Title: Archaeal lineages related to eukaryotes encode functional diterpenoid cyclases.
Short title: Diterpenoid cyclases from Asgardarchaeota.

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Abstract

The first eukaryotic cell originated through the union of an archaeon and a bacterium. Phylogenetic
evidence suggests that the archaeal partner was a member of phylum Asgardarchaeota, likely
within the Heimdallarchaeia clade. Although little is known about the molecular basis of
eukaryogenesis, insight can be gained from studying the unique biochemistry of modern
Asgardarchaeota. Here, we investigated the potential for Asgards to produce cyclic terpenoids.
Phylogenomics coupled with structural prediction revealed that organisms from the clades
Hodarchaeales and Kariarchaeaceae encode for diterpenoid cyclases. We tested the functionality
of these enzymes *in vitro*, showing that they cyclize geranylgeranyl pyrophosphate to form bicyclic
halimadienyl pyrophosphate. Halimadienyl nucleosides have previously been shown to mediate
intracellular persistence of *Mycobacterium tuberculosis* in host endosomes, and may function
similarly in Asgardarchaeota. The characterization of Asgard diterpenoid cyclases provides insight
into the biochemistry of an elusive group of microbes that are pivotal in the evolution of complex
cellular life.

15
16 **Main Text**
17
The discovery and characterization of the Asgardarchaeota (written here as "Asgards," also known
as Promethearchaeati [1]) revolutionized our understanding of the origin of eukaryotes and
eukaryotic cell biology [2,3]. Phylogenetic analysis has shown that Asgards are likely the closest
archaeal relatives of eukaryotes [2,4], supporting the hypothesis that an Asgard ancestor was a
partner in the endosymbiosis [5–7] from which eukaryotic life arose. Analysis of modern Asgard
genomes suggests that these organisms have some archaeal-like physiological features such as
isoprenoidal membranes [8], as well as features otherwise only known in eukaryotes such as
membrane trafficking proteins and a dynamic cytoskeleton [9–11]. However, relatively little study
has been devoted to Asgard proteins that are not ubiquitous in eukaryotes, despite the potential of
these proteins to reveal unique Asgard biology, including processes that may have been important
in eukaryogenesis.

In pursuit of such non-universal proteins, we identified 64 putative diterpenoid cyclase
genes in archaeal genomes [15] and metagenome-assembled genomes (MAGs), including 31 from

Asgards. All but three Asgard sequences are from Heimdallarchaeia, the clade thought to contain
the Asgard ancestor of eukaryotes [4], and most are from hydrothermal vents (see Supplementary
Methods and Table S1). Four are from organisms which have recently been isolated in co-culture
[14]. All archaea, including Asgards, produce the diterpenoid cyclase substrate, geranylgeranyl
pyrophosphate (GGPP), in the course of archaeal membrane lipid biosynthesis [12]. The polycyclic
products of diterpenoid cyclases range in function from communication (e.g. gibberellins in plants
and their symbionts) to conflict (e.g. platencin and terpentecin in soil bacteria) [13]. The relevance
of these functions to Asgard symbiosis, especially the eukaryogenetic symbiosis, motivated us to
determine the biosynthetic products of Asgard diterpenoid cyclases.

To determine the function of putative Asgard cyclases, we performed *in vitro* experiments
with four Asgard proteins found in our initial search. As a control, we also tested the *Mycobacterium*
tuberculosis H37Rv homolog, Rv3377c, which had been previously shown to produce halimadienyl
pyrophosphate [16]. The Asgard cyclases include two homologs from *Candidatus* Hodarchaeales
archaeon S146_22 (WAPZ01000470.1 and WAPZ01000551.1), one (OLS20947.1) from
Candidatus Heimdallarchaeota archaeon LC2 (assigned to Kariarchaeaceae by the Genome
Taxonomy Database), and one from *Candidatus* Lokiarchaeales archaeon CR 4 (OLS12628.1).
These proteins have the acidic DxD motif [17] necessary to catalyze terpenoid cyclization (Fig. S1),
a hydrogen bond donating residue at position H341 (corresponding to sequence position in
Rv3377c), which positions the general acid for proton transfer, and a bulky aromatic residue at
position W380 to protect the general acid when substrate is not bound.. We incubated the purified
protein (Fig. S2) with geranylgeranyl pyrophosphate (GGPP; **1'**) and analyzed the reaction products
by gas chromatography-mass spectrometry (GC-MS). We found that two of the Asgard cyclases
(OLS20947.1 from *Candidatus* Heimdallarchaeota archaeon LC2 and WAPZ01000551.1 from
Candidatus Hodarchaeales archaeon S146_22) convert GGPP to halimadienyl pyrophosphate (**2'**)
(Fig. 1).

