Actinobacteriophage Inteins: Host Diversity, Local Dissemination, and

Non-Canonical Architecture

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Abstract

Intein presence within Actinobacteriophages (within PhagesDB) was last surveyed in 2016, and despite a 5-fold increase in the size of the database, has not been updated since. To address this, we present a modern survey of the current iteration of the PhagesDB database. We developed a new algorithm — Iterative Cluster Expansion BLAST (ICE-BLAST) — to expand our search to more divergent sequences. Nearly 800 inteins were retrieved through this process; the majority of which were previously unreported. We describe the nature of these inteins, their classes, integration target sites, distribution within phage clusters, and explore the geographical location of nearly identical intein sequences found in divergent exteins. Our findings suggest that these inteins recently invaded local phage populations. We also find two instances of a Cas4 exonuclease intein evolving from a terminase large subunit intein, and propose a model by which one of these inteins was able to utilize sequence similarity conferred by a shared nucleotide binding site to jump between genes. Additionally, we find inteins with never-before-reported homing endonucleases, and inteins with homing endonucleases encoded in a reading frame separate from that which encodes the extein and the intein's self-splicing domain. We provide predicted structures for these elements and hypothesize on their evolution and relation to free-standing homing endonucleases within phage genomes. Finally, we provide evidence that these "non-canonical" inteins are still transferring between host genomes, in a fashion similar to other inteins with canonical homing endonucleases within the dataset.

Introduction

Inteins are self-splicing, site-specific, mobile genetic elements that invade protein-encoding genes across all domains of life and viruses, including bacterial viruses (phages) (Gogarten et al. 2002; Lennon and Belfort 2017; Nanda et al. 2020). An intein begins its journey as an intervening DNA sequence at a specific insertion site within the coding sequence of a host gene. Unlike the intervening DNA sequences known as introns which are post-transcriptionally removed at the RNA level, an intein is transcribed and translated together with its host gene. The intein then seamlessly excises itself post-translationally from the host protein, referred to as the extein, through the unique process of protein splicing (Shih et al. 1988; Hirata et al. 1990; Kane et al. 1990). Protein splicing, performed by the self-splicing domain of the intein, yields a functional extein and the free intein. Inteins are divided into three classes depending on their initial steps of splicing, the residues involved, and differences in reaction chemistry (Tori et al. 2010; Nanda et al. 2020).

Once excised from the extein, the intein uses its homing endonuclease (HEN) domain to engage in the self-propagation method known as homing (Goddard and Burt 1999). When in the presence of a copy of its host gene lacking the intein's DNA sequence at its particular insertion site, the HEN makes a double-strand break at the intein insertion site. Through homologous recombination-based repair using the intein-containing copy of the gene as the template, the intein sequence is copied into the previously vacant insertion site (Belfort and Roberts 1997). Homing allows inteins to rapidly rise to saturation in populations, largely through horizontal transfer-based homing, at which point there is no longer selective pressure to maintain the HEN (Goddard and Burt 1999). The HEN degrades, turning the full intein into a mini intein which contains only the self-splicing domain (Iwaï et al. 2017). Sometimes, an intein may acquire an internal stop codon in the HEN as it degrades, generating a split intein (Tavassoli and Benkovic 2007; Dassa et al. 2009; Hoffmann et al. 2021). Eventually, the mini and split inteins are lost and the intein-free allele once again dominates. Newer models propose the co-existence of full, mini, split, and empty alleles as opposed to synchronized progression through the phases (Gogarten and Hilario 2006; Yahara et al. 2009; Barzel et al. 2011). As an alternate evolutionary endpoint to degradation and loss, some inteins have also been inferred to have acquired novel functions, acting as regulatory

elements under stressful conditions (Novikova et al. 2016; Lennon et al. 2018) or evolving into mating-

type switching endonucleases such as HO of *Saccharomyces cerevisiae* (Coughlan et al. 2020).

Several different HEN domain types have been reported in inteins. Most inteins possess

canonical LAGLIDADG homing endonucleases, which are in-frame with the rest of the intein (Gorbalenya

1998; Gimble 2000). Others include: inteins with an in-frame HNH endonuclease (Dalgaard, Klar, et al.

1997), split-inteins possessing VSR-like endonucleases (Dassa et al. 2009), and LAGLIDADG HENs

encoded on the anti-sense strand (Gorbalenya 1998; Gogarten et al. 2002). Interestingly, most of these

non-canonical inteins are found within phage genomes, with actinobacteriophages serving as a particular

hot-spot in research.

Actinobacteriophages are promising therapeutic alternatives to antibiotics in the treatment of

multi-drug resistant actinobacterial infections (Allué-Guardia et al. 2021; Zeynali kelishomi et al. 2022;

Dedrick et al. 2023; Yang et al. 2024). However, much is still unknown about actinobacteriophage

structure, function, and genomic evolution, necessitating increased studies in these areas. In recent

years, teaching programs (SEA-PHAGES and PHIRE) employing citizen-science based approaches to

isolate and sequence actinobacteriophages have added thousands of genomes to databases such as

PhagesDB (Russell and Hatfull 2017). PhagesDB thus serves as an ideal candidate for large-scale

computational analyses aiming to dissect phage genomic evolutionary dynamics, with inteins serving as

particularly powerful tools for tracing genetic exchange between phage populations (Soucy et al. 2014).

Inteins are present within actinobacteriophages (Tori et al. 2010; Tori and Perler 2011; Kelley et

al. 2016). However, since the last survey of these inteins (Kelley et al. 2016) the PhagesDB database has

expanded from 841 to 5075 genomes; providing a unique resource to study the distribution and evolution

of phage inteins. Hence, we conducted a new survey of PhagesDB, accessed in September of 2023. To

capture more divergent inteins within the database, we developed an algorithm, ICE-BLAST, which uses

a combination of PSI-BLAST and USEARCH clustering to iteratively search an amino acid database

using the representative sequences of clusters (centroids) of matches from previous searches. Herein we

discuss the development and testing of this algorithm, and the inteins discovered through its application

to PhagesDB. We investigate intein distribution within phage clusters, variation in host genes, and

geographic dissemination. Through the latter, we found evidence supporting events of rapid

geographically localized propagation of intein alleles into new populations. Our phylogenetic analyses

also uncovered two occurrences of class 3 inteins of terminase large subunits sharing a common

ancestor with Cas 4 exonuclease inteins. Additionally, we report several cases of inteins with HEN

domains and architectures that have rarely been reported on, or were previously unseen. These include

HENs encoded within a separate ORF on the same strand as the intein, and HENs in a separate ORF on

the anti-sense strand.

Results

Intein Distribution across Actinobacteriophage Clusters

Using our program ICE-BLAST, tested using simulated data from our program Invader-Sim, we identified 784 putative intein sequences in PhagesDB (details on the intein retrieval and validation process are provided under Materials and Methods). A table containing information on all 784 inteins including their host phage, host protein (extein), splicing class, and HEN type is available (Supplemental Table 1). These 784 inteins span a wide variety of PhagesDB phage clusters (Figure 1), with the majority being in clusters C1 (415), A1 (92), and E (68), all of which are listed as mycobacteriophage clusters. The other 209 inteins were distributed across 31 phage clusters and several singleton phages (phages not assigned to a cluster). Of the 31 phage clusters, the maximum number of inteins observed in one cluster was 35, and the majority of clusters contained less than 10 inteins. The most intein-rich genomes were all from cluster C1 phages, with phage Cane17 having seven inteins, followed by phages Blubba and Colt with six inteins each.

