.23), our identification of a diagnostic
characteristic of the curvature on the horns of KM (Fig. 2), and the strong posterior support (>99%) of
the currently described species (*U. carovei* comprised of KN and KS and *U. chiltoni* comprised of KM) in
our species delimitation analysis, we suggest that current species delimitations remain unchanged.
Although KN and KS do show some genetic differentiation, they do not show evidence of niche
divergence, and we do not believe that they merit a new species description at this point in time.
529
Conclusion
To look at species diversity and the potential for speciation in a group such as the petaltail dragonflies
offers a unique perspective in evolutionary biology. Analyses suggest that this family is extremely old,
even for an order as ancient as Odonata, with some extant species persisting as independent evolutionary
lineages since the Cretaceous era (Kohli et al., 2021; Suvorov et al., 2022; E. R. Tolman et al., 2024).
With their reliance on fen habitats, Petaluridae are also somewhat limited in their range, compared to
more generalist dragonflies that use ponds or rivers to complete their life cycle. While Kapokapowai have
potentially been separated for significant periods of time, either across islands or in different habitats
within an island, it may be that their adaptations to these specialized habitats has limited the genetic
variability that would lead to speciation. This may help to explain why this group, despite its age,
continues to be the smallest family within Anisoptera.
541

References
Andress, R. (1998). *Description of the larva of Petalura ingentissima Tillyard, 1907 (Anisoptera:*
Petaluridae). http://archive.org/details/odonatologica-27-353-359
Anopheles gambiae 1000 Genomes Consortium, Data analysis group, Partner working group, Sample
collections—Angola:, Burkina Faso:, Cameroon:, Gabon:, Guinea:, Guinea-Bissau:, Kenya:,
Uganda:, Crosses:, Sequencing and data production, Web application development, & Project
coordination. (2017). Genetic diversity of the African malaria vector Anopheles gambiae. *Nature*,
552(7683), 96–100. https://doi.org/10.1038/nature24995
Auton, A., Abecasis, G. R., Altshuler, D. M., Durbin, R. M., Abecasis, G. R., Bentley, D. R., Chakravarti,
A., Clark, A. G., Donnelly, P., Eichler, E. E., Flicek, P., Gabriel, S. B., Gibbs, R. A., Green, E.
D., Hurles, M. E., Knoppers, B. M., Korbel, J. O., Lander, E. S., Lee, C., … National Eye
Institute, N. (2015). A global reference for human genetic variation. *Nature*, *526*(7571), 68–74.
https://doi.org/10.1038/nature15393
Barreto, E., Rangel, T. F., Pellissier, L., & Graham, C. H. (2021). Area, isolation and climate explain the
diversity of mammals on islands worldwide. *Proceedings of the Royal Society B: Biological*
Sciences, *288*(1965), 20211879. https://doi.org/10.1098/rspb.2021.1879

Booth, T. H. (2022). Checking bioclimatic variables that combine temperature and precipitation data before their use in species distribution models. *Austral Ecology*, *47*(7), 1506–1514. https://doi.org/10.1111/aec.13234

Bouckaert, R. R. (2022). An Efficient Coalescent Epoch Model for Bayesian Phylogenetic Inference. *Systematic Biology*, *71*(6), 1549–1560. https://doi.org/10.1093/sysbio/syac015

Brown, J. L., Hill, D. J., Dolan, A. M., Carnaval, A. C., & Haywood, A. M. (2018). PaleoClim, high spatial resolution paleoclimate surfaces for global land areas. *Scientific Data*, *5*(1), Article 1. https://doi.org/10.1038/sdata.2018.254

Buckley, T. R., Hoare, R. J. B., & Leschen, R. A. B. (2024). Key questions on the evolution and biogeography of New Zealand alpine insects. *Journal of the Royal Society of New Zealand*, *54*(1), 30–54. https://doi.org/10.1080/03036758.2022.2130367

