Long-lasting coexistence of multiple asexual lineages alongside their sexual counterparts in a fungal plant pathogen

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Abstract

Sexual-to-asexual transitions within species are crucial for understanding reproductive evolution, yet the coexistence of both modes of fungal species in the same environment is poorly documented. Here, we report this transition in a plant pathogen species that coexists within the same geographical environment. Our biological model is the poplar rust fungus *Melampsora larici-populina,* which displays a complex life cycle typical of rust fungi (Pucciniales). It alternates between two unrelated hosts to complete an obligate sexual life cycle once a year. We conducted a comprehensive population genetic analysis, using 21 microsatellite markers and data from 2,122 individuals gathered over 30 years from various locations in France. Our results demonstrate the existence of many distinct lineages that reproduce asexually through the years, skipping the sexual phase. Clustering analysis identified a group of multilocus lineages that displayed all hallmarks of the genetic consequences of asexual reproduction, including highly negative and large variance among loci of the inbreeding coefficient (F_{IS}). This indirect evidence for asexual reproduction was confirmed by the direct observation of these asexual lineages being repeatedly sampled across multiple years. This result demonstrates the coexistence of these lineages with their sexual counterparts in the same ecological niche, challenging conventional assumptions about geographical sorting of reproductive modes. These considerations are of paramount importance for understanding the contemporary evolution of major pathogen species. This switch from sexual to asexual reproduction has contributed to devastating epidemics worldwide.

27 **Keywords**: sexual-asexual transition | evolution of reproductive modes | population genetics | rust

28 fungi | time samples.

1 Introduction

The evolution of sex remains a fundamental question and an intriguing topic in evolutionary biology (Culotta & Hines, 1998; Otto, 2009; Hartfield & Keightley, 2012). While sexual reproduction may increase genetic diversity, purging deleterious mutations, and enhancing adaptation to changing environments (Schurko et al., 2009; Pierre et al., 2022). It incurs costs such as time and energy to find mates, combined with the biological cost of meiosis (Roughgarden, 1991; Aanen & Hoekstra, 2007; Hörandl, 2013). Asexual reproduction, on the other hand, produces offspring without a mate, allowing for efficient reproduction, maintenance of lineages adapted to local stable conditions, and quick multiplication (Shick et al., 1979; Lodé, 2013). Due to a lack of recombination, asexual lineages are more vulnerable to the accumulation of deleterious mutations (Lynch et al., 1993; Peck, 1994; Keightley & Eyre-Walker, 2000), and more prone to losing allele diversity by genetic drift (Rouger et al., 2016). The study of species displaying both sexual and asexual reproduction enables pinpointing the pros and cons of each mode of reproduction (Butlin, 2002; Simon et al., 2010; Billiard et al., 2012). In particular, the study of cases where there is a dynamic transition from sexual to asexual reproductive modes is likely to highlight scenarios in which each reproductive mode could be favored.

The success of asexual reproduction frequently relies on specific environmental conditions. Invasive species often shift towards asexual reproduction in their exotic range, while reproducing sexually in their native environment (Song et al., 2011; Platt & Jeschke, 2014; Gladieux et al., 2015; Tilquin & Kokko, 2016). Ant species can exhibit such diverse reproductive strategies, with asexual reproduction frequently being dominant in invasive populations, as observed in *Wasmannia auropunctata* and *Cerapachys biroi* (Foucaud et al., 2006; Kronauer et al., 2012). Plant pathogenic fungi also adopt diverse reproductive strategies, with asexual reproduction being responsible for the most damaging epidemics (Ashu & Xu, 2015; Gladieux et al., 2015; Drenth et al., 2019). Potato late blight is a devastating disease caused by the oomycete pathogen *Phytophthora infestans.* Both sexual and asexual reproduction take place in the native range, but it is asexual reproduction only that led to the devastating Irish Potato Famine in the 1840s (Gavino et al., 2000; Ristaino, 2002; Fry, 2020). The wheat yellow rust fungus *Puccinia striiformis* f. sp. *tritici* displays a similar pattern, with different reproductive modes depending on the region. The fungus reproduces sexually in China and Pakistan, which is highlighted by its high genotypic

diversity, whereas it reproduces asexually in the rest of the world (Bahri et al., 2009; Mboup et al.,

2009; Ali et al., 2014). The rice blast fungus *Magnaporthe oryzae* (Syn: *Pyricularia oryzae*)

reproduces sexually in a limited area of Southeast Asia, its native range, whereas epidemics

globally are driven by asexual lineages (Saleh et al., 2012).

The coexistence of both reproductive modes in the same geographical environment is still poorly

documented, despite offering a unique opportunity to decipher the evolutionary and ecological

dynamics of sexually or asexually derived lineages. While rare in fungi, such studies are common

in other taxa, such as in aphids (Halkett et al., 2006; Simon et al., 2010), daphnia (Lehto & Haag,

2010), and a cyprinid fish *Carassius gibelio* (Barbuti et al., 2012). The phytopathogenic fungus

Melampsora larici-populina, causing poplar rust, belongs to the taxonomic order of Pucciniales

(Steenackers et al., 1996). This species follows a complex life cycle with an obligate sexual

reproduction that alternates with rounds of clonal multiplication (Figure 1). Two unrelated host

plants are required to complete the life cycle (Hacquard et al., 2011). Clonal multiplication occurs

on poplar trees (*Populus* spp.), while sexual reproduction takes place on larch trees (*Larix* spp.)

(Lorrain et al., 2019). In the following, lineages that complete the entire life cycle, including the

obligate sexual reproduction phase, are referred to as sexual lineages. The sexual reproduction on

larch ensures genetic recombination and obliterates all traces of the previous rounds of clonal

multiplication (Xhaard et al., 2011; Rouger et al., 2016).

Figure 1

The *Melampsora larici-populina* life cycle. The blue arrows indicate the heteroecious life cycle, the clonal multiplication phase on poplar tree leaves during summer, and the sexual phase on the larch tree. The dashed red arrow indicates pure asexual multiplication on poplar, skipping the sexual phase.

A previous population genetic study (Xhaard et al., 2011) has provided evidence for some variability in reproductive mode, with an obligate sexual genetic group coexisting (i.e., the same geographic region) with the genetically distant group that displays all hallmarks of long-lasting asexual reproduction. This suggests that some lineages of the poplar rust fungus could survive alone through clonal multiplication on poplar. However, this preliminary study did not investigate the ecological and evolutionary dynamics of the asexual group, as it relied on a single sampling year. The coexistence of sexual and asexual reproduction within the same geographic region makes

M. larici-populina an ideal evolutionary model for elucidating the transition from one reproductive
mode to another, as both modes can be found in the same environment.

In this study, we provide an in-depth characterization of the reproductive strategies of *M. larici-*
populina, with an emphasis on the (spatio-temporal) persistence of asexual lineages and their
genetic characteristics. Our study also examines the ecological and spatial factors influencing the
distribution of sexual and asexual populations, thereby offering insights into the evolution of
reproductive modes in rust fungi. This study encompasses the analysis of 2122 individuals sampled
across France over 30 years of population surveys that have been genotyped using 21 microsatellite
markers.

We used a two-fold approach to assess the reproductive mode variation among poplar rust
individuals. First, we employed an indirect population genetic approach based on the clustering
among lineages and subsequent evaluation of the variation in population genetic indices known to
be signatures of asexual reproductive mode, namely: repeated genotypes, linkage disequilibrium,
and overall negative F_{IS} value and large variance over loci (Balloux et al., 2003; Halkett et al.,
2005; Stoeckel & Masson, 2014). Second, we benefited from time-samples to investigate the
resampling through years of the same multilocus lineage (MLL), which is direct evidence for their
asexual survival. Furthermore, the genetic relatedness of all individuals was analyzed to
investigate evolutionary relationships among asexual lineages and with their sexual counterparts,
which points to multiple events of the emergence of asexual lineages. Finally, examining the
coexistence of sexual and asexual lineages, their long-term persistence, and their spatial
distribution suggested that environmental and ecological factors potentially play a role in shaping
the survival and maintenance of asexual lineages in this species.

