10 **The predictive structure of navigation determines mouse CA1 representational structure**
11 **in a multicompartment environment**
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Predictive theories of cognitive mapping propose that these representations encode the predictive
relationships among contents as experienced by the navigator. One hallmark of these theories is
that representational structure in complex environments can be predicted from the behavioral
history of the navigator. Here, we image neural activity in hippocampal CA1 as initially naïve mice
repeatedly navigate a multicompartment environment to test whether representational structure
in this subregion is consistent with these predictions. We find that different mice instantiate
different patterns of remapping across identically shaped compartments. Within mouse,
compartments with more similar predictive navigational histories on a particular spatiotemporal
scale are represented more similarly, accounting for these individual differences. Manipulating
navigational options induces reorganization of the CA1 structure which specifically resembles the
new predictive navigational structure on this scale. Through computational modeling we show that
a combination of predictive encoding and geometrically structured inputs can uniquely account
for this pattern of results. Together, these results demonstrate that the structure of CA1
representations in complex environments can be predicted from the behavioral history of the
navigator, consistent with predictive theories of cognitive mapping.

In order to survive and thrive, organisms from mice to humans rely on mnemonic representations
of the external world and their relationships to it[1]. Referred to as *cognitive maps*, these
representations are instantiated by the coordinated activity of spatially tuned neural populations
in the hippocampus and neighboring cortices[2,3]. Classically, cognitive maps are thought to
represent geometric relationships – distances and/or angles – among their contents, motivated
by the plethora of functional cell types tuned to geometric features throughout the hippocampal
formation and neighboring cortices[2,4–18]. However, recent theories posit instead that cognitive
maps represent the predictive relationships among their contents as experienced by the
navigator[19–22], with geometrically structured inputs providing a scaffold upon which predictive
structure can be learned[3,23–25].

Because the geometric layout of an environment often determines the predictive
relationships that the navigator experiences, and modifying navigation often entails modifying the
layout of the world, the outcomes of traditional and predictive models can be difficult to
disentangle. Nevertheless, there are fundamental differences between these theories. One
hallmark of predictive theories is that the way in which a navigator explores a space is a critical
determinant of the structure of their cognitive map[19]. If navigators explore the same space
differently, then their cognitive maps should be structured differently in a way which can be
predicted from their navigational patterns. These differences should manifest across levels of
explanation from neural instantiations through behavior, from the activity of single cells through
the representational geometry of neural populations.

Motivated in part by these predictions, a growing body of work is contrasting behavioral
and geometric determinants of neural activity in the hippocampus and functionally related
cortices[6,13,26–34]. In many cases this work has demonstrated behavioral modulation of activity at
the level of single cells and moment-to-moment population trajectories which is roughly consistent
with predictive theories[13,29,30,33–35] and which challenges traditional theories. In addition to these
predictions, predictive theories also make concrete claims about how behavior should determine
the representational similarity among locations in complex environments consisting of multiple
subspaces[19]. Prior work has demonstrated that representational similarity in the hippocampus –
or its converse, *remapping* – depends on the features shared between spaces[36–40], past
experience with those spaces[41–45], and the broader reference frames in which those spaces are
situated[43]. Individual differences in the degree of remapping within the same paradigm have been
observed[46], suggesting that remapping is not determined by world features and prior experience
alone and can vary between navigators. Each of these characteristics can be understood within
a predictive framework. However, more specific tests of whether patterns of hippocampal
remapping in complex spaces conform to a predictive framework have failed[19,47], calling into
question whether the structure of hippocampal representations in such spaces can be understood
through this lens.

Notably, this previous test included multiple complexities – such as extensive prior
experience in the environment and short-term changes to navigational options and the reward
landscape – which make deriving concrete predictions from a predictive framework more
challenging. Here, we test whether patterns of hippocampal remapping in complex spaces
conform to a predictive framework in a simpler paradigm which avoids some of these
complexities. Specifically, we characterize population activity in hippocampal CA1 as initially
naïve mice repeatedly explore four identically shaped connected compartments in the absence
of reward. To do so with high fidelity and across long timescales, we rely on chronic miniscope
imaging. We test three hypotheses. If CA1 representational structure conforms to a predictive
framework, then (i) mice that navigate differently should exhibit different patterns of remapping
across compartments, (ii) within mouse, compartments with more similar predictive navigational

histories should be represented more similarly, and (iii) changing the navigational trajectory of a
mouse should change its representational structure to better resemble the new predictive
structure. Evidence against any of these hypotheses would support an alternative theory.
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In all cases our data support the predictive claims. That is, we find that different mice
exhibit different patterns of remapping across compartments. These patterns coincide with the
similarity of predictive navigational histories on a seconds-long timescale, modestly overweighting
early experience. Manipulating the navigational trajectory of the mouse induces a reorganization
of the CA1 representational structure which specifically resembles the new predictive structure
on this timescale, with a reversion to the previous pattern when relaxed. Leveraging
computational modeling, we show that key results can be accounted for by a combination of
predictive encoding operating on geometrically structured inputs; neither predictive encoding nor
geometrically structured inputs were on their own sufficient to reproduce our pattern of results.
Together, these results demonstrate that the structure of hippocampal representations in complex
environments can be predicted from the behavioral history of the navigator, consistent with
predictive theories of cognitive mapping.
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135 **Results**
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137 **Characterizing CA1 representational structure in a multicompartment environment**
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We recorded daily from hippocampal CA1 via miniscope calcium imaging (Fig. 1a) as initially
naïve mice (n = 11; 6 male) navigated a radial multicompartment environment. This environment
consisted of four 20 cm by 35 cm rectangular compartments connected by a central chamber
(Fig. 1b). Each session lasted either 30 or 40 min depending on imaging signal strength to
maximize the amount of data while minimizing the impact of photobleaching. In the first phase of
the experiment, initially naïve mice navigated this environment freely for between five and fifteen
sessions (n = 121 sessions total). We refer to these sessions as *free$_1$*. The environment was not
baited with rewards during any phase of the experiment.
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For each session, imaging data were first motion corrected[48]. Next, cells were segmented,
and their calcium traces were extracted via constrained nonnegative matrix factorization (CNMFE;
Fig. 1a)[49,50]. For each trace, the likelihood of spiking events giving rise to that trace were inferred
through a second-order autoregressive deconvolution algorithm[51] (Fig. 1a). This nonnegative
vector was then binarized such that any frame with a nonzero likelihood was treated as active
(i.e. 1), otherwise it was treated as not active (i.e. 0). All further analysis was conducted on this
binary activity vector.
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To characterize the extent to which spatial location modulated the activity of each cell, we
computed a rate map for each cell summarizing the mean activity of that cell as a function of
location within the environment (Fig. 1c). Next, we computed the split-half stability and spatial
information content of this rate map. To assess the significance of these measures, we compared
these values to surrogate distributions computed by circularly shifting the activity vector relative
to the position vector 1000 times. Cells with split-half stability and spatial information content
exceeding the 95th percentiles of both distributions were considered spatially tuned and included
in further analysis, though relaxing these criteria did not qualitatively change our results.
Generally, the majority of cells met these criteria (Fig. 1d).
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Prior work in multicompartment environments has demonstrated that individual CA1 cells
tend to be active in multiple compartments[52,53], making comparisons between compartments
meaningful. We confirmed this was the case in our data. To do so, for each cell we computed its

mean activity rate in each compartment and sorted these from the most active to least active
compartments. Across sessions, we found that mean activity rates remained high in all
compartments, with on average at least ~1 active frame per second even in dispreferred
compartments (Fig. 1e). The difference in activity rate between preferred and dispreferred
compartments was approximately 3-fold. Interestingly, we also observed a change in mean
activity rates with experience in dispreferred (but not preferred) compartments (Fig. 1f), consistent
with the development of sparsity and/or sharpening of the spatial code across experience[54].
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Figure 1. Characterizing mouse CA1 in a multicompartment environment via miniscope
imaging. *(a) Schematic of miniscope imaging and data processing pipeline. (b) Schematic of the*
multicompartment environment. (c) Example rate maps from simultaneously-recorded cells for
one session from two mice. Rate maps normalized from zero (blue) to peak activity rate (yellow).
Peak activity rate in proportion of active frames noted for each map. (d) Proportion of cells with
split-half stability and spatial information content exceeding the 95th percentile of their shuffled
controls across sessions for each mouse. Median number of cells per session for each mouse
noted on the right. (e) Mean activity rate as a function of most active to least active compartments.
(f) Change in mean event rate versus the first session, as a function of most active to least active
compartments. Soft shaded lines are individual mice. Bold line is the mean across mice. Shaded
box denotes sessions for which p < 0.05. Significance markers denote the outcome of a t-test
versus zero for a given compartment and session. *p<0.05

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191 **Different mice instantiate distinct and persistent patterns of remapping across identical**
192 **compartments**