Conway, M., & Olsen, B. J. (2019). Contrasting drivers of diversification rates on islands and continents across three passerine families. *Proceedings. Biological Sciences*, *286*(1915), 20191757. https://doi.org/10.1098/rspb.2019.1757

Danecek, P., Bonfield, J. K., Liddle, J., Marshall, J., Ohan, V., Pollard, M. O., Whitwham, A., Keane, T., McCarthy, S. A., Davies, R. M., & Li, H. (2021). Twelve years of SAMtools and BCFtools. *GigaScience*, *10*(2), giab008. https://doi.org/10.1093/gigascience/giab008

Di Cola, V., Broennimann, O., Petitpierre, B., Breiner, F. T., D'Amen, M., Randin, C., Engler, R., Pottier, J., Pio, D., Dubuis, A., Pellissier, L., Mateo, R. G., Hordijk, W., Salamin, N., & Guisan, A. (2017). ecospat: An R package to support spatial analyses and modeling of species niches and distributions. *Ecography*, *40*(6), 774–787. https://doi.org/10.1111/ecog.02671

Douglas, J., & Bouckaert, R. (2022). Quantitatively defining species boundaries with more efficiency and more biological realism. *Communications Biology*, *5*(1), 1–10. https://doi.org/10.1038/s42003-022-03723-z

Dray, S., & Dufour, A.-B. (2007). The ade4 Package: Implementing the Duality Diagram for Ecologists. *Journal of Statistical Software*, *22*, 1–20. https://doi.org/10.18637/jss.v022.i04

Durrant, R. (2024). *Invertebrate Justice: Extending The Boundaries of Non-Speciesist Green Criminology*. Springer Nature.

Fick, S. E., & Hijmans, R. J. (2017). WorldClim 2: New 1-km spatial resolution climate surfaces for global land areas. *International Journal of Climatology*, *37*(12), 4302–4315. https://doi.org/10.1002/joc.5086

Fong, H. M. (2024). *A revision of the genus Mecodema Blanchard 1853 (Coleoptera: Carabidae) species groups ducale and laterale from the South Island, Aotearoa New Zealand* [Thesis, ResearchSpace@Auckland]. https://researchspace.auckland.ac.nz/handle/2292/70309

Gleditsch, J., Behm, J., Ellers, J., Jesse, W., & Helmus, M. (2022). *Contemporizing island biogeography theory with anthropogenic drivers of species richness*. https://doi.org/10.22541/au.163955172.21711050/v3

Goodman, A. M., Beatty, C. D., Büsse, S., Ubukata, H., Miyazaki, T., Blair, M. E., & Ware, J. L. (2024). Paleoecological niche modeling of Epiophlebia (Epiophlebioptera: Epiophlebiidae) reveals continuous distribution during the Last Glacial Maximum. *International Journal of Odonatology*, *27*, 60–76. https://doi.org/10.48156/1388.2024.1917262

Grant, E. A. (2014). *Bronze as a non-customary intervention in the interpretation of insects from the natural world of Māori: An exhibition report presented in partial fulfilment of the requirements for the degree of Doctor of Philosophy in Fine Arts at Massey University, Palmerston North, Aotearoa, New Zealand* [Massey University]. http://hdl.handle.net/10179/6512

Harris, C. R., Millman, K. J., Van Der Walt, S. J., Gommers, R., Virtanen, P., Cournapeau, D., Wieser, E., Taylor, J., Berg, S., Smith, N. J., Kern, R., Picus, M., Hoyer, S., Van Kerkwijk, M. H., Brett, M., Haldane, A., Del Río, J. F., Wiebe, M., Peterson, P., … Oliphant, T. E. (2020). Array programming with NumPy. *Nature*, *585*(7825), 357–362. https://doi.org/10.1038/s41586-020-2649-2

Heads, M. J. (2017). *Biogeography and evolution in New Zealand*. CRC press, Taylor & Francis group.

