Cortical motor activity modulates respiration and reduces apnoea in neonates

Coen S. Zandvoort[1], Fatima Usman[1], Shellie Robinson[1], Odunayo Fatunla[1], Eleri Adams[2], Kyle T.S. Pattinson[3,4], Simon F. Farmer[5,6], Caroline Hartley[1,*]

[1] | Department of Paediatrics, University of Oxford, Oxford, United Kingdom

[2] | Newborn Care Unit, John Radcliffe Hospital, Oxford University Hospitals NHS Foundation Trust, Oxford, United Kingdom

[3] | Nuffield Department of Clinical Neurosciences, University of Oxford, Oxford, UK.

[4] | Nuffield Department of Anaesthetics, Oxford University Hospitals NHS Foundation Trust, Oxford, UK.

[5] | Department of Neurology, National Hospital for Neurology and Neurosurgery, London, United Kingdom

[6] | Department of Clinical and Human Neuroscience, UCL Institute of Neurology, London, United Kingdom

[*] | For correspondence: caroline.hartley@paediatrics.ox.ac.uk

Abstract

Respiration is governed by a widespread network of cortical and subcortical structures.

This complex communication between the brain and lungs is altered in pathological

conditions. Apnoea – the cessation of respiration – is a common condition in infants,

particularly those born prematurely. Apnoea in infants is believed to relate to

immaturity of brainstem respiratory centres; involvement of the cortex in respiration in

infants has yet to be explored. We investigated if cortical control of respiration occurs

in newborn humans and whether it relates to apnoea. Using simultaneous

electroencephalography (EEG) and impedance pneumography, we show that cortico-

respiratory coupling is present in premature and term newborns (104 recordings from

68 infants; 34.5 ± 2.6 weeks post-menstrual age), identifying an interplay between

breathing phase and EEG amplitude. We further shed light on the biological meaning

by revealing that the strongest coupling occurs during inspiration and that cortical

activity precedes respiration, with coupling strongest over frontocentral regions. These

findings support the notion that the cortico-respiratory coupling observed here

primarily constitutes communication between cortical motor areas and lung effectors.

Moreover, we show that cortico-respiratory coupling is negatively correlated with the

rate of apnoea, revealing novel mechanistic insight into this common and potentially

life-threatening neonatal pathology.

Introduction

Apnoea – the cessation of respiration – is a potentially life-threatening event[1], affecting more than 50% of preterm infants and some term infants[2]. Apnoeic episodes can result in physiological instability including oxygen desaturation and bradycardia[1], are associated with suppression of brain activity[3], and have been linked to poorer neurodevelopmental outcomes[4]. Apnoea of prematurity is caused by immaturity of the lungs, chest wall, and nervous system[5]. Immaturity of the brainstem plays a significant role in apnoea[1]; however, the relationship with other respiratory-linked brain regions is less clear. In adults with congenital central hypoventilation syndrome[6] or obstructive sleep apnoea[7], a respiratory-linked increase in cortical motor activity suggests that the motor cortex plays an important role in maintaining autonomous respiration.

Recently, it was proposed that communication between the cortex and lungs can be identified and quantified through so-called cortico-respiratory coupling[8,9]. This functional coupling arises when the amplitude of cortical activity is modulated by the respiration phase, or *vice versa*. This phase-amplitude coupling is typically quantified using non-invasive recordings capturing respiratory and neural activity (e.g., magneto- or electroencephalography [EEG]). Studies have demonstrated that the respiratory phase at low frequencies (<1 Hz) relates to higher-frequency oscillations in the cortex[9,10]. Phase-amplitude coupling therefore allows us to quantify the interplay between respiratory and cortical activity. In adults, this coupling can be observed in a wide network of subcortical and cortical regions including motor areas[9]. Moreover, cortico-respiratory coupling is aberrated in pathological conditions[11].

Here, using cortico-respiratory coupling, we investigated whether cortical motor areas play a role in respiration in infants (Figure 1). We hypothesised that the strength of cortico-respiratory coupling is negatively associated with apnoea rate. To this end, we first confirmed the existence of cortico-respiratory coupling in both premature and term infants. To investigate if the coupling relates to the central motor drive, we further studied the spatial localisation, timing throughout the respiratory cycle and coupling directionality. Finally, we examined the relationship between cortico-respiratory coupling and apnoea rate.