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Predictive theories claim that, all else being equal, CA1 representational structure should be
determined by the predictive structure of one's navigational trajectory. If so, then we would expect
mice in our paradigm to instantiate different patterns of remapping between compartments insofar

as they navigate with different trajectories. On the other hand, purely geometric theories might
predict that remapping is dictated by the geometric features of the environment, such as the
distance between compartments or the relative orientations of the compartments[52,53]. If so, then
we would expect mice in our paradigm to instantiate similar patterns of remapping, perhaps
corresponding to key geometric features.
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To address these possibilities, we characterized the degree of remapping between
compartments in our data using a population vector approach. To this end, for each cell we first
computed a separate rate map for each compartment (Fig. 2a). Next, for each session we
computed the population vector (PV) correlations between all pairwise comparisons of
compartment rate maps, aligned by their entryways and excluding unsampled pixels (Fig. 2a). PV
correlations summarize the degree to which two spaces are represented similarly at the level of
the neural population in a way that captures changes in both firing rates and spatial tuning.
Computing all pairwise PV correlations between compartments yields a 4 x 4 matrix which we
refer to as the *PV structure* (Fig. 2a). Finally, we correlated PV structures between sessions and
compared correlations between different sessions from the same mice to correlations between
sessions from different mice (Fig. 2b).
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Figure 2. Individual differences in patterns of remapping across compartments in a
multicompartment environment. (a) Schematic of PV structure analysis pipeline. Rate maps
scaled to peak activity rate across all compartments. This peak rate noted next to each map. (b)
PV structure correlations across all pairwise comparisons of sessions. White boxes bound
session comparisons from the same mice. (c) PV structure correlations for comparisons within
mouse versus between different mice, treating each pairwise session comparison independently
(rank-sum test: z(3143353) = 11.879, p = 1.513e-32). (d) PV structure correlations for
comparisons within mouse versus between mice, treating each mouse as the unit of analysis
(signed-rank test: p = 9.766e-04). Soft lines denote individual mice. (e) Distribution of
compartment-to-compartment PV correlation values for each mouse. (f) PV structures averaged
across sessions for each mouse. ***p<0.001
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228   Consistent with the predictive theory, comparisons of sessions from the same mouse
229 resulted in significantly higher correlations than comparisons between mice (Fig. 2c,d). Moreover,

despite the limited number of comparisons defining the PV structure (i.e. six pairwise comparisons between the four compartments), these differences were so reliable that a simple Euclidean distance based classifier could correctly predict mouse identity well above chance (accuracy = 45.45%, chance = 9.09%, p = ~0; binomial test). This reliable structure was present even though PV correlation values between compartments were generally low in magnitude (Fig. 2e), suggesting that even weak correlations in representational structure can be stable and informative[55]. Even averaging within mouse, PV structures varied widely between mice and qualitatively did not come to resemble a consistent geometric feature of the compartmental layout (i.e. distance, entryway angle, long axis, etc.; Fig. 2f). Similar results were observed when correlating mean activity rates as the measure of remapping (Fig. S1), indicating that our results are not dependent upon the rate map-based assumptions underlying PV correlations. Together, these results demonstrate that mice instantiate idiosyncratic patterns of remapping across compartments in this multicompartment paradigm.

Idiosyncratic CA1 representational structure matches the predictive structure of navigation on a particular spatiotemporal scale

Our previous results demonstrate reliable individual differences in CA1 representational structure in mice navigating our multicompartment environment. While these differences could arise from predictive cognitive mapping, individual differences are not on their own unique to this theory. For example, a geometric theory might predict individual differences if navigators are attending to or naturally emphasize different features of the world. Therefore, we next tested the more specific claim of predictive theories: that individual differences in CA1 representational structure arise due to idiosyncratic navigational differences. If correct, then compartments with more similar predictive navigational structures should be represented more similarly in CA1.

To test this possibility, we leveraged one formalization of predictive cognitive mapping, the successor representation[19]. At a high level, the successor representation defines the behavioral determinants of hippocampal representational structure as the temporally discounted probabilities of transitioning between locations within an environment. This is expressed as a successor matrix M such that

$$M = \sum_{t=0}^{\infty} \gamma^t T^t = (I - \gamma T^1)^{-1} \tag{1}$$

where T^n is the *n*-step transition matrix capturing the likelihood of transitioning from one location to another after *n* timesteps, γ is the temporal discount factor which determines how heavily transitions near in time versus far in time are discounted, and I is the identity matrix.

In our case, for a given session s we can compute the successor matrix $M_{s,k}$ separately for each compartment k. Correlating these matrices between all pairwise comparisons of compartments yields a 4 x 4 matrix which we can compare to the PV structure for that session. If the predictive claim is correct, then these matrices should be correlated. Importantly though, in our paradigm mice explored the environment for multiple sessions, and it is not obvious whether navigation during every session should have an equal impact on CA1 representational structure. While this is certainly a possibility, it is also possible that more recent behavior or early behavior has an exaggerated impact on determining CA1 representational structure. The longitudinal nature of our recording paradigm allows us to test these possibilities. To do so, for each session s and compartment k we computed a cumulative one-step transition matrix $C_{s,k}$ by including all navigational data up to and including that session such that

$$C_{s,k} = \sum_{i=1}^{s} T_{i,k} \, e^{(\tau i)} \tag{2}$$

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where $T_{i,k}$ is the one-step transition matrix for session i and compartment k and τ determines the
exponential weight with which each session contributes to the cumulative transition matrix. A
negative value of τ will overweight early session behavior. A positive value of τ will overweight
more recent behavior. A τ of zero will equally weight data from all sessions. Substituting our
weighted cumulative one-step transition matrix for T^1 in equation (1), we can compute the
cumulative successor matrix $M_{s,k}$ for each session s and compartment k such that
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$$M_{s,k} = (I - \gamma \sum_{i=1}^{s} T_{i,k} \, e^{(\tau i)})^{-1} \tag{3}$$

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Finally, we can correlate $M_{s,k}$ between all pairwise comparisons of compartments (excluding
unsampled pixels), yielding the 4 x 4 matrix which we refer to as the *cumulative successor*
structure (Fig. 3a). This matrix summarizes the similarity in predictive navigational structure
between compartments dependent upon two parameters, γ and τ. These parameters determine
the time horizon of the encoded predictive structure and the weighting of cumulative experience,
respectively.
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To determine whether cumulative successor structures match CA1 PV structures at any
parameterization, we conducted a grid search where γ and τ were varied and the mean successor
structure was correlated with the mean PV structure for each mouse. This analysis revealed that
mean successor structures were highly correlated with mean PV structures at a particular subset
of parameterizations, with a maximum at γ = 0.994 and τ = -0.25 (Fig. 3b,c; Fig. S2a). These
parameters correspond to a seconds-long timescale for encoding predictive structure, modestly
overweighting early experience (Fig. 3d). To assess the significance of the high correlation we
observed at this parameterization (mean r = 0.600), we considered several controls. First, we
compared this value to the distribution we might expect from generic mouse behavior by randomly
shuffling transition probabilities (i.e. T in Equation 3) between compartments, sessions, and mice
prior to computing cumulative successor structures. To account for the selection of the maximizing
parameterization from our grid search, we repeated our grid search for each shuffle and took the
maximum correlation for that shuffle. The true correlation exceeded this control (nonparametric p
~ 0.000, 1,000 shuffles; Fig. 3e). Next, we tested whether the specific order of experience
mattered by randomly shuffling navigational trajectories across sessions separately for each
mouse before computing cumulative successor structures and choosing the maximum correlation
across the grid search for each shuffle. Again, the true correlation exceeded this control
(nonparametric p = 0.011, 1,000 shuffles; Fig. 3f). Repeating these analyses but treating
individual sessions as the unit of analysis rather than individual mice yielded similar results (Fig.
S2). Substituting mean activity rates as the measure of remapping also yielded similar results
(Fig. S2), indicating that the concordance between predictive structure and degree of remapping
we observe is not dependent upon the rate map-based assumptions inherent in PV correlations.
We note that positive values of τ which overweight current session experience yielded numerically
weaker correlations between successor structures and PV structures. This provides additional
evidence that the concordance we observe between these structures at maximizing
parameterizations is not due to low-level within-session confounds. Together, these results
demonstrate that compartments with more similar predictive navigational structures on a

particular spatiotemporal scale are represented more similarly in CA1, consistent with a predictive
theory of cognitive mapping.
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Figure 3. CA1 PV structure matches the similarity of predictive navigational structure on a
particular spatiotemporal scale. *(a) Schematic of pipeline for computing cumulative successor*
structure. (b) Grid search where γ and τ are varied and the mean cumulative successor structure
is correlated with the mean PV structure for each mouse. (c) Mean PV structures and mean
cumulative successor structures for all mice with correlations noted. Ordered from most to least
correlated. (d) Relative weightings derived from γ and τ at maximizing parameterizations. (e)
Correlation between PV structure and cumulative successor structure against a shuffled control
where transition probabilities are shuffled across compartments, mice, and sessions and
controlling for the grid search. (f) Correlation between PV structure and cumulative successor
structure against a shuffled control where transition probabilities are shuffled across sessions
within mouse and controlling for the grid search.
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To gain more insight into how predictive navigational structure varied within and between
mice on this critical timescale (γ = 0.994), we next computed individual session (non-cumulative)
successor structures and correlated these structures between sessions (Fig. 4a). Successor
structures were modestly more correlated within mouse than between mice (Fig. 4b,c), suggesting
some consistency in behavioral patterns across sessions within mouse. However, this varied

