D2-expressing neurons of the anterior paraventricular nucleus of the thalamus modulate

learned aversive and safety salience.

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Abstract

Associative learning provides a mean to learn appropriate valence and salience value signals that

animal can later use to make appropriate value-based choices. By generalizing from prior aversive

experience and comparing motivational salience signals to a current one, animals can learn the relative

aversive ("better or worse than") value or relative positive safety between experiences. Yet, how such

relative value signals are acquired during learning remain poorly known. The paraventricular nucleus

of the thalamus (PVT) is critical for the control of salience and valence signals, therefore it is well-placed

to be involved in learning relative aversive value and learned safety. Using a newly-developed

conditioned place aversive task, we first uncover specific behavioral signatures associated with learning

relative aversive value and learned safety, and related value-based choices. We then show that

neurons expressing dopamine D2 receptor (D2+) in the anterior PVT (aPVT) rather than posterior PVT

(pPVT), are preferentially recruited by both relative aversive value learning and learned safety. By

employing a chemogenetic inactivation strategy, we first demonstrate a role of aPVT D2+ neurons for

learning relative aversive value by regulating motivational aversive salience. Finally, we show that the

same chemogenetic intervention also increases learned safety salience. Overall, our data are the first

to demonstrate that aPVT D2+ neurons modulate the salience of experience independently of valence

to compute adapted learned value signals during learning and promote appropriate value-based

choices.

Introduction

Valence (positive or negative) and salience (e.g. intensity of a reinforcer) of behaviorally relevant signals are critical value information acquired during associative learning that animals can later use to make appropriate choices. [1–3] Nevertheless, past experience can modify how value information is acquired during learning. [4–6] By generalizing predictions [7,8] and comparing the salience of contextual/sensory signals of a prior experience with an ongoing one, animals can also learn the relative appetitive or aversive value between two experiences (i.e. whether one is better or worse than the other). [9–14] When later confronted to a choice between the two options, pre-computed values acquired during learning facilitate decision-making processes promoting more efficient and accurate value-based choices. [4] Critically, overgeneralization, leading to alteration of relative value computation, is maladaptive and serves as a major feature of fear-related disorders including post-traumatic stress disorder (PTSD). [7]

Previous studies show that learning the relative aversive value between two aversive experiences, such that one is perceived as better than the other, involves the reward system.[9,10,15] Similarly, a neutral and safe experience can acquire a relative positive value, or learned safety, when compared to a prior aversive experience, a process that also requires the reward system. [16–18] While fear generalization [6,7] and the interaction between appetitive and aversive systems [10,19–21] are likely crucial players, how the comparison of generalized predicted values and current motivational salience contribute to the encoding of relative aversive value and learned safety remains poorly understood.

The paraventricular nucleus of the thalamus (PVT) is a hub connecting the thalamus with cortical and limbic structures [22–26] that is critically involved in stimulus salience processing [27,28] and value assignment to associative stimuli. [29–32] The PVT is therefore a strong candidate structure for disentangling motivational conflicts and comparing experiences to compute appropriate relative value signals during learning. [5,23,27,32–35] Through its involvement in granting and updating the value of aversive experiences [27,34,36–40], here we hypothesize that the PVT is crucial for a correct assignment of a relative aversive value. In particular, the D2 dopamine receptor-expressing neurons (D2+) of the PVT are sensitive to valence [32] and are implicated in signaling aversive states. [32,37] In contrast, D2-non-expressing neurons (D2-) are modulated by arousal, relaying a salience signal to the cortex. [32] According to this functional dissociation, D2+ neurons could be crucial to inform a "better than expected" relative aversive value.

On the other side, the PVT is also instrumental for the correct assignment of a safe value to an

experience. [40] However, it remains unclear whether D2+ neurons along the antero-posterior axis of the

PVT differentially contributes to encoding of relative aversive value and learned safety.

Here, we addressed this question by designing a novel conditioned place avoidance task that allows

mice to encode either relative aversive value or learned safety. In this task, mice learn to associate one

context with a foot shock punishment, followed by a second context that is either punished with a shock

of lower intensity (learning a relative "better" aversive value), or unpunished (learning a relative positive

safety). After learning, mice are confronted to choose between the two contexts. We show that mice

adapt their defensive responses to each task, exhibiting distinct behavioral signatures during learning

and choice-test. Our data reveal that during learning of the second context, when comparing it to the

previous aversive experience, both tasks preferentially recruit D2+ neurons in the aPVT compared to

pPVT. We found that pharmacogenetic silencing of aPVT D2+ neurons during the exposure to the

second context disrupt relative aversive comparison by increasing learned aversive salience leading to

a suppressed preference towards the less punished-associated context during the choice-test.

Consistently, when mice learn to associate two distinct contexts paired with the same shock intensity,

silencing of aPVT D2+ neurons during the exposure to the second context increases the aversive

salience of the experience, tilting mouse preference towards the first context during choice test. We

also found that silencing aPVT D2+ neurons increases safety salience of the ongoing experience

inducing enhanced preference towards the safe context during the choice test.

Taken together, our data demonstrate that aPVT D2+ neurons modulate motivational salience to enable

accurate assignment of relative aversive value and learned safety during learning, thereby promoting

appropriate value-based choices. These results also provide compelling evidence that PVT neuronal

ensembles expressing D2 receptors, embedded in limbic circuits, are key in the evaluation process for

assigning correct value signals to sensory experience during associative learning.

Results

Mice use different behavioral strategies for learned safety and relative aversive value-based learning and choice.

To study the neural circuits and mechanisms underlying learned safety and relative aversive motivational salience, we designed a novel 1-day version of the conditioned place avoidance (CPA) task **(Figures 1A** and **S1A**). In this task, mice are first subjected to a pretest session ("Pretest"), allowing them to explore two never-encountered contexts A and B (connected via a bridge) followed by two conditioning sessions ("S1" and "S2") in each context, and a test session ("Choice-test") between contexts A and B. Three groups were tested. For the control group, no shock was delivered in any context during S1 and S2. For the safe group (to study learned safety), mice received 4 electric shocks of 0.6 mA in Context A during S1 ("A+0.6"), but no shock in Context B during S2 ("B+0"). For the aversive group (to study learning relative aversive value), mice were subjected to 4 electric shocks of 0.6 mA in Context A during S1 ("A+0.6") as for the safe group, but received 4 electric shocks of 0.3 mA in Context B during S2 ("B+0.3"). As expected, no freezing was observed during Pretest in any group (**Figure S1B**). Both safe and aversive groups exhibited increased freezing levels in S1 and S2 conditioning sessions compared to the control group (**Figure S1C**). However, only mice in the aversive group showed increased freezing from S1 to S2 **(Figure 1B)**, along with heightened anxiety as measured with a decreased time spent in the center during S2 (**Figure S1D**). Looking at the dynamic of freezing during S1 and S2 (**Figure S1E**), we found that freezing before the putative first shock of S2, but not S1, significantly (and similarly) increased for both safe and aversive groups compared to control group (**Figure 1C**). Increased freezing at the beginning of S2 suggests a generalized expectation based on their previous aversive experience in Context A. Importantly, while mice in both aversive and safe groups increased freezing after the 4th (and last) shock in S1 (**Figures 1D** and **S1E**), only mice in the aversive group increased freezing at the end of S2 (i.e. after receiving 4 shocks) (**Figures 1D** and **S1E**). Strikingly, we also observed tail-rattling behavior only in the aversive group immediately after each shock event in S2 (**Figure 1E; video 1**), but not S1 (**Figure S1F**). Therefore, the aversive group displayed an intense defensive response to a threat [41] after the delivery of additional shocks in S2. Overall, our results show that after the context-shock association, the number of aversive events modify mice general state and defensive response.