Figure 1. Total number of inteins found in each PhagesDB phage cluster, depicted with (left) and without (right) intein-rich clusters A1, C1, and E. A1, C1, and E are depicted with dark gray bars and contained 92, 415, and 68 inteins respectively. Singletons are denoted as Si*.

Wide Range of Actinobacteriophage Proteins Invaded by Inteins

By re-annotating the host protein sequences of the 784 putative inteins, we found that they invade 19 distinct proteins (Figure 2). Unsurprisingly, the majority were found in proteins previously reported as containing inteins. These include: Cas4 exonuclease (InterPro IPR013343), DNA helicase

(InterPro IPR006500), DNA methylase N-6/N-4 (Pfam PF01555) and C-5 (InterPro IPR001525), DNA

primase (InterPro IPR006171), nucleotidyltransferase (InterPro IPR018775), phosphoesterase (InterPro

IPR029052), portal protein (InterPro IPR009279), terminase large subunit (Pfam PF04466), and

thymidylate synthase (InterPro IPR003669). The remaining host proteins are new to this study within

actinobacteriophages, and include: DNA polymerase III alpha subunit (InterPro IPR011708), minor capsid

protein (InterPro IPR006528), ribonucleotide reductase subunits alpha (InterPro IPR026459) and beta

(InterPro IPR026494), RtcB-like RNA ligase (InterPro IPR021122), tape measure protein (InterPro

IPR013491), and tRNA splicing RNA ligase (InterPro IPR036025). Two types of intein-containing exteins

identified could not be definitively annotated, but have best matches to baseplate wedge domain

containing proteins (UniProt M9MUF4) and integrin-like proteins respectively. Based on the annotations

of other closely related intein-containing exteins, the former could be a portal protein of some sort, but

does not share considerable similarity to known portal proteins in any NCBI database. The inteins present

within the putative integrin-like exteins were previously reported on in Kelley et al. 2016, but with fewer

intein-containing exteins (Kelley et al. 2016).

Variation in Intein Splicing Domains

The putative inteins were clustered at 70% sequence identity. For each intein cluster, we

determined the intein splicing class (1, 2, or 3). Most of the inteins were class 1 (497) while the remainder

(287) were class 3. The class 1 inteins displayed a much wider range of host proteins (16 different

proteins) than the class 3 inteins which were only found within 4 different extein hosts. More than half of

the inteins within our dataset (431) invaded just two proteins, with 293 inteins within terminase large

subunit sequences, and 138 within Cas4 exonucleases (Figure 2). The size of class 1 inteins ranged from

385 amino acids (aa) (terminase large subunit of cluster DC2 phage Clown) to 134 aa (terminase large

subunit of cluster EN phage C3PO). The size of class 3 inteins ranged from 359 aa (DNA helicase of

cluster C1 phage Alice) to 225 aa (minor capsid of singleton phage REQ3). In addition to classifying the

splicing domain of each intein, we classified their HEN domains, which revealed further diversity and

previously undocumented intein architectures.

Figure 2. Identified inteins, which fall into class 1 and class 3 based on their splicing domains, are distributed across 19 different host genes: baseplate wedge domain protein (InterPro M9MUF4), Cas4 exonuclease (InterPro IPR013343), DNA helicase (InterPro IPR006500), DNA methylase N-6/N-4 (Pfam PF01555) and C-5 (InterPro IPR001525), DNA polymerase III alpha subunit (InterPro IPR011708), DNA primase DnaG-like (InterPro IPR006171), putative integrin-like protein, minor capsid protein (InterPro IPR006528), nucleotidyltransferase DNA polymerase beta superfamily (InterPro IPR018775), phosphoesterase (InterPro IPR029052), portal protein (InterPro IPR009279) ribonucleotide reductase alpha subunit (InterPro PR026459) and beta subunit (InterPro IPR026494), RtcB-like RNA ligase (InterPro IPR021122), tape measure protein (InterPro IPR013491), terminase large subunit (Pfam PF04466), thymidylate synthase (InterPro IPR003669), and tRNA-splicing ligase (InterPro IPR036025).

Variation in Intein Homing Endonuclease (HEN) Domains

Most of the inteins in our database (712) possess classic intein architecture: a self-splicing domain and a canonical LAGLIDADG (InterPro IPR004860) HEN domain. Of the remaining 72 inteins, nine have decaying or completely degraded HENs (mini inteins), and 63 possess unique non-canonical HENs. These non-canonical HENs include in-frame HNH (InterPro IPR002711) HENs (Figure 3A),

LAGLIDADG or VSR-like (InterPro IPR004603) HENs encoded on the anti-sense strand (Figure 3B-C),

and out-of-frame endonuclease VII-like (InterPro IPR004211) HENs (Figure 3D). The inteins possessing

these non-canonical HENs are distributed across four host protein types: DNA polymerase, minor capsid,

terminase large subunit, and thymidylate synthase (Figure 3E). Based on current knowledge, these four

non-canonical HEN architectures are rare, with some being reported on for the first time with this work.

The non-canonical HENs which have already been reported on include the in-frame HNH HEN which is

observed in phages (Dalgaard, Klar, et al. 1997; Dalgaard, Moser, et al. 1997), the anti-sense

LAGLIDADG HENs (Gorbalenya 1998; Gogarten et al. 2002), and the VSR-like HEN which was reported

in the context of split inteins with no other known intein-associated reports (Dassa et al. 2009). The fourth

type of non-canonical HEN we found, an endonuclease VII-like HEN encoded within the HEN region but

in frame 3 relative to the rest of the intein, has not yet been reported to our knowledge.

169   To further assess the identities of the out-of-frame and anti-sense non-canonical HENs,

AlphaFold-predicted structures of the inteins (Supplemental Figure 1) and the HENs in their respective

frames (Supplemental Figure 2) were compared to solved structures. Based on the predicted structures

for the endonuclease VII-like and anti-sense LAGLIDADG HENs, coupled with the fact that these HENs

can be dimeric, the terminal domains of these structures may be linker domains or secondary DNA

binding domains (Raaijmakers et al. 1999; McMurrough et al. 2018). The VSR-like HEN appears to

contain a long stretch of linked turns and alpha helices not present in normal VSR endonuclease

structures (Hennecke et al. 1991). The predicted local distance difference test (pLDDT) — a per-residue

measure of local prediction confidence (Jumper et al. 2021) — for this stretch is relatively high, especially

within helices, but the region lacked meaningful matches against PDB structures. However, an HHPred

search found a significant match (e-value of 0.000046) against protein structure 3R3P from Bacillus

phage 0305phi8-36, which indicates the VSR-like HEN may be an EDxHD HEN (Taylor et al. 2011).

