Transmission Dynamics of Eastern Equine Encephalitis: Global Sensitivity Analysis and SHAP Parameter Importance in an Age-Structured Vector-Host Model

Aurod Ounsinegad

Department of Biostatistics and Medical Informatics, University of Wisconsin–Madison, Madison, Wisconsin 53703, United States

Christopher Mitchell*

Department of Quantitative and Qualitative Health Sciences, University of Texas San Antonio School of Public Health, San Antonio, Texas 78229, United States
mitchellc4@uthscsa.edu

Nicholas Komar

Division of Vector-Borne Diseases, Center for Disease Control and Prevention, Fort Collins, Colorado 80521, United States

Eastern equine encephalitis virus (EEEV) is a deadly arboviral pathogen with 30% severe case fatality. EEEV exhibits pronounced 2–3 year cyclical outbreak patterns in the northeastern United States, linked to shifts in mosquito feeding preferences between hatch-year and adult avians. We developed an age-structured vector-host model incorporating differential feeding patterns of *Culiseta melanura* mosquitoes on European Starlings and American Robins. Global sensitivity analysis revealed mosquito biting rate (a) as the dominant driver in transmission, with avian infectivity (δ) and exposure (α) playing secondary roles. Pairing tree-based machine learning algorithms with SHAP analysis on 100,000 parameter sets identified parameter hierarchies that govern cyclic transmission. Adult avian mortality (μ_a) was identified as the key parameter underlying cyclic and stable transmission patterns. SHAP further revealed these patterns to be grounded on opposite sides of the same epidemiological mechanism. Stable endemics emerge from demographic stability paired with transmission optimization, while cyclic endemics emerge from demographic instability paired with minimal transmission optimization. Numerical simulations illustrated critical threshold dynamics at $0.2 \leq \alpha \leq 0.4$, where heightened hatch-year exposure triggers demographic instability responsible for observed 2–3 year cycles, while balanced exposure ($\alpha \approx 0.5$) leads to stable endemics. These mechanisms provide a foundation for targeted surveillance and control interventions.

Keywords: Age-structured vector-host modeling; Sobol sensitivity analysis; SHAP parameter importance

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*Corresponding Author.

1. Introduction

Eastern equine encephalitis virus (EEEV) is an arthropod-borne alphavirus first detected in humans in the United States in 1938, representing one of the most severe mosquito-borne pathogens in North America [3]. EEEV causes acute neuroinvasive disease with case fatality rates of approximately 30% among patients who develop severe neurological symptoms, and up to 90% in equine cases [3, 35]. The virus is maintained in an enzootic cycle primarily involving the mosquito vector, the Black-Tailed Mosquito (*Culiseta melanura*) in this paper, and avians, with occasional spillover to dead-end hosts such as humans and horses [8].

EEEV transmission exhibits distinct regional patterns across the eastern coast of the United States. In northern latitudes (e.g., Massachusetts, Connecticut, New Hampshire), surveillance data reveals pronounced 2–3 year epidemic cycles characterized by periods of intense transmission followed by apparent dormancy. Long-term surveillance in Connecticut demonstrates clear periodicity in viral activity, with mosquito abundance significantly correlated with EEEV detection (r = 0.69, p < 0.001) [2, 13]. Conversely, southern latitudes, particularly Florida, show more consistent endemic circulation with less pronounced cyclic patterns [12, 33].

Recent field studies have revealed that these transmission dynamics depend on differential mosquito feeding preferences based on the age structure of avian hosts. Throughout this study, we use young host/hatch-year avian, adult host/adult avian, and vector/mosquito interchangeably. Hatch-year avians experience 2.5–3.5 times higher infection rates than adults, creating an amplification effect during breeding seasons. Blood meal analysis reveals that *C. melanura* exhibits temporal feeding shifts, taking 26.9% of blood meals from Northern Cardinals year-round but increasing to over 40% during peak transmission periods in June [26]. This temporal synchronization between mosquito feeding behavior and avian breeding cycles provides a mechanistic explanation for observed outbreak patterns [2, 13].

The quantitative relationship between avian host demographics and EEEV transmission dynamics remains poorly understood. While previous modeling efforts have broadly examined EEEV transmission dynamics [27, 34], they have yet to explore the biological mechanisms driving the pronounced 2–3 year endemic cycles observed in northeastern states versus stable endemic patterns in southeastern states. Understanding these regional differences is critical for predicting outbreak patterns and developing targeted intervention strategies.

Mathematical modeling demonstrates that vector-host interactions can fundamentally alter transmission patterns through complex demographic processes [16, 23]. When vectors exhibit feeding preferences among host age classes, this can produce "diversity amplification" effects that concentrate transmission [23], while other configurations produce "dilution effects" that reduce transmission [16]. The complex interactions between mosquito vectors and avian hosts preceding human and equine cases exhibit various dynamics that influence regional infection patterns [10–12, 24].

Although demographic differences between young and adult avians may influence

transmission [19, 29], the mechanistic drivers of regional outbreak patterns remain unclear. While quantitative analysis demonstrates that vector populations determine outbreak potential [7], the biological parameters governing whether transmission exhibits stable or cyclic dynamics have not been systematically identified.

This study develops an age-structured mathematical model combined with comprehensive sensitivity analysis and tree-based machine learning approaches to identify the biological mechanisms driving EEEV transmission patterns. We focus on the enzootic cycle involving *C. melanura* and two key avian species: European Starlings (*Sturnus vulgaris*) and American Robins (*Turdus migratorius*) [18]. Using Sobol global sensitivity analysis and SHAP feature importance applied to machine learning analysis of 100,000 biologically plausible parameter combinations, we identify distinct biological mechanisms governing transmission potential versus dynamical stability in EEEV. Our findings reveal that transmission efficiency parameters, particularly mosquito biting rate and the avian exposure coefficient, control outbreak potential, while demographic parameters such as adult avian mortality serve as primary drivers of cyclic versus stable dynamics. Model simulations validate these analytical findings by reproducing the three transmission regimes observed across the eastern United States. The resulting age-structured framework provides mechanistic insight into EEEV outbreak patterns and identifies key biological targets for intervention.

Fig. 1: EEEV stage-structured model flow chart. The top left, top right, and bottom center boxes represent hatch-year (y), adult avian (a), and mosquito (M) classes, respectively. Solid directional arrows connect stages within each class, indicate their interactions, and show entrance and exit into the system. Dashed directional arrows show maturation between young and adult avian classes, where it is assumed young hosts recover before maturing. Dashed lines between vectors and hosts show bidirectional viral transmission, where **1.** represents feeding success (successful viral transmission) and host preference.