2 Material and Methods

2.1. Isolate collection and sampling strategy

To characterize the long-term genetic structure of *Melampsora larici-populina* in France, we analyzed a total of 2,122 isolates collected from 61 administrative departments over 30 years (1992-2024) (Figure S1). The dataset (Table S1) combines previously published genotypes with new collections, structured into three distinct sampling categories: (i) National surveys (broad spatial scale): We integrated data from two major countrywide sampling campaigns. The first corresponds to the 2009 survey (23 locations, n=428) previously described by Xhaard et al. (2011). The second is a new dataset from a 2011 survey, extending coverage to 60 locations (n=1,374). All survey samples were collected in late summer (August-September), coinciding with the peak of clonal multiplication but prior to the sexual cycle (Figure 1); (ii) Targeted monitoring and historical collections (temporal scale): We included 246 isolates from a long-term epidemiological monitoring project in the Durance River valley (2004, 2008, 2020) (Xhaard et al., 2012; Becheler et al., 2016). These were supplemented by 34 historical isolates from our laboratory collection, spanning seven years between 1992 and 2014.   (iii) Overwintering assessment: To investigate potential overwintering mechanisms, we performed a specific sampling campaign in February 2024 (n=40) at a unique stand in Cavalaire-sur-Mer (southern France). Unlike typical poplar stands, this site retains green foliage through winter, offering a rare opportunity to sample active rust populations outside the standard growing season.

2.2 Genotyping

The DNA extraction process was guided by a protocol that employs the BioSprint 96 DNA Plant Kit and the BioSprint 96 automated workstation (Qiagen). Samples were genotyped using 21 microsatellite loci [Mlp_91, Mlp_83, Mlp_50, Mlp_71, Mlp_96, Mlp_95, Mlp_55, Mlp_66, Mlp_12, Mlp_68b Mlp_54, Mlp_57, Mlp_58, Mlp_49, Mlp_93, Mlp_82b Mlp_56, Mlp_87, Mlp_97, Mlp_77, Mlp_94 (Xhaard et al., 2009)]. These loci have been extensively utilized in several population genetic studies of this species (Xhaard et al., 2011; Becheler et al., 2017; Persoons et al., 2017; Louet et al., 2023; Saubin et al., 2024). The amplification through PCR with the Multiplex PCR Kit (Qiagen) was performed as described by Xhaard et al. (2011). It was conducted across three multiplex reactions containing eight, five, and eight loci, ranked by

amplification size, respectively. The PCR products were then pooled and analyzed using an ABI

3730 Genetic Analyzers (Applied Biosystems) by the company GenoScreen. Fragment sizing

utilized a LIZ-1200 size standard, and allele scoring was performed with GENEMAPPER V4.0.

Samples that failed to amplify at more than six loci were excluded from further analysis.

2.3 Population genetic and statistical analyses

First, to handle genetic data of microsatellite markers in the R programming language v.4.4.1, we

converted the data format from a data frame to a genuine object using the `df2genind` function

from the **adegenet** package v.2.1.10. This function optimizes multilocus genotype data and

accommodates missing data if it is found in some individuals or loci. The automated workflow

code provides full details on subsequent analyses (https://doi.org/10.5281/zenodo.15100450)

(Abdalrahem et al., 2026) .Moreover, specific analyses were executed using the GenAPoPop 1.0

software due to specific methodological requirements.

Identification of repeated genotypes: The prerequisite to an accurate assessment of reproductive

mode is the proper delineation of repeated genotypes derived from clonal multiplication (Halkett

et al., 2005; Arnaud-Haond et al., 2007). Individuals with the same combination of alleles (the so-

called multilocus genotypes, MLG) were consistently identified through both an ad hoc procedure

in Excel during the manual curation of chromatographs and using `mlg.filter` from the **poppr**

package v.2.9.3 (Kamvar et al., 2014). Genotypes differing by a few alleles were double checked

by returning to the original chromatographs. All remaining differences between genotypes are

therefore confirmed and reliable. We computed (*Pidu*) and (*N_Pidu)* to test for the discrimination

power of our marker set, and to rule out the probability of finding repeated MLGs by chance (that

is deriving from sexual reproduction) using GenAPoPop 1.0 (Evett & Weir, 1998; Waits et al.,

2001; Stoeckel et al., 2024).

Delineation of multilocus lineages: As the accumulation of somatic mutations over time can lead

to slight differences among multilocus genotypes within a given asexual lineage (Arnaud-Haond

et al., 2007), we subsequently grouped MLGs into multilocus lineages (MLL). So as not to apply

an arbitrary threshold for MLL delineation, we first computed the distribution of pairwise shared

allele distance as advised by Arnaud-Haond et al. (2007), and Bailleul et al. (2016). As intra lineage

differentiation can vary across MLLs, we chose to use the UPGMA clustering method, which

aggregates MLGs one by one by examining the nearest neighbors until the MLL boundaries are

found (Kamvar et al., 2014, 2015). The threshold on genetic distance between neighbor genotypes

was set using the function `cutoff_predictor` from **poppr**.

Clustering procedure to indirectly identify asexual lineages: As the asexual reproductive mode

freezes genotypes, it retains ancestral allelic states. Asexual lineages can thus rapidly diverge from

their sexual counterparts, the latter being still influenced by genetic drift (Rouger et al., 2016). In

line with this statement, we previously found large genetic differences between sexual and asexual

groups of *M. larici-populina* (Xhaard et al., 2011). Assigning poplar rust individuals would thus

be an effective approach to identify asexual lineages without any *a priori*. The main pitfall of this

method being that the clustering algorithm should not rely on gathering individuals according to

Hardy-Weinberg expectations, which is the common assumption of any Structure-like approach

(Pritchard et al., 2000). We thus carry out our clustering analysis using a PCA and discriminant

analysis approach based on allelic distribution (Jombart et al., 2010). We used the `find.clusters`

function from the **adegenet** package to reduce the dimensionality of the dataset. Discriminant

analysis of principal components (DAPC) was then used to refine the identification of clusters by

maximizing the variance between two clusters, while minimizing within cluster variance (Jombart

et al., 2010). Cross-validation (`xvalDapc`; 100 replicates, 90% training set) identified 10 principal

components (PCs) as optimal for DAPC, with a mean group assignment success of 99% and 46%

of total genetic variance retained. The optimal number of clusters was determined to be K=2, a

choice supported by two complementary lines of evidence. First, the Silhouette score K=2

(Shahapure & Nicholas, 2020), as a statistical support (Figure S2). Second, this clustering aligns

with our biological question to disentangle the two major reproductive modes (sexual vs. asexual

lineages); individuals were grouped into two clusters using the `dpca` function with *K*=2. To ensure

the reliability of the assignment, only individuals with a high membership coefficient (≥0.8) were

retained for further analyses of within-cluster genetic characteristics.

Resampling multilocus lineages (MLLs) as direct evidence for asexual reproduction. Observing

the same genotype over several years is an indication that this lineage survived asexually - unless

sexual reproduction could have resulted in the same combination of alleles by chance, which can

be ruled out by a *Pidu* analysis (Evett & Weir, 1998; Stoeckel et al., 2024). To track the persistence

of MLLs over time, we recorded the years each MLL was detected and calculated its frequency

across different years. A lineage that persisted for at least two years was considered asexual. We

tested the overlap between the direct and indirect approaches for assessing asexual reproductive

mode by performing Fisher's exact test (Clarkson et al., 1993) on the contingency table of MLL

sorting according to the direct or indirect approach by using the `fisher.test` function from the

stats package v.4.4.1. In the following, a given MLL is considered asexual if it meets at least one

of the two criteria: the assignment to an asexual cluster or the inter-annual resampling over a

minimum of two years.

Computation of classical population genetic indices: As asexual reproductive mode distorts allele

repartition within and among individuals, we computed several classical population genetic indices

used to characterize within cluster and within MLL genetic signatures. Observed heterozygosity

(H_O), unbiased expected heterozygosity (H_E) (Nei, 1978), genotype richness (R), and multilocus

linkage disequilibrium corrected for the number of loci $\bar{r}_d$ were calculated using the **poppr**

package (basic function). Allelic richness (A_R), and *F*-statistics, including the inbreeding

coefficient (F_{IS}) (Weir & Cockerham, 1984) were computed using the **hierfstat** package v.0.5-11.

The clonal evenness was measured by calculating the *Pareto β* index, which summarizes the

distribution in size of MLLs using GenAPoPop 1.0 (Arnaud-Haond et al., 2007; Stoeckel et al.,

2024).

Spatial analysis of the repartition of sexual and asexual lineages: To illustrate the geographical

distribution of sexual and asexual individuals across France, the **maps** package v3.4 was utilized.