Figure 3. A-D. Gene-map depictions of the four observed non-canonical intein HEN organizations.

Regions shaded with diagonal lines indicate HEN-encoding region. **A.** in-frame HNH HEN. **B.** anti-sense

LAGLIDADG HEN. **C.** anti-sense VSR-like HEN. **D.** endonuclease VII-like HEN in reading frame 3 relative

to intein. **E.** Distribution of inteins with canonical and non-canonical HEN architectures across host protein

types.

Actinobacteriophage Intein Phylogenetics

Having characterized the structural architectures of each intein, we sought to investigate how

these inteins have evolved with respect to their host proteins and host phages. Performing such

evolutionary characterizations of inteins can often unveil pathways for genetic exchange within

populations that may have gone unnoticed when focusing on genetic material with primarily orthodox

vertical inheritance (Soucy et al. 2014; Turgeman-Grott et al. 2023). Thus, we constructed phylogenies

from the DNA sequences of all inteins in our database. The extracted subsets of intein sequences used to

generate these phylogenies were separated by HEN type, with the largest set (canonical HENs) split into

two sets based on splicing class (class 1 or class 3). By analyzing the grouping patterns of inteins within

the phylogenies compared to the types of host proteins they invade, potential instances of intein host

protein switching (e.g. a terminase large subunit-targeting intein evolving into a Cas4 exonuclease-

targeting intein) can be identified.

While some inferences could be drawn from the phylogenies of the inteins with non-canonical

HEN domains, current sampling is ultimately too sparse to make any definite claims from the intein

phylogenies alone (Supplemental Figures 3-6). In contrast, the inteins with canonical HENs produced

more informative phylogenies.

The class 1 inteins with a canonical LAGLIDADG HEN produced a phylogeny which provided

evidence for several independent events of intein integration site diversification events (Figure 4). For

example, the terminase large subunit inteins form a polyphyletic clan with at least four separate groups.

Each group is separated, divided from the other terminase large subunit groups by multiple highly

supported splits along the inner nodes. Similarly, portal protein inteins and DNA methylase inteins form

polyphyletic clans with five and three separate groups respectively.

The class 3 inteins with canonical LAGLIDADG HENs are almost entirely found within Cas4

exonuclease and terminase large subunit exteins (Figure 4). Within this phylogeny are two instances of

Cas4 associated inteins diverging from terminase large subunit inteins. One of these groupings had

particularly high bootstrap support, and was thus further investigated as a potential case of extein

switching (an intein evolving to target an insertion site inside of a new type of host protein).

Figure 4. The DNA sequences of the inteins with canonical LAGLIDADG HEN domains were split into two subsets by splicing class (class 1 and class 3), with each subset then used for maximum likelihood phylogenetic reconstruction. Substitution models used were TIM2+F+R3 (class 1) and TPM2+F+R4 (class 3). Ultrafast bootstrap support is listed on internal nodes. Scale bar measures substitutions per site. Clans of inteins within the same extein are collapsed; clade labels indicate the number of inteins in the collapsed group, and which phage clusters they are found in. Branches leading to terminase large subunit inteins are colored in blue, and branches leading to Cas4 exonuclease inteins are colored in red. On the class 3 phylogeny, the potential extein switching event between terminase large subunit and Cas4 exonuclease inteins (involving phages LittleE and ScoobyDoobyDoo) further investigated is indicated with a purple circle.

Extein Switching

The class 3 intein phylogeny (Figure 4) depicts two instances of a clade of Cas 4 exonuclease

inteins grouping inside of a clade of terminase large subunit inteins. This grouping suggests that an intein

which now targets an insertion site in Cas4 exonucleases evolved from an intein which targets an

insertion site in terminase large subunits. We will refer to this type of evolutionary event as extein

switching. To assess whether this extein switching topology was a phylogenetic artifact, an

approximately unbiased (AU) tree topology test (Shimodaira 2002) was used to gauge the strain induced

when one attempts to force the terminase large subunit and Cas4 exonuclease inteins to group by extein

type, as opposed to the observed topology. The same model determined by IQ-TREE's ModelFinder for

the original class 3 intein phylogeny (WAG+F+R4) was used for the AU test. The test produced little

support for the grouping of these inteins by extein type, (bootstrap proportions of 0.998 and 0.002 for tree

constrained by extein switching event and the tree constrained by extein type respectively (Supplemental

Table 2)). After investigating both putative extein switching events, the event involving phages LittleE and

ScoobyDoobyDoo provided a more complete picture of how the extein switching may have occurred. The

terminase large subunit inteins of the other extein switching event (involving phage Frankenweenie

(Figure 4)) all exhibited degradation in their HEN domains, making comparison of predicted intein

structures for this group less informative. Thus, the event involving phages LittleE and ScoobyDoobyDoo

will be the focus of the presented investigation that follows.

Extein Switching from Terminase Large Subunit to Cas4 Exonuclease

Comparing the +/-30 nucleotides at the intein insertion site across the LittleE terminase large

subunit and ScoobyDoobyDoo (SDD) Cas4 exonuclease reveals regions of striking similarity

(Supplemental Figure 7A). Both insertion sites reside in stretches of DNA sequence which encode

important nucleotide binding sites for their respective proteins (Supplemental Figure 7B). Comparison to

the solved structure of a phage terminase large subunit bound to ATP (Zhao et al. 2013) shows the intein

insertion site encodes a region on an alpha-helix within the Rossmann fold constituting the nucleotide

binding site within the ATPase domain of the terminase large subunit. The SDD Cas4 exonuclease intein

insertion site encodes the motif EFKT, which is the RecB-like motif-III in the Cas4 exonuclease, and is

important for nucleotide binding (Shiimori et al. 2018). In addition to assessing insertion site sequence

similarity, we compared the predicted protein structures for the inteins. Alignment of the predicted

structures generated for the terminase large subunit intein of LittleE, the Cas4 exonuclease intein of SDD,

and the terminase large subunit intein of cluster DE4 phage APunk (representing a clade of terminase

large subunit inteins which do not group with LittleE's terminase large subunit intein) reveals them all to

be near identical, barring APunk (Figure 5). These results align with the expected results based on the

phylogeny. The APunk intein does not align as strongly to the LittleE and SDD inteins as they do to each

other, specifically exhibiting a slight structural difference in its HEN (Figure 5). This difference in HEN

structure between the APunk and LittleE terminase large subunit inteins in particular reflects how they

target two separate regions of the terminase large subunit gene (both intein insertion site regions encode

alpha-helices in the ATPase domain, but two distinctly different helices). In contrast, the high similarity

between the HEN domains of the SDD and LittleE predicted intein structures further supports the notion

that these inteins can recognize and cut an extremely similar intein insertion site. To explain the lack of

intein in SDD's terminase large subunit, we later propose a model in which the SDD terminase large

subunit was not a viable host for the LittleE terminase large subunit intein, and the SDD Cas4

exonuclease provided a viable alternate insertion site.

