The Blueprint for Survival: The Blue Dasher Dragonfly as a Model for Urban Adaptation

Ethan R. Tolman[1,2,3]*, Ellie Gamett[2,4]*, Christopher D. Beatty[3,6]*, Aaron Goodman[2,5]*, Brittney Hahn[2], ,
Christian Benischek[3,7], Gracie Castillo[3,7], Ethan Derderian[7], Santiago Fernandez-Juarez[3,7], Ben
Gallafent[3], James Jenson[3,8], Dick Jordan[3], Magnolia Schneider[3,7], Roberto Salazar[1], Towako Tamano[3],
Maleah Wei[3], Jacob Idec[9], Rob Guralnick[9], Jessica L. Ware[2], Manpreet K. Kohli[,2,3,5,10]

Affiliations:
1. Department of Biological Sciences, Virginia Tech, Blacksburg, Virginia, USA
2. Department of Invertebrate Zoology, American Museum of Natural History, New York, New York, USA
3. Conservation Connection Foundation, Boise, Idaho, USA
4. Department of Ecology, Evolution, and Environmental Biology, Columbia University, New York, New York, USA
5. CUNY Graduate Center, New York, New York, USA
6. Program for Conservation Genomics, Department of Biology, Stanford University, Stanford, California, USA
7. The College of Idaho, Caldwell, Idaho, USA
8. Boise State University, Boise, Idaho, USA
9. Florida Museum of Natural History, University of Florida, Gainesville, FL, USA
10. The Department of Natural Sciences, Baruch College, New York, New York, USA

Abstract

Human alteration of natural environments and habitats is a major driver of species decline. However, a handful of species thrive in human altered environments. The biology, distribution, population structure, and molecular adaptations enabling certain species to thrive in human-altered habitats are not well understood. Here, we evaluate the population and functional genomics, ecological niche and distributions, and geometric morphometrics of the blue dasher (*Pachydiplax longipennis*), one of the most ubiquitously observed insects in human altered habitats. Using resequencing data we identify a number of genes involved with the success of the blue dasher in human altered habitats, including loci contributing to immune function and response to oxidative stress. Some genes related to these functions are found in regions of strong population structure, while others are not, potentially indicating both regional and widespread adaptations to urban environments within this species. Using one of the most robust locality datasets for any species to date, we also generate habitat suitability predictions which show that *P. longipennis* has spread with urbanization, suggesting humans have created suitable habitat for this species. These results complement morphological and genomic data showing *P. longipennis* (particularly East of the Rocky Mountains) has the capacity to rapidly disperse to newly suitable habitats. Given the shared barriers to colonizing an urban habitat, we expect that many of the adaptations we have identified in *P. longipennis* can be used for predicting what animals might succeed in urban habitats more generally.

Keywords

Urban ecology, species resiliency, human-modified landscapes, species distribution, Anisoptera, Libellulidae

Introduction

In the Anthropocene, population decline is the norm for many species [1,2]. However, some species are thriving on our changing planet. One such "winner" of the Anthropocene is the blue dasher dragonfly, *Pachydiplax longipennis* (Brauer 1868). Not only is the range of this species thought to be expanding with a warming climate [3], gene family expansions related to the processing of free radicals and immune function are hypothesized to allow this species to live in highly polluted lentic bodies of water near

humans [4]. As a highly photogenic "percher" species which spends a great deal of its time perched on

vegetation [5,6], *P. longipennis* is easily detectable through community science tools, such as iNaturalist.

Due to its proximity to urban habitats, *P. longipennis* was the most observed dragonfly on iNaturalist

through December 2024 and the 40th most observed species in its distribution. As a robust generalist with

the potential to displace specialists [4,7], a comprehensive analysis of this species is important for the

monitoring of aquatic insect communities on a rapidly changing planet, yet no work has been done to

explore regional genetic, genomic and phenotypic differences in *P. longipennis*. Here we consider the life

history, population and functional genomics, geometric morphometrics, and ecological niche of *P.*

longipennis populations in North America to test the hypothesis that this species is expanding its range

and is equipped for colonizing newly altered habitats where more specialist species are unable to persist.

We also identify loci involved in success in urban habitats, and more generally, make a case for

establishing *P. longipennis* as an indicator of altered habitats.

Pachydiplax longipennis life history

P. longipennis is distributed across North America, from southern Canada in the north, to Mexico and

Belize in the south, and is found on either side of the Rocky Mountains [6,8]. It is also found on the

Caribbean islands of Cuba and the Bahamas, as well as Bermuda, although it is currently unclear when or

how it arrived on these islands [8]. Blue dashers are medium to small dragonflies in the family

Libellulidae, roughly 2.5 to 4.5 cm in length, with striped thoraces, dark colouration at the base of their

wings, and waxy blue pruinosity in mature males [6]. *Pachydiplax* is a monotypic genus that has been

consistently recovered in a clade that includes the genera *Micrathyria* and *Zenithoptera* [9–11]. The

features of males, including their size and dispersal [12,13], aggressiveness, feeding and territoriality [14–

18], and nymph microhabitat preference and emergence details [19], have been well studied, but data on

females is generally lacking.

P. longipennis has long been considered a potential migrant species, as it has been found in

swarms of migrating *Anax junius* on several occasions [20–22], and was considered a putative migrant by

the (no longer active) Migratory Dragonfly Partnership [23]. Recently, individuals of *Pachydiplax*

longipennis were observed in a large migratory mixed swarm of dragonflies that flew over Rhode Island

in July of 2024 [20] . Migratory, or even high dispersing, behavior may allow *P. longipennis* to more

rapidly colonize newly suitable habitat, which has been observed anecdotally [20,22], but no molecular or

morphological study has been used to assess gene flow within, or the migratory capabilities of, this

species.

Materials and Methods

Population genomics

We obtained low coverage 150 bp Illumina data from Azenta Life Sciences for 15 individuals, five each

from Boise, Idaho; Newark, New Jersey; and Knoxville, Tennessee. We mapped the trimmed reads,

delivered from the manufacturer, with BWA [24], to the previously assembled reference genome of *P.*

longipennis [4], using default settings. We marked and removed duplicates with Picard [25], and called

variants with bcftools v1.15.1 [26], using the options: `bcftools mpileup -a AD,DP,SP -Ou -`

`f reference.fasta. Input_bam | bcftools call -f GQ,GP -mO z -o output.`

We filtered the resulting vcf file for a minimum quality score of 30, and a minimum and maximum depth

of 1 and 6 respectively. We utilized PLINK v1.90b6.21 [27] to prune linked sites. To estimate population

structure, we performed a principal components analysis (PCA) and calculated the F_{ST} between and

within each of the three populations using the linkage-pruned SNPs with VCFtools v0.1.16 [27]. To

estimate the ancestry in each of our populations, we generated admixture models assuming 2-5 ancestral

populations with ADMIXTURE v1.3.0 [28] and evaluated the models with cross-validation error. The

best fitting admixture-model and the PCA plots were plotted in R using mapmixture v1.1.0 [29].

