***Warming up to a new coat*: moulting king penguins exhibit hyperthermia and increased**

peripheral heat loss

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Abstract

Penguins undergo catastrophic moult, which causes increases in metabolic rate, insulation, and heat loss via thermal windows. Here, we investigated the timing of these physiological changes to test two hypotheses. Under the *thermal challenge* hypothesis, we expected an initial rise in heat dissipation effort (i.e., increased peripheral vasomotion) followed by a rise in core temperature, indicating a failed attempt to prevent hyperthermia. Under the *warm-up* hypothesis, we expected an initial rise of core temperature concomitant or followed by an increase of peripheral vasomotion, indicating regulated hyperthermia, possibly to accelerate feather development. We tested these hypotheses using simultaneous measurements of core and surface temperatures in captive king penguins (*Aptenodytes patagonicus*). Core and surface temperatures increased drastically but concomitantly during moult, providing tentative support for the *warm-up* hypothesis. Moulting penguins were never seen panting, suggesting that increasing peripheral heat loss was sufficient to regulate moulting-induced hyperthermia. Core and subcutaneous temperatures in wild individuals resembled patterns measured in captivity, despite lower heat load and additional options for behavioural thermoregulation. Together, these results indicate that hyperthermia is prevalent during moulting in king penguins, which raises questions regarding the ability of wild penguins to properly thermoregulate during moult under a warming climate.

Introduction

Maintaining thermal insulation is essential for endotherms to thrive in their natural environments [1]. Fur and feathers contribute to thermal insulation in most mammals and birds, but these keratinaceous structures degrade through wear and breakage as they age. Thus, the maintenance of proper insulation from fur or feathers through time requires their replacement through shedding: a protracted, year-round replacement, or moulting: a contracted, punctuated replacement [1]. Feathers are unique among morphological traits due to the variety of functions they serve, with major roles in crypsis, mate choice, species signalling, thermoregulation,

waterproofing, flight, and protection [2]. Moulting is thus considered a key life-history stage

because it conditions the ability of individuals to survive and reproduce in their environment [3].

Moult has been well characterized in various avian models from a metabolic perspective.

Moult requires a massive amount of protein because feathers are composed of approximately

90% protein and can represent or even exceed 10% of body mass and 25% of total body protein

content [4]. Synthesizing new feathers also requires an increase in energy expenditure, with

moulting birds generally exhibiting a metabolic rate *ca.* 9-111% higher than non-moulting

individuals [4]. From a thermoregulatory perspective, moult has been less well characterized to

the best of our knowledge. The few studies available in birds suggest that an increase of *ca.* 20-

60% of thermal conductance occurs during moult [5,6], which may be explained by a decrease in

insulation and/or an increase in evaporative heat transfer due to increased body-water turnover

[4]. Increases in thermal conductance have been shown to raise the lower critical temperature in

long-eared owls (*Asio otus*) from approximately 18°C to 24°C [7] and in emperor penguins

(*Aptenodytes forsteri*) from approximately -10°C to 0°C [8]. Thus, moult has mostly been

considered thermally challenging under cold environmental conditions. Alternatively, it has been

hypothesized that the increased metabolism of moulting birds could more than compensate for

the decrease in insulation, and thus that the increased heat generation may pose a challenge for

moulting birds [4,9,10].

Penguins have specialized vascular arrangements that provide countercurrent heat

exchange in the legs [11], head [12], and wings [13]. Penguins also boast efficient insulation

through a layer of subcutaneous fat [14], and specialized plumage comprised of an insulating

underlayer and a waterproof overlayer [8,15]. Despite penguins' adaptations for heat retention

underwater, they must breed and moult on land, where dissipating heat is more difficult than in

water (thermal conductance is 2.2-4.8 times higher in water than in air; [16]). Penguins undergo

a catastrophic moult once a year, replacing their entire plumage extremely quickly (between 13

and 34 days; [17]), which involves an increase of metabolic rate (and thus of heat production) of

ca. 30-50% [18,19]. Catastrophic moult involves the temporary overlap of new and old feather

layers [20], which increases thermal insulation [10]. Penguins seem relatively sensitive to heat

stress while on land [21–23], and climate change is pronounced in polar and sub-polar regions

[24,25], where many penguin species moult on land during the warmest season [26]. Thus, if heat

dissipation is indeed a challenge during moult [4,9,10], the increasing temperatures penguins are

facing while moulting on land may challenge these iconic species.

Captive Gentoo penguins held within their thermoneutral zone had increased thermal

insulation and heat dissipation effort during the overlap of old and new feather layers [10].