This enabled us to obtain information not only on spatial distribution, but also on the potential

coexistence of the two reproduction modes within the same geographical area. Using the `glmer`

function from the **lme4** v.1.1-35.5 package, a generalized linear mixed model (GLMM) with a

binomial distribution and a logit as a link function was employed to assess the regression

coefficients (Bolker et al., 2009). To further test the relationship between the reproductive mode

variation and the geographic locations, we performed a regression analysis on the count of sexual

and asexual lineages per site. The independent variables (fixed effect) were the geographic location

represented by latitude and longitude. The effect of the year of sampling was added as a random

effect. This analysis was performed on the 2009 and 2011 data originating from the two

countrywide surveys, so as not to overrepresent certain locations. To further assess the distribution

and relative abundance of asexual MLLs across time and space, we performed Fisher's exact test

for fluctuation in MLL abundance across years and locations. Given the large size of the data sets,

we employed a Monte Carlo simulation approach with 100,000 replicates to estimate p-values.

Genetic relationships between and among sexual and asexual lineages: To further elucidate the

evolutionary trajectories of sexual and asexual individuals, and to address the question of the

dynamics of transition from sexual to asexual reproduction and vice versa, a discrete matrix based

on the relative dissimilarity between individuals was calculated by `diss.dist` with the **poppr**

package to create a neighbor-joining tree (Saitou & Nei, 1987) using `nj` from the **ape** package

v5.8.1.

Inference of the rate of clonality: Finally, to estimate the rate of clonality in the *M. larici-populina*

populations, we employed the ClonEstiMate Bayesian method. This method analyzes the change

of genotype frequencies from the same population over generations (Becheler et al., 2017). We

calculated based on two scenarios: i) merging all individuals within a mixed population (that is

any individuals can survive asexually by chance), or ii) considering two distinct sexual and asexual

populations, based on the lineage sorting detailed above. Since the ClonEstiMate method should

be applied on temporal samplings of the same kind of populations, we used the data collected

during the countrywide surveys carried out in the years 2009 and 2011.

3 Results

3.1 Delineation of multilocus lineages

We identified 1470 multilocus genotypes (MLGs) over the 2121 genotyped individuals. The 21 markers we used provided enough statistical power to ensure unambiguous discrimination between genotypes. The overall probability of obtaining the same combination of alleles by chance under panmictic sexual reproduction (*Pidu*) is approximately 10^{-16}. This implies that 118 million individuals would have to be genotyped before observing two sexually produced descendants with identical genotypes (*N_Pidu*). We thus confidently considered that all the repeated genotypes we found were clones. We then grouped closely related MLGs into MLLs to account for the few somatic mutations that could occur during clonal multiplication. The delineation of MLLs was undertaken using a genetic distance threshold of 4.6, as determined by the UPGMA clustering algorithm. This grouping led to the delineation of 1381 MLLs, so slightly fewer than the number of MLGs, showing that the vast majority of MLGs have no closely related genotype.

3.2 Reproductive mode assessment

The clustering approach indicated that almost all individuals exhibited a high probability of membership in one of two genotypic clusters: 450 individuals were unambiguously assigned to cluster one, while 1665 individuals were assigned to cluster two (Figure S3). Only one isolate had membership coefficients below the threshold and was excluded from further analyses.

A comprehensive population genetic analysis of each cluster (Table 1) was conducted to assess the value of key population genetic indices, thereby revealing a contrast between the two clusters. Cluster 1 exhibits a highly negative F_{IS} value, low genotypic diversity, and high linkage disequilibrium (LD), all of which are indicative of a strong rate of clonal reproduction (Balloux et al., 2003; Halkett et al., 2005; Arnaud-Haond et al., 2007). The high variance of F_{IS} across loci further points to rare events of recombination within a population reproducing essentially asexually over the long term. In contrast, cluster 2 exhibited slightly positive F_{IS} values and higher genetic diversity, suggesting a higher degree of allelic recombination, a feature of sexual reproduction. The reduced linkage disequilibrium in this cluster further supported this classification. Consequently, cluster 1 was labeled as asexual, and cluster 2 as sexual. It is

275   noteworthy that similar levels of observed heterozygosity were found in both clusters, but a much

276   reduced allelic richness and gene diversity in the asexual cluster 1. Further, the *Pareto β* index was

277   low (~ 0.2), indicating a small number of dominant MLGs, which was consistent with high

278   clonality in cluster 1. In the sexual cluster 2, the *Pareto β* was greater than 1, reflecting a more

279   even distribution of genotypes due to genetic recombination.

Table 1. Characteristics of the two genetic clusters identified in *M. larici-populina* individuals.

	N	MLG	MLL	R	Pareto β	A_R	H_O	H_E	F_{IS}	Var (F_{IS})	$\bar{r}_d$
Clustering approach											
Cluster 1	450	67	15	0.147	0.226	4.754	0.523	0.361	-0.448	0.093	0.345
Cluster 2	1665	1403	1367	0.843	1.078	9.566	0.506	0.544	0.069	0.004	0.016
Resampling approach											
Multi-years	485	87	18	0.178	0.248	5.364	0.517	0.390	-0.324	0.130	0.339
Single-year	1630	1383	1363	0.848	1.057	9.695	0.508	0.545	0.064	0.004	0.012
Combination of approaches											
Asex	499	96	27	0.191	0.252	5.554	0.517	0.394	-0.314	0.119	0.327
Sex	1616	1374	1354	0.850	1.051	9.760	0.508	0.544	0.067	0.004	0.012

MLG: number of multilocus genotypes, MLL: number of multilocus lineages, *N*: number of individuals, *R*: genotypic diversity, *Pareto β*: evenness among repeated genotypes; A_R: allelic richness, H_O: observed heterozygosity, H_E: expected heterozygosity, F_{IS}: inbreeding coefficient, Var (F_{IS}): variance of inbreeding coefficient, and $\bar{r}_d$: multilocus linkage disequilibrium.

The resampling approach revealed two groups: 488 individuals with lineages spanning more than
one year due to asexual reproduction and 1,633 sexual individuals without resampled lineages
exceeding two years. The contingency table (Table 2), which compares the clustering and
resembling approaches, shows a discrepancy. Among the 54 individuals identified as sexual
through cluster analysis, subsequent resampling evidence validated their clonality. In contrast, 11
individuals were clustered as asexual; however not resampled.

Table 2. Contingency table showing the relationship between clustering and resampling
approaches for reproductive mode definition in the dataset. Values represent the number of
individuals assigned to each category.

	Asex (Resampling approach)	Sex (Resampling approach)
Asex (Clustering approach)	436	14
Sex (Clustering approach)	49	1616

A Fisher's exact test was performed to evaluate the correlation between the assignment of
reproductive modes from clustering and resampling analyses. The results were highly significant
(p-value $< 2.2 \times 10^{-16}$), thereby rejecting the null hypothesis of no association, with a 95%
confidence interval. This indicated a robust relationship between the two classification methods.
Therefore, a combination of clustering and resampling approaches was used to determine the
reproductive mode of each individual. The subsequent analysis involved the calculation of the
population genetic indices for both groups. The inference of reproductive mode using two
approaches as well as a combination of both, yielded consistent results over all population genetic
indices, confirming the robustness of the classification (Table 1). However, there are slight
differences in the values of population genetic indices among classification methods, with the

asexual group determined by the clustering approach revealing a stronger signature of asexual

reproduction.

3.3 Spatial distribution of reproductive modes

The spatial distribution of both sexual and asexual lineages of *M. larici-populina* individuals in France revealed distinct patterns (Figure 2). Sexual lineages were widespread across most regions, predominating in the northern and central parts of the country. In contrast, asexual lineages were more concentrated in the southern regions.

Figure 2

Geographic distribution of sexual and asexual lineages of *Melampsora larici-populina* individuals collected in 2009 and 2011 across France. The pie charts show the proportions of asexual (red) and sexual (blue) lineages at each sampling location. The size of each pie chart corresponds to the total number of individuals collected from that site.

To further investigate the factors that influence the prevalence of asexual lineages, a GLMM

binomial regression analysis was conducted. The analysis (Table 3) revealed a significant effect

of both latitude and longitude (Figure S4) on the proportion of sexual and asexual lineages, with

positive coefficients indicating that the relative abundance of asexual individuals increases with

lower latitude (the closer we get to the south of the country). Additionally, the random effect of

years (variance = 13.44, sd = 3.666) indicated that the temporal distribution of individuals was not

associated with a specific reproductive mode.

Table 3. Results of the binomial regression model predicting the effect of spatial distribution on

the reproductive mode of *M. larici-populina*

Predictor	Estimate	Std. Error	z-value	p-value	Significance
(Intercept)	-22.998	2.476	-9.126	< 2e-16	***
Lat	0.464	0.047	9.798	< 2e-16	***
Long	0.093	0.029	3.199	0.001	***

Estimate (regression coefficient), Std. Error (standard error of the estimation), z-value (z-statistic), p-value

(statistical significance), and Significance power indicated by asterisks (0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1

' ' 1).