In addition to the called SNPs, we also estimated population structure analyses from genotype

likelihoods. We used ANGSD v.0.921 [30] to calculate genotype likelihoods with the options "`-`

`remove_bads 1 -uniqueOnly 1 -only_proper_pairs 1 -C 50 -baq 1 -minMapQ`

`30 -minQ 30 -minInd 13 -doSnpStat 1 -doHWE 1 -sb_pval 1e-6 -hwe_pval`

`1e-6 -hetbias_pval 1e-6 -doMajorMinor 1 -skipTriallelic 1 -doMaf 2 -`

`doPost 1 -minMaf 0.05 -snp_pval 1e-6 -doGeno 8 -doGlf 2`". We then used

ngsLD v1.2.1 [31] to identify unlinked sites, which were used as input for PCangsd [32], to evaluate

population structure through a PCA. We also estimated admixture models from the genotype likelihoods

using NGSadmix [28], with 2-5 ancestral populations. As with the SNP analyses, both the principal

components and admixture analyses were plotted in R using mapmixture [29].

To identify specific genomic regions and genes of high structure between our three populations,

we used VCFtools [26] to calculate Fst across a sliding window of 10,000 bp, with a step size of 1,000bp.

We then identified genes, (previously annotated by Tolman et al. [33]) in windows of high (F_{ST} >.5) and

low (F_{ST} <.01) structure between populations and used REVIGO [34] to find and summarize the

molecular functions of the genes in these regions into treemaps, using the previously published functional

annotations [4] .

Demographic modeling

To model historical gene flow between our sampled populations of *P. longipennis* we generated a

demographic model for *P. longipennis* using GADMA2 v2.0.0 [35]. We used the LD-trimmed SNP file as

input with the default engine moments [36], a mutation rate of 2e-9 (previously shown to be a reasonable

estimate for Odonata [37]), and the local optimizer BFGS_log (see supplementary materials for full

configuration file).

COI Haplotype Structure

To broaden our population sampling we downloaded all COI sequences for *P. longipennis* from BOLD

[38], and also extracted the COI sequence from the reference assembly of *P. longipennis* [33], which was

generated from one individual from Boise, Idaho. We also obtained Sanger sequencing (performed by

Genewiz) for the COI gene (using primers and protocol as specified in Kohli et al [39]) for samples

collected in New Jersey and Tennessee. The sequences were aligned in CLUSTAL OMEGA (online

server https://www.ebi.ac.uk/jdispatcher/msa/clustalo) [40] and then manually corrected in MESQUITE

v1.8 [41]. All together we had sequences from the following locations: Florida (24), Ontario (7),

Arkansas (9), Texas (5), Arizona (11), Oklahoma (1), British Columbia (1), New Jersey (19), Idaho (1),

and Tennessee (2). The population structure and the corresponding haplotype network and population

statistics for COI were estimated in software package PopART [42]. To test for population structure on

either side of the Rocky Mountain divide, we considered Arizona, Idaho, and British Columbia to belong

to a "West Coast" population, with the remaining individuals belonging to an "East Coast" population,

and used PopART [42] to estimate the COI F_{ST} between the West and East coast populations. As the total

sequence length for two Tennessee individuals was less than other samples, we performed all our COI

analyses in PopART both with and without the individuals from Tennessee.

145
146 ***Morphological analysis***
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1) Hindwing length   6) Forewing length   11) Thorax length
2) Hindwing depth   7) Forewing depth   12) Hind femur length
3) Hindwing distal half   8) Forewing distal half
4) Hindwing proximal half   9) Forewing proximal half
5) Hindwing pterostigma   10) Forewing pterostigma

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149 *Figure 1: Morphological measurements taken of P. longipennis. An example of a photograph taken for*
150 *measurements, with the measurements labeled on the figure.*

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To investigate regional morphological differences in the wings of *P. longipennis*, which can have a major

effect on dragonfly dispersal [43], we collected specimens from two locations: Boise, Idaho and Herndon,

Virginia and compared them to museum specimens from Secaucus, New Jersey and Knoxville,

Tennessee. We collected a total of 28 individuals from the combined four locations (14 from Idaho, 6

from Virginia, 9 from Tennessee, and 10 from New Jersey). Live specimens from Virginia and Idaho

were carefully secured by their wings to labeled index cards using paper clips to avoid any damage to the

dragonflies. Each individual was photographed alongside a ruler for scale reference (Fig. 1). Museum

specimens were scanned with an Epson Perfection V550 Photo flatbed color scanner according to a

digitization template created for the Targeted Odonata Wing Digitization Project v2.0 (Supplementary

figure 1). Specimens were imaged with the ventral side of the right pair of wings facing down on the

glass.

Four researchers then independently measured the hindwing length, hindwing depth, hindwing

pterostigma length, length of the hindwing distal half (wing node to tip), length of the hindwing proximal

half (wing node to base), forewing length, forewing depth, forewing pterostigma length, length of the

forewing distal half, length of the forewing proximal to the body, thorax length and femur length (Fig. 1)

in ImageJ [44]. An average of each measurement taken by the four researchers was used in statistical

analyses. We attempted to capture the area of the fore and hindwings using the protocol established by

Idec et al. [45].

To determine whether there were significant morphological differences between the four

geographic populations (Idaho, Virginia, Tennessee, and New Jersey) that may affect the dispersal

capability of *P. longipennis*, we applied the Kruskal-Wallis test in R v4.1.4 [46] by scaling each wing

measurement according to thorax length and femur length to control for differing wing measurements

being a result of a larger body size in general between the populations [47,48]. This non-parametric

method is suitable for comparing more than two groups, as it does not assume a normal distribution.

Variables with p-values below 0.05 were considered to show significant regional differences. As our

gender sampling was uneven (Idaho were all males, all but one individual from New Jersey were females,

and the other populations were mixed) we compared males and females in a combined sample of the three

genetically similar east coast populations with a Wilcoxon rank-sum test. As we determined that our wing

measurements differed by sex when normalized by thorax length, but not when normalized by femur

length, we considered measurements normalized by femur length in data visualization and further

analyses. To further visualize the morphological space inhabited by each population we conducted a

PCA. We imputed 26 missing measurements for each population using the package mice v3.16.0 [49] and

conducted a PCA using the wing measurements normalized by femur length in R [46]. We visualized the

PCA with mapmixture v1.1.4 [29]

Ecological niche modeling

Occurrence Data

We selected occurrences for *P. longipennis* using the Global Biodiversity Information Facility (GBIF),

pulling data from citizen science sources and natural history museum collections (iNaturalist, Odonata

Central, National Museum of Natural History). We selected research-grade occurrences with verified

latitude and longitude coordinates, a photograph of the sighting, observation date, and at least ⅔

agreement on species identification by the community. Furthermore, we filtered occurrences by removing

erroneous localities (natural history museums, middle of the ocean, etc.). We binned our occurrences to 5-

year intervals between the years 1990-2020, which consisted of 6 time intervals, each with differing

numbers of occurrences:

1990-1995: https://doi.org/10.15468/dl.aj6jh6 - 311

1996-2000: https://doi.org/10.15468/dl.anmsxp - 194

2001-2005: https://doi.org/10.15468/dl.5hg78h - 766

2006-2010: https://doi.org/10.15468/dl.bduugx - 2194

2011-2015: https://doi.org/10.15468/dl.uvkeug - 3727

2016-2020: https://doi.org/10.15468/dl.xbtu2p - 29,845

Environmental Data

All modeling and environmental raster processing was conducted using the statistical program R v. 4.1.2

with the packages ENMeval v2.0 [50,51], WALLACE v.1.9 [52], and dismo [53]. We acquired monthly

minimum and maximum temperature and precipitation rasters of the United States from 2000-2020 using

the Worldclim "Historical Monthly Weather Dataset" at 2.5 arcminute resolution. We averaged the

monthly rasters for the 20-year interval and extrapolated the 19 bioclimatic variables using the final

averages of maximum and minimum temperature and precipitation [54].

