Mechanisms of host exploitation by a microsporidian parasite

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Abstract

Parasites exploit their hosts to enhance their growth and reproduction, yet the mechanisms underlying host manipulation remain understudied for many taxa. The microsporidian *Vavraia culicis*, a potential biological control agent for mosquitoes, serves as an excellent model to explore such mechanisms. In this study, we investigate how infection by *V. culicis* lines that vary in virulence alters resource dynamics within the mosquito host *Anopheles gambiae*. Using metallomics and quantification of protein, carbohydrate, and lipid content, we show that infection alters host resource concentrations in ways that depend on parasite virulence. More virulent parasites led to increased protein levels and greater energy demands, evidenced by higher carbohydrate reserves. Additionally, infection with *V. culicis* impacted host metal content, particularly zinc and manganese, used by *V. culicis* independently of its evolutionary background. Iron availability, a key nutrient for parasite growth, enhanced spore production, with selected parasite lines better able to exploit host iron than unselected. These findings provide insight into the mechanisms by which *V. culicis* manipulates host resources, shedding light on the role of host exploitation in parasite virulence and the potential use of microsporidia as biological control agents in vector biology.

Keywords: microsporidia, virulence, iron, *Anopheles gambiae*, resource allocation, metallomics

1. Introduction

The infection of hosts and their consequent exploitation by parasites is a ubiquitous feature of life [1]. The negative impact a parasite has on its host, particularly on host survival, is commonly referred to as virulence [2]. Virulence arises from the interplay between host and parasite traits [3]. Hosts can influence the cost of infection by limiting parasite load or mitigating infection-induced damage - mechanisms known as resistance and tolerance, respectively [4–6]. Parasites, in turn, may exploit host resources to fuel their own growth (host exploitation or proliferation), or invest in toxins and other virulence factors that cause additional harm independent of parasite load (per-parasite pathogenicity or benevolence)[3,7,8].

Since parasites typically sequester resources from their hosts, many hosts have evolved strategies to restrict resource acquisition by parasites through both metabolic and behavioural changes [9]. The mechanisms limiting exploitation have been heavily studied in recent years and have given rise to nutritional immunity [10–13], where the main aim is to enhance host survival by manipulating the parasite's ability to exploit the host. The specific resources targeted by parasites vary depending on the stage of infection (time) and the host tissues involved (space). However, most parasites universally require a ubiquitous set of resources. These range from dietary carbohydrates to essential minerals, which are often hijacked by dedicated parasite transporters. Among these, iron is perhaps the most intensively studied [14–17]. Its critical role in microbial metabolism and structural function is underscored by the widespread evolution of siderophores - iron-chelating molecules used by microbes to extract iron from hosts [18–20]. Iron also acts as a developmental cue in some parasites, such as *Plasmodium* [21–24]. For instance, *Plasmodium falciparum* requires high iron concentrations for gametocyte fusion and successful development within the mosquito vector.

Despite increasing insight into parasite resource demands and sequestration strategies, many gaps remain, especially for obligate intracellular parasites such as microsporidia. Microsporidia are single-celled fungi that infect a wide range of hosts, from humans to dipteran insects [25–30]. Despite their compact genomes [31–33], they exhibit broad ecological diversity, colonising both aquatic and terrestrial environments [34]. Although studied for over 150 years [27], microsporidia have recently gained renewed attention due to their ability to suppress *Plasmodium* infection in *Anopheles* mosquitoes during co-infection [35–39]. Two species, *Vavraia culicis* and *Microsporidia MB*, have emerged as potential malaria control agents [35,40,41], supported by experimental [39,42] and field studies [37], respectively. However, the mechanisms by which these parasites exploit their hosts remain poorly understood. Considering *V. culicis* seems to elicit a limited host response, regardless of the vector host [43,44], here we turned focus to resource competition as a potential method of *Plasmodium*-inhibition. To date, only one study has partially characterised *V. culicis* exploitation of *Anopheles* larvae [45], and another has described limited effects on adult mosquito protein, carbohydrate and lipid content [46].

Understanding how microsporidia exploit their hosts is not only of interest for host-parasite interactions
but also for vector biology, which entails the development of vector control strategies. In light of the
long developmental time *Plasmodium* requires within its vector host, any host capable of transmitting
Plasmodium will inherently favour parasites that also persist longer. Long-lasting parasites, such as
some *V. culicis*, may thus evolve enhanced resource sequestration abilities, potentially outcompeting or
excluding co-infecting parasites such as *Plasmodium spp*. Indeed, a recent study by Silva and Koella
(2024) subjected *V. culicis* to artificial selection for either: i) early transmission and less time within
the host, hereby referred to as *Early* treatment (i.e., parasites that killed the host under 7 days); or ii)
late transmission and more time within the host, hereafter *Late* treatment (i.e., parasites that killed the
host after 20 days) **(Fig. 1)** [44,47,48]. The authors found that lines selected for late transmission, and
consequently persistence within the host, were more virulent, developed faster, and produced higher
spore loads throughout their infective cycle (and the mosquito's life) than those in *Early* or the
unselected stock population (referred to here as *Stock* treatment). However, this came at a cost: *Late*
parasites were less viable outside the host, highlighting a trade-off between adaptation within and
among hosts, in the outside environment [48].

In this study, we took advantage of these selected lines of *V. culicis* to: a) characterise *A. gambiae*
exploitation by *V. culicis*; b) understand which mechanisms can be enhanced upon selection for early
or late transmission (or less and more time within the host, respectively); and c) how changes in host
exploitation relate to the different parasite lines' virulence. To address these questions, we: i) quantified
protein, carbohydrate, and lipid content; ii) performed metallomics; and iii) manipulated iron
availability - known to influence *V. culicis* growth - by supplementing or depleting circulating iron in
the adult mosquito. All measurements were taken on days 5, 10, and 15 of adulthood, following larval-
stage infection with the selected (i.e., *Early* and *Late*) and unselected (i.e., *Stock*) *V. culicis* treatments
(Table 1). These time points correspond to three different phases of *V. culicis* growth: beginning,
highest growth, and slowdown of the growth. We believe our findings will offer important insights into
microsporidian biology, parasite evolution, and disease control, as the molecular machinery studied
here is likely conserved or used by other parasites.

Figure 1. Illustration of the artificial selection experimental setup. An ancestral population of *V. culicis* was used to infect a Kisumu colony of the mosquito host *A. gambiae*. Two selection treatments were generated: i) *Early*, encompassing the parasites that were retrieved from the first third of the mosquitoes that died in the population; and ii) Late, including the parasites that were collected from the last third of the mosquitoes to die. The parasite went through seven serial passages before conducting the experiments in this study. Each selection treatment was replicated five times. As a baseline reference, the unselected parasite kept in a similar rearing parallel to the selection lines was also included in this study (i.e., *Stock*). To facilitate statistical comparisons, the *Stock* was also replicated five times. Further details can be found in Section 2.1 of the Materials and Methods and in the original study describing the selection lines, Silva and Koella (2024), bioRxiv.

Table 1. Experiments conducted and respective replication. Experiments performed in this study (Figures 2 to 4), their treatments, and the number of replicates. The factor(s) of interest are the level at which conclusions are drawn. The scale at which the factor is applied represents the level at which treatments or factors are implemented, whereas the number of replicates at the appropriate scale shows the count of independent replicates for each treatment or factor level. Parasite treatment includes selected treatments, i.e., *Early* and *Late*, and the unselected *Stock*. Time consists of three time-points during mosquito adulthood, 5-, 10-, and 15-day post-adult emergence. Diet encompasses a standard sugar diet, supplemented with iron or chelated with BPS. The replication statement was adapted from

the guidelines provided by the journal Functional Ecology (Wiley-Blackwell and British Ecological Society).

Dependent variable(s)	Factor(s) of interest	Scale at which the factor is applied	Number of replicates at the appropriate scale
Figure 2. Protein, carbohydrate and lipid content throughout infection.			
Protein, carbohydrate and lipid content	Parasite treatment and time	Individual mosquito	10 mosquitoes per replicate (5) per time-point (3)
Figure 3. Metallomic profiles throughout infection.			
Fe, Zn, Cu, Ca and Mn content	Parasite treatment and time	Pool of 10 mosquitoes	1 pool per replicate (5) per time-point (3)
Figure 4. Effect of iron-supplementation and -chelation on parasite growth.			
Spore load	Parasite treatment and diet	Individual mosquito	10 mosquitoes per replicate (5) per time-point (3) per diet (3)

2. Materials and Methods

(a) Experimental model and mosquito rearing

We used the Kisumu strain of the mosquito host *Anopheles gambiae* (s.s.) and differently selected and unselected lines of the microsporidian parasite *Vavraia culicis floridensis* **(Fig. 1)**. To generate the selected lines, we used an ancestral population of *V. culicis* to infect naive *A. gambiae* mosquitoes. From the initial generation (F0), parasites were collected from the first one-third of mosquitoes to die (forming the *Early* treatment) and from the final one-third to die (forming the *Late* treatment). From F1 onwards, these parasite lines were maintained under the same selection protocol - *Early* and *Late* - but evolving independently. This selection process was repeated across seven generations for each of five replicate lines per parasite treatment, allowing only the parasite to evolve, not the host [47]. After seven generations, the selected parasite lines and an unselected reference line (*Stock*), maintained in parallel to the artificial selection experiment, were used in the described experiments. Further details on the selection experiment and its maintenance are described in Silva and Koella (2024). Experiments and rearing were conducted at standard laboratory conditions, 26 ± 1ºC, $70 \pm 5\%$ relative humidity, and 12 h light/dark.

(b) Experimental setup

All experiments were performed similarly **(Table 1)**. In each experiment, freshly hatched *A. gambiae*

larvae were grown in groups of 50 per Petri dish (120 mm diameter x 17 mm) containing 80 ml of

distilled water. Five petri dishes were prepared for each parasite treatment and replicate. Larvae in these

dishes were fed daily with Tetramin Baby fish food according to their age: 0.04, 0.06, 0.08, 0.16, 0.32

and 0.6 mg/larva for 0, 1, 2, 3, 4 and 5 day-old or older larvae, respectively. Two-day-old larvae were

exposed to 10,000 spores per larva of their respective *V. culicis* treatment. Pupae were then transferred

to cages (21 x 21 x 21 cm) and left to emerge as adults. Each cage included individuals from all petri

dishes within a given parasite treatment and replicate. In each experiment, alive mosquitoes were

retrieved for the different assays. The collection was random within the same parasite treatment and

replicate for all traits measured. For the entirety of their adult life, the mosquitoes were provided with

a 6% sucrose solution up until trait measurement. Males were removed as soon as they emerged to

avoid mating-associated costs.

