Cell-cycle-dependent repression of histone gene transcription by histone H4

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Abstract
In all eukaryotes DNA replication is coupled to histone synthesis to coordinate chromatin packaging of the genome. Canonical histone genes coalesce in the nucleus into the Histone Locus Body (HLB), where gene transcription and 3' mRNA processing occurs. Both histone gene transcription and mRNA stability are reduced when DNA replication is inhibited, implying that the Histone Locus Body senses the rate of DNA synthesis. In *Drosophila melanogaster*, the S-phase-induced histone genes are repeated in an ~100 copy repeat unit array, whereas in humans, these histone genes are scattered. In both organisms these genes coalesce into Histone Locus Bodies. We used a transgenic histone gene reporter and RNAi in Drosophila to identify canonical H4 histone as a unique repressor of histone synthesis during the G2 phase in germline primary spermatocytes. Using cytology and CUT&Tag chromatin profiling, we find that histone H4 uniquely occupies histone gene promoters in both Drosophila and human cells. Our results suggest that repression of histone genes by soluble histone H4 is a conserved mechanism that coordinates DNA replication with histone synthesis in proliferating cells.

Introduction
The genome of eukaryotic cells is packaged into nucleosomes, where DNA is wrapped around histone octamers. In animal cells, the genes encoding canonical histone proteins are highly distinctive: These multi-copy genes are abundantly transcribed by RNA Polymerase II (RNAPII) during S phase of the cell cycle, and are the only protein-coding genes that produce transcripts without introns or 3' polyadenylation [Duronio & Marzluff 2017]. Histone genes nucleate a distinctive body within the nucleus termed the Histone Locus Body (HLB), where specific transcription factors and RNA processing proteins localize. The HLBs of Drosophila and mammals share fundamental molecular components, including the Cyclin E/CDK2-activated transcription cofactor Mxc/NPAT, 3' mRNA stem-loops, Stem Loop Binding Protein (SLBP) and the U7 snRNP 3'-end processing machinery. Despite these cytological and compositional similarities, the histone genes are radically different in gene organization: In *Drosophila melanogaster* the five canonical histone genes (H1, H2A, H2B, H3 and H4), are arranged in a unit tandemly repeated ~100 times at one locus [Liu et al 2006], whereas in humans 72 genes are scattered with a major cluster on Chromosome 6 and two minor clusters on Chromosome 1. These human genes are non-repetitive and embedded in euchromatic regions of chromosomes, while the tandemly repeated Drosophila genes are subject to heterochromatic silencing [Elgin 1996], and it has been unclear how many of these 200 gene repeat units in a diploid cell are transcribed. By profiling both *D. melanogaster* and human histone genes, we aim to understand ancient conserved mechanisms of S-phase-dependent histone gene regulation that have endured despite profound genomic and epigenomic changes.

Here, we have taken advantage of Drosophila histone gene rescue constructs [Günesdogan et al 2010; MacKay et al 2015] and fluorescently marked histone transgene reporters [Shindo & Amodeo 2019] to test for histone gene derepression after knockdown of candidate repressors in the synchronized G2 phase gonial cells of testes. We discovered that reduction of histone H4 strongly derepressed histone gene expression outside of S phase, but no other candidate regulator had an effect. Using imaging, we showed that histone H4 localizes to the HLB in Drosophila cells; using CUT&Tag chromatin profiling, we precisely localized histone H4 to histone gene promoters, coinciding with peaks of Mxc/NPAT, initiating RNAPII and of active chromatin. Turning to human K562 cells, we observed similar cytological localization of histone H4 to the HLBs and coincident genomic localization of histone H4, NPAT, and RNAPII at active histone genes. These results imply a direct mechanism whereby excess histone H4 in cells buffers histone gene transcription to coordinate chromatin packaging with DNA replication.

Results
Chromatin features at active and silenced histone genes in Drosophila
As the histone genes in the Histone Locus Body (HLB) are repetitive, and normal cells contain both active and silenced histone genes, mapping of chromatin features to a genome assembly cannot distinguish which features are associated with which expression state. To address this, we profiled chromatin features in two genotypes: wildtype, where some of the ~200 histone genes must be active while others are silenced, and the "12XWT" line, where the histone locus has been deleted and a construct carrying 12 copies of the histone repeat unit (HRU) rescues the flies [Crain et al 2024]. We expect all copies of the histone genes to be active in this second genotype. By comparing chromatin profiles between these two genotypes, we infer the chromatin features of active and of silenced histone genes.

We dissected wing imaginal discs from male larvae as a

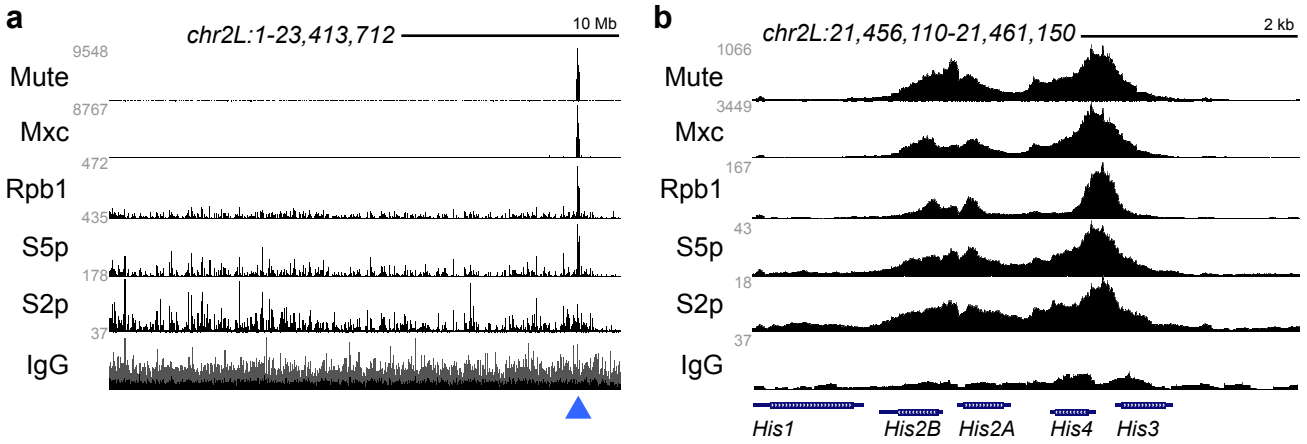


Figure 1. Chromatin factors and RNAPII at the histone locus in wing imaginal disc cells. (a) Browser tracks of chromatin factors and RNAPII isoforms across chromosome *2L* of Drosophila. The blue arrowhead marks the location of the histone locus. **(b)** Browser tracks of chromatin factors and RNAPII isoforms across one Histone Repeat Unit.

sample of proliferating cells, and subjected them to CUT&Tag profiling [Kaya-Okur et al 2019], generating >1 million reads mapped to the dm6 genome assembly for each sample (**Supplementary Table 1**). We first used antibodies to the HLB-specific transcription cofactors Mxc [White et al 2011] and Mute [Bulchand et al 2010]. As expected, these two factors are localized to only the histone locus in the genome (**Figure 1a**). Signal for both factors is broadly dispersed across the *His3-His4* and *His2A-His2B* gene pairs of the 5 kb HRU, with peaks at the divergent promoters of each pair, and no signal over the adjacent *His1* gene (**Figure 1b**). These results are consistent with previous chromatin mapping of Mxc in Drosophila embryos [Hodkinson et al 2024]. The divergent *His4-His3* promoter region nucleates HLB formation [Salzler et al 2013], and these binding profiles are consistent with the idea that Mxc binds at these sites in the histone locus and nucleates HLB formation.

To assess transcriptional activity of the histone genes, we profiled multiple components and isoforms of RNA polymerase II (RNAPII) in larval wing disc samples. As expected, the RNAPII component unphosphorylated Rpb1, and phosphorylated initiating (RNAPII-S5p) and elongating (RNAPII-S2p) isoforms of Rpb1 mark the histone locus in wildtype cells (**Figure 1**). In fact, the histone locus is the major site of Rpb1 and RNAPII-S5p signal across the genome (**Figure 1a**), accounting for 1.8% and 2.5% of mapped reads, respectively, while only 0.2% of reads for the RNAPII-S2p isoform map to this locus. This is consistent with cytological description of enrichment for the unmodified and initiating forms of RNAPII at the HLB in other Drosophila cell types [Huang et al 2021, Kemp et al 2021, Cho et al 2022]. Note that the *His3-His4* and *His2A-His2B* have strong peaks of RNAPII-S5p and RNAPII-S2p isoforms at their promoters and across their gene bodies, indicating high transcription of these genes. In contrast, *His1* has only a low broad distribution of these polymerase isoforms across its length (**Figure 1b**), implying that it is expressed at lower levels.

We then compared signal counts for Mxc, Mute, and RNAPII between wing disc samples from wildtype and from *12XWT* flies. We calculated per-gene counts to adjust for the different numbers of copies in the two strains (100 copies/genome in wildtype, 12 copies/genome in 12XWT). The per-copy sum counts of Mute and Mxc signal are higher in wildtype than in 12X flies, and proportional to the numbers of histone genes in these genotypes (**Figure 2a**). In contrast, signals for the RNAPII-S5p isoform are dramatically increased, and signal for the RNAPII-S2p isoform shows a slight gain. Thus, each histone gene in the 12XWT line must carry more RNAPII than in wildtype. This is consistent with the idea that some histone genes are active and others silenced in wildtype, or alternatively that each gene is active an intermediate level. In either case, all histone genes in the 12XWT strain appear to be more heavily transcribed, presumably to support cell proliferation.

Heterochromatic histone marks at silenced HLB genes

To determine histone modifications associated with histone genes, we first profiled five modifications associated with active gene expression [Barth & Imhof 2010] (**Figure 2b**). In wildtype samples, all three methylation states of the histone H3K4 residue (H3K4me1, H3K4me2, and H3K4me3) and acetylation at histone H3-K27 (H3K27ac) are enriched at the wildtype histone locus at levels comparable to surrounding active enhancers and genes. in contrast, there is little detectable trimethylation of the K36 residue of histone H3 (H3K36me3), consistent with the intronless structure of histone genes [Schwartz et al 2009]. Adjusting for copy number, there is a substantial per-gene copy gain only in the H3K4me3 mark in the 12XWT line (**Figure 2a**) and no or moderate gains in the H3K4me1 and H3K4me2 marks. These changes are consistent with the higher average transcription of histone genes in this genotype.

We then examined five histone modifications typically associated with silencing across histone genes between the two Drosophila lines. Three modifications are strongly enriched at the histone genes in wildtype samples: mono- and di-methylation of histone H3 at lysine-9 (H3K9me1 and H3K9me2 [Ebert et al 2006]), and ubiquitinylation of histone H2A at lysine-118 (uH2A [Barbour et al 2020]) (**Figure 2b**). These first two marks imply that histone genes have a partially heterochromatic character likely due to the repeated

a

	wt	log2(12XWT/12) – log2(wt/100)				
	log_2(CPM)	*His1*	*His2A*	*His2B*	*His3*	*His4*
Mxc	18.50	1.31	-0.05	0.40	0.06	0.33
Mute	18.84	1.78	0.73	1.05	0.87	0.95
RNAPII-S5p	14.00	4.12	3.47	3.81	3.95	4.21
RNAPII-S2p	11.48	2.17	1.59	2.03	1.93	2.40
H3K4me1	11.63	0.61	-0.38	0.31	0.16	0.16
H3K4me2	13.10	2.46	1.77	2.29	1.99	1.81
H3K4me3	12.92	3.91	3.12	3.34	3.06	3.07
H3K27ac	11.06	2.00	1.32	1.77	0.94	1.52
H3K36me3	9.15	3.02	1.19	2.02	1.97	2.09
H3K9me1	12.56	-0.57	-1.23	-0.73	-1.00	-0.82
H3K9me2	10.51	0.95	-1.23	-1.11	-0.94	-2.48
H3K9me3	9.70	2.11	1.44	1.60	-0.05	1.70
H3K27me3	9.36	0.56	0.39	0.86	0.99	1.34
uH2A	11.21	2.21	1.05	1.87	1.43	1.61