Phylogenetic analysis (Fig. 2) revealed that Asgard sequences are monophyletic and sister
diterpenoid cyclases from plant chloroplasts. The chloroplast clade has several bacterial
sequences from the Candidate Phyla Radiation (CPR) on its stem. All sequences in the Asgard-
dominated clade are from Heimdallarchaeia except for one from Lokiarchaeales and one from a
bacterium. Sister to Asgard and plant sequences is a clade of archaeal sequences from
Thermoplasmata, Theionarchaea, and Methanofastidiosia groups of superphylum
Methanobacteriati (also known as Euryarchaeota), with lone sequences from the superphylum
Thermoproteati (TACK) taxa Nitrososphaerota and Bathyarchaeia, two sequences from the Asgard
phylum Thorarchaeota, and two from bacteria. Outside the archaeal-plant group, paraphyletic
bacterial groups are dominated by Chloroflexota, particularly sequences from sponge symbiont
MAGs, suggesting a possible bacterial source for sponge cyclic diterpenoids [18]. Among these
groups are clusters of Actinomycetota sequences, including one which contains the characterized
[16] halimadienyl cyclase Rv3377c from *M. tuberculosis*. The deepest-branching clade is large and
composed primarily of bacterial sequences from Actinomycetota, including characterized proteins
which synthesize precursors to gibberellins and terpenoid antibiotics [19,20]. This phylogeny shows
that unlike the distantly related sterol and hopanoid cyclases, which are near-ubiquitous and
perhaps ancestral in eukaryotes and bacteria respectively, diterpenoid cyclases are quite
taxonomically restricted, but often widespread within those taxa.

Halimadienyl cyclases homologs are prevalent in Heimdallarchaeia (Hodarchaeales and
Kariarchaeaceae) but rare in other Asgards. Heimdall cyclases could have been acquired vertically
from a common ancestor with Methanobacteriati, with gene loss in other Asgard phyla, or via
horizontal transfer on the Heimdall stem. The position of chloroplast cyclases sister to, rather than
nested within, those from Asgards, suggests that these eukaryotes did not inherit their diterpenoid
cyclases from Asgards. However, the position of plant sequences in the cyclase tree and that of
eukaryotes within the Asgard genome tree may change as additional Asgards are sequenced.
Regardless of evolutionary history, the enrichment of diterpenoid cyclases in Heimdallarchaeia may

suggest a function for these enzymes that is unique to Heimdall biology, and that may have been
shared by the Asgard ancestor of eukaryotes.

The biological role of halimadienyl compounds in Asgards is unknown. The only other
microbe known to synthesize halimanes is *M. tuberculosis*, an intracellular bacterial pathogen of
mammalian macrophages. *M. tuberculosis* constitutively secretes 1-halimadienyl-adenosine into
the host endosome, where it acts as a Lewis base to neutralize endosomal pH and halt maturation
of the endosome into a lysosome, facilitating survival and persistence of the bacterium [21]. In
addition to the cyclase, 1-halimadienyl-adenosine biosynthesis requires an adenosyl transferase
[22], which is also found in the genome of *Candidatus* Hodarchaeales S146_22. If this organism
can synthesize the functional antacid, it could stabilize endosomal persistence of the Asgard in a
host cell, or of symbionts acquired through an endomembrane trafficking system [3] in the Asgard
cell. Testing these hypotheses will require characterizing the S146_22 adenosyl transferase
homolog and its product, and ultimately (co-)culturing this Asgard to determine whether it can
participate in endosymbiosis. Currently, endosymbiosis has not been observed in modern Asgards
despite their ancestors' likely role in the eukaryogenetic endosymbiosis. However, the biology of
halimadienyl cyclases in Asgards, as well as that of other Asgard proteins not widespread in
eukaryotes, holds great promise for understanding their unique biology and their potential role in
cellular evolution.