considerably between mice, with some mice showing little to no consistency in successor
structures across sessions. This variability is useful for distinguishing the relative weightings of
cumulative behavior (i.e. τ), as consistent predictive structure across sessions makes it difficult to
parse out the relative contributions of individual sessions. It also suggests that the greater degree
of consistency we observe in the CA1 PV structure across sessions is not due to consistent
behavior from session to session alone but rather reflects integration of experience across time.
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Finally, we asked whether other behavioral determinants might explain CA1 PV structure
equally well. We first tested whether transitions between compartments were equally correlated
with PV structure, as might be expected from some instantiations of a predictive theory[19,47]. To
this end, for each session we computed the cumulative number of transitions between each pair
of compartments across all sessions up to and including that session (Fig. 4d). As before, it is
unclear whether experience during different sessions should be weighted equally. Thus, we
weighted the contributions of each session as in Equation (2) and determined the maximizing τ
through a grid search at the same resolution as before (i.e. Fig. 3b). Even at maximizing τ, these
matrices were significantly less correlated with PV structures than cumulative successor
structures (Fig. 4e).
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Figure 4. Successor structures are modestly stable across sessions and cumulative
successor structures better explains PV structure than other behavioral variables. *(a)*
Noncumulative successor structure correlations across all pairwise comparisons of sessions.
White boxes bound session comparisons from the same mice. Note that some mice exhibit high
correlations across sessions while others do not. (b) Successor structure correlations for
comparisons within mouse versus between different mice, treating each pairwise session
comparison independently (rank sum test: Z(2964100) = 8.466, p = 2.545e-17). (c) Successor
structure correlations for comparisons within mouse versus between different mice, treating each
mouse as the unit of analysis (sign rank test: p = 3.223e-02). (d) Schematic for computing
cumulative weighted transitions between compartments. (e) PV structures were more highly
correlated with cumulative successor structures than cumulative transitions between
compartments (sign rank test: Z(5609) = 4.962, p = 6.963e-07). Sessions treated independently.
(f) As in (e), except for velocity similarity (Z(5405) = 4.435, p = 9.219e-06). (g) As in (e), except
for higher mean velocities (Z(5253) = 4.042, p = 5.309e-05). (h) As in (e), except for the similarity

of occupancy times (Z(4769) = 2.790, p = 5.276e-03). (i) As in (e), except for higher mean
occupancy times (j) As in (e), except for more similar degrees of thigmotaxis (Z(4941) = 3.235, p
= 1.218e-03). (k) As in (e), except for higher mean degrees of thigmotaxis (Z(5285) = 4.124, p =
3.718e-05). (l) As in (e), except for the similarity of sampling distributions (Z(5270) = 4.086, p =
4.397e-05). (m) As in (e), except for the similarity of cumulative sampling distributions (Z(4528) =
2.166, p = 3.029e-02). *p<0.05, **p<0.01, ***p<0.001

Next, we asked whether differences in velocity, time spent in each compartment,
thigmotaxis, or general sampling patterns could equally account for PV structure. Neither the
similarity of velocity – measured as the inverse absolute difference between mean velocity in each
compartment – nor higher mean velocities could account for PV structure as well as cumulative
successor structure (Fig. 4f,g). Neither the similarity of occupancy times – measured as the
inverse absolute difference between total occupancy time in each compartment – nor longer mean
occupancy times could account for PV structure as well as cumulative successor structure (Fig.
4h,i). Neither similar degrees of thigmotaxis – measured as the inverse absolute difference
between the proportion of time spent within 5cm of a wall in each compartment – nor higher mean
degrees of thigmotaxis could account for PV structure as well as cumulative successor structure
(Fig. 4j,k). Even the similarity of sampling patterns – measured as the correlation between pixel
occupancy maps within each compartment – could not account for PV structure as well as
cumulative successor structure. This was true regardless of whether we compared only the
sampling for a given session or the cumulative sampling for all sessions up to and including that
session (Fig. 4l,m). Together, these results demonstrate that cumulative successor structures
uniquely resemble CA1 representational structure, even when compared to other associated
behavioral metrics.

Manipulating navigational options induces reorganization of CA1 representational
structure to match the new predictive structure of navigation

Our previous results demonstrate that CA1 exhibits individual differences in the degree of
remapping between compartments which coincides with the similarity of predictive structure on a
particular spatiotemporal scale. While these results provide correlational evidence consistent with
a predictive theory of cognitive mapping, these theories make a stronger causal claim. That is,
changing the movements of a navigator should induce a corresponding change in
representational structure to match the new predictive structure. We next sought to test this
prediction. To this end, in four of our mice we followed the initial free navigation phase (15
sessions; *free$_1$*) with an additional phase of limited navigation (7 sessions; *limited*) and finally a
subsequent free navigation (21 sessions; *free$_2$; Fig. S3*). During the limited phase, each session
consisted of locking the mouse in each compartment for four minutes at a time, twice per session.
Locking was achieved by the experimenter placing a small barrier in the entryway. This barrier
was made of the same material as the walls and fit snuggly in the entryway, matching its
dimensions. When each four-minute epoch ended, the door was removed and the mouse freely
navigated in clockwise order to the next compartment.

Extensive evidence indicates that manipulating environmental features[37,38,43–45,56] and
behavioral contingencies[57–59] can induce remapping in hippocampal CA1. We therefore reasoned
that limiting navigational options, a novel experience for the mouse, would lead to remapping in
CA1 during which new predictive structure might be encoded. Moreover, although this
manipulation does not enforce a particular change to their navigational trajectories, free
navigation typically involved shorter bouts of exploration within each compartment (median bout
duration = 11.13 ± 4.92 s; mean ± standard deviation across Free$_1$ sessions). Thus, we suspected
that limiting options would also lead mice to deviate from their free navigational patterns.

We first confirmed that our manipulation did indeed induce remapping in CA1. To do so,
we correlated PV structures across all sessions within each mouse and compared correlations
between different phases of this experiment. Limiting navigational options led to a clear
reorganization of PV structure on averaged and separately in each mouse (Fig. 5a). Quantifying
this, correlations between free phases and the limited phase were significantly lower than
correlations within the same phase (rank sum tests: $Z(1891217) = 19.020$, $p = 1.163e-80$) or
between $free_1$-$free_2$ ($Z(926848) = -13.985$, $p = 1.917e-44$; Fig. 5b). Correlations between $free_1$-
$free_2$ were numerically high but significantly weaker than same phase correlations ($Z(1874165) =$
6.447, $p = 1.143e-10$). This pattern of results indicates rapid remapping during the limited
manipulation, followed by a near return to the initial representational structure when this
manipulation was relaxed.

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Figure 5. Limiting navigational options induces remapping which encodes the new
predictive structure. *(a) PV structure correlations across all 43 sessions averaged across mice*
and separately for each mouse. Experimental phase boundaries denoted by white lines. (b) PV
structure correlations for comparisons between sessions from the same phase, between free and
limited phases, and between $Free_1$ and $Free_2$ phases. (c) Difference in the correlation between
PV structure and SR_{Free1} and $SR_{Limited}$ for two example mice. Dots indicate individual session
values. Line depicts these data smoothed with a gaussian kernel with 1.5 session standard
deviation. Correlation between the SR structures denoted in the lower right for each mouse. (d)
Correlation between PV structure and both SR_{Free1} and $SR_{Limited}$ for all sessions, aggregating
across mice. For complete statistical information, see Supplementary Table 1. (e) Change in mean
event rate versus the first session, as a function of most active to least active compartments. Soft
shaded lines are individual mice. Bold line is the mean across mice. Shaded box denotes sessions

Given that we observed rapid remapping during the limited phase, we next tested the key
hypothesis that patterns of remapping during the limited phase would *specifically resemble the*
new predictive structure. To do so, for each mouse we computed the mean cumulative successor
structure for the $free_1$ and limited navigation phases at the critical spatiotemporal scale determined
above (γ = 0.994 and τ = -0.25). We refer to these mean cumulative successor structures as
SR_{Free1} and $SR_{limited}$. These structures were only partially correlated with one another within each
mouse (r_{99} = 0.699, r_{208}=0.299, r_{215} = -0.330, and r_{216} = 0.171). We then correlated each of these
successor structures with the PV structure for each session and compared these correlations
across phases (Fig. 5c,d). This analysis revealed that during the $free_1$ phase, PV structures were
highly correlated with SR_{Free1} but uncorrelated with $SR_{limited}$. Crucially, during the limited phase PV
structures became correlated with $SR_{limited}$, representing a significant increase over $free_1$ phase
correlations. Correlations with $SR_{limited}$ numerically exceeded those with SR_{Free1}, though this
difference was statistically marginal. Finally, during $free_2$ the pattern of correlations reverted to
that of $free_1$. These results did not depend on the inclusion of pixels near the entryway in the
analysis (Fig. S4). Together, these results demonstrate that our manipulation specifically induced
a pattern of remapping which matched the new predictive structure at the critical spatiotemporal
scale, consistent with a predictive framework.

Our findings indicate that limiting navigational options led to rapid encoding of new
predictive structure while the subsequent return to free navigation prompted a return of the $free_1$
representational structure. This pattern suggests that the rate of encoding of predictive structure
appears to be specific to the experimental phase. Though speculative, we asked whether we
could observe other hallmarks of encoding which might corroborate this possibility. We noted that
during $free_1$ activity tended to become sparser over repeated sessions, as reflected by a decrease
in the mean activity rate in dispreferred compartments (Fig. 1f). This decrease roughly matched
the optimal weighting of predictive structure across sessions (τ = -0.25). We therefore
characterized sparsity across the entire experiment utilizing our full dataset (n = 233 sessions).
This analysis revealed that during limited navigation mean activity rates in dispreferred
compartments returned to their initial levels but immediately reverted to their pre-limited rates
during the subsequent $free_2$ (Fig. 5e). Qualitatively, during the limited phase mean activity rates
in dispreferred compartments decreased across sessions in a way echoing $free_1$, while $free_2$
continued the trending decrease from $free_1$. Mean activity rates in the most active compartment
also decreased significantly during the limited phase and returned to their $free_1$ levels during the
subsequent $free_2$ phase. Overall, this pattern provides tentative evidence of phase-specific activity
changes coinciding with enhanced encoding of predictive structure. Moreover, these findings
suggest that the sparsity of the CA1 representation might serve as a proxy for the rate at which
predictive structure is encoded.

