Stratification along tropical forest succession enhances pollinator

diversity via functionally unique canopies

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ABSTRACT

Vertical stratification is among the main drivers of biodiversity in forests. Canopies are a key spatial component for primary consumers due to a higher concentration of resources, but methodological constraints impair a proper assessment of canopy communities. Therefore, the occupancy of key animal groups along the forest's vertical axis and the role of stratification in forest succession remains little understood. In this work, we investigated the stratification of pollinator abundance, (functional) diversity, and composition along a succession chronosequence in a lowland rainforest ecosystem. Moths, social diurnal bees, and nocturnal bees were more abundant and diverse in canopies, while orchid bees showed an inverse trend across parameters, suggesting taxa-specific responses. Late successional and old-growth forests were similarly diverse, suggesting that stratification establishes fast. Moreover, canopy-bearing successional and old-growth forests were more diverse than early successional plots across all groups. For moths and partially for bees, stratification enhanced diversity in old-growth and late-successional forests via species turnover between canopy and understory. Plot isolation, elevation, and canopy height had overall little effect on the variance between canopy and understory, potentially indicating a more prominent role of resource composition than environmental and local plot variables. Moreover, canopies harbored different pollinator compositions than understories. In particular, smaller, shorter-tongued, slower-flying, light-colored bees, and cryptic moths were associated with the canopy, suggesting that the relative role of interaction, dispersal and response traits in pollinator stratification is also group-dependent. Our study underlines the importance of stratification in leveraging diversity along forest succession, with a trait-driven, group-specific divergence of pollinator communities

50   between forest strata. These results highlight the importance of considering canopies

51   to improve sampling completeness and fully assess ecosystem biodiversity.

1. INTRODUCTION

Forests are remarkably diverse ecosystems that harbor most of the Earth's biota (Gaston, 2000). The diversity of biological niches offered by forests can ultimately be linked to their multidimensional complexity in the form of habitat heterogeneity (clearings, edges, and patch-matrix dynamics) (Forman & Godron, 1986; Watson, 1998) and a wide range of microhabitats and resources associated with trees and epiphytes (Nadkarni, 1994). Moreover, a prominent driver of forest heterogeneity and diversity is vertical stratification, with each forest stratum characterized by distinct niches formed by the interplay of microclimatic conditions and microhabitats (Shaw, 2004), as well as different sets of nutritional resources and nesting or roosting sites for animals (Jachowski et al., 2016; Nadkarni et al., 2013).

While the forest under- and midstories offer a variety of niches and resources associated with the soil (e.g., dung, deadwood, and carrion) and with low-laying vegetation (herbs and bushes in deciduous forests or pioneers in clearings) (Basset et al., 2003; Kudo et al., 2008), the concentration of resources in the upper layers is an important driver of animal distribution, particularly in tropical rainforests where a clear degree of vertical stratification is present (Basset et al., 2003). The canopy, a photosynthetic hotspot in the forest due to increased radiation, excels in resource yield for primary consumers (Ozanne et al., 2003). Large and productive trees have a much higher resource yield (e.g., flowers and fruits) than understory plants (Baker et al., 1983), while the accumulation of epiphytes, lianas, and tree crown results in a higher taxonomical and functional diversity of primary resources and turns canopies into optimal foraging areas for herbivores, frugivores and nectarivores (Ramalho, 2004; Schowalter & Chao, 2021).

Nonetheless, canopies are fragile sections of the landscape marked by low

resistance due to the targeted logging of slow-growing, large hardwood trees (Asner et

al., 2006), and low resilience, as such trees sometimes take centuries to establish,

grow, and provide their ecosystem functions (Xu et al., 2015). Abandoned logged

forests may take several decades to achieve a similar structural and biological

complexity as old-growth forests (Finegan, 1996). Tropical forests are particularly

critical in this regard. They foster an estimated three-quarters of Earth's biodiversity

(Stork, 2018) and account for most of the world's current Biodiversity Hotspots

(Myers et al., 2000). In these ecosystems, the loss of vertical complexity through

selective logging further hinders forest succession by filtering out canopy-specialist

pollinators and seed dispersers from the landscape (Silva et al., 2020; Whitworth et al.,

2016).

The sensitivity of fauna to canopy removal and the consequent effects on forest

recovery is, however, context- and taxa-dependent. Enhanced forest connectivity and

concentrated losses of canopy cover may still allow for the occurrence of canopy

specialist and their movement across the landscape (Cadavid-Florez et al., 2020).

Meanwhile, highly fragmented and logged forests will have a perceptive impact on

canopy specialists, even removing canopy-restricted animals with low mobility (e.g.,

canopy primates, Shaw, 2004). Moreover, a higher exposure associated with canopy

cover loss, such as increased heat and light incidence also selectively removes smaller,

desiccation-prone animals from the landscape (Basset et al., 2003), underlining the

role of dispersal and response traits in defining canopy dependence and response to

disturbance.

The effect of canopy loss and the role of traits in defining canopy usage is,

however, still relatively little understood for insects, despite half of the world's

arthropods estimated to occur in canopies (Ozanne et al., 2003; Schowalter & Chao, 2021) and up to a quarter of invertebrates expected to be canopy specialists (Basset et al., 2003). Most of our current knowledge of invertebrate diversity is still based on understory sampling (Nakamura et al., 2017). While there is evidence that several insect communities, including pollinators, are more abundant and diverse in canopies (Brehm, 2007; Roubik, 1992a; Stangler et al., 2016; Watanabe et al., 2017), evidence is sparse and often conflicting. Pollinators are expected to be particularly sensitive to the loss of old-growth canopies. Mass-flowering canopy trees serve as abundant and year-round resources for several groups of canopy-restricted bees and beetles, for instance (Kirmse & Chaboo, 2020; Ramalho, 2004; Roubik, 1992a), underlining the critical role of old forests with dense canopy cover in maintaining pollinator communities. Pollinators, critical for the reproduction of more than 95 % of tropical plants (Ollerton et al., 2011) and consequently essential for forest regeneration (López-Cubillos et al., 2023), are thus an exceptional model group to assess the importance of the canopy for ecosystem processes and the effects of canopy loss on biodiversity, yet the ecological drivers of pollinator stratification are largely understudied (Allen & Davies, 2023; Ashton et al., 2016; Nakamura et al., 2017; Roubik, 1993; Smith et al., 2012).