78
79 **Figure 1. Schematic of the data acquisition setup (left panel) and main findings (right panel).**
80 *Each recording consisted of cortical activity (using electroencephalography) and respiratory activity*
81 *(using impedance pneumography).*

Results and Discussion

Respiratory and cortical activity exhibit phase-amplitude coupling in infants

We analysed simultaneous 8-channel resting-state EEG (to measure cortical activity)

and impedance pneumography (IP; to measure respiration) in 68 infants (28-42 weeks

postmenstrual age [PMA] at time of recording) on 104 occasions. We first assessed

whether and how EEG and IP are interrelated using cross-frequency coupling[9,12,13]. In

line with findings in adults[9], we expected that the amplitude of oscillatory brain activity

is modulated as a function of the respiratory cycle. That is, we expected to detect an

EEG component that systematically varied with timing within the respiratory cycle. To

this end, we identified individual breaths (and thus respiratory phase) from the IP

signal[14]. Through averaging the EEG (time-locked to the IP signal) across breaths, we

observed that EEG amplitude was modulated in the delta- (0.5-4 Hz) and theta-band

(4-8 Hz) frequencies depending on the respiratory phase (Figure 2A-B). This analysis

can be seen as a first indication that the EEG amplitude in specific frequency ranges

changes systematically with the respiratory phase.

99
Figure 2. Cortico-respiratory coupling in infants*. A) Electroencephalographic (EEG) amplitude*
modulation depending on the respiratory cycle and frequency. EEG amplitude is averaged over breaths,
recordings and channels FCz and Cz. The blue graph below is the impedance pneumographic signal
time-locked from breath to breath and averaged over breaths and recordings. Grey and white bars
below indicate expiration (Ex; in grey) and inspiration (In; in white). ***B)*** *The time-resolved amplitude*
modulations averaged within four frequency bands (delta: 0.5-4 Hz, theta: 4-8 Hz, alpha: 8-13 Hz, beta:
13-25 Hz), focusing on channels FCz and Cz again. Shaded light-green area is the standard error
across recordings after pooling over channels. ***C)*** *Spatial map of coherence-based phase-amplitude*
*coupling (PAC) between the respiratory phase (ω*_{respiratory phase}*) and EEG amplitude (ω*_{EEG amplitude}*) – see*
Figure S1 for the pan-spectral PAC estimates. Spectra are the mean over all recordings. ***D)*** *Spatial*
map for the corresponding statistical significance relative to surrogate PAC obtained through epoch
shuffling (reported on a logarithmic scale).

We then statistically confirmed this association between EEG and respiration

using phase-amplitude coupling (PAC). Using cross-frequency coherence, we

observed significant PAC between the phase at the mean respiratory frequency and

the EEG amplitude in delta and theta frequency bands (Figure 2C-D). PAC was

strongest at central electrodes (channels FCz and Cz; Figure 2C-D). In line with the

results presented in Figure 2A, we did not find significant PAC at higher EEG

frequencies such as the alpha (8-13 Hz) and beta (13-25 Hz) frequency bands (Figure

S1).

These findings confirm that cortico-respiratory coupling occurs in infants. Whilst

we are the first to show that cortico-respiratory coupling already exists in early life,

adult studies have revealed widespread brain-respiratory networks covering both
cortical and subcortical areas, as well as pan-spectral components of oscillatory brain
activity (2-150 Hz)[9]. These networks have been linked to a diverse range of perceptual,
cognitive and motor processes[8,15-17]. In these studies, respiratory rhythm has been
found to relate to task performance (e.g., better performances are found during a
certain respiratory phase) as well as neural oscillations in the brain[11]. For example,
during an isometric motor task, the strength of cortico-muscular communication at
beta-band frequencies cyclically changes with the respiratory phase[16]. Moreover, the
probability of initiating goal-directed movements relates to cortical motor activity (here,
measured as the readiness potential amplitude) and respiratory phase[17]. These
relationships between task performance, respiration and cortical activity are well-
established and replicated in a variety of animal and human adult studies,
demonstrating the importance of cortico-respiratory coupling.