Jenkins, T. L. (2024). mapmixture: An R package and web app for spatial visualisation of admixture and population structure. *Molecular Ecology Resources*, e13943. https://doi.org/10.1111/1755-0998.13943

Karger, D. N., Conrad, O., Böhner, J., Kawohl, T., Kreft, H., Soria-Auza, R. W., Zimmermann, N. E., Linder, H. P., & Kessler, M. (2017). Climatologies at high resolution for the earth's land surface areas. *Scientific Data*, *4*(1), Article 1. https://doi.org/10.1038/sdata.2017.122

Karger, D. N., Nobis, M. P., Normand, S., Graham, C. H., & Zimmermann, N. E. (2023). CHELSA-TraCE21k – high-resolution (1 km) downscaled transient temperature and precipitation data since the Last Glacial Maximum. *Climate of the Past*, *19*(2), 439–456. https://doi.org/10.5194/cp-19-439-2023

Kass, J. M., Muscarella, R., Galante, P. J., Bohl, C. L., Pinilla-Buitrago, G. E., Boria, R. A., Soley-Guardia, M., & Anderson, R. P. (2021). ENMeval 2.0: Redesigned for customizable and reproducible modeling of species' niches and distributions. *Methods in Ecology and Evolution*, *12*(9), 1602–1608. https://doi.org/10.1111/2041-210X.13628

Kass, J. M., Vilela, B., Aiello-Lammens, M. E., Muscarella, R., Merow, C., & Anderson, R. P. (2018). Wallace: A flexible platform for reproducible modeling of species niches and distributions built for community expansion. *Methods in Ecology and Evolution*, *9*(4), 1151–1156. https://doi.org/10.1111/2041-210X.12945

Kennedy, C. H. (1922). The morphology of the penis in the genus Libellula (Odonata). *Entomology Newsletter*, *33*, 33–40.

Kohli, M., Letsch, H., Greve, C., Béthoux, O., Deregnaucourt, I., Liu, S., Zhou, X., Donath, A., Mayer, C., Podsiadlowski, L., Gunkel, S., Machida, R., Niehuis, O., Rust, J., Wappler, T., Yu, X., Misof, B., & Ware, J. (2021). Evolutionary history and divergence times of Odonata (dragonflies and damselflies) revealed through transcriptomics. *iScience*, *24*(11), 103324. https://doi.org/10.1016/j.isci.2021.103324

Kuznetsova, V. G., & Golub, N. V. (2020). A checklist of chromosome numbers and a review of karyotype variation in Odonata of the world. *Comparative Cytogenetics*, *14*(4), 501–540. https://doi.org/10.3897/CompCytogen.v14i4.57062

L. S. Wolfe. (1952). A study of the genus Uropetala Selys (Order Odonata) from New Zealand. *Transactions of the Royal Society of New Zealand*, *80*, 245–275.

Leschen, R., Collier, K., Death, R., Harding, J., & Smith, B. (2023). Systematics expertise and taxonomic status of New Zealand's freshwater insects. *New Zealand Science Review*, *73*(3–4), 92–95. https://doi.org/10.26686/nzsr.v73i3-4.8534

Lewis, K. B., Carter, L., & Davey, F. J. (1994). The opening of Cook Strait: Interglacial tidal scour and aligning basins at a subduction to transform plate edge. *Marine Geology*, *116*(3), 293–312. https://doi.org/10.1016/0025-3227(94)90047-7

Li, B., Duan, Y., Du, Z., Wang, X., Liu, S., Feng, Z., Tian, L., Song, F., Yang, H., Cai, W., Lin, Z., & Li, H. (2024). Natural selection and genetic diversity maintenance in a parasitic wasp during continuous biological control application. *Nature Communications*, *15*(1), 1379. https://doi.org/10.1038/s41467-024-45631-2

Li, H., & Durbin, R. (2009). Fast and accurate short read alignment with Burrows–Wheeler transform. *Bioinformatics*, *25*(14), 1754–1760. https://doi.org/10.1093/bioinformatics/btp324