(c) Proteins, carbohydrates and lipids quantification

Mosquitoes were collected 5, 10, or 15 days after emergence and frozen at -80 ºC. We quantified their

proteins, carbohydrates, and lipid content through colourimetry, which we adapted from [49]. Protein,

carbohydrates, and lipid concentrations were estimated from the same sample for each of the parasite

treatments and respective replicates. Each individual body was weighed and then submerged in a 100

µL PBS pH 7.4 solution before homogenisation. To each sample, we added two stainless beads (Ø 2

mm) and homogenized the sample using a Qiagen TissueLyser LT at a frequency of 30 Hz for three

minutes. Then, the protein, carbohydrate, and lipid content were assessed for each mosquito as

described below. It is noteworthy that although we aimed to extract as much of the resource content

from the mosquito, some still hold onto the fibrous tissue. Hence, our measurements concern content

that is rather available and circulating, rather than structural.

Proteins were quantified by centrifuging the samples at 1100 RPM at 4º C for four minutes, adding 200

µL Bradford reagent to each solution, incubating at room temperature with soft agitation for 20 min,

and reading the absorbance at 595 nm with a SpectraMax i3x plate reader. To quantify protein content,

a standard curve of seven concentrations ranging from 400 µg/ml to 10 µg/ml of albumin in extraction

buffer (100 mM monopotassium phosphate, 1 mM DTT, and 1 mM EDTA in Milli-Q water) was also

prepared.

Carbohydrates were measured by first adding 20 µL of sodium sulphate at 20% to the homogenate,

adding 5 µL of the extraction buffer and 1500 µL Chloroform-Methanol (1:2, v:v), vortexing, and

centrifuging at 1100 RPM, 4ºC for 15 minutes. The supernatant was then removed, of which 300 µL

was transferred to a 96-well borosilicate plate and left to evaporate under the hood for approximately

30 minutes at room temperature. 240 µL of anthrone reagent was added to each sample and incubated

at room temperature for 15 minutes. The plate was sealed and incubated in a water bath at 90ºC for 15

minutes before measuring absorbance at 625 nm. A standard curve of seven concentrations ranging

from 40 µg/ml to 1 µg/ml of glucose in Chloroform-Methanol (1:2, v:v) was also prepared to quantify

carbohydrate content.

Lipids were measured by adding 200 µL of the supernatant to a 96-well borosilicate plate, then heated

to 90 ºC for 30 minutes. 10 µL of 98% sulfuric acid was added, and the sample was sealed and incubated

for two minutes in a water bath at 90 ºC. The samples were transferred to ice for three minutes before

adding 190 µL of vanillin reagent. Samples were sealed and incubated at room temperature for 15

minutes with gentle agitation. Their absorbance was measured at 525 nm. A standard curve of seven

concentrations ranging from 200 µg/ml to 5 µg/ml of triolein in Chloroform-Methanol (1:2, v:v) was

also prepared.

A blank sample was also included in all the standard curves to control for contamination. To acquire

individual resource content, the amount of resources measured was divided by the weight of the

mosquito. Details regarding the experiment scale and replication can be found in **Table 1**.

(d) Metals quantification

Five, 10 and 15 days after emergence, 30 alive females per parasite line and replicate were snap-frozen

in a 2 ml microcentrifuge tube at -80ºC. The samples were freeze-dried and submitted to a

mineralisation procedure using a Titan MPS microwave (Perkin Elmer, Waltham, MA). Each sample

was placed into a PTFE digestion vessel, and 2.5 mL HNO3 (Rotipuran Supra 69%, ROTH, Germany)

and 2.5 mL H2O2 (analytical reagent grade, Fisher Scientific, UK) were added. The vessels were heated

following a two-step digestion program consisting of a first step of 25 min until reaching 175°C and a

second step of 10 min at 175°C. After cooling, the digested sample solutions were transferred to 15 mL

Falcon PP tubes and stored in the fridge. A blank solution was prepared using the same procedure

without the sample. The elements iron, zinc, copper, calcium and manganese were simultaneously

analysed using inductively coupled plasma optical emission spectrometry (ICP-OES) [24]. An Avio

550 max equipped with a dual view and an S23 autosampler (PerkinElmer, Waltham, MA) was used.

The instrument was operated in axial view with the following conditions: RF power 1450 W, cyclonic

spray chamber with nebuliser nitrogen gas flow rate 0.7 mL/min, plasma argon gas flow rate 8 mL/min

and auxiliary compressed air gas flow rate 0.2 mL/min. The sample was injected into the plasma at 1.30

mL/min. All the calibration curves were prepared in 2% HNO3 MilliQ water (w/w) in the concentration

range of 0.1-10 mg/L and using SCP Science multi-element standards (SCP Science, France). Yttrium

(Atomic spectroscopy standard, PerkinElmer Pure, 2% HNO3, 1000 ppm) was used as the internal

standard and was added at a concentration of 1 mg/L in the blank standards and samples. The elemental

wavelengths were 238.204 nm for Fe, 206.200 nm for Zn, 327.393 nm for Cu, 317.933 nm for Ca,

257.610 nm for Mn and 371.029 nm for Y. Measurements were performed in triplicates. Details

regarding the experiment scale and replication can be found in **Table 1**.

(e) Iron metabolism assay

In a separate experiment, after rearing larvae and pupae infected with their respective parasite treatment,

all the pupae of a given combinatorial treatment (parasite treatment x replicate) were allowed to emerge

within a cage. The emerging adults were then randomly split into three diet treatments, starting the

experiment. Mosquitoes were fed with daily prepared cotton pads with either: i) iron (1 mM $FeCl_3$ in

6% sucrose solution), ii) chelator (200 µM Bathophenanthroline disulfonic acid disodium salt (BPS) in

6% sucrose solution) or iii) or sugar (6% sucrose solution) for five days after emergence. From day six

onwards, the mosquitoes were provided with cotton pads with only 6% sucrose solution. Alive

individual mosquitoes were collected for spore load quantification 5, 10, or 15 days after emergence.

They were placed individually into 2 ml microcentrifuge tubes with 1 ml of distilled water and a

stainless-steel bead (Ø 5 mm, and homogenised with a Qiagen TissueLyser LT at a frequency of 30 Hz

for one minute. The number of spores was counted in a haemocytometer under a phase-contrast

microscope (400x magnification). Details regarding the experiment scale and replication can be found

in **Table 1**.

(f) Statistical analysis

All the analyses were performed with R version 4.3.1. Statistical analysis and their diagnosis were

conducted using: "DHARMa" [50], "car" [51], "lme4" [52], "muhaz" [53], "emmeans" [54] and

"multcomp" [55] .

First, we tested if the parasite treatment or the day of collection affected the mosquito's body weight.

For this, we used a linear mixed model with log-transformed weight as a dependent variable and parasite

treatment, time of collection and their interaction as explanatory variables and replicate as a random

factor. Body weight was not affected by any of the factors (Parasite treatment: $\chi^2 = 1.38$, $df = 2$, $p =$

501; Time: $\chi^2 = 0.80$, $df = 1$, $p = 0.373$; Parasite treatment * Time: $\chi^2 = 1.04$, $df = 2$, $p = 0.60$).

Differences in resource content were assessed using a linear mixed model for each of the three resources

(proteins, carbohydrates, and lipids), with parasite treatment and time of collection as explanatory

variables and their interaction. Replicate was considered a random factor. Individual resource content

was quantified by measuring the sample's quantity and adjusting it according to the respective

individual weight. The square root of each resource content was calculated to fit the model best.

We ran a linear mixed model for each metal element to test for differences in the metallomes of

differently infected or uninfected mosquitoes across time. The dependent variable was the log-

transformed concentration of the element obtained by ICP-OES, while the parasite treatment and assay

day were explanatory variables and their interaction. Replicate was included as a random factor.

We assessed the effect of the parasite treatment and diet on spore rate and growth for each time point

assayed. For each spore rate comparison, we ran a generalised linear mixed model with a binomial error

structure, where parasite treatment and diet treatment were set as explanatory variables, their

interaction, and the presence or absence of spores as a response variable. Replicate was included as a

random factor. Concerning spore growth analyses, we only considered individuals with at least one

spore. We then ran a linear mixed model with the same explanatory and random factors as the previous

one.

3. Results

(a) Resource and energy use in mosquitoes infected with lines of V. culicis

The protein content was affected by parasite treatment ($\chi^2 = 9.75$, $df = 2$, $p = 0.008$), time ($\chi^2 = 45.28$,

$df = 1$, $p < 0.001$) and their interaction ($\chi^2 = 24.75$, $df = 2$, $p < 0.001$; Fig. 2a). In particular, protein

content remained stable until day 10 of adulthood when it significantly increased for mosquitoes

infected with late-selected spores (i.e., the most virulent ones) compared to the other parasite treatments

($\chi^2 = 9.75$, $df = 2$, $p = 0.008$).

The carbohydrate content increased with infection time ($\chi^2 = 139.65$, $df = 1$, $p < 0.001$; Fig. 2b). It was

higher in mosquitoes infected by the most virulent parasites (i.e., late-selected ones) than in those

infected by the unselected or the early-selected ones ($\chi^2 = 16.56$, $df = 2$, $p < 0.001$).

The lipid content decreased with infection time ($\chi^2 = 8.25$, $df = 1$, $p = 0.004$) irrespective of treatment,

and the mosquitoes infected with selected spores (i.e., early or late) had higher lipid content than the

ones infected with the unselected parasites ($\chi^2 = 6.33$, $df = 2$, $p = 0.042$; Fig. 2c).

(b) Metallomics of mosquitoes infected with lines of V. culicis

All metals (iron, copper, zinc, manganese) and calcium concentrations in the mosquito increased with

time (Fe: $\chi^2 = 18.88$, $df = 2$, $p < 0.001$; Cu: $\chi^2 = 11.93$, $df = 2$, $p = 0.003$; Zn: $\chi^2 = 18.62$, $df = 2$, $p = <$

0.001; Mn: ($\chi^2 = 17.91$, $df = 2$, $p < 0.001$; Ca: $\chi^2 = 34.08$, $df = 2$, $p < 0.001$). While iron, copper, and

calcium concentrations were similar for parasite-exposed and unexposed mosquitoes and for all parasite

lines (Fe: $\chi^2 = 2.26$, $df = 3$, $p = 0.521$; Cu: $\chi^2 = 0.66$, $df = 3$, $p = 0.88$; Ca: $\chi^2 = 3.19$, $df = 3$, $p = 0.36$), the

zinc concentrations were lower in infected mosquitoes, irrespective of parasite line (Zn: $\chi^2 = 13.97$, *df*

$= 3$, $p = 0.003$) and manganese concentration was higher ($\chi^2 = 32.30$, $df = 3$, $p < 0.001$) (**Fig. 3b, e**).