A computational model encoding predictive structure over geometrically structured inputs
uniquely accounts for key results

We have shown that in a complex environment CA1 instantiates idiosyncratic patterns of
remapping between compartments which are correlated with the similarity of predictive
navigational structure on a particular spatiotemporal scale. To gain insight into how this pattern of
results might arise, we implemented a series of computational models intended to mimic the input-
output transformation carried out by CA1. We varied two aspects of these models: the spatial
structure of their inputs and the transformation they compute on these inputs.

We considered inputs with two types of spatial structure. In both cases, inputs consisted
of cells tuned to a single preferred location in each compartment. In the case of *geometrically*
structured inputs, the preferred locations were in geometrically identical locations in each
compartment for a given cell (Fig 6a). In the case of *randomly structured inputs*, the preferred
location randomly differed between each compartment for a given cell (Fig 6a). Contrasting the
two will help us infer whether spatial tuning of inputs alone is sufficient to reproduce our results,
or whether geometric structure plays an important role.
511
We also considered three types of transformations. Firstly, we considered a simple linear
transformation (*linear*), where each CA1 cell received a small number of equally weighted inputs
(see Methods). Secondly, we considered a predictive transformation (*SR*) where CA1 learns an
approximation of the successor representation over its inputs, as explored previously[24]. In this
case, the weight matrix M which determines the strength of the connection from each input cell
to each CA1 cell is an online approximation of the successor representation updated at each
timestep according to
519

$$M \leftarrow M + \alpha[r(t) + \gamma Mr(t+1) - Mr(t)] r(t) \tag{4}$$

520
521   where $r(t)$ is the vector of input firing rates at time t, γ is the temporal discount factor, and α
522   denotes the learning rate. The total input F_i to each CA1 cell i is then determined by
523

$$F_i = \sum_j M(i,j)r_j(t) \tag{5}$$

524
where M is the weight matrix at the current timestep computed in equation (4), and $r_j(t)$ is the
firing rate of the jth input at the current timestep t. Thirdly, we considered a version of the
predictive transformation where a separate successor representation M is learned for each
compartment (*multi-SR*), and F_i for each CA1 cell is computed based on the M corresponding to
the currently occupied compartment. Finally, for all transformations the output of each CA1 cell
was thresholded such that only the top 10% most excited cells across the population were active
during each timestep and normalized to range between zero and one (see Methods).
532
533   To carry out our simulations, we used the actual navigational trajectories of our mice from
534   the Free$_1$ phase of our experiment. For SR and multi-SR simulations, we used γ = 0.994
535   corresponding to the maximizing parameterization estimated above. To capture the decreasing
536   weighting of cumulative experience over repeated sessions (i.e. τ = -0.25), the learning rate α
537   was set to decay exponentially at τ = -0.25 from its initial value on session one (initial α = 0.005).
538
For each combination of input structures and transformations we first asked whether we
observed reliable individual differences in representational structure, as we observed in our actual
data. To this end, for each model we correlated PV structures between all pairwise comparisons
of sessions and contrasted comparisons from within the same mouse versus between different
mice (Fig. 6b). We found that all models with randomly structured inputs (which differed from
mouse to mouse) recapitulated strong individual differences as indicated by higher correlations
for comparisons within mouse than between different mice. On the other hand, geometrically
structured inputs (which were identical across mice) did not recapitulate individual differences in
every case. Only when these inputs were coupled with learning of predictive structure (i.e. an SR
or multi-SR transformation) were significant individual differences observed. These differences
were particularly strong when predictive structure was learned separately for each compartment
(i.e. multi-SR). These results demonstrate that individual differences are consistent with predictive

encoding of geometrically structured inputs, although they may alternatively reflect other between-mouse differences such as randomness in input structure.

***Figure 6. A predictive model with geometrically structured inputs replicates key results.** (a) Schematic depicting possible model configurations. Peak activity rate noted for each rate map. (b) PV structure correlations among sessions separately for comparisons within the same mouse or between different mice, treating each mouse as the unit of analysis and separately for each model (signed-rank tests; geo-multiSR, p = 2.930-03, geo-SR, p = 0.024, geo-linear, p = 0.101; all random input models, ps = 9.766e-04). Soft lines denote individual mice. (c) Correlations between the mean PV structure and the mean cumulative successor for each mouse, separately for each model. Black dots denote the true correlation. Grey box-and-whiskers denote the control distribution computed by shuffling cumulative successor structures across sessions (1,000 shuffles, nonparametric p-values estimated against surrogate distributions: geo-multiSR, p ~ 0.000, geo-SR, p = 0.430, geo-linear, p = 0.065, rand-multiSR, p = 0.389, rand-SR, p = 0.690, rand-linear, p = 0.504). (d) Example rate maps for actual data and geo-multiSR model CA1. Peak activity rate noted for each rate map. (e) PV structure correlations across all pairwise comparisons of sessions for geo-multiSR model CA1. White boxes bound session comparisons from the same mice. (f) Mean PV structures for geo-multiSR model CA1 and mean cumulative successor structures for all mice with correlations noted, ordered as in Fig. 3c. *p<0.01 **p<0.01, ***p<0.001*

Next, for each combination of input structures and transformations we asked whether
model PV structures were correlated with cumulative successor structures, as we observed in our
actual data. Computing cumulative successor structures at the maximizing parameters (γ = 0.994,
τ = -0.25), we found that PV structures were significantly correlated with cumulative successor
structures in only one case: when inputs were geometrically structured and predictive encoding
was learned separately for each compartment (multi-SR). Neither factor was on its own sufficient,
as other models with geometrically structured inputs or predictive encoding learned separately for
each compartment failed to produce significant correlations. Interestingly, environment-wide
encoding of predictive structure (SR) failed to produce significant correlations even when inputs
were geometrically structured. This is because this model encodes only a single predictive
structure for the entire environment, and the geometric repetition of inputs across compartments
leads the model to learn the average predictive structure across all compartments rather than the
predictive structure particular to each compartment. In sum, among models tested, only a model
with geometrically structured inputs and predictive encoding that is learned separately for each
compartment can account for both individual differences in PV structure and the correlation
between PV structure and successor structure that we observe in our actual data. For
completeness, we provide additional visualizations of output from this model (Fig. 6d,e,f).
589
590 **Discussion**
591
Here we leveraged miniscope imaging of mouse CA1 in a multicompartment environment to test
whether patterns of remapping exhibited hallmarks of predictive structure in a complex
environment. We showed that different mice instantiated different patterns of remapping across
identically shaped compartments in this paradigm. These patterns coincided with the similarity of
predictive navigational structure on a seconds-long timescale, modestly overweighting early
experience. Limiting navigational options in this environment induced a new pattern of remapping
which specifically matched the new predictive structure on this spatiotemporal scale. Finally,
through computational modeling, we show that key results can be account for by a specific
combination of predictive encoding over geometrically structured inputs learned separately within
each compartment. Together, these results demonstrate that the structure of the hippocampal
representation in a complex environment can be predicted from the behavioral history of the
navigator, consistent with a predictive theory of cognitive mapping.
604
In this experiment, we compared patterns of remapping across identical unrewarded
compartments from naiveté through extend experience to isolate a potential influence of predictive
navigational structure on the hippocampal representation distinct from confounding differences in
world features, reward structure, or previous experience. Given that we find evidence of such a
determinant under these conditions, we hypothesize that remapping across environments
intentionally disambiguated by external cues[36,41,43,44] reflects both differences in input features as
well as idiosyncratic differences in navigation. Likewise, these results provide evidence that
predictive structure is an ongoing determinant of CA1 representational structure even in the
absence of external rewards. Thus, we hypothesize that influences of reward structure on
hippocampal coding[60–62] may reflect both differences in rewarding input features as well as
differences in goal-directed navigational trajectories. Our findings also suggest that idiosyncratic
predictive structure likely contributed to the results of other multicompartment paradigms[39,47,52,53]
even when individual differences in representational structure may have been masked by
averaging across animals.
619
We relied on the successor representation as an operationalization of a predictive theory
of hippocampal cognitive mapping[19]. In this model, the hippocampus can be understood as
mapping temporally discounted transitions between states of the world, which can be defined not

