Specimen-tailored "lived" climate reveals precipitation onset and amount best predict specimen
phenology, but only weakly predict specimen reproductive success across a clade

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Summary

- Herbarium specimens are broadly distributed in space and time, enabling investigation of impacts of climate on phenology and persistence . Many studies have revealed climate-associated phenological shifts; we ask which of temperature, precipitation, or the timing of precipitation onset best explain specimen phenological stage, and whether the same variables also explain specimen reproduction, a measure closely aligned with fitness.

- We calculated specimen-specific climate from estimated specimen germination date until average collection dates for 14 annual species of *Streptanthus* and *Caulanthus* (Brassicaceae). We also test whether relationships of phenology and reproduction with climate variables are evolutionarily constrained.

- Precipitation amount and onset date predicted specimen phenology, but much less so, specimen reproductive success. Earlier rainfall onset was associated with more phenologically advanced specimens, a relationship that showed evolutionary conservatism; however, such advancement had little effect on specimen reproduction. This lack of correlation may arise from processes like phenological shifts that buffer fitness impacts of climate, interactions with other community members also responding to climate, or challenges in estimating total reproduction from herbarium specimens.

- We make suggestions for collection protocols that could improve the usefulness of herbarium data to understand climate impacts on reproduction, a better predictor than phenology of future population viability.

Introduction

As climate change alters the environmental conditions that species experience, an urgent and open question is how climate affects plant performance and ultimately population stability. A number of studies have addressed how changing rainfall and temperature may affect plant growth, phenology, and reproduction in greenhouse and field experiments, and the results are often complex. In natural settings, diverse antagonists and mutualists may mediate plant responses to climate, contributing to the complexity of outcomes (González-Megías & Menéndez, 2012; Barnett & Facey, 2016; Wade *et al.*, 2017). Herbarium specimens collected from natural plant populations provide valuable historical data and allow for insights into how climate conditions affect plant performance (Davis *et al.*, 2015; Willis *et al.*, 2017; Meineke *et al.*, 2018) by integrating over biotic and abiotic environments. Since specimens are collected over large spatial scales and across diverse habitats, as well as over many years with varying

climate conditions, analyses of specimen characteristics can be complementary to experimental approaches for understanding impacts of climate on plant phenology and fitness.

Numerous studies have analyzed phenological data from herbarium specimens and have revealed signatures of changing climate on phenological responses, as well as differences among species and regions in phenological sensitivity to climate variation (Primack *et al.*, 2004; Miller-Rushing *et al.*, 2006; Calinger *et al.*, 2013; Jones & Daehler, 2018; DeLeo *et al.*, 2020; Love & Mazer, 2021; Pearson *et al.*, 2021; Willems *et al.*, 2022; Dangremond *et al.*, 2022; Ramirez-Parada *et al.*, 2024). In contrast, only a few studies have analyzed measures of fitness or reproductive success from herbarium data, and to date, these studies have focused on integrating individual performance with species distribution models to investigate the climatic factors contributing to species range limits (Bontrager & Angert, 2016; Yim *et al.*, 2024).

Phenology and reproductive success are not just affected by the magnitude but also the timing of weather events (Steyn *et al.*, 1996; Levine *et al.*, 2011; Olliff-Yang & Ackerly, 2021; Martínez-Berdeja *et al.*, 2023; Worthy *et al.*, 2024a). In western North America, recent studies have shown that rainfall onset is occurring later in fall and precipitation in fall is decreasing (Swain *et al.*, 2018; Luković *et al.*, 2021). These trends, layered onto increasing temperature trends over the past century (Pathak *et al.*, 2018), are effectively increasing drought stress, shifting seasonal timing, and potentially decoupling temperature and precipitation patterns (Flint *et al.*, 2013; Swain *et al.*, 2018; Luković *et al.*, 2021). The timing of rainfall may have especially large effects on the growing season in Mediterranean ecosystems where rainfall occurs in only a few months of the year, beginning in the fall and ending with the onset of months-long drought in spring. Annual plant species, in particular, may suffer reduced fitness due to shortened growing seasons with later-arriving rainfall (Steyn *et al.*, 1996; Levine *et al.*, 2011; Olliff-Yang & Ackerly, 2021; Martínez-Berdeja *et al.*, 2023; Worthy *et al.*, 2024a). In a lathhouse experiment (where plants are exposed to ambient temperatures but controlled watering regimes), simulated later rainfall onset caused some native annual species to speed up their phenology, thereby making up for later germination, but these phenological shifts could not fully compensate for the effects of shorter growing seasons on reproductive fitness (Worthy *et al.*, 2024a; see also Steyn *et al.*, 1996; Levine *et al.*, 2011; Olliff-Yang & Ackerly, 2021; Martínez-Berdeja *et al.*, 2023; Zettlemoyer *et al.*, 2024). In the field, effects of delayed rainfall onset also depend on rainfall distribution and total amounts—an early rainfall year, which could benefit plant reproduction by lengthening the growing season, might be disastrous if a mid-season drought ensues (Hatchett *et al.*, 2023).

Here, we relate precipitation, temperature, and the timing of rainfall onset to the phenological
stage and total reproductive output of herbarium specimens and ask whether the same climatic
variables affect both of these reproductive properties. Our methods focus on reconstructing the
"lived climate" for each specimen from the estimated germination date in the fall of the year of
collection until collection in the spring or summer. We examine specimens of 14 annual species
in the native Streptanthoid complex (Brassicaceae) that germinate in the fall and complete
reproduction in early to mid-summer. We studied this clade because we have extensive
knowledge of its ecology (see below) as well as a robust evolutionary hypothesis (Cacho *et al.*,
2014) and thus can ask whether phenological and reproductive responses to climate might be
constrained by evolutionary history.

In this system, the timing and extent of rainfall affect phenology and fitness, and these effects
vary across the clade and in relation to local climate. Our previous experimental work provides
some *a priori* expectations for how these species might respond to changing precipitation levels
or timing in the field and also shows that evolutionary history of the clade may influence these
responses (Pearse *et al.*, 2022; Worthy *et al.*, 2024a). In a common garden study of 11 annual
Caulanthus and *Streptanthus* species from the California Floristic Province (Raven & Axelrod,
1978), we found that higher-aridity *Caulanthus* species flowered relatively earlier than species
from wetter habitats and could maintain reproduction at field-relevant extremely low water
levels, but were unable to extend their flowering periods when ample water was provided. In
contrast, taxa from relatively wetter Mediterranean climates flowered later and extended their
flowering periods substantially with ample water, but were unable to reproduce at the lowest
water levels. A separate experiment manipulating the seasonal timing of rainfall revealed that
fitness generally declined with later rainfall onset, but the magnitude of this effect varied among
species; more mesic species accelerated flowering more than arid species in response to late
rainfall arrival, and such compensatory plasticity resulted in smaller fitness declines (Worthy *et*
al., 2024a). Experiments on germination timing revealed that these species germinate in the fall
with rainfall onset (Worthy *et al.*, 2024b).

Extending these results to herbarium specimens of the same and related species, we predicted
a priori that low rainfall years should result in specimens with lower total reproduction relative to
wetter years (Pearse *et al.*, 2020), and that *Caulanthus* species from arid habitats would be
less responsive to rainfall amount. We also predicted that late rainfall onset should result in
specimens with relatively delayed phenology, especially in arid species with little compensatory
plasticity (Worthy *et al.*, 2024a). More broadly, we ask: 1) Within species, can we detect effects
of climate—precipitation, temperature and climatic water deficit—on the phenology and

reproductive success of herbarium specimens, and do the same climate variables influence both aspects of reproduction? 2) Does later onset of seasonal rainfall reduce specimen reproductive output or alter phenology? 3) Across species, are phenological and fitness responses to climate associated with each other and 4) are they evolutionarily conserved across the clade?