Most of these gaps are due to the logistical limitations of sampling canopies (Ulyshen et al., 2010). While several methods exist, ranging from tree climbing to cranes and scaffoldings (Schowalter & Chao, 2021), they are often cumbersome and expensive, thus only occasionally used and consequently suffer from sample size and replication issues (Barker & Pinard, 2001; Nakamura et al., 2017). Shooting methods used to rig pulley ropes on trees for passive trapping became progressively more popular since early canopy research (Barker & Pinard, 2001), particularly slingshots

and hand catapults (Allen & Davies, 2023; Yusah et al., 2012). Bows and crossbows, however, entail the most precision and potency and allow for placing traps in higher strata (Ellwood & Foster, 2001), while still being relatively simple and cheap. Although the use of bows is frequently mentioned (Ashton et al., 2016; R. L. Kitching et al., 2001; Morris et al., 2015; Sobek et al., 2009), the method has never been described in detail, hindering its reproducibility, and has also not yet been used in large biodiversity assessments.

We employed and provided technical details on the archery-assisted pulley system method in a comprehensive sampling of diurnal and nocturnal pollinators in tropical canopies to assess their abundance, diversity, composition, and functional uniqueness in comparison to the understory, focusing on bees and moths. We aimed to investigate how stratification contributes to and enhances diversity along forest succession in a disturbed landscape. Considering the accumulation of floral resources in the canopy, we expect that (i) pollinator (functional) diversity and abundance will be higher in the canopy than in the understory, particularly as the degree of stratification increases in older forests. Due to higher exposure in the canopy, as well as differing assemblages of floral resources (e.g., epiphytes and crown trees), we expect (ii) differing community compositions of pollinator species between strata. The resulting turnover will lead to (iii) a higher overall diversity in old-growth forests in relation to the early stages of forest recovery. Turnover, in turn, should be driven by (iv) dispersal traits, e.g., larger (less desiccation-prone) insects with a better flying performance (higher wind resistance) insects associated with the canopy; by their (v) response traits, with light-colored or aposematic species associated with the canopy to resist overheating and avoid predation; and by their (vi) interaction traits, with the packing of more variable floral resources allowing for the occurrence of longer proboscises.

2. METHODS

2.1. Study area

The study was conducted between 2022 and 2023 in the Río Canandé Reserve

(0°31'33.4"N, 79°12'46.0"W) and the Tesoro Escondido Reserve (0°32'30.9"N,

79°08'41.9"W), Esmeraldas province, northwestern Ecuador. The site is a lowland

forest ecosystem within the Chocó-Darien ecoregion and a biodiversity hotspot on the

western foothills of the Andes (Fagua & Ramsey, 2019), with a mean annual

temperature of 24°C and annual rainfall of ca. 2100 mm, (Af climate, Köppen scale).

The study was carried out in an ~7800 ha area within the REASSEMBLY research

unit, a chronosequence of 62 50x50 m plots ranging from active disturbance

agricultural sites, to regenerating secondary forests and old-growth forests (Escobar et

al., 2024). From these plots, we sampled the canopy at 27 plots which had a defined

vertical structure, at least one tree 15 m in height or taller, and significant canopy

closure (section *2.2* below). All these plots were located in old-growth (OG) primary

forests (N = 16) and late successional forests (LS), formerly pastures, or cacao

plantations (N = 11, 9-37 years of regeneration). The remaining 35 plots were sampled

at the understory only. These plots were either active plots (AC), i.e. cacao plantations

or pastures (N = 12), or early successional (ES) forests without clear vertical

stratification (N = 23, 1-8 years of regeneration). All plots were separated by at least

350 m (maximum distance 12.8 km) and were within comparable elevations (224-

615m, average 388 ± 119,3).

Four insect sampling campaigns were carried out in total with each plot sampled

twice: two campaigns in 2022, and two campaigns in 2023 (half of the plots were

sampled between March and May and the respective other half between October and

December). Three LS plots were removed from the 2023 campaigns as canopy ropes

could not be installed (the only usable tree fell or there were no more suitable

branches). The permit to conduct research on the site and to sample insects was

granted by the institutions in charge (Ministry of Environment of Ecuador, Contrato

Marco MAE-DNB-CM-2021-0187).

2.2. Installation of pulley system and canopy traps

In each LS and OG plot, a pulley system was installed in a single tree in the

canopy with the assistance of bow-and-arrow shooting (Fig. 1). For the rope

installation we selected an accessible branch without much surrounding clutter or

vegetation from the tallest or among the tallest trees within the plot (mean height 25.8

± 5.3 m, median 25.2 m, range 15 – 37.0 m) to allow for a clean shot and avoid arrow

tangling.

For installing the rope, we used a 29-inches glass fiber and aluminum

Strongbow GECKO compound bow (BogenSportWelt, Germany) and 70 cm glass

fiber arrows with rounded points (Toparchery, Germany). A 0.32 mm 4-strands braided

polyethylene fishing line (Mounchain, Japan) was attached to the arrows and set onto a

TP500 double-drag brake system fishing reel (YJIU, China, see Fig. 1 A for technical

adjustments made to the reel). After selecting a fitting branch and successfully

shooting the fishing line over it (Fig. 1, B – C), the fishing line was replaced by a ca. 5

mm polypropylene braided plastic rope, the final pulley system to which traps were

attached (Fig. 1, D). A troubleshooting guide for the canopy setting can be found in the

Appendix S1: Box S1. After the rope installment, traps (section 2.3 below) were

attached to it using carabiners on the trap roofs and pulled up into the canopy. Traps

were separated by approximately 30 cm from one another and lifted to the maximum

height possible, i.e. ideally directly below the branch where the rope was installed.

2.3. Insect trapping

We employed several types of traps to capture a variety of pollinator groups. In

2022, we sampled nocturnal insects (moths and bees), as well as diurnal bees. On each

plot, we installed one light trap and one vane trap at each stratum level (one set of

traps at AC and ES plots, two sets in LS and OG plots) within the plot area. Traps were

left in the field for 24 hours. At the understories, traps were always ca. 2 m apart and

ca. 1.5 m above ground. At the canopy, vane traps were always placed directly below

light traps, separated by ca. 1 m. The traps employed were as follows:

(i) Light vane traps: target nocturnal insects; they were modified funnel traps

consisting of a roof, three white vanes and a funnel of white polypropylene (Singh et

al., 2022), equipped with a LepiLED mini Switch light (Brehm, 2017) and connected

to a power bank battery. We used the mixed light setting (UV and visible light

spectrums). The funnel traps were attached to a collection chamber which was

connected to a 5 L bucket (wrapped in aluminum foil) with chloroform used to fast-

kill trapped insects. The traps were timed to start light emission at 1800h and stop at

0600h. All trapped insects were collected the following day.