2. Model Development

In developing the stage-structured model, the host avian class is split into hatch-year avians (y) and adult avians (a) to better understand host age-specific transmission dynamics where both avian classes contain three stages: susceptible (S), infected (I), and recovered (R) (Fig. 1). Both classes have recovered stages where they are classified as non-competent hosts, R_y (Eq. 2.3) and R_a (Eq. 2.6), respectively, implying an inability for transmission. For this model, it is assumed that once an individual from either avian class enters the recovered stage, they remain recovered for life. This is assumed as both avian species considered for this study maintain recovery status for the same amount of time as their average lifespan [17, 25].

Young hosts mature into adults at a rate m_y from the susceptible and recovered stages, but not the infected stage. It is assumed that young hosts must recover and enter the recovered stage before they can mature. It is also assumed that there is a constant birth rate, ψ_B, within the avian population paired with constant natural mortality rates for both hosts in each stage given by μ_y and μ_a, respectively. It should be noted that the carrying capacity term, $\left(1 - \frac{S_y}{c}\right)$, keeps logistic growth of the avian population within the model while simultaneously keeping the population bounded, where c denotes the numerical carrying capacity.

Through the development of the vector transmission dynamics within this model, it was determined that the mosquito class has a different stage structure compared to that of the avian host; the mosquito class moves between a susceptible (S), latent (L), and infected (I) stage. It is assumed that all mosquitoes are recruited into the susceptible class, S_M (Eq. 2.7), and that a vector must move from the latent stage, L_M (Eq. 2.8), into the infected stage, I_M (Eq. 2.9), before they are considered infectious. This is because an extrinsic incubation period must pass before infection can take place [30, 34]. Additionally, once a mosquito has made it to the infectious stage, it is assumed that it remains infectious for life. A constant birth rate, ψ_M, and mortality rate, μ_M in and out of the mosquito class is also assumed.

It was observed that the relative number of mosquito bites on each avian species is determined by the number of female mosquitoes present and the abundance of avian hosts within the model [24]. However, the value for the biting rate parameter, a is simplified with this model, whereas the realistic transmission of EEEV occurs between a variety of different host and vector species.

Mosquito bites are distributed among both the young and adult host classes, where the exposure coefficient of the hosts to mosquito bites is given by $1 - \alpha$ and α, respectively. If $\alpha \approx 0.5$, then each avian class is bitten in proportion to the abundance of its respective population, and all avians receive a similar number of bites. Higher α values correlate with increased adult host exposure, whereas lower α values correlate with increased young host exposure.

Susceptible hosts, S_y (Eq. 2.1) and S_a (Eq. 2.4), can move into their respective infectious class, I_y (Eq. 2.2) and I_a (Eq. 2.5), once bitten by an infectious mosquito vector, I_M, dependent on their exposure to the virus (α). Although it has been

observed that EEEV can be transmitted vertically within the avian population [30], vertical transmission is ignored within the model, and the principal method of transmission is observed to occur between vectors and hosts during the viral amplification season [19, 22]. Transmission of the virus from infectious mosquito vectors to susceptible hosts is partially determined by the proportion of susceptible hosts to the total population of avians N_B, $\frac{S_y}{N_B}$ and $\frac{S_a}{N_B}$. The class-dependent virulence is given by ν_y and ν_a and the class-dependent recovery rate, is given by γ_y and γ_a. To analyze the independent effect of differential avian class viral exposure, it is assumed that infectivity and recovery rate baseline values of each avian class are equal.

Mosquitoes within the susceptible stage can move into the latent stage, L_M, after successfully biting an infectious host. Transmission of the virus from infectious avian hosts to susceptible mosquito vectors is partially determined by the proportion of susceptible mosquitoes to the total population of mosquitoes (N_M), $\frac{S_M}{N_M}$. Susceptible mosquitoes take a bites per day, with respect to both host classes, where only a fraction of the bites taken are from infectious hosts, $\frac{I_y}{N_B}$ and $\frac{I_a}{N_B}$. The probability that vector-host transmission will successfully occur per bite, also known as avian infectivity, is given by δ. Latent mosquitoes move into the infectious stage at a rate k, where $\frac{1}{k}$ is the extrinsic incubation period of the virus that must pass before a mosquito can be considered fully infectious and transmit the virus. Although it has been observed that EEEV can affect the reproductive fitness of *C. melanura* [6], due to short life span, it is assumed that infectious mosquitoes do not have a virulence, never recover, and can only exit the system through natural death.

$$\frac{dS_y}{dt} = \psi_B(S_a + R_a)\left(1 - \frac{S_y + R_y + S_a + R_a}{c}\right) - a(1-\alpha)I_M\left(\frac{S_y}{N_B}\right) - (m_y + \mu_y)S_y \tag{2.1}$$

$$\frac{dI_y}{dt} = a(1-\alpha)I_M\left(\frac{S_y}{N_B}\right) - (\gamma_y + \mu_y + \nu_y)I_y \tag{2.2}$$

$$\frac{dR_y}{dt} = \gamma_y I_y - (m_y + \mu_y)R_y \tag{2.3}$$

$$\frac{dS_a}{dt} = m_y S_y - a\alpha I_M\left(\frac{S_a}{N_B}\right) - \mu_a S_a \tag{2.4}$$

$$\frac{dI_a}{dt} = a\alpha I_M\left(\frac{S_a}{N_B}\right) - (\gamma_a + \mu_a + \nu_a)I_a \tag{2.5}$$

$$\frac{dR_a}{dt} = m_y R_y + \gamma_a I_a - \mu_a R_a \tag{2.6}$$

$$\frac{dS_M}{dt} = \psi_M - a\left(\frac{S_M}{N_M}\right)\delta\left((1-\alpha)I_y + \alpha I_a\right) - \mu_M S_M \tag{2.7}$$

$$\frac{dL_M}{dt} = a\left(\frac{S_M}{N_M}\right)\delta\left((1-\alpha)I_y + \alpha I_a\right) - (k + \mu_M)L_M \tag{2.8}$$

$$\frac{dI_M}{dt} = kL_M - \mu_M I_M \tag{2.9}$$

All model parameters are assumed positive. The system of ordinary differential equations the model is comprised of is presented above, with complete parameter descriptions and values in Table 1.