Model Building

We thinned our occurrence dataset to 10 km to match the resolution of our raster dataset, with the aim of

filtering to 1-2 occurrences per city, and to reduce spatial autocorrelation, artificial clustering, and to even

sample sizes across our time intervals. We used the spThin package [55]. We defined a study extent

consisting of the contiguous United States. Within our extent, we randomly sampled 50,000 background

points for modeling and extracted their environmental values. We used these values to calculate

correlations between variables using the 'vifcor' and 'vifstep' functions in the 'usdm' package [56] and

filtered out variables with correlation coefficients higher than 0.7 and a Variance Inflation Factor (VIF)

threshold of 10.

MaxEnt (Maximum entropy algorithm) was chosen not only for its ability to incorporate spatial

biases into the modelization process but is recommended for presence-only occurrence datasets [57–60].

MaxEnt compares the environmental conditions at locations of occurrence records with randomly selected

points from a background extent to create a machine-learning model of habitat suitability. Species

occurrences were spatially partitioned using the 'checkerboard2' method [51], whereby occurrence data is

geographically split into equal partitions by overlaying two sets of checkerboards, using 25% of the

localities to calibrate the models ('training' data) and 75% to evaluate the models' predictive accuracy

('evaluation' data).

Feature classes were incorporated into the model, which determine the shape of available

modeled relationships in environmental space [59]. To objectively compare modeled suitability of *P.*

longipennis, and due to the high number of occurrence records (making computation with multiple feature

classes unfeasible) we restricted our model relationship to linear feature classes and a regularization of 1.

Both feature classes and regularization multipliers can strongly influence MaxEnt model outputs [61,62].

Optimal models were assessed using threshold independent (AUC curve) and threshold

dependent metrics (10% omission). AUC (Area Under the Curve) is the measure of the ability of the

MaxEnt model generated to discriminate between false positives and negatives. AUC ranges from 0 to 1,

with 1 being considered a "perfect model", 0.5 as random, and < 0.5 considered worse than random

[63,64]. Optimal models were based on the averaged validation AUC, which is the mean of the AUC of

the testing data for all partitions (AUC_{test}). Omission rate is another method of evaluation whereby a

threshold is applied to a continuous model prediction. Application of the threshold makes the model

binary, in which points are categorized as either within or outside the prediction. We checked model

performance using the Akaike criterion for small sample sizes (AICc) [62], as well as the number of

nonzero coefficients (parameters) from each model.

For optimal models within each period, we documented the importance of variables and plotted

response curves. Maxent calculates permutation importance by changing the values of each

environmental variable at random, then calculating the difference using the AUC from the 'training data'.

The values of each environmental variable are randomly permuted within the training presence and

background data, the resultant drop in training AUC is calculated, then normalized to percentages. The

larger the percentage, the more importance that variable possesses in suitability. Response curves show

how each environmental variable individually affects the maxent prediction in terms of increasing or

decreasing suitability. Behavior of response curve is dictated by model complexity (feature classes and

regularization multipliers) [58].

Maxent raw predictions were transformed to a scale of 0–1 to approximate probability of

occurrence using the 'cloglog' transformation [58] and displayed on a QGIS *v.3.34*.

RESULTS

Sequencing and depth

With the exception of one individual from Tennessee with an average depth of 0.45, which was excluded

from downstream analyses, all sequenced samples had an average sequencing depth between 2.00 and

3.40.

Figure 2: Population Structure of P. longipennis. Summary of population structure analyses calculated
with whole genome resequencing data. Admixture analyses calculated using (A) genotype likelihoods,
and (B) called SNPs. The best model was chosen by cross validation and is shown here. PCA plots of
sampled P. longipennis calculated with (C) genotype likelihoods, and (D) called SNPs. (E) COI haplotype
network of P. longipennis from publicly available COI sequences, including the COI sequence from the
reference genome. (F) A male P. longipennis from Boise, Idaho mate guarding a female (credit: Kim
Chmura).

Table one: F_{ST} Estimates				
Weighted and unweighted global F_{ST} estimates between sampled populations of P. longipennis using SNP calls and genotype likelihoods from re-sequenced populations				
	Unweighted F_{ST} (SNPs)	Weighted F_{ST} (SNPs)	Unweighted F_{ST} (Likelihoods)	Weighted F_{ST} (Likelihoods)
Idaho and New Jersey	0.13	0.19	0.13	0.24
Idaho and Tennessee	0.12	0.18	0.07	0.24
New Jersey and Tennessee	<0.01	<0.01	0.06	0.07
COI F_{ST} measures between individuals East and West of the Rocky Mountains. (Considered with and without Tennessee as a shorter sequence fragment was available for this population).				
Population comparison				F_{ST}
East of Rocky Mountains (Excluding Tennessee) and West of Rocky Mountains				0.34
East of Rocky Mountains (Including Tennessee) and West of Rocky Mountains				0.38

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Figure 3: Molecular functions of genes from highly segregating (Fst >.5) and highly unstructured (Fst < .01) regions of the genome, calculated pairwise between the Idaho and New Jersey and Idaho and Tennessee populations with a sliding window of 10kb, with a step of 1kb.

Population structure

The best-fitting admixture analyses identified two ancestral populations from genotype likelihoods, and four ancestral populations from SNPs (Fig 2A-B). In the SNPs admixture analysis, ancestral populations one and three were unique to Tennessee and New Jersey (Fig. 2B). All individuals from Idaho had ancestry from ancestral populations two and four, which were also found at low levels in Tennessee (Fig. 2B). In the likelihood admixture analysis all individuals from Idaho had 100% of their ancestry assigned ancestral population two, one individual from Tennessee split its ancestry between ancestral populations

one and two, and the remainder of the individuals had 100% of their ancestry assigned to ancestral

population one (Fig. 2A).

The PCA, admixture, and weighted and unweighted Fst analyses, using both the called SNPs and

genotype likelihoods, showed little divergence between New Jersey and Tennessee ($F_{ST} < .01$, Fig. 2A-D,

Table 1). Between these two populations, ~88% of genes were in regions of high genomic conservation

($F_{st} < 0.01$), and $< 0.01\%$ of genes were in highly structured regions ($F_{st} > 0.5$). None of the 31

structured genes ($F_{ST} > .50$) between New Jersey and Tennessee had a functional annotation, and the

large number of unstructured genes between these populations was not tractable for visualization with

REVIGO. The Idaho population showed comparatively average high divergence with both East Coast

populations ($F_{ST} > 0.18$, Fig. 2A-D, Table 1). The COI haplotype analyses also showed an East/West

divide, with largely shared haplotypes between East Coast populations, and differing, but closely related

haplotypes in the West Coast populations (Fig. 2E), and a high F_{ST} (> 0.30) compared to the genome wide

averages (Table 1). In comparing Idaho and Tennessee, we found that ~ 31% of genes were in regions of

high genomic conservation between the two populations ($F_{st} < 0.01$), and ~ 9% of genes were in regions

of high divergence ($F_{st} > 0.5$). 236 biological processes were summarized from the functional annotation

of genes in highly unstructured regions between Idaho and Tennessee, including 86 cellular components

and 307 molecular functions (Fig. 3). Genes from highly diverged regions of the genome were

summarized into 76 biological processes, 31 cellular components, and 100 molecular functions (Fig. 3).