Figure 5. Subtree from phylogeny of class 3 inteins (Figure 4) depicting one instance in which a Cas4 exonuclease intein emerges from a group of terminase large subunit inteins. For each intein indicated with a star (cluster DE4 representative phage APunk terminase large subunit intein, LittleE terminase large subunit intein, and ScobyDoobyDoo (SDD) Cas4 exonuclease intein) a predicted protein structure was generated with AlphaFold. The predicted intein structures (right) are APunk (top) LittleE (middle) and SDD (bottom). Alignments of the predicted intein structures using the Matchmaker tool in ChimeraX produce the following results: LittleE aligned to SDD yields a sequence alignment score of 995.6, and RMSD of 2.716; APunk aligned to LittleE yields a sequence alignment score of 644.9, and RMSD of 3.097; APunk aligned to SDD yields a sequence alignment score of 643.1, and RMSD of 2.994.

Intein Dissemination within Local Areas

Typically, copies of a particular intein-invaded extein in different individuals will have higher

sequence similarity than the inteins that invade them. This is due to the types of genes invaded by inteins

often being of vital function to their hosts and thus highly conserved, as well as the increased evolutionary

rates experienced by homing elements such as inteins (Goddard and Burt 1999). We further discuss the

relative sequence conservation and selection pressures on intein and extein sequences in Supplemental

Materials under the section Supplemental Results and Discussion: Sequence Conservation in Inteins and

Exteins. Thus, two individuals having less similar exteins but nearly identical inteins is cause for further

investigation, as this can indicate a recent transfer event since the intein has not yet had time to diverge

from its origin copy despite the higher evolutionary rates of inteins. Geographic data regarding host phage

isolation paired with these events can be further investigated to assess the locality of such recent

invasion events.

Following this line of analysis, we previously found inteins which appeared to be proliferating into

divergent phage extein sequences within local geographic areas (Gosselin et al. 2023). We chose to

investigate our dataset for additional cases of local dissemination and proliferation of inteins into new

phage populations. We filtered our data in two stages: first to find clusters of closely related inteins

(97.5% similarity), then to discover clusters within the filtered set whose extein sequences were more

divergent from each other than their respective intein sequences. These clusters were used to construct

phylogenetic trees, and their geographic positions were plotted using metadata from PhagesDB. Of the

resulting 13 clusters of intein sequences, we focused on three cases which displayed high levels of

incongruence between the extein and intein phylogenies during a visual inspection, suggesting recent

intein transfer between divergent hosts.

First, we investigated a set of inteins which were found in nucleotidyltransferases (Figure 6). This

group of inteins had a global distribution, and of particular interest to us, two groups of co-localized

inteins. One group of 5 inteins isolated from California, and a second group found in Durban, South

Africa. In both of these locations, inteins exhibited nearly identical nucleotide sequences, but were found

in comparatively divergent exteins. Furthermore, these extein sequences were separated in the extein

phylogeny by non-intein containing sequences.

Figure 6. **A.** Maximum likelihood phylogeny of intein-free and intein-containing nucleotidyltransferase extein-encoding nucleotide sequences, using TIM+F+R5 as the substitution model. Double slashes indicate branches shortened for readability. Circles to the right of taxa names indicate intein presence, with orange and purple denoting two different intein clusters. Ultrafast bootstrap support is listed on internal nodes. There is a split intein in this dataset, from phage Zeenon, indicated by asterisks on the extein phylogeny since its extein is split into two separate open reading frames by the split intein. Scale bar measures substitutions per site. **B**. Maximum likelihood phylogeny of the intein-encoding sequences from the orange cluster, using HKY+F+I as the substitution model. Bootstrap support is shown as it was in **A**. Geographic maps show the distribution of inteins from the orange cluster within: California (**C**), Natal South Africa (**D**), the city of Durban (**E**), and the whole world (**F**). Note that the phages from South Africa (**E**) have identical or nearly identical intein sequences (**B**) but that their exteins do not group together and are separated by exteins harboring no intein, or inteins from a different cluster (**A**). Similarly, the phages isolated in California have nearly identical inteins (**B**), but their exteins are divergent and do not group together (**A**).

We repeated this analysis with a second group of inteins from Ohio and Pennsylvania, found in

minor capsid proteins (Figure 7, red cluster). These 7 inteins possess an anti-sense LAGLIDADG HEN.

These inteins, despite being identical to each-other (for 5 of the seven), are once again found in divergent

extein sequences which were separated from each-other on the phylogenetic tree with non-intein

containing taxa, and taxa with completely different inteins.

Figure 7. **A.** Maximum likelihood phylogeny of the nucleotide sequences of intein-containing and intein-

free minor capsid proteins using TN+F+R3 as the substitution model. Colored circles to the right of taxa

names indicate intein presence, and each color corresponds to a different intein cluster. Ultrafast

bootstrap support is listed on internal nodes. Scale bar measures substitutions per site. **B.** Maximum

likelihood phylogeny of the intein nucleotide sequences from the red cluster only, using F81+F as the

substitution model. Bootstrap support is shown as it was previously. The geographic maps indicate the

distribution of inteins from the red cluster in the Ohio-Pennsylvania area (**C**), Ohio State University and

surrounding area (**D**), and the University of Pittsburgh and its surrounding area (**E**). Note that the three

phages depicted in panel **E** have identical intein sequences, but that the extein from phage Nordenberg

diverges from the exteins in Lennon and Sitar and that the three exteins do not form a clan.

Finally, we investigated the inteins found within exteins annotated as putative integrin-like

proteins (Figure 8). Once again, these inteins were found in divergent exteins separated on the

phylogenetic tree by non-intein containing taxa. Many of these inteins were found specifically within the

grounds of Howard University in Washington DC; however, despite the small geographic distance

between the phages found there, the exteins showed significant divergence. Furthermore, inteins from

each of the three clusters were found on this campus. The vast majority of the inteins from the bottom

clan of the intein phylogeny (Figure 8B, gold) were present predominantly on the eastern shoreline of the

USA. The clan at the top (Figure 8B, blue) had a much wider distribution — mostly confined to the

Mississippi River Valley, and the Midwest. The last of the three clans (Figure 8B, red) was found across

the USA.

Figure 8. **A.** Maximum likelihood phylogeny based on the extein-encoding nucleotide sequences of intein-free and intein-containing putative integrin-like proteins using TIM2+F+I+G4 as the substitution model. Colored circles to the right of taxa names indicate intein presence, and the color indicates which cluster (**B**) said intein belongs to. Ultrafast bootstrap support is listed on internal nodes. An asterisk indicates that the phage was isolated within Washington DC. Scale bar measures substitutions per site. **B.** Maximum likelihood phylogeny of all intein-encoding nucleotide sequences from **A**, using TN+F as the substitution model. Colored bars on the three clans correspond to the intein colors in **A**. Bootstrap support is shown as it was previously. **C.** Geographic maps indicating the distribution of these inteins. First within the entire United States of America, and then gradually zooming in on Washington DC, and eventually Howard University. Isolation locations where more than one intein containing phage were collected are

360   indicated with a plus sign and the corresponding number of additional inteins. The color of the circles

361   corresponds to the clans in **B**.