(ii) Color vane traps: target diurnal bees; the traps were in principle adaptations

of a common and commercially-available used vane trap model (Prendergast et al.,

2020) but due to the rainy environment, they had a roof and were equipped with both

a yellow and a blue plastic vane attached transversally, forming an alternating color

pattern designed to visually attract bees. Insects were collected in a connected 1 L

plastic bucket wrapped in aluminum foil.

In 2023, color vane traps were set up once more in all plots, now with the

addition of sugar baits (small quantities of honey smeared on the interior walls of the

funnels) to increase capture rates. Additionally, we set up (iii) fragrance traps targeting

fragrance-collecting male orchid bees (Apidae: Euglossini) (Eltz et al., 1999), a

dominant pollinator group in the Neotropics (Michener, 2007). These traps were

adaptations of those used by (Ferreira et al., 2013), using two lateral entrance holes

(ca. 3 cm) equipped with plastic funnels and a piece of cotton imbibed in household

insecticide (Raid, USA) as a fast-killing agent. On each plot, we placed four fragrance

traps at the canopy and understory, each containing one of the following compounds:

1,8-cineole, eugenol, methyl-salicylate, and skatol (Carl Roth GmbH, Germany). This

combination was chosen to attract a diverse range of orchid bee species (Cancino &

Damon, 2007). As in the previous year, each set of traps (one vane trap, four fragrance

traps) was simultaneously installed at the canopy and understory. In total, 348 traps

(54 light vane traps, 102 color vane traps, and 192 fragrance traps) were set up, half at

each strata level of the 27 LS and OG plots, and 245 (35 light vane traps, 70 color

vane traps, and 140 fragrance traps) in the understory of AC and ES plots.

2.4. Focal insect groups and processing

After collection, insect samples were sorted, and target groups were selected

for light traps: hawkmoths (Sphingidae) and tiger moths (Erebidae: Arctiinae). The

first are large, fast-flying moths known for their importance as pollinators (Renner &

Feil, 1993), while the latter are a large and diverse clade of conspicuous and often

aposematic moths, which are likely underestimated as pollinators (Van Zandt et al.,

2020). After sorting, insects were identified to the lowest possible level. Moths were

sorted to morphospecies (referred to as species hereafter) and identified using the

collections in the Phyletisches Museum Jena, Germany and the research collection of

GB from Costa Rica, Ecuador, and Perú, as well as photographs taken by GB in the

Natural History Museum (London). Identification supported by COI barcoding of ca.

90% of all moth species (GB unpublished catalogue). Nearly all moth species could be

assigned a genus, and most were identified to species level. Bees were identified using

specialized literature (Bonilla-Gómez & Nates-Parra, 1992; Michener, 2007; Roubik,

1992b; L. M. Santos & Melo, 2015). All bees could be assigned a genus, and nearly all

(> 95%) a (morpho)species.

From each insect species, we collected four morphological traits: body size

(BS), proboscis length (PL), effective proboscis length (EL), and wing aspect ratio

(AR). BS is an indicator of dispersal ability and resistance to desiccation in insects

(Bailey et al., 2018) and was measured as the length between the tip of the head and

the tip of the last abdominal segment of moths. The inter-tegular distance was used as

a proxy for size in bees (Cane, 1987). PL is related to morphological coupling with

flowering plants and is an interaction trait associated with feeding specialization

(Johnson et al., 2017). It was measured as the combined extended length of the

prementum and glossa for bees, and the entire extended length of the galeae. EL was

calculated as the ratio between PL and BS, which accounts for the allometric

relationship between PL and BS. AR is a measure of flight performance, with a higher

AR indicating a better gliding capability (faster flyers) (Bhat et al., 2019). It was

measured as the ratio between the square root of the forewing length and the wing area

for moths; and for bees, the formula was simplified to the ratio between wing length

and the wing's mean chord (widest wing width) due to a relatively conserved wing

shape across groups (Bhat et al., 2019). Traits were measured manually with a caliper

for larger bees (e.g., Euglossini) and with the aid of microscopy and linear

morphometric analysis via the ImageJ software (Collins, 2007) for smaller (> 5 mm of

body length) bees. For the latter, the proboscises were dissected and mounted on slides

for further image analysis. All moth traits were collected via a standardized

photography (Brehm, submitted) and processed via image analysis, using a modified

version of the mothseg algorithm (Jaimes Nino et al., 2019). We supplemented this

data with individual measurements of photographs with scale bars for proboscis length

(Brehm, submitted). We used measurement data from all species and used mean values

when more than one individual was present (up to ten).

We additionally measured coloration, which is linked to thermoregulation and

predation avoidance (Badejo et al., 2020). Coloration is thus an important response

trait that might affect the vertical distribution of species. For moths, we measured two

variables, also via standardized moth photographs and the modified version of the

mothseg algorithm: (1) Lightness was measured as the median pixel value of four

main color channels (red, green, blue, and UV) of the dorsal side of fully spread

individuals, which represents overall brightness (e.g., high values for white species

and low values for dark species), (2) Contrast, measured as the interquartile distance

$(25^{th}-75^{th}$ percentiles) of the distribution of pixel values, representing coloration

variability with high values often associated with conspicuousness to predators and

aposematism. Trait measurements will be described in detail separately (GB, DB,

Yenny Correa Carmona, Dimitri Korsch, unpublished data). Finally, external visual

appearance was established as a set of categorical variables. Following Fiedler and

Brehm (2021), moths were classified as aposematic, hymenopteran mimic, beetle

mimic, and cryptic (none of the aforementioned). Bees, which did not show strong

mimicry patterns, were visually assessed and separated into five categories based on

the predominant color of their bodies, with the dorsal side of the head, thorax, and

abdomen used as reference: "dull-dark" (matte bees ranging from dark brown to

black), "dull-clear" (matte bees of orange or yellow color), "iridescent-blue",

"iridescent-green", "iridescent-red" and "aposematic" (the latter four strictly

Euglossines).

2.4. Data analysis

All analyses were performed in R Studio 4.1.0 (R Core Team, 2021). To assess

the variation in the occurrence of pollinators between canopies and understories, we

tested for differences in four community parameters as response variables: (i)

abundance (A), or the total sum of individuals in each sample; (ii) exponential

Shannon diversity (effective diversity, $q = 1$), which corresponds to the effective

number of common species (Magurran, 2013). To account for the effect of abundance

and allow for a better cross-sample comparison, the asymptotic diversity of each

sample was estimated via abundance-based rarefaction (Chao et al., 2014); (iii)

functional richness (FRic) and (iv) functional dispersal (FDis), both measured via a

distance-based and multi-trait framework to estimate functional diversity through the

FD package (Laliberté & Legendre, 2010; Villéger et al., 2008). FRic estimates

functional richness as the volume of the minimum convex hull encompassing all

species in a PCoA reduction (Villéger et al., 2008), while FDis measures the distance

of all species to a common centroid, thus estimating the average dispersal in functional

profiles in comparison to a common mean (Laliberté & Legendre, 2010). All numeric

traits were used to calculate FDis and FRic. Because these two indices could not be

properly estimated for communities with less than three species, species-poor groups

in the samples (i.e., *Megalopta* bees) did not undergo functional analysis. These four

parameters were calculated for the canopy and understory of all LS and OG plots, and

each main pollinator group was analyzed separately. Paired t-tests were used to assess

whether the canopies and understories differed significantly for each of these

parameters. The normality of the data was checked in all instances.