In the pairwise comparison between Idaho and New Jersey, ~29% of genes were highly unstructured, ~

9% of genes were greatly diverged. The unstructured genes consisted of 240 biological processes, 77

cellular components, and 271 molecular functions (Fig. 3). The structured genes consisted of 57

biological processes, 24 cellular components, and 82 molecular functions (Fig. 3).

Figure 4: Historical Demography of P. longipennis. (A) Demographic model of the three P. longipennis
populations as estimated by GADMA2 [35] demonstrating little contact between populations on either
side of the Rocky Mountains following separation until bursts of gene flow occurred following the last
glacial maxima. (B) Empirical and simulated pairwise site frequency spectra from the analysis.

The demographic model estimated a split between Idaho and the two East Coast populations

~750ka with Tennessee and New Jersey splitting ~50 Ka (Fig. 4A). Since their divergence, Tennessee is

estimated to have been the recipient of frequent gene flow from New Jersey, and Idaho was a recipient of

gene flow from the ancestral East coast population ~400,000 years ago, and has been a recipient of gene

flow from both populations more recently, in the past ~50k years (Fig. 4A). The model identified one

small burst of gene flow from Idaho to New Jersey (Fig. 4A). In this analysis, the simulated site

frequency spectra (SFS) reasonably approximated the observed (Fig. 4B).

 Morphological analysis

Table three: Significance of Kruskal Wallace Test for each measurement when normalized by either femur or thorax length.		
Measurement	p-value (normalized by femur length)	p-value (normalized by thorax length)
Hindwing length	<.01	<.01[†]
Hindwing depth	.12	<.01[†]
Hindwing pterostigma	.27	.02[†]
Hindwing distal half	<.01	<.01[†]
Hindwing proximal half	.01	.02[†]
Hindwing aspect ratio	.02	.04
Forewing length	<.01	<.01[†]
Forewing depth	.22	.02[†]
Forewing pterostigma	.33	.05
Forewing distal half	<.01	<.01[†]
Forewing proximal half	<.01	<.01[†]
Forewing aspect ratio	.04	.36
Thorax length	.29[†]	-
Femur length	-	.29[†]
[†]Significantly differs ($p < .05$) between males and females (calculated via a Wilcoxon rank-sum test)		

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We were not able to successfully capture wing area, as different measurements of the same individual

varied considerably due to a flaw in our semi-automated workflow that made it sensitive to scale bar

variation (Supplementary Table 1). We found that the other wing measurements differed between the

sexes when normalized by thorax length, but not when normalized by femur length (Table 2). When

normalized by femur length the hindwing length, hindwing distal half, hindwing proximal half, hindwing

aspect ratio, forewing length, forewing distal half, forewing proximal half and forewing aspect ratio all

significantly differed between populations. The distribution of these measurements in the Idaho

population skewed smaller than the other sites (Fig. 5), and Idaho was distal to the other populations

across PC1 (loadings: hindwing length: 0.37; hindwind distal half: 0.37; forewing length: 0.37; hindwing

depth: 0.36; forewing proximal half: .36), but not PC2 (loadings: hindwing depth: 0.82; forewing depth:

0.39; hindwing pterostigma: 0.36; forewing proximal half: 0.16, Fig. 5).

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379   *Figure 5: Wing measurements of P. longipennis. Violin plots of significantly differing wing*
380   *measurements, when normalized by femur length, and PCA plot generated from wing measurements*
381   *normalized by femur length.*
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Ecological niche models

Optimal models exhibited robust results in 10-percentile omission rate but exhibited subpar

results in mean validation AUC. Our mean validation AUC was low across time periods (0.63-

0.7), while 10-percentile omission rate was low (0.08-0.15) relative to its expected value of 0.1.

Furthermore, models possessed a stark increase in AICc as the number of occurrences increased

(AICc: 26660.67 for year 2016-2020).

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Table Four: Sample size of occurrence localities before and after spatial thinning and maxent ENM settings for *P. longipennis* for the 5-yr intervals from 1990-2020. Settings shown are feature classes (features). Statistics shown are mean validation AUC (AUC_{val}), mean omission rates for 10 percentile training values (OR10), Akaike Information Criterion (AICc), and delta Akaike Information criterion (delta.AICc), and number of non-zero coefficients (ncoef).

Year	Raw Occurrences	Thinned Occurrences	Features	Regularization	AUC_{val}	AICc	delta.AICc	OR10	ncoef
1990-1995	311	147	L	1	0.70	2497.03	0	0.15	6
1996-2000	194	93	L	1	0.63	1737.37	0	0.08	6
2001-2005	766	412	L	1	0.69	6196.174	0	0.11	6
2006-2010	2194	1061	L	1	0.67	11633.92	0	0.11	5
2011-2015	3727	1935	L	1	0.67	16417.95	0	0.11	6
2016-2020	29845	8819	L	1	0.66	26660.67	0	0.10	6

393   Mean diurnal range (bio02) possessed the highest variable importance for the years 1990-1995, 2011-

394   2015, and 2016-2020 (range: ~35-55%), maximum temperature of the warmest month (bio05) for the

395   years 2006-2010 (~35%), mean temperature of the warmest quarter (bio10) for the years 2001-2005

396   (~50%), and precipitation of the driest month for the years 1996-2000 (~34%). Mean diurnal range

397   (bio02), temperature seasonality (bio04), maximum temperature of the warmest month (bio05), mean

temperature of the warmest quarter (bio10), and precipitation of the driest month (bio14) possessed the

second and third highest permutation importance throughout our models.

Suitability Predictions

For the years 1990-1995, suitability was highest throughout the eastern United States, tapering off at the

western half of Texas, with a pocket of suitability occurring in the Pacific Northwest (Fig. 6). For the

years 1996-2000, suitability was highest along the southern and western coasts of the US (Fig. 6). For the

years 2001-2005, suitability moves back to the entire eastern US, excepting Maine (Fig. 6). Throughout

the years 2006-2020, suitability remains consistent, with the highest suitability along the southern and

eastern coasts of the US, as well as the eastern US (Fig. 6).

Fig. 6: *Maxent predictions for P. longipennis from the years 1990-2020, subdivided into 5 year time intervals. Predictions were derived using linear feature classes and 1x regularization. Predictions were transformed using the 'cloglog' function and converted to a range of 0-1 to approximate a probability of occurrence (Phillips et al., 2017). Brighter colors (yellow, green, blue) indicate areas of higher suitability (higher probability of occurrence), while darker colors (violet) indicate areas of lower suitability (lower probability of occurrence). Orange points are our thinned dataset for P. longipennis for each time slice.*

Discussion

Dispersal capabilities of Pachydiplax

Our findings support the hypothesis that *P. longipennis* has a high dispersal capability, but it is not

completely panmictic, as some migratory dragonflies are thought to be [65]. The two sampled East Coast

populations are genetically quite similar (Fig. 2A-D, table 1), even more so in coding regions, and the

demographic model shows some structure between Tennessee and New Jersey, but with a high degree of

gene flow. Genetically, the population from Idaho does not appear to be a part of a migratory complex. It

is genetically distinct from New Jersey and Tennessee (Fig. 2A-D, Table 1), and has primarily served as a

sink population (Fig. 4A), all providing evidence that it is not dispersing towards eastern populations.