Materials and Methods

Study system, species selection, and image acquisition

Streptanthus (s.l.) is a Brassicaceae clade that arose in the southwestern desert of western North America (Cacho *et al.*, 2014) and continued to speciate as species moved west and northward as part of what Raven and Axelrod (1978) termed the Madro-Tertiary flora (Cacho *et al.*, 2014, 2021). The Streptanthus I clade is comprised of ca. 40 species in the *Streptanthus* and *Caulanthus* genera that occupy a diversity of habitats, spanning desert ecosystems to mid-montane habitats (Cacho *et al.*, 2014). They are primarily annual species and all share an affinity for open, bare, rocky or sandy habitat (Cacho & Strauss, 2014). Species encompass a remarkable diversity of range sizes, latitudinal ranges, and elevational distributions (Fig. S1). Thus, the clade occupies a variety of climatic conditions (Pearse *et al.*, 2020; Cacho *et al.*, 2021; Worthy *et al.*, 2024b), and within some of the more wide-ranging taxa, populations can span a large amount of intraspecific climatic variation (Gremer *et al.*, 2020; Love & Mazer, 2021).

From the ~40 species in *Streptanthus* I major clade (Cacho *et al.*, 2014), we chose 14 annual species to include in this study. The selection of species was based on three criteria: 1) sampling subclades across the phylogeny, 2) sampling species from across the latitudinal and elevational space that the genus occupies, and 3) sampling species with an adequate sample size of individuals (>40 specimens). The last criterion resulted in the exclusion of the most narrow-ranged endemic species, which comprise about 20% of the group.

To assess phenological stage and reproductive success at the time of collection, we counted and categorized reproductive structures on high resolution digital images of herbarium sheets available via the Consortium of California Herbaria specimen data portal (cch2.org). We supplemented these images with herbarium sheets from the California Academy of Sciences herbarium that we photographed ourselves. These efforts resulted in inventories of 53-195 herbarium sheets per species (average = 145). When >200 sheets were available for a given species, we focused our counting efforts on approximately 200 sheets, subsampling by county

to maximize geographic coverage. All data were collected at the individual plant level. Often, multiple plants were mounted on a single sheet (average of 2.4 specimens/sheet, range = 1-12; non-independence was addressed with random effect structure, see statistical methods). Before counting an individual, we checked for the presence of roots, rosette leaves or rosette leaf scars and only counted individuals that appeared to be entire plants.

Counting and measuring specimens

Because machine learning has been shown to undercount reproductive structures of herbarium specimens (Love *et al.*, 2021), a team of fifteen people counted and classified reproductive structures on 1939 specimens. Each counter was trained on the same set of specimens by the same team member using an annotated guide to the different types of structures. Counts were spot-checked for consistency across species and counters. We used a custom plugin in ImageJ (modified from CellCounter) to count and categorize reproductive structures; this plugin made counting more accurate by allowing the counter to select structure types by name— e.g., buds, flowers, fruits—and place color coded counters on the images. These counts were then saved in .xml files for collation and spot-checking. Reproductive structures were classified in detail, including missing structures that we inferred to be buds, flowers and fruits, based on persistent empty pedicels and the position of those pedicels relative to other buds, flowers and fruits (see Table S1). "Empty" pedicels could arise from selective abortion of structures, natural damage before collection, or loss during specimen preparation. Because flowers produce pollen and are part of male fitness in these species, we included all pedicels in our reproductive structure count, even if they did not produce fruit. Some species have sterile buds forming a showy tuft at the top of the plant; the structures comprising this tuft were not counted. Species varied in the dates and mean phenological stages at which they were collected (Fig. S2).

Georeferencing

Geographic data and locality descriptions for each herbarium sheet were downloaded from the Consortium of California Herbaria and checked for consistency with described collection sites. We used Google Earth, complemented by the USGS Historical Topo Map database (https://apps.nationalmap.gov/downloader/#/maps) and results from google searches of location descriptions. We added error radius distances when they were missing. These were estimated by the georeferencer as the distance at which the provided description would no longer be the most logical. When specimens were recent and the label indicated that coordinates were recorded in the field with a GPS, rather than by another party after specimen accession, we tended to assign a smaller error radius if missing (though in most cases these

collections already had an error radius recorded). Specimens for which we could not determine

coordinates with <=5 km error radius (n = 112) were not used (though the large majority of

specimens had much more precise coordinates, mean = 1.1 km, median = 0.5 km).

Estimating phenological stage and reproductive output

To quantify phenological stage, w e used a weighted sum of reproductive structures (0*number

of buds + 0.33*number of flowers + 0.66*number of immature fruits + 1*number of mature

fruits). This is similar to the metric used in Love *et al.* (2019), but scaled so that a binomial

response distribution can be used (in our statistical models, the weighted sum is used as the

number of "successes", total reproduction as the number of "trials", and in figures and text the

"phenological stage" refers to successes divided by trials). Based on our metric, a specimen

with a phenological stage of 0.1 would mostly have buds, while a specimen with a stage of 0.9

would be mostly in mature fruit. To quantify reproductive output, we used the sum of all

reproductive structures, generously assuming that every reproductive structure would make a

fruit, and bearing in mind that flowers contribute to male fitness of plants.

In our statistical models, we accounted for the day of year that a specimen was collected, as

we expected that individuals that grew for longer periods of time had more time to advance in

phenology and to reproduce, regardless of the climate conditions they experienced. The

collection day-of-year variable was calculated as the number of days starting from September 1

in the fall prior to collection until the collection date the following spring/summer. Collection

day-of-year was included in every model as it was frequently positively related to our response

variables of phenology and total reproduction (Figs. 1, S3).

Estimating growing season climate

In Mediterranean ecosystems, precipitation starts in the fall, typically peaks in the winter and is

very rare in the summer months (Di Castri & Mooney, 2012) . For annual species in

Mediterranean climates, fall precipitation of sufficient quantity stimulates germination and

species complete their life cycles in a single season before surviving the summer drought as

seeds (Keeley, 1991). Prior experiments on germination behavior in the *Streptanthus* clade

(Worthy *et al.*, 2024b) and other winter annual plants have found that most species have their

greatest germination during early fall rain events (Levine *et al.*, 2008; Hänel & Tielbörger, 2015;

Martínez-Berdeja *et al.*, 2023), which can start as early as September. To estimate the timing of

germination, we identified relevant rain events, which we considered to be when a total of 25

mm or more rain fell. This threshold precipitation amount was chosen based on prior work

showing that 25 mm of rain was needed to trigger biological activity (Went, 1949; Tevis Jr.,
1958; Beatley, 1974). We extracted climate data from the time period preceding the collection
of each specimen from two different sources which offered two different temporal resolutions.

Monthly climate data were extracted from the California Basin Characterization Model (Flint &
Flint, 2014). We used monthly data for each specimen beginning with the first month after
August with >25 mm precipitation and ending with the average collection month across all
specimens of a species. We used the average collection month of each species, rather than
the specific collection month of each specimen, to avoid confounding our climate
characterizations with collection timing. We extracted maximum temperature (tmx), minimum
temperature (tmn), precipitation (ppt), and climatic water deficit (CWD). We calculated average
temperature (monthly (min + max)/2), summed CWD, and summed precipitation for the time
period preceding collection of the specimen.

We also used climate data available at a daily resolution from the PRISM database (PRISM
Climate Group, Oregon State University, https://prism.oregonstate.edu , accessed 14 Dec
2023). We used these daily data to estimate germination timing at a finer temporal scale than
the monthly data allowed, using the same >25mm threshold. We omitted eight specimens from
this subset of the data because there was no rain event >25mm until after March 1, which is no
longer a plausible germination time (Worthy *et al*. 2024). Unfortunately, daily climate records
are only available starting in 1981, and thus only specimens collected after 1981 could be used
for analyses that included putative germination timing as a predictor (n = 1082) .