Table 1: EEEV stage-structured model parameter list. Note that total population counts for avians and mosquitoes (N_B and N_M, respectively) don't have references due to their constant change in model simulations.

Parameter	Description	Value	Reference
ψ_B	Hatch-year avian birth rate	0.05 avians day^{-1}/avians	[17, 25]
a	Mosquito biting rate	2.5 day^{-1}	[24]
α	Avian exposure coefficient	$0 \leq \alpha \leq 0.5$	Varied
γ_y	Hatch-year avian recovery rate	$\frac{1}{3}$ day^{-1}	[19]
γ_a	Adult avian recovery rate	$\frac{1}{3}$ day^{-1}	[19]
μ_y	Hatch-year avian death rate	0.002 day^{-1}	[17, 25]
μ_a	Adult avian death rate	$\frac{1}{547}$ day^{-1}	[17, 25]
μ_M	Mosquito death rate	0.1 day^{-1}	[6]
ν_y	Hatch-year avian virulence	0.5 day^{-1}	[22]
ν_a	Adult avian virulence	0.5 day^{-1}	[22]
ψ_M	Mosquito birth rate	3,000 mosquitoes day^{-1}	[6]
δ	Avian infectivity	0.2	[19]
k	Extrinsic viral incubation period	$\frac{1}{3}$ day^{-1}	[30]
m_y	Avian maturation rate	0.04 day^{-1}	[32]
N_B	Total population of avians	Varies	
N_M	Total population of mosquitoes	Varies	
c	Avian population carrying capacity	Varies	[17, 25]

Susceptible young hosts (S_y) are recruited at a rate of ψ_B, where it is assumed that a majority of female avians are capable of laying two clutches of eggs per year with a mean clutch size of five [17, 25]. To bound the growth of the avian population, a carrying capacity c is used, where it is assumed that the carrying capacity of the avian population will remain constant.

3. Sensitivity Analysis

A comprehensive sensitivity analysis was conducted to identify and quantify the relative influence of model parameters on peak infection levels across three key populations: hatch-year avians (I_y), adult avians (I_a), and mosquitoes (I_M). The basic reproduction number $\mathcal{R}_0$ was derived using the next generation matrix approach:

$$\mathcal{R}_0 = \frac{a\sqrt{k}\sqrt{\delta}\sqrt{(m_Y + \mu_A)((1-\alpha)^2\mu_A(\gamma_A + \mu_A + \nu_A) + m_Y\alpha^2(\gamma_Y + \mu_Y + \nu_Y))}}{(m_Y + \mu_A)\sqrt{\mu_M}\sqrt{k + \mu_M}\sqrt{\gamma_A + \mu_A + \nu_A}\sqrt{\gamma_Y + \mu_Y + \nu_Y}}$$

$$(3.1)$$

3.1. *Parameter Sensitivity Patterns and Mechanistic Interpretation*

Sobol sensitivity analysis is a variance-based global sensitivity method that quantifies the contribution of each input parameter to the total variance in model outputs [31]. The method decomposes output variance into components attributable to individual parameters (first-order indices, S1) and their interactions with other parameters (total-effect indices, ST). First-order indices measure the direct effect of a parameter when varied independently, while total-effect indices capture both direct effects and all interactions involving that parameter. This approach is particularly valuable for complex biological models where parameter interactions may be substantial, as it provides model-independent measures of parameter importance across the entire parameter space rather than at a single point.

Fig. 2: Local sensitivity analysis for EEEV transmission showing percentage changes in model outputs when parameters are varied by $\pm 10\%$ and $\pm 20\%$. **Top row:** Total peak infections, peak young infected (I_y), and peak adult infected (I_a). **Bottom row:** Peak infected mosquitoes (I_M) and basic reproduction number $(\mathcal{R}_0)$. Red bars indicate negative parameter effects while blue bars indicate positive effects.

Both local (Fig. 2) and global sensitivity (Fig. 3) analyses demonstrate that mosquito biting rate (a) emerges as the most important parameter across all model outputs, with Sobol total-effect indices (ST) ranging from 0.368 to 0.491. This dominance validates the central role of vector-host contact frequency in arboviral transmission [4, 24]. Avian infectivity (δ) consistently ranks second (ST values

0.120–0.231), matching patterns reported for other vector-borne diseases where transmission efficiency parameters dominate system behavior [29].

Fig. 3: Global sensitivity analysis using Sobol indices for EEEV transmission. Red bars represent the five highest paramters indices, while blue/green are lower values. Shows first-order effects (S1: light-red) and total effects (ST: dark-red) for each parameter across multiple model outputs. Higher Sobol index values indicate greater parameter importance in determining model output variability.

The parameter hierarchy shows agreement between local and global sensitivity analyses for the top parameters (a, δ, μ_M), confirming that parameter influences reflect genuine biological processes rather than methodological artifacts. The influence of demographic parameters $(ST < 0.11)$ compared to transmission parameters validates that EEEV outbreaks are driven by transmission efficiency rather than virulence alone, consistent with field observations [10].

3.2. *Age-Structured Population Responses and Vector Dynamics*

Different sensitivity patterns between hatch-year and adult avian populations validate our age-structured approach. Both avian classes show strong sensitivity to mosquito biting rate, with adults being more responsive (ST = 0.491) than young avians (ST = 0.390). However, hatch-year avians show unique sensitivity to carrying capacity (c; ST = 0.171) while adults show minimal response (ST = 0.046).

The avian exposure coefficient (α) reveals important age-specific patterns: young

avians show minimal sensitivity (ST = 0.011), adults demonstrate moderate sensitivity (ST = 0.100), and α strongly influences $\mathcal{R}_0$ (ST = 0.212). This pattern indicates that differential exposure between age classes becomes critical for determining overall transmission potential, validating biological evidence that mosquito feeding preferences significantly alter outbreak dynamics [24].

For vector populations, while mosquito biting rate maintains dominance (ST = 0.368), the mosquito death rate emerges as the second most influential parameter (ST = 0.257), validating the critical importance of vector longevity for EEEV transmission. This confirms that vectors must survive long enough for viral development and subsequent transmission, aligning with field observations that environmental factors affecting mosquito survival strongly influence outbreak probability [7, 26].