COI analyses also provided evidence of higher dispersal in eastern populations, which largely shared one

COI haplotype, compared to western populations which were more disjointed (Fig. 2E).

Morphologically, the comparatively longer wings of the eastern populations may be related to

selection for lower energy expenditure during gliding style flight [43], a possible adaptation for long

distance travel. The molecular functions summarized from genes in highly structured regions between

Idaho and Tennessee and Idaho and New Jersey provide evidence that Tennessee and New Jersey are

equipped for longer-duration flight than *P. longipennis* from Idaho. Highly structured genes involved in

the metabolism of carbohydrates and lipids, both of which are under heavy selection in migratory insects

[66–68], and the selective pressures of high dispersal could be driven by positive selection associated with

differing dispersal capabilities in *P. longipennis*, explaining the patterns observed in our F_{ST} scans (Fig.

3).

Much work remains to fully understand the dispersal patterns of *P. longipennis*. Broader

sampling is needed to determine how *P. longipennis* dispersed to the Caribbean Islands and Bermuda, and

the speed at which it should be expected to colonize newly available habitats. As a common pond dweller,

it is possible that humans are transporting eggs and larvae with pond substrate, which could impact our

estimates of gene flow and divergence times and facilitate dispersal to new habitats, but it cannot be ruled

out that *P. longipennis* arrived at these islands through a natural dispersal process not directly involving

human activity. Further genome wide resequencing data is needed to confirm the broad East/West

population structure identified with COI, and to determine if any populations transverse the relatively

inhospitable Rocky Mountains (Fig. 6).

Molecular and morphological barriers between populations

Our molecular and morphological analyses revealed potential reproductive barriers between Idaho and the

sampled eastern populations. One of the biological functions summarized from genes in genomic

windows of high structure between Idaho and Tennessee was "circadian regulation of gene expression"

(Fig. 3). Such genes have been implicated in temporal reproductive isolation in insects such as

Drosophila [69], melon flies [70], Lepidoptera [71,72] and aphids [73], but no such work has been done

in Odonata. Also of note are genes involved in "immune response" and "innate immune response" which

segregate between Idaho and East Coast populations. In insects, immune response can have a strong

influence on sexual reproduction and behavior [74,75].

Morphologically, it has been demonstrated that mate guarding, a behavior undertaken by *P.*

longipennis [6] where a male guards the female until she deposits eggs (Fig. 2F), has a heavy effect upon

front wings; indeed, forewings may be broader in species that utilize tandem mate guarding where males

and females fly in copula to oviposit [76]. As female blue dashers on the East Coast have evolved with

the longer-winged males (Fig. 5), it is unclear if the shorter- winged males from Idaho could carry out this

behavior with East Coast females.

The implications of these findings on the species delimitation in the genus *Pachydiplax* should be

further explored with more robust population and phenomic sampling.

P. longipennis as an urban "extraordinaire"

Difficulties in modeling an abundant species

Our work confirms the hypothesis that *P. longipennis* is very well adapted to life in urban habitats. With

the increased popularity of citizen science datasets starting approximately in 2010, the number of

sightings of common species, like *P. longipennis*, increased dramatically. Indeed, as of December 2024,

in its range, *P. longipennis* was the most widely observed aquatic insect on iNaturalist, the fifth most

observed insect, and the 40th most observed species on iNaturalist. Although such increases in data

provide the opportunity for highly robust niche models, a consequence is the overfitting of model

performance due to the widespread abundance and density of *P. longipennis*. Predicted ENMs of *P.*

longipennis revealed low AUC values across time slices. AUC values are a continuous metric, which

provide discriminative ability in discerning between false positives and false negatives. Since *P.*

longipennis is found throughout the continental United States, our models were unable to discern

gradients of high and low suitability. This pattern is also observed in our AICc values. For the years

2016-2020, our thinned dataset consisted of 8,000+ occurrences. As a result, our AICc value was

exceptionally high compared to the other models. This same observation can be seen on islands where

occurrences are high for singular species [77]. Interestingly, our 10-percentile omission rate metrics were

highly robust as most values were close to the expected value of 0.1. The 10-percentile omission rate

metric excludes the lowest 10 percent of raw occurrence values, thus creating a threshold within

suitability. Our low omission and AUC values suggest minimal outliers in our dataset, suggesting that our

predictions are not overly generalist.

Adaptations for success in human altered environments

Our models estimate that extremes in temperature, precipitation, and temperature fluctuation are the main

drivers of the response of *P. longipennis* to their distribution. Due to their widespread abundance

throughout the contiguous United States, it seems this species has an exceptionally high thermal tolerance

and desiccation threshold, which elucidates why they are so widespread in urban environments. In reared

experiments, larval development of *P. longipennis* was unaffected by increases in temperature or resource

level [78]. Furthermore, mesocosm experiments [79] revealed that larval *P. longipennis* are more likely to

engage in intraguild predation with increases in temperature, which may allow them to diversify prey

types in warmer habitats. Urban environments have significantly higher ground temperatures than rural

areas, a consequence of the higher demand of power plants providing energy for in-home cooling units

and impervious surfaces creating urban heat islands [80]. We propose that *P. longipennis* is distributed

widely in urban environments due to their optimal habitats being warmer, a condition that will only

increase as the urban heat sink phenomenon continues to worsen.

Our suitability predictions suggest that the distribution of *P. longipennis* is widening as time

progresses and urbanization increases throughout the United States. However, one caveat of our

distribution sampling is the impact of the increase of documented *P. longipennis* sightings as the result of

increasingly popular community science programs. Although our models may be slightly skewed towards

increases in distribution of *P. longipennis*, our genomic analyses support the biological realism of the

models. One molecular function previously identified in expanding gene families in the genome assembly

of *P. longipennis* was the "Peptidoglycan Catabolic Process" [4]. Peptidoglycan is an integral compound

in most bacterial cell walls [81], and genes associated with peptidoglycan catabolism are thought to be

involved in both immune and desiccation response in insects [82]. It is intriguing that between Idaho and

the East Coast populations of *P. longipennis,* examples of highly structured and highly unstructured genes

can *both* be tied to this molecular process (Fig. 3). Our population genomics analysis provides further

evidence that these gene families play a key role in the dispersal success of *P. longipennis* in polluted

habitats and are a driver of the expanding niche of *P. longipennis*. In addition to enabling *P. longipennis*

to survive amidst the higher desiccation risk found in warmer habitats, the lentic water bodies they inhabit

are expected to carry a high bacterial load. It is possible that certain peptidoglycan variants will be

encountered by all populations of *P. longipennis*, explaining unstructured genes related to these functions.

Each population of *P. longipennis* could also encounter unique variants of peptidoglycan, explaining the

high segregation in some of these genes. Such a dynamic could be extended to an identical phenomenon

observed involving genes which are associated with "immune response" and "innate immune response."

Sampling of *P. longipennis* alongside bacterial communities will be an important step in testing this

hypothesis.

Furthermore, *P. longipennis* likely faces high levels of exposure to free radicals, similar to other

animals in urban environments [83]. Expanded gene families involved in the processing of free radicals

have been hypothesized as an important mechanism enabling P. longipennis to successfully colonize

urban environments [33]. Our finding that genes involved in this process are highly unstructured between

populations (Fig. 3) provides further evidence that these gene families are key to the success of *P.*

longipennis, in addition to its ability to respond to desiccation and a high pathogen load. Future research

with these genes will likely include transcriptome sequencing to understand patterns of regulation in

association with oxidative stress.