Therefore, moulting seems to represent a challenge for heat dissipation in Gentoo penguins, but

the efficiency or inefficiency of their thermoregulatory response remains unknown because core

body temperature was not measured. We define the *thermal challenge* hypothesis as the

scenario where core body temperature rises because of failure to dissipate heat. Alternatively, it

has been suggested that birds may elevate core body temperature while moulting, hypothetically

under physiological control, to favour feather growth [3], an idea we are referring to as the *warm-*

up hypothesis. To test these hypotheses, we simultaneously recorded panting behaviour, heat

dissipation effort (*i.e.* surface temperature of thermal windows such as flippers, bill and feet),

insulation (*i.e.* surface temperature of the trunk) and core body temperature in captive king

penguins before, during and after the moult. To address ecological relevance of the results

obtained in captivity, we also include behavioural and physiological data of wild king penguins,

although limited in sample size.

If heat dissipation effort increases but core temperature does not, this would indicate that

heat dissipation capacity is sufficient to offset the increased metabolism [4] and insulation [10]

that are known to occur during moult. If heat dissipation effort increases and body temperature

increases afterwards, this would indicate a failure to regulate body temperature, thus supporting

the *thermal challenge* hypothesis. Under the thermal challenge hypothesis, we predict that wild

king penguins will be less likely to exhibit hyperthermia because environmental conditions are

colder in the field and because wild king penguins have additional methods to cool themselves

(*e.g.,* immersing feet in cold water). If core body temperature increases before heat dissipation

effort, or if both increase simultaneously, this would indicate a regulated increase in core body

temperature, thus supporting the *warm-up* hypothesis. Under the *warm-up* hypothesis, we

predict that both captive and wild king penguins will exhibit hyperthermia despite environmental

and behavioural variation.

Materials and Methods

Captive king penguins

We studied 8 captive adult king penguins (*Aptenodytes patagonicus*) in the spring of 2024 at Océanopolis aquarium, Brest, France. The birds were individually identified by their coloured plastic band, and six individuals moulted during the study period. Individuals were maintained indoors at 8.12 ± 0.07°C (mean ± SE), well within their estimated thermoneutral zone (*i.e.* -5 to +22 °C; [27]). They had permanent access to free water to swim/dive, but no possibility to only immerse their feet, and there were consistently scheduled visits from researchers and keepers. Researchers were the first to enter the enclosure after a full night with no disturbance to collect thermal images at 8:00 AM, and keepers entered afterwards throughout the day for regular enclosure cleanings, feeding non-moulting birds (*i.e.* moulting birds are naturally fasting and stay on land even in captivity). The lighting program used in the enclosure includes a monthly variation in artificial light, with exposure varying between 11 and 13 h of light per day during the study. Following the 7-stage moult categorization [10], we assigned moult stage each day for each individual from uniform old plumage (M1) to uniform new plumage (M7) before surface temperature data extraction. To investigate the continuous changes that occur within the visibly distinct moult stages, we tracked the moulting process numerically by assigning a moult day for each bird, which began at day 0 when we visually detected changes in the plumage structure (*i.e.* the start of M2, [10]). Moult actually starts before any visual changes occur (*i.e.* feather growth below the skin), which occurs at sea in wild penguins and has been estimated to last *ca.* 10 days [20].

Individuals were fed with an ingestible core temperature (T_{core}) data logger (Bodycap, Anipill, France, sampling rate = 5min) hidden in their normal diet (*i.e.* without handling) when they began exhibiting hyperphagia, which precedes the fasting period during catastrophic moult [28]. Individuals were equipped at this stage because retention time in the stomach is quite variable (*ca.* 7 to 80 days in wild king penguins, *pers. obs.*). Five moulting individuals were successfully equipped before the onset of visible plumage change, and one at the time of early plumage change (*i.e.* early M2 phase, [10]). Only one (non-moulting) individual lost the logger

before the completion of the study. Moulting individuals were equipped on average 16.5 ± 4.8

days before visible changes in plumage occurred. Data were downloaded remotely with an

Anilogger® monitor placed inside the enclosure throughout the study.

We collected thermal images in the mornings at 8:00 AM (*i.e.* 30 minutes after lights

turned on, and before any keepers had entered the enclosure). Upon entering the enclosure, the

presence/absence of panting behaviour was recorded for each individual [22], and we used a

thermal imaging camera (three different cameras were used due to technical issues: E96, E8, or

T640; resolutions provided in Electronic Supplementary Materials, *ESM*) to collect images of each

individual from a distance of 1 m [10]. We collected images of birds in the profile orientation [29].