log_2FC
-2.5 0 +4.2

b

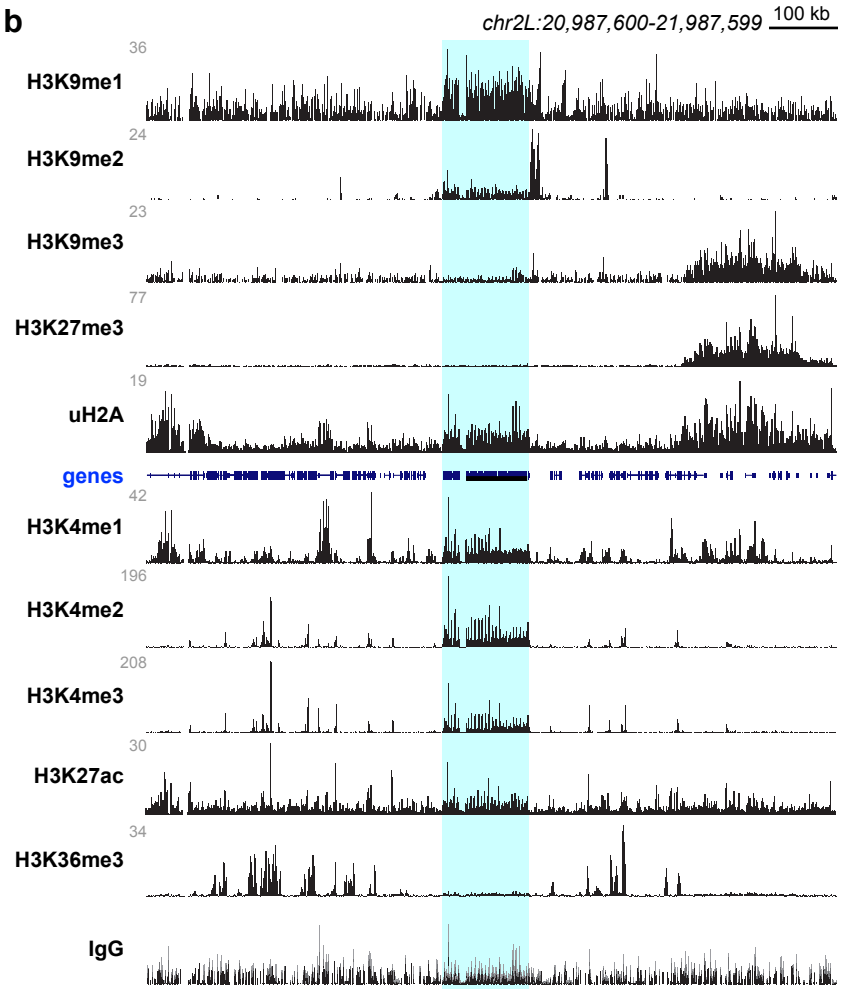


Figure 2. Histone modifications at the histone locus in wing imaginal disc cells. (a) Per-gene copy abundance of chromatin features and histone modifications in wildtype cells (log_2 CPM), and the fold-change in per-gene copy abundance between the *12XWT* strain (12 histone gene copies) and wildtype (100 copies). **(b)** Browser tracks of histone modifications around the histone locus in wildtype wing imaginal disc cells. Blue shading marks the histone locus.

genes, although the locus lacks tri-methylation of histone H3 at lysine-9 (H3K9me3 [Ebert et al 2006]) (**Figure 2b**). The presence of uH2A at histone genes suggests that these are sites of PRC1 activity, although the canonical trimethylation mark of histone H3 at lysine-27 (H3K27me3 [Cao et al 2002]) mark of Polycomb-silenced domains is absent.

Since any histone modification associated with histone gene silencing should be present in wildtype but absent in the 12XWT line, we profiled silencing histone modifications in this rescue line, and calculated per-gene copy changes (**Figure 2a**). Previous results have implicated H3K9 methylation in histone gene silencing [Ner et al 2002; Ito et al 2012]. Indeed, the per-gene copy coverage of the H3K9me1 and H3K9me2 marks drop in the 12XWT strain compared to wildtype. In contrast, per-gene density of the uH2A modification is slightly

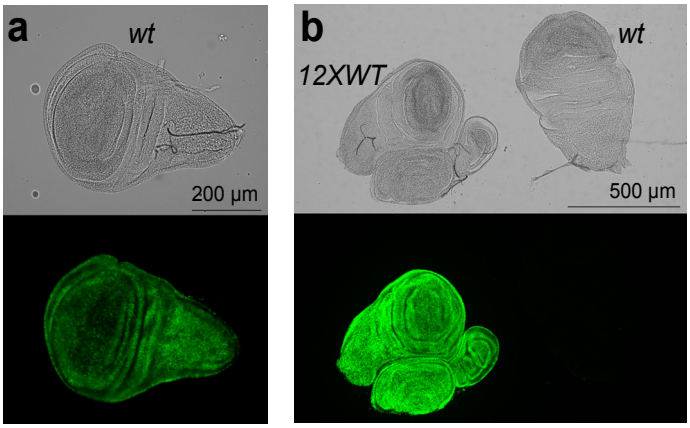


Figure 3. Expression of a *His2ADendra2* reporter Histone Repeat Unit in wildtype and 12XWT strains. (a) Fluorescence of the *His2ADendra2* reporter in a wildtype larval wing imaginal disc, adjusted to display the weak fluorescence expressed from the reporter in this genotype. **(b)** Fluorescence of *His2ADendra2* in larval wing imaginal discs from wildtype (wt) and from 12XWT larvae. Fluorescence in 12XWT background is 37X that in the wildtype background (average summed fluorescence of wildtype discs 5,908,005 ± 1,096,058 a.u. *versus* 158,704 + 26,029 a.u. in the 12XWT background).

increased. There is very little of the H3K9me3 or H3K27me3 marks in either the wildtype or 12XWT strains. These results support the idea that inactive histone genes are marked with mono- and demethylation of the H3K9 residue, where the wildtype histone locus is a mixture of transcriptionally active histone genes mixed with silenced histone genes. This is analogous to the functional organization of ribosomal RNA genes, where actively transcribed units are intermixed with silenced units [Grummt & Pikaard 2003]. In both cases a mixture of active and silenced genes may allow cells to fine-tune histone production to the needs of cell proliferation and growth, the rates of which vary between tissues and life stages.

A visual reporter for histone gene silencing

Extra histone repeat unit transgenes are repressed in proportion to the number of total histone genes in a genotype [Chaubal et al 2023], an effect that appears similar to the reduced expression of histone genes in wildtype. To visualize histone gene expression in living animals, we used HRU reporter constructs where either the *His3* or the *His2A* gene is fused to the octocoral *Dendra2* fluorescent protein coding sequence [Shindo & Amodeo 2019; **Methods**]. These constructs express fluorescently tagged histones in eggs and developing embryos [Shindo & Amodeo 2019], and at low levels in proliferating cells of later stages such as in larval imaginal wing discs (**Figure 3a**). We reasoned that if these transgenes are partially repressed, then genetically interfering with histone gene silencing would produce more fluorescent protein. Indeed, the expression of the *His2ADendra2* HRU transgene is dramatically increased ~37X in the 12XWT background compared to its expression in wildtype in wing imaginal discs (**Figure 3b**). This implies that cells with reduced histone gene numbers sense a dearth of histones, and upregulate all histone genes to compensate and provide for chromatin duplication.

We wished to identify the mechanism by which histone genes are repressed, but genetic reduction of such a factor might inhibit viability in the growing wing imaginal disc. Therefore, we turned to a non-essential tissue where we could still assess reporter expression with RNAi. The adult testis is dispensable for organismal viability. It also spatially organized so that the developmental and cell cycle stage of cells can be identified by their position in the tissue [White-Cooper 2010]. Germline stem cells are located at the apical testis tip and undergo four rounds of mitotic division in this proliferating zone, producing G2 phase spermatogonial cells. These then grow for ~4 days before meiosis and sperm differentiation. We imaged these stages by dissecting and fixing testes from males carrying a *bamGAL4* driver with an inducible *UAS-RFP* transgene. This combination produces RFP specifically in gonial cells (**Figure 4a**). We immunostained these testes with antibodies to Mxc, a constitutive component of the HLB, and to phospho-Mxc, which is catalyzed by Cyclin E/Cdk2 kinase in S phase of the cell cycle [White et al 2007]. The proliferating zone at the apical tip of the testis is marked with large HLBs containing phosphorylated Mxc, and abut RFP-stained G2 phase cells (**Figure 4a**). Slightly more distal in the testis the Mxc-labeled HLB is divided into 2-4 smaller dots, consistent with the unpairing of homologous loci in this developmental stage [Vazquez et al 2002], and Mxc staining disappears from nuclei in later primary spermatocytes before meiosis.

We first imaged fluorescence from a control *His2AVDendra2* histone variant gene (located on chromosome *3* outside of the histone locus), and this variant protein is abundant throughout the apical tip of the testis (**Figure 4b,c**). In contrast, the HRU transgenes are repressed in the testis. No fluorescence from the *His2ADendra2* transgene is apparent in the very apical tip including in bam-positive gonial cells, but then weakly appears in later stages **(Figure 4d,e**). Fluorescent protein from the *His3Dendra2* HRU is undetectable throughout the testis, with signal only in the nuclei of somatic sheath cells (**Figure 4f,g**). We attribute the difference in pattern of expression between *His2ADendra2* and *His3Dendra2* transgenes to the deposition of histone H2A in post-mitotic cells, while histone H3 cannot [Jackson & Chalkley 1985, Kimura & Cook 2001]. This repression is sensitive to the demand for histones, because animals with reduced numbers of histone genes and the *His2ADendra2* reporter HRU now show intense fluorescence throughout the apical tip of the testis, and gonial cells (**Figure 4h**). Thus, we infer that germline cells – like somatic cells – upregulate histone gene expression when these genes are limiting.

Reduced histone H4 activates histone gene reporters

Previous work has implicated the H3K9-methyltransferase *Suppressor of variegation 3-9* (*Su(var)3-9*) in histone gene silencing [Ner et al 2002; Ito et al 2012], and indeed we find that the histone locus is enriched for H3K9 methylation (**Figure 2b**). We constructed viable null *Su(var)3-9* flies from transheterozygous point mutation alleles [Schotta et al 2002], however neither *His2ADendra2* nor *His3Dendra2* reporters are derepressed in this background (**Figure 4i**). To identify