All mice were tested during a choice-test session one hour after conditioning. Importantly, mice were first placed in their most preferred context (defined during Pretest), i.e. the context A_0 for the control group and the most punished context $A_{0.6}$ for both safe and aversive groups. We first looked at their latency to cross from context A to context B (i.e. 1st latency, **Figure S2A**). While the first latency was similar between all 3 groups during the choice-test (**Figure S2A**), only mice from the aversive group spent more time in the bridge before the 1st cross (**Figures 2A** and **2B**). In addition, most of mice in the aversive group (10 out of 12) exhibited a "turn-back" behavior (**Figure 2C; video 2**) and a tendency for more tail rattling (**Figure S2B**). Such behavioral signature is reminiscent of the "vicarious trial and error" behavior with animals moving back and forth in direction of different choices at decision zones. [42,43]

After crossing the bridge for the first time (i.e. 1st cross), mice were allowed to explore the two contexts for 15 min (as in [44,45]) (**Figure S2C**). We calculated a ☐CPA score comparing the time spent in each context during Pretest (15 min, **Figure S2D**) vs Choice-test (15 min after 1st cross, **Figure 2D**). As expected, control mice showed no preference between the two contexts during choice-test (**Figure 2D**). Only mice of the safe and aversive groups reverted their initial preference and spent more time in the less aversive context (**Figure 2D**), along with a reduced exploratory behavior (**Figure S2E**). This preference was similar for both safe and aversive groups during the first 5 min of the test, but was maintained for the aversive group throughout the choice-test (**Figure S2F**). For the aversive group, this preference was accompanied with increased freezing in the bridge and context $B_{0.3}$ (**Figure 2E**), along with higher stress marked by more pee and poo (**Figure S2G**). In line with these results, we found that the aversive group stayed longer in the bridge after the 1st cross (**Figure 2F**) and therefore took longer to go back to context $A_{0.6}$ (i.e. 2nd latency, **Figure 2G**). Mice in the aversive group therefore utilized prior learning information to avoid the most punished context $A_{0.6}$ and potentially approach context $B_{0.3}$ [15] while being anxious and have difficulties to make a choice between the two contexts. Overall, we reveal that mice adopt specific behavioral signatures induced by prior learned experience to enable adapted value-based choices.

Both learned safety and relative aversive learning recruit D2-expressing neurons in the aPVT

We hypothesize that mice compare the relative saliency of the two experiences in conditioning S1 vs S2 and assign a relative "better than" aversive value to Context B paired 0.3mA shocks for the aversive group or a relative safety to context B unpaired for the safe group. However, how mice compare the

motivational salience of each experience to encode a relative (aversive or safe) value between experiences and influence ensuing value-based choices is unclear. The PVT is central for modulating anxiety-related conflicting appetitive-aversive information. [35,46] We therefore propose that the PVT can be pivotal in the encoding of relative (aversive or safe) value. Notably, the functional antero-posterior axis and expression D2 dopaminergic receptor (D2+ neurons) subdivision of the PVT in salience and valence coding [32,47,48] could extricate the different motivational salience of prior and actual information learned during the conditioning session S2. We first tested this hypothesis by asking whether PVT neurons, in particular D2+ neurons, are differentially recruited during learning using D2-Cre:Ai14 mice (see Methods). First, our *ex vivo* whole-cell recording experiments confirmed that td-Tomato (i.e. Ai14) cells are responding to quinpirole (D2R-like agonist, 1µM; Action potential (AP) frequency baseline = 0.10 +/- 0.20 Hz, AP frequency quinpirole = 0.37 +/- 0.21 Hz, n= 5 cells, paired t-test p= 0.0047 **) validating the use of this mouse line. We then performed a c-Fos immunostaining in the PVT of D2-tagged animals of 3 groups: home-cage (HC), safe and aversive (**Figures 3A** and **3B**). To reflect c-Fos activity at the time of the comparison between the two conditioning sessions, mice were perfused 90 min after the conditioning session S2. Behaviorally, we verified that both safe and aversive groups showed distinct freezing patterns at the end of S2 (**Figures S3A** and **S3B**), as we previously described (**Figures 1C** and **1D**). Looking at PVT c-Fos positive neurons, we found that both safe and aversive groups show similar levels of activated neurons, but significantly higher compared to home-cage group (**Figure 3C**). In both groups, such recruitment was more prominent in the anterior PVT (aPVT) than the posterior PVT (pPVT) (**Figures 3C** and **S3C**) and incorporated a higher proportion of D2+ than D2-neuronal population (**Figure 3D**). Importantly, this was not due to an overall increased D2+ cell density in the aPVT, nor to a difference of D2+ cell densities between groups (**Figure S3D**). These results indicate that both learned safety and relative aversive learning recruit preferentially D2-expressing neurons in the aPVT. Mice were subjected to a choice-test immediately before being perfused (**Figures 3A** and **S3E).** In line with our previous findings (**Figure 2D)**, both safe and aversive groups significantly preferred the non-punished (Context B_0) or less punished compartment (Context $B_{0.3}$) during the choice-test, respectively (positive □CPA score in **Figure S3E**). We also showed that, as in **Figure 2**, the relative choice between the two previously (and differently) punished contexts $A_{0.6}$ and $B_{0.3}$ led mice to spend a long time in the bridge likely assessing the two aversive options to make an appropriate choice (**Figure S3F**). Considering the role of the PVT and the neuronal activation of D2+ neurons during the

conditioning session S2, we hypothesize that mice behavior during the choice test is tightly linked with

the activity of aPVT D2+ neurons during learning in S2. Accordingly, we found that in the aversive

group, the percentage of activated D2+ neurons of the aPVT (but not pPVT) during the conditioning

session S2 is negatively correlated with the time spent in the bridge (i.e. the decision zone) during the

choice test (**Figure 3E**). In other words, the less aPVT D2+ neurons were activated during S2, the more

time mice will spend in the bridge at the beginning of the choice test. Strikingly, such correlation was

not observed for aPVT D2-negative activated neurons (**Figure 3E**) nor for the safe group (**Figure 3F**).