Liu, S., & Hansen, M. M. (2017). PSMC (pairwise sequentially Markovian coalescent) analysis of RAD (restriction site associated DNA) sequencing data. *Molecular Ecology Resources*, *17*(4), 631–641. https://doi.org/10.1111/1755-0998.12606

Lobo, J. M., Jiménez-Valverde, A., & Real, R. (2008). AUC: A misleading measure of the performance of predictive distribution models. *Global Ecology and Biogeography*, *17*(2), 145–151. https://doi.org/10.1111/j.1466-8238.2007.00358.x

Malinsky, M., Matschiner, M., & Svardal, H. (2021). Dsuite—Fast D-statistics and related admixture evidence from VCF files. *Molecular Ecology Resources*, *21*(2), 584–595. https://doi.org/10.1111/1755-0998.13265

May, M. L. (1997). Reconsideration of the status of the genera Phyllomacromia and Macromia
(Anisoptera: Corduliidae). *Odonatologica*, *26*(4), 405–414.

McKinney, W. (2010). *Data Structures for Statistical Computing in Python*. 56–61.
https://doi.org/10.25080/Majora-92bf1922-00a

Naimi, B. (2023). *usdm: Uncertainty Analysis for Species Distribution Models* (Version 2.1-6) [Computer
software]. https://cran.r-project.org/web/packages/usdm/index.html

Noskova, E., Abramov, N., Iliutkin, S., Sidorin, A., Dobrynin, P., & Ulyantsev, V. (2023). *GADMA2:
More efficient and flexible demographic inference from genetic data* (p. 2022.06.14.496083).
bioRxiv. https://doi.org/10.1101/2022.06.14.496083

Oksanen, J., Simpson, G. L., Blanchet, F. G., Kindt, R., Legendre, P., Minchin, P. R., O'Hara, R. B.,
Solymos, P., Stevens, M. H. H., Szoecs, E., Wagner, H., Barbour, M., Bedward, M., Bolker, B.,
Borcard, D., Carvalho, G., Chirico, M., Caceres, M. D., Durand, S., … Weedon, J. (2024). *vegan:
Community Ecology Package* (Version 2.6-6.1) [Computer software]. https://cran.r-
project.org/web/packages/vegan/index.html

O'Neill, S. B., Chapple, D. G., Daugherty, C. H., & Ritchie, P. A. (2008). Phylogeography of two New
Zealand lizards: McCann's skink (*Oligosoma maccanni*) and the brown skink (*O. zelandicum*).
Molecular Phylogenetics and Evolution, *48*(3), 1168–1177.
https://doi.org/10.1016/j.ympev.2008.05.008

Peterson, A. T., & Lieberman, B. S. (2012). Species' Geographic Distributions Through Time: Playing
Catch-up with Changing Climates. *Evolution: Education and Outreach*, *5*(4), 569–581.
https://doi.org/10.1007/s12052-012-0385-2

Phillips, S. J., Anderson, R. P., Dudík, M., Schapire, R. E., & Blair, M. E. (2017). Opening the black box:
An open-source release of Maxent. *Ecography*, *40*(7), 887–893.
https://doi.org/10.1111/ecog.03049

Purcell, S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M. A. R., Bender, D., Maller, J., Sklar, P.,
de Bakker, P. I. W., Daly, M. J., & Sham, P. C. (2007). PLINK: A Tool Set for Whole-Genome
Association and Population-Based Linkage Analyses. *American Journal of Human Genetics*,
81(3), 559–575.