Statistical analyses

Q1: How does lived climate affect the phenology and reproduction of herbarium specimens?

The climate variables we used were average temperature, summed precipitation, and summed
CWD, calculated as described above to capture what each specimen experienced in its
lifetime. CWD is an overall measure of drought stress, and incorporates temperature,
precipitation, soil water holding capacity, and aspect. Because CWD is derived in part from
temperature and precipitation, it is correlated with both of these variables (positively with
temperature and negatively with precipitation). We fit two different models for each response
variable (phenological stage and total reproduction): one that included specimen-experienced
temperature and precipitation together, and another with CWD alone. The model with CWD
alone allows us to explore the effects of drought stress specifically, while the model with

temperature and precipitation allows us to see the relative importance of these two climate variables for plant reproduction.

To estimate the effects of climate on phenology, we fit generalized linear mixed effects models with a binomial response distribution and a logit link function for each species separately. To estimate the effects of climate on reproductive output, we fit generalized linear mixed effects models with a negative binomial (gamma-Poisson) response distribution and a log link function, again using data from each species separately. In both cases, models were fit with the Bayesian regression package brms (Bürkner, 2017) and included a covariate of collection day-of-year. We could imagine scenarios in which a quadratic relationship might better explain the relationship between total reproduction and climate; for example, under low precipitation, total reproduction could be reduced by drought, while at high precipitation, total reproduction could be reduced by inter- or intraspecific competition (Hänel & Tielbörger, 2015). To explore these possibilities, we checked whether a quadratic function better explained the relationship between total reproduction and climate variables than a linear one; in the vast majority of cases, quadratic terms for temperature, precipitation, and CWD were not significant (the exceptions were *C. cooperi*, which had highest reproduction at intermediate CWD, *C. anceps*, which had highest reproduction at intermediate precipitation and high/low temperatures, *S. insignis*, which had highest reproduction at intermediate temperatures) and thus only linear relationships were tested in the models reported here.

For analyses related to questions 1 and 2, we included a group-level (random) effect of collection location, which was generated by placing buffers around each collection locality with a radius of 2.5 km (or their coordinate uncertainty, if larger than 2.5 km) and assigning points with overlapping buffers to the same group level (this method allowed "daisy chain" effects where two points without overlapping buffers were joined by a common neighbor). Multiple herbarium sheets are sometimes collected at the same location (e.g., to deposit in different herbaria) and so the non-independence of the specimens on these sheets is also represented in this effect. Predictors (collection day-of-year and climate variables) were centered and scaled before analysis. We used a weak normal prior on betas, the default student-t prior on the intercept, the default bounded student-t prior on the standard deviation of group-level intercepts, and the default inverse-gamma prior on the shape parameter. All parameters were assessed for convergence (Rhat = 1), parameter estimates and convergence metrics for all regression models are provided in Appendix 1 (see online supporting information). We interpret effects to be different from zero when the 95% confidence interval of the estimate does not include 0. We visualised predicted regression slopes and confidence intervals using the

ggemmeans() function from the package ggeffects (Lüdecke, 2018). For each model for each
species, we checked whether predictor variables were correlated with each other. In all cases
except three, correlation coefficients were <0.6, in the remaining three they were <0.7.

Q2: Does later onset of seasonal rainfall timing alter phenology or reduce reproductive output?

A priori, we predicted from results of our lathhouse experiments that late rainfall would result in
later reproduction (represented by lower phenology scores, controlling for collection date), and
ultimately would decrease fitness (Worthy *et al.*, 2024a). Conversely, early rainfall should lead
to earlier reproduction (higher phenology score) and increase fitness by lengthening the
growing season, independent of rainfall amount and temperature. To test these hypotheses, we
used the subset of specimens for which we had estimated germination date at the daily scale
(see Methods: Estimating growing season climate). Using this subset of the data, we ran
models with average temperature, total precipitation, and collection day-of-year as described
for Q1 but added estimated germination date as an additional predictor. All other model details
are as described for Q1. We again checked each species for correlations between the four
predictors used. Most pairs of predictors had correlation coefficients below 0.6, six species had
one to three pairs of predictors with correlation coefficients >0.6 and <0.76.

Q3: Across species, are phenological and total reproduction responses to climate variables
correlated?

Plasticity in phenology in response to climate may be adaptive and may buffer fitness from the
effects of climate variation. Species with phenologies that respond to climate variation may be
buffered from effects of climate on total reproduction; in contrast, species that are unable or do
not buffer climate variation via phenological timing—that is, have more fixed phenologies
regardless of climate experienced—should be more affected by climate variation (Worthy et al.,
2024a). Across species, we tested for correlations between slopes of the relationships between
species' phenological and reproductive responses to each climate variable using phylogenetic
independent contrasts implemented with the pic() function in the R package ape (Paradis &
Schliep, 2019) and the phylogeny from Cacho et al. (2014).

Q4: Are responses to climate evolutionarily conserved across the clade based on phylogenetic
relationships?

To evaluate whether there might be evolutionary constraints in how species respond to climate
(temperature, precipitation, CWD, and the timing of the first rainfall event) in their phenology
and reproductive output, we tested for phylogenetic signal in effect size and direction from the

models above. For this, we used Blomberg's K (Blomberg *et al.*, 2003), calculated with the
phylosig function in the phytools package (Revell, 2012). We used the estimated error in our
model effect sizes as the sampling error. We estimated the significance of phylogenetic signal
using 10,000 simulations in a randomization test. When significant, we then tested whether the
estimate of Blomberg's K was significantly greater than one by comparing our estimate of K to
a null distribution of 10,000 K values generated under Brownian motion (Revell & Harmon,
2022). K has an expected value of one under evolution by Brownian motion (Revell & Harmon,
2022), where values of K significantly greater than one indicate that species tend to have more
similar phenology and reproductive output responses to climate than expected based on a
Brownian motion model of evolution, i.e. more evolutionary constraint. Significant effects of
phylogenetic relationships on the size and direction of climate effects might also indicate
climate adaptation during species divergence.

Results

Q1: How does lived climate affect the phenology and reproductive success of herbarium
specimens?

Phenology
We found the effects of total precipitation, average temperature, and CWD experienced by
specimens on the phenological stage of the specimen (Figs. 2, S4). The direction and
magnitude of effects varied among species and climate variables. Precipitation amounts
significantly affected phenology for 10 species: increased annual precipitation significantly
delayed phenology in eight species, slightly accelerated phenology in two species, and had no
effect on four species (Fig. 2ab), all while taking into account effects of average temperature
and collection timing. Eight species adjusted phenological stage in response to temperature,
with seven accelerating phenology with warmer temperatures, and one species delaying
phenology with warmer temperatures (Figs. 2b, S4a). Seven species also significantly
responded to CWD, albeit more idiosyncratically than to temperature or precipitation, with four
species slightly accelerating phenology, and three slightly delaying phenology with increasing
CWD (Figs. 2c, S4b). These analyses all included collection day-of-year as a covariate to
account for the period of time that a specimen grew (Figs. 1a, S4). Precipitation overall had
stronger effects than temperature or CWD on phenology, based on the frequency of significant
slopes and the magnitude of effect sizes relating phenology to climate variables.

<u>Total number of reproductive structures</u>

Specimens collected later in the season had more reproductive structures for eight of fourteen

species—the remaining six species showed no effect of collection day (Figs. 1b, S3b).