Discussion

Overarching Trends

As in previous studies, the inteins we discovered within actinobacteriophages are found

predominantly within DNA-binding extein sequences (Kelley et al. 2016). While the majority are within

terminase large subunits, the inteins we discovered show a remarkable range of host proteins. In addition

to ubiquitous phage genes (such as the terminase large subunit), we find inteins within phage genes with

more limited distributions such as thymidylate synthase, or ribonucleotide reductase subunits. Even

genes that lack nucleotide-associated enzymatic activity, the minor capsid proteins, were found within the

set of invaded phage proteins. Interestingly, the inteins invading minor capsid proteins were exclusively

inteins with non-canonical HENs.

Many inteins in our dataset had sporadic distributions across their extein phylogenies, and

phylogenies of the intein sequences frequently conflicted with the phylogeny of their extein. We believe

that many of these inteins are actively moving between different hosts. However, exceptions do exist in

cases of mini inteins (an example of which can be found in Supplemental Figure 8). In addition, inteins

that persist in clans of extein sequences, at first sight might be considered to have decaying HENs, and

like mini-inteins be vertically inherited (see the purple intein group in Figure 7). However, while the exteins

show some sequence divergence, the fact that the inteins are identical likely illustrates a recent invasion

event (see below for further discussion).

The many intein-free homologs accompanying intein-containing genes in our dataset indicate that

intein homing into new empty target site must be balanced by the loss of inteins in other homologs. Given

the seemingly rapid dissemination of inteins into local phage populations, intein loss might also be

assumed to occur frequently. This could occur via gradual decay and recombination of the intein, or be

the consequence of a fitness cost imparted by the intein on the phage. Inteins are known to cause a drop

in fitness; in the archaeon *Haloferax volcanii,* a DNA polymerase intein caused a 7% drop in fitness for

the host (Naor et al. 2016). Thus, the alternative loss scenario would act at the population level — the

intein imparts enough of a fitness cost such that the intein-containing phage is out-competed by intein-

free phages, and the intein containing phages would be slowly selected out of the population. This

process would be enhanced if the non-invaded phages evolve at least a partial resistance to intein

propagation (perhaps via mutations to the HEN target site). This potential impact on host fitness is a

mechanism worth exploring in future work.

The Nature of Non-Canonical Inteins

In this study we described several inteins with previously unseen, or extremely rare architectures

in relation to their HEN domains. To our knowledge, only the anti-sense LAGLIDADG HEN has been

reported on in prior literature within the context of a fully functional intein (Gorbalenya 1998; Gogarten et

al. 2002). These anti-sense LAGLIDADG HEN containing inteins were discovered in non-phage genomes

within the *dnaX* gene of a *Synechocystis* species, and the SpoVR-like protein from *Chloroflexus*

aurantiacus J-10-fl (sequences deposited in the InBase database (Perler 2002)). Hence, while not entirely

novel, we show that this intein architecture is present within at least one other extein sequence, and

importantly, that the intein and anti-sense HEN appear to work together to facilitate ongoing intein

invasion events.

VSR-like HEN-containing inteins have been reported on previously, within split-inteins (Dassa et

al. 2009). Our findings show that anti-sense VSR-like HEN-containing full inteins also exist and are not

merely a point of speculation. If the theory of split-intein evolution outlined by Dassa et al. 2009 is true,

then perhaps these newly discovered inteins represent one of the potential off-tracks within their model. It

should also be noted here, that Dassa et al. 2009 report that the VSR-like HENs they discovered were

related to freestanding HENs within their database (a fact whose importance will be discussed below).

Finally, the endonuclease VII-like HEN containing intein appears to be completely novel, with no

known reports. Like the VSR-like HENs, this endonuclease has been reported prior in a free-standing

context (Pope et al. 2013).

Given the ubiquity of freestanding HEN genes within actinobacteriophages, the frequency with

which these genes seem to proliferate (Edgell et al. 2010; Pope et al. 2013; Zhang et al. 2017; Hatfull

2020; Barth et al. 2023), and the presence of free-standing HENs related to those found within our intein

sequences, we believe that the three intein architectures described above originate via invasion of an

intein sequence by a free-standing HEN. Given the seeming rarity of these elements, we further propose

(in line with the model laid out by Dassa et al. 2009 (Dassa et al. 2009)), that successful integration of

these HENs into the region of an intein between splicing associated blocks is extremely rare. The

breaking of, and subsequent integration into the intein CDS by a free-standing HEN in this fashion may

be relatively common, but most of the time this integration is likely to result in split-inteins, or introduce

nonsense mutations which prevent post-translational rescue of extein function via intein splicing.

Assuming the free-standing HEN's integration into the intein CDS is successful, the path forward

is unclear. It is possible the HEN becomes a pseudo-gene, but there have been prior examples of phage

genes with nested HEN open reading frames (ORFs) that still translate both proteins (Gibb and Edgell

2007; Wright et al. 2021). In our cases, the secondary ORFs typically lie outside the bounds of the intein

splicing domain, and given the splicing domain's high conservation, it is probable that the spicing domain

still functions properly. Additionally, many of these non-canonical inteins were found in the crucial

terminase large subunit gene, and it is therefore unlikely that the intein is not splicing itself out — these

phages would otherwise be unable to properly package their DNA into their capsid (a certain end for that

phage lineage). Our phylogenetic reconstructions of the anti-sense LAGLIDADG-containing inteins

provided evidence for continued invasion. These inteins are present in multiple locations within divergent

host exteins, and importantly are separated with meaningful support by relatives that lack the intein. Such

a pattern is not reasonably explained via vertical inheritance and loss, and it is therefore likely that the

splicing domain and anti-sense HEN domain of that particular intein still function as a single unit. This

may also be true of two groups of inteins with VSR-like HENs (Supplemental Table 1, clusters 60 and 91)

which likewise show a pattern of inheritance more in-line with horizontal gene transfer. However, the

other cases of VSR-like HEN-containing inteins exhibit patterns suggestive of vertical inheritance (or were

singletons and thus no evolutionary inference could be drawn). The endonuclease VII-like HEN-

containing inteins presented a more ambiguous distribution. While most are clustered together, there are

a number of closely related extein alleles without an intein. This, plus the sequence similarity of these

inteins (for example, the inteins of cluster 47 — which includes phages Angelicase, Affeca, Keitabear,

and Sedona — are all identical with the exception of Angelicase), suggests that these inteins may still be

transferring between phages horizontally (Supplemental Figure 9, Supplemental Figure 4).

If these inteins can still splice, but can no longer successfully jump, then selection will likely lead

to these secondary ORF's very quickly accumulating stop codons and ceasing to function. This would be

especially likely in the case of anti-sense ORFs since they would not benefit from the purifying selection

acting upon the host extein to remove nonsense mutations. Mini-inteins and split-inteins would be the

most likely observable outcome of this process. While we do find nine inteins which appear to have

decaying HEN domains, and a further nine mini-inteins, a survey strategy fine-tuned towards finding split

and mini inteins would be needed to look for these potential relics of decay. Potentially, the secondary

ORFs encoded on the same strand could transit into the extein's reading frame via one or two base pair

indels, resulting in a fully contiguous intein with a novel HEN.