Radosavljevic, A., & Anderson, R. P. (2014). Making better Maxent models of species distributions:
Complexity, overfitting and evaluation. *Journal of Biogeography*, *41*(4), 629–643.
https://doi.org/10.1111/jbi.12227

Rambaut, A., Drummond, A. J., Xie, D., Baele, G., & Suchard, M. A. (2018). Posterior Summarization in
Bayesian Phylogenetics Using Tracer 1.7. *Systematic Biology*, *67*(5), 901–904.
https://doi.org/10.1093/sysbio/syy032

Schoener, T. W. (1968). The Anolis Lizards of Bimini: Resource Partitioning in a Complex Fauna.
Ecology, *49*(4), 704–726. https://doi.org/10.2307/1935534

Shepherd, L., Simon, C., Langton-Myers, S., & Morgan-Richards, M. (2024). Insights into Aotearoa New
Zealand's biogeographic history provided by the study of natural hybrid zones. *Journal of the
Royal Society of New Zealand*, *54*(1), 55–74. https://doi.org/10.1080/03036758.2022.2061020

Skotte, L., Korneliussen, T. S., & Albrechtsen, A. (2013). Estimating individual admixture proportions
from next generation sequencing data. *Genetics*, *195*(3), 693–702.
https://doi.org/10.1534/genetics.113.154138

Stevens, M. I., & Hogg, I. D. (2004). Population genetic structure of New Zealand's endemic corophiid
amphipods: Evidence for allopatric speciation. *Biological Journal of the Linnean Society*, *81*(1),
119–133. https://doi.org/10.1111/j.1095-8312.2004.00270.x

Stoltz, M., Baeumer, B., Bouckaert, R., Fox, C., Hiscott, G., & Bryant, D. (2021). Bayesian Inference of
Species Trees using Diffusion Models. *Systematic Biology*, *70*(1), 145–161.
https://doi.org/10.1093/sysbio/syaa051

Suvorov, A., Scornavacca, C., Fujimoto, M. S., Bodily, P., Clement, M., Crandall, K. A., Whiting, M. F.,
Schrider, D. R., & Bybee, S. M. (2022). Deep Ancestral Introgression Shapes Evolutionary

History of Dragonflies and Damselflies. *Systematic Biology*, *71*(3), 526–546. https://doi.org/10.1093/sysbio/syab063

Taylor, R. (1855). *Te Ika a Maui, or, New Zealand and its inhabitants: With the geology, natural history, productions, and climate of the country, &c* (Vol. 1855, pp. 1–554). https://doi.org/10.5962/bhl.title.161768

Taylor, R. (with Smithsonian Libraries). (1870). *Te Ika a Māui, or, New Zealand and its inhabitants: Illustrating the origin, manners, customs, mythology, religion, rites, songs, proverbs, fables and language of the Māori and Polynesian races in general ; together with the geology, natural history, productions, and climate of the country*. London : William Macintosh ; Wanganui, N.Z. : H. Ireson Jones. http://archive.org/details/teikamauiornewze00tayl

Thomas, A., Meudt, H. M., Larcombe, M. J., Igea, J., Lee, W. G., Antonelli, A., & Tanentzap, A. J. (2023). Multiple origins of mountain biodiversity in New Zealand's largest plant radiation. *Journal of Biogeography*, *50*(5), 947–960. https://doi.org/10.1111/jbi.14589

Tolman, E. (2024). Systematics and Comparative Genomics of the Petaluridae (Odonata: Anisoptera). *Dissertations, Theses, and Capstone Projects*. https://academicworks.cuny.edu/gc_etds/5758

Tolman, E. R., Beatty, C. D., Frandsen, P. B., Bush, J., Bruchim, O. R., Driever, E. S., Harding, K. M., Jordan, D., Kohli, M. K., Park, J., Park, S., Reyes, K., Rosari, M., Ryu, J. L., Wade, V., & Ware, J. L. (2023). *Newly Sequenced Genomes Reveal Patterns of Gene Family Expansion in select Dragonflies (Odonata: Anisoptera)* (p. 2023.12.11.569651). bioRxiv. https://doi.org/10.1101/2023.12.11.569651