These results strongly suggest a crucial role of aPVT D2+ neurons for learning relative aversive value

during conditioning to subsequently making an appropriate relative aversive value-based choice.

aPVT D2-expressing neurons are crucial for learning relative aversive value

Next, we directly tested the role of aPVT D2+ neurons for learning relative aversive value during

conditioning in S2. We virally injected D2-Cre mice with a Cre-dependent DREADD (hM4Di) or control

(mCherry) viral vector in the aPVT (**Figure 4A**). Four weeks later mice underwent the aversive CPA

task and were intra-peritoneally injected CNO 30 min before S2 to silence aPVT D2+ neurons when

mice relatively compare the previous S1 experience with the current S2 one (**Figure 4B**). To ascertain

the effect of viral transduction, we performed *ex vivo* whole-cell recordings in control- and hM4Di-D2-

expressing neurons of the aPVT, identified by their mCherry fluorescence (**Figure S4A**). Our data

confirmed that bath application of 10□M CNO to aPVT brain sections expressing inhibitory DREADD

(hM4Di) hyperpolarizes D2+ neurons by drastically decreasing spike frequency. In contrast, bath

application of CNO was ineffective on aPVT D2-expressing neurons transduced with the control virus

mCherry (**Figure S4A**). In line with previous reports, we also confirmed projection targets of aPVT D2+

neurons such as Nucleus Accumbens (NAc) and Amygdala (**Figure S4B**). [32] First and as expected,

both groups showed a similar preference of enclosure during Pretest before CNO injection (**Figure**

S4C). While silencing aPVT D2+ neurons prior to and during S2 did not change the amount of freezing

during conditioning (**Figures S4D** and **S4E**), it led to an increased tail-rattling behavior in S2

immediately after receiving each shock (**Figure 4C**). Moreover, while we observed a decreased

average distance travelled after each shock between S1 and S2 in the control mCherry, inhibiting aPVT

D2-expressing neurons lead to a maintenance of this distance during S2 (**Figure 4D**). Such findings

suggest that silencing aPVT D2+ neurons enhanced the shock perceived by the mice during S2

therefore increasing the aversive salience of context B+0.3. As a consequence, before their first cross

from context $A_{0.6}$ to context $B_{0.3}$ during the choice-test mice display a higher number of turnback (**Figure**

4E) and tail rattling (**Figure 4F**) in the bridge without changing their first latency (**Figure S4F**) nor the

time they spent in the bridge (**Figure S4G**). These results seem to indicate that mice perceive context

$B_{0.3}$ as threatening as context $A_{0.6}$. In line with this hypothesis, once mice first cross from context $A_{0.6}$ to

context $B_{0.3}$, the preference of the mCherry controls towards the previously less punished context $B_{0.3}$

was fully suppressed in hM4Di mice when aPVT D2+ neurons were inactivated during S2 (ΔCPA score

around 0; **Figure 4G**). We also found that hM4Di mice spent more time in the bridge after their first

cross from context $A_{0.6}$ to context $B_{0.3}$ (**Figure 4H**), indicating more hesitation to choose between the

two contexts. Moreover, mice froze less in the supposedly better option of context $B_{0.3}$ (**Figure 4I;** also

see **Figure 2E**) and returned to context $A_{0.6}$ faster (**Figure 4J**). Overall, our results suggest that silencing

aPVT D2+ neurons enhanced the aversive salience of the "B+0.3 mA" experience to reach a similar

aversive salience as in context A+0.6 mA. This will therefore prevent mice to learn the relative difference

between the two aversive experiences and to assign a relative "better than" aversive value to context

B+0.3 mA. In line with this hypothesis, conditioning mice to contexts A and B both paired with 0.6 mA

(**Figure S4H**) led to similar results with notably an increased tail rattling during S2 and a similar

distanced travelled after shock between S1 and S2 (**Figures S4I-L**). In addition, we observed similar

consequences during the choice-test with notably a ΔCPA score not different from 0 and similar freezing

in $A_{0.6}$, bridge and $B_{0.6}$ (**Figures S4M-R**). Considering the role of the PVT for regulating stimulus

salience [27,34,49–51], we propose that aPVT D2+ neurons play a critical role for maintaining the relative

salience between the two aversive experiences and therefore for participating to the encoding of relative

aversive value triggering adapted threat-related behavior.

aPVT D2+ neurons modulate the salience of an aversive experience.

To test whether silencing aPVT D2+ neurons can enhance the salience of an aversive experience we

conditioned mCherry and hM4Di mice to associate the two contexts A and B with the same series of

low 0.3 mA shock intensity and injected CNO prior to S2 to potentially enhance the aversive salience

of context B+0.3 mA (**Figure 5A**). Confirming our hypothesis, we found that silencing aPVT D2+

neurons drastically increased the number of tail rattling during S2 (**Figure 5B**) inducing a slight

reduction of time freezing after the 4th shock between S1 and S2 (**Figure 5C**). However, it did not modify

the distance travelled after shock (**Figure S5A**). Furthermore, and as previously shown in **Figure S4D**,

silencing aPVT D2+ neurons prior to S2 did not change mice freezing before the first shock during S2

(**Figure S5B**). These results indicate that mice perceive the conditioning session S2 as more aversively

salient therefore engaging into higher threat predicting behavior. As a consequence of context B+0.3

mA being perceived as more salient and therefore more aversive than context A+0.3 mA, we found that

hM4Di mice first placed in $A_{0.3}$ during the choice-test display more turnback in the bridge before their

first cross to context $B_{0.3}$ (**Figure 5D**). We also observe a strong trend toward spending more time before

their first cross to context $B_{0.3}$ (1st latency; **Figure 5E**) but not in the bridge (**Figure 5F**). However, hM4Di

mice do not display more tail rattling before their first cross to context $B_{0.3}$ (**Figure S5C**). We also found

that after their first cross to context $B_{0.3}$ while hM4Di mice spent the same amount of time in the bridge

than control (**Figure S5D**), they tend to freeze more in the bridge (**Figure 5G**) without changing the

latency to cross back to context $A_{0.3}$ (**Figure S5E**). Moreover, while control mice spend more time in

context $B_{0.3}$ (ΔCPA positive score; **Figure 5H**), hM4Di mice had not preference between the two

contexts (ΔCPA score not different from 0; **Figures S5F** and **5H**). This behavior is likely due to the

stronger aversiveness of context $B_{0.3}$ compared to that of context $A_{0.3}$. Combined with our results in

Figures 4 and **S4**, we found that aPVT D2+ neurons can modulate the salience of an aversive

experience and therefore are essential in the comparison between the intensity of aversive experiences

during learning to enable correct assignment of relative aversive value crucial to promote future

appropriate value-based choices.

aPVT D2+ neurons are involved in the incentive salience of learned safe experience.

Our results in **Figures 3C** and **3D** strongly suggest that aPVT D2+ neurons are involved in both learned

relative aversive value and relative safety. PVT neurons have been shown to regulate the salience of

both aversive and appetitive behavior [27], a crucial information for both fear and reward learning. [17,52]

We therefore hypothesize that silencing aPVT D2+ neurons prior to and during S2 could also enhance

the salience of learned safety of context B+0 mA when it follows conditioning of context A+0.6 mA. To

test our hypothesis, we subjected mCherry and hM4Di mice to the safe conditioning (**Figure 1A**) and

injected CNO prior to conditioning S2 in context B+0 mA (**Figure 6A**). Silencing aPVT D2+ neurons

prior to and during S2 did not change mouse threat-related behavior during conditioning (**Figures 6B-**

C and **S6A-B**). However, mice behavior was significantly altered during the choice test between context

$A_{0.6}$ and B_0. As expected, threat-predicting behavior such as the number of turnback and tail-rattling

behavior in the bridge before going to context B_0 (i.e. before 1st cross) was unchanged (**Figures 6D-E**).

However, mice tend to exhibit a shorter latency to first cross to the safe enclosure B_0 (i.e. 1st latency,

Figure 6F) and a longer latency to go back to the punished context $A_{0.6}$ (**Figure 6G**), without changing

the time spent in the bridge after their first cross (**Figure S6C**). In addition, and in line with our

hypothesis, hM4Di mice exhibited a higher percentage of freezing in the safe context B_0 (**Figure 6H**)

as they showed a higher preference for the safe context B_0 compared to control mCherry mice (**Figures**

S6D and **6I**). This indicates that silencing aPVT D2+ neurons prior to and during S2 increased the

incentive salience of context B+0 mA thus inducing a higher preference towards the safe context B_0

during the choice test.