Cortico-respiratory coupling is strongest during inspiration and driven by the cortex

To further shed light on the functional meaning of cortico-respiratory coupling in
neonates, we next examined the timing and directionality of the coupling. To study
whether coupling strength fluctuates throughout the respiratory cycle, we focused on
the EEG channels with strongest coupling (FCz and Cz), and frequencies where
statistically significant coupling occurs (Figure 2D; alpha level set at 0.001). We
confirmed that PAC changes in a time-dependent manner, observing the strongest
PAC during the inspiratory phase (Figure 3A-B). Such within-breath modulation is
evident for both delta- and theta-frequencies (Figure 3A-B). This indicates that the
strength of cortico-respiratory coupling changes throughout the respiratory cycle.
During the inspiratory phase, neural activation from the central nervous system is
required to drive respiratory motoneurons causing contraction of respiratory muscles.
Given the phasic PAC modulation as seen in Figure 3A-B, this suggests that cortical
areas centred around the vertex play an important role in the neural activation of
respiratory muscles.

153
Figure 3. Timing and directionality of the cortico-respiratory coupling. A) *Phase-amplitude*
coupling modulation (estimated as cross-frequency coherence) throughout the respiratory cycle.
Graphs are colour-coded for the delta- and theta-frequency bands (dark- and light-blue colours
respectively), which are the EEG's primary frequency bands for which we observed statistically
significant PAC (Figure 2C-D). Shaded regions indicate standard error over recordings. PAC was
estimated for EEG channels FCz and Cz. Grey waves at the top and grey bar at the bottom indicate the
phase of the respiratory cycle. Coloured dots indicate points that are statistically significant (alpha level
was set to 0.001). ***B)*** *Corresponding statistical significance analogous to the coloured dots at the top of*
panel A (black dashed line indicates the alpha level). ***C)*** *Spatial map of the cross-frequency phase-*
slope index between respiratory phase ($\omega_{respiratory\ phase}$) and EEG amplitude ($\omega_{EEG\ amplitude}$) (see Figure S2
for the pan-spectral estimates) and ***D)*** *its statistical significance (reported on a logarithmic scale). The*
spectra show the mean over all recordings.

Standard coherence (i.e., our PAC measure) only quantifies linear coupling in
a bidirectional manner and cannot directly show any directionality, meaning that it is
not possible to distinguish whether cortical activity precedes or follows respiratory
activity. To unravel whether the cortical activity drives respiration or *vice versa*, we
employed a cross-frequency version of the phase-slope index[13]. This revealed that
delta- and theta-band EEG amplitude leads respiratory activity (Figure 3C),
demonstrating that the cortical activity drives respiration. We observed the strongest
phase-slope index around central areas, in particular for channel FCz (Figure 3D).
Moreover, in line with the PAC analysis (Figure 2), phase-slope index was not
statistically significant for alpha- and beta-band frequencies (Figure S2).

Considering the cortico-respiratory coupling's spatial location, its timing in the
respiratory cycle and its directionality of action leads us to conclude that this coupling
observed represents the respiratory drive from higher-order motor centres, primarily
encompassing the cortical motor areas. Structurally, the motor cortex connects to

respiratory muscles both directly and indirectly[18]. This allows for functional control over
respiratory muscles (comprising diaphragm and intercostal muscles) via the
corticospinal[19] and reticulospinal tracts[20]. In cat preparations, transection of the
ventrolateral part of the spinal cord (which essentially blocks the brainstem input to
respiratory motoneurons) does not prevent cortical stimulation from eliciting responses
in the respiratory intercostal muscle motoneurons, signifying the cortico-spinal drive[20].
Such motor-evoked potentials can also be elicited by stimulating the cortical motor
areas of humans using transcranial magnetic stimulation[21,22].

Whilst we believe that it is likely that our coupling reflects cortical motor drive,
one might also argue that cortico-respiratory coupling could reflect changes in blood
flow or oxygen saturation which induces 'coupling', where blood flow/oxygen
saturation acts as a confounding factor essentially modulating respiratory and brain
activity in a cycle-dependent manner. It is, however, unlikely that this is the case for
our results given the focal spatial localisation around central electrodes and narrow
spectral bandwidth comprising EEG frequencies between 1-8 Hz. Moreover, the
amplitude modulation of blood flow reveals a different time-frequency representation
over the respiratory cycle (Figure S3) when compared to cortical activity (Figure 2A).

