# Protein Structure Description with ρ, θ and φ: A Case Study with Caenopore-5

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## Abstract

Since its establishment in 1971, the Protein Data Bank (PDB) has been using Cartesian coordinate system (CCS) as the standard framework for protein structure description with $x$, $y$, $z$. Despite the interconvertibility of CCS and spherical coordinate systems (SCS, ρ, θ and φ), CCS remains to date the default and the only framework for protein structure description in PDB. Recent advances in protein structure prediction (e.g., AlphaFold) revolutionized the field by integrating deep learning algorithms with experimental structural data, achieving unprecedented accuracy of protein structure prediction and relying on Cartesian representation of protein structures to extract geometric features, such as inter-residue distances and dihedral angles, which are essential for building and visualizing protein structures with $x$, $y$, $z$. To this end, questions remain about the potential for further improvement in the performance of protein structure prediction, i.e., what is going to drive the next stage of its continued development with improved accuracy and efficiency. Therefore, this article introduces an alternative coordinate system for protein structure description and feature extraction. Using Caenopore-5 as an example, this article redefines protein backbone structures using atomic bonding networks (ABN) within the SCS framework (ABN-SCS), leading to the extraction of a set of spherical parameters (ρ, θ and φ) from the NMR ensemble of Caenopore-5, encompassing 477 covalent bonds and 80 peptide bonds within its backbone for each structural model in its NMR ensemble. Overall, this work offers an alternative spherical framework (ABN-SCS), i.e., a new language (ρ, θ, φ), for protein structure description and ABN-SCS-based extraction of spherical geometric features for protein structure prediction, aiming to contribute a little bit to its performance and application in drug discovery and design.

## Keywords

Protein Data Bank; Spherical Coordinate System; Atomic Bonding Network-based Spherical Coordinate System; Protein structure prediction; Feature extraction.

# Introduction

## $[x,\ y,\ z]$ and $[\rho,\ \theta,\ \phi]$ for protein structure description

In 1971, Protein Data Bank (PDB) was established as the single global repository for experimentally determined biomolecular 3D structures [1–4]. Since then, PDB has been acting as a catalyst [5] for structural biology, i.e., understanding [6–8], determination [9–12], description [13–18] and prediction [19–24] of the structure of protein, the essential building block of life. To this end, protein structure determination and description are like the two sides of a coin, both essential for understanding its function, and uncovering disease mechanisms caused by structural abnormalities, which drive advancements in protein engineering and structural modeling for drug discovery and design [25–27].

Since 1971 [1, 2], Cartesian coordinate system (CCS) has been the default framework (Figure 1) to describe protein 3D structure, i.e., the specification of spatial atomic positions with Cartesian coordinates $[x,\ y,\ z]$. In biochemistry, the primary structure of protein is described with an alphabet of twenty letters, each letter representing one standard amino acid, and the secondary structure of protein a mini alphabet of three letters for α-helix, β-strand and random coil, and the tertiary and the quaternary structures of protein a set of $[x,\ y,\ z]$. That is, for over half a century [28], $[x,\ y,\ z]$ has been used as the standard language (i.e, the default Cartesian coordinate system) for protein structure description, reflecting an intuitive simplification and conceptualization of protein 3D structure as a set of discretely distributed points in 3D space, where each point corresponds to an atom in the protein, and its position is defined by the $[x,\ y,\ z]$. In this intuitive simplification and conceptualization with $[x,\ y,\ z]$, nonetheless, one key aspect is not taken into account, i.e., the fact that structurally, protein is a set of chemically bonded points in 3D space, consisting of an intrinsic inter-atomic bonding network, instead of a set of discretely distributed points.