4. Machine Learning Analysis and Parameter Importance

Fig. 4: Parameter correlation matrix showing very low correlation values across all parameters ($0.0 \leq |r| \leq 0.25$), indicating negligible collinearity between parameters, supporting independent parameter influence on system behavior. Grey to blue boxes show negative correlations, and orange to red boxes show positive correlations.

To quantify parameter interactions and identify biological mechanisms driving different endemic equilibria, we employed tree-based machine learning algorithms on 100,000 biologically plausible parameter sets. Based on epidemiological observations

of transmission patterns across different regions [2,7], we classified endemic equilibria into two categories: Stable Endemic Equilibria (Stable EE), seen as persistent non-oscillatory transmission comprising $\sim 98.5\%$ of all endemic equilibria; and Unstable Endemic Equilibria (Unstable EE), seen as rare cyclic dynamics with 2–3 year periodicity comprising $\sim 1.5\%$ of all endemic equilibria. The extreme class imbalance between stable and unstable EE reflects the rarity of cyclic EEEV behavior.

Parameter correlation analysis showed very low correlation and confirmed independence between all biological parameters ($0.0 \leq |r| \leq 0.25$), validating that each parameter contributes unique information without confounding analyses (Fig. 4).

4.1. *Methodology and Model Performance*

Three machine learning algorithms were chosen for their ability to capture non-linear parameter interactions while providing interpretable parameter importance measures: Random Forests (RF) [5,14], eXtreme Gradient Boosted trees (XGB) [9], and Light Gradient Boosted Machine (LGBM) [15]. Tree-based algorithms excel at modeling threshold effects and parameter interactions characterizing biological systems, making them ideal for understanding EEEV's complex transmission dynamics [21].

Fig. 5: Model performance comparison: AUROC (top right), AUPRC (top center), AUROC Bar Plot comparison between cross-validated data and test data performance (top left), and Confusion Matrices (bottom row of plots; from right to left: RF, XGB, and LGBM). Given that all models overfit due to class imbalance, LGBM performed the best (AUROC: 0.991; AUPRC: 0.625), followed by XGB (AUROC: 0.991; AUPRC: 0.608), and lastly RF (AUROC: 0.978; AUPRC: 0.426).

All models were optimized using Optuna Bayesian hyperparameter tuning and

5-fold stratified cross-validation over 30 trials to maintain proper class distribution [1, 28]. Particularly, due to the extreme class imbalance, we utilized Area Under the Precision-Recall Curve (AUPRC) in addition to Area Under the Receiver Operating Curve (AUROC) metrics to track any indication of overfitting. AUROC metrics can be misleading when there is an extreme class imbalance present in the data, which is where AUPRC becomes a more desirable performance metric.

With unstable EE set as the target variable, all models overfit due to class imbalance, with LGBM overfitting least, showing better overall generalization (Fig. 5).

4.2. *Shapley Value Parameter Importance Analysis*

Fig. 6: Unstable EE LGBM Model SHAP Summary Plot showing SHAP value distributions for all parameters with μ_a, α, and ν_a ranking highest (parameter importance is read from top to bottom). Color gradients indicate parameter values with blue (cooler colors) being low, purple (warmer colors) being mid, and pink (hottest colors) being high parameter values. Positive/Negative SHAP values represent positive/negative impact on the model's output, respectively. The combination of each point's color and SHAP value provides information about whether a parameter has negative/positive/no correlation with model output. **Note:** The color gradient label, "Feature Value", is the same as "Parameter Value".

SHAP (SHapley Additive exPlanations) value analysis, a model-agnostic parameter importance algorithm based on game theory [20], revealed distinct mechanistic drivers for both types of endemic equilibria. SHAP values offer local explanations for individual predictions and global parameter importance rankings, enabling comprehensive understandings of how each parameter drives endemic behavior across the parameter space. To allow for meaningful comparison across models, SHAP was normalized by dividing each feature's SHAP value by the maximum absolute value within that model. This normalization approach preserves the relative importance rankings while standardizing the scale across models: $\text{SHAP}_{\text{norm}}(f_i) = \frac{\text{SHAP}(f_i)}{\max_j |\text{SHAP}(f_j)|}$, where f_i represents the i-th feature. This ensures parameter importance rankings reflect biological influence rather than model-specific architectures.

One common way to visualize SHAP values is through the use of SHAP Summary Plots (Fig. 6). In summary plots, each point represents one case (e.g., an unstable or stable EE), and each case will have exactly one point for each of the parameters shown on the y-axis. As can be seen in Fig. 6, the color of the points represents the value for each respective parameter, with blue (cooler colors) being low parameter values, purple (warmer colors) being mid parameter values, and pink (hottest colors) being high parameter values. Lastly, the x-axis shows the SHAP value for each respective point, with negative SHAP values representing a negative impact on the model's output (e.g., the parameter is pushing the model to predict "Stable EE" for the respective case) and positive SHAP values representing a positive impact on the model's output (e.g., the parameter is pushing the model to predict "Unstable EE" for the respective case). The combination of the color of each point (parameter value) and the respective SHAP value (negative/positive) in the plot provides information about whether a parameter has negative/positive/no correlation with model output.

Demographic instability mechanisms tended to dominate across all models with the adult death rate (μ_a), avian exposure coefficient (α), adult virulence (ν_a), mosquito death rate (μ_m), and mosquito biting rate (a) surfacing as the top five most important parameters in order (Fig. 6 shows this in the best-performing model, LGBM). μ_a, α, and a all show a negative correlation between parameter and SHAP values with average absolute SHAP values of ~ 0.455, 0.221, and 0.107, respectively. ν_a and μ_m; however, show clear positive correlation between parameter and SHAP values with average absolute SHAP values of ~ 0.182 and 0.140 respectively.

This hierarchy uncovers two main drivers in both types of endemic equilibria. First, with μ_a achieving importance values up to twice those of the second most important parameter, this shows that adult avian population stability in the system is crucial in maintaining cyclic transmission patterns. Second, α exhibits a more complex negative correlation than μ_a or a, demonstrating that higher exposure in adults ($\alpha \geq 0.7$) results in more stable EE; however, when exposure begins to favor young avians around the mid-to-low range ($0.2 \leq \alpha \leq 0.4$), you see more unstable EE. Overall, the hierarchy of parameter importance indicates that cyclic dynamics emerge more through population-level demographics rather than transmission optimization.

The SHAP ridge plot further reveals the underlying distribution of the demographic instability mechanism through a fine-tuned balance between population-level parameters—parameters that can be modulated through habitat conservation and demographic support during vulnerable periods (Fig. 7 shows this in the best-performing model, LGBM).