The ClonEstiMate Bayesian analysis yielded insights into the rate of asexual reproduction in *M.*

larici-populina. The 99% credible interval from the posterior probability obtained for each group's

clonal rates. In the context of a mixed group, where sexual and asexual lineages are merged into

one population, the estimated clonal rate was between 0.22 to 0.31, indicating that about 27% of

the population reproduces asexually. When the analysis was performed under the assumption of

two distinct populations, the clonal rate was between 0.93 to 0.99 for the asexual group and

between 0 to 0.21 for the sexual group.

3.4 Genetic relationships between sexual and asexual lineages

The dendrogram tree shows different clades for sexual and asexual lineages (Figure 3), highlighting the seven main asexual lineages. However, some asexual lineages also appeared as closely related sister branches from the sexual clade. Barring MLL9, one of the most abundant asexual lineages, all of these asexual MLLs, closely related to the sexual clade are of small size. These small asexual lineages were thus sporadic, unlike the seven most abundant asexual MLLs that show persistence through time (Figure 3, Table 4). These main asexual MLLs also displayed some reticulation, indicating they consist of different MLGs. In contrast, MLL5 (a most prevalent MLL but classified as sexual) consists of individuals sampled only in 2024 that have exactly the same genotype. These observations underscored the evolutionary relationships and temporal dynamics between reproductive modes within and among *M. larici-populina* lineages.

 Figure 3

Circular Neighbor-joining (NJ) tree based on microsatellite data (21 markers) illustrating the genetic
relationships among *M. larici-populina* individuals. The color of the branches indicates the reproductive mode,
with blue representing sexual and red representing asexual modes. The outer rings represent the year of sample
collection, with each height corresponding to a year. The nine most abundant multilocus lineages (MLLs) are
color-coded.

3.5 Genetic structure and persistence of asexual lineages

A more detailed analysis of the asexual group identified several distinct genetic lineages. Among
these, seven lineages were found to be the most abundant. The dominant lineages exhibited varying
levels of persistence over the years, as indicated by their range of collection years (Table 4). In
particular, MLL1 was consistently detected in almost all collection years and across all regions,
highlighting its long-term persistence and widespread occurrence.

Table 4. Characteristics of the seven most abundant asexual multilocus lineages (MLLs) of *M.*
larici-populina.

MLL	MLG	N	Year range	R	A_R	H_O	H_E
1	36	279	1992:2024	0.122	1.274	0.519	0.274
2	3	49	2014:2020	0.042	1.290	0.571	0.287
3	4	44	1996:2009	0.070	1.316	0.619	0.312
4	9	38	1996:2020	0.216	1.266	0.499	0.263
6	6	23	1996:2020	0.227	1.196	0.354	0.192
8	5	14	1993:2014	0.308	1.306	0.575	0.296
9	3	14	2009:2011	0.154	1.189	0.350	0.183

358 *MLL:* number of multilocus lineages, *MLG:* number of multilocus genotypes, N: number of individuals, *R*:
359 Genotypic diversity, A_R: Allelic richness, H_O: Observed heterozygosity, H_E: Expected heterozygosity.

The distribution of these lineages in France differed with some lineages being widespread and others more restricted to some areas (Figure 4).

Figure 4

Distribution of asexual lineages over time and across locations. (A) The abundance of asexual MLLs in different French administrative regions. (B) The abundance of asexual MLLs in different years.

Fisher's exact test revealed a highly significant association between MLL distribution and both temporal and spatial factors (P = 1e-05). The analysis revealed that certain MLLs were repeatedly observed in the same geographic regions and over multiple sampling years, suggesting a pattern of persistence. Conversely, other MLLs appeared sporadic.

4 Discussion

The poplar rust fungus *M. larici-populina* is commonly regarded as an obligate sexual species that needs the presence of both poplars and larches to perform its annual life cycle (Pinon & Frey, 2005; Hacquard et al., 2011). However, a previous population genetic study suggested the existence of a distinct group of individuals that would survive asexually only on poplar hosts (Xhaard et al., 2011). Here i) we characterized the existence of an asexual group maintaining on the poplar hosts over the years, ii) we found that this asexual group is made up of several genetic lineages. We documented that some of these asexual lineages have persisted for nearly three decades despite their coexistence with sexual lineages in some places. These findings challenge conventional assumptions about the evolutionary dominance of sexual recombination in this species and offer a unique opportunity to investigate the transitions from sexual to asexual reproduction in a plant pathogenic fungus.

In the present study, two complementary approaches were employed to reveal and characterize asexual individuals. A clustering approach based on differences in allelic frequency and probability of membership was combined with a resampling approach aiming to identify the persistence of lineages over the years. We observed a strong overlap between these indirect and direct approaches, which enabled a sound delineation of asexual lineages among *M. larici-populina* individuals. This result further supports the reliability of the clustering approach performed in the initial study of (Xhaard et al., 2011) to identify asexual lineages indirectly. Applying such an individual based clustering approach is all the more relevant that sexual and asexual lineages can be admixed in the same location (Halkett et al., 2006). Consequently, the clustering approach can be regarded as a robust and independent method for detecting cryptic population genetic structures. In addition to the genetic differentiation between sexual and asexual clusters caused by the lack of gene flow, several genetic signatures clearly distinguished asexual lineages thereby reflecting the fundamental differences in evolutionary trajectories between these reproductive modes (Balloux et al., 2003; Halkett et al., 2005; Rouger et al., 2016). Sexual reproduction ensures the convergence toward Hardy-Weinberg equilibrium (HWE) with recombination among loci that reduces linkage disequilibrium (LD). Asexual reproduction, on the other hand, distorts both allelic and genotypic frequencies mostly because it freezes genotypes (Stoeckel & Masson, 2014; Rouger et al., 2016). As a result, the best way to identify asexual

lineages is to apply a multi-criteria method (Kronauer et al., 2012) in which the (diploid)

population displays the following hallmarks (Halkett et al., 2005; Arnaud-Haond et al., 2007): a

reduced genotypic diversity, combined with a significant excess of heterozygotes (negative F_{IS}

values). It should be noted that we also observed a very high variance of F_{IS} across loci, as expected

when some recombination events seldom occur (Balloux et al., 2003; Stoeckel & Masson, 2014;

Stoeckel et al., 2021). Here, the values found for these indices are consistent with a very high

asexuality rate as inferred by ClonEstiMate analysis. The asexual group also displayed significant

linkage disequilibrium among loci, which is often reported in asexual and partially asexual species,

despite not being consistently expected in neutrally evolving populations (Stoeckel et al., 2021).

Linkage disequilibrium was also observed in the sexual group along with a positive F_{IS} value as

the result of a Wahlund effect. Indeed, the sexual group encompasses different genetic subgroups

that have been identified by previous population genetics studies (Xhaard et al., 2011; Persoons et

al., 2017) and which are not considered here. We observed clone mates in the sexual group too, as

these can form lineages as abundant as those in the asexual group, with the notable difference that

they occur always at a unique sampling site and that, above all, they are never resampled over the

years in spite of their local abundance. This local abundance of clone-mate lineages is in

accordance with the sampling mostly done in the autumn, at the end of the clonal epidemic phase

of *M. larici-populina* life cycle. With regards to this, it should be recalled that the mandatory sexual

reproduction that takes place on larch resets the population genetic indices in the sexual group

each year (Xhaard et al., 2011; Rouger et al., 2016).The contrast in population genetic indices

provide a sound framework for distinguishing reproductive modes, supporting the reliable

assignment of individuals into either sexual or asexual groups and providing the first insights into

their respective evolutionary history (Xhaard et al., 2011).

In accordance with the previous study of Xhaard et al. (2011), we found that the asexual group

was unevenly distributed across the country, and mostly occurred in southern France. Asexual

lineages have often been described as distributed at the margins of a species' natural range (Simon

et al., 1999; Halkett et al., 2005; Drenth et al., 2019; Pereyra et al., 2023), a phenomenon called

the geographic parthenogenesis (Peck et al., 1998). Poplar rust is intimately dependent upon the

presence of its two hosts. Although the native range of European Larch (*Larix decidua*) is restricted

to the Alps and some other Central-European Mountain ranges, it has been widely planted in many

parts of France in mountains and plains since the 18th century. Nevertheless, it is less frequent in
the Mediterranean area, which can therefore be considered as a margin for the poplar rust fungus.
If we compare our results with those of other rust fungi, particularly those infecting wheat, we can
see a larger spatial segregation between the sexual lineages found only in the disease's center of
origin and the asexual lineages that have invaded the crops worldwide. Beyond rusts, this
observation is common to many pathogenic species as for the ascomycete fungus responsible for
rice blast (Saleh et al., 2014), or for the oomycete responsible for potato late blight (Ristaino,
2002). Interestingly, asexual lineages in these cases are specialized on different cultivars, and
therefore on different host genotypes (Goyeau et al., 2007), and the relative abundance of asexual
lineages therefore fluctuates over time with the use or disuse of these cultivars (Fontyn et al.,
2022). This shows to which extent the asexual evolution of these plant pathogen species is
associated with human practices. By contrast, in *M. larici-populina*, asexual lineages are sampled
mostly on wild poplar trees and are thus found on different host genotypes. We thus can argue that
this rust species enables the incipient stage of sexual to asexual transition to be documented.