Inteins created by the mechanism described above thereby provide meaningful support for a

"protein splicing first" theory of intein evolution (Derbyshire and Belfort 1998; Gogarten et al. 2002; Dassa

et al. 2009; Green et al. 2018). The number of intein-associated HEN's (including HNH, VSR-like, GIY-

YIG, LAGLIDADG, etc. (Dalgaard, Klar, et al. 1997; Gorbalenya 1998; Gimble 2000; Dassa et al. 2009))

suggests that there have been multiple separate invasions of intein coding sequences by free-standing

HENs (Derbyshire and Belfort 1998). Perhaps most, if not all, of these HENs came into intein sequences

via a free-standing HEN, proliferated through the intein's self-splicing machinery, and were shifted by an

indel or an inversion into a contiguous orientation.

Frequent Horizontal Transfer of Inteins

Extein phylogenies with intein presence mapped onto them, such as in Figures 6A, 7A, and 8A,

can reveal likely cases of vertical inheritance, recent dissemination, and horizontal transfer of inteins.

Using an extein phylogeny with intein presence mapped will differentiate between horizontal transfer and

vertical inheritance or recent dissemination; however, intein phylogenies and assessment of intein and

extein conservation are required to differentiate between vertical inheritance with frequent intein loss

events and recent dissemination.

A set of intein-containing exteins which all group together initially indicates vertical inheritance of

that intein, such as the group in Figure 7A containing phage ViaConlectus. While the phylogeny is

unrooted, if one follows the presented arrangement one would infer that ViaConlectus was invaded by the

purple cluster intein, and this intein was then vertically inherited by phages APunk, Abblin, Scioto,

Sampson, and Natkenzie. To assess whether the event is a case of vertical inheritance or recent rapid

dissemination, the degree of similarity between inteins in these cases must be further analyzed, as well

as geographic proximity between hosts when such information is available. In such cases, extremely

similar inteins in geographically-close but less similar exteins may indicate a recent intein dissemination

event.

In contrast to the phylogenetic patterns suggesting vertical inheritance or recent local

dissemination are cases where we find inteins within extein sequences that lack a direct common

ancestor and are separated on the extein phylogeny by many nodes leading to other non intein-

containing extein sequences. In such cases, a vertical inheritance pattern would require many loss events

to be an accurate model given these phylogenies. To supplement findings from extein phylogenies with

intein presence mapping, the phylogeny of the intein of interest can be reconstructed and its topology

compared to that of the extein phylogeny to identify incongruencies, which often provide further evidence

of a non-vertical inheritance pattern. For example, in the extein phylogeny with intein presence mapping

in Figure 7A, we see a well-supported (99% ultrafast bootstrap support) split between a group of exteins

from phages including phage SchottB, and a group of exteins from phages including phages Kamashten,

Thing3, Barsten, Lennon, Sitar, and Nordenberg. Despite the evolutionary distance between exteins,

these phages all contain near identical inteins at the same position in their extein (inteins from the red

cluster in Figure 7A). To supplement this observation, Figure 7B depicts the intein phylogeny for this red

cluster of inteins which shows the SchottB intein grouping closely with the Barsten intein, despite the

large well-supported evolutionary distance between their exteins. The most parsimonious model

explaining this observation is a horizontal transfer event between SchottB and a member of the

Kamashten group, as opposed to vertical inheritance with many independent loss events. Applying this

phylogenetic analysis to each of our extein-intein phylogeny pairings thus reveals a multitude of cases for

the horizontal transfer of inteins between phages.

For highly similar inteins which possess a wide geographic distribution, the next question

inevitably becomes how did they get there? While humans are almost certainly acting as a secondary or

tertiary vector for the intein, we propose that these inteins are likely dispersing in part via the phages we

find them in and in part, via the phage's host bacteria. Previous literature, and our own anecdotal

observations, find that many phage-associated inteins are found inside of bacterial genomes, most likely

within prophages (25). Thus, inteins experience increased genetic mobility, taking advantage of both the

viral and bacterial vectors. As a prophage, the intein can propagate not only when the bacterial cell

replicates, but also through the numerous avenues of horizontal gene transfer that bacterial cells engage

in (Soucy et al. 2015), and by transfer into closely related phages upon infection.

Geographic Spread of Inteins

Our dataset contains several groups of inteins which are extremely similar on the nucleotide level

(and in the cases described, often identical), these inteins must have diverged from one another relatively

recently. This presents two options: rapid host proliferation with direct vertical inheritance, or rapid intein

proliferation via horizontal gene transfer. For some of the inteins in our dataset, the former may be

considered a possibility (see the clan of intein containing exteins colored purple in Figure 7); however,

where the intein containing exteins form a well-supported clan with limited geographic distribution, the

observation that the inteins are identical, whereas the exteins are divergent, suggests that the inteins

have recently invaded the divergent exteins instead of being vertically inherited. This argument is

strengthened by the observation that inteins found in divergent exteins experience less selection for

sequence conservation than the exteins that harbor them (see Supplemental Materials: Supplemental

Results and Discussion – Sequence Conservation in Inteins and Exteins). The finding of nearly identical

inteins found in divergent exteins in close geographical proximity suggests that intein invasion represents

an example of a highly effective gene drive acting on the local level following the introduction of a new

founder intein allele. The forces that limit this drive (after all, not all homologous exteins within a given

location contain the intein) need further study (see the above discussion of fitness cost to the host that

can be associated with inteins). Phages, because of their rapid propagation and large population sizes

provide an excellent opportunity to study intein population dynamics in more detail.

Nucleotide binding sites as an avenue for extein-switching

The instance of Cas4 exonuclease inteins evolving from terminase large subunit inteins on the

class 3 intein phylogeny involving phages LittleE and ScoobyDooByDoo (SDD) (Figure 4) provides a

compelling argument for the use of highly similar nucleotide binding sites as a means of intein movement

between otherwise dissimilar genes. The stretch of identical nucleotides between the two intein insertion

sites (10 bases on the 5' side, 2 bases on the 3' side) in conjunction with the other shared positions within

the +/-30bp surrounding the site (Supplemental Figure 7A) may be sufficient for recognition and invasion

by the same intein. The proposed range for HEN recognition sites is ~12-40bp, with recognition sites in

smaller genomes such as phage genomes being on the lower end of the range (Jurica and Stoddard

1999). Additionally, many types of phage recombinases can tolerate some dissimilarity between regions

used for homologous recombination (Lopes et al. 2010) suggesting this intein could possibly invade either

site if given the chance.

To further complete the picture and investigate why the SDD terminase large subunit and LittleE

Cas4/RecB-like exonuclease are intein-free, we analyzed the regions corresponding to the intein-insertion

sites on the intein-free proteins (Supplemental Figure 7B) and find SDD's Cas4 exonuclease gene to be a

more hospitable landing pad for the LittleE terminase large subunit intein by metrics of insertion site

recognition and residues at the insertion site required for intein self-splicing (the SDD terminase large

subunit encodes an alanine at the position which would need to be a serine, threonine, or cysteine for

successful intein splicing to occur) (Supplemental Figure 7B) (Mills et al. 2014; Turgeman-Grott et al.