(c) Iron effect on parasite growth

In the iron supplementation or chelation experiment, all mosquitoes harboured spores by day 15. At

days 5 and 10, the proportion of mosquitoes with spores was higher if the parasite had been late-selected

than if it had been early-selected or not selected (day 5: $\chi^2 = 15.43$, $df = 2$, $p < 0.001$; day 10: $\chi^2 = 8.46$,

$df = 2$, $p = 0.015$). On both days, iron supplementation increased while chelation decreased the spore

rate (day 5: $\chi^2 = 15.43$, $df = 2$, $p < 0.001$; day 10: $\chi^2 = 8.46$, $df = 2$, $p = 0.015$). A similar pattern was

observed for the spore load in individuals that harboured spores, with higher spore load after an iron-

supplemented diet and lower spore load after a chelator-supplemented diet (day 5: $\chi^2 = 12.45$, $df = 2$, p

$= 0.002$; day 10: $\chi^2 = 37.60$, $df = 2$, $p < 0.001$; day 15: $\chi^2 = 79.65$, $df = 2$, $p < 0.001$; **Fig. 4b**). Individuals

infected with late-transmitted spores also showed the highest number of spores regardless of the time

point or diet (day 5: $\chi^2 = 37.75$, $df = 2$, $p < 0.001$; day 10: $\chi^2 = 37.60$, $df = 2$, $p < 0.001$; day 15: $\chi^2 =$

26.73, $df = 2$, $p < 0.001$; **Fig. 4b**), except for time-point 15 days where infection by the unselected

parasite reached the same load as the late-transmitted spores for the iron-supplemented diet (interaction

between parasite and diet treatments: $\chi^2 = 18.60$, $df = 4$, $p < 0.001$; **Fig. 4b**).

Figure 2. Mean protein, carbohydrate and lipid content through infection. Mean square root of

the **(a)** protein, **(b)** carbohydrate and **(c)** lipid content at day 5, 10 and 15 of adulthood upon infection

with early-, late-selected, or unselected stock parasite. Each time-point consists of individual resource

content from 50 live females per parasite treatment. Statistical factors and respective *p*-values are

shown on the right of each plot. Letters denote differences from multiple comparisons tests for an

interaction between "parasite treatment" and "time". For statistical details, see SI Appendix Table S1.

Figure 3. Metallomic profiles at adulthood. Metal concentrations determined by ICP-OES in
uninfected *An. gambiae* and infected with one of three *V. culicis* parasite lines: early, late or unselected
stock. Iron **(a)**, zinc **(b)**, copper **(c)**, calcium **(d),** and manganese **(e)** concentrations were quantified
across adult age at days 5, 10 and 15 post emergences. For each time point, a pool of 30 females was
used to assess metal concentration for each selection treatment x replicate combination (i.e., *n* = 5 data
points per selection treatment). Statistical factors and respective *p*-values are shown on the right of each
plot. Letters denote differences from multiple comparisons tests for an interaction between "parasite
treatment" and "time". The mean and standard error of the mean were plotted. For further statistical
details, see SI Appendix Table S2.

Figure 4. The effect of iron-supplementation and -chelation on parasite growth. The impact of a
standard sugar diet (6% Sucrose), supplemented with an iron-chelator (6% Sucrose + 200 µl BPS) or
iron itself (6% Sucrose + FeCl₃ 1mM) on the presence of detectable spores (i.e., the proportion of
females with spores secreted) **(a-c)** and spore load **(d-f)** of differently selected, or unselected, *V. culicis*
treatments. Each selection treatment has a sample size of 50 females split by five experimental replicates

321 for each time point. Letters denote differences from multiple comparisons tests for an interaction
322 between "parasite treatment" and "diet" for each independent time point. For further statistical details,
323 see SI Appendix Table S3.

4. Discussion

Parasites are masters at host exploitation, sequestering, and manipulating host resources to their benefit. In this study, we show how infection by the microsporidian parasite *V. culicis* alters host resource concentrations, particularly metals and energy sources stored in the mosquito host *A. gambiae*. We could also link virulence better to host exploitation by using parasite lines that differ in virulence - that is, the degree of harm inflicted on the host. The findings discussed below reveal microsporidia-specific mechanisms of host exploitation that may be inherently present or selectively enhanced under particular conditions. In addition to characterizing these mechanisms, we consider and discuss how their differential selection influences the potential role of microsporidia as biological control agents within vector biology.

(a) Parasite virulence drives host energy use during infection

First, we observed that protein content in mosquitoes increased over time, particularly between days 10 and 15 of adulthood **(Fig. 2a)**. As most immune effectors and signalling pathways are protein-based, this increase may indicate a host shift to parasite recognition and consequent immune response. If so, our findings support the threshold hypothesis [56], which proposes that an immune response is triggered only once parasite load surpasses a certain threshold. This would explain the delayed rise in proteins and potential immune effectors, such as antimicrobial peptides.

Moreover, the magnitude of the increase varied with parasite virulence: infection with the most virulent parasite line (i.e., *Late*) led to higher host protein levels by day 15 **(Fig. 2a)**. A similar pattern has been observed in several other studies [7,57,58], and is likely linked to the much higher parasite burdens reached by *Late* parasites, as shown in our previous work [47]. These results also highlight the mosquito host's ability to detect and adjust life history investments in immunity or other fitness traits, such as fecundity, based on the virulence of the infecting parasite. For instance, *Late* spores have been shown to induce earlier reproduction in *A. gambiae* mosquitoes [47].

We also measured two fundamental energetic reserves in mosquitoes: carbohydrates and lipids. Carbohydrates are typically more accessible and readily used, whereas lipids are a last resort or emergency energy source. Over time, we found that lipid reserves declined slightly **(Fig. 2c)**, while carbohydrate reserves increased for most parasite lines **(Fig. 2b)**. This pattern likely reflects our

experimental conditions, where adult mosquitoes had ad libitum access to sucrose (a carbohydrate) but no lipid-based diet. Consequently, although both energy reserves are utilised during infection, as shown previously for this microsporidian species [46], carbohydrate stores are replenished, whereas lipid reserves are steadily depleted or sequestered. Interestingly, we do find a difference in energetic use from a previous study that only focused on infection by the unselected line of *V. culicis* [46]. While in this latter study, Zeferino and colleagues (2024) observed a shift in energy use later in mosquito life from carbohydrates to lipids, here we observed an almost immediate use of lipids and a gradual increase in carbohydrates, particularly in the selected parasite lines, *Late* and *Early*. Although perhaps speculative, we can hypothesize that this difference might be driven by the selection process, which selects for better-adapted parasites to *A. gambiae* and therefore, better host exploitation. To aggravate this, parasite lines that better exploit the host, such as *Early* and particularly *Late*, and produce higher numbers of infective spores, might have higher energy demands. Previous studies on this model system have shown the ability of this parasite to change the host behaviour [59,60]. Hence, it might be conceivable that either the host or the parasite is driving the mosquito to feed more and have its carbohydrates replenished for the duration of the infection.

Therefore, parasite treatment influenced the dynamics of these energy sources **(Fig. 2bc)**. Carbohydrates increased proportionally to parasite virulence, which can reflect the host's increased energetic demand for immunity [61–63] and damage control [64], or the parasite's needs, as mentioned above. Additionally, *V. culicis* draws upon the host's carbohydrate reserves for its own consumption early in adulthood [46]. Thus, more virulent parasites, which elicit stronger immune responses and produce more infective spores, impose greater energy demands on the host. The proportional increase in carbohydrates with parasite virulence may therefore represent a behavioural adaptation by the host to fuel investment in immunity.

Finally, parasite treatment also influenced baseline lipid reserves **(Fig. 2c)**, with mosquitoes infected by selected parasites exhibiting higher lipid levels. The cause of this is unclear, but it may reflect greater parasite growth in these treatments compared to the unselected *Stock* parasite, leading to more lipid incorporation into spore production. It is also worth noting that our previous work showed that lipids are heavily exploited by the parasite later in infection [46], after day 22 of adulthood - beyond the time points measured here. Hence, lipid usage by the parasite may not yet have been strong enough to be detectable during the sampled stages, and due to the considerable mortality in some of the treatments, we could not measure later time points.

(b) V. culicis infection alters host zinc and manganese levels independently of parasite selection

We observed notable changes in host metal content during infection **(Fig. 3)** and its usage by the parasite **(Fig. 4)**. Among the metals measured, only zinc **(Fig. 3b)** and manganese **(Fig. 3e)** were significantly

affected by infection. Infections with any of the parasite treatments led to a decrease in host zinc content.
This could reflect heavy utilization of zinc by the microsporidian parasite or represent a mechanism of
immune evasion [44]. Zinc is essential for parasite growth and its proper metabolism [10,33], and is
often linked to the production of virulence factors [65–67]. Furthermore, some fungal parasites can
manipulate host immunity by inducing localised zinc deficits that impair immune responses [11,67–
69]. However, our selection protocol had no detectable impact on zinc-related processes, as no
differences were observed between parasite treatments.

In contrast, manganese levels increased following infection with any of the parasite lines **(Fig. 3e)**. This
may indicate a parasite response to host immune defences, as many microsporidian antioxidant
enzymes, such as manganese superoxide dismutases, depend on manganese for their activity and are
particularly relevant for this taxon [70–72]. As with zinc, this increase in manganese was independent
of the parasite's selection history.

Across all treatments, we also noted a slight temporal increase in metal concentrations. Although the
underlying mechanisms remain unclear, metals may become more bioavailable as infection progresses,
potentially benefiting the host, the parasite, or both **(Fig. 3)**. From the parasite's perspective, enhanced
spore production over time would increase demand for metals sequestered from the host, making their
accumulation more detectable at later stages. Future studies will be essential to better understand metal
mobilization and dynamics during microsporidian infection and within the microsporidian cell.

(c) Iron availability enhances parasite growth and spore production

Given the crucial role of metals in infection dynamics, and particularly the known importance of iron
for parasite development, we next focused specifically on how iron availability affects *V. culicis* growth
within the *A. gambiae* host.

Iron plays a vital role as a nutrient for growth in many parasite species [16,17,21,24,39,73], and it is
particularly abundant in mosquitoes due to their hematophagous ecology [23,74]. In fungal parasites,
iron acquisition is a critical determinant of virulence [75,76]. Many fungi have evolved sophisticated
mechanisms to overcome host-imposed iron limitation, including the secretion of siderophores - small,
high-affinity iron-transporter compounds - or the expression of more specialized iron transporter
systems [18,77,78]. Although microsporidia lack many canonical metabolic pathways due to their
highly reduced genomes [31], recent genomic studies suggest they have retained or adapted specific
mechanisms for iron acquisition, including transporters likely involved in scavenging iron from the host
cytoplasm [79], which was not the focus of the previously conducted metallomics.

Hence, in our study, we experimentally manipulated host iron levels by supplementing or chelating
circulating iron in the mosquito haemolymph, consequently increasing and reducing its absorption by

host cells and the replicating parasitic cells inside them. We then assessed the growth of different *V.*
culicis treatments **(Fig. 4)**. Five-day iron supplementation and chelation led to an increase and decrease,
respectively, in the production of infective spores at days 5 and 10 **(Fig. 4ab)**, but not at day 15, by
which all mosquitoes harboured spores **(Fig. 4c)**. These effects were mirrored in parasite burden under
the two dietary treatments **(Fig. 4d-f)**.