To understand whether diversity in stratified LS and OG is enhanced via

species turnover between strata, we compared the rarefied diversity of canopy

samples, understory samples, and pooled canopy and understory samples of both

forest legacies through generalized linear models (GLMs) fit with a gaussian error

family and log link function, accounting for residual overdispersion. The same

analysis was used along the entire chronosequence gradient, i.e., rarefied diversity of

pollinators was compared among AC, ES, LS (pooled), and OG (pooled) to assess

whether canopy-bearing plots were more diverse in pollinators than early successional

and active plots. Main pollinator groups were analyzed separately so group-specific

trends could be considered. To ensure the lack of effect of plot proximity on diversity,

the lack of spatial autocorrelation of residuals of the diversity models for each group

was checked via Moran's I index calculated with the *ape* package (Paradis & Schliep,

2019) (Appendix S1: Table S1).

To find possible drivers of pollinator community stratification in the canopy-

bearing plots, we assessed how absolute differences in abundance, diversity, FDis, and

FRic between canopy and understory were affected by the following environmental

and landscape variables: (i) disturbance legacy (LS or OG), which may affect the

forest vertical structure and plant community found in the plot; (ii) canopy height,

using the height of canopy traps as a proxy; (iii) elevation a.s.l., which may affect plot

temperature and thus the microclimatic niches in both strata (Rapp & Silman, 2012);

and (iv) the proportion of forest within a 500 m buffer around the plot, a proxy for

landscape isolation and fragmentation, which affects pollinator occurrence and

dispersal (Didham et al., 1996). The ESRI World Imagery basemap was used as source

for recent landcover and sourced from Escobar et al. (2024). We examined the effect

estimates of the four normalized community parameters by GLMs adjusted with a

quasibinomial error distribution to account for the ratio values of the community

parameters and residual overdispersion. In order to assess legacy-specific patterns, the

data from the three disturbance legacies (old-growth, former cacao plantations, and

former pastures) were separated and analyzed independently as described above.

Finally, we examined whether species composition differed significantly between

canopies and understories though Non-metric Multidimensional Scaling (NMDS)

using the Bray-Curtis distance. A PERMANOVA (1000 permutations) followed to

assess whether the canopy and understory groups differed significantly in composition.

To understand how species composition was influenced by functional traits (PL, BS,

EL, and AR, plus lightness and contrast for moths), we employed a Canonical

Correspondence Analysis (CCA) using each trait's community-weighted mean as

constraining. The significance variance explained by the main CCA axes, as well as

the six constraining variables, were assessed via Monte Carlo permutation tests (1000

permutations). The main pollinator groups were analyzed separately. The occurrence

(presence of absence) of visual categories was analyzed independently, with estimates

of the effect of stratification on color occurrence obtained via GLMs set with a

binomial error family. Similarly to the previous section, the three disturbance legacies

were separated and analyzed independently (old growth, former cacao plantations, and

former pastures).

3. RESULTS

3.1. Stratification and its effect on pollinator diversity

A total of 13997 target insects were collected in the 27 LS and OG plots,

encompassing 7614 moths from 302 species and 6383 bees from 89 species. Of these,

5727 (41%) were captured in the understory, and 8270 (59%) in the canopy. Moths

were represented by the two focal groups Arctiinae and Sphingidae, with the former

accounting for the vast majority of individuals (98.1%). Therefore, all results

concerning moths hereafter refer to both groups analyzed together. The samples from

diurnal traps were dominated by stingless bees (Apidae: Meliponini) (51% of bee

individuals) collected exclusively from vane traps, orchid bees (Apidae: Euglossini)

(43%) collected from fragrance traps, and nocturnal *Megalopta* (Halictidae:

Augochlorini) bees (6%) from light traps. Other bee groups (e.g., Colletidae, diurnal

Halictidae) accounted for 0.27% of the total bee captures and were thus not included

in further analyses.

Abundance and diversity were significantly higher in the canopy relative to the

understory for moths (abundance: $t = 4.48$, $p < 0.001$; diversity: $t = 7.16$, $p < 0.001$),

stingless bees ($t = 2.95$, $p < 0.01$; $t = 3.19$, $p < 0.01$) and *Megalopta* bees ($t = 3.01$, $p <$

0.01; $t = 2.97$, $p < 0.01$). Moths showed the strongest preference towards the canopy

(Fig. 2). Orchid bees, however, showed an inverse pattern, with a stark decrease in

abundance ($t = -5.23$, $p < 0.001$) and diversity ($t = -3.36$, $p < 0.01$) in the canopy

relative to the understory. Moreover, moths showed a significantly higher functional

richness ($t = 3.85$, $p < 0.001$) and dispersion ($t = 4.13$, $p < 0.001$) in the canopy, while

orchid bees again showed a strong and inverse pattern (FRic, $t = -5.60$, $p < 0.001$;

FDIs, $t = -2.29$, $p < 0.05$) (Fig. 2). There was no significant difference between the

strata for stingless bees ($p > 0.05$ for both variables).

Additionally, 4997 bees and 4263 moths were collected in the remaining AC

and ES plots, summing up to 23257 target insects captured in total. Along the

succession gradient, the canopy-bearing LS and OG forests always had similar species

diversity vales, and both forest legacies were significantly more diverse than ES and

AC for Meliponini bees and more diverse than ES for moths (Fig. 3, Tab. 1). Moths

were also highly diverse in AC plots and did not differ between late and old-growth

forest plots (Fig. 3 A-B, Tab.1). Overall, nocturnal *Megalopta* bees were also more

diverse in older forests, particularly OGs (Fig. 3 D). However, Euglossines showed no

significant pattern along the successional gradient, although they were on average

most diverse in LS and OG (Fig. 3 C).

In terms of the turnover between canopy and understory, pooled LS and OG

samples were significantly more diverse than individual canopy and/or understory

samples for moths, particularly in late successional forests (Fig. 3 A, Tab. 1),

suggesting high turnover and a complementary interaction between strata and increase

in total diversity. Meliponini showed no turnover between canopy and understory in

both forest types (Fig. 3, B). The diversity of Euglossini in forest plots was similar to

that of understory samples, especially in OG plots (Fig. 3 C). *Megalopta* showed an

inverse trend (Fig. 3, D), although their low diversities likely impaired proper

estimation.