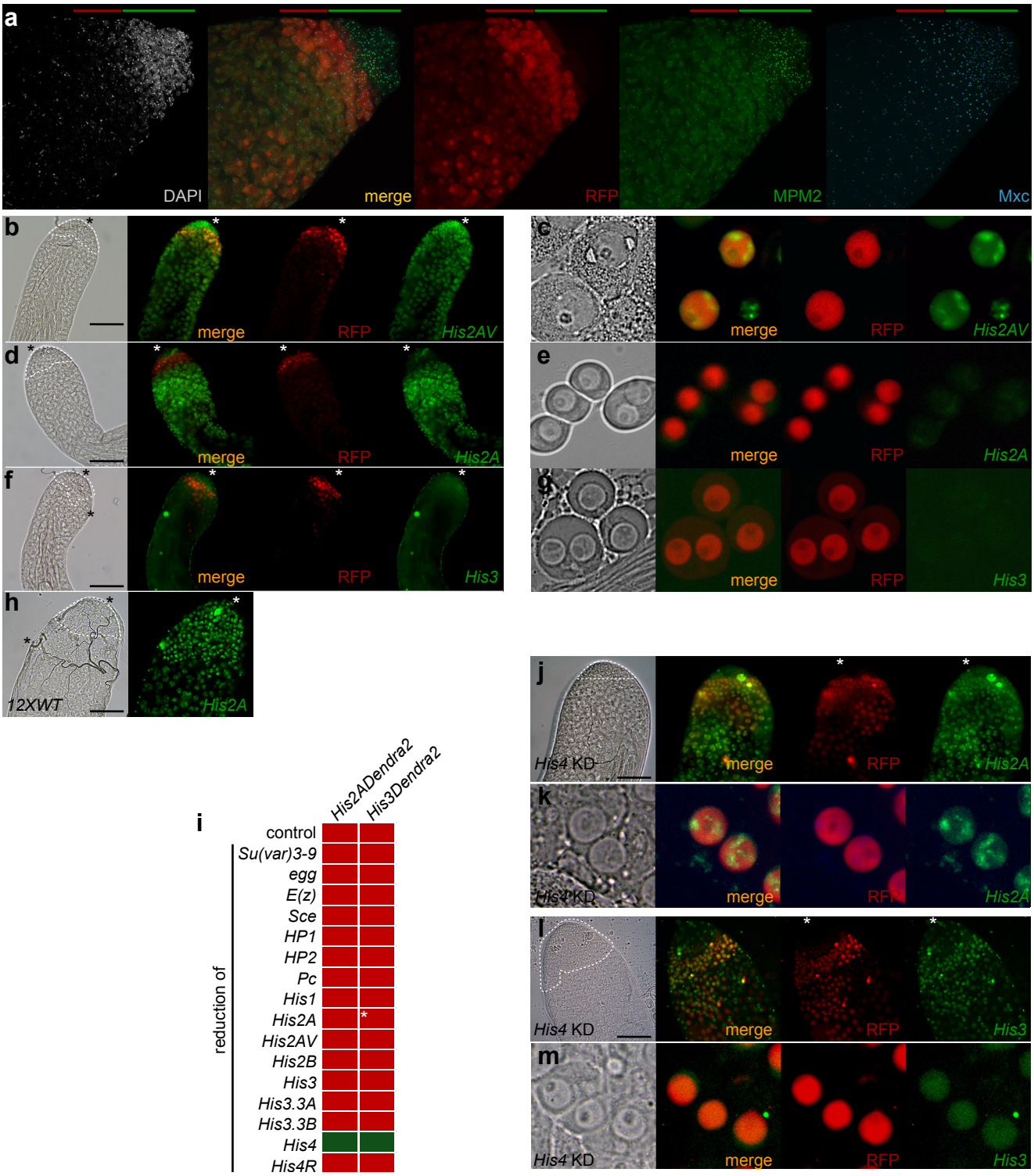


Figure 4. Silencing and derepression of extra histone genes in the Drosophila male germline. (a) The HLB in testis cells is marked by Mxc staining (blue). Cells in the proliferating zone at the apical tip of the testis are marked with phospho-Mxc detected by the MPM2 antibody at the HLB (green). Post-mitotic G2 phase germline cells are marked by *bamGAL4*-induced *UASRFP* expression (red). The proliferating zone of the testes is marked by the green bar, and the RFP-marked G2 gonial cells by the red bar. **(b-h)** Fluorescence of histone-Dendra2 reporter constructs (green) in live testes. The apical tip is marked with an asterisk, the proliferating zone is outlined with white dashed lines on phase contrast images, and G2 phase gonial cells are identified with RFP. (**b,c**) *His2AVDendra2* expression in wildtype males. Dendra2 fluorescence is apparent throughout the testis, including in the apical tip, and intensely stains the nuclei of isolated and squashed gonial cells, marked with *bamGAL4*-induced RFP expression. **(d,e)** *His2ADendra2* Histone Repeat Unit expression in wildtype males. Fluorescence is absent from the proliferating zone and from RFP-positive gonial cells but is present in later germline cells. **(f,g)** *His3Dendra2* Histone Repeat Unit expression in wildtype males. Staining is absent throughout the germline; the few fluorescent nuclei are in somatic cells of the testis sheath. **(h)** Fluorescence of the *His2ADendra2* HRU reporter in 12XWT males. This line does not carry *bamGAL4* and *UAS-RFP* constructs; the proliferating zone was identified by position in the testis (dashed line). Strong H2ADendra2 fluorescence is apparent throughout the apical tip of the testis and in later

(Figure 4 continued)
stages. **(i)** Results of tests for derepression of *His2ADendra2* and *His3Dendra2* Histone Repeat Unit reporters in testes with reductions in chromatin factors. *Su(var)3-9* was tested in *Su(var)3-9*^{1}/*Su(var)3-9*^{2} homozygotes, all other factors were tested by *bamGAL4*-induced knockdown in testes. Red squares represent repression similar to wildtype, green squares indicate derepression. **(j-m)** Derepression of *His2ADendra2* (j,k, green) or of *His3Dendra2* (l,m, green) in cells with *bamGAL4*-induced RFP expression (red) and *His4* knockdown (KD). Fluorescence is absent in the apical tip of the testis but appears in the post-mitotic stage where knockdown occurs.

mechanisms responsible for HRU repression, we targeted a selection of chromatin proteins for RNAi knockdown in the gonial cell stage. None of these knockdowns derepressed *His2ADendra2* or *His3Dendra2* reporters, including the histone H3 lysine-9 methyltransferase *eggless* (*egg* [Clough et al 2007]), the H3K9me2/3-binding proteins HP1 [James et al 1989] or HP2 [Shaffer et al 2002], the histone H2A ubiquitin ligase *Sex combs extra* (*Sce* [Gutierrez et al 2012]), the PRC1 component *Polycomb* (*Pc* [Lanzuolo & Orlando 2012]), or the histone H3 lysine-27 methyltransferase *Enhancer of zeste* (*E(z)* [Cao & Zhang 2004]). Thus, it appears that while silenced HRUs carry modifications associated with heterochromatic silencing, these modifications do not determine expression of the histone gene reporter; instead, the predominant mechanism of regulating the histone genes is in response to the demand for chromatin packaging in proliferating cells.

Since changes in histone gene number do alter reporter expression, we used RNAi to knockdown histones in gonial cells of the male germline. We tested knockdown of the linker histone gene *His1*, the core histone genes *His2A*, *His2B*, *His3*, and *His4*, of the histone variant genes *His2AV*, *His3.3A*, and *His3.3B*, as well as the orphan gene *His4R*. Testes for all of these genotypes contain sperm and males are fertile, implying that these knockdowns only partially reduce histone gene expression. The HRU reporters remained repressed in 8/9 of these knockdowns. Only knockdown of the *His4* gene resulted in dramatic derepression of both *His2ADendra2* and *His3Dendra2* in gonial cells (**Figure 4i-m**). This implies that cells measure the demand for histones based only on the H4 histone.

Histone H4 localizes to the HLB in Drosophila cells

It is surprising that knockdown of only one histone modulates HRU silencing, since histones associate in dimers and in octamers as nucleosomes are assembled, although some examples where monomeric histone are complexed with protein chaperones have been identified [Apta-Smith et al 2018], and instances where a singular histone is used to measure chromatin in both Drosophila and in human cells [Shindo & Amodeo 2021; Kobiyama et al 2010]. To test if histone H4 localizes to the HLB on its own, we examined the localization of different histones within the male germline using inducible GFP-tagged constructs [Schwartz & Ahmad 2005; **Methods**]. Induction of a tagged H3 or tagged H3.3 histones in gonial cells broadly labels the nuclei of these cells (**Figure 5a,b**). In contrast, tagged H4 histone shows a distinct subnuclear pattern, with one major dot in each nucleus (**Figure 5c**). This dot coincides with Mxc protein at the HLB in gonial cells (**Figure 5d**). Since the *bamGAL4*-induced H4GFP construct is not expressed in early stages, we stained testes with an antibody to histone H4. This revealed an H4 dot in cells throughout the earlier proliferating zone, including most cells actively undergoing DNA replication (**Figure 5e**) and interphase cells **(Figure 5f).** Only 7% (1/14) of prophase cells show the dot (**Figure 5g**), consistent with the partial disassembly of the HLB in mitosis [White et al 2011]. Thus, this histone is in the right place to directly affect histone gene expression both in S phase and in gap phase cells.

We examined Drosophila cultured *Kc167* cells to determine if histone H4 localizes to the HLB in other cell types. In immunostained samples, histone H4 staining colocalizes with Mxc at HLBs (**Figure 5h**). In chromatin profiling, histone H4 shows high signal only at the histone locus, coincident with Mxc, H3K27 acetylation, and RNAPII-S5p signal (**Figure 5i**). As *Kc167* cells are derived from somatic embryonic cells, this implies that non-nucleosomal histone H4 is a general component of the HLB.

Loss of histone H4 alters HLB activity

The HLB is enriched for the initiating RNAPII-S5p isoform in embryos and in the female germline [Kemp et al 2021, Cho et al 2022, Lu et al 2024]. In the testis, RNAPII-S5p is distributed through nuclei and forms a bright focal spot at the HLB in proliferating zone nuclei with bright phospho-Mxc staining, and slightly weaker focal staining in RFP-positive G2 phase gonial cells (**Figure 6a-c**). This is the major isoform of RNAPII engaged at histone genes, as staining for the RNAPII-S2p isoform is broadly distributed throughout nuclei with no greater enrichment at the HLB (**Figure 6d**). By the early primary spermatocyte stage the HLB is no longer enriched for any RNAPII isoform. We infer that active histone loci in the proliferating zone are engaged with high levels of the RNAPII-S5p isoform, and a reduced amount of this isoform persists in G2 gonial phase cells when histone genes are no longer expressed. Overall, the HLB in the testis progresses from having high levels of engaged RNAPII in the proliferating zone, to less engaged and non-transcribing RNAPII in G2 phase gonial cells, and to loss of RNAPII in primary spermatocytes. Dissolution of the HLB occurs in later stages, as Mxc staining is eventually lost, as cells do not need histone gene expression as they proceed to meiosis and sperm differentiation. This progression and the spatial arrangement of the testis is an easily tractable setting to follow changes in the HLB.

The *bamGAL4*-induced H4-GFP dot is intense in G2 phase gonial cells, and this coincides with reduced RNAPII-S5p staining in G2 gonial cells (**Figure 7a,b**), consistent with a role in limiting histone gene expression. We then examined testes where *His4* was knocked down in these cells. In

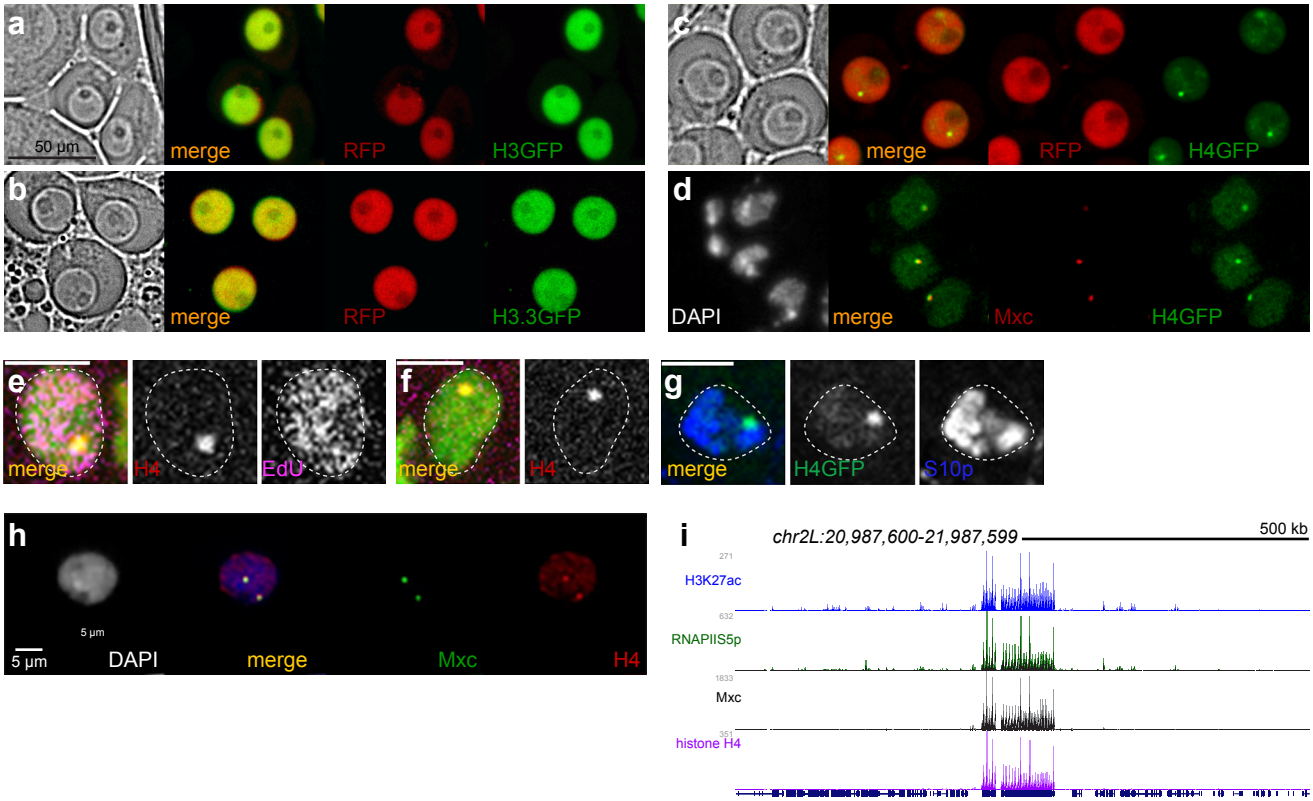


Figure 5. Histone H4 is a component of the Histone Locus Body in male germline cells. (**a-c**) Fresh squashes of testes with *bamGAL4*-induced expression of GFP-tagged histones (green) Induced RFP expression (red) marks G2 phase gonial cells. Histone H3-GFP **(a)** and histone H3.3-GFP **(b)** broadly label the nuclei of gonial cells. In contrast, induced histone H4-GFP **(c)** labels one bright dot with a low broad background in G2 phase nuclei. **(d)** The induced histone H4-GFP (green) dot coincides with the HLB, marked by Mxc staining (red). **(e,f)** Representative spermatogonial cells labeled with EdU (pink) to mark nuclei with ongoing DNA replication or in gap phase. 94% (47/50) S phase cells show focal staining of histone H4 (green) and the yellow HLB dot is also visible in gap phase cells. **(g)** Representative spermatogonial cell marked with the M phase epitope H3S10p (blue). The histone H4 dot is visible in only 7% (1/14) of mitotic cells. **(h)** *Kc167* nucleus stained with antibodies to histone H4 (red) and Mxc (green). Histone H4 is enriched in HLBs. **(i)** Browser tracks of histone modifications around the histone locus in *Kc167* cells. The histone locus is enriched for the active H3K27 acetylation modification (blue), the RNAPII-S5p isoform (green), Mxc (black), and for histone H4 (purple).

contrast to wildtype controls (**Figure 7c,e**), knock-down testes have elevated intensity of the RNAPII-S5p isoform in G2 gonial cells and increased staining of phospho-Mxc at HLBs (**Figure 7d,f**). These defects implicate histone H4 in the switch from active to inactive forms of the HLB.