Overall, selected parasites exhibited a greater capacity to utilise circulating iron within the mosquito
compared to the unselected *Stock*, resulting in accelerated production of infective spores. This result
was not evident in the results from the metallomics, likely due to the small fraction of iron diverted to
the parasite compared to the whole-body iron content that is not only circulating but also incorporated
into many host cell structures. Differences in iron usage likely reflect adaptations arising during the
selection experiment, favouring parasites that more efficiently sequester and use host iron. Enhanced
iron acquisition is a known evolutionary strategy among intracellular parasites to sustain rapid
proliferation under resource-limited conditions. However, the extent of these differences appeared
inversely related to parasite growth: *Stock* parasites, initially with lower burdens, showed a larger
relative increase in response to iron supplementation than selected parasites, which already maintained
high burdens without supplementation. This trend was evident at day 15 **(Fig. 4f)**, when the parasite
usually reaches its chronic persistent load, and suggests the lower increase in selected lines might be
due to either limiting costs in the assimilation of iron by the parasite or limited evolvability at this time
for the mechanism that is allowing a better iron sequestration by the parasite.

Interestingly, iron manipulation by fungal parasites is not only associated with growth but also with
evasion of host immune responses [16,75,76]. In particular, the ability to modulate host iron
homeostasis can impair the oxidative burst or other immune functions that are heavily dependent on
iron-regulated pathways [80]. Whether microsporidia, including *V. culicis*, directly manipulate
mosquito iron homeostasis or simply compete with the host for this resource remains an open question.
Nevertheless, our findings provide evidence that iron availability is a critical driver of microsporidian
growth within the host, and that virulence evolution in *V. culicis* may be partly linked to improved iron
sequestration strategies.

(d) Conclusion and perspectives

In conclusion, this study provides important insights into how *V. culicis* manipulates its host, *A.*
gambiae, to exploit host resources for its growth and reproduction. We have shown that the parasite
alters the dynamics of proteins, carbohydrates, lipids, and metals in a manner that is strongly influenced
by the virulence of the parasite. More virulent strains of *V. culicis* seem to induce stronger immune
responses in the host, resulting in higher protein levels and altered energy metabolism. Additionally,
the study highlights the critical role of metal homeostasis, particularly zinc, manganese, and iron, in

this host-parasite interaction, with iron availability being a key factor in parasite growth and spore production. These findings underscore the complex interplay between parasite, virulence, host resource allocation, and immune response.

Furthermore, the results suggest that the evolutionary adaptation of *V. culicis* to more efficiently exploit host resources, including iron, may play a significant role in driving within-host virulence and consequent host exploitation strategies. As the role of microsporidia as potential biological control agents in vector biology continues to be explored, these insights have implications for understanding the dynamics of parasitic disease and the potential for manipulating these interactions for disease control. Particularly concerning malaria, our findings suggest the inhibitory effect of microsporidian parasites on *Plasmodium spp.* might be due to competition for a common resource, rather than collateral immunity.

Author contributions

LMS conceived the overall idea, JCK and LMS designed the experiments, LMS collected and analysed the data, AV performed the ICP-OES elemental analysis, and LMS wrote the first draft of the manuscript. All authors contributed critically to the drafts.

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Supplementary Information

Table S1. Protein, carbohydrate, and lipid content through infection.

Table S2. Metallomic profiles at adulthood.

Table S3. Iron-supplementation and -chelation on parasite growth.

Supplementary Information

"Mechanisms of host exploitation by a microsporidian parasite"

by Luis M. Silva, Armelle Vallat and Jacob C. Koella

Table S1. Protein, carbohydrate and lipid content through infection.

Table S2. Metallomic profiles at adulthood.

Table S3. Iron-supplementation and -chelation on parasite growth.

Table S1. Protein, carbohydrate and lipid content through infection. Each resource was analysed
independently. Differences in resource content were assessed using a linear mixed model for each of
the three resources (proteins, carbohydrates, and lipids), with parasite treatment and time of collection
as explanatory variables and their interaction. Replicate was considered a random factor. Individual
resource content was quantified by measuring the sample's quantity and adjusting it according to the
respective individual weight. The square root of each resource content was calculated to fit the model
best. Multiple comparisons tests were conducted to examine the interaction between "parasite
treatment" and "time" and show pairwise differences between all combinatorial treatments.

Tested effect			
Model 1a - Protein content	χ2	*df*	*p*
Parasite treatment	9.752	2	**0.008**
Time	45.283	1	**< 0.001**
Parasite treatment: Time	24.748	2	**< 0.001**
Multiple comparisons for Model 1a (PT: T)	*df*	*t-ratio*	*p*
Early, day 5 - Late, day 5	437	-0.719	0.999
Early, day 5 - Stock, day 5	437	-0.584	0.999
Early, day 5 - Early, day 10	437	-0.583	0.999
Early, day 5 - Late, day 10	437	-1.692	0.752
Early, day 5 - Stock, day 10	437	-2.108	0.469
Early, day 5 - Early, day 15	437	-6.996	**< 0.001**
Early, day 5 - Late, day 15	437	-11.761	**< 0.001**
Early, day 5 - Stock, day 15	437	-4.326	**< 0.001**
Late, day 5 - Stock, day 5	437	0.135	1.000
Late, day 5 - Early, day 10	437	0.132	1.000
Late, day 5 - Late, day 10	437	-0.973	0.988
Late, day 5 - Stock, day 10	437	-1.386	0.903
Late, day 5 - Early, day 15	437	-6.277	**< 0.001**
Late, day 5 - Late, day 15	437	-11.042	**< 0.001**
Late, day 5 - Stock, day 15	437	-3.607	**0.010**
Stock, day 5 - Early, day 10	437	-0.002	1.000
Stock, day 5 - Late, day 10	437	-1.108	0.973
Stock, day 5 - Stock, day 10	437	-1.521	0.845
Stock, day 5 - Early, day 15	437	-6.412	**< 0.001**
Stock, day 5 - Late, day 15	437	-11.177	**< 0.001**
Stock, day 5 - Stock, day 15	437	-3.742	**0.006**
Early, day 10 - Late, day 10	437	-1.100	0.974
Early, day 10 - Stock, day 10	437	-1.512	0.850
Early, day 10 - Early, day 15	437	-6.377	**< 0.001**
Early, day 10 - Late, day 15	437	-11.118	**< 0.001**
Early, day 10 - Stock, day 15	437	-3.721	**0.007**
Late, day 10 - Stock, day 10	437	-0.408	1.000
Late, day 10 - Early, day 15	437	-5.304	**< 0.001**
Late, day 10 - Late, day 15	437	-10.069	**< 0.001**
Late, day 10 - Stock, day 15	437	-2.635	0.175
Stock, day 10 - Early, day 15	437	-4.022	**< 0.001**
Stock, day 10 - Late, day 15	437	-9.710	**< 0.001**
Stock, day 10 - Stock, day 15	437	-2.239	0.382
Early, day 15 - Late, day 15	437	-4.765	**< 0.001**
Early, day 15 - Stock, day 15	437	2.670	0.162
Late, day 15 - Stock, day 15	437	7.435	**< 0.001**
Model 1b - Carbohydrate content	χ2	*df*	*p*
Parasite treatment	16.558	2	**< 0.001**
Time	139.651	1	**< 0.001**
Parasite treatment: Time	0.644	2	0.725
Multiple comparisons for Model 1b (PT: T)	*df*	*t-ratio*	*p*

	df	t-ratio	p
Early, day 5 - Late, day 5	437	-1.903	0.612
Early, day 5 - Stock, day 5	437	1.429	0.886
Early, day 5 - Early, day 10	437	-5.564	**< 0.001**
Early, day 5 - Late, day 10	437	-5.987	**< 0.001**
Early, day 5 - Stock, day 10	437	-4.771	**< 0.001**
Early, day 5 - Early, day 15	437	-7.421	**< 0.001**
Early, day 5 - Late, day 15	437	-8.193	**< 0.001**
Early, day 5 - Stock, day 15	437	-5.614	**< 0.001**
Late, day 5 - Stock, day 5	437	-3.331	**0.026**
Late, day 5 - Early, day 10	437	-3.672	**0.008**
Late, day 5 - Late, day 10	437	-4.084	**0.002**
Late, day 5 - Stock, day 10	437	-2.859	0.102
Late, day 5 - Early, day 15	437	-5.518	**< 0.001**
Late, day 5 - Late, day 15	437	-6.290	**< 0.001**
Late, day 5 - Stock, day 15	437	-3.712	**0.007**
Stock, day 5 - Early, day 10	437	-6.986	**< 0.001**
Stock, day 5 - Late, day 10	437	-7.415	**< 0.001**
Stock, day 5 - Stock, day 10	437	-6.207	**< 0.001**
Stock, day 5 - Early, day 15	437	-8.849	**< 0.001**
Stock, day 5 - Late, day 15	437	-9.621	**< 0.001**
Stock, day 5 - Stock, day 15	437	-7.043	**< 0.001**
Early, day 10 - Late, day 10	437	-0.391	1.000
Early, day 10 - Stock, day 10	437	0.845	0.996
Early, day 10 - Early, day 15	437	-1.818	0.670
Early, day 10 - Late, day 15	437	-2.586	0.200
Early, day 10 - Stock, day 15	437	-0.021	1.000
Late, day 10 - Stock, day 10	437	1.245	0.950
Late, day 10 - Early, day 15	437	-1.434	0.884
Late, day 10 - Late, day 15	437	-2.206	0.403
Late, day 10 - Stock, day 15	437	0.372	1.000
Stock, day 10 - Early, day 15	437	-2.686	0.156
Stock, day 10 - Late, day 15	437	-3.462	**0.018**
Stock, day 10 - Stock, day 15	437	-0.871	0.994
Early, day 15 - Late, day 15	437	-0.772	0.998
Early, day 15 - Stock, day 15	437	1.807	0.678
Late, day 15 - Stock, day 15	437	2.579	0.199

Model 1c - Lipid content	χ^2	df	p
Parasite treatment	6.333	2	**0.042**
Time	8.251	1	**0.004**
Parasite treatment: Time	0.924	2	0.630

Multiple comparisons for Model 1c (PT: T)	df	t-ratio	p
Early, day 5 - Late, day 5	437	0.127	1.000
Early, day 5 - Stock, day 5	437	1.906	0.610
Early, day 5 - Early, day 10	437	2.003	0.542
Early, day 5 - Late, day 10	437	2.666	0.163
Early, day 5 - Stock, day 10	437	3.528	**0.014**
Early, day 5 - Early, day 15	437	2.050	0.509
Early, day 5 - Late, day 15	437	2.193	0.412
Early, day 5 - Stock, day 15	437	2.783	0.124

Late, day 5 - Stock, day 5	437	1.779	0.696
Late, day 5 - Early, day 10	437	1.877	0.637
Late, day 5 - Late, day 10	437	2.539	0.216
Late, day 5 - Stock, day 10	437	3.401	**0.021**
Late, day 5 - Early, day 15	437	1.924	0.598
Late, day 5 - Late, day 15	437	2.066	0.497
Late, day 5 - Stock, day 15	437	2.656	0.167
Stock, day 5 - Early, day 10	437	0.107	1.000
Stock, day 5 - Late, day 10	437	0.760	0.998
Stock, day 5 - Stock, day 10	437	1.613	0.798
Stock, day 5 - Early, day 15	437	0.144	1.000
Stock, day 5 - Late, day 15	437	0.287	1.000
Stock, day 5 - Stock, day 15	437	0.877	0.994
Early, day 10 - Late, day 10	437	0.649	0.999
Early, day 10 - Stock, day 10	437	1.496	0.857
Early, day 10 - Early, day 15	437	0.037	1.000
Early, day 10 - Late, day 15	437	0.178	1.000
Early, day 10 - Stock, day 15	437	0.765	0.998
Late, day 10 - Stock, day 10	437	0.849	0.997
Late, day 10 - Early, day 15	437	-0.615	1.000
Late, day 10 - Late, day 15	437	-0.473	1.000
Late, day 10 - Stock, day 15	437	0.117	1.000
Stock, day 10 - Early, day 15	437	-1.467	0.870
Stock, day 10 - Late, day 15	437	-1.324	0.924
Stock, day 10 - Stock, day 15	437	-0.732	0.998
Early, day 15 - Late, day 15	437	0.142	1.000
Early, day 15 - Stock, day 15	437	0.732	0.998
Late, day 15 - Stock, day 15	437	0.590	1.000

Table S2. Metallomic profiles at adulthood. Similarly to the previous resources, each metal was analysed independently. We ran a linear mixed model for each metal element to test for differences in the metallomes of differently infected or uninfected mosquitoes across time. The dependent variable was the log-transformed concentration of the element obtained by ICP-OES, while the parasite treatment and assay day were explanatory variables and their interaction. Replicate was included as a random factor. Multiple comparisons tests were conducted to examine the interaction between "parasite treatment" and "time" and show pairwise differences between all combinatorial treatments.