Air temperature (T_a) and relative humidity (RH) inside the enclosure were measured using a

weather station (Kestrel 5400; Kestrel® instrument, United Kingdom). Thermal imaging collection

started on average 23.3 ± 5.4 days before visible changes in plumage occurred. Pictures of

individuals being wet at the time of thermal imaging were excluded from the analysis (*i.e.* n = 36

images from non-moulting individuals, or moulting ones with moult day > 20). This approach

resulted in a sample size of 291 datapoints with body surface and core temperatures measured

simultaneously.

After collecting thermal images, we extracted surface temperatures using FLIR Thermal

Studio Pro. We set emissivity to 0.98 [10] and we specified T_a and RH for each image using

measurements collected during each thermal imaging session. We then measured the average

surface temperatures of the bill, flipper, and foot (T_{bill}, $T_{flipper}$, and T_{foot}, respectively) by drawing

separate region of interest (ROI) polygons for each area. We measured the maximum surface

temperature of the periorbital region (henceforth T_{eye}; [30]) using an ellipse. We measured

ground temperature (T_{ground}) below the bird using a line segment the length of the foot. To

determine a representative trunk surface temperature (T_{trunk}) despite irregular feather loss, we

measured the temperature of both the old and newly grown plumage (T_{old} and T_{new}) using a

square with side length equal to ~one third the length of the flipper. We used visual landmarks to

measure T_{ground}, T_{old}, and T_{new} because image resolution varied between the three cameras. After

standardizing between the cameras (see *ESM*), we used values of T_{old} and T_{new} to calculate T_{trunk}

according to [10].

Data analysis was conducted in R Version 4.3.3 [31]. We first calculated wet-bulb

temperature (T_w) in R package HeatStress (https://rdrr.io/github/anacv/HeatStress/) using the

measurements of T_a and RH collected while sampling. Following guidance from two external (*i.e.,*

non-author) experts in the field of thermography, we standardized measurements made with the

three cameras using two approaches (*i.e.* see ESM S0 for details). We then modelled T_{core}, T_{eye},

T_{bill}, $T_{flipper}$, T_{foot}, and T_{trunk} as a function of moult day. To capture the non-linear changes of body

temperatures over the course of the moult (in moulting birds only), we created segmented

models based on calculated changes in slope (*i.e.,* breakpoints; [32]). We determined the

presence/timing of breakpoints and the associated 95% intervals using the R package segmented

(https://cran.r-project.org/web/packages/segmented/index.html). After identifying breakpoints,

we fitted separate linear mixed models ((lmer()) in R package lme4 [33]) for each time period as

determined by the breakpoints for moulting birds. Although thermal conditions were held

relatively constant (Fig. S1), we included T_w as a fixed effect in all global models to account for any

subtle variation in thermal conditions; we used T_{ground} instead of T_w for the models of foot

temperature according to [10]. Non-moulting birds were analysed in separate mixed models and

were assigned an arbitrary moulting day for graphical purpose only (Fig. 1). We included the bird's

assigned identification number (bird ID) as a random intercept in all models to account for

repeated measures. Model support was determined using AICc [34] using the function dredge()

in R package MuMIn (https://CRAN.R-project.org/package=MuMIn). We report the top-ranked

model with parameter estimates for moult day and T_w or T_{ground} (Table S1). We assessed the fit of

the top models by calculating the conditional Nakagawa's R^2 using function r.squaredGLMM() in

R package MuMIn. We visually represented findings from our models by plotting model

predictions obtained from package ggeffects [35] along with raw data for each response variable

using package ggplot2 [36]. Finally, predicted values and the associated 95% confidence intervals

were extracted from key stages based on breakpoint (*i.e.* pre-moult, peak-moult, post-moult) as

well as non-moulting individuals using semi-parametric bootstrapping (n = 1000 simulations) with

confint() and bootMer() functions of R package lme4 [33]).