A puzzling observation is that asexual lineages of *M. larici-populina* coexist with their sexual
counterparts in most sampling sites. The absence of larch trees in some areas can be compensated
for by the very high dispersal abilities of this species (Barrès et al., 2008), which can be wind
dispersed over hundreds of kilometers in a year (Saubin et al., 2024).This can readily explain why
sexual and asexual lineages are admixed even if no larch tree exists in the close vicinity of the
sampling site. Another non-exclusive hypothesis to account for the long-lasting coexistence of
sexual and asexual lineages is that of a temporal fluctuation in the environmental conditions that
favor the ones over the others. This temporal fluctuation could be responsible for the fluctuations
in abundance we observed, with almost half of the asexual lineages sampled in the year 2020 alone.
These fluctuations in relative sexual vs. asexual abundances could be linked to peculiar
meteorological events that would desynchronize the alternation between poplar and larch during
the sexual life cycle and disadvantage sexual lineages that have to complete this complex life cycle.
An outstanding example of the effect of a hazardous event on the fate of sexual and asexual
lineages was documented in the red alga *Agarophyton chilense* along the Chilean coasts (Becheler
et al., 2020). This study monitored wild (mostly sexual) and cultivated (asexually propagated)
populations of this red alga before and after a devastating earthquake. Although sexual populations

paid a large tribute immediately after the catastrophe, the artificial maintenance of an asexual
population caused the algae culture to collapse in the long term, suggesting that each mode of
reproduction plays a role in maintaining a dynamic equilibrium.

The distribution of asexual lineages of *M. larici-populina* prevailing in southern France points to
the first mechanism that can explain their maintenance and diversification. In milder climates the
asexual survival could be facilitated by a longer growing period of poplars, creating a so-called
green bridge that would enable the rust lineages to jump over the winter from poplar leaves to
poplar leaves. According to this hypothesis, the asexual lineages would passively bypass the sexual
phase on larch, without the need for any genetic determinism. In support of this hypothesis, it
should be noted that *M. larici-populina* urediniospores can survive for 10 months and still be able
to infect poplar leaves (Taris, 1968). But in these experiments, rust-infected poplar leaves were
stored either in a greenhouse, or in a wire cage 2 meters above ground level, which does not mimic
real natural overwintering conditions. According to this neutral hypothesis, both the onset and the
survival of asexual lineages would be a matter of chance only. In support of this hypothesis, we
observed that some asexual lineages branched among the sexual cluster and that these lineages
have only been revealed through the resampling over years, indicating they are in very close
genetic proximity with the lineages that undergo obligate sexual reproduction each year.

The persistence of several asexual lineages over decades, attesting to their evolutionary
maintenance and relative success, could also be indicative of some genetic determinism favoring
their survival over winter. In support of this hypothesis, the trade-off between overwintering ability
and multiplication during the epidemic season has been documented in other fungal species
(Hamelin et al., 2016; Suffert et al., 2018), and proven to account for the coexistence and
evolutionary maintenance of sexual and asexual lineages (Castel et al., 2014). It is also likely that
those long-lasting asexual lineages could have lost, at least to some degree, their ability to invest
in sexual reproduction, maybe to the benefit of greater asexual survival. Our population genetic
analysis, and especially the high variance of F_{IS}, would nonetheless indicate that recombination
does not occur among asexual lineages and thus that they do not evolve under strict asexuality but
rather as a succession of clones maintaining on poplar leaves over winter, renewed over the years.
Furthermore, the rare engagement of asexual lineages into the sexual reproduction can bring
additional explanations for the diversity of asexual lineages we observed. In line with the concept

of contagious asexuality observed in other organisms with mixed reproductive modes, such as
aphids and daphnia (Simon et al., 2002, 2003; Halkett et al., 2006, 2008), asexual lineages that
invest in sexual reproduction could be able to produce offspring that inherit their propensity to
survive asexually (Halkett et al., 2008). Deciphering further the history and evolutionary success
of these asexual lineages calls for a dedicated experiment to test their performance and investment
throughout the life cycle. While the complex life cycle of *M. larici-populina* can be implemented
in the laboratory (Pernaci et al., 2014), it remains a challenging experiment that is still ongoing.

The main original result of our study was to document the diversity among asexual lineages in *M.*
larici-populina, with some lineages persisting over decades and others appearing more
sporadically. The fact that two sampling campaigns covered the entire French territory, and that
other sites were sampled regularly over more than 30 years is evidence for this pattern not being
due to sampling bias. Notably, differences in abundance correlate with differences in the genotypic
structure between lineages. We documented that asexual MLLs can consist of different genotypes
(MLGs), unlike lineages within the sexual group that primarily consist of a singular repeated MLG.
While some persistent asexual lineages, such as MLL1, showed relatively high genotypic diversity
over time. Here it is important to state that only slight differences distinguish MLGs within asexual
MLLs, in the vast majority of cases up to three loci differ only. Moreover, the changes consist in
either heterozygous to homozygous conversion or to increase or decrease in allele length by a one
step mutation. This regular pattern of allele changes, found across all MLGs within MLLs,
supports the hypothesis that the asexual lineages evolved through single mutation steps. Although
mutation rates for the microsatellite markers used in *M. larici-populina* have not yet been
estimated, this regular pattern could present an opportunity to apply a tip calibration approach of
molecular clock analysis to estimate the timing of the emergence and persistence of these asexual
lineages (Rieux & Balloux, 2016). Yet, the differential abundance and genotypic diversity among
asexual lineages reflects heterogeneity within asexual population dynamics and emphasizes the
necessity to consider each lineage independently, rather than presuming that the whole asexual
population follows the same evolutionary trajectory. The most parsimonious hypothesis is that this
diversity has been generated by the repeated emergence of asexual lineages through the
evolutionary mechanisms previously discussed (chance of survival or contagious asexuality).
Among all asexual lineages, MLL1 stood out as the most abundant lineage, being repetitively

sampled from 1992 and showing an outbreak in 2020. It was detected in all regions where asexual

lineages were sampled, and forms the only contribution to the unexpected occurrence of asexual

lineages northward in the center of France, nearby Paris. The genome of a reference asexual isolate,

representative of this MLL, has recently been sequenced, and analysis of its genome structure,

polymorphisms, and gene content will further document the tale of this dominant asexual lineage.

Data

To ensure reproducibility, all analyses were executed using R version 4.4.1. The relevant scripts and data can be accessed at https://doi.org/10.5281/zenodo.15100450 (Abdalrahem et al., 2026).

Author Contributions

Sébastien Duplessis, Pascal Frey, and Fabien Halkett conceived and designed the study. Axelle Andrieux designed and performed most of the molecular experiments. Ronan Becheler, Ammar Abdalrahem, Axelle Andrieux, and Fabien Halkett performed the manual curation of genotyping results. Axelle Andrieux, Ronan Becheler, and Fabien Halkett did the preliminary analyses of the data. Ammar Abdalrahem performed the final data analyses with help and advice from Benoit Marçais, Ronan Becheler, Solenn Stoeckel, and Fabien Halkett. Ammar Abdalrahem, Kadiatou Schiffer-Forsyth, and Fabien Halkett prepared the manuscript. All authors revised and approved the manuscript.

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We have used AI tools to proofread, suggest synonyms, check grammar and spelling, and debug R script. All suggestions made by these tools have been scrutinized and cross-checked by the authors.

553 **Conflicts of Interest**

554 The authors declare no conflicts of interest

555 **Benefit-Sharing Statement**

556 The advantage of this study is that its analysis code is an automated workflow and data are publicly

557 accessible.