Histone H4 localizes to active histone gene promoters in human cells

Since histone genes are repeated and we cannot distinguish between gene copies, we cannot define whether histone H4 binds all histone genes or only active or silent genes in Drosophila. But histone H4 is an ancient protein, and if it plays a role in limiting histone gene expression we expect HLB localization to be conserved across species. In the human genome the multiple copies of histone genes are not in a repeat array; instead they are comparatively separated and scattered across four clusters, termed *HIST1-4* [Dhahri et al 2024], and these aggregate into HLBs containing the transcription cofactor NPAT, the human homolog of Mxc [Ma et al 2000; Ghule et al 2008]. Indeed, immunostaining of human K562 cells reveals that NPAT and histone H4 colocalize at HLBs (**Figure 8a**). We then profiled the distribution of histone H4 and the active histone modification H3K27 acetylation in K562 cells and compared these to previously published profiling of NPAT [Kaya-Okur et al 2019] and RNAPII-S5p [Janssens et al 2021]. As expected, 64 canonical histone genes in the *HIST* on chromosome *6* are marked with both RNAPII and with H3K27mac modifications, identifying them as active genes (**Figure 8b-d**). Each of these active histone genes are also marked with NPAT and histone H4 (**Figure 8b**), and at high resolution these four chromatin features coincide at promoters (**Figure 8c**). In contrast, the 8 non-transcribed canonical histone genes lack both NPAT and histone H4 (**Figure 8b,d**), as do the 5 active histone variant genes that are not S phase-regulated (**Figure 8d**). Actively transcribed non-histone genes neighboring *HIST* clusters also lack NPAT and histone H4 signal (**Figure 8b**). These results implicate histone H4 in the S phase regulation of canonical histone gene promoters in human cells.

Discussion

The invention of eukaryotic chromatin required coordination of histone protein synthesis with DNA replication in S phase cells. DNA replication and histone synthesis must be coupled because even small imbalances are detrimental, given the large amounts of chromatin duplicated each S phase. Underproduction of histones will result in incomplete chromatin packaging, and this leads to exposure and damage

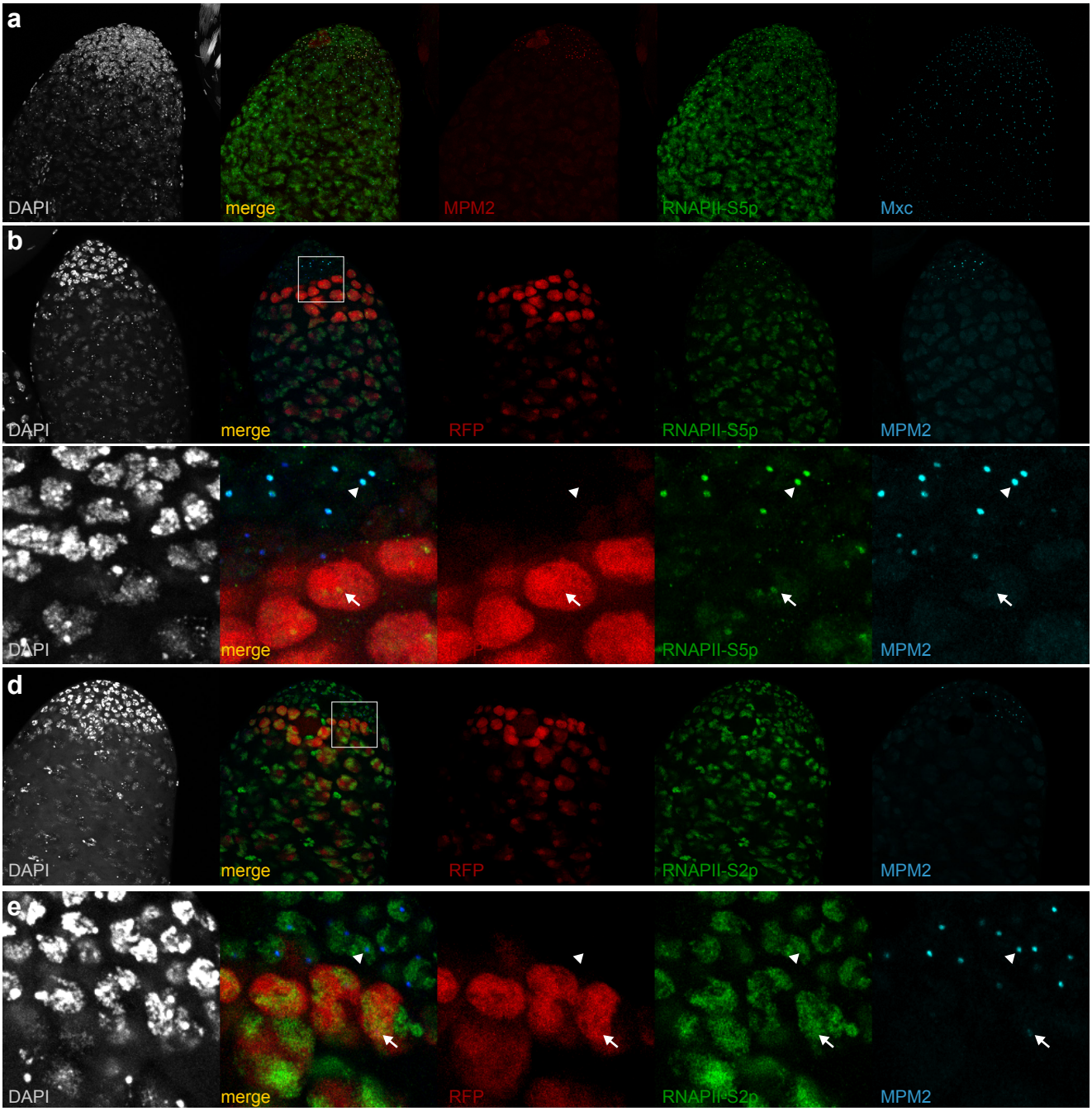


Figure 6. RNAPII isoforms at the HLB in the Drosophila testis. (a) RNAPII-S5p (green) stains the HLB in the proliferating zone marked by phospho-Mxc staining (red) and and the HLB in more distal cells. Mxc staining (blue) marks the HLB. **(b)** The proliferating zone is marked with phospho-Mxc (blue) and the G2 phase gonial cells by *bamGAL4*-induced RFP (red). RNAPII-S5p staining (green) strongly stains the HLB in the proliferating zone, and more weakly stains the HLB in G2 phase cells. **(c)** zoom of the boxed area in (b). The arrowhead points to an HLB in proliferating zone, and the arrow to an HLB in a G2 phase cell. **(d)** RNAPII-S2p staining (green) strongly stains nuclei in the proliferating zone and in RFP-labeled G2 phase cells, but is not enriched in HLBs. **(e)** zoom of the boxed area in (d). The arrowhead points to an HLB in proliferating zone, and the arrow to an HLB in a G2 phase cell.

of new DNA [Dreyer et al 2024]. Histone overproduction results in chromosome loss [Meeks-Wagner & Hartwell 1986] and is cytotoxic [Gunjan & Verreault 2003]. Feedback between these two processes provides just enough histones to package newly replicated DNA [Weintraub 1972; Duronio & Marzluff 2017]. Feedback control implies that S phase cells measure both ongoing DNA replication and the need for more histones. Histone production is modulated through two main controls: CDK2-catalyzed phosphorylation of Mxc/NPAT induce transcription canonical histone genes when cells commit to S phase [Armstrong & Spencer 2023], and cell cycle-regulated associations between histone mRNA processing factors stabilize transcripts in S phase [Marzluff et al 2008]; both these processes take place in HLBs [Romeo & Schümperli 2016]. S phase-induced histone mRNAs are the only protein coding transcripts that are not poly-adenylated in animals; instead, these transcripts have a terminal stem-loop structure that is bound by stem-loop binding protein (SLBP), directing 3' processing [Marzluff et al 2008]. Mathematical modeling suggested that feedback

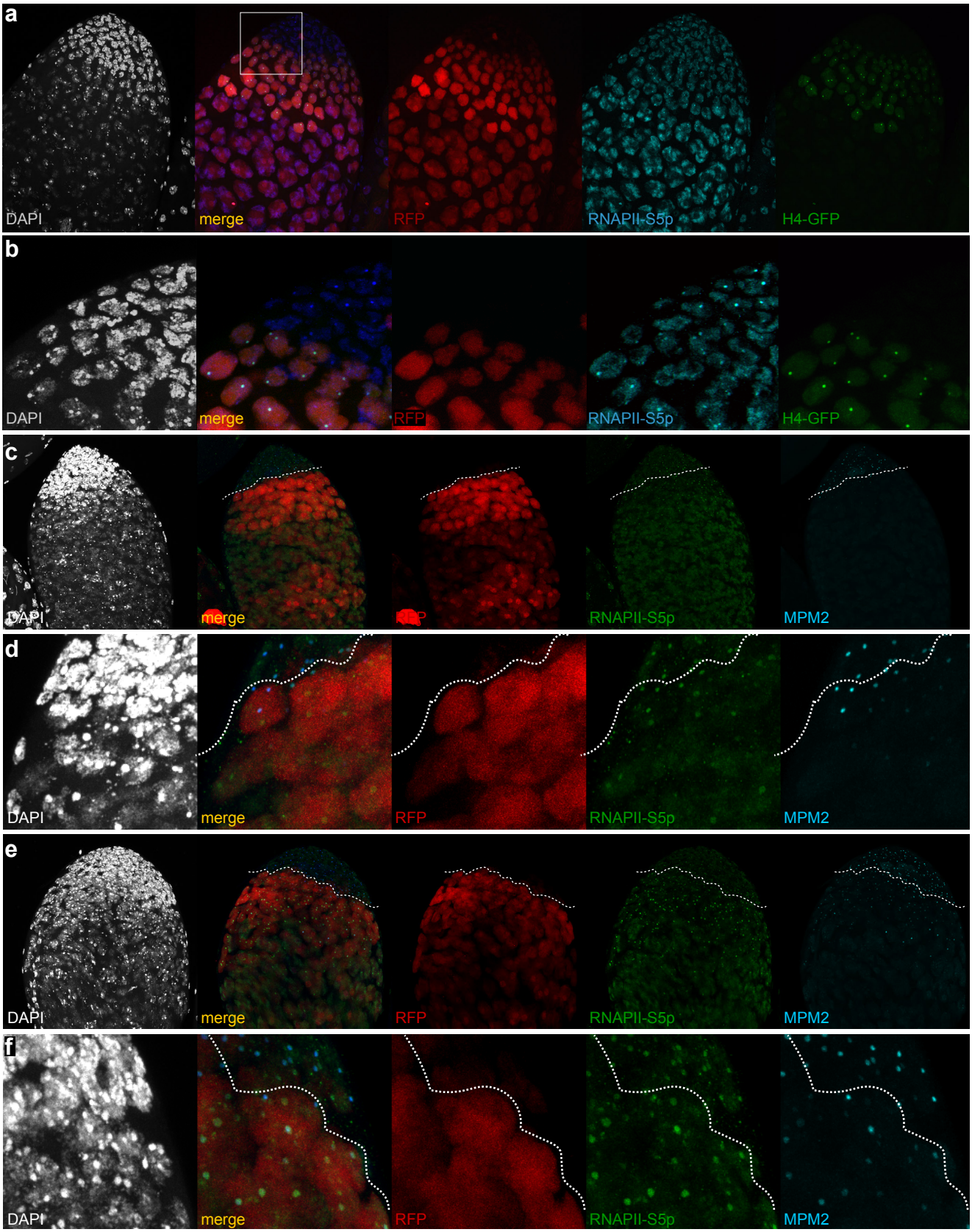


Figure 7. Histone H4 localizes with reduced RNAPII in HLBs. (a) Immunostaining for RNAPII-S5p (blue) in the apical tip of a testis with *bamGAL4*-induced expression of histone H4-GFP (green). G2 phase gonial cells are marked by induced RFP expression (red). The intense dot of H4-GFP coincides with focal RNAPII-S5p signal in G2 phase gonial cells. **(b)** zoom of the box marked in (a) showing costaining of HLBs with RNAPI-S5p and H4-GFP in RFP-positive G2 phase gonial cells. RNAPII staining in these cells is lower than that in more apical cells in the proliferating zone. **(c-f)** Testes with *bamGAL4*-induced RFP expression (red) marking G2 gonial cells and stained for phospho-Mxc (blue) and the RNAPII-S5p isoform (green). Dashed lines demarcate the proliferative zone from G2 phase cells based on RFP expression. A wildtype testis **(c,d)** with high phospho-Mxc in the proliferative zone and low staining in G2 phase cells. RNAPII-S5p signal in the proliferative zone is high at some HLBs and moderate at others, and moderate at