3.2. The role of forest legacy, canopy height, and environmental features

Plot environmental variables and disturbance legacy only explained little

variation between canopy and understory in terms of abundance, diversity, and

functional diversity for most pollinator groups. Neither canopy height, legacy,

elevation nor the proportion of forest around the plots had a significant effect on the

variation of Meliponines and moths ($p > 0.05$ for all variables) (Fig. 4). The proportion

of forested area around sampling plots, however, had a significant and negative effect

on the variation in diversity ($\chi^2 = 9.47$, $p = 0.002$) and the functional dispersal ($\chi^2 =$

3.88, $p = 0.04$) of Euglossines (Fig. 4), i.e., canopy and understory communities

became more similar taxonomically and functionally with increasing proportions of

forest around plots. *Megalopta* bees showed a sharp decrease in the variation in

abundance ($\chi^2 = 4.83$, $p = 0.02$) and diversity ($\chi^2 = 6.06$, $p = 0.01$) between canopy and

understory in LS plots, i.e., strata were more similar in successional forests than in

old-growth forests. Abundance variation was further positively affected by plot

elevation ($\chi^2 = 5.43$, $p = 0.02$); and diversity variation was negatively affected by

canopy height ($\chi^2 = 6.63$, $p = 0.01$) (Fig. 4), i.e., bees were more evenly distributed

between strata at lower elevations and in plots with shorter canopies.

3. 5. Species composition and the influence of traits

438   Species composition differed significantly between canopy and understory for

both bees (F = 8.63, $R^2 = 0.15$, p < 0.001) and moths (F = 4.29, $R^2 = 0.008$, p <

0.001), with a most noticeable divergence within bees following a phylogenetic signal

due to differences in strata preferences of Meliponini and Euglossini. The entire bee

community varied significantly in their functional profiles, with larger, longer-

tongued, and faster-flying bees strongly associated with the understory (CCA1: Eigenv

= 0.54, F = 6.63, p < 0.001; CCA2: Eigenv = 0.28, F = 3.45, $p < 0.01$). All traits

significantly affected the distribution of pooled bee species (Fig. 5, A, Tab. 2).

Proboscis length and wing aspect ratio, which are likely associated with the long-

tongued and fast-flying orchid bees, explained the most variance, although with an

overall low variance explained (Tab. 2).

449   Meliponini, when isolated, showed a stronger association with the understory

for all traits (CCA1: Eigenv = 0.72, F = 10.60, $p < 0.001$; CCA2: Eigenv = 0.69, F =

10.20, $p < 0.001$) (Fig. 5, B; Tab. 2). Euglossini presented a less clear yet significant

signal of functional trait distribution between canopy and understory (CCA1: Eigenv =

0.33, F = 6.39, $p < 0.001$; CCA2: Eigenv = 0.19, F = 3.71, $p < 0.001$), yet bees in the

canopy tended to be larger, slower flyers with shorter effective proboscis lengths (Fig.

5, C; Tab. 2). Traits significantly influenced moth composition (CCA1: Eigenv = 0.26,

F = 3.93, $p < 0.001$; CCA2: Eigenv = 0.21, F = 3.20, $p < 0.001$), yet to a smaller extent

and with the least clear distinction between canopy and understory. Most notably,

moths in canopy samples tended to show a lighter coloration, while contrast did not

significantly vary across samples (Fig. 5, D; Tab. 2).

The different color categories of bees and moths differed significantly between

canopy and understory. Canopy-associated bees fell into the dull bright category,

followed by the dull dark bees. Across all forest legacies, both groups were

represented mainly by small social stingless bees (Fig. 6, A). All other colors tied to

euglossines were more associated with the understory, i.e., aposematism on old-growth

plots, and iridescent colors, particularly red, in all legacies (Fig. 6, A). Cryptic moths

were much more common in the canopy across all three legacies, while aposematic

and beetle mimics were associated with the understory (Fig. 6, B). These patterns were

stronger in old-growth forests and relatively weak in former cacao plantations.

Hymenopteran mimics were strongly and significantly associated with the canopy in

old-growth plots, but not in successional forests (Fig. 6, B).

4. DISCUSSION

4.1 Patterns of pollinator stratification

Our study provides new insight into the vertical stratification of insect

pollinators along forest succession. We found an overall increase in pollinator

abundance and diversity in canopies, as well as different species composition between

strata, which led to a diversity-enhancing effect in successional and primary forests for

moths and, to a lesser extent for orchid and stingless bees. Our expectations (i-iii)

were, however, only partially confirmed, as canopy preferences were group-dependent

and driven by taxa-specific functional traits.

Positive associations with canopies were observed for moths, meliponines, and

Megalopta bees, all of which are driven, to some extent, by the vertical stratification

of resources. Arctiinae moths increase in abundance and diversity in canopies, likely

influenced by the availability of resources for liana- and vine-specialized caterpillars

(Brehm, 2007), suggesting a bottom-up regulation of this group by vegetative

resources. In addition, a concentration of floral resources in the canopy has also been

suggested as a potential driver of stratification in tropical Arctiinae and Sphingidae

(Schulze et al., 2001). Such associations have not been observed for other groups of

moths (e.g., Geometridae), indicating that group-specific canopy preferences operate

even within a given feeding guild (Brehm & Axmacher, 2006). In addition, in

temperate forests, the inverse pattern was observed for several groups including tiger

moths (Böttger et al., 2025), suggesting that the stratifying effect of resource

availability is more prominent in the tropics and likely related to components not

present in temperate forests (e.g., lianas and epiphytes). Meanwhile, a high abundance

and diversity of Meliponines is likely related to their specialization on mass-flowering

canopy trees (Calaça et al., 2022; Ramalho, 2004), which provide abundant food

resources for their partly large colonies. *Megalopta*, in turn, is a highly understudied

group but scarce evidence shows a dietary preference towards a few veteran bat-

pollinated canopy trees (Smith et al., 2012), which is further supported here by their

spike in occurrence in old-growth canopies.