Tested effect			
Model 2a - Iron (Fe)	**χ2**	**df**	**p**
Parasite treatment	2.258	3	0.521
Time	18.880	2	**< 0.001**
Parasite treatment: Time	10.555	6	0.103
Multiple comparisons for Model 2a (PT: T)	**df**	**t-ratio**	**p**
Early, day 5 - Late, day 5	43	-1.942	0.727
Early, day 5 - Stock, day 5	43	-0.226	1.000
Early, day 5 - Uninfected, day 5	43	-2.073	0.643
Early, day 5 - Early, day 10	43	-0.746	1.000
Early, day 5 - Late, day 10	43	-0.666	1.000
Early, day 5 - Stock, day 10	43	0.434	1.000
Early, day 5 - Uninfected, day 10	43	0.810	1.000
Early, day 5 - Early, day 15	43	-1.680	0.868
Early, day 5 - Late, day 15	43	-1.734	0.843
Early, day 5 - Stock, day 15	43	-2.488	0.374
Early, day 5 - Uninfected, day 15	43	-2.925	0.169
Late, day 5 - Stock, day 5	43	1.716	0.851
Late, day 5 - Uninfected, day 5	43	-0.131	1.000
Late, day 5 - Early, day 10	43	1.195	0.990
Late, day 5 - Late, day 10	43	1.276	0.978
Late, day 5 - Stock, day 10	43	2.376	0.443
Late, day 5 - Uninfected, day 10	43	2.752	0.237
Late, day 5 - Early, day 15	43	0.142	1.000
Late, day 5 - Late, day 15	43	0.208	1.000
Late, day 5 - Stock, day 15	43	-0.546	1.000
Late, day 5 - Uninfected, day 15	43	0.983	0.998
Stock, day 5 - Uninfected, day 5	43	-1.847	0.783
Stock, day 5 - Early, day 10	43	-0.520	1.000
Stock, day 5 - Late, day 10	43	-0.440	1.000
Stock, day 5 - Stock, day 10	43	0.660	1.000
Stock, day 5 - Uninfected, day 10	43	1.036	0.996
Stock, day 5 - Early, day 15	43	-1.469	0.942
Stock, day 5 - Late, day 15	43	-1.508	0.930
Stock, day 5 - Stock, day 15	43	-2.262	0.517
Stock, day 5 - Uninfected, day 15	43	-2.699	0.262
Uninfected, day 5 - Early, day 10	43	1.326	0.971
Uninfected, day 5 - Late, day 10	43	1.407	0.956
Uninfected, day 5 - Stock, day 10	43	2.506	0.363
Uninfected, day 5 - Uninfected, day 10	43	2.882	0.184
Uninfected, day 5 - Early, day 15	43	0.265	1.000
Uninfected, day 5 - Late, day 15	43	0.339	1.000
Uninfected, day 5 - Stock, day 15	43	-0.415	1.000
Uninfected, day 5 - Uninfected, day 15	43	-0.852	0.999
Early, day 10 - Late, day 10	43	0.081	1.000
Early, day 10 - Stock, day 10	43	1.180	0.988
Early, day 10 - Uninfected, day 10	43	1.556	0.915
Early, day 10 - Early, day 15	43	-0.979	0.997
Early, day 10 - Late, day 15	43	-0.987	0.997

Early, day 10 - Stock, day 15	43	-1.741	0.839
Early, day 10 - Uninfected, day 15	43	-2.178	0.573
Late, day 10 - Stock, day 10	43	1.099	0.993
Late, day 10 - Uninfected, day 10	43	1.475	0.939
Late, day 10 - Early, day 15	43	-1.055	0.995
Late, day 10 - Late, day 15	43	-1.068	0.995
Late, day 10 - Stock, day 15	43	-1.822	0.797
Late, day 10 - Uninfected, day 15	43	-2.259	0.519
Stock, day 10 - Uninfected, day 10	43	0.376	1.000
Stock, day 10 - Early, day 15	43	-2.087	0.634
Stock, day 10 - Late, day 15	43	-2.167	0.580
Stock, day 10 - Stock, day 15	43	-2.921	0.170
Stock, day 10 - Uninfected, day 15	43	-3.358	0.063
Uninfected, day 10 - Early, day 15	43	-2.440	0.403
Uninfected, day 10 - Late, day 15	43	-2.543	0.342
Uninfected, day 10 - Stock, day 15	43	-3.297	0.073
Uninfected, day 10 - Uninfected, day 15	43	-3.734	**0.024**
Early, day 15 - Late, day 15	43	0.053	1.000
Early, day 15 - Stock, day 15	43	-0.654	1.000
Early, day 15 - Uninfected, day 15	43	-1.064	0.995
Late, day 15 - Stock, day 15	43	-0.754	1.000
Late, day 15 - Uninfected, day 15	43	-1.191	0.988
Stock, day 15 - Uninfected, day 15	43	-0.437	1.000

Model 2b - Zinc (Zn)	χ^2	df	p
Parasite treatment	13.970	3	**0.003**
Time	18.622	2	**< 0.001**
Parasite treatment: Time	8.840	6	0.183

Multiple comparisons for Model 2b (PT: T)	df	t-ratio	p
Early, day 5 - Late, day 5	43	-0.712	1.000
Early, day 5 - Stock, day 5	43	-0.429	1.000
Early, day 5 - Uninfected, day 5	43	-2.346	0.462
Early, day 5 - Early, day 10	43	-0.934	0.998
Early, day 5 - Late, day 10	43	-1.012	0.997
Early, day 5 - Stock, day 10	43	0.229	1.000
Early, day 5 - Uninfected, day 10	43	-0.820	1.000
Early, day 5 - Early, day 15	43	-1.983	0.702
Early, day 5 - Late, day 15	43	-1.392	0.959
Early, day 5 - Stock, day 15	43	-2.119	0.613
Early, day 5 - Uninfected, day 15	43	-4.918	**< 0.001**
Late, day 5 - Stock, day 5	43	0.282	1.000
Late, day 5 - Uninfected, day 5	43	-1.635	0.886
Late, day 5 - Early, day 10	43	-0.223	1.000
Late, day 5 - Late, day 10	43	-0.300	1.000
Late, day 5 - Stock, day 10	43	0.941	0.999
Late, day 5 - Uninfected, day 10	43	-1.315	1.000
Late, day 5 - Early, day 15	43	-0.108	0.973
Late, day 5 - Late, day 15	43	-0.680	1.000
Late, day 5 - Stock, day 15	43	-1.407	0.956
Late, day 5 - Uninfected, day 15	43	-4.206	**0.006**

Stock, day 5 - Uninfected, day 5	43	-1.917	0.743
Stock, day 5 - Early, day 10	43	-0.505	1.000
Stock, day 5 - Late, day 10	43	-0.582	1.000
Stock, day 5 - Stock, day 10	43	0.659	1.000
Stock, day 5 - Uninfected, day 10	43	-0.390	1.000
Stock, day 5 - Early, day 15	43	-1.580	0.907
Stock, day 5 - Late, day 15	43	-0.963	0.998
Stock, day 5 - Stock, day 15	43	-1.690	0.863
Stock, day 5 - Uninfected, day 15	43	-4.488	**0.003**
Uninfected, day 5 - Early, day 10	43	1.412	0.955
Uninfected, day 5 - Late, day 10	43	1.335	0.970
Uninfected, day 5 - Stock, day 10	43	2.576	0.324
Uninfected, day 5 - Uninfected, day 10	43	1.527	0.925
Uninfected, day 5 - Early, day 15	43	0.219	1.000
Uninfected, day 5 - Late, day 15	43	0.954	0.998
Uninfected, day 5 - Stock, day 15	43	0.227	1.000
Uninfected, day 5 - Uninfected, day 15	43	-2.571	0.327
Early, day 10 - Late, day 10	43	-0.077	1.000
Early, day 10 - Stock, day 10	43	1.164	0.989
Early, day 10 - Uninfected, day 10	43	0.115	1.000
Early, day 10 - Early, day 15	43	-1.106	0.993
Early, day 10 - Late, day 15	43	-0.458	1.000
Early, day 10 - Stock, day 15	43	-1.185	0.988
Early, day 10 - Uninfected, day 15	43	-3.983	**0.012**
Late, day 10 - Stock, day 10	43	1.241	0.982
Late, day 10 - Uninfected, day 10	43	0.192	1.000
Late, day 10 - Early, day 15	43	-1.033	0.996
Late, day 10 - Late, day 15	43	-0.380	1.000
Late, day 10 - Stock, day 15	43	-1.107	0.993
Late, day 10 - Uninfected, day 15	43	-3.906	**0.015**
Stock, day 10 - Uninfected, day 10	43	-1.049	0.995
Stock, day 10 - Early, day 15	43	-2.198	0.560
Stock, day 10 - Late, day 15	43	-1.621	0.892
Stock, day 10 - Stock, day 15	43	-2.349	0.460
Stock, day 10 - Uninfected, day 15	43	-5.147	**< 0.001**
Uninfected, day 10 - Early, day 15	43	-1.214	0.985
Uninfected, day 10 - Late, day 15	43	-0.572	1.000
Uninfected, day 10 - Stock, day 15	43	-1.299	0.975
Uninfected, day 10 - Uninfected, day 15	43	-4.098	**0.009**
Early, day 15 - Late, day 15	43	0.677	1.000
Early, day 15 - Stock, day 15	43	-0.006	1.000
Early, day 15 - Uninfected, day 15	43	2.631	0.294
Late, day 15 - Stock, day 15	43	-0.727	1.000
Late, day 15 - Uninfected, day 15	43	-3.526	**0.042**
Stock, day 15 - Uninfected, day 15	43	-2.799	0.217
Model 2c - Copper (Cu)	$\chi 2$	*df*	*p*
Parasite treatment	0.663	3	0.882
Time	11.925	2	**0.003**
Parasite treatment: Time	6.333	6	0.387