(Figure 7 continued) HLBs in the RFP-marked cells. **(e,f)** A testis with knockdown of *His4* (KD). High phospho-Mxc signal is apparent in the proliferative zone and persists in the RFP-labeled gonial cells. The RNAPII-S5p signal is more intense at the HLB in gonial cells than in cells of the proliferative zone.

from soluble histone pools is necessary for precise coupling between DNA replication and histone synthesis [Christopher et al 2016]. Our observations suggest a simple model where soluble histone H4 protein directly represses histone gene transcription. Ongoing DNA replication and chromatin packaging use up soluble histones, but once DNA replication ceases, soluble histones including histone H4 accumulate. Soluble histone H4 then localizes to the HLB and represses transcription. Although histone H4 localizes to histone gene promoters, repression might occur by modulating transcription or 3' processing since both processes occur within the HLB.

This appears to be the ancestral organization of S phase-induced histone genes throughout Eukaryota, because both stem-loop mRNA structures and SLBP homologs have been identified in protozoa at the base of the eukaryotic tree [Lopez & Samuelsson 2008]. The evolutionary origin of this system was unclear, but recently similarities to 3' processing of transcripts in bacteria were pointed out [Shine et al 2024]. Thus, the histone 3' processing system appears to be one of the few relics in eukaryotic genomes of their origin and may explain why these genes are sequestered in their own nuclear body. The eukaryotic histones themselves were derived from bacterial or viral proteins in the last eukaryotic common ancestor [Talbert et al 2022], with eukaryotic histone H4 being a sister lineage to both the HMfB archaeal histones and the doublet H4-H3 histones of the Nucleocytoviricota giant viruses [Grau-Bové et al 2022], that later diversified into the four core histone subtypes [Malik and Henikoff 2003]. Thus, it is conceivable that histone H4 has been used as a negative regulator of core histone gene expression all this time. Indeed, histone H4 is distinctive in that variants for this isotype rarely occur across eukaryotic evolution. One exception is a variant histone H4 encoded by some symbiotic bracoviruses; braconid wasps harboring this virus inject viral DNA into host moth larvae, where production of the variant histone suppresses host histone H4 mRNA production [Gad & Kim 2009]. This unusual variant appears to have weaponized the normal negative feedback loop of histone H4 on histone gene regulation for parasite life history.

Histone gene regulation has been implicated in cancer progression in human patients, and histone overproduction is predictive of cancer malignancy [Henikoff et al 2025]. There are multiple theories to explain the initiation of a cancer, invoking genetic mutation, changes in epigenetic marks, and defects in developmental signaling. Although the relative importance of these theories is now debated [Jassim et al 2023; Huang et al 2025], in any scenario progenitor cells must maintain a relatively undifferentiated state and proliferate. We have described that widespread chromosome arm aneuploidies are common in cancers and number of arm losses scales with malignancy [Zheng et al 2025]. Such aneuploidies in tumors were suggested to inhibit cellular differentiation and thereby trap anaplastic cells in a proliferative stage [Hansemann 1890; reviewed in Henikoff & Ahmad 2025]. We have proposed that these events are causally linked: that histone overexpression during S phase compromises histone variant-based centromere assembly and also accelerates cell proliferation; these effects would directly cause mitotic chromosome errors and result in aneuploidies [Zheng et al 2025; Henikoff & Ahmad 2025]. In support of this scenario, a very recent study identified reduced cell cycle duration as the only common feature of multiple distinct cancers, showing that tumorigenesis could be blocked by mutations affecting CDK2 activity [Chen et al 2025]. CDK2 is the kinase that phosphorylates Mxc/NPAT and histone gene activation, linking conserved HLB regulation described here to a deeper understanding of cancer. As such, detailing the mechanisms by which histone synthesis is normally restrained should provide insights into originating oncogenic events and opportunities for intervention.

Methods

Fly strains

All crosses were performed at 25˚C. All mutations and chromosomal rearrangements used here are described in Flybase (http://www.flybase.org). The w^{1118} strain was used as a wildtype control. The *12XWT* strain is *w ; DHis^{C} ; 12XWT* [MacKay et al 2015]. The HRU reporter *His3Dendra2* was previously described [Shindo & Amodeo 2019], and the *His2ADendra2* reporter was constructed similarly. Inducible histone lines were *UAS-H3-GFP* [Schwartz & Ahmad 2005], *UAS-H3.3-GFP* [Schwartz & Ahmad 2005], and the *UAS-H4-GFP* construct [Ahmad & Henikoff 2002] was injected into fly embryos for P element transformation [Rubin & Spradling 1982] by BestGene Inc. (Chino Hills CA). A similar UAS-H4-eGFP construct used here for some experiments was previously published [Wooten et al 2019]. Additional constructs used for cytological characterization were *y w P[bam-GAL4:VP16,w^{+}]1/Y ; P[UAS-RFP,w^{+}]2*. Inducible knockdown constructs and Bloomington Drosophila Stock Center IDs for histones and chromatin regulators are listed in the *Key Resources Table*.

Antibodies

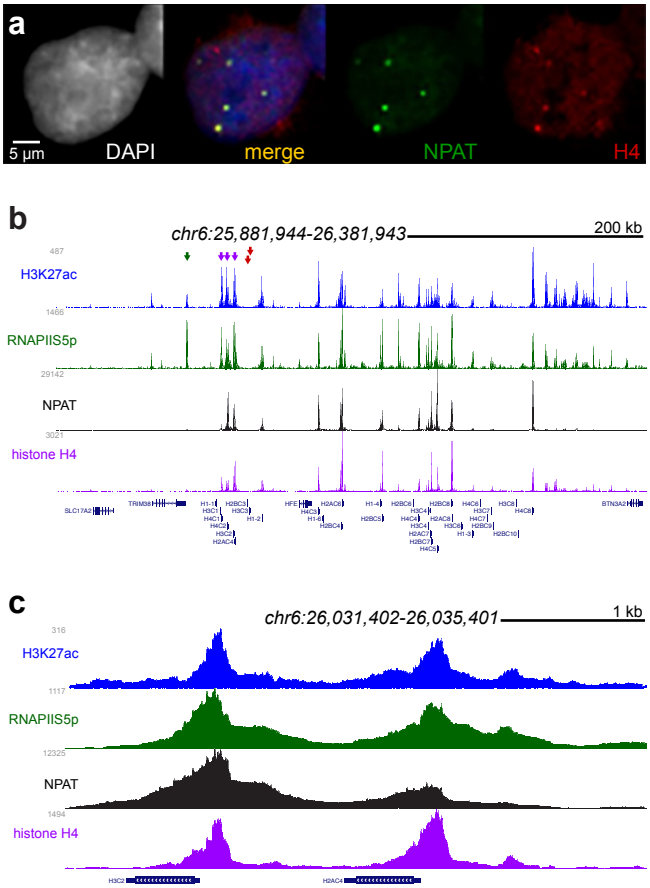
Antibodies used for CUT&Tag profiling and for immunocytology are listed in the *Key Resources Table*.

Imaging fresh tissues

Dissected tissues from larvae or adults were mounted in PBS on slide and imaged by epifluorescence on an EVOS FL Auto 2 inverted microscope (Thermo Fisher Scientific) with a 10X, 20X, or 40X objective. Pseudo-colored images were adjusted and composited in Adobe Photoshop and Adobe Illustrator.

Imaging immunostained testes

Testes from one-day old adult males were dissected in PBS, incubated in Accutase (Stemcell Technologies, #07920) for 10 minutes at room temperature to permeabilize the tissue,



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Isotype	Geneid	S5p	K27ac	NPAT	H4	location
H1	H1-4					chr6-HIST1
H1	H1-2					chr6-HIST1
H1	H1-3					chr6-HIST1
H1	H1-5					chr6-HIST1
H1	H1-6					chr6-HIST1
H1	H1-1					chr6-HIST1
H1	H1-10					chr3
H1	H1-8					chr3
H1	H1-0					chr22
H1	H1-7					chr12-HIST4
H2A	H2AC11					chr6-HIST1
H2A	H2AC12					chr6-HIST1
H2A	H2AC8					chr6-HIST1
H2A	H2AW					chr1-HIST3
H2A	H2AC20					chr1-HIST2
H2A	H2AC21					chr1-HIST2
H2A	H2AC6					chr6-HIST1
H2A	H2AC13					chr6-HIST1
H2A	H2AC18					chr1-HIST2
H2A	H2AC19					chr1-HIST2
H2A	H2AC17					chr6-HIST1
H2A	H2AC7					chr6-HIST1
H2A	H2AC4					chr6-HIST1
H2A	H2AC15					chr6-HIST1
H2A	H2AC16					chr6-HIST1
H2A	H2AC14					chr6-HIST1
H2A	H2AC1					chr6-HIST1
H2A	H2AJ					chr12-HIST4
H2A	H2AZ2					chr7
H2A	H2AX					chr11
H2A	H2AZ1					chr4
H2A	H2AB2					chrX
H2A	H2AL3					chrX
H2A	H2AB1					chrX
H2A	H2AP					chrX
H2A	H2AB3					chrX
H2B	H2BC4					chr6-HIST1
H2B	H2BC12					chr6-HIST1
H2B	H2BC11					chr6-HIST1
H2B	H2BC5					chr6-HIST1
H2B	H2BC18					chr1-HIST2
H2B	H2BC21					chr1-HIST2
H2B	H2BC8					chr6-HIST1
H2B	H2BU1					chr1-HIST3
H2B	H2BC17					chr6-HIST1
H2B	H2BC13					chr6-HIST1
H2B	H2BC7					chr6-HIST1
H2B	H2BC15					chr6-HIST1
H2B	H2BC9					chr6-HIST1
H2B	H2BC14					chr6-HIST1
H2B	H2BC1					chr6-HIST1
H2B	H2BC10					chr6-HIST1
H2B	H2BC6					chr6-HIST1
H2B	H2BC3					chr6-HIST1
H2B	H2BN1					chr17
H2B	H2BK1					chr7
H2B	H2BC12L					chr21
H2B	H2BW1					chrX
H2B	H2BW2					chrX
H3	H3C4					chr6-HIST1
H3	H3C2					chr6-HIST1
H3	H3C15					chr1-HIST2
H3	H3C14					chr1-HIST2
H3	H3C12					chr6-HIST1
H3	H3C10					chr6-HIST1
H3	H3C13					chr1-HIST2
H3	H3C7					chr6-HIST1
H3	H3-7					chr1-HIST2
H3	H3C1					chr6-HIST1
H3	H3C6					chr6-HIST1
H3	H3C8					chr6-HIST1
H3	H3C11					chr6-HIST1
H3	H3C3					chr6-HIST1
H3	H3-4					chr1-HIST3
H3	H3-3B					chr17
H3	H3-3A					chr1
H3	H3-5					chr12
H3	H3Y2					chr5
H3	H3Y1					chr5
H4	H4C15					chr1-HIST2
H4	H4C3					chr6-HIST1
H4	H4C4					chr6-HIST1
H4	H4C8					chr6-HIST1
H4	H4C14					chr1-HIST2
H4	H4C5					chr6-HIST1
H4	H4C1					chr6-HIST1
H4	H4C2					chr6-HIST1
H4	H4C9					chr6-HIST1
H4	H4C12					chr6-HIST1
H4	H4C11					chr6-HIST1
H4	H4C7					chr6-HIST1
H4	H4C6					chr6-HIST1
H4	H4C13					chr6-HIST1
H4	H4-16					chr12-HIST4