Wild king penguins

Wild king penguins were studied in 2023-24 (as part of a long-term observatory) in the
'Baie du Marin' colony on Possession Island [37], Crozet Archipelago, in the Southern Indian
Ocean (46°26' S, 51°52' E) from courtship in November 2023 to the 17th of October 2024, when
all scientific activities were shut down because of an outbreak of highly pathogenic avian
influenza (HPAI) at the study site [38]. At breeding initiation (Nov-Dec 2023), 48 breeding pairs
were temporarily marked with hair dye on the breast feathers (*i.e.* lasting until the post-breeding
moult) and permanently marked with thermosensitive pit-tags (BioTherm13 tag, Biomark, USA)
enabling long-term individual identification and subcutaneous temperature measurements [22].
Subcutaneous temperature measurements were conducted during incubation (3rd day of an
incubation shift on land) as part of other long-term monitoring procedures. A subsample of birds
was equipped with core temperature ingestible loggers (Bodycap, Anipill, France, sampling rate =
5 or 15 min) during incubation. At the time of post-breeding moult (Sep-Oct 2024), the colony
was searched daily to identify marked penguins being back on land to moult (N = 43 penguins
observed 1 to 10 times during moult), scoring their moult according to ([10]; M1 to M7 stages),
noting the presence/absence of panting behaviour [22] and the usage of the river as a potentially
cooling substrate (*i.e.* feet in the river: yes/no). Approximately on their 3rd day on land, 3 moulting
individuals (*i.e.* stage M1 for two individuals and stage M2 for another one) were recaptured, had
their subcutaneous temperature recorded and were equipped with a core temperature ingestible
logger. At moult completion (stage M7), individuals were re-captured to download core body
temperature data, which was unfortunately only possible for one individual before field site
closure linked to HPAI outbreak. Considering the limited sample size, no statistical tests were
performed and individual data points are presented. To quantify the environmental heat load
faced by wild *vs.* captive individuals (*i.e.* including solar radiation and wind speed in addition to
air temperature and humidity used for T_w in captivity), Wet Bulb Globe Temperature (WBGT)
values were computed using the R package HeatStress function wbgt.Liljegren(). Environmental
heat load in the wild was lower during moult than breeding, and captive penguins experienced
an environmental heat load being intermediate between moulting and breeding individuals in the
wild (Fig. S1).

Results

Captive penguins

Moulting and non-moulting birds were never observed panting during the study. The

visible part of moult lasted on average 24.3 ± 0.9 days in captive individuals.

Figure 1. Variation in core (a) and surface temperatures (b: eye, c: bill, d: flipper, e: foot and f: trunk) of captive king penguins in relation to moult dynamics. The dashed line at day 0 indicates the first visible signs of moult, and the one at *ca.* day 24 indicates the completion of plumage renewal. Coloured circles

and solid lines represent moulting birds, whereas white circles and dashed lines represent non-moulting

birds. Lines are predictions from the top ranked models (Table S1). n = 6 moulting and n = 2 non-moulting

individuals.

Changes in body temperatures during moult followed a non-linear pattern (Fig. 1). While

T_{core} was best fitted with 2 breakpoints (*i.e.* an increase followed by a decrease during moult and

then stabilisation post-moult, Fig 1a, Table S1), all surface temperatures fitted best with at least

3 breakpoints that corresponded to 3 main phases: pre-moult, moult (with 2 segments

surrounding peak-moult: early-moult and late-moult) and post-moult (Fig. 1b to 1f). While surface

temperatures (T_{eye}, T_{bill}, $T_{flipper}$, T_{foot} and T_{trunk}) during pre- and post-moult periods were not always

constant, the magnitude of changes during these periods were limited (Fig. 1 and Table S1).

During moult, T_{eye}, T_{bill}, $T_{flipper}$ and T_{foot} initially increased during early-moult (although non-

significantly for T_{foot}) and then decreased during late-moult, so post-moult levels were

comparable to pre-moult and non-moulting levels (Fig. 1b to 1e; Fig 2b to 2e; Table S1).

Conversely, T_{trunk} (*i.e.* an inverse proxy of insulation provided by the plumage) initially decreased

and then increased back to levels comparable to pre-moult and non-moulting levels (Fig. 1f and

2f, Table S1).

Figure 2. Predicted core (a) and surface body temperatures (b: eye, c: bill, d: flipper, e: foot and f: trunk)

of captive king penguins at key stages during moult (*i.e.* pre-moult, peak-moult and post-moult based

on breakpoint analyses) and in non-moulting individuals. Error bars represent 95% CI; n = 2-6 moulting

and n = 2 non-moulting individuals.

Body temperatures were mostly indistinguishable between pre-moult, post-moult and
non-moulting individuals (Fig. 2). Yet, the changes in body temperatures during moult were
drastic, with an increase at peak moult of *ca.* +1.4°C for T_{core} (Fig. 2a), +4.0°C for T_{eye} (Fig. 2b),
+11.5°C for T_{bill} (Fig. 2c), +10.2°C for $T_{flipper}$ (Fig. 2d), +5.0°C for T_{foot} although 95% CI overlapped
with most other stages (Fig. 2e), and a decrease of *ca.* -5.0°C for T_{trunk} (Fig. 2f).

Figure 3. Temporal dynamics of changes in body temperatures during moult in captive king penguins
based on breakpoint analyses. The dashed line at day 0 indicates the first visible signs of moult, and the
one at *ca.* day 24 indicates the visual completion of plumage renewal. Error bars represent 95% CI, N = 6
captive individuals.