Tolman, E. R., Beatty, C. D., Kohli, M. K., Abbott, J., Bybee, S. M., Frandsen, P. B., Stephen Gosnell, J., Guralnick, R., Kalkman, V. J., Newton, L. G., Suvorov, A., & Ware, J. L. (2024). A molecular phylogeny of the Petaluridae (Odonata: Anisoptera): A 160-Million-Year-Old story of drift and extinction. *Molecular Phylogenetics and Evolution*, *200*, 108185. https://doi.org/10.1016/j.ympev.2024.108185

Trewick, S. A., Pilkington, S., Shepherd, L. D., Gibb, G. C., & Morgan-Richards, M. (2017). Closing the gap: Avian lineage splits at a young, narrow seaway imply a protracted history of mixed population response. *Molecular Ecology*, *26*(20), 5752–5772. https://doi.org/10.1111/mec.14323

Veloz, S. D. (2009). Spatially autocorrelated sampling falsely inflates measures of accuracy for presence-only niche models. *Journal of Biogeography*, *36*(12), 2290–2299. https://doi.org/10.1111/j.1365-2699.2009.02174.x

W. J. Winstanley. (1981). AN EMERGENCE STUDY ON UROPETALA CAROVEI CAROVEI (ODONATA: PETALURIDAE) NEAR WELLINGTON, NEW ZEALAND, WITH NOTES ON THE BEHAVIOUR OF THE SUBSPECIES. *Tuatara*, *25*(1).

Wallis, G. P., & Buckley, T. R. (2024). Evolutionary biogeography of Aotearoa. *Journal of the Royal Society of New Zealand*, *54*(1), 1–7. https://doi.org/10.1080/03036758.2023.2260539

Ware, J. L. (2008). *MOLECULAR AND MORPHOLOGICAL SYSTEMATICS OF LIBELLULOIDEA (ODONATA: ANISOPTERA) AND DICTYOPTERA* [PhD Thesis, Rutgers University]. https://rucore.libraries.rutgers.edu/rutgers-lib/24985/PDF/1

Ware, J. L., Beatty, C. D., Herrera, M. S., Valley, S., Johnson, J., Kerst, C., May, M. L., & Theischinger, G. (2014). The petaltail dragonflies (Odonata: Petaluridae): Mesozoic habitat specialists that survive to the modern day. *Journal of Biogeography*, *41*(7), 1291–1300. https://doi.org/10.1111/jbi.12273

Warren, D. L., & Seifert, S. N. (2011). Ecological niche modeling in Maxent: The importance of model complexity and the performance of model selection criteria. *Ecological Applications*, *21*(2), 335–342. https://doi.org/10.1890/10-1171.1

Waters, J. M., & Craw, D. (2006). Goodbye Gondwana? New Zealand Biogeography, Geology, and the Problem of Circularity. *Systematic Biology*, *55*(2), 351–356. https://doi.org/10.1080/10635150600681659

759   Wilson, E. O. (2009). Island Biogeography in the 1960s: Theory and Experiment. In *The Theory of Island
760   *Biogeography Revisited* (pp. 1–12). Princeton University Press.
761   https://doi.org/10.1515/9781400831920.1
762   Winstanley, W. J., & Rowe, R. J. (1980). The larval habitat of Uropetala carovei carovei (Odonata:
763   Petaluridae) in the North Island of New Zealand, and the geographical limits of the subspecies.
764   *New Zealand Journal of Zoology*, *7*(1), 127–134.
765   https://doi.org/10.1080/03014223.1980.10423769
766   Wyckhuys, K. A. G., Sanchez Garcia, F. J., Santos, A. M. C., Canal, N. A., Furlong, M. J., Melo, M. C.,
767   Gc, Y. D., & Pozsgai, G. (2022). Island and Mountain Ecosystems as Testbeds for Biological
768   Control in the Anthropocene. *Frontiers in Ecology and Evolution*, *10*.
769   https://doi.org/10.3389/fevo.2022.912628
770