Increased precipitation had a negative effect on total reproduction for five of the

species—perhaps counterintuitively in California's mediterranean climate and considering

results from Pearse *et al.* (2020)—and a positive effect on two of the species (Fig. 3ab). As with

phenology, temperature had fewer impacts on reproduction than did precipitation, with only two

species showing significant responses (Figs. 3b, S5a). Higher temperatures were associated

with lower reproduction for *S. diversifolius,* and were positively related to reproduction for *S.*

insignis. Our separate model examining the relationships between CWD and reproductive

output also showed varying effect sizes and directions (Figs. 3c, S5b). In four species

increased CWD had positive effects on total reproduction and in two species increased CWD

had negative effects on total reproduction.

Q2: Does later onset of seasonal rainfall timing alter phenology or reduce reproductive output?

For nine species, earlier rainfall onset resulted in specimens that were collected at a more

advanced phenological stage relative to specimens that experienced later rainfall onset (Fig 4).

These effects were present when also taking into account collection timing, average

temperature, and total precipitation during the specimens' growing seasons (Fig S6), and did

not change substantially when temperature and total precipitation . In contrast, with these same

covariates in the model, the timing of germination-triggering rains had no effects on specimen

reproductive output except for one species, *S. drepanoides*, which produced relatively more

fruit with later rainfall onset (Figs. 5, S7). This result suggests that individuals may be able to

buffer fitness effects in the face of variable rainfall timing via phenology, or that effects of other

community members responding to the same climatic conditions result in a lack of

correspondence between timing of rainfall onset and total reproduction.

Q3: Are phenological and total reproduction responses to climate variables correlated?

Across species, we asked whether species with greater phenological plasticity had lower

fitness sensitivity to climate. Slopes of phenological responses were not significantly related to

slopes of total reproduction responses for any climate variable. Thus, phenological and

reproductive sensitivities to climatic variables have not evolved together across the clade (Fig.

S8).

Q4: Are responses to climate evolutionarily conserved across the clade based on phylogenetic relationships?

We found no evidence of evolutionary history (phylogenetic signal) influencing phenological or reproductive responses to temperature, precipitation, or CWD (Table S3). However, there was significant phylogenetic signal in both phenological and reproductive responses to the timing of first rain events,which are likely to trigger germination (K>1, P<0.001 for phenology, and <0.026 for total reproduction; Figs. 4, 5; Table S3). This result further illustrates the value of our lived climate approach, as these data are based on daily climate records specific to the places and times these specimens were growing. Phylogenetic signal arises when traits (e.g. slope of relationship between specimen phenology and rainfall timing) of closely related species are more similar to each other than to those of more distantly related species; for example, the subclade of southern 'true' *Caulanthus* species share very similar shallow slopes in their phenological responses to rainfall onset timing, and these differ from the magnitude of slopes of the subclade of closely related *S. breweri* and *S. drepanoides,* which between themselves similar responses to rainfall onset. (Fig. 4). These results might indicate more evolutionary constraint in adapting to the timing of rainfall onset than to absolute amounts of precipitation (no phylogenetic signal in slopes relating phenology or reproduction to total precipitation; Figs. 2, 3).

Discussion

Herbarium collections offer incredible resources for understanding the impacts of climate change on plant performance as they span many climatological conditions, large geographic regions and integrate over biotic as well as abiotic environments. A growing number of studies have shown that the phenological stages of herbarium specimens at collection often reflect the climate in which they grew (Primack *et al.*, 2004; Miller-Rushing *et al.*, 2006; Calinger *et al.*, 2013; Jones & Daehler, 2018; DeLeo *et al.*, 2020; Love & Mazer, 2021; Pearson *et al.*, 2021; Willems *et al.*, 2022; Dangremond *et al.*, 2022; Ramirez-Parada *et al.*, 2024). Our study contributes to this literature by relating climate variables not only to specimen phenology but also to specimen total reproductive success, which, in annual species, is a key fitness component. Importantly, we also consider climate-performance relationships in light of specimen-specific tailored estimates of climate (see also Bontrager & Angert, 2016; Hereford *et al.*, 2017; Love & Mazer, 2021; Mazer *et al.*, 2021 for other studies exploring more tailored climate windows). The lived climate of a specimen was characterized from estimated germination date to species average collection date using fine-scaled temporal and spatial

climate records and knowledge of previously determined germination cues (Worthy *et al.*,
2024b). In the course of analyzing data with these aims, we came to the realization that
collection protocols aimed at accurate species identification could be supplemented with
protocols that would improve our ability to measure climate impacts on plants and population
viability using specimen data. Thus, we also propose collection protocol additions that could
complement the current practices for specimen collection.

Effects of lived climate on plant phenology and reproduction

Of four potentially important climate variables (average temperature, total precipitation, timing
of precipitation onset and total CWD), timing of onset and total amount of precipitation had the
greatest impacts on plant phenology and reproduction, based on the number of significant
relationships and the magnitude of slopes of these relationships (Figs. 2-6). Ten species
exhibited a significant relationship between total precipitation and phenological stage, and eight
species demonstrated a relationship between total precipitation and total reproduction. In
contrast, seven species had relationships with phenology and temperature, and only two
showed relationships between temperature and reproduction (Fig. 3). A recent meta-analysis of
climate impacts on phenology in field experiments finds temperature to be more important than
precipitation in predicting phenological shifts (Zhou et al., 2023); similarly, a meta-analysis of 21
plant traits measured in the field also found temperature to be more important than precipitation
in influencing trait values (Moles *et al.*, 2014). However, in water-limited ecosystems,
precipitation may be the dominating force influencing phenology (see meta-analysis of Lu *et al.*,
2023), especially for annual species with less access to deep water reserves (Currier & Sala,
2022). For example, phenological change in grasses in an arid ecosystem was more impacted
by precipitation than temperature (Currier & Sala, 2022).

Lack of correspondence between phenological and total reproduction responses

A large number of processes may decouple phenological from fitness responses. Rapid
phenological shifts can compensate to some degree for climatic stress, reducing the impact of
climate on total reproduction (Martínez-Berdeja *et al.*, 2023; Miller & Stuble, 2024; Worthy *et*
al., 2024a and see below). In California mediterranean ecosystems, fall precipitation onset,
which is the cue for germination in many annual species (e.g., Keeley, 1991; Levine *et al.*,
2008; Worthy *et al.*, 2024b) is occurring later, and providing less water (Swain *et al.*, 2018;
Luković *et al.*, 2021). We found that later rainfall onset resulted in specimens with more
delayed phenology in 9 of our 14 study species, after the effects of growing season
temperature and precipitation amounts were accounted for. These effects on phenology,

however, did not translate into significant effects on reproductive success (Figs. 3, 4), suggesting the possibility of compensatory buffering of phenological responses to stabilize fitness responses (Martínez-Berdeja *et al.*, 2023; Miller & Stuble, 2024; Worthy *et al.*, 2024a). Our previous experimental work showed that many of these species can speed phenological phases, for example by accelerating flowering rate in response to later germination timing (Worthy *et al.*, 2024a), as has also been observed in other annual species in Mediterranean climates (Steyn *et al.*, 1996; Olliff-Yang & Ackerly, 2021; Martínez-Berdeja *et al.*, 2023). Interestingly, the responses of species to rainfall onset timing in both phenology and total reproduction were the only relationships that were evolutionarily conserved (phenology P<0.001, Figs. 4, S5, Table S3; reproduction P<0.02, Figs. 5, S6; Table S3). Moreover, the general pattern of responses, with the southern *Caulanthus* subclade being less responsive phenologically to rainfall, is consistent with results of our previous watering experiment (Pearse *et al.*, 2020). Our results suggest that adaptation to rainfall timing might be more constrained than adaptation to total amounts of rainfall.