Considering the timing of changes in body temperatures, it is important to note that we
were unable to detect a pre-moulting stable period for T_{core} (Fig. 1a), and thus a proper moult
start, despite birds being equipped with T_{core} logger on average 16.5 ± 4.8 days before visible
changes in plumage occurred (*i.e.* moult day 0). For body surface temperatures, the initial rise
(*i.e.* moult start) in T_{eye}, T_{bill}, $T_{flipper}$ and T_{foot} occurred simultaneously around 10 days before moult
day 0 (Fig. 3). The initial drop in T_{trunk} occurred slightly later (*ca.* moult day -6), but not significantly
so considering 95% CI overlapped (Fig. 3). The peak in $T_{flipper}$ was reached approximately 10 days
earlier than the peaks in T_{core}, T_{eye}, T_{bill}, and T_{foot}, while the peak in plumage insulation (*i.e.* lowest
T_{trunk}) occurred approximately 5 days later (Fig. 3, non-overlapping 95% CI). The peak in T_{core}

occurred approximately 3 days before the peaks in T_{eye}, T_{bill} and T_{foot}, but not significantly so

considering 95% CI overlapped (Fig. 3). T_{foot} was the first to return to baseline levels at *ca.* moult

day 14, followed by T_{trunk}, T_{bill}, T_{eye} (*ca.* moult day 27), then T_{core} (*ca.* moult day 34) and then only by

$T_{flipper}$ (*ca.* moult day 44). While 95% CI overlapped between T_{core} and T_{bill}, T_{eye}, $T_{flipper}$, this was not

the case between T_{core} and T_{trunk} (Fig. 3).

A glimpse at the natural scenario

Out of 87 observations from 43 wild individuals (mostly during early moulting stages),

none were observed panting, whereas breeding individuals have been reported to pant during

ca. 20% of behavioural observations in the same population [22]. Moulting birds immersed their

feet in the river during 75.8% of our observations. The visible part of moult lasted 13 days in the

only individual that completed its moult before the HPAI-related closure of the field site.

T_{subcut} was approximately 1.4°C higher in early-moulting birds compared to the same

individuals measured while breeding (Fig. 4a). Average daily T_{core} was overall 1.2°C higher during

moulting compared to breeding in one wild individual (Fig. 4b), despite environmental heat load

(*i.e.* WBGT) being lower during moult (Fig. 4b) and this individual being consistently observed

with its feet immersed in the river. This wild individual had T_{core} within the range of captive non-

moulting individuals during breeding (except the first day), and within the range of captive peak-

moult between approximately 4 and 11 days of its moulting fast on land (Fig. 4b). T_{core} of this

moulting individual did not return to breeding or captive non-moulting values, even once moult

was visibly completed (Fig. 4b).

Figure 4. Subcutaneous (a) and core (b) body temperatures of moulting *vs.* breeding wild king

penguin(s). The same individual(s) were measured both incubating and moulting during the same

breeding cycle (2023-24). (a) N = 3 individuals measured during incubation and early moult (*ca.* 3rd day of

fasting on land), with individual values presented in grey and mean ± SE in black. (b) N = 1 individual,

coloured symbol and error bars represent daily average core body temperature ± SE, lines represent

environmental heat load measured as Wet Bulb Globe Temperature, and shaded areas represent 95% CI

of core body temperature for captive non-moulters (blue, n = 2) and during peak moult (red, n = 6).

Discussion

Warming up to a new coat

Our results provide evidence that moulting elicits thermovascular responses that increase

heat loss in king penguins, as previously described in gentoo penguins [10]. We now also provide

evidence that core body temperature in king penguins markedly increases during moult (*i.e.* up

to +1.4°C), suggesting either a failure to dissipate excess heat generated by the up-regulation of

metabolism (*thermal challenge hypothesis*), or a regulated increase of body temperature during

moult (*warm-up hypothesis*). By assessing the timing of changes in surface and core body

temperature, we found that surface temperatures increased rapidly, and likely after the rise in

core temperature occurred, thereby providing support for the *warm-up* hypothesis. Coupled with

the lack of panting observed in the study period, these findings suggest that increasing peripheral

heat dissipation was likely enough to maintain core temperature below a maximum threshold,

but that this threshold was likely finely modulated throughout moult. The limited evidence we

gathered from wild king penguins (*i.e.* +1.4°C in subcutaneous and +1.2°C in core body

temperature) suggests that the hyperthermic pattern we observed in captivity also occurs under

natural conditions, where behavioural heat dissipation was less constrained because individuals

had access to cold water, and used it for partial immersion.