Multiple comparisons for Model 2c (PT: T)	df	t-ratio	p
Early, day 5 - Late, day 5	43	-1.152	0.990
Early, day 5 - Stock, day 5	43	-0.412	1.000
Early, day 5 - Uninfected, day 5	43	-1.937	0.730
Early, day 5 - Early, day 10	43	-0.983	0.997
Early, day 5 - Late, day 10	43	-0.343	1.000
Early, day 5 - Stock, day 10	43	-0.138	1.000
Early, day 5 - Uninfected, day 10	43	-0.249	1.000
Early, day 5 - Early, day 15	43	-2.271	0.511
Early, day 5 - Late, day 15	43	-1.409	0.955
Early, day 5 - Stock, day 15	43	-2.668	0.277
Early, day 5 - Uninfected, day 15	43	-2.059	0.653
Late, day 5 - Stock, day 5	43	0.740	1.000
Late, day 5 - Uninfected, day 5	43	-0.786	1.000
Late, day 5 - Early, day 10	43	0.268	1.000
Late, day 5 - Late, day 10	43	0.809	1.000
Late, day 5 - Stock, day 10	43	1.014	0.997
Late, day 5 - Uninfected, day 10	43	0.903	0.999
Late, day 5 - Early, day 15	43	-1.190	0.988
Late, day 5 - Late, day 15	43	-0.258	1.000
Late, day 5 - Stock, day 15	43	-1.516	0.928
Late, day 5 - Uninfected, day 15	43	-0.907	0.999
Stock, day 5 - Uninfected, day 5	43	-1.526	0.925
Stock, day 5 - Early, day 10	43	-0.572	1.000
Stock, day 5 - Late, day 10	43	0.069	1.000
Stock, day 5 - Stock, day 10	43	0.274	1.000
Stock, day 5 - Uninfected, day 10	43	0.163	1.000
Stock, day 5 - Early, day 15	43	-1.885	0.762
Stock, day 5 - Late, day 15	43	-0.998	0.997
Stock, day 5 - Stock, day 15	43	-2.256	0.521
Stock, day 5 - Uninfected, day 15	43	-1.647	0.881
Uninfected, day 5 - Early, day 10	43	0.954	0.998
Uninfected, day 5 - Late, day 10	43	1.594	0.902
Uninfected, day 5 - Stock, day 10	43	1.799	0.810
Uninfected, day 5 - Uninfected, day 10	43	1.688	0.864
Uninfected, day 5 - Early, day 15	43	-0.453	1.000
Uninfected, day 5 - Late, day 15	43	0.528	1.000
Uninfected, day 5 - Stock, day 15	43	-0.730	1.000
Uninfected, day 5 - Uninfected, day 15	43	-0.121	1.000
Early, day 10 - Late, day 10	43	0.640	1.000
Early, day 10 - Stock, day 10	43	0.845	0.999
Early, day 10 - Uninfected, day 10	43	0.734	1.000
Early, day 10 - Early, day 15	43	-1.348	0.967
Early, day 10 - Late, day 15	43	-0.426	1.000
Early, day 10 - Stock, day 15	43	-1.684	0.865
Early, day 10 - Uninfected, day 15	43	-1.075	0.994
Late, day 10 - Stock, day 10	43	0.205	1.000
Late, day 10 - Uninfected, day 10	43	0.094	1.000
Late, day 10 - Early, day 15	43	-1.949	0.723

Late, day 10 - Late, day 15	43	-1.066	0.995
Late, day 10 - Stock, day 15	43	-2.325	0.476
Late, day 10 - Uninfected, day 15	43	-1.716	0.851
Stock, day 10 - Uninfected, day 10	43	-0.111	1.000
Stock, day 10 - Early, day 15	43	-2.141	0.598
Stock, day 10 - Late, day 15	43	-1.271	0.979
Stock, day 10 - Stock, day 15	43	-2.530	0.350
Stock, day 10 - Uninfected, day 15	43	-1.921	0.740
Uninfected, day 10 - Early, day 15	43	-2.037	0.667
Uninfected, day 10 - Late, day 15	43	-1.160	0.989
Uninfected, day 10 - Stock, day 15	43	-2.419	0.416
Uninfected, day 10 - Uninfected, day 15	43	-1.810	0.804
Early, day 15 - Late, day 15	43	0.948	0.998
Early, day 15 - Stock, day 15	43	-0.232	1.000
Early, day 15 - Uninfected, day 15	43	0.339	1.000
Late, day 15 - Stock, day 15	43	-1.258	0.980
Late, day 15 - Uninfected, day 15	43	-0.649	1.000
Stock, day 15 - Uninfected, day 15	43	0.609	1.000

Model 2d - Calcium (Ca)	χ^2	df	p
Parasite treatment	3.190	3	0.363
Time	34.084	2	**< 0.001**
Parasite treatment: Time	10.268	6	0.114

Multiple comparisons for Model 2d (PT: T)	df	t-ratio	p
Early, day 5 - Late, day 5	43	-1.999	0.691
Early, day 5 - Stock, day 5	43	-0.654	1.000
Early, day 5 - Uninfected, day 5	43	-1.295	0.975
Early, day 5 - Early, day 10	43	-1.399	0.957
Early, day 5 - Late, day 10	43	01.662	0.875
Early, day 5 - Stock, day 10	43	-0.208	1.000
Early, day 5 - Uninfected, day 10	43	0.730	1.000
Early, day 5 - Early, day 15	43	-2.786	0.222
Early, day 5 - Late, day 15	43	-3.075	0.122
Early, day 5 - Stock, day 15	43	-4.147	**0.008**
Early, day 5 - Uninfected, day 15	43	-3.234	0.085
Late, day 5 - Stock, day 5	43	1.346	0.968
Late, day 5 - Uninfected, day 5	43	0.705	1.000
Late, day 5 - Early, day 10	43	0.600	1.000
Late, day 5 - Late, day 10	43	0.337	1.000
Late, day 5 - Stock, day 10	43	1.791	0.814
Late, day 5 - Uninfected, day 10	43	2.730	0.247
Late, day 5 - Early, day 15	43	-0.910	0.999
Late, day 5 - Late, day 15	43	-1.076	0.994
Late, day 5 - Stock, day 15	43	-2.148	0.593
Late, day 5 - Uninfected, day 15	43	-1.234	0.983
Stock, day 5 - Uninfected, day 5	43	-0.641	1.000
Stock, day 5 - Early, day 10	43	-0.745	1.000
Stock, day 5 - Late, day 10	43	-1.009	0.997
Stock, day 5 - Stock, day 10	43	0.446	1.000
Stock, day 5 - Uninfected, day 10	43	1.384	0.961

	df	t-ratio	p
Stock, day 5 - Early, day 15	43	-2.172	0.577
Stock, day 5 - Late, day 15	43	-2.421	0.414
Stock, day 5 - Stock, day 15	43	-3.494	**0.045**
Stock, day 5 - Uninfected, day 15	43	-2.580	0.322
Uninfected, day 5 - Early, day 10	43	-0.104	1.000
Uninfected, day 5 - Late, day 10	43	-0.367	1.000
Uninfected, day 5 - Stock, day 10	43	1.087	0.994
Uninfected, day 5 - Uninfected, day 10	43	2.025	0.675
Uninfected, day 5 - Early, day 15	43	-1.571	0.910
Uninfected, day 5 - Late, day 15	43	-1.780	0.819
Uninfected, day 5 - Stock, day 15	43	-2.853	0.195
Uninfected, day 5 - Uninfected, day 15	43	-1.939	0.729
Early, day 10 - Late, day 10	43	-0.263	1.000
Early, day 10 - Stock, day 10	43	1.101	0.987
Early, day 10 - Uninfected, day 10	43	2.129	0.606
Early, day 10 - Early, day 15	43	-1.473	0.940
Early, day 10 - Late, day 15	43	-1.676	0.869
Early, day 10 - Stock, day 15	43	-2.748	0.239
Early, day 10 - Uninfected, day 15	43	-1.835	0.790
Late, day 10 - Stock, day 10	43	1.454	0.945
Late, day 10 - Uninfected, day 10	43	2.392	0.432
Late, day 10 - Early, day 15	43	-1.226	0.984
Late, day 10 - Late, day 15	43	-1.413	0.955
Late, day 10 - Stock, day 15	43	-2.485	0.376
Late, day 10 - Uninfected, day 15	43	-1.571	0.910
Stock, day 10 - Uninfected, day 10	43	0.938	0.998
Stock, day 10 - Early, day 15	43	-2.590	0.316
Stock, day 10 - Late, day 15	43	-2.867	0.190
Stock, day 10 - Stock, day 15	43	-3.939	**0.014**
Stock, day 10 - Uninfected, day 15	43	-3.026	0.136
Uninfected, day 10 - Early, day 15	43	-3.471	**0.047**
Uninfected, day 10 - Late, day 15	43	-3.805	**0.020**
Uninfected, day 10 - Stock, day 15	43	-4.878	**< 0.001**
Uninfected, day 10 - Uninfected, day 15	43	3.964	**0.013**
Early, day 15 - Late, day 15	43	-0.100	1.000
Early, day 15 - Stock, day 15	43	-1.106	0.993
Early, day 15 - Uninfected, day 15	43	-0.249	1.000
Late, day 15 - Stock, day 15	43	-1.072	0.994
Late, day 15 - Uninfected, day 15	43	-0.159	1.000
Stock, day 15 - Uninfected, day 15	43	0.914	0.999

Model 2e - Manganese (Mn)	χ^2	df	p
Parasite treatment	32.303	3	**< 0.001**
Time	17.909	2	**< 0.001**
Parasite treatment: Time	4.655	6	0.589
Multiple comparisons for Model 2e (PT: T)	df	t-ratio	p
Early, day 5 - Late, day 5	43	-0.803	1.000
Early, day 5 - Stock, day 5	43	-0.669	1.000
Early, day 5 - Uninfected, day 5	43	1.700	0.859
Early, day 5 - Early, day 10	43	-0.302	1.000