Another potential mechanism underlying the lack of correspondence between phenology and total reproduction entails interactions with other species (Farías *et al.*, 2021; Van Dyke *et al.*, 2022; Armitage, 2024; Terry, 2024). These include climate-induced changes in competitive hierarchies (e.g., Niu & Wan, 2008), mismatch or facilitation of pollination (e.g., Kudo & Ida, 2013; Gezon *et al.*, 2016; Tiusanen *et al.*, 2020; Freimuth *et al.*, 2022), changes in interactions with herbivores (Fabina *et al.*, 2010; Meineke *et al.*, 2018; Hamann *et al.*, 2021), or other interactors like microbes (Ware *et al.*, 2021). On top of all these ecological complexities is the possibility that specimens may not have been collected at an advanced enough phenological stage to provide a good estimate of the true total reproductive potential of the specimen. The average specimen was 28% fruit at time of collection. Despite this limitation, we did find significant effects of precipitation on total reproduction in five species. All of these complexities could reduce the linkages between climate and total reproductive success in herbarium specimens, even when climate may be impacting plant phenology. Revised collection protocols could make linkages of specimen reproduction to climate more useful for projecting resilience to climate change (see *Making collections even more valuable for climate change research* below).

Making collections even more valuable for climate change research

If we want to draw on the incredible extent of temporal and spatial sampling that herbarium specimens provide, and if we want to use specimens to answer questions like the ones posed here—specifically, assessing fitness responses to ongoing climate change—we suggest some

changes to collection protocols that could make collections more valuable for these purposes.

Most of them include targeted increased sampling effort within species and across years.

Increase the number of stages at which specimens are collected . The aim of most

botanists in specimen collection is to capture all the reproductive organs and stages of a

specimen to best enable identification (.e.g,

https://www.usu.edu/herbarium/education/learning-about-plants/making-specimens). However,

if we want to use specimen performance for understanding responses to climate change,

ideally we would also want to collect specimens later in the season as well, when most of the

population is predominantly in fruit and individuals are closer to their full reproductive potential.

Collect a number of representative specimens based on size and phenological stage.

Protocols for collecting specimens already emphasize that specimens should be representative

of the population e.g.,

(https://brahmsonline.kew.org/Content/Projects/msbp/resources/Training/15-making-her

barium-specimens.pdf ;

https://www.usu.edu/herbarium/education/learning-about-plants/making-specimens)**,** but

explicitly including a place on the collection data form where this assessment could be asserted

would provide more confidence that specimens really are representative. Collecting enough

individuals to get a rough estimate mean and variance in specimen traits (e.g., 8-10 individuals)

would also be very valuable when logistically feasible (e.g., with small-statured species).

Collect in the same places and every year . While there is tremendous value in expansive

samping across a species' range, there is also value in having at least a few populations that

are consistently collected every year. More complete time series from the same localities will

allow better estimation of impacts of climate because they will be measured over the most

similar ecological contexts—e.g, other community members, soil environments—through time.

Provide a community context. Because local plant density also tells us about plant

performance, crude estimates of population density in a representative 0.25 meter square

would be helpful for interpreting changes in reproduction across years—even if it were a crude

estimate of singles, tens, or hundreds of individuals. To understand whether populations are

stable, we would want to be able to distinguish among various scenarios that could result in the

same specimen fitness. For example, specimens could have a relatively lower individual fitness

because there were high densities of conspecifics suppressing individual performance or

because conditions were suboptimal, with low densities of conspecifics. In addition, linking a

digital image of a 0.25 meter square centered on where the specimen was collected to the

digital specimen would situate the specimen ecologically.

Conclusions

Herbarium specimens are valuable resources that preserve a record of the effects of historical

conditions on plant traits and performance. Our study shows that amounts of precipitation and

timing of precipitation onset are important climate factors influencing herbarium specimen

phenology across this clade. While phenological responses to climate are important, linking

those changes to plant fitness will ultimately determine the usefulness of herbarium specimens

to inform climate impacts on the viability of plant populations. Our results suggest that

specimen total reproduction is substantially more buffered from climate effects than plant

phenology, a result which could stem from phenology buffering plant fitness, interactions with

other species, or other mechanisms. Ultimately, this buffering may signal robustness of these

populations in the face of changing precipitation and temperature regimes, but to increase

confidence in estimating climate impacts on plant fitness from herbarium specimens, we

suggest additional collection practices. Our study also contributes to methodological

approaches by characterizing climate tailored to the lifespan of the specimen and estimating

germination timing based on our knowledge of germination cues. In sum, our results highlight

how herbarium specimens can lend more insight into effects of climate variation on plant

performance.

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and specimen sampling protocols, MGB and SJW conducted data analyses in substantial

consultation with all authors; MGB and LCL oversaw specimen reproductive data collection, JAM

contributed to image analysis protocols; SYS wrote the first draft, with contributions from MGB and

SJW. All authors commented substantively on the draft.

Data availability. Upon acceptance all data and code will be publicly archived. All data and code will

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References

Armitage DW. **2024**. To remain modern the coexistence program requires modern statistical rigour. *Nature* **632**: E15–E20.

Barnett KL, Facey SL. **2016**. Grasslands, Invertebrates, and Precipitation: A Review of the Effects of Climate Change. *Frontiers in Plant Science* **7**.

Beatley JC. **1974**. PHENOLOGICAL EVENTS AND THEIR ENVIRONMENTAL TRIGGERS IN MOJAVE-DESERT ECOSYSTEMS. *Ecology* **55**: 856–863.

Blomberg SP, Garland T, Ives AR. **2003**. TESTING FOR PHYLOGENETIC SIGNAL IN COMPARATIVE DATA: BEHAVIORAL TRAITS ARE MORE LABILE. *Evolution* **57**: 717–745.

Bontrager M, Angert AL. **2016**. Effects of range-wide variation in climate and isolation on floral traits and reproductive output of Clarkia pulchella. *American Journal of Botany* **103**: 10–21.

Bürkner P-C. **2017**. brms: An R Package for Bayesian Multilevel Models Using Stan. *Journal of Statistical Software* **80**: 1–28.

Cacho NI, McIntyre PJ, Kliebenstein DJ, Strauss SY. **2021**. Genome size evolution is associated with climate seasonality and glucosinolates, but not life history, soil nutrients or range size, across a clade of mustards. *Annals of Botany* **127**: 887–902.

Cacho NI, Millie Burrell A, Pepper AE, Strauss SY. **2014**. Novel nuclear markers inform the systematics and the evolution of serpentine use in Streptanthus and allies (Thelypodieae, Brassicaceae). *Molecular Phylogenetics and Evolution* **72**: 71–81.

Cacho NI, Strauss SY. **2014**. Occupation of bare habitats, an evolutionary precursor to soil specialization in plants. *Proceedings of the National Academy of Sciences* **111**: 15132–15137.

Calinger KM, Queenborough S, Curtis PS. **2013**. Herbarium specimens reveal the footprint of climate change on flowering trends across north-central North America. *Ecology Letters* **16**: 1037–1044.

Currier CM, Sala OE. **2022**. Precipitation versus temperature as phenology controls in drylands. *Ecology* **103**: e3793.

Dangremond EM, Hill CH, Louaibi S, Muñoz I. **2022**. Phenological responsiveness and fecundity decline near the southern range limit of Trientalis borealis (Primulaceae). *Plant Ecology* **223**: 41–52.

Davis CC, Willis CG, Connolly B, Kelly C, Ellison AM. **2015**. Herbarium records are reliable sources of phenological change driven by climate and provide novel insights into species' phenological cueing mechanisms. *American Journal of Botany* **102**: 1599–1609.

DeLeo VL, Menge DNL, Hanks EM, Juenger TE, Lasky JR. **2020**. Effects of two centuries of global environmental variation on phenology and physiology of *Arabidopsis thaliana*. *Global*

Change Biology **26**: 523–538.