The observed canopy preferences suggest a key role of resource availability in

the vertical distribution of tropical insects, as often reported in temperate forests

(Sobek et al., 2009; Ulyshen et al., 2010). The stratifying effect of resources has also

been observed in a variety of functional groups, including seed dispersers and non-

obligate floral visitors, ranging from non-flying mammals (Rader & Krockenberger,

2006), birds and frugivorous bats (Chmel et al., 2016; Thiel et al., 2021) to butterflies

(Fermon et al., 2005), as well as ground predators and flying insectivores (Basham et

al., 2023; Kalko & Handley, 2001), indicating that resource stratification permeates

different ecosystem processes. In our study site, several slow-growth, mass-flowering

trees and palms with accessible flowers were likely driving the increased abundance of

pollinators, particularly stingless bees, in canopies, including *Miconia*

(Melastomataceae), *Wettinia* (Arecaceae), *Banara* (Salicacaea), and resin-rewarding

Clusia (Clusiaceae) (Lozano et al., 2024). On top, the abundant night-blooming *Inga*

(Fabaceae) and Meliaceae (e.g., *Guarea*, *Tapirira*) (Lozano et al., 2024) might further

explain the canopy preference of *Megalopta* bees in OG plots and why stratification in

this group was lower in successional forests.

Euglossines, in contrast, showed a notable aversion towards the canopy, as

frequently noted (Feitosa Ribeiro et al., 2024; Ramos et al., 2022; Vega-Hidalgo et al.,

2020), but underlying causes are unknown. A prominent hypothesis is the dissipation

of floral volatiles (synthetic and natural) by sunlight and wind in the canopy, which

hinders detection by Euglossines and would explain why these bees avoid the canopy

(Ramos et al., 2022), as well as the low occurrence of Euglossini-pollinated orchids in

canopies (Hernández-Pérez et al., 2018). A further and more detailed examination of

resource (functional) diversity stratification might shed light on the role of various

resources (food, nests, scent) on the vertical distribution of orchid bees and other

pollinators in general (e.g., Silva et al., 2021). Beyond the role of resources, the

composition shift from larger, longer-tongued Euglossines with good flying

performance towards small and light-colored eusocial Meliponines in the canopy may

shed some light onto the role of traits on the stratification of pollinators and the

influence of abiotic factors.

4.2 The influence of traits

Our results also only partially confirmed our expectations (iv-vi) of large, light-

colored, or aposematic pollinators with longer proboscises and better flying

performance in the canopy. Size had the most notable variation between strata, and

against our expectations, the canopy was dominated by small Meliponines with

relatively broad wings and smaller proboscis. Most of this variation reflects the

canopy aversion of the large, dispersal-capable, and long-tongued Euglossines, but this

pattern repeats also within stingless bees. Canopy specialists mainly comprised small

(ca. 2 mm), gregarious, and wood-nesting *Trigonisca* bees (Michener, 2007),

especially the bright-colored and highly abundant *T. atomaria*, while larger genera,

such as *Partamona,* were more common in the understory (as also seen by Roubik,

1993).

This outcome is the opposite of the "shade-loving" bees associated with the

understory described by Bawa (1990) and Kato (1996) and suggests that the interplay

between dispersal and response traits and abiotic factors is not always straightforward

and may be related to other variables to the usage of the landscape by a given animal,

such as finding food and nesting resources. For instance, nesting and foraging close to

the canopy may enable small Meliponines with very short flight ranges (e.g.,

Trigonisca, less than 100 m from colonies) to account for both smaller size and lower

mobility (Grüter & Hayes, 2022) and may consequently represent a tradeoff between

exposure, such as stronger winds, low humidity, and heat (which may be partially

buffered by brighter colors), and foraging optimization. Establishing a nest may also

be easier in upper strata, as the abundance of natural substrate for cavity bees (e.g.,

natural crevices in wood) (Sobek et al., 2009) provides an advantage to most

Meliponines and many solitary bees (Batra, 1984; Wille & Michener, 1973).

Therefore, while functional traits may explain bee stratification to some extent, much

of the variation seems to be tied to group- and genera-specific preferences. Within

Euglossini, for instance, a slight shade-loving bee pattern (more robust bees in the

exposed canopy) emerged mainly due to large *Eulaema* bees that often ventured

beyond the understory, as also seen by Roubik (1993). The importance of accounting

for genera or species when assessing canopy preference is thus clear, as suggested for

organisms ranging from bees (Roubik, 1993) to bats (Kalko & Handley, 2001).

Life-history traits may provide additional insight. The nomadic and long-range

movement of solitary Euglossini males, as opposed to the localized colony life of

Meliponini, makes this group vulnerable to habitat fragmentation if patch sizes are too

small (Powell & Powell, 1987). In our study system, this negative effect of

fragmentation manifested in more homogeneous strata, with less (functionally) diverse

canopies in more isolated plots. This outcome suggests an interaction between canopy

usage and fragmentation for isolation-sensitive groups, as assumed by Ramos et al.

(2022). Nocturnal bees, in turn, naturally intolerant to heat and desiccation (Gonzalez

et al., 2023), were less stratified when canopies were higher (thus potentially drier)

and a higher stratification in more elevated plots, where colder temperatures likely

buffered the effect of exposure on these bees.

As for moths, specific adult functional traits showed a limited influence on the

community composition, further suggesting a stronger role of larval stages in

stratification. An exception was the occurrence of visual deception. Light-colored,

cryptic, and hymenopteran mimic moths were associated with the canopy, indicating

that canopy-specialist adult moths require some form of protection from predators to

compensate for their exposure (Brehm, 2007) when, e.g., searching for ovipositing

sites. The occurrence of this specific form of mimicry in the canopy might be related

to cavity-nesting wasps and bees, common in canopies (Sobek et al., 2009), while the

frequent occurrence of coleopteran mimicry and aposematism in the understory further

reinforces genus-specific preferences for strata (e.g., Coleoptera mimicry is mainly

represented by genera *Correbia*, *Correbidia*, and *Lycomorphodes*, Rabl et al., 2020)

possibly driven by the vertical stratification of different predator guilds (Basham et al.,

2023).

Therefore, the varying importance of dispersal, response, and protective traits

across taxa, allied to tribe- or genus-specific patterns, suggest accounting for both

morphological and life-history traits (e.g., roosting and nesting requirements, feeding

specialization of adults and larval stages, sociality, period of activity) for a more

comprehensive understanding of animal stratification and recovery (see Montoya-

Pfeiffer et al., 2020).