Recent niche models of odonates have suggested high-dispersing species will be the ecological

"winners" in a changing global climate [84–86]. It seems that *P. longipennis* is definitively a winner that

will flourish as humans expand. We do not expect the genes involved in the success of *P. longipennis* in

urban environments to be restricted to this species. Further work needs to be done in other such species to

determine if there is convergence in species that inhabit these environments.

Conclusion

Our work presents compelling evidence that *P. longipennis* is a successful habitat generalist. Studies of

lentic habitats in California between the early Twentieth and early Twenty-first Centuries found a shift in

odonate assemblages away from specialists and towards generalists [7]. Among the species showing the

greatest expansion was *P. longipennis*. This study confirms that *P. longipennis* is well equipped to deal

with the stress of urban habitats. East Coast populations have a relatively higher dispersal capability (Fig.

2) , which could allow them to more rapidly colonize newly suitable habitat. While the specific dynamics

between bacterial loads, oxidative stress, and the establishment *P. longipennis* need further study we posit

that the dominance of the highly observable *P. longipennis* at a water body can be considered evidence of

contamination and biodiversity loss at a given habitat.

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Data availability

The "Gadma" parameters file, rMarkdown script used for statistical analysis, images, and phyllip file use

for COI analyses are available on figshare (10.6084/m9.figshare.28020914). The raw sequencing reads

have been uploaded to the sequence read archive (bioproject PRJNA1199395).

Conflicts of interest

The authors declare no conflicts of interest.

References

1. Wagner DL, Grames EM, Forister ML, Berenbaum MR, Stopak D. Insect decline in the Anthropocene: Death by a thousand cuts. *PNAS*. National Academy of Sciences; 2021; doi: 10.1073/pnas.2023989118.
2. Hallmann CA, Sorg M, Jongejans E, Siepel H, Hofland N, Schwan H, et al.. More than 75 percent decline over 27 years in total flying insect biomass in protected areas. *PLOS ONE*. Public Library of Science; 2017; doi: 10.1371/journal.pone.0185809.
3. Catling PM. Climate Warming as an Explanation for the Recent Northward Range Extension of Two Dragonflies, *Pachydiplax longipennis* and *Perithemis tenera*, into the Ottawa Valley, Eastern Ontario. *The Canadian Field-Naturalist*. 2016; doi: 10.22621/cfn.v130i2.1846.
4. Tolman ER, Beatty CD, Frandsen PB, Bush J, Bruchim OR, Driever ES, et al.. Newly Sequenced Genomes Reveal Patterns of Gene Family Expansion in select Dragonflies (Odonata: Anisoptera). bioRxiv;
5. Corbet PS, May ML. Fliers and perchers among Odonata: dichotomy or multidimensional continuum? A provisional reappraisal. *International Journal of Odonatology*. 2008; doi: 10.1080/13887890.2008.9748320.
6. Paulson D. Dragonflies and Damselflies of the West. Princeton: Princeton University Press;
7. Ball-Damerow JE, M'Gonigle LK, Resh VH. Changes in occurrence, richness, and biological traits of dragonflies and damselflies (Odonata) in California and Nevada over the past century. *Biodivers Conserv*. 2014; doi: 10.1007/s10531-014-0707-5.
8. Rosser W. Garrison PD, Natalia von Ellenrieder PD, Jerry A. Louton PD. Dragonfly Genera of the New World. Johns Hopkins University Press;
9. Ware J, May M, Kjer K. Phylogeny of the higher Libelluloidea (Anisoptera: Odonata): An exploration of the most speciose superfamily of dragonflies. *Molecular Phylogenetics and Evolution*. 2007; doi: 10.1016/j.ympev.2007.05.027.
10. Pilgrim EM, Von Dohlen CD. Phylogeny of the Sympetrinae (Odonata: Libellulidae): further evidence of the homoplasious nature of wing venation. *Systematic Entomology*. 2008; doi: 10.1111/j.1365-3113.2007.00401.x.
11. Carle F, Kjer K, May M. A molecular phylogeny and classification of Anisoptera (Odonata). *Arthropod Systematics & Phylogeny*. Senckenberg Gesellschaft für Naturforschung; 2015; doi: 10.3897/asp.73.e31805.
12. McCauley SJ. Body size and social dominance influence breeding dispersal in male Pachydiplax longipennis (Odonata). *Ecological Entomology*. 2010; doi: 10.1111/j.1365-2311.2010.01191.x.
13. Penn GH. Seasonal Variation in the Adult Size of Pachydiplax Longipennis (Burmeister) (Odonata, Libellulidae)1. *Annals of the Entomological Society of America*. 1951; doi: 10.1093/aesa/44.2.193.
14. Baird J, May M. Foraging behavior of Pachydiplax longipennis (Odonata: Libellulidae). *Journal of Insect Behavior - J INSECT BEHAV*. 1997; doi: 10.1007/BF02765385.
15. Baird J, May M. Fights at the Dinner Table: Agonistic Behavior in Pachydiplax longipennis (Odonata: Libellulidae) at Feeding Sites. *Journal of Insect Behavior*. 2003; doi: 10.1023/A:1023963717997.
16. Johnson C. A Study of Territoriality and Breeding Behavior in Pachydiplax longipennis Burmeister