To make sure that silencing aPVT D2+ neurons before context B+0 mA did not assign a positive value

to this context during conditioning we exposed mCherry and hM4Di mice to two novel and distinct

contexts (C and D contexts, **Figure S6E**) without shock delivered, and injected CNO 30 min before S4

in context D+0. As expected, we did not find any change in any mice behavior during conditioning or

choice test (**Figures S6F-P**). Altogether, these findings strongly suggest that aPVT D2+ neurons

modulate learned safety during conditioning markedly influencing ensuing value-based choice

behavior.

Discussion

Our study reveals the critical role of aPVT D2-expressing neurons in modulating salience for learning

relative aversive value and relative positive safety to promote appropriate value-based choices. Using

a novel conditioned place avoidance task, we found that distinct behavioral signatures are associated

with learning relative aversive value and relative positive safety, and related value-based choices. We

then demonstrated that learning relative aversive value and relative positive safety preferentially recruit

aPVT D2-expressing neurons. When comparing an ongoing experience with a previous aversive one,

inactivating aPVT D2-expressing neurons increases the motivational salience of the ongoing

experience, i.e. enhancing the aversiveness or the safety of the experience. Such salience

enhancement modifies learning and then the selection of subsequent behavior when mice have to

choose between the two available options. This work provides key findings that may increase our

understanding of how context-dependent relative value and motivational salience are encoded in the

brain.

aPVT D2+ neurons for learning relative aversive value.

Extensive work focused on brain mechanisms for relative appetitive learning related to reward and gain.

[14,53,53,54,54–64] Surprisingly, the neural circuits underling relative aversive learning and choice remain

poorly understood. Here, we designed a novel 1-day conditioned place avoidance task in which mice

learn the association of two distinct contexts with different shock intensities, 0.6mA paired with the first

context and a second context with less intense 0.3mA shocks. Later, mice are confronted to a choice

between the two contexts. Our data showed that this relative aversive task is associated with increased

freezing, apparition of tail rattling and decreased time in the center during the second conditioning

session. Strikingly, tail rattling is a behavior typically associated with aggression in male mice before

fighting. [65] In our set of experiments, tail rattling may therefore be associated with a negative emotional

state and a reaction against a possible threat, occurring during learning (i.e. immediately after a

negative shock event) but also during memory retrieval (when mice need to choose between two

aversive contexts). Previous work has suggested a role of PVT for modulating the value and salience

of stimuli. [27,32] Moreover, previous studies highlighted its crucial role during motivational conflict

scenarios where both approach and defensive behaviors compete. [27,35,46,50] We found that

pharmacogenetic inactivation of aPVT D2+ neurons enhances the saliency of the ongoing aversive

experience demonstrated by increased tail-rattling and running immediately after each shock. Since tail

rattling behavior was recently shown to increase visual and auditory saliency of the mouse to predators

[41], our data suggest that aPVT D2+ neuronal inactivation caused a shift in the defensive behavior

strategy, favoring saliency-enhancing responses to threat. Our results also reveal that during the

relative choice between the two previously punished contexts, mice increased time spent and numerous

turnback when located in the bridge zone. Such behavior is reminiscent of the "Vicarious Trial and Error"

(VTE) phenomenon [42,66] observed at decision point when animals pause and move their head back and

forth to "scan" and evaluate the possible outcomes before making a choice between two or more options

with uncertain outcomes. [43] Importantly, VTE behavior is preferentially exhibited in hippocampal-

dependent tasks and during complex decision-making processes. [67–69] Given the critical role of

contextual information in our new relative task and the observed PVT-hippocampus anatomo-functional

connectivity [22], our newly designed relative aversive task can serve to expand our knowledge of the

neuronal circuits underlying VTE behavior.

aPVT D2+ neurons modulate predictive signals for learning aversive value and learned safety.

The PVT is widely responsive to aversive states and modulates fear-related behaviors [22,27,32,34,36,39,70–

72], but is also involved in appetitive associative learning [27,51] and reward-seeking behavior. [49,73,74]

Moreover, PVT neurons are activated by cues or contexts previously associated with positive or

negative emotional outcomes. [22,34–36,49,71,75–80] Here, we also designed a task in which the first context

is paired with 0.6 mA shocks followed by a second unpaired context, likely acquiring a learned safety.

[7] Our c-Fos staining revealed that the aPVT is activated by both a second context paired with a mild

shock (relative aversive) or the absence of shock (learned safety). As mice likely generalize expected

negative prediction from a previous aversive experience to learn relative aversive value or safety, our

results are reminiscent of previous reports showing that the omission of expected reward activates the

aPVT, but not the pPVT [49] The aPVT is therefore strongly involved in generating predictive value signals

independent of valence. To compute these predictive value signals, the PVT receives inputs from

regions involved in learned safety such as the infralimbic, prelimbic cortex [26,81–84] and ventral

hippocampus [85,86], that convey information about motivational state and arousal, respectively. To

modulate predictive value signals the PVT projects to the ventral tegmental area (VTA) and substantia

nigra pars compacta (SNc) [87] where computation of prediction error (comparison between predicted

and actual information) is crucial to signal correct value to sensory experience during learning.[88,89]

Notably VTA dopaminergic neurons expressing D2 receptors (auto-receptors) can regulate the salience

of threat signals. [90] As the reward system is involved is both relative aversive value [9,10] and learned

safety [16,18,91,92], the PVT could modulate the computation of relative value signals in the VTA. The aPVT

also projects to regions modulating approach-avoidance behaviors such as PFC [32,93] and ventral

subiculum [93], extended amygdala [93,94], nucleus accumbens (NAc) shell [25,93,95] and NAc core. Since NAc

integrates dopaminergic inputs from the VTA with glutamatergic inputs from the PVT [96,97], inactivation

of aPVT D2+ neurons could deregulate dopamine signaling in the NAc. This could potentially lead to

an aberrant assignment of salience to neutral stimuli [98] and therefore disrupt the ability to modulate

aversive and safety-related learning.