4.3 The diversity-leveraging effect of stratification

Despite the different stratification patterns and signal of functional traits across

groups, the overall increased pollinator diversity in canopy-bearing underlines the

relevance of both primeval and successional forests in maintaining animal diversity in

disturbed landscapes, as attested for a range of ecosystems (Taki et al., 2013; Ulyshen

et al., 2024). Pollinator diversity was consistently similar between LS and OG forests

and significantly higher than early successional plots, except for Euglossini bees. Male

Euglossines require wide dispersal and foraging ranges to search for particular and

specific volatile compounds used in courtship that are sparsely distributed across the

forest (Eltz et al., 1999). On top of being an essential part of their reproductive cycle, a

good dispersal capability grants them a notably high resistance to disturbance (Aguiar

& Gaglianone, 2012; Botsch et al., 2017), consequently leading to a low response to

the successional gradients (Rasmussen, 2009). High resistance and resilience to the

loss of habits have been linked to dispersal ability, more vagile being often less

sensitive (Meyer et al., 2009). Low dependence on canopy-specific resources and a

solitary life likely also played a role in their prolific nature, unlike other social bee

groups that are more dependent on resources closer to colonies.

612   For instance, social Meliponines and facultatively social *Megalopta* (Smith,

2021) are more tied to forested plots and are much more sensitive to disturbance.

Consequently, these groups must be more strongly tied to the resources, in terms of

both quantity and composition, found in LS and OG forests. Although the plant

composition of primary forests may take centuries to recover (Finegan, 1996), older

secondary forests (20-35 years) harbor insect communities similar to that of old-

growth forests, successfully sustaining associated ecosystem processes (Taki et al.,

2013). Most Meliponines and many solitary bees are known to be polylectic, i.e., to

consume nectar and pollen from a wide range of plant species (Michener, 2007;

Wcislo & Cane, 1996), thus allowing for the colonization of LS plots when canopy

closure starts to establish and plant diversity increases (10-25 years, Chazdon, 2008)

as observed for stingless bees and *Megalopta*.

Moths showed the strongest turnover and complementary effect between

canopy and understory, particularly in LS forests. This group may serve as a good

example to illustrate the effect of resource turnover during forest succession on

pollinator diversity. The high moth diversity seen in disturbed plots, which has

repeatedly been found for Arctiinae and Sphingidae moths, is probably associated to a

high mobility of most species that allows movement across the matrix during milder

nighttime conditions, as well as with the feeding generalism of caterpillars of many

species (Fiedler et al., 2007; Jaimes Nino et al., 2019; Kitching et al., 2000; Nöske et

al., 2008). Colonizing bushes and herbs, however, are then quickly removed during

early forest succession due to the establishment of the first light-tolerant pioneer trees,

and plant diversity only significantly increases again and is at its highest after the first

decade during the stem exclusion phase (*sensu* Chazdon, 2008), when plant species

suppression, turnover, and growth are the highest stratification starts to take place.

Thus, high plant (functional) diversity may explain both the stronger stratification of

moths in LS plots and the lower diversity in ES plots, where a structured plant

community is still not established.

Whether due to the higher availability of leaf or floral resources, a high

pollinator diversity starting at recently stratified LS forests underlines the importance

of structural complexity in secondary forests in maintaining biodiversity (Hilmers et

al., 2018). Moreover, it signals that the positive effects of forest regeneration in

recovering ecosystem processes are quick to reap. Whenever selective logging cannot

be regulated, allowing patches of forest to naturally recover may thus significantly

increase the resilience of disturbed landscapes (Meli et al., 2017). In conclusion, the

varying patterns of stratification and recovery across taxa highlight the need for

comprehensive biodiversity assessments that consider the different forest strata. For

this purpose, we stress that the archery-assisted pulley system was highly suitable for

the quick setup of traps at heights up to 37 m. Note that this limit was set by the study

site and longer shooting ranges are certainly possible. This method can thus be applied

for primary or successional rainforests with no access to infrastructure, such as canopy

cranes and platforms, or where topography (e.g., inclination) precludes their setup,

thereby aiding in reducing sample size issues (Barker & Pinard, 2001).

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AUTHOR CONTRIBUTIONS

SDL, GB, AK, and UMD conceptualized the research. SDL, AK and GB acquired and

managed the funding. UMD, DB, SVL, and JW performed fieldwork and data

collection. UMD, DB, SVL, JW, MP and KF performed sample processing and data

curation. CR, GB, UMD, DB, SVL, JW, MP, KF identified insects. UMD performed

data analysis and wrote the manuscript draft. All authors contributed critically to the

last manuscript draft.

CONFLICT OF INTERST

The authors declare no conflicts of interest.

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  TABLES

Table 1. Estimates (β) and standard errors (SE) of differences in diversity (q = 1)

between plot types along the successional gradient (top), and between isolated strata

and pooled samples of late successional forests (middle) and old-growth forests

(bottom). The reference plot type is always the first in the couplet, followed by the

compared plot type, hence positive and negative values indicate an increase or

decrease in diversity in the compared plot type in relation to the reference plot,

respectively.

Succession gradient	β (SE)					
	AC - ES	AC - LS	AC - OG	ES – LS	ES - OG	LS - OG
Moths	**-0.29 (0.11)**	0.11 (0.11)	0.04 (0.10)	**0.40 (0.10)**	**0.33 (0.10)**	-0.08 (0.10)
Meliponini	**1.06 (0.36)**	**2.18 (0.45)**	**2.47 (0.39)**	**1.12 (0.40)**	**1.41 (0.33)**	0.28 (0.42)
Euglossini	0.10 (0.09)	0.19 (0.11)	0.19 (0.10)	0.09 (0.09)	0.09 (0.07)	-0.01 (0.09)
Megalopta	0.34 (0.21)	**0.68 (0.26)**	0.90 (0.22)	0.34 (0.23)	**0.57 (0.19)**	0.23 (0.24)

Late succession (LS)	β (SE)		
	Pooled - Understory	Pooled - Canopy	Understory - Canopy
Moths	**-0.22 (0.10)**	**-0.20 (0.09)**	0.03 (0.10)
Meliponini	-0.41 (0.44)	-0.47 (0.43)	-0.06 (0.43)
Euglossini	-0.48 (0.87)	**-2.96 (0.87)**	**-2.49 (0.87)**
Megalopta	-0.38 (0.26)	-0.43 (0.26)	-0.06 (0.26)

Old growth (OG)	β (SE)		
	Pooled - Understory	Pooled - Canopy	Understory - Canopy
Moths	**-0.33 (0.09)**	-0.15 (0.08)	0.18 (0.09)
Meliponini	-0.16 (0.13)	-0.07 (0.12)	0.09 (0.13)
Euglossini	-0.11 (0.10)	**-0.32 (0.11)**	-0.21 (0.11)
Megalopta	**-0.64 (0.19)**	-0.04 (0.19)	**0.59 (0.18)**

Notes: Bold values indicate significant values (p <0.05).

Abbreviations: AC – active disturbance; ES – Early succession; LS – late succession, OG –

old-growth forests.