Figure 8. Histone H4 localizes to the promoters of active histone genes in human K562 cells. (a) Immunostaining human K562 cells for the HLB factor NPAT (green) and histone H4 (red). **(b)** Distribution of histone H3K27acetylation, RNAPII-Sp5, NPAT, and histone H4 across a portion of the HIST1 histone cluster on chromosome *6*. Purple arrowheads mark three active canonical histone genes on one side of this cluster, the red arrowheads marks two inactive histone genes, and the green arrowhead marks the promoter of an adjacent active non-histone gene. NPAT and histone H4 coincide only at active histone genes, and are absent from inactive histone genes and from active non-histone genes. **(c)** Zoom showing coincidence of the H3K27 acetylation mark, RNAPII-S5p, NPAT, and histone H4 at the promoters of two active histone genes. **(d)** Summary of summed signal for chromatin features at the 94 histone genes in the human genome. Variant histone genes are labeled in orange. Localization of anti-histone H4 signal to the multiple HLBs in a K562 nucleus. Each histone isotype and variant is ordered by expression (RNAPIIS5p signal).

fixed in 4% formaldehyde/PBS with 0.1% Triton-X100 (PBST) for 10 minutes, incubated twice in 0.3% sodium deoxycholate/PBST for 10 minutes each [Lim & Fuller 2012], incubated with primary antibodies in A+t buffer at 4˚C overnight, and then with fluorescently-labeled secondary antibodies (1:200 dilution, Jackson ImmunoResearch). Testes were stained with 0.5 µg/mL DAPI/PBS and mounted in 80% glycerol on slides, and imaged on a Stellaris 8 confocal microscope (Leica) with 20X or 63X objectives. Pseudo-colored images were adjusted and composited in ImageJ, Adobe Photoshop, and Adobe Illustrator.

Imaging tissue culture cells

Drosophila *Kc167* and human K562 cells were swelled with a hypotonic 0.5% sodium citrate solution, then smashed onto glass slides in a Cytospin 4 centrifuge (Thermo). Slides were fixed with 4% formaldehyde/PBST and incubated with primary antisera in A+t buffer, then with fluorescently-labeled secondary antibodies (1:200 dilution, Jackson ImmunoResearch), stained with 0.5 µg/mL DAPI/PBS, mounted in 80% glycerol, and imaged by epifluorescence on an EVOS FL Auto 2 inverted microscope (Thermo Fisher Scientific) with a 40X objective. Pseudo-colored images were adjusted and composited in Adobe Photoshop and Adobe Illustrator.

CUT&Tag chromatin profiling

To perform CUT&Tag [Kaya-Okur et al 2019], we dissected 20 imaginal wing discs from male 3rd instar larvae in PBS buffer, and transferred them to a tube containing Accutase (Stemcell Technologies, #07920) at 25˚C for 30 minutes. We then added an equal volume of 30% BSA to block proteases, and ran the material through a 30 ½ gauge needle once to dissociate tissue. Tissue suspensions were divided between 4-8 reaction tubes, and bound with BioMag Plus ConA (Bangs Inc., #531) magnetic beads. Tissue culture samples in media were added directly to ConA beads for binding, and then lightly fixed onto beads with 0.1% formaldehyde at room temperature for 1'. All samples were incubated with the following CUT&Tag solutions sequentially: primary antibodies diluted in Wash+ buffer (20 mM HEPES pH 7.5, 150 mM NaCl, 0.5 mM spermidine, 0.05% triton-X100, 2 mM EDTA, 1% BSA, with Roche cOmplete protease inhibitor) overnight at 4˚, secondary antibodies (in Wash+ buffer) for 1 hour at room temperature, and then incubated with pAGTn5 (Epicypher 15-1017) in 300Wash+ buffer (20 mM HEPES pH 7.5, 300 mM NaCl, 0.5 mM spermidine, 0.05% triton-X100 with cOmplete protease inhibitor) for 1 hour. After one wash with 300Wash+ buffer, samples were incubated in 300Wash+ buffer supplemented with 10 mM $MgCl_2$ for 1 hour at 25˚C to tagment chromatin. Tissue culture samples were tagmented in CUTAC buffer (10 mM TAPS pH 8.5, 20% DMF) supplemented with 5 mM $MgCl_2$ to enhance tagmentation efficiency. Samples were washed with 10 mM TAPS pH 8.5 and DNA was released with 0.1% SDS, 0.012 U/µL thermolabile protease K (NEB P8111S) in 10 mM TAPS at 37˚ and inactivated at 55˚C. Libraries were enriched with 14 cycles of PCR as described [Kaya-Okur et al 2019], and sequenced in dual-indexed paired-end mode (PE50) on the Illumina NextSeq 2000 platform at the Fred Hutchinson Cancer Center Genomics Shared Resource. Paired-end reads were mapped to this assembly using Bowtie2 using parameters, e.g.: --end-to-end --very-sensitive --no-mixed --no-discordant -q --phred33 -I 10 -X 700.

Gene score tables

To summarize the enrichment of profiling features across histones, we counted mapped reads from the start to the end of each gene in .bam files using subreads/feature_counts with option '_o'. Counts for each replication-coupled histone were summed, and counts for all genes were scaled by total mapped reads to give Counts per Million (CPM) reads. These values are provided in **Supplementary Table 2**.

Genomic display

Files of mapped reads were converted to genome coverage with bedtools/bamcoverage [Quinlan & Hall 2010] and displayed in the UCSC genome browser [Kent et al 2002]. Selected regions were exported as PDFs and formatted with Adobe Illustrator.

Data availability

Sequencing data have been deposited in GEO under accession code GSE280833. Data for NPAT profiling in K562 cells (SH_Hs_NPA1_20190217, SH_Hs_NPA2_20190217, SH_Hs_NPA4_20190217, SH_Hs_NPA8_20190217, SH_Hs_NPB1_20190217, SH_Hs_NPB2_20190217, SH_Hs_NPB4_20190217, and SH_Hs_NPB8_20190217 was previously published in Kaya-Okur et al 2019, and were merged here into one file (SH_Hs_NPB4_20190217). Data for RNAPII-S5p profiling in K562 cells (SH_Hs_K5xlin_PolS5P_3cy_0320, SH_Hs_K5xlin_PolS5P_6cy, SH_Hs_K5xlin_PolS5P_9cy_0320, SH_Hs_K5xlin_PolS5P_12cy_0320, and SH_Hs_K5xlin_PolS5P_20k_0320 were previously published in Janssens et al 2021, and were merged here in one file (SH_Hs_K5xlin_PolS5P_20k_0320).

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Author contributions

K.A. and S.H. conceived the study. K.A., M.W., B.N.T., and V.V. performed the experiments. K.A., S.H., and V.V. performed data analysis. K.A. wrote the manuscript. K.A., S.H., and X.C. reviewed and edited the manuscript. All authors approved the manuscript.

Competing interests

S.H. and K.A. have filed patent applications on related work.

References

Ahmad, K. and S. Henikoff (2002). "The histone variant H3.3 marks active chromatin by replication-independent nucleosome assembly." Mol Cell9(6):1191-200.

Amatori S, Tavolaro S, Gambardella S and Fanelli M. (2021). The dark side of histones: genomic organization and role of oncohistones in cancer. Clin Epigenetics 13(1):71. PMID: 33827674

Apta-Smith, M. J., J. R. Hernandez-Fernaud and A. J. Bowman (2018). “Evidence for the nuclear import of histones H3.1 and H4 as monomers.” Embo j 37(19). PMC6166134

Armstrong, C., Passanisi, V.J., Ashraf, H.M., and Spencer, S.L. (2023). “Cyclin E/CDK2 and feedback from soluble histone protein regulate the S phase burst of histone biosynthesis.” Cell Rep. 42(7):112768. PMID: 37428633

Barbour, H., S. Daou, M. Hendzel and E. B. Affar (2020). “Polycomb group-mediated histone H2A monoubiquitination in epigenome regulation and nuclear processes.” Nat Commun 11(1): 5947. PMC7683540

Barth, T. K. and A. Imhof (2010). “Fast signals and slow marks: the dynamics of histone modifications.” Trends Biochem Sci 35(11): 618-626.

Bonaccorsi, S., C. Pisano, F. Puoti and M. Gatti (1988). “Y chromosome loops in Drosophila melanogaster.” Genetics 120(4): 1015-1034. PMC1203565

Bulchand, S., S. D. Menon, S. E. George and W. Chia (2010). “Muscle wasted: a novel component of the Drosophila histone locus body required for muscle integrity.” J Cell Sci 123(Pt 16): 2697-2707.

Cao, R., L. Wang, H. Wang, L. Xia, H. Erdjument-Bromage, P. Tempst, R. S. Jones and Y. Zhang (2002). “Role of histone H3 lysine 27 methylation in Polycomb-group silencing.” Science 298(5595): 1039-1043.

Cao, R. and Y. Zhang (2004). “The functions of E(Z)/EZH2-mediated methylation of lysine 27 in histone H3.” Curr Opin Genet Dev 14(2): 155-164.

Chaubal, A., Waldern, J.M., Taylor, C., Laederach, A., Marzluff, W.F., and Duronio, R.J. (2023). “Coordinated expression of replication-dependent histone genes from multiple loci promotes histone homeostasis in Drosophila.” Mol Biol Cell 34(12):ar118. PMID: 37647143

Chen, D., Lu, S., Huang, K., Pearson, J.D., Pacal, M., Peidis, P., McCurdy, S., Yu, T., Sangwan, M., Nguyen, A., Monnier, P.P., Schramek, D., Zhu, L., Santamaria, D., Barbacid, M., Akeno, N., Wikenheiser-Brokamp, K.A., and Bremner, R. (2025). “Cell cycle duration determines oncogenic transformation capacity.” Nature (*Online ahead of print*). PMID: 40307557

Cho, C.Y., Kemp, J.P. Jr., Duronio, R.J., and O’Farrell, P.H. (2022). “Coordinating transcription and replication to mitigate their conflicts in early Drosophila embryos.” Cell Rep. 41(3):111507. PMID: 36261005

Christopher, A., Hameister, H., Corrigall, H., Ebenhöh, O., Müller, B., and Ullner, E. (2016). “Modelling Robust Feedback Control Mechanisms That Ensure Reliable Coordination of Histone Gene Expression with DNA Replication.” PLoS One 11(10):e0165848. PMID: 27798685

Clough, E., W. Moon, S. Wang, K. Smith and T. Hazelrigg (2007). “Histone methylation is required for oogenesis in Drosophila.” Development 134(1): 157-165.

Crain, A. T., M. Nevil, M. P. Leatham-Jensen, K. B. Reeves, A. G. Matera, D. J. McKay and R. J. Duronio (2024). “Redesigning the Drosophila histone gene cluster: an improved genetic platform for spatiotemporal manipulation of histone function.” Genetics 228(1). PMC11373521

Dávila López M, Samuelsson T. (2008). Early evolution of histone mRNA 3’ end processing. RNA 14(1):1-10. PMID: 17998288

Davis, F. M., T. Y. Tsao, S. K. Fowler and P. N. Rao (1983). “Monoclonal antibodies to mitotic cells.” Proc Natl Acad Sci U S A 80(10): 2926-2930. PMC393946

Dhahri, H., Saintilnord, W.N., Chandler, D., and Fondufe-Mittendorf, Y.N. (2024). “Beyond the Usual Suspects: Examining the Role of Understudied Histone Variants in Breast Cancer.” Int J Mol Sci. 25(12):6788. PMID: 38928493

Dreyer, J., Ricci, G., van den Berg, J., Bhardwaj, V., Funk, J., Armstrong, C., van Batenburg, V., Sine, C., VanInsberghe, M.A., Tjeerdsma, R.B., Marsman, R., Mandemaker, I.K., di Sanzo, S., Costantini, J., Manzo, S.G., Biran, A., Burny, C., van Vugt, M.A.T.M., Völker-Albert, M., Groth, A., Spencer, S.L., van Oudenaarden, A., and Mattiroli, F. (2024). “Acute multi-level response to defective de novo chromatin assembly in S-phase.” Mol Cell 84(24):4711-4728.e10. PMID: 39536749

Duronio, R. J. and W. F. Marzluff (2017). “Coordinating cell cycle-regulated histone gene expression through assembly and function of the Histone Locus Body.” RNA Biol 14(6): 726-738. PMC5519241

Ebert, A., S. Lein, G. Schotta and G. Reuter (2006). “Histone modification and the control of heterochromatic gene silencing in Drosophila.” Chromosome Res 14(4): 377-392.