References

Aanen, D. K., & Hoekstra, R. F. (2007). Why Sex Is Good: On Fungi and Beyond. In *Sex in Fungi* (pp. 527–534). John Wiley & Sons, Ltd. https://doi.org/10.1128/9781555815837.ch32

Abdalrahem, A., Halkett, F., & KadiatouSF. (2026). *ammarabdalrahem/poplar_rust: V3.0.0* [Computer software]. Zenodo. https://doi.org/10.5281/zenodo.15100450

Ali, S., Gladieux, P., Rahman, H., Saqib, M. S., Fiaz, M., Ahmad, H., Leconte, M., Gautier, A., Justesen, A. F., Hovmøller, M. S., Enjalbert, J., & de Vallavieille-Pope, C. (2014). Inferring the contribution of sexual reproduction, migration and off-season survival to the temporal maintenance of microbial populations: A case study on the wheat fungal pathogen *Puccinia striiformis* f.sp. *tritici*. *Molecular Ecology*, *23*(3), 603–617. https://doi.org/10.1111/mec.12629

Arnaud-Haond, S., Duarte, C. M., Alberto, F., & Serrão, E. A. (2007). Standardizing methods to address clonality in population studies. *Molecular Ecology*, *16*(24), 5115–5139. https://doi.org/10.1111/j.1365-294X.2007.03535.x

Ashu, E. E., & Xu, J. (2015). The roles of sexual and asexual reproduction in the origin and dissemination of strains causing fungal infectious disease outbreaks. *Infection, Genetics and Evolution*, *36*, 199–209. https://doi.org/10.1016/j.meegid.2015.09.019

Bahri, B., Leconte, M., Ouffroukh, A., De Vallavieille-Pope, C., & Enjalbert, J. (2009). Geographic limits of a clonal population of wheat yellow rust in the Mediterranean region. *Molecular Ecology*, *18*(20), 4165–4179. https://doi.org/10.1111/j.1365-294X.2009.04267.x

Bailleul, D., Stoeckel, S., & Arnaud-Haond, S. (2016). RClone: A package to identify MultiLocus
Clonal Lineages and handle clonal data sets in r. *Methods in Ecology and Evolution*, *7*(8),
966–970. https://doi.org/10.1111/2041-210X.12550

Balloux, F., Lehmann, L., & de Meeûs, T. (2003). The Population Genetics of Clonal and Partially
Clonal   Diploids.   *Genetics*,   *164*(4),   1635–1644.
https://doi.org/10.1093/genetics/164.4.1635

Barbuti, R., Mautner, S., Carnevale, G., Milazzo, P., Rama, A., & Sturmbauer, C. (2012).
Population dynamics with a mixed type of sexual and asexual reproduction in a fluctuating
environment. *BMC Evolutionary Biology*, *12*(1), 49. https://doi.org/10.1186/1471-2148-
12-49

Barrès, B., Halkett, F., Dutech, C., Andrieux, A., Pinon, J., & Frey, P. (2008). Genetic structure of
the poplar rust fungus *Melampsora larici-populina*: Evidence for isolation by distance in
Europe and recent founder effects overseas. *Infection, Genetics and Evolution*, *8*(5), 577–
587. https://doi.org/10.1016/j.meegid.2008.04.005

Becheler, R., Guillemin, M.-L., Stoeckel, S., Mauger, S., Saunier, A., Brante, A., Destombe, C.,
& Valero, M. (2020). After a catastrophe, a little bit of sex is better than nothing: Genetic
consequences of a major earthquake on asexual and sexual populations. *Evolutionary
Applications*, *13*(8), 2086–2100. https://doi.org/10.1111/eva.12967

Becheler, R., Masson, J.-P., Arnaud-Haond, S., Halkett, F., Mariette, S., Guillemin, M.-L., Valero,
M., Destombe, C., & Stoeckel, S. (2017). ClonEstiMate, a Bayesian method for quantifying

rates of clonality of populations genotyped at two-time steps. *Molecular Ecology*

Resources, *17*(6), e251–e267. https://doi.org/10.1111/1755-0998.12698

Becheler, R., Xhaard, C., Klein, E. K., Hayden, K. J., Frey, P., De Mita, S., & Halkett, F. (2016).

Genetic signatures of a range expansion in natura: When clones play leapfrog. *Ecology and*

Evolution, *6*(18), 6625–6632. https://doi.org/10.1002/ece3.2392

Billiard, S., López-Villavicencio, M., Hood, M. E., & Giraud, T. (2012). Sex, outcrossing and

mating types: Unsolved questions in fungi and beyond. *Journal of Evolutionary Biology*,

25(6), 1020–1038. https://doi.org/10.1111/j.1420-9101.2012.02495.x

Bolker, B. M., Brooks, M. E., Clark, C. J., Geange, S. W., Poulsen, J. R., Stevens, M. H. H., &

White, J.-S. S. (2009). Generalized linear mixed models: A practical guide for ecology and

evolution. *Trends in Ecology & Evolution*, *24*(3), 127–135.

https://doi.org/10.1016/j.tree.2008.10.008

Butlin, R. (2002). The costs and benefits of sex: New insights from old asexual lineages. *Nature*

Reviews Genetics, *3*(4), 311–317. https://doi.org/10.1038/nrg749

Castel, M., Mailleret, L., Andrivon, D., Ravigné, V., & Hamelin, F. M. (2014). Allee Effects and

the Evolution of Polymorphism in Cyclic Parthenogens. *The American Naturalist*, *183*(3),

E75–E88. https://doi.org/10.1086/674828

Clarkson, D. B., Fan, Y., & Joe, H. (1993). A remark on algorithm 643: FEXACT: an algorithm

for performing Fisher's exact test in r x c contingency tables. *ACM Trans. Math. Softw.*,

19(4), 484–488. https://doi.org/10.1145/168173.168412

Culotta, E., & Hines, P. (1998). The Evolution of Sex. *Science*, *281*(5385), 1979–1979.

https://doi.org/10.1126/science.281.5385.1979

Drenth, A., McTaggart, A. R., & Wingfield, B. D. (2019). Fungal clones win the battle, but

recombination wins the war. *IMA Fungus*, *10*(1), 18. https://doi.org/10.1186/s43008-019-

0020-8

Evett, I., & Weir, B. S. (Bruce S.). (1998). *Interpreting DNA evidence: Statistical genetics for*

forensic scientists. Sinauer Associates. https://cir.nii.ac.jp/crid/1130282269131400192

Fontyn, C., Zippert, A.-C., Delestre, G., Marcel, T. C., Suffert, F., & Goyeau, H. (2022). Is

virulence phenotype evolution driven exclusively by Lr gene deployment in French

Puccinia   triticina   populations?   *Plant   Pathology*,   *71*(7),   1511–1524.

https://doi.org/10.1111/ppa.13599

Foucaud, J., Jourdan, H., Breton, J. L., Loiseau, A., Konghouleux, D., & Estoup, A. (2006). RARE

SEXUAL REPRODUCTION EVENTS IN THE CLONAL REPRODUCTION SYSTEM

OF INTRODUCED POPULATIONS OF THE LITTLE FIRE ANT. *Evolution*, *60*(8),

1646–1657. https://doi.org/10.1111/j.0014-3820.2006.tb00509.x

Fry, W. E. (2020). Phytophthora infestans: The itinerant invader; "late blight": the persistent

disease. *Phytoparasitica*, *48*(1), 87–94. https://doi.org/10.1007/s12600-019-00778-3

Gavino, P. D., Smart, C. D., Sandrock, R. W., Miller, J. S., Hamm, P. B., Lee, T. Y., Davis, R. M.,

& Fry, W. E. (2000). Implications of Sexual Reproduction for Phytophthora infestans in

638  the United States: Generation of an Aggressive Lineage. *Plant Disease*, *84*(7), 731–735.

639  https://doi.org/10.1094/PDIS.2000.84.7.731

640 Gladieux, P., Feurtey, A., Hood, M. E., Snirc, A., Clavel, J., Dutech, C., Roy, M., & Giraud, T.

641  (2015). The population biology of fungal invasions. *Molecular Ecology*, *24*(9), 1969–1986.

642  https://doi.org/10.1111/mec.13028

643 Goyeau, H., Halkett, F., Zapater, M.-F., Carlier, J., & Lannou, C. (2007). Clonality and host

644  selection in the wheat pathogenic fungus *Puccinia triticina*. *Fungal Genetics and Biology*,

645  *44*(6), 474–483. https://doi.org/10.1016/j.fgb.2007.02.006

646 Hacquard, S., Petre, B., Frey, P., Hecker, A., Rouhier, N., & Duplessis, S. (2011). The Poplar-

647  Poplar Rust Interaction: Insights from Genomics and Transcriptomics. *Journal of*

648  *Pathogens*, *2011*, 716041. https://doi.org/10.4061/2011/716041

649 Halkett, F., Kindlmann, P., Plantegenest, M., Sunnucks, P., & Simon, J. C. (2006). Temporal

650  differentiation and spatial coexistence of sexual and facultative asexual lineages of an

651  aphid species at mating sites. *Journal of Evolutionary Biology*, *19*(3), 809–815.