only by location but also extra-spatial features. Indeed, one of the most compelling successes of
this model is the ability to make quantitative predictions across scales (single cells, neural
populations, behavior), species (rodents, humans), and domains (physical space, visual spaces,
conceptual spaces)[19,63–65]. Although for simplicity we characterize transitions between locations
as the representational determinant in this work, we imagine that in practice the CA1
representational structure may be better understood as reflecting transitions among its spatially
varying inputs, including those from CA3 and entorhinal cortex[24]. Our modeling reflects this. In
line with this reasoning, we would hypothesize that predictive structure determines CA1 structure
when navigating extra-spatial domains, in both rodents and humans. Additionally, we note that
the successor representation is not the only theory consistent with a predictive determinant of
CA1 representational structure[20–22,66,67]. While our results are consistent with the predictions of
this model, our experiment was not designed to adjudicate between competing theories of
predictive cognitive mapping.
636
Our results demonstrate that CA1 representational structure most closely matches
predictive structure when computed with a seconds-long temporal discount factor and
overweighting early sessions. These characteristics are reminiscent of a unique form of
hippocampal plasticity known as behavioral timescale synaptic plasticity (BTSP)[68,69]. BTSP is a
mechanism by which new place fields can be endogenously or artificially induced in previously
silent CA1 cells by evoking a calcium-mediated plateau potential. Properties of induced place
fields indicate that inputs active within a few seconds of this event are potentiated, that these
fields exhibit skew which depends on the navigational trajectory during the initial plateau potential,
and that endogenous plateau potentials are most common during initial experience[68,70]. It is thus
possible that the predictive determinants of CA1 representational structure we characterize here
are a product of BTSP, and more generally that BTSP provides a biological basis for the normative
claims of predictive mapping theories. If so, then we would expect to observe an increase in
plateau potentials not only in novel environments but also during novel experiences which
provoke representational change[71]. Our observations of changes in sparsity within and across
experimental phases are roughly consistent with this. However, we interpret this finding with
caution due to limitations inherent in our recording technique which do not allow us to distinguish
plateau potentials from typical action potentials.
654
The critical spatiotemporal scale of predictive structure which we characterize here may
also depend upon recording location within the hippocampus. It is known that the scale of spatial
representations differs along the long dorsoventral axis of the hippocampus, progressing from
finely tuned cells in the dorsal hippocampus to coarsely-tuned cells in the ventral[72]. Entorhinal
inputs mirror this arrangement[73–75]. Even if the timescale of plasticity which encodes predictive
structure were to be the same across this axis, differences in the scale of inputs might produce
differences in the scale of encoded predictive structure[76]. In our work here, all populations were
recorded from the dorsal hippocampus. Future work recording from the ventral hippocampus
under similar conditions might address this possibility.

References

1. Tolman, E. C. Cognitive maps in rats and men. *Psychol Rev* **55**, 189–208 (1948).
2. O'Keefe, J. & Nadel, L. *The Hippocampus as a Cognitive Map*. (Oxford University Press, 1978).
3. McNaughton, B. L., Battaglia, F. P., Jensen, O., Moser, E. I. & Moser, M.-B. Path integration and the neural basis of the 'cognitive map'. *Nat Rev Neurosci* **7**, 663–678 (2006).
4. Poulter, S., Lee, S. A., Dachtler, J., Wills, T. J. & Lever, C. Vector trace cells in the subiculum of the hippocampal formation. *Nat Neurosci* **24**, 266–275 (2021).
5. O'Keefe, J. Place units in the hippocampus of the freely moving rat. *Exp Neurol* **51**, 78–109 (1976).
6. O'Keefe, J. & Burgess, N. Geometric determinants of the place fields of hippocampal neurons. *Nature* **381**, 425–428 (1996).
7. Høydal, Ø. A., Skytøen, E. R., Andersson, S. O., Moser, M.-B. & Moser, E. I. Object-vector coding in the medial entorhinal cortex. *Nature* **568**, 400–404 (2019).
8. Deshmukh, S. S. & Knierim, J. J. Influence of local objects on hippocampal representations: Landmark vectors and memory. *Hippocampus* **23**, 253–67 (2013).
9. Knierim, J. J., Neunuebel, J. P. & Deshmukh, S. S. Functional correlates of the lateral and medial entorhinal cortex: objects, path integration and local–global reference frames. *Philosophical Transactions of the Royal Society B: Biological Sciences* **369**, 20130369 (2014).
10. Sargolini, F. *et al.* Conjunctive representation of position, direction, and velocity in entorhinal cortex. *Science* **312**, 758–762 (2006).
11. Hafting, T., Fyhn, M., Molden, S., Moser, M.-B. & Moser, E. I. Microstructure of a spatial map in the entorhinal cortex. *Nature* **436**, 801–806 (2005).
12. Lever, C., Burton, S., Jeewajee, A., O'Keefe, J. & Burgess, N. Boundary Vector Cells in the Subiculum of the Hippocampal Formation. *The Journal of Neuroscience* **29**, 9771–9777 (2009).
13. Ormond, J. & O'Keefe, J. Hippocampal place cells have goal-oriented vector fields during navigation. *Nature* **607**, 741–746 (2022).
14. Barry, C. *et al.* The boundary vector cell model of place cell firing and spatial memory. *Rev Neurosci* **17**, 71–97 (2006).
15. Zhou, Y.-Q. *et al.* Vector coding and place coding in hippocampus share a common directional signal. *Nat Commun* **15**, 10630 (2024).
16. Taube, J. S. & Burton, H. L. Head direction cell activity monitored in a novel environment and during a cue conflict situation. *J Neurophysiol* **74**, 1953–1971 (1995).
17. Taube, J. S. Head direction cells recorded in the anterior thalamic nuclei of freely moving rats. *J Neurosci* **15**, 70–86 (1995).
18. Solstad, T., Boccara, C. N., Kropff, E., Moser, M.-B. & Moser, E. I. Representation of geometric borders in the entorhinal cortex. *Science* **322**, 1865–1868 (2008).
19. Stachenfeld, K. L., Botvinick, M. M. & Gershman, S. J. The hippocampus as a predictive map. *Nat Neurosci* **20**, 1643–1653 (2017).
20. George, D. *et al.* Clone-structured graph representations enable flexible learning and vicarious evaluation of cognitive maps. *Nat Commun* **12**, 2392 (2021).
21. Whittington, J. C. R. *et al.* The Tolman-Eichenbaum Machine: Unifying Space and Relational Memory through Generalization in the Hippocampal Formation. *Cell* **183**, 1249-1263.e23 (2020).
22. Levenstein, D., Efremov, A., Eyono, R. H., Peyrache, A. & Richards, B. Sequential predictive learning is a unifying theory for hippocampal representation and replay. Preprint at https://doi.org/10.1101/2024.04.28.591528 (2024).

23. Chandra, S., Sharma, S., Chaudhuri, R. & Fiete, I. Episodic and associative memory from
spatial scaffolds in the hippocampus. *Nature* **638**, 739–751 (2025).
24. de Cothi, W. & Barry, C. Neurobiological successor features for spatial navigation.
Hippocampus **30**, 1347–1355 (2020).
25. Bennett, L. *et al.* Unifying Subicular Function: A Predictive Map Approach. *bioRxiv* (2024)
doi:10.1101/2024.11.06.622306.
26. Derdikman, D. *et al.* Fragmentation of grid cell maps in a multicompartment environment.
Nat Neurosci **12**, 1325–1332 (2009).
27. Keinath, A. T., Epstein, R. A. & Balasubramanian, V. Environmental deformations
dynamically shift the grid cell spatial metric. *Elife* **7**, (2018).
28. Moore, J. J., Cushman, J. D., Acharya, L., Popeney, B. & Mehta, M. R. Linking
hippocampal multiplexed tuning, Hebbian plasticity and navigation. *Nature* **599**, 442–448
(2021).
29. Zutshi, I. *et al.* Hippocampal neuronal activity is aligned with action plans. *Nature* **639**,
153–161 (2025).
30. Nieh, E. H. *et al.* Geometry of abstract learned knowledge in the hippocampus. *Nature*
595, 80–84 (2021).
31. Climer, J. R., Davoudi, H., Oh, J. Y. & Dombeck, D. A. Hippocampal representations drift
in stable multisensory environments. *Nature* (2025) doi:10.1038/s41586-025-09245-y.
32. Hardcastle, K., Maheswaranathan, N., Ganguli, S. & Giocomo, L. M. A Multiplexed,
Heterogeneous, and Adaptive Code for Navigation in Medial Entorhinal Cortex. *Neuron*
94, 375-387.e7 (2017).
33. Liberti, W. A., Schmid, T. A., Forli, A., Snyder, M. & Yartsev, M. M. A stable hippocampal
code in freely flying bats. *Nature* **604**, 98–103 (2022).
34. Wiener, S. I., Paul, C. A. & Eichenbaum, H. Spatial and behavioral correlates of
hippocampal neuronal activity. *J Neurosci* **9**, 2737–63 (1989).
35. Sun, C., Yang, W., Martin, J. & Tonegawa, S. Hippocampal neurons represent events as
transferable units of experience. *Nat Neurosci* **23**, 651–663 (2020).
36. Leutgeb, S. *et al.* Independent codes for spatial and episodic memory in hippocampal
neuronal ensembles. *Science* **309**, 619–23 (2005).
37. Bostock, E., Muller, R. U. & Kubie, J. L. Experience-dependent modifications of
hippocampal place cell firing. *Hippocampus* **1**, 193–205 (1991).
38. Jeffery, K. J. & Anderson, M. I. Dissociation of the geometric and contextual influences on
place cells. *Hippocampus* **13**, 868–72 (2003).
39. Fuhs, M. C., Vanrhoads, S. R., Casale, A. E., McNaughton, B. & Touretzky, D. S.
Influence of path integration versus environmental orientation on place cell remapping
between visually identical environments. *J Neurophysiol* **94**, 2603–16 (2005).
40. Tanni, S., de Cothi, W. & Barry, C. State transitions in the statistically stable place cell
population correspond to rate of perceptual change. *Curr Biol* **32**, 3505-3514.e7 (2022).
41. Bostock, E., Muller, R. U. & Kubie, J. L. Experience-dependent modifications of
hippocampal place cell firing. *Hippocampus* **1**, 193–205 (1991).
42. Lever, C., Wills, T., Cacucci, F., Burgess, N. & O'Keefe, J. Long-term plasticity in
hippocampal place-cell representation of environmental geometry. *Nature* **416**, 90–4
(2002).
43. Colgin, L. L. *et al.* Attractor-map versus autoassociation based attractor dynamics in the
hippocampal network. *J Neurophysiol* **104**, 35–50 (2010).
44. Wills, T. J., Lever, C., Cacucci, F., Burgess, N. & O'Keefe, J. Attractor dynamics in the
hippocampal representation of the local environment. *Science* **308**, 873–6 (2005).
45. Plitt, M. H. & Giocomo, L. M. Experience-dependent contextual codes in the
hippocampus. *Nat Neurosci* **24**, 705–714 (2021).