2023). Additionally, LittleE is a temperate phage of the same strain of bacteria that SDD is a lytic phage of

(*Mycobacterium smegmatis* mc2155) (Russell and Hatfull 2017).

The presented evidence taken together suggest a model in which the recipient genome was

exposed to the donor's terminase large subunit intein during co-infection or infection of a host containing

the donor's prophage. Then, either homing of the donor terminase large subunit intein into the recipient's

terminase large subunit gene was thwarted by the lack of similarity between these terminase large

subunits, or homing was successful but nonviable due to lack of a splicing-compliant residue on the C-

terminal side of the insertion site. By happenstance, a region of similarity to the donor terminase large

subunit present in the recipient Cas4 exonuclease gene conferred by a shared nucleotide binding motif

provided a viable alternate insertion site for the intein (Figure 9). Being in a brand-new context also lets

this copy of the intein begin a fresh evolutionary trajectory, as the majority of homologous genes in the

population will initially lack this intein. This may explain the presence of SDD's Cas4 exonuclease intein in

over 130 other phage Cas4 exonucleases (Figure 4). As PhagesDB and other phage genome sequence

resources continue to grow, increased sampling will tell a clearer story of the events that unfolded in the

evolution of these inteins.

Figure 9: Three different scenarios for attempted intein invasion by the LittleE terminase large subunit

(TLS) intein, explaining simultaneous presence of intein in LittleE TLS and ScoobyDoobyDoo (SDD) Cas4

exonuclease genes, but lack of intein in LittleE Cas4/RecB-like endonuclease and SDD TLS genes. TLS

exteins are depicted as thin blue boxes, Cas4 exonuclease exteins are depicted as thin red boxes, and

the intein sequence within invaded genes is depicted as a thin purple box. Nucleotide binding site A

(depicted as a thick purple box) is the insertion site recognized by the LittleE TLS intein, and is present in

both the LittleE TLS gene and the ScoobyDoobyDoo (SDD) Cas4 exonuclease gene. Nucleotide binding

site B (depicted as a thick light red box) is not recognized by the LittleE TLS intein, and is present in the

LittleE Cas4/RecB-like exonuclease gene in place of nucleotide binding site A. Thus, the LittleE TLS

intein cannot engage in homing with the LittleE Cas4/RecB-like exonuclease gene and invasion cannot

occur. Nucleotide binding site C (depicted as a thick dark blue box) is not recognized by the LittleE TLS

intein, and is present in the SDD TLS gene in place of nucleotide binding site A. Thus, the LittleE TLS

intein cannot engage in homing with the SDD's TLS gene and invasion cannot occur. Since the SDD

Cas4 exonuclease gene contains nucleotide binding site A, the LittleE TLS intein can engage in homing,

allowing the invasion of the LittleE TLS intein into the SDD Cas4 exonuclease gene.

Conclusions

The work outlined above shines a light on the previously unknown breadth of inteins within

Actinobacteriophage genes. Throughout this research we repeatedly find disjunct distributions of intein

containing exteins in extein phylogenies, and propose that the conflict between extein and intein

phylogenies suggests that inteins within Actinobacteriophages frequently home into target sites within

intein-free genes. We find evidence for multiple cases of rapid geographically local episodes of intein

invasion that support the prior assertion. In addition, we describe several previously unseen or rare intein

architectures, and provide a hypothesis to explain their potential occurrence and apparent continued

dissemination. We also describe a model for how inteins may home into new target sites, within

previously un-invaded gene families, based on the intein phylogenies and the similarities and

dissimilarities between target sites.

Finally, we wish to stress that these non-canonical inteins may be significantly more common

than even what we have outlined here. Given their overlapping ORFs, these inteins are likely missed in

automated annotation pipelines, especially within phages — current SEA-PHAGES training argues for

"...typically only one frame in one strand is used for a protein-coding gene…" (SEA PHAGES). Even to

the trained eye, these inteins may currently be glossed over as 'decaying' canonical inteins or

misclassified as mini-inteins. Significant work will be needed to confirm that inteins listed as decaying are

correctly classified, and to discover the true extent of these inteins within phages and their bacterial hosts.

Materials and Methods

Code Availability

All code used in this project is available at https://github.com/sophiagosselin/intein_distribution along with usage information. The two exceptions to this are ICE-BLAST and Invader-Sim which are kept in separate repositories detailed below.

Iterative Cluster Expansion BLAST: ICE-BLAST

To increase efficiency in mass intein sequence retrieval, we developed "Iterative Cluster Expansion BLAST" (ICE-BLAST). ICE-BLAST is a Perl (v 5.36.0) based program using PSI-BLAST (Altschul et al. 1997) as the principal sequence search tool and USEARCH clustering (Edgar 2010) to identify new queries from previous iterations. Through this process, ICE-BLAST retrieves highly divergent homologous protein sequence matches. In short, ICE-BLAST iteratively searches amino acid databases with PSI-BLAST, clusters filtered outputs using USEARCH, then uses the representative sequence of each match cluster (centroids) as the next query sequences (Figure 10A).

ICE-BLAST is available at https://github.com/sophiagosselin/ICE_BLAST along with detailed usage, dependencies, and other information. Algorithm details and parameters used for this study are available in Supplemental Materials.

Simulations of Intein Evolution: Invader-Sim

Since ICE-BLAST involves multiple individual searches, one must take proper precautions against false-positives. False-positives can introduce errors that will propagate through all downstream iterations. Hence, it was important to test ICE-BLAST against known data to assess overall sensitivity and specificity. We developed a Perl (v 5.36.0) based sequence simulation program, Invader-Sim, to generate sequences simulating intein invasion into extein sequences. The program uses a combination of PAML-EVOLVER (v4.9j) (Yang 2007) and a Markov chain to simulate datasets of intein-containing extein sequences (Figure 10B). ICE-BLAST performed as well as PSI-BLAST on the simulated datasets. Results and additional discussion can be found in Supplementary Materials.

Invader-Sim is available at https://github.com/sophiagosselin/invader_sim. Information on the

method and parameters used for this study are available in the Supplemental Materials.

Figure 10. Flow charts describing the algorithms of ICE-BLAST and Invader-Sim. **A.** ICE-BLAST begins

by taking amino acid query sequences and iteratively searching against a training amino acid sequence

databank using PSI-BLAST. The matches resulting from this iterative search are clustered by sequence

similarity, where a representative amino acid sequence is selected from each cluster (referred to as the

centroid sequence for that cluster). The centroids of each cluster are then used as input for a new round

of PSI-BLAST searches, with the process repeated until either the iteration limit is reached, there are no

new centroids, or there are new centroids but no new matches over all. **B.** Invader-Sim begins by using

PAML-EVOLVER to simulate an extein phylogeny and an intein phylogeny. From the phylogeny of

potential exteins, a random individual is chosen to be invaded by an intein from the intein phylogeny. The

closest relative of the randomly selected intein is given a new extein to invade, with the extein selection

proposed by a Markov model. The process is repeated until all exteins are invaded by an intein.