**Fig 1.** Over half-a-century burial of $[\rho,\ \theta,\ \phi]$ in the Protein Data Bank [2, 3, 5].

In geometry, CCS ($[x,\ y,\ z]$) and spherical coordinate system (SCS, $[\rho,\ \theta,\ \phi]$) are interconvertible, both applicable in the specification of atomic positions [17, 18, 28]. In 2011, for instance, it was proposed for the first time that protein structure be represented in spherical coordinates [14, 15], where a global spherical coordinate system (G-SCS, Figure 1) was proposed and used for the separation of the protein outer layer from its inner core, and for the identification of protrusions and invaginations on the protein surface [14, 15]. By definition, G-SCS (Figure 1) requires three parameters ($R$, θ and φ), and takes the geometric centroid (an imaginary point) of the protein structure as origin ($[0,\ 0,\ 0]$), where $R$ is used to signify the distance between the imaginary centroid and any atom of the protein [14, 15].

## A Cartesian Protein Data Bank for protein structure prediction

As of January 8, 2025, more than 229,000 biomolecular structures have been determined and deposited in PDB [29, 30]. These structures were predominantly resolved using experimental techniques such as X-ray crystallography [9, 12], NMR spectroscopy [10, 12], cryo-electron microscopy [11, 12, 31], etc [12, 32]. Nonetheless, 229,000 represents only a small fraction of the billions of known protein sequences [33]. To address this disparity, computational tools become indispensable for large-scale structural bioinformatics, driving advances in protein structure prediction in the past few years [23, 34–39]. For instance, AlphaFold (Figure 1) [34] and RoseTTAFold (Figure 1) [35] transformed protein structure prediction through the increasing adoption of deep learning methodologies [33, 40–42], which use neural networks to replace traditional energy models and sampling procedures, resulting in significant improvements in prediction accuracy [23, 24, 43–46].

As described above, the Protein Data Bank that we have now is essentially a Cartesian Protein Data Bank due to CCS's long-standing role as the default system for specifying atomic positions with $[x, y, z]$ [1, 2]. As the primary repository of protein structure data, PDB [1, 2] serves as the foundation for the training and the validation of most protein structure prediction tools [23, 34–39]. Take AlphaFold (Figure 1) [34] for instance, which takes Cartesian-format PDB files as input for the extraction of geometric features, such as inter-residue distances and dihedral angles. Subsequently, along with the protein structure data in PDB, these structural features are plugged into deep learning algorithm(s) for the prediction of discrete probability distributions of inter-residue distances, resulting in an $L \times L$ distance map for the protein of interest [33, 40, 47]). This $L \times L$ distance map serves as the foundation for constructing Cartesian format ($[x, y, z]$) structural models, subsequent visualization with PyMol [48] or VMD [49], or further analysis and use in computational drug discovery and design [25–27].

## Aim

Traditional protein structural modeling in the 1980s is based on known structural templates [50–52], followed by advancements in molecular dynamics and energy minimization techniques [53]. Recent years saw a paradigm shift with the advent of AlphaFold, which leverages deep learning to achieve unprecedented accuracy in predicting protein structures [20–22, 34, 54, 55]. However, questions remain about the potential for further improvement in the performance (e.g., accuracy, efficiency) of protein structure prediction, i.e., what is going to drive the next stage of the development in this field. This article, therefore, reports an atomic bonding network-based spherical coordinate system (ABN-SCS) [16–18] for protein structure description with $[\rho, \theta, \phi]$, instead of $[x, y, z]$. Aiming to provide an alternative framework rooted in the intrinsic geometry of atomic bonding network of protein structure, this article builds on this ABN-SCS framework, using Caenopore-5 [56–58] as an example to demonstrate the use of ABN-SCS in redefining protein structure ($[\rho, \theta, \phi]$) and extracting spherical structural features, in addition to those Cartesian structural features used by current protein structure prediction tools [59–65].

## Materials and Methods

### Two geometrically inter-convertabile coordinate systems

This section is described in the first and the second sections of supplemtary file **S2.pdf**.

## An ABN-SCS description of the structures of twenty stand-alone natural amino acids

This section is the same as the third section of supplemtary file **S2.pdf**, and is described in supplemtary file **S1.pdf**.

## An ABN-SCS description of protein backbone structure: a case study with a model peptide with five amino acids

This section is described in the fourth section of supplemtary file **S2.pdf**.

## An ABN-SCS description of protein peptide bond structure: a case study with a model peptide with five amino acids

This section is described in the fifth section of supplemtary file **S2.pdf**.