46.   Nilchian, P., Wilson, M. A. & Sanders, H. Animal-to-Animal Variability in Partial
766   Hippocampal Remapping in Repeated Environments. *The Journal of Neuroscience* **42**,
767   5268–5280 (2022).
768   47.   Duvelle, É. *et al.* Hippocampal place cells encode global location but not connectivity in a
769   complex space. *Curr Biol* **31**, 1221-1233.e9 (2021).
770   48.   Pnevmatikakis, E. A. & Giovannucci, A. NoRMCorre: An online algorithm for piecewise
771   rigid motion correction of calcium imaging data. *J Neurosci Methods* **291**, 83–94 (2017).
772   49.   Pnevmatikakis, E. A. *et al.* Simultaneous Denoising, Deconvolution, and Demixing of
773   Calcium Imaging Data. *Neuron* **89**, 285–99 (2016).
774   50.   Zhou, P. *et al.* Efficient and accurate extraction of in vivo calcium signals from
775   microendoscopic video data. *Elife* **7**, (2018).
776   51.   Friedrich, J., Zhou, P. & Paninski, L. Fast online deconvolution of calcium imaging data.
777   *PLoS Comput Biol* **13**, e1005423 (2017).
778   52.   Spiers, H. J., Hayman, R. M. A., Jovalekic, A., Marozzi, E. & Jeffery, K. J. Place field
779   repetition and purely local remapping in a multicompartment environment. *Cereb Cortex*
780   **25**, 10–25 (2015).
781   53.   Grieves, R. M., Jenkins, B. W., Harland, B. C., Wood, E. R. & Dudchenko, P. A. Place
782   field repetition and spatial learning in a multicompartment environment. *Hippocampus* **26**,
783   118–134 (2016).
784   54.   Karlsson, M. P. & Frank, L. M. Network Dynamics Underlying the Formation of Sparse,
785   Informative Representations in the Hippocampus. *The Journal of Neuroscience* **28**,
786   14271–14281 (2008).
787   55.   Fenton, A. A. Remapping revisited: how the hippocampus represents different spaces.
788   *Nat Rev Neurosci* **25**, 428–448 (2024).
789   56.   Anderson, M. I. & Jeffery, K. J. Heterogeneous modulation of place cell firing by changes
790   in context. *J Neurosci* **23**, 8827–35 (2003).
791   57.   Krishnan, S., Heer, C., Cherian, C. & Sheffield, M. E. J. Reward expectation extinction
792   restructures and degrades CA1 spatial maps through loss of a dopaminergic reward
793   proximity signal. *Nat Commun* **13**, 6662 (2022).
794   58.   Sosa, M., Plitt, M. H. & Giocomo, L. M. A flexible hippocampal population code for
795   experience relative to reward. *Nat Neurosci* **28**, 1497–1509 (2025).
796   59.   Pettit, N. L., Yuan, X. C. & Harvey, C. D. Hippocampal place codes are gated by
797   behavioral engagement. *Nat Neurosci* **25**, 561–566 (2022).
798   60.   Jarzebowski, P., Hay, Y. A., Grewe, B. F. & Paulsen, O. Different encoding of reward
799   location in dorsal and intermediate hippocampus. *Current Biology* **32**, 834-841.e5 (2022).
800   61.   Sato, M. *et al.* Distinct Mechanisms of Over-Representation of Landmarks and Rewards
801   in the Hippocampus. *Cell Rep* **32**, 107864 (2020).
802   62.   Gauthier, J. L. & Tank, D. W. A Dedicated Population for Reward Coding in the
803   Hippocampus. *Neuron* **99**, 179-193.e7 (2018).
804   63.   Momennejad, I. *et al.* The successor representation in human reinforcement learning. *Nat
805   Hum Behav* **1**, 680–692 (2017).
806   64.   de Cothi, W. *et al.* Predictive maps in rats and humans for spatial navigation. *Current
807   Biology* **32**, 3676-3689.e5 (2022).
808   65.   Garvert, M. M., Saanum, T., Schulz, E., Schuck, N. W. & Doeller, C. F. Hippocampal
809   spatio-predictive cognitive maps adaptively guide reward generalization. *Nat Neurosci* **26**,
810   615–626 (2023).
811   66.   Savelli, F. & Knierim, J. J. Origin and role of path integration in the cognitive
812   representations of the hippocampus: computational insights into open questions. *J Exp
813   Biol* **222**, (2019).
814   67.   Zutshi, I. *et al.* Hippocampal neuronal activity is aligned with action plans. Preprint at
815   https://doi.org/10.1101/2024.09.05.611533 (2024).

68. Bittner, K. C., Milstein, A. D., Grienberger, C., Romani, S. & Magee, J. C. Behavioral time
scale synaptic plasticity underlies CA1 place fields. *Science* **357**, 1033–1036 (2017).
69. Milstein, A. D. *et al.* Bidirectional synaptic plasticity rapidly modifies hippocampal
representations. *Elife* **10**, (2021).
70. Madar, A. D., Dong, C. & Sheffield, M. E. J. BTSP, not STDP, Drives Shifts in
Hippocampal Representations During Familiarization. *bioRxiv* (2023)
doi:10.1101/2023.10.17.562791.
71. Monaco, J. D., Rao, G., Roth, E. D. & Knierim, J. J. Attentive scanning behavior drives
one-trial potentiation of hippocampal place fields. *Nat Neurosci* **17**, 725–31 (2014).
72. Kjelstrup, K. B. *et al.* Finite scale of spatial representation in the hippocampus. *Science*
321, 140–3 (2008).
73. Brun, V. H. *et al.* Progressive increase in grid scale from dorsal to ventral medial
entorhinal cortex. *Hippocampus* **18**, 1200–1212 (2008).
74. Stensola, H. *et al.* The entorhinal grid map is discretized. *Nature* **492**, 72–78 (2012).
75. Giocomo, L. M. *et al.* Topography of head direction cells in medial entorhinal cortex. *Curr
Biol* **24**, 252–262 (2014).
76. Momennejad, I. & Howard, M. W. Predicting the Future with Multi-scale Successor
Representations. Preprint at https://doi.org/10.1101/449470 (2018).
77. Pnevmatikakis, E. A. & Giovannucci, A. NoRMCorre: An online algorithm for piecewise
rigid motion correction of calcium imaging data. *J Neurosci Methods* **291**, 83–94 (2017).
78. Pnevmatikakis, E. A. *et al.* Simultaneous Denoising, Deconvolution, and Demixing of
Calcium Imaging Data. *Neuron* **89**, 285–99 (2016).
79. Mathis, A. *et al.* DeepLabCut: markerless pose estimation of user-defined body parts with
deep learning. *Nat Neurosci* **21**, 1281–1289 (2018).
80. Skaggs, W. E., McNaughton, B. L. & Gothard, K. M. An information-theoretic approach to
deciphering the hippocampal code. in *Advances in neural information processing systems*
1030–1037 (1993).
81. McKenzie, S. *et al.* Preexisting hippocampal network dynamics constrain optogenetically
induced place fields. *Neuron* **109**, 1040-1054.e7 (2021).

Methods

Subjects

Eleven naive mice (C57Bl/6, Charles River; 6 male, 5 female) were housed in pairs in 20 cm x 40
cm cages which included running wheels and additional enrichment. Mice were kept on a 14-hour
light/10-hour dark cycle at 23°C and 30% humidity with food and water ad libitum. All experiments
were carried out during the light portion of the light/dark cycle, and in accordance with University
of Illinois Chicago Animal Use and Care Committee (protocol #23008) and with AAALAC
guidelines.

Surgeries

During all surgeries, mice were anesthetized via inhalation of a combination of oxygen and 5%
Isoflurane before being transferred to the stereotaxic frame (David Kopf Instruments), where
anesthesia was maintained via inhalation of oxygen and 0.5-2.5% Isoflurane for the duration of
the surgery. Body temperature was maintained with a heating pad and eyes were hydrated with
gel (Optixcare). Meloxicam (2 mg kg^{-1}) and saline (0.5 ml) were administered subcutaneously at
the beginning of each surgery. Preparation for recordings involved three surgeries per mouse.

First, at the age of six to ten weeks, each mouse was transfected with a 400 nl injection of the calcium reporter GCaMP8f via the viral construct AAV9.syn.GCaMP8f.WPRE.SV40 with an original titre of 2.3×10^{13} GC ml^{-1} (Addgene) diluted at a 1 part virus to 7 parts sterile artificial cerebrospinal fluid before surgical microinjection. While expression in both principal cells and inhibitory populations is possible under the syn promoter, it is likely that the large majority of cells we record are excitatory given their sparse transients.

Three weeks post-injection, a 1.8mm diameter gradient refractive index (GRIN) lens (Edmund Optics) was implanted above dorsal CA1 (Referenced to bregma: ML = 2.0 mm, AP = -2.1 mm; Referenced to brain surface: DV = -1.35 mm). Implantation required aspiration of intervening cortical tissue. In addition to the GRIN lens, two stainless steel screws were threaded into the skull above the contralateral hippocampus and prefrontal cortex to stabilize the implant. Dental cement (C&B Metabond) was applied to secure the GRIN lens and anchor screws to the skull. A silicone adhesive (Kwik-Sil, World Precision Instruments) was applied to protect the top surface of the GRIN lens until the next surgery.