652  https://doi.org/10.1111/j.1420-9101.2005.01055.x

653 Halkett, F., Plantegenest, M., Bonhomme, J., & Simon, J.-C. (2008). Gene flow between sexual

654  and facultatively asexual lineages of an aphid species and the maintenance of reproductive

655  mode variation. *Molecular Ecology*, *17*(12), 2998–3007. https://doi.org/10.1111/j.1365-

656  294X.2008.03798.x

Halkett, F., Simon, J.-C., & Balloux, F. (2005). Tackling the population genetics of clonal and

partially clonal organisms. *Trends in Ecology & Evolution*, *20*(4), 194–201.

https://doi.org/10.1016/j.tree.2005.01.001

Hamelin, F. M., Bisson, A., Desprez-Loustau, M., Fabre, F., & Mailleret, L. (2016). Temporal

niche differentiation of parasites sharing the same plant host: Oak powdery mildew as a

case study. *Ecosphere*, *7*(11), e01517. https://doi.org/10.1002/ecs2.1517

Hartfield, M., & Keightley, P. D. (2012). Current hypotheses for the evolution of sex and

recombination. *Integrative Zoology*, *7*(2), 192–209. https://doi.org/10.1111/j.1749-

4877.2012.00284.x

Hörandl, E. (2013). Meiosis and the Paradox of Sex in Nature. In *Meiosis*. IntechOpen.

https://doi.org/10.5772/56542

Jombart, T., Devillard, S., & Balloux, F. (2010). Discriminant analysis of principal components:

A new method for the analysis of genetically structured populations. *BMC Genetics*, *11*(1),

94. https://doi.org/10.1186/1471-2156-11-94

Kamvar, Z. N., Brooks, J. C., & Grünwald, N. J. (2015). Novel R tools for analysis of genome-

wide population genetic data with emphasis on clonality. *Frontiers in Genetics*, *6*.

https://www.frontiersin.org/articles/10.3389/fgene.2015.00208

Kamvar, Z. N., Tabima, J. F., & Grünwald, N. J. (2014). Poppr: An R package for genetic analysis

of populations with clonal, partially clonal, and/or sexual reproduction. *PeerJ*, *2*, e281.

https://doi.org/10.7717/peerj.281

Keightley, P. D., & Eyre-Walker, A. (2000). Deleterious Mutations and the Evolution of Sex.

Science, *290*(5490), 331–333. https://doi.org/10.1126/science.290.5490.331

Kronauer, D. J. C., Pierce, N. E., & Keller, L. (2012). Asexual reproduction in introduced and

native populations of the ant Cerapachys biroi. *Molecular Ecology*, *21*(21), 5221–5235.

https://doi.org/10.1111/mec.12041

Lehto, M. P., & Haag, C. R. (2010). Ecological differentiation between coexisting sexual and

asexual strains of Daphnia pulex. *Journal of Animal Ecology*, *79*(6), 1241–1250.

https://doi.org/10.1111/j.1365-2656.2010.01726.x

Lodé, T. (2013). Adaptive Significance and Long-Term Survival of Asexual Lineages.

Evolutionary Biology, *40*(3), 450–460. https://doi.org/10.1007/s11692-012-9219-y

Lorrain, C., Gonçalves dos Santos, K. C., Germain, H., Hecker, A., & Duplessis, S. (2019).

Advances in understanding obligate biotrophy in rust fungi. *New Phytologist*, *222*(3),

1190–1206. https://doi.org/10.1111/nph.15641

Louet, C., Saubin, M., Andrieux, A., Persoons, A., Gorse, M., Pétrowski, J., Fabre, B., De Mita,

S., Duplessis, S., Frey, P., & Halkett, F. (2023). A point mutation and large deletion at the

candidate avirulence locus AvrMlp7 in the poplar rust fungus correlate with poplar RMlp7

resistance breakdown. *Molecular Ecology*, *23*(10), 2472–2483.

https://doi.org/10.1111/mec.16294

Lynch, M., Bürger, R., Butcher, D., & Gabriel, W. (1993). The Mutational Meltdown in Asexual

Populations.   *Journal   of   Heredity*,   *84*(5),   339–344.

https://doi.org/10.1093/oxfordjournals.jhered.a111354

Mboup, M., Leconte, M., Gautier, A., Wan, A. M., Chen, W., De Vallavieille-Pope, C., &

Enjalbert, J. (2009). Evidence of genetic recombination in wheat yellow rust populations

of a Chinese oversummering area. *Fungal Genetics and Biology*, *46*(4), 299–307.

https://doi.org/10.1016/j.fgb.2008.12.007

Nei, M. (1978). Estimation of Average Heterozygosity and Genetic Distance from a Small Number

of Individuals. *Genetics*, *89*(3), 583–590.

Otto, S. P. (2009). The Evolutionary Enigma of Sex. *The American Naturalist*, *174*(S1), S1–S14.

https://doi.org/10.1086/599084

Peck, J. R. (1994). A Ruby in the Rubbish: Beneficial Mutations, Deleterious Mutations and the

Evolution of Sex. *Genetics*, *137*(2), 597–606.

Peck, J. R., Yearsley, J. M., & Waxman, D. (1998). Explaining the geographic distributions of

sexual   and   asexual   populations.   *Nature*,   *391*(6670),   889–892.

https://doi.org/10.1038/36099

Pereyra, R. T., Rafajlović, M., De Wit, P., Pinder, M., Kinnby, A., Töpel, M., & Johannesson, K.

(2023). Clones on the run: The genomics of a recently expanded partially clonal species.

Molecular Ecology, *32*(15), 4209–4223. https://doi.org/10.1111/mec.16996

Pernaci, M., De Mita, S., Andrieux, A., Pétrowski, J., Halkett, F., Duplessis, S., & Frey, P. (2014).

Genome-wide patterns of segregation and linkage disequilibrium: The construction of a

linkage genetic map of the poplar rust fungus *Melampsora larici-populina. Frontiers in*

Plant Science, 5. https://www.frontiersin.org/articles/10.3389/fpls.2014.00454

Persoons, A., Hayden, K. J., Fabre, B., Frey, P., De Mita, S., Tellier, A., & Halkett, F. (2017). The

escalatory Red Queen: Population extinction and replacement following arms race

dynamics in poplar rust. *Molecular Ecology, 26*(7), 1902–1918.

https://doi.org/10.1111/mec.13980

Pierre, J.-S., Stoeckel, S., & Wajnberg, E. (2022). The advantage of sex: Reinserting fluctuating

selection in the pluralist approach. *PLOS ONE, 17*(8), e0272134.

https://doi.org/10.1371/journal.pone.0272134

Pinon, J., & Frey, P. (2005). Interactions between poplar clone and Melampsora populations and

their implications for breeding for durable resistance. In *Rust Diseases of Willow and*

Poplar (pp. 139–154). https://doi.org/10.1079/9780851999999.0139

Platt, V., & Jeschke, J. M. (2014). Are exotic species red queens? *Ethology Ecology & Evolution,*

26(2–3), 101–111. https://doi.org/10.1080/03949370.2014.898699

Pritchard, J. K., Stephens, M., & Donnelly, P. (2000). Inference of Population Structure Using

Multilocus Genotype Data. *Genetics, 155*(2), 945–959.

https://doi.org/10.1093/genetics/155.2.945

Rieux, A., & Balloux, F. (2016). Inferences from tip-calibrated phylogenies: A review and a

practical guide. *Molecular Ecology*, *25*(9), 1911–1924. https://doi.org/10.1111/mec.13586

Ristaino, J. B. (2002). Tracking historic migrations of the Irish potato famine pathogen,

Phytophthora infestans. *Microbes and Infection*, *4*(13), 1369–1377.

https://doi.org/10.1016/S1286-4579(02)00010-2

Rouger, R., Reichel, K., Malrieu, F., Masson, J. P., & Stoeckel, S. (2016). Effects of complex life

cycles on genetic diversity: Cyclical parthenogenesis. *Heredity*, *117*(5), Article 5.

https://doi.org/10.1038/hdy.2016.52

Roughgarden, J. (1991). The Evolution of Sex. *The American Naturalist*, *138*(4), 934–953.

https://doi.org/10.1086/285261

Saitou, N., & Nei, M. (1987). The neighbor-joining method: A new method for reconstructing

phylogenetic trees. *Molecular Biology and Evolution*, *4*(4), 406–425.