King penguins must accomplish moulting efficiently to avoid reproductive failure because

food availability decreases rapidly after the moulting period [39]. It is known that king penguins

that arrive from the sea later in the season initiate feather loss more quickly and undergo a

contracted moult period relative to individuals that arrive earlier [40]. Hyperthermia may benefit

moulting penguins by accelerating the process of feather growth, thus enabling them to better

match high food availability. Assuming that king penguins in our study had a Q10 equal to 2.3 [41,

Roussel *pers. com.*], the observed increase in body temperature of ~1.4°C would yield a 12.4%

increase in metabolic rate, which may shorten moult by *ca.* 2.5 days, equal to a 9.3% reduction in

the total duration of moult.

Positive correlations between feather growth and body temperature have been observed

in chickens, where individuals with an early feathering genotype display higher body

temperatures and faster feather growth during the first six weeks post-hatching [42]. A similar

pattern has been observed indirectly in turkeys, where individuals held at higher ambient

temperature had longer feathers at 14 days post-hatching [43], although this may partially be

explained by metabolic costs being diverted to thermoregulation instead of feather growth at the

lower temperature treatment used in that study. Further supporting the association between

high body temperature and accelerated feather growth, the majority of feather growth occurs

during the day when body temperature and metabolic rate are highest [44]. In penguins,

moulting-induced hyperthermia occurs alongside increases in blood perfusion at the surface [10;

this study], which likely helps to deliver nutrients to peripheral tissues that is necessary for feather

growth [45].

Thermoregulation in captivity and in the wild during catastrophic moult

Core and surface temperatures in captive non-moulting birds remained stable during the

study period relative to moulting birds, which exhibited distinct changes over time. Core

temperature was the first to increase under captive conditions as we were not able to detect a

pre-moulting stable period. During the gradual rise in core temperature, we observed increases

in surface temperature in thermal windows (flippers, bill, and feet), indicating increased effort to

dissipate heat. Interestingly, the flippers were the only thermal window to reach the maximum

surface temperature before core body temperature reached its maximum, suggesting that the

flippers may function as a first line of defense for peripheral heat dissipation in penguins, likely

due to their large surface area and vascularization [46]. On the other hand, the surface

temperatures of the bill and feet closely tracked fluctuations in core temperature, as indicated by

the similarities in the timing and direction of changes in these temperatures. That heat dissipation

via thermal windows increased after core temperature had already been rising indicates that

hyperthermia was likely regulated and maintained below a maximum threshold. This

interpretation is further supported by the lack of panting observed throughout the study period.

We found that the surface temperature of the trunk declined in captive moulting birds, indicating

increased insulation due to the overlap in new and old layers of plumage [10,47]. Despite the

increased insulation during the moult, core temperature had already begun declining by the time
insulation reached its maximum (i.e., the minimum of T_{trunk}). This observation suggests that the
rise in core temperature was primarily driven by increased metabolic heat generation, not by
increased insulation as hypothesized in [10].

Conducting this research in captivity enabled us to eliminate variation that would be
introduced by more dynamic thermal environments in the wild, thus isolating the effects of
moulting on thermoregulation. On one hand, captivity prevented heating from solar radiation; on
the other hand, captivity prevented cooling from wind chill and precipitation, and it limited
behavioural thermoregulation in the form of habitat selection (i.e., partial immersion in water).
However, behavioural thermoregulation and environmental conditions, such as solar radiation,
wind, and humidity remain essential considerations for wild animal thermoregulation [22,48–50].
Therefore, to provide insights about the relevance of our finding in captivity, we measured
thermoregulatory behaviour, subcutaneous temperature, and core temperature in a limited
number of wild king penguins. During 75.8% of our observations, free-living moulting king
penguins had their feet immersed in the river, confirming that wild birds likely take advantage of
additional means to dissipate heat. Despite this behaviour and greater potential for wind chill,
core temperature measured in one wild moulting king penguin showed a similar hyperthermic
pattern (+1.2°C compared to incubation) as the one found in captive moulting king penguins
(+1.4°C). Although this result was obtained in only one individual, it is supported by the similar
pattern observed in the subcutaneous temperature of three wild king penguins measured during
both incubation and early-moult (+1.4°C). It is interesting to note that the core body temperature
of captive individuals was similar to the wild scenario, both during ($T_{core} \simeq 38.7$°C) and outside
($T_{core} \simeq 37.3$°C) the moulting period, thereby suggesting that captivity does not markedly affect
thermoregulation in king penguins.

Seeking baselines: Operating points of body and surface temperatures

The data we report conform well to a negative feedback-based model of thermoregulation
with multiple autonomous thermo-effectors [51]. The elevation of metabolic heat generation

while moulting represents a positive (i.e., heat) thermal challenge that ultimately triggers an

increase in heat dissipation via the periphery. The rises in core, and subsequently surface,

temperatures represent the changes in multiple operating points that would be necessary to

balance heat loss with elevated heat generation. The thermal challenge introduced by moulting

was large enough to elicit changes in peripheral heat loss, but not panting, suggesting that the

thermoregulatory response observed lies within an inter-threshold zone [52–55]. We interpret

the difference in timing and magnitude of change in flipper temperature relative to other thermal

windows to represent varying thresholds for response across body regions, a pattern supported

by the specialization of regional heterothermy in birds [56], and indeed by the widespread

presence of regional heterothermy in endotherms [57].