Di Castri F, Mooney HA . **2012**. *Mediterranean Type Ecosystems: Origin and Structure* .
Springer Science & Business Media.

Fabina NS, Abbott KC, Gilman RT . **2010**. Sensitivity of plant–pollinator–herbivore
communities to changes in phenology. *Ecological Modelling* **221**: 453–458.

Farías AA, Armas C, Gaxiola A, Cea AP, Luis Cortés J, López RP, Casanoves F, Holmgren
M, Meserve PL, Gutiérrez JR, *et al.*2021. Species interactions across trophic levels mediate
rainfall effects on dryland vegetation dynamics. *Ecological Monographs* **91**: e01441.

Flint LE, Flint AL . **2014**. California Basin Characterization Model: A dataset of historical and
future hydrologic response to climate change. *USGS data release* **Version 1.1**.

Flint L, Flint A, Thorne J, Boynton R . **2013**. Fine-scale hydrologic modeling for regional
landscape applications: the California Basin Characterization Model development and
performance. *Ecological Processes* **2**: 25.

Freimuth J, Bossdorf O, Scheepens JF, Willems FM . **2022**. Climate warming changes
synchrony of plants and pollinators. *Proceedings of the Royal Society B: Biological Sciences*
289: 20212142.

Gezon ZJ, Inouye DW, Irwin RE . **2016**. Phenological change in a spring ephemeral:
implications for pollination and plant reproduction. *Global Change Biology* **22**: 1779–1793.

González-Megías A, Menéndez R . **2012**. Climate change effects on above- and below-ground
interactions in a dryland ecosystem. *Philosophical Transactions of the Royal Society B:*
Biological Sciences **367**: 3115–3124.

Gremer JR, Chiono A, Suglia E, Bontrager M, Okafor L, Schmitt J . **2020**. Variation in the
seasonal germination niche across an elevational gradient: the role of germination cueing in
current and future climates. *American Journal of Botany* **107**: 350–363.

Hamann E, Wadgymar SM, Anderson JT . **2021**. Costs of reproduction under experimental
climate change across elevations in the perennial forb Boechera stricta. *Proceedings of the*
Royal Society B: Biological Sciences **288**: 20203134.

Hänel S, Tielbörger K . **2015**. Phenotypic response of plants to simulated climate change in a
long-term rain-manipulation experiment: a multi-species study. *Oecologia* **177**: 1015–1024.

Hatchett BJ, Koshkin AL, Guirguis K, Rittger K, Nolin AW, Heggli A, Rhoades AM, East
AE, Siirila-Woodburn ER, Brandt WT, *et al.*2023. Midwinter Dry Spells Amplify Post-Fire
Snowpack Decline. *Geophysical Research Letters* **50**: e2022GL101235.

Hereford J, Schmitt J, Ackerly DD . **2017**. The seasonal climate niche predicts phenology and
distribution of an ephemeral annual plant, Mollugo verticillata. *Journal of Ecology* **105**:
1323–1334.

Jones CA, Daehler CC. **2018**. Herbarium specimens can reveal impacts of climate change on plant phenology; a review of methods and applications. *PeerJ* **6**: e4576.

Keeley JE. **1991**. Seed germination and life history syndromes in the California chaparral. *The Botanical Review* **57**: 81–116.

Kudo G, Ida TY. **2013**. Early onset of spring increases the phenological mismatch between plants and pollinators. *Ecology* **94**: 2311–2320.

Levine JM, McEachern AK, Cowan C. **2008**. Rainfall effects on rare annual plants. *Journal Of Ecology* **96**: 795–806.

Levine JM, McEachern AK, Cowan C. **2011**. Seasonal timing of first rain storms affects rare plant population dynamics. *Ecology* **92**: 2236–2247.

Love NLR, Bonnet P, Goëau H, Joly A, Mazer SJ. **2021**. Machine Learning Undercounts Reproductive Organs on Herbarium Specimens but Accurately Derives Their Quantitative Phenological Status: A Case Study of Streptanthus tortuosus. *Plants* **10**: 2471.

Love NLR, Mazer SJ. **2021**. Region-specific phenological sensitivities and rates of climate warming generate divergent temporal shifts in flowering date across a species' range. *American Journal of Botany* **108**: 1873–1888.

Love NLR, Park IW, Mazer SJ. **2019**. A new phenological metric for use in pheno-climatic models: A case study using herbarium specimens of *Streptanthus tortuosus*. *Applications in Plant Sciences* **7**: e11276.

Lu C, Zhang J, Min X, Chen J, Huang Y, Zhao H, Yan T, Liu X, Wang H, Liu H. **2023**. Contrasting responses of early- and late-season plant phenophases to altered precipitation. *Oikos* **2023**: e09829.

Lüdecke D. **2018**. ggeffects: Tidy Data Frames of Marginal Effects from Regression Models. *Journal of Open Source Software* **3**: 772.

Luković J, Chiang JCH, Blagojević D, Sekulić A. **2021**. A Later Onset of the Rainy Season in California. *Geophysical Research Letters* **48**: e2020GL090350.

Martínez-Berdeja A, Okada M, Cooper MD, Runcie DE, Burghardt LT, Schmitt J. **2023**. Precipitation timing and soil substrate drive phenology and fitness of Arabidopsis thaliana in a Mediterranean environment. *Functional Ecology* **37**: 2471–2487.

Mazer SJ, Love NLR, Park IW, Ramirez-Parada T, Matthews ER. **2021**. PHENOLOGICAL SENSITIVITIES TO CLIMATE ARE SIMILAR IN TWO CLARKIA CONGENERS: INDIRECT EVIDENCE FOR FACILITATION, CONVERGENCE, NICHE CONSERVATISM, OR GENETIC CONSTRAINTS. *Madroño* **68**.

Meineke EK, Davis CC, Davies TJ. **2018**. The unrealized potential of herbaria for global change biology. *Ecological Monographs* **88**: 505–525.

Miller CN, Stuble KL. **2024**. Warm Spring Days are Related to Shorter Durations of
Reproductive Phenophases for Understory Forest Herbs. *Ecology and Evolution* **14**: e70700.
Miller-Rushing AJ, Primack RB, Primack D, Mukunda S. **2006**. Photographs and herbarium
specimens as tools to document phenological changes in response to global warming.
American Journal of Botany **93**: 1667–1674.
Moles AT, Perkins SE, Laffan SW, Flores-Moreno H, Awasthy M, Tindall ML, Sack L,
Pitman A, Kattge J, Aarssen LW, *et al.***2014**. Which is a better predictor of plant traits:
temperature or precipitation? *Journal of Vegetation Science* **25**: 1167–1180.
Niu S, Wan S. **2008**. Warming changes plant competitive hierarchy in a temperate steppe in
northern China. *Journal of Plant Ecology* **1**: 103–110.
Olliff-Yang RL, Ackerly DD. **2021**. LATE PLANTING SHORTENS THE FLOWERING PERIOD
AND REDUCES FECUNDITY IN LASTHENIA CALIFORNICA. *Madroño* **68**: 377–387.
Paradis E, Schliep K. **2019**. ape 5.0: an environment for modern phylogenetics and
evolutionary analyses in R. *Bioinformatics* **35**: 526–528.
Pathak TB, Maskey ML, Dahlberg JA, Kearns F, Bali KM, Zaccaria D. **2018**. Climate
Change Trends and Impacts on California Agriculture: A Detailed Review. *Agronomy* **8**: 25.
Pearse IS, Aguilar JM, Strauss SY. **2020**. Life-History Plasticity and Water-Use Trade-Offs
Associated with Drought Resistance in a Clade of California Jewelflowers. *The American*
Naturalist **195**: 691–704.
Pearse IS, McIntyre P, Cacho NI, Strauss SY. **2022**. Fitness homeostasis across an
experimental water gradient predicts species' geographic range and climatic breadth. *Ecology*
103: e3827.
Pearson KD, Love NLR, Ramirez-Parada T, Mazer SJ, Yost JM. **2021**. PHENOLOGICAL
TRENDS IN THE CALIFORNIA POPPY (ESCHSCHOLZIA CALIFORNICA): DIGITIZED
HERBARIUM SPECIMENS REVEAL INTRASPECIFIC VARIATION IN THE SENSITIVITY OF
FLOWERING DATE TO CLIMATE CHANGE. *Madroño* **68**: 343–359.
Primack D, Imbres C, Primack RB, Miller-Rushing AJ, Del Tredici P. **2004**. Herbarium
specimens demonstrate earlier flowering times in response to warming in Boston. *American*
Journal of Botany **91**: 1260–1264.
Ramirez-Parada TH, Park IW, Record S, Davis CC, Ellison AM, Mazer SJ. **2024**. Plasticity
and not adaptation is the primary source of temperature-mediated variation in flowering
phenology in North America. *Nature Ecology & Evolution* **8**: 467–476.
Raven PH, Axelrod DI. **1978**. *Origin and Relationships of the California Flora*. University of
California Press.
Revell LJ. **2012**. phytools: an R package for phylogenetic comparative biology (and other