Fig. 7: Unstable EE LGBM Model SHAP Ridge Plot comparison showing SHAP value distributions for all model parameters with top parameters μ_a, α, and ν_a (parameter importance is read from bottom to top—opposite of Fig. 6). Each parameter's respective ridge plot shows the broad distribution of SHAP values, with higher ridges indicating a greater frequency of the respective SHAP value.

4.3. *Biological Validation and Intervention Implications*

The unique parameter importance hierarchy shows that stable and unstable EE are both grounded in the same epidemiological demographic mechanism, but in fundamentally different ways. Stable EE depends more on demographic stability and transmission optimization through higher mosquito biting rates as well as higher natural death and exposure rates in adult avians paired with lower death in mosquitoes and virulence in adult avians (reflecting true transmission patterns in the southeastern United States) [33]. Unstable EE depends on the polar opposite: demographic instability and minimal transmission optimization through lower natural death, but higher virulence, in adults, higher exposure in young avians, and higher death, but lower biting rates, in mosquitoes (reflecting the true complex transmission patterns in the northeastern United States) [2, 13].

Mann-Whitney U-tests revealed significant parameter distribution differences ($p < 0.001$) between stable and unstable EE for over half the model parameters. Sensitivity and SHAP analysis approaches reveal complementary yet distinct parameter importance hierarchies. Sobol sensitivity analysis consistently identifies mosquito biting rate (a) as the most dominant parameter across model outputs ($ST > 0.4$), reflecting its control over transmission intensity. In contrast, SHAP analysis highlights demographic parameters as primary drivers of model behavior, with adult mortality (μ_a) showing the highest parameter importance (SHAP = 0.455), followed by exposure coefficient (α; SHAP = 0.221) and adult virulence (ν_a; SHAP = 0.182). Both methods generally agreed on the critical importance of the exposure coefficient, though they emphasized different aspects of its influence on system dynamics. This methodological divergence suggests that global sensitivity analysis captures transmission threshold effects while machine learning approaches emphasize parameters governing dynamical stability and oscillatory behavior within endemic states.

Since both types of endemic equilibria are grounded in the same epidemiological demographic mechanism, targeted intervention strategies are quite straightforward: emphasize host population stability (habitat conservation, disease surveillance, focus on adult avian health, etc.). The rarity of unstable EE (1.5% of endemic equilibria) reflects the biological reality that cyclic transmission represents exceptional rather than typical EEEV behavior.

5. Model Simulations and Results

The host stage-structured EEEV model was implemented using parameter values from Table 1 with systematic variation of key parameters identified through sensitivity and data analysis. Model simulations validate both the theoretical $\mathcal{R}_0$ framework and parameter importance hierarchies revealed through global sensitivity analysis and machine learning approaches.

Numerical simulations confirm that the calculated $\mathcal{R}_0$ (Eq. 3.1) accurately predicts transmission outcomes across the parameter space. Fig. 8 demonstrates how the

avian exposure coefficient (α) interacts with key transmission parameters identified in previous sections. Panels (A) and (C) show nearly vertical $\mathcal{R}_0 = 1$ boundaries, indicating that the exposure coefficient dominates transmission viability over both adult avian virulence (ν_a) and mortality (μ_a). Panel (B) reveals that higher mosquito mortality (μ_M) requires a correspondingly higher exposure coefficient to maintain $\mathcal{R}_0 > 1$, confirming vector survival as a critical constraint. Panel (D) shows that despite wide variation in mosquito biting rate (a), viable transmission requires exposure coefficients within a narrow window, creating a steep threshold that validates the importance of balanced age-structured exposure. Panel (E) demonstrates that avian infectivity (δ) operates within an exceedingly narrow range, with little influence from the exposure coefficient, suggesting a limited threshold effect despite its secondary importance in sensitivity analyses.

Fig. 8: $\mathcal{R}_0$ heatmap showing interactions between avian exposure coefficient (α) and key transmission parameters. **(A)** Adult avian virulence (ν_a) versus exposure coefficient showing demographic effects. **(B)** Mosquito mortality (μ_M) versus exposure coefficient demonstrating vector survival constraints. **(C)** Adult avian mortality (μ_a) versus exposure coefficient showing demographic effects. **(D)** Mosquito biting rate (a) versus exposure coefficient revealing the primary transmission threshold. **(E)** Avian infectivity (δ) versus exposure coefficient displaying transmission efficiency boundaries. White lines indicate $\mathcal{R}_0 = 1$ thresholds separating viable transmission from extinction.

Systematic parameter exploration confirms three distinct dynamical patterns predicted by $\mathcal{R}_0$ analysis and stability theory. Fig. 9 illustrates these dynamics using biologically realistic parameter values, where outcomes are determined by $\mathcal{R}_0$ and demographic stability characteristics.

Fig. 9: Population dynamics of total infected vectors/hosts across dynamical patterns. **Top row:** Total infected avians (filled curves) illustrating distinct dynamical patterns over extended temporal periods. **Bottom row:** Total infected mosquitoes (filled curves) showing corresponding vector dynamics. The three dynamical patterns illustrate fundamentally different long-term behaviors: extinction following initial outbreak (DFE), convergence to stable endemic levels (Stable EE), and persistent oscillations with 2–3 year periodicity (Cyclic/Unstable EE), consistent with observed EEEV outbreak patterns in nature.

Disease-Free Equilibrium ($\mathcal{R}_0 = 0.950$): Parameter combinations with low mosquito biting rate ($a = 0.8\,\mathrm{day}^{-1}$) and high mosquito mortality ($\mu_M = 0.18\,\mathrm{day}^{-1}$) produce realistic outbreak dynamics that peak around 5–6 months, and end up falling into complete extinction over 2–3 years. This pattern matches observed EEEV outbreaks in marginal habitats where environmental conditions prevent sustained transmission despite initial amplification [2, 26].

Stable Endemic Equilibrium ($\mathcal{R}_0 = 1.570$): Balanced parameter values ($a = 2.2\,\mathrm{day}^{-1}$, $\alpha = 0.5$, $\mu_M = 0.08\,\mathrm{day}^{-1}$) result in stable transmission that rapidly converge to an equilibrium within 2–3 years. While this simulation demonstrates *standard* stable dynamical patterns, SHAP analyses revealed that *optimal* stable endemic conditions are characterized by higher mosquito biting rates and exposure coefficients combined with lower mosquito mortality. It is further verified that these balanced parameters better represent a standard rather than an optimal stable

configuration, as seen in Fig. 6, since the stable EE (negative SHAP values) favors parameter combinations that result in enhanced transmission efficiency.