Three weeks after lens implantation, an aluminum baseplate was affixed via dental cement (C&B Metabond) to the skull of the mouse, which would later secure the miniaturized fluorescent endoscope (miniscope) in place during recording. The miniscope/baseplate was mounted to a stereotaxic arm for lowering above the implanted GRIN lens until the field of view contained visible cell segments and dental cement was applied to affix the baseplate to the skull. A polyoxymethylene cap was affixed to the baseplate when the mice were not being recorded to protect the baseplate and lens.

After surgery, animals were continuously monitored until they recovered. For the initial two days after surgery mice were provided with additional doses of meloxicam for pain management. One week following baseplating, to familiarize mice with the recording procedure and to monitor the quality of cell activity, we recorded mice daily in a 75 cm x 75 cm square open field for 10 min to 30 min per day. When recording quality was deemed of sufficiently high quality based on a stable number of cells across days, rapid transients, and a high proportion of cells with stable place fields within day (>50% of cells versus shuffled controls), mice began the multicompartment experiment. Mice typically met these criteria 2+ weeks following baseplating.

Apparatus

The recording environment was constructed of white Lego base and white acrylic walls (Professional Plastics). All walls had a height of 20 cm. All compartments were 20 cm x 35 cm, with a 20cm central chamber. No other internal directional cues were provided, but the apparatus was closer to one external grey room wall which could serve as an external directional cue. During recording, the environment was dimly lit by an LED lamp positioned to reduce shadows, and a white-noise machine was used to mask any uncontrolled sounds. All sessions were 30 or 40 min (depending on signal strength to minimize photobleaching), and only one session was recorded per day. On rare occasions (~2%) sessions were impacted by equipment failures, and sessions were terminated early; data from these sessions were not analyzed. During all free exploration sessions, mice were placed in the center of the environment at the start of the session facing the nearer external grey room wall. During limited navigation sessions, mice were placed in the first room at the start of the session also facing the nearer external grey wall. Mice were not intentionally disoriented prior to recording, and the stability of patterns of remapping across days suggests that they remained oriented throughout. The recording environment was cleaned between recordings with veterinarian-grade disinfectant.

Data acquisition

In vivo calcium videos were recorded with a UCLA miniscope (v3; miniscope.org) containing a monochrome CMOS imaging sensor (MT9V032C12STM, ON Semiconductor) connected to a custom data acquisition (DAQ) box (miniscope.org) with a lightweight, flexible coaxial cable. The DAQ was connected to a PC with a USB 3.0 SuperSpeed cable and controlled with Miniscope custom acquisition software (miniscope.org; software version v4). The outgoing excitation LED was set to between 5-30%, depending on the mouse to maximize signal quality with the minimum possible excitation light to mitigate the risk of photobleaching. Gain was adjusted to match the dynamic range of the recorded video to the fluctuations of the calcium signal for each recording to avoid saturation. Behavioral video data were recorded by a webcam mounted above the environment. The DAQ simultaneously acquired behavioral and cellular imaging streams at 30 Hz as FFV1 losslessly-compressed AVI files and all recorded frames were timestamped for post-hoc alignment.

Data preprocessing

Calcium imaging data were preprocessed prior to analyses via a pipeline of open source MATLAB (MathWorks; version R2024a) functions to correct for motion artifacts[77], segment cells and extract transients[78]. The motion-corrected calcium imaging data were manually inspected to ensure that motion correction was effective and did not introduce additional artifacts. Following this preprocessing pipeline, the spatial footprints of all cells were manually verified to remove lens artifacts. Finally, to estimate the spike trains which gave rise to the transients we recorded, we implemented a second-order autoregressive deconvolution algorithm[51]. This nonnegative vector was then binarized such that any frame with a nonzero likelihood was treated as active (i.e. 1), otherwise it was treated as not active (i.e. 0). All further analysis was conducted on this binary activity vector.

Position data were inferred from behavioral videos via DeepLabCut[79] tracking the nose, ears, and tail base of the mouse. The average between the two ears was taken as the position of the mouse in all analyses. Periods were estimates of either ear positions were low-confidence (<0.90) or the estimated distance between the ears was unrealistic were excluded and position was linearly interpolated across these timepoints (<0.5% of timepoints). Position data were then resampled via linear interpolation based on system clock timestamping to estimate the position at each time when imaging frames were collected.

Data analysis

All analyses were conducted using the binarized nonnegative spike train estimates inferred via a second-order autoregressive deconvolution, henceforth the *activity* vector.

Rate maps. Rate maps for each cell were computed by dividing the environment (or each compartment) into 2.5 cm x 2.5 cm pixels, computing the mean activity rate at each pixel for that cell, and then smoothing the result map with an isotropic gaussian kernel with standard deviation of 2.5 cm. These maps were then corrected for edge artifacts and artifacts induced by smoothing over unsampled pixels as follows (MATLAB function *nanconv* available at https://www.mathworks.com/matlabcentral/fileexchange/41961-nanconv). First, a second map where sampled pixels are set to one and all other pixels are set to zero was computed. This maps was then smoothed with the same kernel, and each rate map was divided by this smoothed map. Unsampled pixels were excluded from further comparisons.

Split-half stability. To characterize the stability of each cell's spatial tuning during a session,
we computed its split-half stability by computing whole environment rate maps using only the first
half or second half of the data from that session and correlating the two rate maps. To determine
the statistical significance of these correlations, we compared this true correlation to a surrogate
distribution computed by randomly circularly shifting the position vector relative to the activity
vector by at least 30 seconds and recomputing the resulting correlation 1000 times. Cells with
true correlations exceeding the 95% of this surrogate distribution were considered spatially-stable
and included in further analysis.
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961   *Spatial information content*. To characterize the specificity of each cell's spatial tuning
962 during a session, we computed its spatial information content from its whole environment rate
963 map as described previously[80] via the equation:
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$$\text{Spatial information content} = \sum_i \mathbf{p}_i \mathbf{a}_i \log\left(\frac{\mathbf{a}_i}{\bar{\mathbf{a}}}\right) \tag{6}$$

965
where i is the rate map pixel index, $\mathbf{p}_i$ is the probability of sampling pixel i, $\mathbf{a}_i$ is the mean firing
rate at pixel i, and $\bar{\mathbf{a}}$ is the mean firing rate across all pixels. To determine the statistical
significance of these values, we compared the true spatial information content to a surrogate
distribution computed by randomly circularly shifting the position vector relative to the activity
vector by at least 30 seconds and recomputing the resulting correlation 1000 times. Cells with
true correlations exceeding the 95% of this surrogate distribution were considered spatially tuned
and included in further analysis.

Similarity structures. Population vector correlations between compartments were
computed by aligning these rate maps by the entryway, concatenating the linearized compartment
rate maps of all cells from each compartment, and computing the correlation between these
vectors. This measure of takes into account both changes in the spatial distributions of firing and
relative changes in firing rate. Mean activity rate correlations between compartments were
computed by correlating the vector of mean activity rates for all included cells between
compartments. Successor structure and cumulative successor structure were computed as
described in the main text. To compute successor matrices, position data were binned to 5 cm x
5 cm pixels, and the timestep was set to 1/30th of a second to match the frame rate of data
acquisition.

Computational modeling

To compute activity vectors of model inputs, first each compartment was divided into a
grid of possible preferred locations at 1.25 cm x 1.25 cm increments (448 possible preferred
locations). Each cell was assigned one of these preferred locations (448 input cells total). For
geometrically structured inputs, each cell had the same preferred location across all
compartments, aligned by their entryways. For randomly structured inputs, the assignment of
preferred locations was randomly shuffled across cells for each compartment. Once assigned,
each cell kept its preferred locations for all sessions. At each timestep, the activity of each cell
was determined from its distance to its nearest preferred location according to a gaussian
distribution with standard deviation 5 cm. Finally, input activity vectors were normalized to scale
from [0, 1] inclusive. Each transformation received identical input traces for the geometrically
structured inputs and randomly structured inputs.

For the linear transformation, each CA1 cell received a random number of inputs on the
range [2, 16] drawn from a Poisson distribution with $\lambda = 4$. These inputs were chosen from the
input population at random and given equal weight (i.e. 1). All other inputs were assigned a weight
of 0 for that CA1 cell. Once assigned, these weights were constant across all sessions. SR and
multi-SR transformations are described in the main text. In all cases, the successor matrices were
initialized as the identity matrix.

Finally in all cases the output O_i from CA1 cell i was determined at each timestep by

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$$O_i = \frac{\Theta(F_i - T_{90\%})}{\max_j \left(\Theta\left(F_j - T_{90\%}\right) \right)} \tag{7}$$

1003
1004   where F_i is the total input to CA1 cell i, $T_{90\%}$ is a dynamic threshold corresponding to the F of the
1005   cell with the 90th percentile F across the population at that timestep, and
1006

$$\Theta(\text{x}) = \begin{cases} x & \text{if } x > 0 \\ 0 & \text{otherwise} \end{cases} \tag{8}$$

1007
The result of this thresholding and normalization is that only 10% of CA1 cells are active during
each timestep with output rates ranging from [0, 1]. This dynamic normalization is intended to
mimic competition among principal cells in CA1[81], and is especially useful when changes in
connectivity during learning lead to large changes in excitatory drive as is often the case in our
SR and multi-SR simulations. For each model, we simulated 448 CA1 cells, matching the number
of input cells. Output vectors were treated like the activity vectors of our actual data in all further
analyses. SR and multi-SR transformations were deterministic and therefore a single iteration is
presented in the main text. A single iteration of the linear transformation model was also included
to match the presentation of SR and multi-SR simulations, despite some additional randomness
in its implementation. However, we note that we have run approximately 10 iterations at the
identical parameterization and in all cases the results closely resemble those presented in the
main text.
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Histological validation of expression and recording targets

After experiments, animals were perfused to verify GRIN lens placement. Mice were deeply
anesthetized and intracardially perfused with 4% paraformaldehyde in PBS. Brains were
dissected and post-fixed with the same fixative. Coronal sections (50 µm) of the entire
hippocampus were cut using a vibratome and sections were mounted directly on glass slides.
Sections were split and half of all sections were stained for DAPI and mounted with Fluoromount-
G (Southern Biotechnology) to localize GRIN lens placement and to evaluate viral expression.
Due to the large imageable surface but restricted miniscope field of view (~0.5 mm x ~0.8 mm),
we were unable to determine more specific localization of populations within the hippocampus for
mice recorded with 1.8 mm lenses.