Furthermore, our results revealed a preferential activation of aPVT D2-expressing neurons during

learning relative aversive value or safety, although we and others found D2+ neurons are preferentially

located in the pPVT. [32,47]. Importantly, generalized threat prediction, involved here for learning relative

aversive value or safety, also involve the activation of D2 type receptors in both extended amygdala

substructures i.e.e the central amygdala and the bed nucleus of the stria terminalis BNST that have

been shown to contribute to anxiety-like behaviors. [99] Noteworthy, aPVT D2-expressing neurons

activation was tightly linked with the time spent in the bridge during choice-test, a crucial behavioral

hallmark of the decision-making process between different options. Indeed, the less recruited aPVT

D2+ neurons, but not pPVT, the more animals spent time in the bridge in the relative aversive, but not

safe, condition. Consistent with this observation, we found that the selective inactivation of aPVT D2-

expressing neurons increases the time spent in the bridge in the relatively aversive, but not the safe

condition, thus preventing mice to select the most appropriate behavior when facing two aversive

situations.

aPVT D2+ neuronal modulation of motivational salience

Emotionally salient events, regardless whether positive or negative, are encoded and remembered with

greater accuracy compared to emotionally neutral ones. [100–104] The PVT is involved in arousal [32,105,106]

and can encode salience of both conditioned and unconditioned stimuli. [27] Our results showed a role

for aPVT D2-expressing neurons for modulating the salience of an experience, with increasing its

aversiveness or its safety. It remains unclear whether the conditioned stimulus (here, the context) or

the outcome (shock or no shock delivered) become more salient. For the relative aversive task, such

manipulation increased tail rattling behavior during conditioning, but not freezing. Consistently, a recent

study identified that tail rattling, but not freezing behavior, positively scales with arousal level. [41]

Inactivation of aPVT D2-expressing neurons also increased the number of turnback and time in the

bridge (i.e. hesitation) during the choice-test, thus leading to the absence of preference for the less

punished context. Importantly, the same inactivation procedure during the safe conditioning did not

affect tail rattling behavior nor the time in the bridge, but it led to in an increased preference for the safe

context. The efficiency of the value-based choices is therefore influenced by the salience of the

experience and the computation and comparison of values assigned at the time of learning. [4]

Inactivation of aPVT D2+ neurons could have led to an incorrect value assignment during the context-

outcome association, thus favoring the contribution of D2-negative neurons for the attribution of

salience to the ongoing experience. [32,78] This could hamper (for the relative aversive task) or favor (for

learned safety) the ability of animals to distinguish between the options associated with each context.

Inactivating aPVT D2+ neurons could therefore enhance the formation of memory of any otherwise low-

salient stimuli.

Overall, our findings extend current knowledge about the role of PVT as an integrator of emotional,

contextual and sensory information to guide appropriate behavior. This work provides new evidence

about the role of aPVT D2-expressing neurons for contextual aversive learning that supports value-

based decisions. Here, we show that this population modulates the salience of contextual

cues/information, which allows mice to detect the aversiveness or safety of an experience. A better

understanding of neuronal circuit mechanisms responsible of the balance between value information

and salience is therefore critical for motivated behaviors. Indeed, an excessive stimulus evaluation

and/or salience processing during traumatic events (that could involve the PVT) might lead to

associative learning impairment and contribute to the genesis of anxiety disorders such as post-

traumatic stress disorder (PTSD). [107,108]

Resource availability

Further information and requests for resources and reagents should be directed to, and will be fulfilled

by, the lead contact, Emmanuel Perisse (emmanuel.perisse@igf.cnrs.fr) and Stéphanie Trouche

(stephanie.trouche@igf.cnrs.fr).

Materials availability

All original reagents presented in this study are available from the Lead Contacts upon request.

Data and code availability

All code used in this paper using Fiji will be shared by the Lead Contacts upon request.

METHODS

Animals

We used C57BL/6J (Charles River Laboratories, France) and three transgenic heterozygous adult male mouse lines: D2-Cre mice (STOCK Tg(*Drd2*-cre) ER44Gsat/Mmucd, provided by E. Valjent), D2-Cre:Ribotag mice [109] (provided by E. Valjent) and D2-Cre:Ai14 mice, that were generated by crossing Drd2-Cre mice with Ai14 mice (B6.Cg-*Gt(ROSA)26Sor*$^{tm14(CAG-tdTomato)Hze}$/J, stock #007914). The latter transgenic mouse line expressed the red fluorescent protein double-tomato (tdTom) under endogenous regulatory elements of the D2 receptor. All transgenic mice were maintained on a C57BL/6J background. Animals were housed with their littermates and were singly housed in their home cage after the surgical procedure on a 12-h light/dark cycle (7 a.m. to 7 p.m.) in stable temperature (22°C) and humidity (60%) conditions with free access to food and water in a dedicated housing room. Mice were randomly assigned to behavioral groups. All experimental procedures were conducted in agreement with the guidelines of the French Agriculture and Forestry Ministry for handling animals (authorization number/license A3417241) and approved by the local and national ethic committees (authorizations APAFIS #23555).

Immunofluorescent staining and c-Fos analysis

Mice (D2-Cre:Ai14 or D2-Cre:Ribotag, see Figures 3 and S3) were anaesthetized and transcardially perfused with PBS followed by 4% PFA in PBS solution 90 min after the second session of conditioning of the CPA task. Brains were extracted and kept in 4% PFA for at least 24 h before slicing using the vibratome and coronal sections (30µm thick) were stored in PBS-azide at 4°C. For the immunohistochemistry procedures, free-floating sections were rinsed extensively in PBS with 0.25% Triton X-100 (PBS-T) and blocked for 1 h at room temperature in PBS-T with 10% normal donkey serum (NDS). An average of 3.5 sections per animal of either aPVT or pPVT were used. Sections were then incubated overnight at room temperature with primary antibodies diluted in 3% NDS blocking solution. The primary antibodies used were: anti-RFP (MBL, rabbit, 1/2000, #PM005), *c-Fos* antibody (Synaptic Systems, rat, 1/5000, #226017), anti-HA (rabbit, 1/500, Rockland #600-401-384). After three 15 min washes using PBS, sections were incubated overnight at room temperature in PBS containing a mixture of secondary antibodies. Secondary antibodies used were: donkey 488-coupled anti-rabbit (Jackson Laboratory, 1/1000), donkey Cy3-coupled anti-rabbit (Jackson Laboratory #711-165-152, 1/1000),

donkey Cy5-coupled anti-rat (Jackson Laboratory #712-605-150, 1/1000), donkey 488-coupled anti-rat

(Jackson Laboratory #712-545-153, 1/500) and donkey Cy3-coupled anti-rat (Jackson Laboratory

#711-545-152, 1/1000). After three 15 min washes in PBS, sections were mounted on slides and stored

at 4°C. Images were taken using a ZEISS Apotome Microscope at 16bits and either 10X or 20X.

c-Fos expression was examined in D2 tagged cells using the D2-Cre:ai14 mice or D2-Cre:Ribotag (in

a balanced manner by group) using ImageJ based automated analysis. For each section, a region of

interest was assigned for both aPVT and pPVT and a threshold was selected to count both the amount

of c-Fos+ cells, D2+ cells and c-Fos+ D2+ colocalization. Sections from the anterior part of the PVT

(aPVT) were taken at AP:-0.22 to -1.22 while the posterior part of the PVT (pPVT) were taken at AP:-

1.34 to -2.18.

In vivo **pharmacogenetic inhibition**

For *in vivo* pharmacogenetic (Designer Receptors Exclusively Activated by Designer Drugs, DREADD)

inhibition in CPA experiments, Clozapine N-Oxide (2 mg/kg CNO, Enzo Life Science, #BML-NS105-

0025) was intraperitoneally (i.p.) injected 30 min before the second conditioning session S2.

Stereotaxic viral injections

The following adeno-associated viral vectors (AAV) were used: AAV2/1-hSyn-DIO-hM4D(Gi)-mCherry,

(7.1E12 vg/mL, #44362 Addgene), AAV2/1-hsyn-DIO-mCherry (1.7E13 vg/mL, #50459 Addgene). Mice

were anaesthetized with Isoflurane (0.5–3.5% oxygen gas mixture 2 l/min) and buprenorphine (0.1mg

per kg of body weight) for stereotaxic injections. Viral injections were targeted unilaterally to aPVT

(coordinates: AP -0.4 mm; ML -0.6 mm; DV -3.25 mm, 10° angle; 200nl per site). The viral vectors were

delivered at a rate of 100nl/min using a glass micropipette lowered to the target site and that remained

for 5min after injection before it was withdrawn. Mice were tested for behavior 4 weeks after viral

injections. At least 7 days before testing, animals were singled caged to prepare them for the behavioral

experiment.