Early, day 5 - Late, day 10	43	-0.448	1.000
Early, day 5 - Stock, day 10	43	0.663	1.000
Early, day 5 - Uninfected, day 10	43	2.786	0.222
Early, day 5 - Early, day 15	43	-2.078	0.640
Early, day 5 - Late, day 15	43	-1.490	0.935
Early, day 5 - Stock, day 15	43	-2.874	0.187
Early, day 5 - Uninfected, day 15	43	0.864	0.999
Late, day 5 - Stock, day 5	43	0.134	1.000
Late, day 5 - Uninfected, day 5	43	2.503	0.365
Late, day 5 - Early, day 10	43	0.501	1.000
Late, day 5 - Late, day 10	43	0.356	1.000
Late, day 5 - Stock, day 10	43	1.466	0.942
Late, day 5 - Uninfected, day 10	43	3.589	**0.035**
Late, day 5 - Early, day 15	43	-1.324	0.971
Late, day 5 - Late, day 15	43	-0.687	1.000
Late, day 5 - Stock, day 15	43	-2.071	0.645
Late, day 5 - Uninfected, day 15	43	1.667	0.873
Stock, day 5 - Uninfected, day 5	43	2.369	0.447
Stock, day 5 - Early, day 10	43	0.367	1.000
Stock, day 5 - Late, day 10	43	0.221	1.000
Stock, day 5 - Stock, day 10	43	1.332	0.970
Stock, day 5 - Uninfected, day 10	43	3.455	**0.050**
Stock, day 5 - Early, day 15	43	-1.450	0.946
Stock, day 5 - Late, day 15	43	-0.821	1.000
Stock, day 5 - Stock, day 15	43	-2.205	0.555
Stock, day 5 - Uninfected, day 15	43	1.533	0.923
Uninfected, day 5 - Early, day 10	43	-2.002	0.690
Uninfected, day 5 - Late, day 10	43	-2.147	0.594
Uninfected, day 5 - Stock, day 10	43	-1.037	0.996
Uninfected, day 5 - Uninfected, day 10	43	-1.087	0.994
Uninfected, day 5 - Early, day 15	43	-3.673	**0.028**
Uninfected, day 5 - Late, day 15	43	-3.190	0.094
Uninfected, day 5 - Stock, day 15	43	-4.574	**0.002**
Uninfected, day 5 - Uninfected, day 15	43	-0.836	0.999
Early, day 10 - Late, day 10	43	-0.145	1.000
Early, day 10 - Stock, day 10	43	0.965	0.998
Early, day 10 - Uninfected, day 10	43	3.088	0.119
Early, day 10 - Early, day 15	43	-1.794	0.812
Early, day 10 - Late, day 15	43	-1.188	0.987
Early, day 10 - Stock, day 15	43	-2.572	0.326
Early, day 10 - Uninfected, day 15	43	1.166	0.989
Late, day 10 - Stock, day 10	43	1.111	0.993
Late, day 10 - Uninfected, day 10	43	3.234	0.085
Late, day 10 - Early, day 15	43	-1.658	0.877
Late, day 10 - Late, day 15	43	-1.043	0.996
Late, day 10 - Stock, day 15	43	-2.426	0.411
Late, day 10 - Uninfected, day 15	43	1.312	0/.973
Stock, day 10 - Uninfected, day 10	43	2.123	0.610
Stock, day 10 - Early, day 15	43	-2.700	0.260

Stock, day 10 - Late, day 15	43	-2.153	0.590
Stock, day 10 - Stock, day 15	43	-3.537	**0.040**
Stock, day 10 - Uninfected, day 15	43	0.201	1.000
Uninfected, day 10 - Early, day 15	43	-4.692	**0.001**
Uninfected, day 10 - Late, day 15	43	4.277	**0.005**
Uninfected, day 10 - Stock, day 15	43	-5.660	**< 0.001**
Uninfected, day 10 - Uninfected, day 15	43	-1.922	0.739
Early, day 15 - Late, day 15	43	-.680	1.000
Early, day 15 - Stock, day 15	43	-0.619	1.000
Early, day 15 - Uninfected, day 15	43	2.889	0.187
Late, day 15 - Stock, day 15	43	-1.384	0.961
Late, day 15 - Uninfected, day 15	43	2.354	0.457
Stock, day 15 - Uninfected, day 15	43	3.738	**0.024**

Table S3. The effect of iron-supplementation and -chelation on parasite growth. We assessed the
effect of the parasite treatment and diet on spore rate and growth for each time point assayed. For each
spore rate comparison, we ran a generalised linear mixed model with a binomial error structure, where
parasite treatment and diet treatment were set as explanatory variables, their interaction, and the
presence or absence of spores as a response variable. Replicate was included as a random factor.
Concerning spore growth analyses, we only considered individuals with at least one spore. We then ran
a linear mixed model with the same explanatory and random factors as the previous one. Multiple
comparisons tests were conducted to examine the interaction between "parasite treatment" and "diet"
and show pairwise differences between all combinatorial treatments.

Tested effect			
Model 3a - Spore rate at day 5	χ2	*df*	*p*
Diet treatment	15.430	2	**< 0.001**
Parasite treatment	15.546	2	**< 0.001**
Diet treatment: Parasite treatment	4.986	4	0.289
Multiple comparisons for Model 3a (PT: D)	*df*	*t-ratio*	*p*
Early, Sugar - Late, Sugar	inf	-1.396	0.900
Early, Sugar - Stock, Sugar	inf	0.203	1.000
Early, Sugar - Early, Sugar + BPS	inf	1.460	0.874
Early, Sugar - Late, Sugar + BPS	inf	0.000	1.000
Early, Sugar - Stock, Sugar + BPS	inf	1.460	0.874
Early, Sugar - Early, Sugar + FeCl₃	inf	-1.790	0.689
Early, Sugar - Late, Sugar + FeCl₃	inf	-2.976	0.072
Early, Sugar - Stock, Sugar + FeCl₃	inf	0.823	0.996
Late, Sugar - Stock, Sugar	inf	1.594	0.808
Late, Sugar - Early, Sugar + BPS	inf	2.792	0.118
Late, Sugar - Late, Sugar + BPS	inf	1.396	0.900
Late, Sugar - Stock, Sugar + BPS	inf	2.792	0.118
Late, Sugar - Early, Sugar + FeCl₃	inf	-0.405	1.000
Late, Sugar - Late, Sugar + FeCl₃	inf	-1.656	0.773
Late, Sugar - Stock, Sugar + FeCl₃	inf	2.192	0.411
Stock, Sugar - Early, Sugar + BPS	inf	1.262	0.942
Stock, Sugar - Late, Sugar + BPS	inf	-0.203	1.000
Stock, Sugar - Stock, Sugar + BPS	inf	1.262	0.942
Stock, Sugar - Early, Sugar + FeCl₃	inf	-1.986	0.553
Stock, Sugar - Late, Sugar + FeCl₃	inf	-3.160	**0.042**
Stock, Sugar - Stock, Sugar + FeCl₃	inf	0.621	1.000
Early, Sugar + BPS - Late, Sugar + BPS	inf	-1.460	**0.874**
Early, Sugar + BPS - Stock, Sugar + BPS	inf	0.000	1.000
Early, Sugar + BPS - Early, Sugar + FeCl₃	inf	-3.160	**0.042**
Early, Sugar + BPS - Late, Sugar + FeCl₃	inf	-4.241	**< 0.001**
Early, Sugar + BPS - Stock, Sugar + FeCl₃	inf	-0.648	0.999
Late, Sugar + BPS - Stock, Sugar + BPS	inf	1.460	0.874
Late, Sugar + BPS - Early, Sugar + FeCl₃	inf	-1.790	0.689
Late, Sugar + BPS - Late, Sugar + FeCl₃	inf	-2.976	0.072
Late, Sugar + BPS - Stock, Sugar + FeCl₃	inf	0.823	0.996
Stock, Sugar + BPS - Early, Sugar + FeCl₃	inf	-3.160	**0.042**
Stock, Sugar + BPS - Late, Sugar + FeCl₃	inf	-4.241	**< 0.001**
Stock, Sugar + BPS - Stock, Sugar + FeCl₃	inf	-0.648	0.999
Early, Sugar + FeCl₃ - Late, Sugar + FeCl₃	inf	-1.262	0.942
Early, Sugar + FeCl₃ - Stock, Sugar + FeCl₃	inf	2.574	0.200
Late, Sugar + FeCl₃ - Stock, Sugar + FeCl₃	inf	3.705	**0.007**
Model 3b- Spore rate at day 10	χ2	*df*	*p*

	χ2	df	p
Diet treatment	8.464	2	**0.015**
Parasite treatment	16.867	2	**< 0.001**
Diet treatment: Parasite treatment	0.504	4	0.973

Multiple comparisons for Model 3b (PT: D)	df	t-ratio	p
Early, Sugar - Late, Sugar	inf	-0.001	1.000
Early, Sugar - Stock, Sugar	inf	1.275	0.9387
Early, Sugar - Early, Sugar + BPS	inf	2.209	0.3993
Early, Sugar - Late, Sugar + BPS	inf	0.972	0.9883
Early, Sugar - Stock, Sugar + BPS	inf	2.785	0.1195
Early, Sugar - Early, Sugar + FeCl$_3$	inf	-0.002	1.000
Early, Sugar - Late, Sugar + FeCl$_3$	inf	-0.002	1.000
Early, Sugar - Stock, Sugar + FeCl$_3$	inf	0.575	1.000
Late, Sugar - Stock, Sugar	inf	0.001	1.000
Late, Sugar - Early, Sugar + BPS	inf	0.001	1.000
Late, Sugar - Late, Sugar + BPS	inf	0.001	1.000
Late, Sugar - Stock, Sugar + BPS	inf	0.001	1.000
Late, Sugar - Early, Sugar + FeCl$_3$	inf	0.000	1.000
Late, Sugar - Late, Sugar + FeCl$_3$	inf	-0.002	1.000
Late, Sugar - Stock, Sugar + FeCl$_3$	inf	0.001	0.878
Stock, Sugar - Early, Sugar + BPS	inf	1.451	1.000
Stock, Sugar - Late, Sugar + BPS	inf	-0.391	0.252
Stock, Sugar - Stock, Sugar + BPS	inf	2.459	1.000
Stock, Sugar - Early, Sugar + FeCl$_3$	inf	-0.002	1.000
Stock, Sugar - Late, Sugar + FeCl$_3$	inf	-0.002	0.997
Stock, Sugar - Stock, Sugar + FeCl$_3$	inf	-0.826	0.706
Early, Sugar + BPS - Late, Sugar + BPS	inf	-1.764	0.961
Early, Sugar + BPS - Stock, Sugar + BPS	inf	1.180	1.000
Early, Sugar + BPS - Early, Sugar + FeCl$_3$	inf	-0.002	1.000
Early, Sugar + BPS - Late, Sugar + FeCl$_3$	inf	-0.002	0.507
Early, Sugar + BPS - Stock, Sugar + FeCl$_3$	inf	-2.051	0.154
Late, Sugar + BPS - Stock, Sugar + BPS	inf	2.682	1.000
Late, Sugar + BPS - Early, Sugar + FeCl$_3$	inf	-0.002	1.000
Late, Sugar + BPS - Late, Sugar + FeCl$_3$	inf	-0.002	1.000
Late, Sugar + BPS - Stock, Sugar + FeCl$_3$	inf	-0.456	1.000
Stock, Sugar + BPS - Early, Sugar + FeCl$_3$	inf	-0.003	1.000
Stock, Sugar + BPS - Late, Sugar + FeCl$_3$	inf	-0.002	1.000
Stock, Sugar + BPS - Stock, Sugar + FeCl$_3$	inf	-2.837	0.105
Early, Sugar + FeCl$_3$ - Late, Sugar + FeCl$_3$	inf	-0.002	1.000
Early, Sugar + FeCl$_3$ - Stock, Sugar + FeCl$_3$	inf	0.002	1.000
Late, Sugar + FeCl$_3$ - Stock, Sugar + FeCl$_3$	inf	0.002	1.000