Statistics and reproducibility

All statistical tests are noted where the corresponding results are reported throughout the main
text and supplement. All tests were uncorrected 2-tailed tests unless otherwise noted. Z-values
for nonparametric rank-sum and signed-rank tests were not estimated or reported for
comparisons with fewer than 15 datapoints. Box plots portray the minimum and maximum

(whiskers), upper and lower quartiles (boxes), and median (cinch/bolded line). All correlations are
Pearson's correlations unless otherwise noted.

Code availability

All custom code written for reported analyses are publicly available at [insert Github link] or via
request to the corresponding authors.

Data availability

The complete dataset for all experiments are publicly available at [insert Dryad link] or via request
to the corresponding authors.

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Acknowledgements

This work was supported by startup funding provided by the University of Illinois Chicago, as well as a UIC LAS CSSR seed grant. We would also like to thank J. Quinn Lee, Rachel Donka, and Rachel Barrett for feedback when drafting this manuscript.

Author Contributions

SAP and ATK contributed to experimental design, surgeries, recordings, analysis of data, as well as revising the manuscript. ATK drafted the manuscript.

Competing interests.

The authors declare no competing interests.

1067 Supplementary information for:
1068
1069 **The predictive structure of navigation determines mouse CA1 representational structure**
1070 **in a multicompartment environment**
1071
1072 Pecirno & Keinath
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Supplementary Figure 1. Individual differences in patterns of remapping across
compartments in a multicompartment environment as assessed by comparing mean
activity rate. *(a) Schematic of mean activity rate (MAR) structure analysis pipeline. (b) MAR*
structure correlations across all pairwise comparisons of sessions. White boxes bound session
comparisons from the same mice. (c) MAR structure correlations for comparisons within mouse
versus between different mice, treating each pairwise session comparison independently (rank-
sum test: z(3395800) = 16.687, p = 16.687). (d) MAR structure correlations for comparisons within
mouse versus between mice, treating each mouse as the unit of analysis (signed-rank test: p =
9.766e-04). Soft lines denote individual mice. (e) Distribution of compartment-to-compartment
MAR correlation values for each mouse. (f) MAR structures averaged across sessions for each
mouse. ***p<0.001
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1088 ***Supplementary Figure 2. Patterns of remapping in CA1 resemble the similarity of predictive***
1089 ***navigational structure on a particular spatiotemporal scale.*** *(a) Grid search where γ and τ*
1090 *are varied and the mean cumulative successor structure is correlated with the mean PV structure*
1091 *shown separately for each mouse. (b) Grid search where γ and τ are varied and the cumulative*
1092 *successor structure is correlated with the PV structure, treating each session as an independent*
1093 *statistical unit. (c) Correlation between PV structure and cumulative successor structure against*
1094 *a shuffled control where transition probabilities are shuffled across compartments, mice, and*
1095 *sessions and controlling for the grid search, treating each session as an independent statistical*
1096 *unit. (d) Correlation between PV structure and cumulative successor structure against a shuffled*
1097 *control where transition probabilities are shuffled across sessions within mouse and controlling*
1098 *for the grid search, treating each session as an independent statistical unit. (e) Grid search where*
1099 *γ and τ are varied and the mean cumulative successor structure is correlated with the mean*
1100 *activity rate (MAR) structure for each mouse. (f) Correlation between MAR structure and*
1101 *cumulative successor structure against a shuffled control where transition probabilities are*
1102 *shuffled across compartments, mice, and sessions and controlling for the grid search, averaging*
1103 *within mouse prior to correlation. (g) Correlation between PV structure and cumulative successor*
1104 *structure against a shuffled control where transition probabilities are shuffled across sessions*
1105 *within mouse and controlling for the grid search, averaging within mouse prior to correlation.*

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Supplementary Figure 3. Example rate maps for all experimental phases. *Example rate*
maps for the first and last session of each phase for Mouse 208. Examples chosen from the
equivalent percentile of the spatial information distribution for each session, ordered from
highest to lowest. Peak activity rate noted for each rate map in black. Spatial information noted
for each rate map in red.

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Supplementary Figure 4. Excluding data recorded near the entryway did not substantially change the results. *(a) For all analyses in this figure, data from the pixels near the entryway were excluded from both the PV structure and successor structure computations. (b) PV structure correlations across all pairwise comparisons of sessions when excluding entry pixels. White boxes bound session comparisons from the same mice. (c) PV structure correlations for comparisons within mouse versus between different mice when excluding entry pixels, treating each pairwise session comparison independently (rank-sum test: z(3148018) = 11.968, p = 5.207e-33). (d) PV structure correlations for comparisons within mouse versus between mice when excluding entry pixels, treating each mouse as the unit of analysis (signed-rank test: p = 9.766e-04). Soft lines denote individual mice. (e) Mean correlation between PV structure and cumulative successor structure at the parameterization γ = 0.994, τ = -0.25 when excluding entry pixels. Shuffled distribution is the mean correlation across 10,000 random shuffles of transition matrices across compartments, sessions, and mice prior to computing cumulative successor structures. P-value estimated nonparametrically from this distribution. (f) Correlation between PV structure and both SR_{Free1} and SR_{Limited} for all sessions, aggregating across mice. For complete statistical information, see Supplementary Table 1. **p<0.01, ***p<0.001*

Experimental Phase	Comparisons	Statistical test	Statistic	Outcome
Main text results (Figure 4c)				
$Free_1$	SR_{Free1} vs. 0	Signed rank	1762	4.510e-10
$Free_1$	$SR_{Limited}$ vs. 0	Signed rank	1151	8.233e-02
$Free_1$	SR_{Free1} vs. $SR_{Limited}$	Signed rank	1528	6.402e-06
Limited	SR_{Free1} vs. 0	Signed rank	346	1.129e-03
Limited	$SR_{Limited}$ vs. 0	Signed rank	363	2.690e-04
Limited	SR_{Free1} vs. $SR_{Limited}$	Signed rank	124	7.203e-02
$Free_2$	SR_{Free1} vs. 0	Signed rank	3484	3.532e-14
$Free_2$	$SR_{Limited}$ vs. 0	Signed rank	1942	4.838e-01
$Free_2$	SR_{Free1} vs. $SR_{Limited}$	Signed rank	3272	3.319e-11
$Free_1$ vs. Limited	SR_{Free1}	Rank sum	3064	4.231e-04
$Free_1$ vs. Limited	$SR_{Limited}$	Rank sum	2385	1.081e-02
$Free_1$ vs. $Free_2$	SR_{Free1}	Rank sum	4263	7.260e-01
$Free_1$ vs. $Free_2$	$SR_{Limited}$	Rank sum	4431	7.443e-01
Limited vs. $Free_2$	SR_{Free1}	Rank sum	1011	1.264e-04
Limited vs. $Free_2$	$SR_{Limited}$	Rank sum	1987	6.568e-03
With pixels near the entrance removed (Supplementary Figure 3)				
$Free_1$	SR_{Free1} vs. 0	Signed rank	1771	2.947e-10
$Free_1$	$SR_{Limited}$ vs. 0	Signed rank	1088	2.028e-01
$Free_1$	SR_{Free1} vs. $SR_{Limited}$	Signed rank	1553	p = 2.644e-06
Limited	SR_{Free1} vs. 0	Signed rank	344	1.324e-03
Limited	$SR_{Limited}$ vs. 0	Signed rank	379	6.129e-05
Limited	SR_{Free1} vs. $SR_{Limited}$	Signed rank	122	6.511e-02
$Free_2$	SR_{Free1} vs. 0	Signed rank	3504	1.770e-14
$Free_2$	$SR_{Limited}$ vs. 0	Signed rank	1967	4.170e-01
$Free_2$	SR_{Free1} vs. $SR_{Limited}$	Signed rank	3020	3.633e-08
$Free_1$ vs. Limited	SR_{Free1}	Rank sum	3007	2.573e-03
$Free_1$ vs. Limited	$SR_{Limited}$	Rank sum	2292	7.199e-04
$Free_1$ vs. $Free_2$	SR_{Free1}	Rank sum	4390	8.728e-01
$Free_1$ vs. $Free_2$	$SR_{Limited}$	Rank sum	4374	9.241e-01
Limited vs. $Free_2$	SR_{Free1}	Rank sum	1172	5.930e-03
Limited vs. $Free_2$	$SR_{Limited}$	Rank sum	2015	3.659e-03

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1130 *Supplementary Table 1. Statistical outcomes for comparisons of correlation between PV*
1131 *structures and both SR_{Free1} and $SR_{Limited}$ as in Figure 4c and Supplementary Figure 3.*