Behavioral procedures

We developed a 1-day aversive task (Conditioned Place Avoidance, CPA, task) based on a previously

published conditioned place preference task. [44,45] The apparatus contained two electric grids on the

floor of each context and consists of two square-walled (46 cm × 46 cm × 30 cm) contexts with distinct

inside building block configurations. The two contexts were connected via a bridge (8-cm length, 7-cm

width) during both pre-test and test sessions. The access to the bridge was blocked during each

conditioning session. All mice were handled for at least 3 days before the first experiment and, when

experiments involved viral injections, habituated to the injection procedure by mock injections during

these days.

During pretest, mice freely explored the two distinct contexts (e.g. one square and one circle) for 15

min and their baseline (spontaneous) preference for one of the two contexts was determined. Next,

mice were subjected to a first 10min-conditioning session (S1), in which 4 shocks (0.6 mA or 0.3mA, at

198 s, 278 s, 358 s and 438 s, of 2-s duration each, as in [110]) were delivered in their most preferred

context (Context A, "A+0.6"). Mice returned to their homecage at the end of session S1 for 30 min.

During the second 10min-conditioning session S2, mice received, in their least preferred enclosure

(Context B), either no shock (0mA, "B+0") or 4 shocks (of 0.6 mA for "B+0.6" or 0.3mA for "B+0.3" at

198 s, 278 s, 358 s and 438 s, of 2-s duration each). The control group (0 vs 0) went through the two

sessions S1 and S2 without receiving any shock. During the choice-test session 1 hour later without

any shock delivered, mice were placed in their most preferred context A (as defined during pre-test)

and were free to visit the two enclosures for 15 min after their first cross from context A to Context B.

Mice that displayed more than 75% preference during pretest were excluded for the rest of the

procedure.

Mice were tracked and several behavioral parameters were measured during pre-test, conditioning and

choice-test. A conditioned place preference score (CPA score) was calculated for each mouse during

both pre-test and choice-test as in [45]. Mice that did not cross to the non-preferred context B in less than

25 min were excluded from further analysis (i.e. they did not perform the following 15 min choice-test).

CPA was calculated as $CPA = \frac{t_{non\,preferred} - t_{preferred}}{t_{non\,preferred} + t_{preferred}}$. The first latency to cross was the time the animal

takes to first cross from the originally preferred context A (where they were first placed) to the originally

non-preferred context B. The second latency to cross was the time the animal takes to cross back to

the preferred context A. Time in the bridge was measured during the first latency (time in the bridge

before first cross) and during the first 5 min after the first cross. Freezing behavior was systematically

measured using a custom-made software (Imetronic) and was calculated as the percentage of time for

which mice remained immobile for more than 2 s. The percentage of time spent freezing during the

entire duration of the pretest and test session were reported. For the conditioning session, the

percentage of freezing before the 1st shock (0-3min) and after the 4th shock (8-10min) were calculated.

For Fig 1B, the difference between the total percentage of freezing in S2 minus the total percentage of

freezing in S1 was calculated for each group (total percentage of freezing for S1 and S2 used for this

calculation are presented in Fig. S1). All relative measurements (ΔCPA, ΔTime in bridge, etc) were

calculated by subtracting Choice-test behavioral measurements with the corresponding measurement

during the Pre-test. For example, ΔTime in the bridge was calculated as the time spent in the bridge

during the first 5 min after crossing during the choice test minus the time spent in the bridge during the

first 5 min of the pre-test session, ie. $\Delta Time\ bridge = Time\ bridge\ Choice_{0-5min} -$

$Time\ bridge\ Pretest_{0-5min}$. We quantified whether the animal engaged in turn back behavior (i.e. when

the whole body turns at 180deg) before 1st crossing from A to B when the animal was in the bridge.

Tail rattling was measured as a rapid vibration of the tail either after each shock delivered during

conditioning sessions S1 and S2 or before the first cross during the choice test. For experiments shown

in Fig. S6, animals from the 0-0 control group were animals that went through the 0.3-0.3 condition one

week before. A different set of contexts (C and D) were used in which the floor was plain and without

grids. For measuring the distance travelled post shock delivery as done in [111], we averaged for each

conditioning session the distance travelled for 8 seconds from the time of each shock epoch (ie. 198-

206 s, 278-286 s, 358-366 s, 438-446 s).

Brain slice electrophysiology

Patch-clamp recordings, in whole-cell configuration, were performed on PVT D2-labeled cells in current

clamp mode. Briefly, D2-Cre:Ai14 mice were anesthetized and perfused intracardially with ice-cold

dissection buffer (25 mM NaHCO3, 1.25 mM, NaH2PO4, 2.5 mM KCl, 0.5 mM CaCL2, 7 mM MgCl2,

25 mM D-glucose, 110 mM choline chloride, 11.6 mM ascorbic acid, 3.1 mM Pyruvic acid maintained

in 5% CO2/ 95% O2). Coronal brain slices (350μm) were cut in dissection buffer using a vibratome

(Campden Instruments). Sections were incubated in artificial cerebrospinal fluid (ACSF) containing the

following (in mM) 125 NaCl, 2.7 KCl, 11.6 NaHCO3, 0.4 NaH2PO4, 1 MgCl2, 2 CaCl2, 5.6 D-glucose

(pH 7.4, osmolarity 305 mOsm) at 32°C for 5 min and then kept in room temperature for at least 1h

before recording. For the experiment, slices were superfused continuously at 1.5 ml/min with an ACSF

solution. Recordings were obtained using a Multiclamp 700B amplifier (Molecular Devices). Recording

electrodes made of borosilicate glass had a resistance of 4-6 MΩ (Warner Instruments, USA) and were

filled with K-gluconate solution containing (in mM) 125 K-gluconate, 5 KCl, 10 Hepes, 1 EGTA, 5 Na-

phosphocreatine, 0.07 CaCl2, 2 Mg-ATP, 0.4 Na-GTP (pH 7.2, osmolarity 300 mOsm). Only cells with

access resistance < 20 MΩ and a change of resistance < 25% over the course of the experiment were

analyzed. Data were filtered with a Hum Bug (Quest Scientific, Canada), digitized at 2 kHz (Digidata

1444A, Molecular Devices, Sunnyvale, CA, USA), and acquired using Clampex 10 software (Molecular

Devices). Whole cell recordings were made from mCherry positive PVT D2 cells visualized with an

Olympus microscope (equipped with an mCherry filter). CNO was dissolved in ACSF (10μM, 30 s) and

applied in the bath after 10 min baseline recording. The number of action potentials were analyzed

before and after CNO application with Clampfit 10 software (Axon Instruments).

Statistical Analysis

Data were shown as the mean ± SEM (standard error of the mean). GraphPad Prism v7.0 software was

used for statistical analyses. D'Agostino Pearson test is used as a test for normality. When this

assumption was not met, non-parametric tests were used (see Table S1). Student's t-test (paired or

unpaired, 2-sided) was used to compare two sets of data. Multiple comparisons were performed by 1-

way or 2-way ANOVA followed by either Sidak's, Tukey's, Uncorrected Fisher's LSD or Dunn's multiple

comparisons were used for *post-hoc* analysis. For comparisons against 0, either one sample t test or

Wilcoxon signed rank test (for non-normal data) were used. All statistical comparisons are presented in

Table S1.