Model 3d - Spore load at day 5	χ2	df	p
Diet treatment	37.751	2	**< 0.001**
Parasite treatment	12.451	2	**0.002**
Diet treatment: Parasite treatment	6.925	4	0.140

Multiple comparisons for Model 3d (PT: D)	df	t-ratio	p
Early, Sugar - Late, Sugar	190	-1.257	0.942
Early, Sugar - Stock, Sugar	189	1.284	0.935
Early, Sugar - Early, Sugar + BPS	190	1.844	0.653
Early, Sugar - Late, Sugar + BPS	190	0.494	1.000

Comparison	df	t-ratio	p
Early, Sugar - Stock, Sugar + BPS	190	2.225	0.394
Early, Sugar - Early, Sugar + $FeCl_3$	188	0.558	1.000
Early, Sugar - Late, Sugar + $FeCl_3$	189	-3.197	**0.042**
Early, Sugar - Stock, Sugar + $FeCl_3$	189	2.128	0.458
Late, Sugar - Stock, Sugar	189	2.613	0.188
Late, Sugar - Early, Sugar + BPS	192	3.041	0.066
Late, Sugar - Late, Sugar + BPS	188	1.791	0.689
Late, Sugar - Stock, Sugar + BPS	190	3.456	**0.019**
Late, Sugar - Early, Sugar + $FeCl_3$	190	1.982	0.558
Late, Sugar - Late, Sugar + $FeCl_3$	189	-2.040	0.518
Late, Sugar - Stock, Sugar + $FeCl_3$	189	3.439	**0.020**
Stock, Sugar - Early, Sugar + BPS	190	0.676	0.999
Stock, Sugar - Late, Sugar + BPS	189	-0.795	0.997
Stock, Sugar - Stock, Sugar + BPS	190	1.057	0.980
Stock, Sugar - Early, Sugar + $FeCl_3$	188	-0.841	0.995
Stock, Sugar - Late, Sugar + $FeCl_3$	188	-4.591	**< 0.001**
Stock, Sugar - Stock, Sugar + $FeCl_3$	188	0.889	0.993
Early, Sugar + BPS - Late, Sugar + BPS	191	-1.398	0.898
Early, Sugar + BPS - Stock, Sugar + BPS	189	0.351	1.000
Early, Sugar + BPS - Early, Sugar + $FeCl_3$	190	-1.476	0.865
Early, Sugar + BPS - Late, Sugar + $FeCl_3$	190	-4.790	**< 0.001**
Early, Sugar + BPS - Stock, Sugar + $FeCl_3$	190	0.157	1.000
Late, Sugar + BPS - Stock, Sugar + BPS	189	1.786	0.691
Late, Sugar + BPS - Early, Sugar + $FeCl_3$	190	0.020	1.000
Late, Sugar + BPS - Late, Sugar + $FeCl_3$	190	-3.753	**0.007**
Late, Sugar + BPS - Stock, Sugar + $FeCl_3$	189	1.660	0.770
Stock, Sugar + BPS - Early, Sugar + $FeCl_3$	190	-1.883	0.626
Stock, Sugar + BPS - Late, Sugar + $FeCl_3$	190	-5.217	**< 0.001**
Stock, Sugar + BPS - Stock, Sugar + $FeCl_3$	190	-0.210	1.000
Early, Sugar + $FeCl_3$ - Late, Sugar + $FeCl_3$	189	-4.193	**0.001**
Early, Sugar + $FeCl_3$ - Stock, Sugar + $FeCl_3$	189	1.764	0.706
Late, Sugar + $FeCl_3$ - Stock, Sugar + $FeCl_3$	189	5.343	**< 0.001**

Model 3e - Spore load at day 10	χ^2	df	p
Diet treatment	37.601	2	**< 0.001**
Parasite treatment	48.705	2	**< 0.001**
Diet treatment: Parasite treatment	3.038	4	0.552

Multiple comparisons for Model 3e (PT: D)	df	t-ratio	p
Early, Sugar - Late, Sugar	404	-3.455	**0.018**
Early, Sugar - Stock, Sugar	404	0.192	1.000
Early, Sugar - Early, Sugar + BPS	404	2.299	0.345
Early, Sugar - Late, Sugar + BPS	404	-0.281	1.000
Early, Sugar - Stock, Sugar + BPS	404	1.247	0.945
Early, Sugar - Early, Sugar + $FeCl_3$	404	-2.241	0.381
Early, Sugar - Late, Sugar + $FeCl_3$	404	-5.341	**< 0.001**
Early, Sugar - Stock, Sugar + $FeCl_3$	404	-1.372	0.908
Late, Sugar - Stock, Sugar	404	3.592	**0.048**
Late, Sugar - Early, Sugar + BPS	404	5.606	**< 0.001**
Late, Sugar - Late, Sugar + BPS	404	3.137	**0.048**
Late, Sugar - Stock, Sugar + BPS	404	4.429	**< 0.001**

Late, Sugar - Early, Sugar + FeCl$_3$	404	1.220	0.952
Late, Sugar - Late, Sugar + FeCl$_3$	404	-1.896	0.618
Late, Sugar - Stock, Sugar + FeCl$_3$	404	2.058	0.504
Stock, Sugar - Early, Sugar + BPS	404	2.083	0.487
Stock, Sugar - Late, Sugar + BPS	404	-0.466	1.000
Stock, Sugar - Stock, Sugar + BPS	404	1.054	0.980
Stock, Sugar - Early, Sugar + FeCl$_3$	404	-2.398	0.288
Stock, Sugar - Late, Sugar + FeCl$_3$	404	-5.448	**< 0.001**
Stock, Sugar - Stock, Sugar + FeCl$_3$	404	-1.541	0.835
Early, Sugar + BPS - Late, Sugar + BPS	404	-2.546	0.214
Early, Sugar + BPS - Stock, Sugar + BPS	404	-0.932	0.991
Early, Sugar + BPS - Early, Sugar + FeCl$_3$	404	-4.448	**< 0.001**
Early, Sugar + BPS - Late, Sugar + FeCl$_3$	404	-7.405	**< 0.001**
Early, Sugar + BPS - Stock, Sugar + FeCl$_3$	404	-3.599	**0.011**
Late, Sugar + BPS - Stock, Sugar + BPS	404	1.495	0.858
Late, Sugar + BPS - Early, Sugar + FeCl$_3$	404	-1.935	0.589
Late, Sugar + BPS - Late, Sugar + FeCl$_3$	404	-5.002	**< 0.001**
Late, Sugar + BPS - Stock, Sugar + FeCl$_3$	404	-1.078	0.977
Stock, Sugar + BPS - Early, Sugar + FeCl$_3$	404	-3.313	**0.028**
Stock, Sugar + BPS - Late, Sugar + FeCl$_3$	404	-6.163	**< 0.001**
Stock, Sugar + BPS - Stock, Sugar + FeCl$_3$	404	-2.5-6	0.232
Early, Sugar + FeCl$_3$ - Late, Sugar + FeCl$_3$	404	-3.116	0.050
Early, Sugar + FeCl$_3$ - Stock, Sugar + FeCl$_3$	404	-0.850	0.996
Late, Sugar + FeCl$_3$ - Stock, Sugar + FeCl$_3$	404	3.934	**0.003**

Model 3f - Spore load at day 15	χ2	df	p
Diet treatment	7.271	2	**0.0264**
Parasite treatment	79.910	2	**< 0.001**
Diet treatment: Parasite treatment	18.102	4	**0.001**

Multiple comparisons for Model 3f (PT: D)	df	t-ratio	p
Early, Sugar - Late, Sugar	437	-1.128	0.970
Early, Sugar - Stock, Sugar	437	1.553	0.830
Early, Sugar - Early, Sugar + BPS	437	7.691	**< 0.001**
Early, Sugar - Late, Sugar + BPS	437	2.497	0.236
Early, Sugar - Stock, Sugar + BPS	437	5.713	**< 0.001**
Early, Sugar - Early, Sugar + FeCl$_3$	437	-0.074	1.000
Early, Sugar - Late, Sugar + FeCl$_3$	437	-2.633	0.176
Early, Sugar - Stock, Sugar + FeCl$_3$	437	-3.095	0.053
Late, Sugar - Stock, Sugar	437	2.681	0.158
Late, Sugar - Early, Sugar + BPS	437	8.8219	**< 0.001**
Late, Sugar - Late, Sugar + BPS	437	3.625	**0.010**
Late, Sugar - Stock, Sugar + BPS	437	6.841	**< 0.001**
Late, Sugar - Early, Sugar + FeCl$_3$	437	1.053	0.980
Late, Sugar - Late, Sugar + FeCl$_3$	437	-1.505	0.853
Late, Sugar - Stock, Sugar + FeCl$_3$	437	-1.967	0.567
Stock, Sugar - Early, Sugar + BPS	437	6.139	**< 0.001**
Stock, Sugar - Late, Sugar + BPS	437	0.944	0.990
Stock, Sugar - Stock, Sugar + BPS	437	4.160	**0.001**
Stock, Sugar - Early, Sugar + FeCl$_3$	437	-1.627	0.790
Stock, Sugar - Late, Sugar + FeCl$_3$	437	-4.186	**0.001**

Stock, Sugar - Stock, Sugar + FeCl$_3$	437	-4.648	< 0.001
Early, Sugar + BPS - Late, Sugar + BPS	437	-5.194	< 0.001
Early, Sugar + BPS - Stock, Sugar + BPS	437	-1.979	0.559
Early, Sugar + BPS - Early, Sugar + FeCl$_3$	437	-7.766	< 0.001
Early, Sugar + BPS - Late, Sugar + FeCl$_3$	437	-10.325	< 0.001
Early, Sugar + BPS - Stock, Sugar + FeCl$_3$	437	-10.787	< 0.001
Late, Sugar + BPS - Stock, Sugar + BPS	437	3.216	0.037
Late, Sugar + BPS - Early, Sugar + FeCl$_3$	437	-2.572	0.202
Late, Sugar + BPS - Late, Sugar + FeCl$_3$	437	-5.130	< 0.001
Late, Sugar + BPS - Stock, Sugar + FeCl$_3$	437	-5.592	< 0.001
Stock, Sugar + BPS - Early, Sugar + FeCl$_3$	437	-5.787	< 0.001
Stock, Sugar + BPS - Late, Sugar + FeCl$_3$	437	-8.346	< 0.001
Stock, Sugar + BPS - Stock, Sugar + FeCl$_3$	437	-8.808	< 0.001
Early, Sugar + FeCl$_3$ - Late, Sugar + FeCl$_3$	437	-2.559	0.207
Early, Sugar + FeCl$_3$ - Stock, Sugar + FeCl$_3$	437	-3.021	0.066
Late, Sugar + FeCl$_3$ - Stock, Sugar + FeCl$_3$	437	-0.462	1.000