## An ABN-SCS description of protein backbone and peptide bond structures: Caenopore-5 as an example

To illustrate the redefinition of protein backbone structure with $\rho$, $\theta$ and $\phi$, this article takes Caenopore-5 (Figure 1) as an example, which adopts a helical conformation, forming oligomeric pores upon interaction with lipid bilayers, with five helices stabilized by three disulfide bridges (Figure 3).

RSALSCQMCELVVKKYEGSADKDANVIKKDFDAECKKLFHTIPFGTRECDHYVNSKVDPIIHELEGGTAPKDVCTKLNECP

**Fig 2.** Amino acid sequence and secondary structure of Caenopore-5 (81 amino acids) [56–58]. This figure was prepared by the PSIPRED server [66].

**Fig 3.** Three-dimensional structure of Caenopore-5 (PDB ID: 2JSA) stabilized by three disulfide bonds (red sticks) [56–58]. This figure is prepared by VMD [49].

As the structure of Caenopore-5 was determined by liquid-state NMR spectroscopy [56,57], the structural data deposited in PDB [2] is an NMR ensemble of 15, instead of 1, structural models calculated with restraints from experimental NMR data. As a result, the redefinition of the backbone structure of Caenopore-5 with $[\rho, \theta, \phi]$ is for all 15 structural models [56–58]. Specifically, the backbone structure of Caenopore-5 is described with $[\rho, \theta, \phi]$ according to the fourth section of supplemtary file **S2.pdf**, and the peptide bond structure of Caenopore-5 is described with $[\rho, \theta, \phi]$ according to the fifth section of supplemtary file **S2.pdf**.

# Results

This article used Caenopore-5 [56–58] as an example to define an alternative framework, i.e., ABN-SCS, for protein structure description with [ρ, θ, φ] [67] and the extraction of spherical structural features in addition to those Cartesian structural features used by current protein structure prediction tools [59–65]. In short, for Caenopore-5 [56–58], two sets of spherical parameters (ρ, θ and φ) were extracted from its NMR ensemble (PDB ID: 2JSA) and summarized in supplemtary file **S4.pdf**. Specifically, for the backbone structure of Caenopore-5 [56–58], a set of spherical parameters (a $3 \times 477$ matrix) were extracted from its NMR ensemble (PDB ID: 2JSA) and visualized with Figures 1, 2 and 3 in supplemtary file **S3.pdf** for ρ, θ and φ, respectively; And for the peptide bond structures of Caenopore-5 [56–58], a set of spherical parameters (a $3 \times 80$ matrix) were extracted from its NMR ensemble (PDB ID: 2JSA) and visualized with Figures 4, 5 and 6 in supplemtary file **S3.pdf** for ρ, θ and φ, respectively. Further details of the results and analysis of them are included in supplementary file **S3.pdf**.

# Conclusion

To sum up, this article used Caenopore-5 [56–58] as an example for protein 3D structure description with a spherical coordinate system based on the intrinsic inter-atomic bonding (ABN-SCS) network of protein. Geometrically, this spherical ABN-SCS framework is an uncharted territory, different from the default Cartesian approach since 1971 [1–4], in the sense that ABN-SCS (ρ, θ and φ) not only is a new language for protein structure description [13–18] and understanding [6–8], but also paves a new way for the extraction of protein structure features beyond conventional Cartesian feature extraction (e.g., inter-atomic and inter-residue distances) by computational tools such as AlphaFold [34] and RoseTTAFold [35].

**Fig 4.** Towards a spherical Protein Data Bank in addition to its Cartesian counterpart [1, 2]. This figure represents a simplified example of the PDB-format ATOM record with Cartesian coordinates ([$x$, $y$, $z$]) replaceable by ABN-SCS coordinates [ρ, θ, φ].

Essentially, this spherical ABN-SCS approach is more of a geometric algorithm, through which spherical structural features (ρ, θ and φ) of protein can be extracted from the entire collection of experimentally determined protein structures inside PDB [1–4]. Therefore, this article here proposes a spherical Protein Data Bank on top (Figure 4) of its 54-year-old Cartesian counterpart [1–4], such that additional spherical structural features are extracted and can be incorporated into deep learning algorithms for continued improvement of the performance of protein structure prediction tools [19–22].