Cyclic Dynamics ($\mathcal{R}_0 = 1.170$): Base parameter values from Table 1 ($\psi_B = 0.05$, $c = 50{,}000$, $a = 2.5$ day^{-1}, $\alpha = 0.3$) produce pronounced 2–3 year oscillatory behavior matching northeastern surveillance patterns. Despite having a moderate $\mathcal{R}_0$, intermediate exposure coefficient values create demographic instability where preferential hatch-year exposure leads to periodic infection outbreaks followed by population recovery cycles. Consequently, this generates sustained oscillations rather than a stable endemic transmission.

5.1. *Age-Structured Mechanisms*

Fig. 10: Age-structured population dynamics across dynamical patterns. **Top row:** Young (orange) and adult (purple) infected avian populations demonstrate differential age-structured responses. The disease-free equilibrium ($\mathcal{R}_0 = 0.950$) shows rapid extinction after the initial outbreak around month 5–6. The stable EE ($\mathcal{R}_0 = 1.570$) exhibits sustained transmission after the first peak with adults maintaining higher infection levels through time. Cyclic dynamics ($\mathcal{R}_0 = 1.170$) display pronounced oscillations every 2–3 years, driven by age-structured amplification effects. **Bottom row:** Corresponding mosquito stage dynamics showing latent (green) and infected (red) populations, with cyclic dynamical patterns demonstrating synchronized periodic outbreaks in both compartments.

Detailed analysis of age-structured dynamics reveals the biological mechanisms underlying each dynamical pattern. Fig. 10 demonstrates how differential exposure between young and adult hosts drives system-level behavior, validating theoretical predictions about the critical role of host age-structure in EEEV transmission.

In the disease-free equilibrium, both age classes show a similar rapid convergence to extinction following initial exposure, confirming that $\mathcal{R}_0 < 1$ prevents sustained transmission regardless of age-specific differences. Both age classes experience initial infection peaks, with adults reaching higher infection levels than young, before infections completely die out.

Stable endemic dynamics illustrate sustained transmission with adult avians facing higher infection levels than young avians due to longer lifespans and slower demographic turnover. This age-structured pattern emerges under balanced parameter conditions that avoid demographic oscillations seen in cyclic dynamics, consistent with SHAP analysis indicating higher α values favor transmission stability (Fig. 6).

Cyclic dynamics reveal the mechanistic basis of oscillatory transmission through pronounced age-structured instability. Young avians reach peak infections of approximately 120 individuals during outbreak years before declining during inter-epidemic periods, while adult avians display larger amplitude oscillations (~ 280 peak infections) with similar decline patterns. This demographic instability emerges when $\alpha = 0.3$, indicating preferential exposure in young hosts, consistent with SHAP analysis identifying lower α values as drivers of cyclic behavior. The resulting oscillatory patterns exhibit biologically plausible periodic behavior with multi-year cycle characteristics. Mosquito infection dynamics exhibit fluctuations between epidemic peaks (~ 600 peak infected vectors) and inter-epidemic minimums approaching near-extinction levels, reflecting the characteristic cyclical population dynamics of this transmission pattern.

These simulation results demonstrate strong correspondence with parameter importance rankings identified through sensitivity analysis and machine learning approaches. The mosquito biting rate (a) exhibits its sensitivity-predicted dominant influence by controlling transitions across the $\mathcal{R}_0 = 1$ threshold that separates persistence from extinction. Meanwhile, the exposure coefficient (α) confirms its critical role in determining stability versus oscillatory dynamics above the same threshold. The quantitative agreement between theoretical predictions and simulated outcomes supports the EEEV age-structured model framework's ability to capture essential biological mechanisms governing transmission. Distinct parameter combinations produce the three dynamical patterns at thresholds consistent with $\mathcal{R}_0$ analysis and mechanistic pathways identified through SHAP analysis.

6. Discussion

Our integration of Sobol global sensitivity analysis, SHAP parameter importance, and dynamical simulations provides complementary insights into EEEV transmission mechanisms. Sobol analysis quantifies parameter influence on transmission potential,

SHAP analysis identifies demographic drivers of endemic behavior, and model simulations validate theoretical predictions against observed patterns. This multi-method approach underscores the necessity of integrating diverse analytical perspectives to achieve a complete understanding of complex biological systems [31].

Sensitivity analyses present different parameter hierarchies that reveal distinct biological mechanisms. Sobol analysis ranks mosquito biting rate (a) as dominant (ST = 0.368–0.491), followed by avian infectivity (δ; ST = 0.120–0.231) and mosquito mortality (μ_M; ST = 0.257). In contrast, SHAP analysis ranks adult avian mortality (μ_a) as the most important parameter, followed by exposure coefficient (α), adult virulence (ν_a), mosquito mortality (μ_m), and mosquito biting rate (a). Remarkably, while μ_a achieves SHAP importance values up to twice those of the second most important parameter, it exhibits minimal Sobol sensitivity. This provides critical biological insight: while μ_a barely affects whether transmission can occur, it fundamentally determines whether endemic transmission exhibits stable or oscillatory dynamics within the viable parameter space.

The exposure coefficient (α) demonstrates consistent importance values across both analytical approaches and aligns well with simulation results. This convergence is evident across all analyses: Sobol analysis demonstrates α's strong influence on $\mathcal{R}_0$ (ST = 0.212), SHAP analysis (SHAP = 0.221) identifies threshold behavior at intermediate values (0.2–0.4) that generate demographic instability, and simulations confirm these predictions by reproducing cyclic dynamics exclusively within this critical range. The agreement between methods validates α as a key parameter, with simulations confirming that preferential exposure in young avians generates the demographic instability characteristic of northeastern EEEV patterns, while balanced exposure ($\alpha = 0.5$) yields stable dynamics in both analytical frameworks.

Cyclic dynamics prove remarkably rare, occurring in only 1.5% of endemic equilibria across 100,000 parameter combinations analyzed using tree-based machine learning algorithms. Model simulations validate this finding by illustrating three distinct transmission patterns: disease-free equilibria/extinction ($\mathcal{R}_0 = 0.950$), stable endemics ($\mathcal{R}_0 = 1.570$), and cyclic dynamics ($\mathcal{R}_0 = 1.170$). SHAP ridge plots (Fig. 7) reveal that cycles emerge through precise demographic balance rather than transmission optimization, with the threshold effects for both μ_a and α proving essential for generating oscillatory patterns reproduced in dynamical simulations.