Elgin, S. C. (1996). “Heterochromatin and gene regulation in Drosophila.” Curr Opin Genet Dev 6(2): 193-202.

Gad W and Kim Y. (2009). N-terminal tail of a viral histone H4 encoded in Cotesia plutellae bracovirus is essential to suppress gene expression of host histone H4. Insect Mol Biol. 18(1):111-8. PMID: 19196351

Ghule, P.N., Dominski, Z., Yang, X.C., Marzluff, W.F., Becker, K.A., Harper, J.W., Lian, J.B., Stein, J.L., van Wijnen, A.J., and Stein, G.S. (2008). “Staged assembly of histone gene expression machinery at subnuclear foci in the abbreviated cell cycle of human embryonic stem cells.” Proc Natl Acad Sci U S A. 105(44):16964-9. PMID: 18957539

Grau-Bové X, Navarrete C, Chiva C, Pribasnig T, Antó M, Torruella G, Galindo LJ, Lang BF, Moreira D, López-Garcia P, Ruiz-Trillo I, Schleper C, Sabidó E, and Sebé-Pedrós A. (2022). “A phylogenetic and proteomic reconstruction of eukaryotic chromatin evolution.” Nat Ecol Evol. 6(7):1007-1023. PMID: 35680998

Grummt, I. and C. S. Pikaard (2003). “Epigenetic silencing of RNA polymerase I transcription.” Nat Rev Mol Cell Biol 4(8): 641-649.

Günesdogan, U., H. Jäckle and A. Herzig (2010). “A genetic system to assess in vivo the functions of histones and histone modifications in higher eukaryotes.” EMBO Rep 11(10): 772-776. PMC2948182

Gunjan, A., & Verreault, A. (2003). “A Rad53 kinase-dependent surveillance mechanism that regulates histone protein levels in S. cerevisiae.” Cell 115(5):537-49. PMID: 14651846

Gutiérrez, L., K. Oktaba, J. C. Scheuermann, M. C. Gambetta, N. Ly-Hartig and J. Müller (2012). “The role of the histone H2A ubiquitinase Sce in Polycomb repression.” Development 139(1): 117-127. PMC3253035

Hansemann, D. (1890). Ueber asymmetrische Zelltheilung in Epithelkrebsen und deren biologische Bedeutung. Arch. für Pathol. Anat. Physiol. für Klin. Med. 119, 299–326. doi:10.1007/bf01882039

Henikoff, S., Zheng, Y., Paranal, R.M., Xu, Y., Greene, J.E., Henikoff, J.G., Russell, Z.R., Szulzewsky, F., Thirimanne, H.N., Kugel, S., Holland, E.C., and Ahmad, K. (2025). “RNA polymerase II at histone genes predicts outcome in human cancer.” Science 387(6735):737-743. PMID: 39946483

Henikoff, S. & Ahmad, K. (2025). “Hansemann’s anaplastic theory of cancer after 135 years.” Frontiers in Epigenetics and Epigenomics (*in press*). doi:10.3389/freae.2025.1607433

Hodkinson, L.J., Gross, J., Schmidt, C.A., Diaz-Saldana, P.P., Aoki, T., and Rieder, L.E. (2024). “Sequence reliance of the Drosophila context-dependent transcription factor CLAMP.” Genetics 227(3):iyae060. PMID: 38775472

Huang, S.-K., P. H. Whitney, S. Dutta, S. Y. Shvartsman, C. A. Rushlow (2021). “Spatial organization of transcribing loci during early genome activation in Drosophila.” Current Biology 31(22):5102-5110. PMC8612988

Huang, S., Soto, A.M., and Sonnenschein, C. (2025). “The end of the genetic paradigm of cancer.” PLoS Biol. 23(3):e3003052. PMID: 40100793

Hur W, Kemp JP Jr, Tarzia M, Deneke VE, Marzluff WF, Duronio RJ and Di Talia S. (2020). CDK-Regulated Phase Separation Seeded by Histone Genes Ensures Precise Growth and Function of Histone Locus Bodies. Dev Cell 54(3):379-394. PMID: 32579968

Ito, S., S. Fujiyama-Nakamura, S. Kimura, J. Lim, Y. Kamoshida, Y. Shiozaki-Sato, S. Sawatsubashi, E. Suzuki, M. Tanabe, T. Ueda, T. Murata, H. Kato, F. Ohtake, R. Fujiki, T. Miki, A. Kouzmenko, K. Takeyama and S. Kato (2012). “Epigenetic silencing of core histone genes by HERS in Drosophila.” Mol Cell 45(4): 494-504.

Jackson, V. & Chalkley, R. (1985). “Histone synthesis and deposition in the G1 and S phases of hepatoma tissue culture cells.” Biochemistry 24(24):6921-30. PMID: 3935167

James, T. C., J. C. Eissenberg, C. Craig, V. Dietrich, A. Hobson and S. C. Elgin (1989). “Distribution patterns of HP1, a heterochromatin-associated nonhistone chromosomal protein of Drosophila.” Eur J Cell Biol 50(1): 170-180.

Janssens, D.H., Meers, M.P. Wu, S.J., Babaeva, E., Meshinchi, S., Sarthy, J.F., Ahmad, K., and Henikoff, S. (2021). “Automated CUT&Tag profiling of chromatin heterogeneity in mixed-lineage leukemia.” Nat Genet. 53(11):1586-1596. PMID: 34663924

Jassim, A., Rahrmann, E.P., Simons, B.D., and Gilbertson, R.J. (2023). “Cancers make their own luck: theories of cancer origins.” Nat Rev Cancer 23(10):710-724. PMID: 37488363

Kaya-Okur, H. S., S. J. Wu, C. A. Codomo, E. S. Pledger, T. D. Bryson, J. G. Henikoff, K. Ahmad and S. Henikoff (2019). “CUT&Tag for efficient epigenomic profiling of small samples and single cells.” Nat Commun 10(1): 1930. PMC6488672

Kemp, J.P. Jr., Yang, X.C., Dominski, Z., Marzluff, W.F., and Duronio, R.J. (2021). “Superresolution light microscopy of the Drosophila histone locus body reveals a core-shell organization associated with expression of replication-dependent histone genes.” Mol Biol Cell 32(9):942-955. PMID: 33788585

Kent, W. J., C. W. Sugnet, T. S. Furey, K. M. Roskin, T. H. Pringle, A. M. Zahler and D. Haussler (2002). “The human genome browser at UCSC.” Genome Res 12(6): 996-1006. PMC186604

Kimura, H. & Cook, P.R. (2001). “Kinetics of core histones in living human cells: little exchange of H3 and H4 and some rapid exchange of H2B.” J Cell Biol. 153(7):1341-53. PMID: 11425866

Kobiyama, K., Takeshita, F., Jounai, N., Sakaue-Sawano, A., Miyawaki, A., Ishii, K.J., Kawai, T., Sasaki, S., Hirano, H., Ishii, N., Okuda, K., and Suzuki, K. (2009). “Extrachromosomal histone H2B mediates innate antiviral immune responses induced by intracellular double-stranded DNA.” J Virol. 84(2):822-32. PMID: 19906922

Lanzuolo, C. and V. Orlando (2012). “Memories from the polycomb group proteins.” Annu Rev Genet 46: 561-589.

Lim, J. G. and M. T. Fuller (2012). “Somatic cell lineage is required for differentiation and not maintenance of germline stem cells in Drosophila testes.” Proc Natl Acad Sci U S A

109(45): 18477-18481. PMC3494938

Liu, J. L., C. Murphy, M. Buszczak, S. Clatterbuck, R. Goodman and J. G. Gall (2006). “The Drosophila melanogaster Cajal body.” J Cell Biol 172(6): 875-884. PMC2063731

Lu, F., B. J. Park, R. Fujiwara, J. E. Wilusz, D. S. Gilmour, R. Lehmann and T. Lionnet (2024). “Integrator-mediated clustering of poised RNA polymerase II synchronizes histone transcription.” bioRxiv. PMC10592978

Ma, T., B. A. Van Tine, Y. Wei, M. D. Garrett, D. Nelson, P. D. Adams, J. Wang, J. Qin, L. T. Chow, and J. W. Harper (2000). “Cell cycle-regulated phosphorylation of p220(NPAT) by cyclin E/Cdk2 in Cajal bodies promotes histone gene transcription.” Genes & Dev 14(18): 2298-2313. PMC316935

MacAlpine, D. M. and G. Almouzni (2013). “Chromatin and DNA replication.” Cold Spring Harb Perspect Biol 5(8): a010207. PMC3721285

Malik, H.S. & Henikoff, S. (2003). “Phylogenomics of the nucleosome.” Nat Struct Biol. 10(11):882-91. PMID: 14583738

Marzluff, W.F., Wagner, E.J., and Duronio, R.J. (2008). “Metabolism and regulation of canonical histone mRNAs: life without a poly(A) tail.” Nat Rev Genet. 9(11):843-54. PMID: 18927579

McKay, D. J., S. Klusza, T. J. Penke, M. P. Meers, K. P. Curry, S. L. McDaniel, P. Y. Malek, S. W. Cooper, D. C. Tatomer, J. D. Lieb, B. D. Strahl, R. J. Duronio and A. G. Matera (2015). “Interrogating the function of metazoan histones using engineered gene clusters.” Dev Cell 32(3): 373-386. PMC4385256

Ner, S. S., M. J. Harrington and T. A. Grigliatti (2002). “A role for the Drosophila SU(VAR)3-9 protein in chromatin organization at the histone gene cluster and in suppression of position-effect variegation.” Genetics 162(4): 1763-1774. PMC1462387

Okazaki, R., K. Yamazoe and Y. H. Inoue (2020). “Nuclear Export of Cyclin B Mediated by the Nup62 Complex Is Required for Meiotic Initiation in Drosophila Males.” Cells 9(2). PMC7072204

Quinlan, A. R. and I. M. Hall (2010). “BEDTools: a flexible suite of utilities for comparing genomic features.” Bioinformatics 26(6): 841-842. PMC2832824

Ragusa, D. & Vagnarelli, P. (2023). “Contribution of histone variants to aneuploidy: a cancer perspective.” Front Genet. 14:1290903. PMID: 38075697 histone overexpression in cancers

Ray, J. and K.A. Maggert (2021). “Symmetric inheritance of histones H3 in Drosophila male germline stem cell divisions.” bioRxiv doi.org/10.1101/2021.10.22.465494

Romeo, V. & Schümperli, D. (2016). “Cycling in the nucleus: regulation of RNA 3’ processing and nuclear organization of replication-dependent histone genes.” Curr Opin Cell Biol. 40:23-31. PMID: 26895140

Rubin, G. M. and A. C. Spradling (1982). “Genetic transformation of Drosophila with transposable element vectors.” Science 218(4570):348-53.

Salzler, H. R., D. C. Tatomer, P. Y. Malek, S. L. McDaniel, A. N. Orlando, W. F. Marzluff and R. J. Duronio (2013). “A sequence in the Drosophila H3-H4 Promoter triggers histone locus body assembly and biosynthesis of replication-coupled histone mRNAs.” Dev Cell 24(6): 623-634. PMC3613982

Schotta, G., A. Ebert, V. Krauss, A. Fischer, J. Hoffmann, S. Rea, T. Jenuwein, R. Dorn and G. Reuter (2002). “Central role of Drosophila SU(VAR)3-9 in histone H3-K9 methylation and heterochromatic gene silencing.” Embo j 21(5): 1121-1131. PMC125909

Schwartz, B. E. and K. Ahmad (2005). “Transcriptional activation triggers deposition and removal of the histone variant H3.3.” Genes Dev 19(7): 804-814. PMC1074318

Schwartz, S., E. Meshorer and G. Ast (2009). “Chromatin organization marks exon-intron structure.” Nat Struct Mol Biol 16(9): 990-995.