Previous studies have suggested that moulting poses a negative (i.e., cold)

thermoregulatory challenge due to reduced insulation during feather loss, which is compensated

by an increase in heat production [17–19,58]. If moulting poses a negative thermal challenge, one

would expect a reduction in body temperature followed by upregulation of metabolic heat

generation until the thermal challenge passes (i.e., hypothermia, not hyperthermia [51]).

However, moulting in penguins [59,60], present study] and other birds [61,62] can elicit elevations

in body temperature (i.e., hyperthermia), the opposite of the pattern predicted if moulting was a

negative thermal challenge. Moulting-induced hyperthermia may be less pronounced in species

where heat loss occurs too efficiently for body temperature to be affected by upregulated

metabolism (e.g., small passerines; [63]), or if the sampling period does not align with the onset

of hyperthermia. In the wild, penguins arrive onshore with new feathers developed to between

18% to 42% of their final length [18,64], and it is possible that studies in wild individuals may not

have been able to detect early changes in core temperature occurring during the pre-moult or

early-moult stages. Pre-moult elevation of core temperature is not mutually exclusive with

increased energy cost of thermoregulation in later stages of the moult when insulation decreases

quickly, as observed in [19].

Conducting this research in captivity enabled us to measure moulting and non-moulting

individuals simultaneously throughout the entire moulting process. Despite starting to measure

core temperature more than two weeks before the first visible change in plumage, our sampling

period was too late to capture a stable pre-moult phase of core temperature. In gentoo penguins,
lower surface temperatures were observed at the end of the study, possibly because the sampling
period may have begun after body temperatures had already begun increasing [10]. This and
previous studies highlight the difficulty of establishing a baseline for normothermia, and these
differences in sampling period may explain some of the differences between studies. We advise
care when planning sampling periods for physiological studies of moulting because physiological
changes in preparation for moulting that underly visible change in plumage may have an earlier
start than what was previously appreciated in the literature.

Warming up in captivity – heating up in the wild?

We found that the increase in core temperature in moulting king penguins was likely
regulated, as evidenced by the lack of influence from plumage insulation, the absence of panting
behavior, and the delay in vascular perfusion. These patterns suggest that hyperthermia was likely
employed as a regulated thermoregulatory strategy [65]. Air temperature remained stable and
far below body temperature during our study, and the warming effect of solar radiation was
absent in captivity. Since environmental conditions are unlikely to be as forgiving in the wild, the
current study raises concerns regarding the vulnerability of moulting penguins in the context of
global climate change. High mortality rates have already been reported in multiple penguin
species immediately after moulting due to the energetic challenge of this fasting period [18,66–
68]. More recently, in 2019, a mortality event was recorded during early moult in little penguins
following a heatwave on land (*Eudyptula minor*; [69]). Based on our finding that moulting poses
a positive (i.e., heat) thermal challenge, we caution that moulting may increase the susceptibility
of wild penguins to heat stress, especially as regions near the poles warm at a disproportionately
rapid rate [24,70]. Further investigation of thermoregulatory responses of penguins during
moulting in the wild is thus deeply needed.

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Author's contribution

Study design: ALew, AS, JZ. Funding acquisition: ALew, AS, ALey, JZ. Data collection in captivity: JZ, EP, ALew. Data collection in the field: EM, AS. Data analysis: JZ, EP, AS. Writing original draft: JZ, ALew and AS. Writing review and editing: ALey, EP, EM.

Ethical note

The study in captive penguins did not include scientific procedures. Scientific procedures for wild king penguins were approved by the French Ethical Committee (*APAFIS#31268-2021042117037897 v3*) and the Terres Australes et Antarctiques Françaises (Arrêtés TAAF A-2023-88).

Data availability

The datasets and R code used in this manuscript are available on FigShare:

https://figshare.com/articles/dataset/Data_from_Warming_up_to_a_new_coat_moul_ng_king_peng

uins_exhibit_hyperthermia_and_increased_peripheral_heat_loss_/28629305

Ethical note

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and the Terres Australes et Antarctiques Françaises (Arrêtés TAAF A-2023-88).