# Discussion $\quad$ <sub>139</sub>

## $[\rho, \theta, \phi]$: a new language for protein structure description $\quad$ <sub>140</sub>

As mentioned above, since 1971 [1,2], $[x, y, z]$ has been used as the standard language $\quad$ <sub>141</sub>
(i.e, the default Cartesian coordinate system) for protein structure description $\quad$ <sub>142</sub>
(Figure 1), reflecting an intuitive simplification and conceptualization of protein 3D $\quad$ <sub>143</sub>
structure as a set of discretely distributed points in 3D space, where each point $\quad$ <sub>144</sub>
corresponds to an atom in the protein, and its position is defined by the $[x, y, z]$. From $\quad$ <sub>145</sub>
a linguistic point of view, language shapes and constrains thought, according to the $\quad$ <sub>146</sub>
Sapir-Whorf Hypothesis [68]. In protein structural biology, thus, there is an $\quad$ <sub>147</sub>
interdependence between protein structure understanding [6–8] and protein structure $\quad$ <sub>148</sub>
description [13–18], which is mirrored in the relationship between signifier (description) $\quad$ <sub>149</sub>
and signified (understanding). Therefore, the tools and terminologies used to describe $\quad$ <sub>150</sub>
protein structures shape how scientists conceptualize molecular configurations with $\quad$ <sub>151</sub>
coordinate systems (e.g., CCS [1], G-SCS [14,15], ABN-SCS [16–18]). These descriptive $\quad$ <sub>152</sub>
frameworks not only represent our understanding but actively influence it by focusing $\quad$ <sub>153</sub>
our attention on certain structural aspects while potentially obscuring others. $\quad$ <sub>154</sub>

$\quad$ For instance, here, as the standard Cartesian language, $[x, y, z]$ offers a Cartesian $\quad$ <sub>155</sub>
perspective on how we understand and describe protein structure. In addition to this $\quad$ <sub>156</sub>
Cartesian perspective, nonetheless, $[x, y, z]$ constrains our understanding and $\quad$ <sub>157</sub>
description of protein structure with an intuitive Cartesian simplification and $\quad$ <sub>158</sub>
conceptualization, where one key aspect is not taken into account, i.e., the fact that $\quad$ <sub>159</sub>
structurally, protein is a set of chemically bonded points in 3D space, consisting of an $\quad$ <sub>160</sub>
intrinsic inter-atomic bonding network, instead of a set of discretely distributed points, $\quad$ <sub>161</sub>
which is one of the reasons why this article aims to redefine protein structure with an $\quad$ <sub>162</sub>
atomic bonding networks (ABN) within the SCS framework (ABN-SCS). Moreover, $\quad$ <sub>163</sub>
protein structural feature extraction [69,70], particularly through spherical feature $\quad$ <sub>164</sub>
extraction ($[\rho, \theta, \phi]$) [67], offers a new paradigm for the integration of this ABN-SCS $\quad$ <sub>165</sub>
framework with existing protein structure prediction tools, improving both accuracy $\quad$ <sub>166</sub>
and computational efficiency [17]. $\quad$ <sub>167</sub>

## An ABN-SCS definition of covalent bond length with $\rho$ $\quad$ <sub>168</sub>

In the past few years, while AlphaFold has significantly advanced protein structure $\quad$ <sub>169</sub>
prediction with homologous single-chain sequences [34], it has limitations in its $\quad$ <sub>170</sub>
performance while handling complex systems and dynamic interactions, which $\quad$ <sub>171</sub>
underscore the need for continued development in this field. Here, this article proposes $\quad$ <sub>172</sub>
an ABN-SCS-based protein structure description, which incorporates chemically $\quad$ <sub>173</sub>
accurate constraints to avoid unrealistic bond stretching or compression in protein $\quad$ <sub>174</sub>
structure prediction, because ABN-SCS encodes protein structures with spherical $\quad$ <sub>175</sub>
coordinates $[\rho, \theta, \phi]$, where $\rho$ represents equilibrium bond length, i.e., the inter-nuclear $\quad$ <sub>176</sub>
distances at which system energy is minimized [71,72]. $\quad$ <sub>177</sub>