Shaffer, C. D., G. E. Stephens, B. A. Thompson, L. Funches, J. A. Bernat, C. A. Craig and S. C. Elgin (2002). “Heterochromatin protein 2 (HP2), a partner of HP1 in Drosophila heterochromatin.” Proc Natl Acad Sci U S A 99(22): 14332-14337. PMC137884

Shindo, Y. and Amodeo, A.A. (2019). “Dynamics of Free and Chromatin-Bound Histone H3 during Early Embryogenesis.” Curr Biol 29(2): 359-366.e354.

Shindo, Y. & Amodeo, A.A. (2021). “Excess histone H3 is a competitive Chk1 inhibitor that controls cell-cycle remodeling in the early Drosophila embryo.” Curr Biol. 31(12):2633-2642.e6. PMID: 33848457

Shine M, Gordon J, Schärfen L, Zigackova D, Herzel L and Neugebauer KM. (2024). Co-transcriptional gene regulation in eukaryotes and prokaryotes. Nat Rev Mol Cell Biol. 25(7):534-554. PMID: 38509203

Talbert PB, Armache KJ and Henikoff S. (2022). “Viral histones: pickpocket’s prize or primordial progenitor?” Epigenetics Chromatin 15(1):21. PMID: 35624484

Vazquez, J., A. S. Belmont and J. W. Sedat (2002). “The dynamics of homologous chromosome pairing during male Drosophila meiosis.” Curr Biol 12(17): 1473-1483.

Weintraub H. (1972). “A possible role for histone in the synthesis of DNA.” Nature 240(5382):449-53. PMID: 4565937

White, A. E., B. D. Burch, X. C. Yang, P. Y. Gasdaska, Z. Dominski, W. F. Marzluff and R. J. Duronio (2011). "Drosophila histone locus bodies form by hierarchical recruitment of components." J Cell Biol 193(4): 677-694. PMC3166876

White, A. E., M. E. Leslie, B. R. Calvi, W. F. Marzluff and R. J. Duronio (2007). "Developmental and cell cycle regulation of the Drosophila histone locus body." Mol Biol Cell 18(7): 2491-2502. PMC1924828

White-Cooper, H. (2010). "Molecular mechanisms of gene regulation during Drosophila spermatogenesis." Reproduction 139(1): 11-21.

Wooten, M., Snedeker, J., Nizami, Z.F., Yang, X., Ranjan, R., Urban, E., Kim, J.M., Gall, J., Xiao, J., and Chen, X. (2019). "Asymmetric histone inheritance via strand-specific incorporation and biased replication fork movement." Nat Struct Mol Biol. 26(8):732-743. PMID: 31358945

Zhang, W., X. Zhang, Z. Xue, Y. Li, Q. Ma, X. Ren, J. Zhang, S. Yang, L. Yang, M. Wu, M. Ren, R. Xi, Z. Wu, J. L. Liu, E. Matunis, J. Dai and G. Gao (2019). "Probing the Function of Metazoan Histones with a Systematic Library of H3 and H4 Mutants." Dev Cell 48(3): 406-419.e405. PMC6595499

Zheng Y, Ahmad K, Henikoff S. (2025). "Total whole-arm chromosome losses predict malignancy in human cancer." Proc Natl Acad Sci U S A 122(18):e2505385122. PMID: 40314975

Supplementary Information

Supplementary Table 1. Sample IDs and sequencing results.

Attached spreadsheet.

Supplementary Table 2. Histone gene count tables.

Attached spreadsheet.

Bioinformatics for figures

For Figure 1: mapped reads for Mute (BT2579), Mxc (BT3065), Rpb1 (BT542), RNAPII-S5p (BT1781), and RNAPII-S2p (BT3067) were converted to bigwigs with bedtools/genome coverage and displayed in the UCSC genome browser.

For Figure 2: (a) .bam files were counted using subReads/featureCounts with option "-o" across a list of all annotated UCSC knownGenes in the dm6 genome assembly. Reads for each histone subtype were summed for each genotype, and counts in each histone subtype in the *12XWT* line were divided by counts in the wildtype line. **(b)** Mapped reads for H3K9me1 (BT1777), H3K9me2 (BT1778), H3K9me3 (BT1175), H3K27me3 (BT1173), uH2A (BT827), H3K4me1 (BT3072), H3K4me2 (BT3068), H3K4me3 (BT3074), H3K27ac (BT3069), and H3K36me3 (BT826) were converted to bigwigs with bedtools/genome coverage and displayed in the UCSC genome browser.

For Figure 5i: Mapped reads for Mapped reads for H3K27ac (BT3505), RNAPII-S5p (BT3417), Mxc (BT3195), and histone H4 (BT3510) were converted to bigwigs with bedtools/genome coverage and displayed in the UCSC genome browser.

For Figure 8: (b,c) K562 tracks Mapped reads for H3K27ac (BT3514), RNAPII-S5p (SH K5xlin_PolS5P_0320), NPAT (SH NPAT_0217_pool), and histone H4 (BT3518) were converted to bigwigs with bedtools/genome coverage and displayed in the UCSC genome browser. **(d)** Gene scores were counted from .bam files for K562 cell profiling using subReads/featureCounts with option "-o" across a list of all annotated genes in the hg19 (February 2009) genome assembly from -200 bp of annotated gene starts to gene ends. Histone genes are displayed shaded from maximum to minimum counts for each epitope.

Key Resources Table

Reagent type	Designation	Source	Identifiers	Additional info
Antibody	anti-RNAPII (mouse)	Covance Research Products	MMS-126R	(1:100 CUT&Tag)
	anti-RNAPII-S5p (rabbit)	Cell Signalling Technology	D9N5I	(1:100 CUT&Tag)
	anti-RNAPII-S2p (rabbit)	Cell Signalling Technology	E1Z3G	(1:100 CUT&Tag)
	anti-Mxc (guinea pig)	RJ Duronio	Mxc-C-1	(1:100 CUT&Tag)
	anti-Mute (guinea pig)	RJ Duronio	Mute	(1:100 CUT&Tag)
	anti-H3K9me1 (rabbit)	Epicypher	13-0029	(1:100 CUT&Tag)
	anti-H3K9me2 (mouse)	EMD Millipore	05-1249	(1:100 CUT&Tag)
	anti-H3K9me3 (rabbit)	Abcam	ab8898	(1:100 CUT&Tag)
	anti-H3K27me3 (rabbit)	Cell Signalling Technology	C36B11	(1:100 CUT&Tag)
	anti-uH2A (rabbit)	Cell Signalling Technology	D27C4	(1:100 CUT&Tag)
	anti-H3K4me1 (rabbit)	Epicypher	13-0057	(1:100 CUT&Tag)
	anti-H3K4me2 (rabbit)	Epicypher	13-0027	(1:100 CUT&Tag)
	anti-H3K4me3 (rabbit)	Epicypher	13-0060	(1:100 CUT&Tag)
	anti-H3K27ac (rabbit)	Epicypher	13-0059	(1:100 CUT&Tag)
	anti-H3K36me3 (rabbit)	Epicypher	13-0058	(1:100 CUT&Tag)
	anti-MPM2 (mouse)	DAKO	M3514	(1:100 IF)
	anti-GFP (rabbit)	Cell Signalling Technology	D5.1	(1:100 IF)
	anti-GFP (mouse)	Thermo Fisher Scientific	3E6	(1:100 IF)
	anti-rabbit IgG (guinea pig)	Antibodies Online	ABIN101961	(1:100 CUT&Tag)
	anti-mouse IgG (rabbit)	Abcam	ab46540	(1:100 CUT&Tag)
	anti-guinea pig IgG (rabbit)	Thermo Fisher Scientific	PA1-28549	(1:100 CUT&Tag)
	anti-rabbit-FITC (goat)	Jackson ImmunoResearch	111-095-144	(1:200 IF)
	anti-mouse-FITC (goat)	Jackson ImmunoResearch	115-095-166	(1:200 IF)
	anti-mouse-RRX (goat)	Jackson ImmunoResearch	115-295-166	(1:200 IF)
	anti-mouse-Cy5 (goat)	Jackson ImmunoResearch	115-175-166	(1:200 IF)
	anti-guinea pig-TRITC (goat)	Jackson ImmunoResearch	706-025-148	(1:200 IF)
	anti-guinea pig-Cy5 (donkey)	Jackson ImmunoResearch	706-175-148	(1:200 IF)
	anti-histone H4 (mouse)	Abcam	ab31830	(1:200 IF)

	anti-histone H3-S10p (mouse)	Abcam	Ab14955	(1:5000 IF)
	anti-NPAT (rabbit)	Thermo Fisher Scientific	PA565419	(1:200 IF)
Genetic reagent (*D melanogaster*)	*w*^{1118}	BDSC	3605	
	12XWT	RJ Duronio		VK33 insert, MacKay et al 2015
	w ; D*HisC ; 12XWT*	RJ Duronio		MacKay et al 2015
	His2ADendra2	this paper		P40 insert
	His2ADendra2	this paper		VK33 insert
	His3Dendra2	A Amodeo		P40 insert, Shindo & Amodeo 2019
	His2AvDendra2	this paper		CRISPR integrant
	His3.3ADendra2	A Amodeo		CRISPR integrant, Shindo & Amodeo 2019
	bamGAL4	BDSC	80579	
	UASRFP	BDSC	30556	
	GFPMxc	B Duronio		White et al 2011
	UAS-H3-GFP	K Ahmad		Schwartz & Ahmad 2005
	UAS-H3.3-GFP	K Ahmad		Schwartz & Ahmad 2005
	UAS-H4-GFP	this paper		M1A insert
	Su(var)3-9^{1}	BDSC	6209	
	Su(var)3-9^{2}	BDSC	6210	
	GD-27042 (*His1* RNAi)	VDRC	27042	Dietzl et al 2007
	P[TRiP.GL00081] attP2 (*His1* RNAi)	BDSC	35205	
	P[TRiP. HMS02879]attP2 (*His1* RNAi)	BDSC	44582	
	P[TRiP. HMS02959]attP40 (*His1* RNAi)	BDSC	50723	
	P[TRiP. HMS03724]attP2 (*His1* RNAi)	BDSC	54463	
	P[TRiP.GL00226] attP2 (*His2A* RNAi)	BDSC	35320	
	P[TRiP. HMS02295]attP2 (*His2B* RNAi)	BDSC	41881	
	P[TRiP. HMS02387]attP40 (*His2B* RNAi)	BDSC	41988	
	P[TRiP. HMS00162] attP2 (*His2AV* RNAi)	BDSC	34844	
	P[TRiP. HMS02773] attP40 (*His2AV* RNAi)	BDSC	44056	

	P[TRiP.GL00255] attP2 (*His3* RNAi)	BDSC	35344	
	P[TRiP. HMS02385] attP40 (*His3* RNAi)	BDSC	41986	
	P[UAS-His3.3A-IR]5825R-3 (*His3.3A* RNAi)	NIG-FLY	5825R-3	
	GD-12771 (*His3.3B* RNAi)	VDRC	12771	
	P[TRiP. HMC06602] attP40 (*His4* RNAi)	BDSC	80366	
	P[TRiP. HMS00126] attP2 (*His4r* RNAi)	BDSC	34816	
	P[TRiP. HMJ21134] attP40 (*His4r* RNAi)	BDSC	51172	
	P[TRiP. HMS00279] attP2 (*Su(var)3-9* RNAi)	BDSC	33401	
	P[TRiP.GL01471]	BDSC	43661	
	P[TRiP. HMS00112] attP2 (*egg* RNAi)	BDSC	34803	
	P[TRiP.GL01002] attP40 (*egg* RNAi)	BDSC	36797	
	P[TRiP. HMS00066] attP2 (*E(z)* RNAi)	BDSC	33659	
	P[TRiP.GL00371] attP2 (*Sce* RNAi)	BDSC	35446	
	P[TRiP. HMJ23146] attP40 (*Sce* RNAi)	BDSC	61312	
	P[TRiP. HMS00278] attP2 (*HP1* RNAi)	BDSC	33400	
	P[TRiP.GL00531] attP40 (*HP1* RNAi)	BDSC	36792	
	P[TRiP. HMS01699] attP40 (*HP2* RNAi)	BDSC	38255	
	PcGFP	BDSC	9593	
	UAS-H4-eGFP	X Chen		Wooten et al 2019
Biological sample (*D melanogaster*)	Male larval wing imaginal discs	this paper		
	Male adult testes	this paper		

Biological sample (*H sapiens*)	K562 cell culture	ATCC	CCL-243	
Commercial reagent	pAGTn5	Epicypher	15-1017	
	EdU Click-iT kit	Invitrogen	C10340	
Software	bowtie2			
	bedtools/ bamcoverage			
	subReads/ featureCounts			