The biological implications are clear from the methodological convergence seen between Sobol sensitivity, SHAP importance, and model simulations. Sobol analysis confirms that transmission viability depends on critical demographic thresholds, SHAP analysis reveals how specific age-structured exposure patterns generate instability, and simulations demonstrate that only rare parameter combinations reproduce the cyclic dynamics observed in northeastern EEEV surveillance data. Stable dynamics emerge when parameters optimize transmission and maintain demographic stability: high mosquito biting rates (a) and adult avian mortality (μ_a), balanced exposure ($\alpha \geq 0.5$), and low mosquito mortality (μ_M) and adult virulence (ν_a). Unstable dynamics require the converse: demographic instability through low a

and μ_a, lower exposure to young avians ($0.2 \leq \alpha \leq 0.4$), and high μ_M and ν_a. Age-structured sensitivity patterns present further validation through simulation results, with young and adult avians showing differential responses to exposure and demographic parameters, reflected in the simulated oscillations where young avians peak at approximately 120 infections while adults peak at approximately 280 during cyclic outbreaks (Fig. 10).

This methodological integration advances EEEV disease modeling by demonstrating that traditional sensitivity analysis, machine learning approaches, and dynamical simulations address different but complementary biological questions. Our analytical approaches reveal critical threshold effects with direct intervention guidance. Within the critical exposure range ($0.2 \leq \alpha \leq 0.4$), sensitivity analysis predicts—and simulations confirm—that minor changes in mosquito feeding behavior could dramatically shift regional transmission patterns from stable to oscillatory dynamics [7, 13].

7. Conclusions and Future Work

Our findings establish that EEEV transmission dynamics emerge from two distinct biological mechanisms: vector-host interaction parameters that determine outbreak potential and host demographic parameters that govern whether transmission exhibits stable or oscillatory dynamics. The identification of adult avian mortality as the dominant driver of cyclic dynamics, despite minimal influence on transmission thresholds, provides new insights into EEEV transmission mechanisms. Model simulations confirm this by showing how demographic instability within critical parameter ranges generates the 2–3 year cycles documented in northeastern surveillance data [2, 13]. The methodological framework presented in this paper demonstrates that a comprehensive understanding of complex biological systems requires analytical approaches that capture both threshold effects and dynamical mechanisms.

This framework opens multiple avenues for advancing EEEV research across temporal, spatial, and environmental dimensions. Temporal extensions could incorporate seasonal variation in parameter importance analyses to reveal how demographic and transmission mechanisms shift throughout annual cycles, identifying optimal intervention timing. Spatial applications could extend SHAP analysis to geographically explicit models, explaining regional transmission differences and enabling the development of targeted management strategies. Environmental investigations could examine how climate change alters the identified parameter hierarchies, informing long-term surveillance planning, given the sensitivity of demographic stability parameters to environmental conditions. Beyond its application to EEEV, this multi-method approach could enhance understanding of other arboviral systems where demographic processes and transmission efficiency operate through distinct parameter sets [16, 29].

Our findings directly inform EEEV management by pinpointing adult host mortality and age-structured exposure as the fundamental drivers that determine cyclic versus stable transmission patterns. The dominance of these population-level

parameters in determining transmission dynamics suggests that habitat conservation and close monitoring of host populations may be more effective than traditional vector control approaches. By demonstrating that EEEV cycles emerge from intrinsic host population dynamics rather than external drivers, this research redirects management paradigms toward population-based interventions and early warning systems grounded in the critical biological parameters identified here [10, 24].

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ORCID

Aurod Ounsinegad - `https://orcid.org/0000-0001-7886-3015`
Christopher Mitchell - `https://orcid.org/0000-0001-8792-9370`
Nicholas Komar - `https://orcid.org/0000-0002-7855-1473`

References

[1] T. Akiba, S. Sano, T. Yanase, T. Ohta and M. Koyama, Optuna: A next-generation hyperparameter optimization framework, in *Proceedings of the 25th ACM SIGKDD international conference on knowledge discovery & data mining* (Association for Computing Machinery, 2019), pp. 2623–2631.

[2] P. M. Armstrong and T. G. Andreadis, Ecology and epidemiology of eastern equine encephalitis virus in the northeastern united states: An historical perspective, *Journal of Medical Entomology* **59**(1) (2022) 1–13.

[3] C. Banda and D. Samanta, Eastern equine encephalitis, in *StatPearls [Internet]*, ed. StatPearls Publishing (StatPearls Publishing, 2022).

[4] J. S. Blackmore and R. P. Dow, Differential feeding of culex tarsalis on nestling and adult birds, *Mosquito News* **18**(1) (1958) 15–17.

[5] L. Breiman, Random forests, *Machine Learning* **45**(1) (2001) 5–32.

[6] E. Buckner, A. Showman and C. R. Connelly, Black-tailed mosquito culiseta melanura (coquillett) (insecta: Diptera: Culicidae): Eeny-536/in950, 9/2012, *EDIS* **2012**(10) (2012).

[7] N. D. Burkett-Cadena, J. F. Day and T. R. Unnasch, Ecology of eastern equine encephalitis virus in the southeastern united states: Incriminating vector and host species responsible for virus amplification, persistence, and dispersal, *Journal of Medical Entomology* **59**(1) (2021) 41–48.

[8] Centers for Disease Control and Prevention, Eastern equine encephalitis virus `www.cdc.gov/easternequineencephalitis/index.html` (May, 2024).

[9] T. Chen and C. Guestrin, Xgboost: A scalable tree boosting system, in *Proceedings of the 22nd ACM SIGKDD International Conference on Knowledge Discovery and Data Mining* (Association for Computing Machinery, 2016), pp. 785–794.

[10] L. K. Estep, C. J. McClure, P. Vander Kelen, N. D. Burkett-Cadena, S. Sickerman, J. Hernandez, J. Jinright, B. Hunt, J. Lusk, V. Hoover, K. Armstrong, L. M. Stark, G. E. Hill and T. R. Unnasch, Risk of exposure to eastern equine encephalomyelitis virus increases with the density of northern cardinals, *PLOS One* **8**(2) (2013) 1–9.