In Fig. 4, 1 mCherry animal was excluded for not crossing, 2 hM4Di and 1 mCherry animals were

excluded for viral injections outside PVT. In Fig. 5, 1 hM4Di and 1 mCherry animal were excluded for

not crossing during the choice-test. In Fig. 6, 1 hM4Di animal was excluded for viral injections outside

PVT. In Fig. S6, 2 animals were excluded for spending more than the 75% criterion in one compartment

during Pretest.

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AUTHOR CONTRIBUTIONS

M.M., E.P. and S.T. conceived the project and designed all the experiments. M.M. performed surgical

and anatomical experiments. M.M., L.V., E.K. and M.D. performed behavioral experiments. P.T and

J.S. performed and analyzed electrophysiological experiments. The manuscript was written by M.M.,

E.P. and S.T and commented by all authors.

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FIGURE LEGENDS

Figure 1. Mice adapt their defensive behavioral responses to safe and aversive conditioning.

(A) Schema of the 1-day conditioned place aversion (CPA) task for the control, safe and aversive

groups. Contexts A (square) and B (circle) are connected via a bridge and contain electric shock grids.

During conditioning session S1, context A is paired with either 0 mA or four 0.6 mA electric shocks.

During conditioning session S2 (30min after S1), context B is paired with either 0 mA or four 0.3 mA

electric shocks.

(B) Only mice from the aversive group increased their total freezing level during conditioning session

S2 compared to S1.

(C) Safe and aversive groups, but not the control group, increased freezing before the 1st shock of the

conditioning session S2 compared to S1.

(D) Freezing after the 4th shock significantly decreased for the safe group between conditioning

sessions S1 and S2 while it was maintained for the aversive group.

(E) Only the aversive group exhibited tail rattling behavior immediately after the shocks delivered during

the conditioning session S2.

All data are mean ± SEM. Individual data points are displayed as dots. Control (n=11); safe (n=11);

aversive (n=12); *p<0.05; **p<0.01; ***p<0.001; ##p<0.01 against 0, ns: non-significant.

See also Figure S1.

Figure 2. Mice use prior knowledge and display specific behavioral signatures to make

appropriate safe and aversive choices.

(A) Percentage of time mice spent in the bridge during the choice-test before and after their 1st cross

from context A to context B for the control, safe and aversive groups.

(B) Mice from the aversive group spent significantly more time in the bridge before their 1st cross from

context $A_{0.6}$ to context $B_{0.3}$ than the control group.

(C) Mice from the aversive group made significantly more turnback in the bridge before making their 1st

cross from context $A_{0.6}$ to context $B_{0.3}$ than the control and safe groups.

(D) After their 1st cross to context B, mice from the safe and aversive groups, but not from the control

group, reverted their initial preference (measured during pretest) during the 15 min choice-test, by

spending more time in the safest context B as indicated by a positive ΔCPA score (CPA score during

the choice test minus CPA score during the pretest).

(E) Mice from the aversive group had a specific pattern of freezing during the entire choice-test

compared to that of the safe and control groups in the context A (left), bridge (center) and context B

(right). The aversive group froze more in the bridge than the safe and control groups. These mice also

froze more in context B than the control group.

(F) Only mice from the aversive group spent more time in the bridge during the choice test - pretest (Δ

Time) after their 1st cross from context A to context B.

(G) Mice from both safe and aversive groups spent more time than the control group to cross back from

context B to context A during the choice test (i.e. 2nd latency).

All data are mean ± SEM. Individual data points are displayed as dots. Control (n=11); safe (n=11);

aversive (n=12); *p<0.05; **p<0.01; ***p<0.001. ## p<0.01 and ### p<0.001 against 0; ns: non-

significant.

See also Figure S2.

Figure 3. D2-expressing neurons (D2+) in the anterior PVT are preferentially recruited during

both safe and aversive conditioning.

(A) Schema of the 1-day conditioned place aversion (CPA) task for the control home cage (HC), safe and aversive groups. Mice were sacrificed 90 min after the conditioning session S2 to assess cFos expression in PVT brain sections.

(B) Schematics and representative image of the anterior PVT (aPVT, antero-posterior axis AP: -0.34) showing examples of D2+ (in red) and cFos+ (in white) cells. Scale bar: 100 μm.

(C) Both safe and aversive conditioning led to a significant increased cFos density (number of neurons per mm^2) in the aPVT compared to that of the pPVT.

(D) D2+ neurons in the aPVT were preferentially recruited after both safe and aversive conditioning compared to home cage control mice.

(E-F) The recruitment of D2+ (cFos+D2+ among D2+ cells), but not D2- (cFos+D2- among D2- cells), neurons during aversive conditioning was negatively correlated with the time spent in the bridge during the first 5 minutes of the choice test after the 1st cross. Δ time in the bridge = time in bridge during the choice test minus pretest for the first 5 min after the 1st cross from context A to context B for the aversive (up) and safe (down) groups. Line represents linear regression with confidence interval.

All data are mean ± SEM. Individual data points are displayed as dots. Home cage control (n=8); safe (n=10); aversive (n=10); *p<0.05; **p<0.01.

See also Figure S3.

Figure 4. D2-expressing neurons are required for aversive evaluation and choice.

(A) D2-Cre mice were injected in the aPVT with either DIO-mCherry or DIO-hM4Di-mCherry viral vector (left panel). Representative image of mCherry expression in the aPVT (right panel). Scale bar: 100 μm.

(B) Schema of the 1-day condition place aversion (CPA) task for the aversive group. Control mCherry and hM4Di mice were i.p. injected with CNO (2mg/kg) 30 min before the conditioning session S2 in Context B$_{0.3}$ and were tested 1h later during a choice-test.

(C) hM4Di mice exhibited higher tail rattling after each shock of the conditioning session S2.

(D) While the distance travelled immediately after shock decreased during the conditioning S2 compared to S1 for mCherry control mice, hM4Di mice did not show such reduced distance during S2.

(E) hM4Di mice showed increased turnback in the bridge before their 1st cross from context A$_{0.6}$ to context B$_{0.3}$.

(F) hM4Di mice exhibited increased tail rattling behavior before their 1st cross from context $A_{0.6}$ to

context $B_{0.3}$.

(G) ΔCPA score (CPA score during the choice test - CPA score during the pretest) was significantly

decreased for hM4Di mice during the 15 min choice test, with mice preferring none of the 2 contexts.

(H) hM4Di mice spent significantly more time in the bridge after their 1st cross from context $A_{0.6}$ to

context $B_{0.3}$.

(I) hM4Di mice significantly decreased their freezing during the 15 min choice test in context $B_{0.3}$. All

tests are comparisons between mCherry control and hM4Di for each compartment.

(J) hM4Di mice significantly decreased their 2nd latency to cross back from context $B_{0.3}$ to context $A_{0.6}$

during the choice test.

All data are mean ± SEM. Individual data points are displayed as dots. mCherry control (n=8); hM4Di

(n=8); *$p<0.05$; **$p<0.01$; ***$p<0.001$; ###: $p<0.001$ difference from 0; ns: non-significant.