$\quad$ By constraining the value of $\rho$ to the equilibrium bond lengths in a spherical $\quad$ <sub>178</sub>
coordinate system, this ABN-SCS framework enhances the realism of the predicted $\quad$ <sub>179</sub>
protein structure by maintaining chemically accurate distances between covalently $\quad$ <sub>180</sub>
bonded atoms. To this end, the ABN-SCS framework ensures that the system is aligned $\quad$ <sub>181</sub>
with chemical bonding principles, making sure that inter-atomic distances reflect the $\quad$ <sub>182</sub>
energetically favored configurations of atoms in protein structure, which could be $\quad$ <sub>183</sub>
particularly relevant in the context of force-field-based energy minimization in NMR $\quad$ <sub>184</sub>
data-driven structural calculation or existing protein structure prediction $\quad$ <sub>185</sub>
algorithms [69,70,73,74]. $\quad$ <sub>186</sub>

## Covalent Bond Length in ABN-SCS: is ρ a real constant?

By definition, Cartesian Coordinate System (CCS) requires three parameters, i.e., $[x, y, z]$, to describe protein structures. Here, in this article, the atomic bonding network-based spherical coordinate system (ABN-SCS) also requires three parameters, i.e., $[\rho, \theta, \phi]$, for protein structure description. In theory, $\rho$ is a physical constant, since it is defined as the equilibrium bond length—a physical constant representing inter-nuclear distances that minimize system energy [71, 72]. Thus, the ABN-SCS framework proposed here facilitates a dimensionality reduction by transforming a three-dimensional matrix ($[\rho, \theta, \phi]$) into a two-dimensional spherical vector comprising polar ($\theta$) and azimuthal ($\phi$) angles. As a result, despite using only two spherical parameters, ABN-SCS is able to theoretically preserve the structural information encoded by $[x, y, z]$ in the Cartesian framework [1–4].

**Fig 5.** Distribution of the lengths ($\rho$) of 80 peptide bonds in Caenopore-5 [56–58]. This data corresponds to the first structural model in the NMR ensemble (PDB ID: 2JSA), depicted as the top-left subfigure in Supplementary File **S3.pdf**.

In practice, however, $\rho$ is not a truly fixed value [71, 72]. By definition in the ABN-SCS framework, $\rho$ is the equilibrium bond length, i.e., the distance at which the attractive and repulsive forces between two nuclei are balanced, resulting in the lowest energy configuration [71, 72]. In fact, the value of the equilibrium bond length is highly sensitive to environmental conditions, in a way similar to how atomic nucleus chemical shift (i.e., Larmor or precessional frequency) is sensitive to chemical environmental conditions. For example, factors such as temperature [75, 76] can influence atomic vibrations or bond stretching, while pH variations [58, 77–80] can alter protonation states and electrostatic interactions within protein structure. Moreover, the composition of the buffer and the ionic strength [81] of the surrounding medium can affect the local environment, influencing the forces between atoms and slightly altering the bond

lengths. Taken together, ρ varies dynamically depending on these environmental conditions, rather than remaining constant under all circumstances.

| Model | Mean (Å) | Std (Å) | Min (Å) | Max (Å) | Std/Mean |
|-------|----------|---------|---------|---------|----------|
| 1 | 1.3260 | 0.0092 | 1.3200 | 1.3600 | 0.0070 |
| 2 | 1.3266 | 0.0091 | 1.3200 | 1.3600 | 0.0069 |
| 3 | 1.3265 | 0.0092 | 1.3200 | 1.3600 | 0.0069 |
| 4 | 1.3261 | 0.0092 | 1.3200 | 1.3600 | 0.0069 |
| 5 | 1.3272 | 0.0090 | 1.3200 | 1.3600 | 0.0068 |
| 6 | 1.3261 | 0.0092 | 1.3200 | 1.3600 | 0.0069 |
| 7 | 1.3262 | 0.0092 | 1.3200 | 1.3600 | 0.0069 |
| 8 | 1.3262 | 0.0092 | 1.3200 | 1.3600 | 0.0069 |
| 9 | 1.3274 | 0.0090 | 1.3200 | 1.3600 | 0.0068 |
| 10 | 1.3267 | 0.0091 | 1.3200 | 1.3600 | 0.0069 |
| 11 | 1.3265 | 0.0092 | 1.3200 | 1.3600 | 0.0069 |
| 12 | 1.3272 | 0.0090 | 1.3200 | 1.3600 | 0.0068 |
| 13 | 1.3270 | 0.0091 | 1.3200 | 1.3600 | 0.0068 |
| 14 | 1.3275 | 0.0089 | 1.3200 | 1.3600 | 0.0067 |
| 15 | 1.3267 | 0.0091 | 1.3200 | 1.3600 | 0.0069 |