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Electronic Supplementary Material (ESM)

Supplementary methods S0: Standardization between IR cameras

We standardized measurements made with the three cameras using two separate linear models

that were built using the surface temperature of the plumage of non-moulting birds. We selected

the plumage because of its lack of vascularization to avoid confounding differences between

cameras with natural variation in peripheral vasoconstriction, and we used data from non-

moulting birds to avoid confounding differences between cameras with changes associated with

the moulting period. The linear models included the camera as a fixed effect to quantify the effect

of each camera on the measured surface temperature, and T_w to account for variation introduced

from environmental conditions (temperature and humidity). We built the first regression using

data collected daily during the study when one camera was available at a time (*i.e.*, non-

simultaneous comparisons), and we built the second regression using data collected at the end

of the experiment when all three cameras were available for simultaneous comparisons. Both

models yielded similar results, and we made corrections using the parameter estimates from the

models built with simultaneous comparisons because we expect that this dataset is more

resistant to temporal variation in surface temperature.

Figure S0. Differences in surface temperature measurements between cameras. Data are of plumage temperature over the course of the study (left) and at the end of the study when all three cameras were available for simultaneous comparisons (right).

Table S0-A. Parameter estimates for the effects of the two replacement cameras on surface temperature measurements. Linear models were built using simultaneous comparisons between the E96 and each of the replacements (E8 and T640), and the resulting parameter estimates were used to standardize measurements made with the different cameras.

Camera	Parameter estimate	Standard error	p-value
E8	2.20	0.594	0.0138
T640	-1.90	0.389	0.0164

Table S0-B. Resolutions of the three thermal imaging cameras used in this study.

Camera	Resolution
E96	640X480
E8	320X240
T640	640X480

Supplementary results S1:

Figure S1. Environmental heat load evaluated as Wet Bulb Globe Temperature (WBGT) between captive conditions (yellow) and wild conditions during breeding (blue) and moulting (red). Individual daily data points are represented in colour along with mean + SE in black.

Table S1. Parameter estimates for fixed effects in top ranked linear models for T_{core}, T_{eye}, T_{bill}, $T_{flipper}$, T_{foot} and T_{trunk} in relation to moult day. Time periods have been defined according to breakpoint analyses (see methods and results), except for non-moulting individual for which the entire study period has been considered. Significant p-values (<0.05) are denoted with (*).

Region	Time period (moult days)	Intercept (°C)	Moult day (°C/day)	p-value	T_w (°C/°C$_w$) or T_{ground} (°C/°C$_{ground}$)	p-value	R^2_c
T_{core}	-28 to 3	38.48	0.064	<0.0001*	NA	NA	0.78
	4 to 32	38.90	-0.045	<0.0001*	NA	NA	0.70
	34 to 62	37.35	NA	NA	NA	NA	0.26
	Non-moulting	37.31	NA	NA	NA	NA	0.14
T_{eye}	-37 to -10	26.36	-0.08	0.0011*	NA	NA	0.35
	-9 to 6	30.05	0.27	<0.0001*	NA	NA	0.63
	7 to 27	32.92	-0.17	<0.0001*	NA	NA	0.45
	28 to 62	28.08	NA	NA	NA	NA	0.11
	Non-moulting	25.05	NA	NA	0.34	0.0252*	0.17
T_{bill}	-37 to -12	15.34	NA	NA	NA	NA	0.24
	-11 to 6	17.85	0.68	<0.0001*	0.63	0.129	0.52
	7 to 26	29.23	-0.45	<0.0001*	NA	NA	0.33
	27 to 62	16.08	NA	NA	NA	NA	0.30
	Non-moulting	8.90	NA	NA	0.57	0.0058*	0.12
$T_{flipper}$	-37 to -11	9.43	-0.08	0.0511	NA	NA	0.069
	-10 to -5	19.50	1.65	0.00775*	1.36	0.05486	0.74
	-4 to 43	21.70	-0.22	<0.0001*	NA	NA	0.54
	45 to 62	4.09	NA	NA	0.87	0.00016*	0.80
	Non-moulting	7.21	NA	NA	0.54	0.0150*	0.080
T_{foot}	-37 to -10	11.31	NA	NA	NA	NA	0.47
	-9 to 6	9.37	0.21	0.12	0.25	0.014*	0.65
	7 to 13	23.35	-0.77	0.04*	NA	NA	0.44
	15 to 62	13.59	NA	NA	-0.087	0.0028*	0.45
	Non-moulting	7.89	NA	NA	0.069	0.0203*	0.10
T_{trunk}	-37 to -7	19.53	NA	NA	-0.68	0.051	0.15
	-6 to 11	11.94	-0.32	<0.0001*	0.29	0.054	0.55
	12 to 25	5.68	0.42	<0.0001*	NA	NA	0.33
	26 to 62	17.83	-0.054	0.0153*	NA	NA	0.55
	Non-moulting	11.22	NA	NA	0.33	0.0097	0.093