Simulated sequences are then generated using PAML-EVOLVER.

Dataset Curation: Applying ICE-BLAST to PhagesDB

Using PhagesDB data (Russell and Hatfull 2017), two databases were constructed: one from

amino acid sequence files, and one from whole genomes, with associated metadata from the PhagesDB

API. The genomic and amino acid databases were downloaded and formatted on September 28 2023

and November 2 2023 respectively. The databases were not updated over the course of this work, due to

reverse incompatibility of PhagesDB updates.

The initial ICE-BLAST query set for this study consisted of 78 intein sequences drawn from three

sources: InBase (final update 2010) (Perler 2002), known Actinobacteriophage inteins (Kelley et al.

2016), and inteins found by the Gogarten Lab in the PhagesDB database between 2020 and 2024

(Gosselin et al. 2023) (Supplemental Table 3). Initial PSSMs were trained on Uniref50 (Suzek et al. 2015)

downloaded November 7 2021. For the ICE-BLAST search, a percent identity of 60%, an e-value of 1e-

20, and a domain specific cutoff bound threshold of 25% were used as inputs for filtering.

Initial results from ICE-BLAST contained 1994 sequences which were then clustered by

USEARCH (v11.0.667) (Edgar 2010) at 70% identity using the cluster_fast algorithm. Associated intein-

invaded extein sequences were then extracted from the BLAST database using blastdbcmd. Putative

intein sequences were aligned to the invaded extein sequences using MAFFT (v7.490) (Katoh and

Standley 2013). When intein identity was uncertain, BLAST against NCBI's non-redundant database

(nrdb) (Altschul et al. 1990), and HHPred against PDB, Pfam, and NCBI CDD (Zimmermann et al. 2018),

were used to determine whether the sequence was actually an intein, and where its boundaries laid.

False positives were removed from the dataset at this point. This screening reduced the dataset to 784

confirmed intein-containing sequences (Supplemental Table 1). The large was attributed entirely to a

false positive match to a DNA Primase sequence; causing over 1000 matches to DNA primase extein

domains which were falsely incorporated. Extein sequences were re-annotated using BLAST and HHPred

as described above. Inteins were classified as class 1, 2, or 3, by comparing to self-splicing domain

consensus sequences (Tori et al. 2010). In-house Perl scripts were used to extract associated coding

sequences from the mentioned PhagesDB genome database. Intein and extein sequence files are

available in Supplemental Data.

HENs were classified using the method above, with two exceptions. First, we used the PHROG

(Terzian et al. 2021) database within HHPred (Zimmermann et al. 2018) (especially for more divergent

sequences) to identify potentially related HENs as specified in the PFAM section of its entries. Second,

we used InterProScan (as of August 2024) (Jones et al. 2014) when BLAST and HHPred were

inconclusive.

Geographic Maps

First, intein nucleotide sequences were clustered using USEARCH (97.5% sequence identity)

resulting in 91 clusters. Clusters with less than four members were excluded, as a meaningful

phylogenetic tree cannot be generated. If exteins within a cluster were more self-similar than their intein

sequences were, the cluster was removed. Remaining clusters were then used to build intein nucleotide

phylogenies, and their extein sequences (with intein-less homologs from PhagesDB) were used to

construct an extein phylogeny. Only clusters whose intein and extein phylogenies were in-congruent were

selected for further study.

Extein phylogenies were marked for intein presence, and the inteins present were then mapped

onto their geographic isolation location. Phage isolation locations were extracted from PhagesDB's API.

Maps of intein isolation locations were created using R (v4.4.1) and the following libraries: sf (v1.0-16)

(Pebesma 2018), ggplot2 (v3.5.1) (Wickham 2016), maps (v3.4.2) (Deckmyn 2023), mapdata (v2.3.1)

(Brownrigg 2022), mapproj (v1.2.11) (Minka and Bivand 2023), rnaturalearthdata (1.0.0) (South et al.

2024a), and rnaturalearthhires (v1.0.0.9000) (South et al. 2024b).

Phylogenetic Trees and AU Tests

Phylogenies in this study were built from MAFFT sequence alignments. Unless specified, MAFFT

was allowed to select the best alignment algorithm for a given set of sequences (strategy: 1. --auto). IQ-

TREE (v2.0.7) using ModelFinder (-m MFP) and 1000 ultrafast bootstraps (-bb 1000) (Minh et al. 2020)

was used for phylogenetic reconstruction. Models selected are included in phylogeny figure legends.

Trees were visualized using Dendroscope (v3.8.10) (Huson et al. 2007), Figtree (v1.4.4) (Rambaut 2012),

and Inkscape (v1.3.2) (Developers). Clans with extremely similar or identical sequences were collapsed in

some phylogenies for readability. Un-collapsed versions are included in the supplemental material (see

Figure 4 and Supplemental Figure 10 for examples of collapsed and un-collapsed respectively).

AU tests (Shimodaira 2002) were performed via the method built into IQ-TREE2 (Minh et al.

2020).

Predicted Protein Structures

Protein structures were predicted for non-canonical inteins and their associated secondary ORFs

using AlphaFold (Jumper et al. 2021) through UCSF ChimeraX (Pettersen et al. 2021; Meng et al. 2023).

Structures were colored according to the predicted local distance difference test (pLDDT) value per-

residue. Each predicted structure was used as query in a DALI search of the PDB database (Holm et al.

2023).

AlphaFold (Jumper et al. 2021) was also used to generate predicted structures in the extein

switching study. UCSF ChimeraX (Pettersen et al. 2021; Meng et al. 2023) was used to compare the

predicted structure of the LittleE terminase large subunit to the solved structure of a phage terminase

large subunit bound to ATP (Zhao et al. 2013).

Using Predicted Structures to Investigate and Classify Non-Canonical HENs

For the inteins with non-canonical HENs, predicted structures suggest that the portion of the

intein which should be an in-frame HEN does not make any recognizable structures (Supplemental Figure

1), and cannot be matched to any known structure using the DALI server PDB search tool. However,

predicted structures of the anti-sense LAGLIDADG HEN, anti-sense VSR-like HEN, and out-of-frame

endonuclease VII-like HEN, all matched to their associated predicted domains (Supplemental Figure 2). It

is important to note that these matches were only to the core catalytic domain of these HENs; significant

portions of these structures could not be matched to known proteins. For the anti-sense LAGLIDADG

HEN, the LAGLIDADG core sits between residues 40 and 185, matching to PDB structure 2O7M (Prieto

et al. 2007) (CreI — a known LAGLIDADG HEN) with a Z-score of 14.8. The VSR-like HEN predicted

structure matched to PDB structure 1CW0 (Tsutakawa et al. 1999) (VSR) with a Z-score of 10.6 across

the residues 144 to 267. Finally, the endonuclease VII-like HEN matches to PDB structure 1E7D

(Raaijmakers et al. 2001) (a recombination endonuclease VII) with a Z-score of 6.5 across residue 1 to

residue 67.

Intein Insertion Site Consensus Sequence Plots

WebLogo (Crooks et al. 2004) was used to generate consensus sequence plots in this study.

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