(Odonata:Libellulidae). *The Southwestern Naturalist*. Southwestern Association of Naturalists; 1962; doi:
10.2307/3668841.
17. May ML, Baird JM. A Comparison of Foraging Behavior in Two "Percher" Dragonflies, Pachydiplax
longipennis and Erythemis simplicicollis (Odonata: Libellulidae). *Journal of Insect Behavior*. 2002; doi:
10.1023/A:1021171306468.
18. Sherman KJ. The adaptive significance of postcopulatory mate guarding in a dragonfly, Pachydiplax
longipennis. *Animal Behaviour*. Netherlands: Elsevier Science; 1983; doi: 10.1016/S0003-
3472(83)80018-9.
19. Van Buskirk J. Density-dependent population dynamics in larvae of the dragonfly Pachydiplax
longipennis: a field experiment. *Oecologia*. 1987; doi: 10.1007/BF00379272.
20. Pfeiffer B. A Wild Swarm of Dragonflies. Chasing Nature.
21. Russell RW, May ML, Soltesz KL, Fitzpatrick JW. Massive Swarm Migrations of Dragonflies
(Odonata) in Eastern North America. *The American Midland Naturalist*.
22. St. John K. Swarm of Dragonflies on the Move. Outside My Window.
23. Migratory Dragonfly Partnership. Field Guide to Migratory Dragonflies. The Hawk Migration
Association of North America;
24. Li H, Durbin R. Fast and accurate short read alignment with Burrows–Wheeler transform.
Bioinformatics. 2009; doi: 10.1093/bioinformatics/btp324.
25. . broadinstitute/picard. Broad Institute;
26. Danecek P, Bonfield JK, Liddle J, Marshall J, Ohan V, Pollard MO, et al.. Twelve years of SAMtools
and BCFtools. *Gigascience*. 2021; doi: 10.1093/gigascience/giab008.
27. Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MAR, Bender D, et al.. PLINK: A Tool Set
for Whole-Genome Association and Population-Based Linkage Analyses. *Am J Hum Genet*. 81:559–
752007;
28. Skotte L, Korneliussen TS, Albrechtsen A. Estimating individual admixture proportions from next
generation sequencing data. *Genetics*. 2013; doi: 10.1534/genetics.113.154138.
29. Jenkins TL. mapmixture: An R package and web app for spatial visualisation of admixture and
population structure. *Mol Ecol Resour*. 2024; doi: 10.1111/1755-0998.13943.
30. Korneliussen TS, Albrechtsen A, Nielsen R. ANGSD: Analysis of Next Generation Sequencing Data.
BMC Bioinformatics. 2014; doi: 10.1186/s12859-014-0356-4.
31. Fox EA, Wright AE, Fumagalli M, Vieira FG. ngsLD: evaluating linkage disequilibrium using
genotype likelihoods. *Bioinformatics*. 2019; doi: 10.1093/bioinformatics/btz200.
32. Meisner J, Albrechtsen A. Inferring Population Structure and Admixture Proportions in Low-Depth
NGS Data. *Genetics*. 2018; doi: 10.1534/genetics.118.301336.
33. Tolman ER, Beatty CD, Frandsen PB, Bush RJ, Burchim OR, Driever ES, et al.. Newly Sequenced
Genomes Reveal Patterns of Gene Family Expansion in select Dragonflies (Odonata: Anisoptera).
bioRxiv;
34. Supek F, Bošnjak M, Škunca N, Šmuc T. REVIGO Summarizes and Visualizes Long Lists of Gene
Ontology Terms. Gibas C, editor. *PLoS ONE*. 2011; doi: 10.1371/journal.pone.0021800.
35. Noskova E, Abramov N, Iliutkin S, Sidorin A, Dobrynin P, Ulyantsev V. GADMA2: more efficient
and flexible demographic inference from genetic data. bioRxiv;
36. Jouganous J, Long W, Ragsdale AP, Gravel S. Inferring the Joint Demographic History of Multiple
Populations: Beyond the Diffusion Approximation. *Genetics*. 2017; doi: 10.1534/genetics.117.200493.
37. Tolman ER, Bruchim OR, Driever ES, Jordan D, Kohli MK, Montague L, et al.. Changes in effective
population size of Odonata in response to climate change revealed through genomics. *IJO*. 2023; doi:
10.48156/1388.2023.1917241.
38. Ratnasingham S, Hebert PDN. bold: The Barcode of Life Data System
(http://www.barcodinglife.org). *Molecular Ecology Notes*. 2007; doi: 10.1111/j.1471-8286.2007.01678.x.
39. Kohli MK, Schneider T, Müller O, Ware JL. Counting the spots: a molecular and morphological
phylogeny of the spotted darner Boyeria (Odonata: Anisoptera: Aeshnidae) with an emphasis on
European taxa. *Systematic Entomology*. 2014; doi: 10.1111/syen.12049.

40. Madeira F, Madhusoodanan N, Lee J, Eusebi A, Niewielska A, Tivey ARN, et al.. The EMBL-EBI
Job Dispatcher sequence analysis tools framework in 2024. *Nucleic Acids Res*. 2024; doi:
10.1093/nar/gkae241.
41. Maddison DR, Wheeler TJ, Maddison WP. Align: a Mesquite package for aligning sequence data.
42. Leigh JW, Bryant D. popart: full-feature software for haplotype network construction. *Methods in*
Ecology and Evolution. 2015; doi: 10.1111/2041-210X.12410.
43. Suárez-Tovar CM, Sarmiento CE. Beyond the wing planform: morphological differentiation between
migratory and nonmigratory dragonfly species. *J Evol Biol*. 2016; doi: 10.1111/jeb.12830.
44. Schneider CA, Rasband WS, Eliceiri KW. NIH Image to ImageJ: 25 years of image analysis. *Nat*
Methods. Nature Publishing Group; 2012; doi: 10.1038/nmeth.2089.
45. Idec J, Bybee S, Ware J, Abbott J, Ferreira RG, Suvorov A, et al.. Interactions between sexual
signaling and wing size drive ecology and evolution of wing colors in Odonata. *Sci Rep*. Nature
Publishing Group; 2024; doi: 10.1038/s41598-024-73612-4.
46. R Core Team. R: A language and environment for statistical computing. Vienna, Austria: R
Foundation for Statistical Computing,;
47. Kaspari M, Weiser MD. The size–grain hypothesis and interspecific scaling in ants. *Functional*
Ecology. 1999; doi: 10.1046/j.1365-2435.1999.00343.x.
48. Teuscher M, Brändle M, Traxel V, Brandl R. Allometry between leg and body length of insects: lack
of support for the size–grain hypothesis. *Ecological Entomology*. 2009; doi: 10.1111/j.1365-
2311.2009.01124.x.
49. Buuren S van, Groothuis-Oudshoorn K. mice: Multivariate Imputation by Chained Equations in R.
Journal of Statistical Software. 2011; doi: 10.18637/jss.v045.i03.
50. Kass JM, Muscarella R, Galante PJ, Bohl CL, Pinilla-Buitrago GE, Boria RA, et al.. ENMeval 2.0:
Redesigned for customizable and reproducible modeling of species' niches and distributions. *Methods in*
Ecology and Evolution. 2021; doi: 10.1111/2041-210X.13628.
51. Muscarella R, Galante PJ, Soley-Guardia M, Boria RA, Kass JM, Uriarte M, et al.. ENMeval: An R
package for conducting spatially independent evaluations and estimating optimal model complexity for
Maxent ecological niche models. *Methods in Ecology and Evolution*. 2014; doi: 10.1111/2041-
210X.12261.
52. Kass JM, Vilela B, Aiello-Lammens ME, Muscarella R, Merow C, Anderson RP. Wallace: A flexible
platform for reproducible modeling of species niches and distributions built for community expansion.
Methods in Ecology and Evolution. 2018; doi: 10.1111/2041-210X.12945.
53. Hijmans RJ, Phillips S, Leathwick J, Elith J. dismo: Species Distribution Modeling.
54. Fick SE, Hijmans RJ. WorldClim 2: new 1-km spatial resolution climate surfaces for global land
areas. *International Journal of Climatology*. 2017; doi: 10.1002/joc.5086.
55. Aiello-Lammens ME, Boria RA, Radosavljevic A, Vilela B, Anderson RP. spThin: an R package for
spatial thinning of species occurrence records for use in ecological niche models. *Ecography*. 2015; doi:
10.1111/ecog.01132.
56. Naimi B. usdm: Uncertainty Analysis for Species Distribution Models.
57. Myers CE, Stigall AL, Lieberman BS. PaleoENM: applying ecological niche modeling to the fossil
record. *pbio*. The Paleontological Society; 2015; doi: 10.1017/pab.2014.19.
58. Phillips SJ, Anderson RP, Dudík M, Schapire RE, Blair ME. Opening the black box: an open-source
release of Maxent. *Ecography*. 2017; doi: 10.1111/ecog.03049.
59. Phillips SJ, Dudík M. Modeling of species distributions with Maxent: new extensions and a
comprehensive evaluation. *Ecography*. 2008; doi: 10.1111/j.0906-7590.2008.5203.x.
60. Stolar J, Nielsen SE. Accounting for spatially biased sampling effort in presence-only species
distribution modelling. *Diversity and Distributions*. 2015; doi: 10.1111/ddi.12279.
61. Radosavljevic A, Anderson RP. Making better Maxent models of species distributions: complexity,
overfitting and evaluation. *Journal of Biogeography*. 2014; doi: 10.1111/jbi.12227.
62. Warren DL, Seifert SN. Ecological niche modeling in Maxent: the importance of model complexity
and the performance of model selection criteria. *Ecol Appl*. 2011; doi: 10.1890/10-1171.1.