things). *Methods in Ecology and Evolution* **3**: 217–223.

Revell LJ, Harmon LJ. **2022**. *Phylogenetic Comparative Methods in R*. Princeton University

Press.

Steyn HM, van Rooyen N, van Rooyen MW, Theron GK. **1996**. The phenology of

Namaqualand ephemeral species. The effect of water stress. *Journal of Arid Environments* **33**:

49–62.

Swain DL, Langenbrunner B, Neelin JD, Hall A. **2018**. Increasing precipitation volatility in

twenty-first-century California. *Nature Climate Change* **8**: 427–433.

Terry JCD. **2024**. Uncertain competition coefficients undermine inferences about coexistence.

Nature **632**: E9–E14.

Tevis Jr. L. **1958**. Germination and Growth of Ephemerals Induced by Sprinkling a Sandy

Desert. *Ecology* **39**: 681–688.

Tiusanen M, Kankaanpää T, Schmidt NM, Roslin T. **2020**. Heated rivalries: Phenological

variation modifies competition for pollinators among arctic plants. *Global Change Biology* **26**:

6313–6325.

Van Dyke MN, Levine JM, Kraft NJB. **2022**. Small rainfall changes drive substantial changes

in plant coexistence. *Nature* **611**: 507–511.

Wade RN, Karley AJ, Johnson SN, Hartley SE. **2017**. Impact of predicted precipitation

scenarios on multitrophic interactions. *Functional Ecology* **31**: 1647–1658.

Ware IM, Van Nuland ME, Yang ZK, Schadt CW, Schweitzer JA, Bailey JK. **2021**.

Climate-driven divergence in plant-microbiome interactions generates range-wide variation in

bud break phenology. *Communications Biology* **4**: 1–9.

Went FW. **1949**. Ecology of Desert Plants. II. The Effect of Rain and Temperature on

Germination and Growth. *Ecology* **30**: 1–13.

Willems FM, Scheepens JF, Bossdorf O. **2022**. Forest wildflowers bloom earlier as Europe

warms: lessons from herbaria and spatial modelling. *New Phytologist* **235**: 52–65.

**Willis CG, Ellwood ER, Primack RB, Davis CC, Pearson KD, Gallinat AS, Yost JM, Nelson

G, Mazer SJ, Rossington NL, *et al.***2017**. Old Plants, New Tricks: Phenological Research

Using Herbarium Specimens. *Trends in Ecology & Evolution* **32**: 531–546.

Worthy SJ, Ashlock SR, Miller A, Maloof JN, Strauss SY, Gremer JR, Schmitt J. **2024a**.

Accelerated phenology fails to buffer fitness loss from delayed rain onset in a clade of

wildflowers. *bioRxiv*: 2024–06.

**Worthy SJ, Miller A, Ashlock SR, Ceviker E, Maloof JN, Strauss SY, Schmitt J, Gremer

JR**. **2024b**. Germination responses to changing rainfall timing reveal potential climate

vulnerability in a clade of wildflowers. *Ecology* **n/a**: e4423.

Yim C, Bellis ES, DeLeo VL, Gamba D, Muscarella R, Lasky JR . **2024**. Climate biogeography of *Arabidopsis thaliana* : Linking distribution models and individual variation. *Journal of Biogeography* **51**: 560–574.

Zettlemoyer MA, Conner RJ, Seaver MM, Waddle E, DeMarche ML . **2024**. A Long-Lived Alpine Perennial Advances Flowering under Warmer Conditions but Not Enough to Maintain Reproductive Success. *The American Naturalist*: E000–E000.

Zhou H, Min X, Chen J, Lu C, Huang Y, Zhang Z, Liu H . **2023**. Climate warming interacts with other global change drivers to influence plant phenology: A meta-analysis of experimental studies. *Ecology Letters* **26**: 1370–1381.

727 Figures

Figure 1. Relationship between collection day-of-year (DOY) and a) phenology and b)

reproductive output. Collection day-of-year was represented in models as the number of

days since September 1 in the year prior to collection until the collection date. In a),

Specimens entirely in bud would have a phenological score of 0, and specimens with only

mature fruits would have a phenological score of 1.Overall, collection day-of-year was

strongly related to phenology, and less so to total reproduction; in each case, there remains

much variation to be explained by climate variables (Fig. S3, this figure).

Figure 2. (a) Effects of precipitation on specimen phenological stage for 14 focal species.
Regression slopes from models that include temperature and collection day-of-year (see Fig 1,
Fig S1). Eight species had significantly more advanced phenology with less precipitation (red
lines), while two species had delayed phenology with less precipitation (blue lines); gray lines
are not significant. Similar plots for temperature and CWD are in Fig S4. (b) Mean slope with
95% confidence interval of the regression of precipitation and temperature (in the same model)
and (c) climatic water deficit on the phenological stage of specimens. The precipitation column
in (b) represents the slopes depicted in (a). None of these responses to climate showed
significant phylogenetic signal (K<1.0, P>0.4 in all cases; Table S3).

Figure 3. (a) Effects of precipitation on total reproductive structures for 14 focal species. Regression slopes are from models that include effects of temperature and collection day-of-year. Five species had significantly less reproduction with more precipitation (red lines), while two species had increased reproduction (blue lines). Plots for temperature and climatic water deficit (CWD) are in Figure S5. (b and c) Summarized effects of (b) precipitation and temperature (in the same model) and (c) CWD on reproductive output. Point-ranges show the mean and 95% confidence interval of the slope of the regression of total reproduction on each climate variable. Grey point ranges indicate that the effects of climate are not different from zero. None of these responses to climate showed significant phylogenetic signal (Table S3).

Figure 4. Effects of the timing of germination-triggering rain events on phenology of herbarium specimens. In (a) red point ranges indicate species for which later estimated germination timing delayed phenology. Grey point ranges show effects not different from zero. In (b) these slopes are plotted over the raw data points; phenological scores range from 0 (entirely in bud) to 1 (entirely mature fruit). Collection day-of-year, total precipitation and average temperature were also included in the model as predictors and these effects are shown in Fig S6.

Figure 5. Effects of timing of germination triggering rainfall based on daily precipitation data on reproductive output of herbarium specimens. Blue symbols/lines show a positive effect of later germination on reproduction and grey lines are not different from zero. Collection date, total precipitation and average temperature were included in the model as predictors (see Figure S7). In (b) these slopes are plotted over the raw data points. There was significant phylogenetic signal in the slopes of reproduction responses to the timing of germination-triggering rainfall (first rain event DOWY; Table S3, Fig. S8).