Table 2. Summary of variance explained by different functional traits obtained by a

Canonical Correspondence Analysis (CCA) analyzing differences in the distribution of

all pollinator groups between canopy and understory. F-statistics and p-values are

based on 999 random Monte Carlo permutations.

Parameter	Moths		Bees (pooled)		Meliponini		Euglossini	
	Variance (%)	F (p)	Variance (%)	F (p)	Variance (%)	F (p)	Variance (%)	F (p)
PL	6.17	3.71 **(.001)**	9.64	5.75 **(.001)**	12.16	9.35 **(.001)**	6.53	3.77 **(.001)**
BS	3.70	2.23 **(.009)**	4.44	2.65 **(.001)**	12.03	9.25 **(.001)**	6.25	3.61 **(.001)**
EPL	3.79	2.28 **(.001)**	4.10	2.45 **(.002)**	7.94	6.10 **(.001)**	4.38	2.53 **(.003)**
AR	3.03	1.83 **(.001)**	6.46	3.86 **(.001)**	10.65	8.19 **(.001)**	8.29	4.78 **(.001)**
Lightness	3.23	1.95 **(.001)**	-	-	-	-	-	-
Contrast	2.04	1.43 (.085**)**	-	-	-	-	-	-
Total	21.96		24.65		42.78		25.46	

1074 Abbreviations: PL - proboscis length, BL - body size (body length for moths, intertegular

1075 distance for bees), AR - wing aspect ratio, EL - effective proboscis length, Lightness -

1076 coloration lightness (moths only), Contrast - coloration contrast (moths only), Total – total

1077 cumulative variance explained

 FIGURE CAPTIONS

1079

Figure 1. A step-by-step process of installing the canopy rope that constitutes the

pulley system used to lift the traps. **a** – preparations for the shot and set up of the

fishing reel and arrow. The inset box lists the technical requirements of the fishing

reel, bow, and arrow for a successful shot. **b** – Selection of a fitting branch at the

canopy, shooting, and arrow retrieval. This step entails most of the difficulties that

may arise during the process. See Box for a list of issues that can occur and respective

solutions. **c** – replacement of the fishing line with the final pulley rope. **d** – Overview

of the successfully set rope and traps. Persons, traps, and trees are not to scale. Created

in BioRender (https://BioRender.com/i24e291).

1089

Figure 2. Paired differences in abundance, diversity (rarefied exponential Shannon

diversity), functional richness (FRic) and functional dispersion (FDis) of moth and bee

pollinators between the understory and the canopy of the sampled plots. Asterisks

indicate significance levels reported by the t-tests (ns = not significant, * = < 0.05, **

= < 0.01, *** < 0.001), and lines between paired points indicate the plot legacy.

Functional richness and dispersion values are not shown for *Megalopta* bees due to

low diversity in most plots (<3 species). Horizontal lines within boxes indicate the

distribution's median, box edges the 25th and 75th percentiles, and whiskers the

minimum and maximum values.

1099

Figure 3. Rarefied diversity (q = 1) of the four major pollinator groups captured along

the successional gradient: active disturbances (AC), early successional plots (ES), late

successional forests (LS), and old-growth forests (OG). LS and OG are further

separated into understory (U) samples, canopy samples (C), and pooled samples, for

which diversity was measured independently. Asterisks indicate significant differences

($p < 0.05$) between the means of pooled, understory, and canopy samples within the

latter two groups. The dashed lines indicate the mean diversity across all plots. A:

moths, B: Meliponini, D: Euglossini, E: *Megalopta.* Horizontal lines within boxes

indicate the distribution's median, box edges the 25th and 75th percentiles, whiskers the

minimum and maximum values, and the shaded area datapoint density.

1110

Figure 4. Model estimates for the effects of plot variables on the community

parameters (abundance, Shannon's exponential diversity, functional dispersal (FDis)

and functional richness (FRic)) of the four main pollinator groups sampled. Points and

bars depict estimate means and error bars, respectively, which are colored according to

the direction of the association between variables [positive (blue) or negative (red)]

and significance ($p < 0.05$). Plot variables: **Leg** – disturbance legacy (successional

forests compared to old-growth forests), **Hei** – canopy height, For – proportion of

forest in a 500 m buffer, **Ele** – plot elevation. For the legacy variables, old-growth

plots are set as the reference group.

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Figure 5. The effect of functional parameters on the divergence of all bee groups

pooled (a), as well as specifically on the two dominant bee groups Meliponini (b) and

Euglossini (c), and moths (d). Canopy and understory plot samples are separated by

shape and color, and grey crosses represent species. Axes correspond to the two main

components of the Canonical Correspondence Analysis (CCA), and percentages are

the variance explained by each axis. Vectors indicate the direction and intensity

(length) of the constrained variables: **PL** = proboscis length, **BS** = body size, **EPL** =

effective proboscis length, **AR** = wing aspect ratio, **contrast** = coloration contrast,

lightness = coloration lightness.

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Figure 6. Model estimates of the occurrence probability of different visual categories

of bees (A) and moths (B) at the canopies of the studied plots relative to the

understories (positive and negative values indicate an association with canopy and

understory, respectively) in the three disturbance legacy classes (old-growth forests,

former cacao plantations, and former pastures). Bars indicate standard errors.

Estimates are significant (p < 0.05) unless stated otherwise (n.s. on top of the

estimate). Categories: **Apo** – aposematic, **Dar** – dull dark, **Bri** – dull bright, **Blu** –

iridescent blue, **Gre** – iridescent green, **Red** – iridescent red, **Cry** – cryptic, **Col** –

coleoptera mimic, **Hym** – hymenoptera mimic.

1140

1141   **FIGURES**

1142   **Figure 1**

Figure 2

1146   **Figure 3**

1148   **Figure 4**

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1150   **Figure 5**

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1152   **Figure 6**

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a

Preparing the shot

1. Line tightly rolled onto the spool - no loose sections.

2. Line between the spool and the arrow straight, but not tight.

3. Completely loose drag adjustment.

4. Bail disarmed.

5. Line tied to the arrow shaft, before the fletching.

6. Reel facing the nock of the arrow, at the same angle.

b

Relatively isolated, horizontal branch with little vegetation or clutter around it.

Arrow is shot above the branch from a diagonal, almost vertical angle - never from a completely vertical angle.

Arrow might be difficult to find, bright-colored markings are used on the shaft.

c

Large knots that might get stuck at the branch are avoided.

Drag adjustment fully tightened up and bail re-armed.

Line is slowly pulled back.

The fishing line is severed from the arrow and tied to a resistant, lightweight (e.g., polypropylene) rope.

d

Traps are placed above one another and above the knot made to connect the rope

The two ends of the rope are tied together; the rope is secured at the tree itself.