[11] H. K. Hassan, E. W. Cupp, G. E. Hill, C. R. Katholi, K. Klingler and T. R. Unnasch, Avian host preference by vectors of eastern equine encephalomyelitis virus, *The American Journal of Tropical Medicine and Hygiene* **69**(6) (2003) 641–647.

[12] L. A. Heberlein-Larson, Y. Tan, L. M. Stark, A. C. Cannons, M. H. Shilts, T. R. Unnasch and D. S. R., Complex epidemiological dynamics of eastern equine encephalitis virus in florida, *The American Journal of Tropical Medicine and Hygiene* **100**(5) (2019) 1266–1274.

[13] V. Hill, R. T. Koch, S. M. Bialosuknia, K. Ngo, S. D. Zink, C. A. Koetzner, J. G. Maffei, A. P. Dupuis, P. B. Backenson, J. Oliver, A. B. Bransfield, M. J. Misencik, T. A. Petruff, J. J. Shepard, W. J. L., M. S. Gill, G. Baele, C. B. F. Vogels, G. Gallagher, P. Burns, A. Hentoff, S. Smole, C. Brown, M. Osborne, L. D. Kramer, P. M. Armstrong, A. T. Ciota and N. D. Grubaugh, Dynamics of eastern equine encephalitis virus during the 2019 outbreak in the northeast united states, *Current Biology* **33**(12) (2023) 2515–2527.

[14] T. K. Ho, Random decision forests, in *Proceedings of 3rd International Conference on Document Analysis and Recognition* **1**, (IEEE Computer Society, 1995), pp. 278–282.

[15] G. Ke, Q. Meng, T. Finley, T. Wang, W. Chen, W. Ma, Q. Ye and T. Liu, Lightgbm: A highly efficient gradient boosting decision tree, in *Proceedings of the 31st International Conference on Neural Information Processing Systems* (Curran Associates, Inc., 2017), pp. 3149–3157.

[16] F. Keesing, R. D. Holt and R. S. Ostfeld, Effects of species diversity on disease risk, *Ecology Letters* **9**(4) (2006) 485–498.

[17] B. Kessel, A study of the breeding biology of the european starling (sturnus vulgaris l.) in north america, *The American Midland Naturalist* **58**(2) (1957) 257–331.

[18] N. Komar, D. J. Dohm, M. J. Turell and A. Spielman, Eastern equine encephalitis virus in birds: relative competence of european starlings (sturnus vulgaris), *The American Journal of Tropical Medicine and Hygiene* **60**(3) (1999) 387–391.

[19] N. Komar, S. Langevin, S. Hinten, N. Nemeth, E. Edwards, D. Hettler, B. S. Davis, R. A. Bowen and M. L. Bunning, Experimental infection of north american birds with the new york 1999 strain of west nile virus, *Emerging Infectious Diseases* **9**(3) (2003) 311–322.

[20] S. M. Lundberg, G. Erion, H. Chen, A. DeGrave, J. M. Prutkin, B. Nair and S. Lee, From local explanations to global understanding with explainable ai for trees, *Nature Machine Intelligence* **2**(1) (2020) 56–67.

[21] S. M. Lundberg and S. Lee, A unified approach to interpreting model predictions (2017).

[22] R. G. McLean, W. J. Crans, D. F. Caccamise, J. McNelly, L. J. Kirk, C. J. Mitchell and C. H. Calisher, Experimental infection of wading birds with eastern equine encephalitis virus, *Journal of Wildlife Diseases* **31**(4) (1995) 502–508.

[23] E. Miller and A. Huppert, The effects of host diversity on vector-borne disease: The conditions under which diversity will amplify or dilute the disease risk, *PLOS ONE* **8**(11) (2013) 1–8.

[24] G. Molaei, M. C. Thomas, T. Muller, J. Medlock, J. J. Shepard, P. M. Armstrong and T. G. Andreadis, Dynamics of vector-host interactions in avian communities in four eastern equine encephalitis virus foci in the northeastern u.s., *PLOS Neglected Tropical Diseases* **10**(1) (2016) 1–23.

[25] F. Morneau, C. Lépine, R. Décarie, M. Villard and J. DesGranges, Reproduction of american robin (turdus migratorius) in a suburban environment, *Landscape and Urban Planning* **32**(1) (1995) 55–62.

[26] S. J. Mundis, S. Harrison, D. Pelley, S. Durand and S. J. Ryan, Spatiotemporal environmental drivers of eastern equine encephalitis virus in central florida: Towards a predictive model for a lethal disease, *Journal of Medical Entomology* **59**(5) (2022) 1805–1816.

[27] A. Petrucciani, G. Yu and M. Ventresca, Multi-season transmission model of eastern equine encephalitis, *PLOS ONE* **17** (08 2022) 1–23.

[28] S. Raschka, Model evaluation, model selection, and algorithm selection in machine learning (2018).

[29] S. L. Robertson and K. A. Caillouët, A host stage-structured model of enzootic west nile virus transmission to explore the effect of avian stage-dependent exposure to vectors, *Journal of Theoretical Biology* **399** (2016) 33–42.

[30] T. W. Scott and L. H. Lorenz, Reduction of culiseta melanura fitness by eastern equine encephalomyelitis virus, *The American Journal of Tropical Medicine and Hygiene* **59**(2) (1998) 341–346.

[31] I. M. Sobol, Global sensitivity indices for nonlinear mathematical models and their monte carlo estimates, *Mathematics and Computers in Simulation* **55**(1) (2001) 271–280, The Second IMACS Seminar on Monte Carlo Methods.

[32] The Royal Society for the Protection of Birds, Baby birds: Baby bird survival `www.rspb.org.uk/birds-and-wildlife/natures-home-magazine/birds-and-wildlife-articles/features/how-do-baby-birds-survive/`, (2022).

[33] T. F. Tsai, Arboviral infections in the united states, *Infectious Disease Clinics of North America* **5**(1) (1991) 73–102.

[34] R. S. Unnasch, T. Sprenger, C. R. Katholi, E. W. Cupp, G. E. Hill and T. R. Unnasch, A dynamic transmission model of eastern equine encephalitis virus, *Ecological Modelling* **192**(3) (2006) 425–440.

[35] A. Young and F. C., Eastern equine encephalitis (eee) `https://ceh.vetmed.ucdavis.edu/health-topics/eastern-equine-encephalitis-eee` (August, 2020).