**Table 1.** A summary of the statistics for the lengths (ρ) of 80 peptide bonds in Caenopore-5 (PDB ID: 2JSA). Each row represents one structural model from the NMR ensemble, with Mean, Std, Min, and Max indicating average bond length, standard deviation, minimum, and maximum values of the lengths (ρ) of 80 peptide bonds, respectively.

Here, with Caenopore-5 [56–58] an example, its 80 peptide bond lengths (ρ) appear rather stable (Figure 5) but also exhibit variations of certain degree across its 81 amino acid residues, as shown in Figure 5 and fifteen subfigures of Figure 4 in supplementary file **S3.pdf**. These variations are represented by the distribution pattern as shown in Figure 5 and quantitatively described by a statistics summary for the lengths (ρ) of 80 peptide bonds in Caenopore-5 (Table 1). Specifically, the 80 bond lengths (ρ) possess an average value of approximately 1.326 Å (Figure 5) and a standard deviation of ∼0.009 Å, yielding a ratio of **Std/Mean** of ∼0.7% (Table 1). Assume that the distribution of ρ is a standard normal one and its **Mean** value equals its equilibrium bond length in theory, its **Mean ± 3 × Std** is able to represent 99.74% of the entire collection of the real situation for the lengths (ρ) of 80 peptide bonds in Caenopore-5. Finally, given that [ρ, θ, φ] is a three-dimension description of protein 3D structure, ρ represents one degree of dimensionality, the variation of ρ represents 3 × **Std/Mean** (Table 1), i.e., 3 × 0.7% = 2.1%. Since the equilibrium bond length in theory is by definition a physical constant, for Caenopore-5 (PDB ID: 2JSA), its structure description does not require a three-dimension description with $[x, y, z]$. Instead, its structure description does not require a 2.021-dimension description with [Std, θ, φ].

Taken together, since ρ is not a truly fixed value [71, 72], the ABN-SCS framework proposed here could not induce a dimensionality reduction by transforming a three-dimensional matrix ([ρ, θ, φ]) into a two-dimensional spherical vector comprising polar (θ) and azimuthal (φ) angles. Yet, the ABN-SCS framework induces a dimensionality reduction from 3 to 2.021 in the case of the 80 peptide bonds of Caenopore-5 [56–58], and this article argues that this dimensionality reduction is applicable not just to the entire 3D structure of Caenopore-5 [56–58], but also to the entire collection of protein structures deposited in the Protein Data Bank [1–4], and that this dimensionality reduction is able to enhance computational efficiency in fields such as molecular dynamics simulations [82].

# Supporting information 

**S1 File.** **S1.pdf** ABN-SCS-based descriptions of the structures of twenty stand-alone 240
natural amino acids. 241

**S2 File.** **S2.pdf** Supplementary Materials and Methods of this manuscript. 242

**S3 File.** **S3.pdf** Supplementary Results of this manuscript. 243

**S4 File.** **S4.pdf** Two sets of spherical parameters ($\rho$, $\theta$ and $\phi$) extracted from the 244
NMR ensemble (PDB ID: 2JSA) of Caenopore-5 [56–58]. 245

**S5 File.** **spherical.py** Python script for ABN-SCS-based extraction of spherical 246
geometric features, i.e., [$\rho$, $\theta$, $\phi$]. 247

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