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H.S.M.: conceptualization, data curation, investigation, methodology, formal analysis,
visualization, writing – original draft, writing – review & editing; V.D.A.: conceptualization,
investigation, writing – review & editing; J.J.B.: conceptualization, writing – reviewing & editing;
P.V.W.: conceptualization, supervision, writing – reviewing & editing; B.J.B.: conceptualization,
supervision, writing – reviewing & editing.

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190
191

Figures

A

1 R = OH
1' R = PO$_4$(PO$_3$)$^{2-}$
Geranylgeranyl pyrophosphate

2 R = OH
2' R = PO$_4$(PO$_3$)$^{2-}$
Halimadienyl pyrophosphate

B

1

No enzyme

2

Rv3377c *Mycobacterium tuberculosis* halimadienyl cyclase

1

2

OLS20947.1 *Candidatus* Heimdallarchaeota archaeon LC2

1

2

WAPZ01000551.1 *Candidatus* Hodarchaeales archaeon S146 22

20.8   21.0   21.2   21.4

Retention time (minutes)

193
Figure 2. Asgard cyclases catalyze the cyclization of geranylgeranyl pyrophosphate (GGPP)
to form halimadienyl pyrophosphate. (*A*) Cyclization reaction and dephosphorylation products,
identified by comparison to published gas chromatography-mass spectrometry (GC-MS) spectra
(Fig. S3). (*B*) Extracted ion chromatograms (*m/z* 119, 189, 191, 290, 362) of *in vitro* reactions
performed with GGPP and cyclases. Reaction products were dephosphorylated by incubation with
phosphatase, extracted with hexanes, and trimethylsilylated before GC-MS. The alcohols

geranylgeraniol (**1**) and halimadienol (**2**) were only detected after dephosphorylation, suggesting
that the cyclization product is halimadienyl pyrophosphate (**2'**).

Actinobacterial diterpenoid cyclases (n=206)

Fungal and Cyanobacterial homologs (n=28)

Proteobacteria, Cyanobacteria, and Chloroflexota homologs (n=43)

Diterpenoid cyclase homologs
from Chloroflexota (n=274),
Actinomycetota (n=64,
including 3 from *Mycobacterium*),
and other taxa (n=11)

Mycobacterium tuberculosis
halimadienyl cyclase
(Rv3377c)

Archaeal homologs (n=37)

Candidate phyla radiation
homologs

Chloroplast homologs (n=115)

Heimdall.

WAPZ01000470.1 *Candidatus* Hodarchaeales archaeon S146 22

Heimdall.

Hod.

Heimdall.

WAPZ01000551.1 *Candidatus* Hodarchaeales archaeon S146 22

Heimdall.

Chryseobacterium wanjuense

Heimdall.

Archaeon

Heimdall.

Heimdall.

Heimdall.

Heimdall.

Heimdall.

Archaeon

Heimdall.

OLS20947.1 *Candidatus* Heimdallarchaeota archaeon LC2

Heimdall.

Hod.

Hod.

Candidatus Flexarchaeum multiprotrusionis SC1

Kari.

Candidatus Margulisarchaeum peptidophila HC1

Candidatus Margulisarchaeum peptidophila HC1

Candidatus Margulisarchaeum peptidophila HC1

OLS12628.1 *Candidatus* Lokiarchaeales archaeon CR 4

Hod.

Heimdall.

Heimdall.

Eukaryotes
Bacteria
Archaea
Asgard
Function tested

1 substitution per site

Figure 1. Asgard cyclase homologs are nested within the diterpenoid cyclases. The maximum likelihood phylogeny was rooted by treating all non-diterpenoid cyclases (e.g., sterol and hopanoid cyclases) as an outgroup (see [23]). Nodes are marked with ultrafast bootstrap support. Enzymes tested in this study are marked with stars, which are filled for those that were functional. Asgard homologs not tested are marked with abbreviations indicating Asgard phylum, where Heimdall. = Heimdallarchaeales, Hod. = Hodarchaeales, Kari = Kariarchaeaceae. Supplemental Data 1 contains this tree in Newick format.