63. Peterson AT, Papeş M, Soberón J. Rethinking receiver operating characteristic analysis applications
in ecological niche modeling. *Ecological Modelling*. 2008; doi: 10.1016/j.ecolmodel.2007.11.008.
64. Peterson AT, Soberón J. Species Distribution Modeling and Ecological Niche Modeling: Getting the
Concepts Right. *NatCon*. 2012; doi: 10.4322/natcon.2012.019.
65. Ware J, Kohli MK, Mendoza CM, Troast D, Jinguji H, Hobson KA, et al.. Evidence for widespread
gene flow and migration in the Globe Skimmer dragonfly Pantala flavescens. *IJO*. 2022; doi:
10.48156/1388.2022.1917166.
66. Jin M, Liu B, Zheng W, Liu C, Liu Z, He Y, et al.. Chromosome-level genome of black cutworm
provides novel insights into polyphagy and seasonal migration in insects. *BMC Biol*. 2023; doi:
10.1186/s12915-022-01504-y.
67. Olsen L, Thum E, Rohner N. Lipid metabolism in adaptation to extreme nutritional challenges.
Developmental Cell. Elsevier; 2021; doi: 10.1016/j.devcel.2021.02.024.
68. Li X, Zhou Y, Wu K. Biological Characteristics and Energy Metabolism of Migrating Insects.
Metabolites. Multidisciplinary Digital Publishing Institute; 2023; doi: 10.3390/metabo13030439.
69. Sandrelli F, Costa R, Kyriacou CP, Rosato E. Comparative analysis of circadian clock genes in
insects. *Insect Molecular Biology*. 2008; doi: 10.1111/j.1365-2583.2008.00832.x.
70. Miyatake T, Matsumoto A, Matsuyama T, Ueda HR, Toyosato T, Tanimura T. The period gene and
allochronic reproductive isolation in Bactrocera cucurbitae. *Proceedings of the Royal Society of London
Series B: Biological Sciences*. The Royal Society; 2002; doi: 10.1098/rspb.2002.2152.
71. Groot AT. Circadian rhythms of sexual activities in moths: a review. *Front Ecol Evol*. Frontiers;
2014; doi: 10.3389/fevo.2014.00043.
72. Schöfl G, Heckel DG, Groot AT. Time-shifted reproductive behaviours among fall armyworm
(Noctuidae: Spodoptera frugiperda) host strains: evidence for differing modes of inheritance. *Journal of
Evolutionary Biology*. 2009; doi: 10.1111/j.1420-9101.2009.01759.x.
73. Abbot P, Withgott JH. PHYLOGENETIC AND MOLECULAR EVIDENCE FOR ALLOCHRONIC
SPECIATION IN GALL-FORMING APHIDS (PEMPHIGUS). *Evolution*. 2004; doi: 10.1111/j.0014-
3820.2004.tb01677.x.
74. Nunes C, Sucena É, Koyama T. Endocrine regulation of immunity in insects. *The FEBS Journal*.
2021; doi: 10.1111/febs.15581.
75. Schwenke RA, Lazzaro BP, Wolfner MF. Reproduction–Immunity Trade-Offs in Insects. *Annual
review of entomology*. 2015; doi: 10.1146/annurev-ento-010715-023924.
76. Johansson F, SÖDERQUIST M, BOKMA F. Insect wing shape evolution: independent effects of
migratory and mate guarding flight on dragonfly wings. *Biological Journal of the Linnean Society*. 2009;
doi: 10.1111/j.1095-8312.2009.01211.x.
77. Goodman AM, Beatty CD, Büsse S, Ubukata H, Miyazaki T, Blair ME, et al.. Paleoecological niche
modeling of Epiophlebia (Epiophlebioptera: Epiophlebiidae) reveals continuous distribution during the
Last Glacial Maximum. *IJO*. 2024; doi: 10.48156/1388.2024.1917262.
78. Chavez MY, Mabry KE, McCauley SJ, Hammond JI. Differential larval responses of two ecologically
similar insects (Odonata) to temperature and resource variation. *International Journal of Odonatology*.
Taylor & Francis; 2015; doi: 10.1080/13887890.2015.1082946.
79. Tituskin JR, Waddell SM, Mabry KE. Species-specific responses to warming alter community
composition. *Ecological Entomology*. 2022; doi: 10.1111/een.13112.
80. Santamouris M. Recent progress on urban overheating and heat island research. Integrated assessment
of the energy, environmental, vulnerability and health impact. Synergies with the global climate change.
Energy and Buildings. 2020; doi: 10.1016/j.enbuild.2019.109482.
81. Vollmer W, Blanot D, De Pedro MA. Peptidoglycan structure and architecture. *FEMS Microbiology
Reviews*. 2008; doi: 10.1111/j.1574-6976.2007.00094.x.
82. Kim H, Kim H-W, Lee JH, Park J, Lee H, Kim S, et al.. Gene family expansions in Antarctic winged
midge as a strategy for adaptation to cold environments. *Sci Rep*. Nature Publishing Group; 2022; doi:
10.1038/s41598-022-23268-9.
83. Salmón P, Watson H, Nord A, Isaksson C. Effects of the Urban Environment on Oxidative Stress in

Early Life: Insights from a Cross-fostering Experiment. *Integr Comp Biol*. 2018; doi: 10.1093/icb/icy099.

84. Abbott JC, Bota-Sierra CA, Guralnick R, Kalkman V, González-Soriano E, Novelo-Gutiérrez R, et

al.. Diversity of Nearctic Dragonflies and Damselflies (Odonata). *Diversity*. Multidisciplinary Digital

Publishing Institute; 2022; doi: 10.3390/d14070575.

85. Bush A, Hermoso V, Linke S, Nipperess D, Turak E, Hughes L. Freshwater conservation planning

under climate change: demonstrating proactive approaches for Australian Odonata. *Journal of Applied*

Ecology. 2014; doi: 10.1111/1365-2664.12295.

86. Kalkman VJ, Boudot J-P, Futahashi R, Abbott JC, Bota-Sierra CA, Guralnick R, et al.. Diversity of

Palaearctic Dragonflies and Damselflies (Odonata). *Diversity*. Multidisciplinary Digital Publishing

Institute; 2022; doi: 10.3390/d14110966.

1) Hindwing length
2) Hindwing depth
3) Hindwing distal half
4) Hindwing proximal half
5) Hindwing pterostigma

6) Forewing length
7) Forewing depth
8) Forewing distal half
9) Forewing proximal half
10) Forewing pterostigma

11) Thorax length
12) Hind femur length

Molecular functions of highly conserved and segregating genes

Idaho vs Tennessee

Idaho vs New Jersey

Segregated (Fst > .5)

Conserved (Fst <.01)

A

Demographic model of New Jersey, Tennessee, and Idaho populations

B

Observed vs. simulated site frequency spectra