https://doi.org/10.1093/oxfordjournals.molbev.a040454

Saleh, D., Milazzo, J., Adreit, H., Fournier, E., & Tharreau, D. (2014). South-East Asia is the

center of origin, diversity and dispersion of the rice blast fungus, Magnaporthe oryzae. *New*

Phytologist, *201*(4), 1440–1456. https://doi.org/10.1111/nph.12627

Saleh, D., Xu, P., Shen, Y., Li, C., Adreit, H., Milazzo, J., Ravigné, V., Bazin, E., Nottéghem, J.-

L., Fournier, E., & Tharreau, D. (2012). Sex at the origin: An Asian population of the rice

blast fungus Magnaporthe oryzae reproduces sexually. *Molecular Ecology*, *21*(6), 1330–

1344. https://doi.org/10.1111/j.1365-294X.2012.05469.x

Saubin, M., Coville, J., Xhaard, C., Frey, P., Soubeyrand, S., Halkett, F., & Fabre, F. (2024). A
mechanistic-statistical approach to infer dispersal and demography from invasion
dynamics, applied to a plant pathogen. *Peer Community Journal*, *4*.
https://doi.org/10.24072/pcjournal.356

Schurko, A., Neiman, M., & Logsdon, J. (2009). Signs of sex: What we know and how we know
it. *Trends in Ecology & Evolution*, *24*, 208–217. https://doi.org/10.1016/j.tree.2008.11.010

Shahapure, K. R., & Nicholas, C. (2020). Cluster Quality Analysis Using Silhouette Score. *2020*
IEEE 7th International Conference on Data Science and Advanced Analytics (DSAA), 747–
748. https://doi.org/10.1109/DSAA49011.2020.00096

Shick, J. M., HOFFMANN, R. J., & LAMB, A. N. (1979). Asexual Reproduction, Population
Structure, and Genotype-Environment Interactions in Sea Anemones. *American Zoologist*,
19(3), 699–713. https://doi.org/10.1093/icb/19.3.699

Simon, J.-C., Baumann, S., Sunnucks, P., Hebert, P. D. N., Pierre, J.-S., Le Gallic, J.-F., &
Dedryver, C.-A. (1999). Reproductive mode and population genetic structure of the cereal
aphid Sitobion avenae studied using phenotypic and microsatellite markers. *Molecular*
Ecology, *8*(4), 531–545. https://doi.org/10.1046/j.1365-294x.1999.00583.x

Simon, J.-C., Delmotte, F., Rispe, C., & Crease, T. (2003). Phylogenetic relationships between
parthenogens and their sexual relatives: The possible routes to parthenogenesis in animals:
ROUTES TO PARTHENOGENESIS IN ANIMALS. *Biological Journal of the Linnean*
Society, *79*(1), 151–163. https://doi.org/10.1046/j.1095-8312.2003.00175.x

Simon, J.-C., Rispe, C., & Sunnucks, P. (2002). Ecology and evolution of sex in aphids. *Trends in*

Ecology & Evolution, *17*(1), 34–39. https://doi.org/10.1016/S0169-5347(01)02331-X

Simon, J.-C., Stoeckel, S., & Tagu, D. (2010). Evolutionary and functional insights into

reproductive strategies of aphids. *Comptes Rendus. Biologies*, *333*(6–7), 488–496.

https://doi.org/10.1016/j.crvi.2010.03.003

Song, Y., Scheu, S., & Drossel, B. (2011). Geographic parthenogenesis in a consumer-resource

model for sexual reproduction. *Journal of Theoretical Biology*, *273*(1), 55–62.

https://doi.org/10.1016/j.jtbi.2010.12.020

Steenackers, J., Steenackers, M., Steenackers, V., & Stevens, M. (1996). Poplar diseases,

consequences on growth and wood quality. *Biomass and Bioenergy*, *10*(5), 267–274.

https://doi.org/10.1016/0961-9534(95)00121-2

Stoeckel, S., Becheler, R., Bocharova, E., & Barloy, D. (2024). GenAPoPop 1.0: A user-friendly

software to analyse genetic diversity and structure from partially clonal and selfed

autopolyploid organisms. *Molecular Ecology Resources*, *24*(1), e13886.

https://doi.org/10.1111/1755-0998.13886

Stoeckel, S., & Masson, J.-P. (2014). The Exact Distributions of FIS under Partial Asexuality in

Small Finite Populations with Mutation. *PLOS ONE*, *9*(1), e85228.

https://doi.org/10.1371/journal.pone.0085228

Stoeckel, S., Porro, B., & Arnaud-Haond, S. (2021). The discernible and hidden effects of clonality

on the genotypic and genetic states of populations: Improving our estimation of clonal

rates. *Molecular Ecology Resources*, *21*(4), 1068–1084. https://doi.org/10.1111/1755-
0998.13316

Suffert, F., Goyeau, H., Sache, I., Carpentier, F., Gélisse, S., Morais, D., & Delestre, G. (2018).
Epidemiological trade-off between intra- and interannual scales in the evolution of
aggressiveness in a local plant pathogen population. *Evolutionary Applications*, *11*(5),
768–780. https://doi.org/10.1111/eva.12588

Taris, B. (1968). Contribution à l'étude des rouilles des Populus observées en France. *Annales Des*
Épiphyties, *19*, 5–54.

Tilquin, A., & Kokko, H. (2016). What does the geography of parthenogenesis teach us about sex?
Philosophical Transactions of the Royal Society B: Biological Sciences, *371*(1706),
20150538. https://doi.org/10.1098/rstb.2015.0538

Waits, L. P., Luikart, G., & Taberlet, P. (2001). Estimating the probability of identity among
genotypes in natural populations: Cautions and guidelines. *Molecular Ecology*, *10*(1), 249–
256. https://doi.org/10.1046/j.1365-294X.2001.01185.x

Weir, B. S., & Cockerham, C. C. (1984). Estimating F-Statistics for the Analysis of Population
Structure. *Evolution*, *38*(6), 1358–1370. https://doi.org/10.2307/2408641

Xhaard, C., Andrieux, A., Halkett, F., & Frey, P. (2009). Characterization of 41 microsatellite loci
developed from the genome sequence of the poplar rust fungus, *Melampsora larici-*
populina. *Conservation Genetics Resources*, *1*, 21–25. https://doi.org/10.1007/s12686-
009-9005-z

Xhaard, C., Barrès, B., Andrieux, A., Bousset, L., Halkett, F., & Frey, P. (2012). Disentangling

the genetic origins of a plant pathogen during disease spread using an original molecular

epidemiology approach. *Molecular Ecology*, *21*(10), 2383–2398.

https://doi.org/10.1111/j.1365-294X.2012.05556.x

Xhaard, C., Fabre, B., Andrieux, A., Gladieux, P., Barrès, B., Frey, P., & Halkett, F. (2011). The

genetic structure of the plant pathogenic fungus *Melampsora larici-populina* on its wild

host is extensively impacted by host domestication. *Molecular Ecology*, *20*(13), 2739–

2755. https://doi.org/10.1111/j.1365-294X.2011.05138.x

Supplementary figures

Figure S1

Geographic distribution of *Melampsora larici-populina* isolates collected across France. Each circle represents a sampling site, with size proportional to the number of isolates collected (5, 20, or 50+ isolates). Colors indicate the sampling campaign: blue for the 2009 national survey, orange for the 2011 national survey (n = 45 sites), and teal for the long-term monitoring program in the Durance River valley (surveys conducted in 2004, 2008, and 2020). The red asterisk marks the collection at Cavalaire-sur-Mer in February 2024. Historical isolates collected before 2004 are not shown on this map but were included in the population genetic analyses.

Figure S2

Silhouette analysis for determining the optimal number of genetic clusters (k) in *Melampsora larici-populina*.
Average silhouette width was computed for k = 1 to 20 using K-means clustering applied to the first 10 principal
components of the microsatellite allele frequency matrix. The dashed vertical line indicates the optimal k = 2,
which achieved the highest average silhouette width (≈ 0.4), indicating the most likely distinct and well-separated
cluster structure.

Figure S3

Scatter plot illustrating the membership probability for individuals within each genotypic cluster, with the names of individuals of lower probability.

Figure S4

The regression plot illustrates the relationship between latitude and the proportion of sexual reproduction. This relationship was analyzed using a binomial generalized linear mixed model (GLMM). It was based on data from 2009 and 2011. The size of the points represents sample sizes.

Supplementary Tables

Table S1. The data set for 2,122 *Melampsora larici-populina* individuals used in population genetic analyses includes collection year, geographic location, and genotypes across 21 microsatellite markers.