See also Figure S4.

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Figure 5. Inhibiting aPVT D2 neurons increases contextual aversiveness and related threat-

predicting behavioral traits.

(A) Schema of the 1-day condition place aversion task in which both A and B contexts are paired with

0.3mA. Control mCherry and hM4Di mice were i.p. injected with CNO (2mg/kg) 30 min before the

conditioning session S2.

(B) hM4Di mice displayed a greater number of tail rattling during conditioning session 2 compared to

mCherry control mice.

(C) Freezing changes between S1 and S2 after the 4th shock tend to be similar between hM4Di and

mCherry mice.

(D) hM4Di displayed more turnback in the bridge before the 1st cross from context $A_{0.3}$ to context $B_{0.3}$

than mCherry control mice.

(E) hM4Di and mCherry control mice showed a similar 1st latency to cross from context $A_{0.3}$ to context

$B_{0.3}$.

(F) hM4Di mice and mCherry control mice spent a similar amount of time in the bridge before their 1st

cross between context $A_{0.3}$ to context $B_{0.3}$.

(G) hM4Di and mCherry control mice froze similarly in the context $A_{0.3}$, bridge and $B_{0.3}$ during the choice

test. All tests are comparisons between mCherry control and hM4Di for each compartment.

(H) ΔCPA score (CPA score during the choice test - CPA score during the pretest) during the 15 min

choice test is significantly lower and not different from 0 in hM4Di mice compared to that of mCherry

control mice.

All data are mean ± SEM. Individual data points are displayed as dots. mCherry control (n=8); hM4Di

(n=7); (*) p=0.0565; *P<0.05; ***p<0.001; [#]P<0.05; ns: non-significant.

See also Figure S5.

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Figure 6. Inhibiting aPVT D2 neurons increases contextual safeness and related safety-

predicting behavioral traits.

(A) Schema of the 1-day condition place aversion task for the safe group. Control mCherry and hM4Di

mice were i.p. injected with CNO (2mg/kg) 30 min before the conditioning session S2.

(B) hM4Di mice did not change the number of tail rattling during conditioning.

(C) hM4Di and control mCherry mice similarly showed a decreased distance after shock between S1

and S2.

(D) hM4Di and control mCherry mice showed a similar number of turnback in the bridge before 1st cross

from context $A_{0.6}$ to context B_0.

(E) hM4Di and control mCherry mice exhibited a similar number of tail rattling before 1st cross from

context $A_{0.6}$ to context B_0.

(F) hM4Di mice did not change the 1st latency to cross from context $A_{0.6}$ to context B_0.

(G) hM4Di mice significantly increased their 2nd latency to cross back from the context B_0 to the context

$A_{0.6}$ during the choice test.

(H) During the choice test, hM4Di mice significantly decreased freezing in context $A_{0.6}$, increased their

freezing in context B_0, with no change in freezing in the bridge. All tests are comparisons between

mCherry control and hM4Di for each compartment.

(I) hM4Di mice significantly increased their ΔCPA score (CPA score during the choice test - CPA score

during the pretest) during the 15 min choice test.

All data are mean ± SEM. Individual data points are displayed as dots. mCherry control (n=8); hM4Di

(n=8); [##]P<0.01 against 0; [###] p<0.001 against 0; *p<0.05; **p<0.01; ns: non-significant.

See also Figure S6.

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Figure 1

Figure 2

A 1st cross

B Choice test before 1st cross

C Choice test before 1st cross

D Choice test after 1st cross

E Choice test

F Choice test after 1st cross

G Choice test after 1st cross

Figure 3

A

control home cage (HC)

Pretest   Conditioning   Choice test
         S1   S2

safe   A → B → A+0.6 → B+0 → A$_{0.6}$ → B$_0$ →

aversive   A → B → A+0.6 → B+0.3 → A$_{0.6}$ → B$_{0.3}$ →

perfusion & cFos immunostaining

B

aPVT   D2-Cre:Ai14

D2   cFos   aPVT

C

- control HC
- safe
- aversive

cFos cells per mm^2 (normalized to HC)

**

aPVT   pPVT

D   **aPVT**

- control HC
- safe
- aversive

% cFos+ (normalized to HC)

**

D2+   D2-

E

aPVT   aversive

% cFos+ (normalized to HC)

D2+: R^2=0.423 *
D2-: R^2=0.045

ΔTime in the bridge (0-5min)

pPVT   aversive

% cFos+ (normalized to HC)

D2+: R^2=0.247
D2-: R^2=0.001

ΔTime in the bridge (0-5min)

F

aPVT   safe

% cFos+ (normalized to HC)

D2+: R^2=0.110
D2-: R^2=0.021

ΔTime in the bridge (0-5min)

pPVT   safe

% cFos+ (normalized to HC)

D2+: R^2=0.067
D2-: R^2=0.015

ΔTime in the bridge (0-5min)

Figure 4

A DIO-mCherry or DIO-hM4Di-mCherry — aPVT — D2-Cre mice

mCherry / DAPI / aPVT

B Pretest · Conditioning S1 S2 · Choice test

CNO i.p. 2mg/kg

mCherry or hM4Di

A — B → A+0.6 → B+0.3 → $A_{0.6}$ — $B_{0.3}$

C Conditioning — Number of tail rattling — S1 S2 mCherry / S1 S2 hM4Di — ** / ***

D Conditioning — Distance after shock (cm) — S1 S2 mCherry ** / S1 S2 hM4Di ns

E Choice test before 1st cross — Turnback in bridge — mCherry / hM4Di **

F Choice test before 1st cross — Number of tail rattling — mCherry / hM4Di *

G Choice test after 1st cross — ΔCPA score — mCherry / hM4Di *** ###

H Choice test after 1st cross — Time in bridge (0-5min) — mCherry / hM4Di *

I Choice test — % Freezing — $A_{0.6}$ Bridge $B_{0.3}$ — mCherry / hM4Di — *

J Choice test after 1st cross — 2nd latency from $B_{0.3}$ to $A_{0.6}$ (min) — mCherry / hM4Di *

Figure 5

A Pretest   Conditioning S1 S2   Choice test

B Conditioning

C Conditioning after 4th shock

D Choice test before 1st cross

E Choice test before 1st cross

F Choice test before 1st cross

G Choice test

H Choice test after 1st cross

Figure 6

A Pretest   Conditioning   Choice test
     S1   S2

CNO i.p.
2mg/kg

mCherry
or
hM4Di

A — B → $A+0.6$ → $B+0$ → $A_{0.6}$ — B_0

B Conditioning
Number of tail rattling
S1 S2 (mCherry)   S1 S2 (hM4Di)

C Conditioning
Distance after shock (cm)
* S1 S2 (mCherry)   ** S1 S2 (hM4Di)

D Choice test before 1st cross
Turnback in bridge
mCherry hM4Di

E Choice test before 1st cross
Number of tail rattling
ns
mCherry hM4Di

F Choice test before 1st cross
1st latency from $A_{0.6}$ to B_0 (min)
ns
mCherry hM4Di

G Choice test
2nd latency to cross from B_0 to $A_{0.6}$ (min)
*
mCherry hM4Di

H Choice test
%Freezing
mCherry hM4Di
* $A_{0.6}$ Bridge   * B_0

I Choice test
ΔCPA score
*

mCherry hM4Di