Supplemental Figures

Figure S1. Elevational and geographic distributions for the 14 *Streptanthus* (s.l.) species in our study. (a) Density ridges of elevational data for the 891 specimens in our dataset with elevations recorded. (b) Locations (points) of the specimens in our dataset, colored by species. Convex hulls are drawn around the specimen locations for each species.

Figure S2. (a) Phenological stages of specimens: these are calculated based on a weighted sum of flowers, immature, and mature fruits, divided by a total count of reproductive structures (including buds). Specimens at a phenological stage of 0 are entirely in bud, those with a phenological stage of 1 have only mature fruits, and intermediate values indicate a combination of buds, flowers, immature fruits, and mature fruits. (b) Specimen collection timing varied within and between species, with the majority of specimens collected in March-June.

Figure S3. Effects of collection day-of-year covariate on phenology and reproduction. These estimates are from models that also included covariates of temperature and precipitation (Figure 3). Blue point ranges indicate species for which specimens collected later in the year had significantly (a) more advanced phenology or (b) greater total reproduction relative to specimens collected earlier in the year. Grey point ranges indicate species for which the effect of collection day-of-year on phenology or reproduction was not different from 0; there were no cases where specimens with later collection day-of-year had more delayed phenology or less reproduction. These effects are also shown plotted over the raw data in Figure 1.

Figure S4. (a) Effects of average temperature on phenological stage for 14 focal species from the *Strepthanthus* clade. One species had significantly less advanced phenology under warm temperatures (red line), while seven species had more advanced phenology under warm temperatures (blue lines). These regression slopes are from models that also include effects of precipitation and collection day-of-year. (b) Effects of total climatic water deficit (CWD) on phenological stage. Four species had significantly more advanced phenology with high CWD (blue lines), while three species had less advanced phenology under high CWD (red lines). Regression slopes are from models that also include effects of collection day-of-year.

Figure S5. (a) Effects of average temperature on total reproductive structures for 14 focal species from the *Strepthanthus* clade. Regression slopes are from models that also include effects of precipitation and collection day-of-year. One species had significantly less reproduction withunder warm temperatures (red line), while one species had increased reproduction under warm temperatures (blue line). (b) Effects of climatic water deficit (CWD) on total reproductive structures. Regression slopes are from models that also include effects of collection day-of-year. Four species had significantly more reproduction with high CWD (blue lines), while two species had decreased reproduction with high CWD (red lines).

Figure S6. Summarized effects of collection day-of-year, timing of first rain event, precipitation, and temperature on specimen phenological scores. Point-ranges show the mean and 95% confidence interval of the slope of the regression of phenological score on each variable. Blue point-ranges indicate that higher values of the variable advanced phenology, red point-ranges indicate that higher values of the variable delayed phenology, and grey point ranges indicate where effects are not different from zero. These results are from a model using only specimens for which daily precipitation data was available—a smaller dataset than we used in our primary analysis of temperature and precipitation effects.

Figure S7. Summarized effects of collection day-of-year, timing of first rain event, precipitation, and temperature on the number of reproductive structures on each specimen. Point-ranges show the mean and 95% confidence interval of the slope of the regression of reproduction on each variable. Blue point-ranges indicate that higher values of the variable are associated with increased reproduction, red point-ranges indicate that higher values of the variable are associated with decreased reproduction, and grey point ranges indicate where effects are not different from zero. These results are from a model using only specimens for which daily precipitation data was available—a smaller dataset than we used in our primary analysis of temperature and precipitation effects.

Figure S8. Scatterplots relating how species responded phenologically to climate versus how they responded in total reproduction to the same to climate variables; these relationships are asking whether there is any systematic across-species correlation in how species respond phenologically and reproductively that might indicate evolutionary constraint. : (a) temperature, (b) total precipitation, (c) total climatic water deficit (CWD), and (d) the day-of-year of the first rainfall event. Slopes in (a) are also shown in figures 2b, S4a, 3b and S5a. Slopes in (b) are also shown in figures 2ab and 3ab. Slopes in (c) are also shown in figures 2c, S4b, 3c and S5b. Slopes in (d) are also shown in figures 4 and 5. We found no significant association between phenological and reproductive responses for these climate variables (Table S2).

841 **Supplemental tables**

842 **Table S1.** Categorization rules of reproductive structures on herbarium specimens.

Category	Description
Buds	unopened flowers with no petals visible
Inferred buds	contextual placement indicates a bud (i.e., buds are above and below), but only the pedicel remains
Flowers	anthers are visible, petals are still attached
Inferred flowers	contextual placement indicates a flower, but only the pedicel remains
Immature fruits	styles which have not yet widened and elongated with seeds; these occur in the section of the stem below flowers
Inferred immature fruits	contextual placement indicates an immature fruit, but only the pedicel remains
Filled fruit	fruit has elongated and swelling from seeds may be visible
Unfilled fruits	fruits that have not elongated or swollen with seeds; morphologically similar to immature fruits but distinguished by their context (filled fruits higher on the stem indicate that these have had time to mature
Inferred fruits	contextual placement indicates either a filled or unfilled fruit, but only the pedicel remains
Unknown reproductive structure	a pedicel is present, but no structure is present, and there is not adequate context to assume what type of structure it was

844 **Table S2.** Results from phylogenetically independent contrasts between phenological and

845 reproductive response slopes for four climate variables.

Variable	Estimate	SE	t-value	P-value
Temperature	0.091	0.29	0.31	0.76
Precipitation	0.22	0.34	0.66	0.52
Climatic water deficit	-0.19	0.31	-0.61	0.55
First rain event DOWY	-0.021	0.25	-0.085	0.93

Table S3. Tests for phylogenetic signal in the slopes of regressions of reproduction and

phenology to climate and collection date. We report Blomberg's K and estimate the significance

of phylogenetic signal using 10,000 simulations in a randomization test—this significance is

reported in the P-value column. When P < 0.05, we go on to test whether the estimate of K was

significantly greater than one by comparing our estimate of K to a null distribution of 10,000 K

values generated under Brownian motion. Values of K significantly greater than one indicate

that species tend to have more similar phenology and reproductive output responses to climate

than expected based on a Brownian motion model of evolution, i.e. more evolutionary

constraint. DOWY = day of water year (days counted from September 1). All models also

included a random effect of collection location (see methods).

Variable	K	P-value	Notes
Model: Reproduction ~ average temperature + log(total precipitation) + collection DOWY			
Average temperature	0.895293	1	
log(Total precipitation)	0.885941	0.4807	
Model: Phenological stage ~ average temperature + log(total precipitation) + collection DOWY			
Average temperature	0.797422	0.8456	
log(Total precipitation)	0.859549	0.7453	
Model: Reproduction ~ total climatic water deficit (CWD) + collection DOWY			
Total CWD	0.811949	0.9147	
Model: Phenological stage ~ total climatic water deficit (CWD) + collection DOWY			
Total CWD	0.961103	0.2662	
Model: Reproduction ~ first rain event DOWY + average temperature + log(total precipitation) + collection DOWY			
First rain event DOWY	**1.96889**	**0.0001**	**Tested if significantly >1 by comparison to null distribution. P-value < 0.001, so significantly greater than 1.**
Average temperature	0.78497	0.1032	
log(Total precipitation)	0.897138	0.573	
Model: Phenological stage ~ first rain event DOWY + average temperature + log(total precipitation) + collection DOWY			
First rain event DOWY	**1.31321**	**0.0252**	**Tested if significantly >1 by comparison to null distribution. P-value = 0.0216, so significantly greater than 1.**
Average temperature	1.06296	0.433	
log(Total precipitation)	1.0037	0.2951	

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