To further our understanding of whether the coupling in infants relates directly
to central motor drive, future work should examine potential coupling in the cortico-
muscular domain. Electrical activity of the diaphragm (Edi) in infants can be recorded
using an Edi catheter[23], as is used in ventilators with neurally adjusted ventilatory
assist[24]. Correlating this electromyographic (EMG) activity from the diaphragm with
brain activity (e.g., EEG) will enable cortico-muscular assessments that can quantify
whether oscillatory components in the EEG and diaphragmatic EMG are phase-locked.
It is of particular interest whether linear coherence can be observed in the frequency
range of 1-8 Hz, which is expected from our PAC findings presented in Figure 2.
Interestingly, such coherence analysis has been conducted at the muscular level in
rodents. Invasively recording activity from the diaphragm and intercostal muscles
revealed strong inter-muscular coherence in the lower frequencies between 1 and 8
Hz[25]. Future work should also establish to what extent the cortical motor areas provide
control over other respiratory functionalities such as altering respiratory rate. In the
current investigation, we focused on how cortical amplitude directly related to the
respiratory phase but phase-amplitude coupling of cortical activity has also been

revealed to play a role in adjusting respiratory rate[26] (i.e., respiratory rate rather than
individual breaths).

Cortico-respiratory coupling relates to apnoea in infants

Having identified significant cortical-respiratory coupling in infants, we then tested our
main hypothesis - that the strength of the cortico-respiratory coupling is negatively
associated with apnoea. Apnoea of prematurity can lead to periods of physiological
instability when apnoea is accompanied by bradycardia and desaturation[5]. These
episodes may have long-term neurological effects[4] and, hence, it is important to better
understand the pathophysiology. In addition to pulmonary immaturity, apnoea of
prematurity has been primarily attributed to immaturity and weakened neural drive
from the respiratory centres in the brainstem. These respiratory centres are principally
located in the medulla oblongata and pons within the brainstem, which ensure the
periodical activation of lung muscles to enable respiration. However, as our results
indicate, further central input to respiration is provided by higher-order brain areas
such as the cortical motor areas.

To test our hypothesis, apnoea rate was estimated over a time window of up to
24 hours (infants have their vital signs continuously monitored at the neonatal care
unit), whereas cortical-respiratory coupling was assessed on the same day from EEG
recordings with an average duration of 1.7 ± 0.6 hours. Choosing different time
windows ensured that any relationship between apnoea rate and cortical-respiratory
coupling was not the mere result of the number of breaths included in the analysis.
For instance, when the same respiratory signal is used in both analyses, a higher
apnoea rate would mean that fewer windows are used for the PAC estimation which
could result in an increased magnitude. We selected PAC pooled over EEG channels
Cz and FCz since their coupling magnitudes were the highest.

We found support for our hypothesis that apnoea and cortico-respiratory-
coupling are related. PAC was negatively associated with apnoea rate (t(98):-2.28,
p:0.02, ρ:-0.22; Figure 4A; this relationship was also identified when focusing on
individual channels [Figure S4] and preterm infants younger than 36 weeks [Figure
S5]). This suggests that a stronger cortico-respiratory coupling may reduce the
likelihood of experiencing apnoeas in infants. This negative association provides

further support that our measures of cortico-respiratory coupling (Figures 2-3) are
related to central motor drive. In this case, cortical motor activity assists the
brainstem's central pattern generators (CPGs) to establish increased control over the
respiratory muscles, which in our analysis, is reflected as an association with apnoea
rate. The motor cortex can exert coordination over the CPGs through its downstream
projections to the thalamus[27], which, in turn is connected to the preBötzinger complex
(the structure that primarily hosts the CPGs[28]).

253
Figure 4. Cortico-respiratory coupling dependency on apnoea rate and postmenstrual age. A)
Relationship between apnoea rate and statistically significant phase-amplitude coupling (PAC; defined
as coherence encompassing delta- and theta-band frequencies). Each blue dot is the data of an
individual recording (data of 68 infants were included from 104 test occasions). Black linear graph is
the best fit of the linear mixed-effects model (fixed factors: coupling averaged over statistically
significant samples from channels FCz and Cz [see Figure S4 for channel-specific relationships], data
length to determine apnoea rate, mode of ventilation, and postmenstrual age; random factor: infant).
Panel title contains the test statistic of the predictor's regression slope (t), its significance (p), and partial
correlation coefficient (ρ). B-C) PAC spectra between the respiratory phase ($\omega_{respiratory\ phase}$) and EEG
amplitude ($\omega_{EEG\ amplitude}$) for B) preterm (58 recordings; 33.0-36.86 weeks), and C) very preterm (28
recordings; 28.0-32.86 weeks) infants. Both spectra are averaged over recordings and EEG channels
FCz and Cz.

In theory, the strength of cortical-respiratory coupling and its inverse
relationship with apnoeas could reflect a shared relationship with PMA, where both

coupling strength and apnoea rate change depending on PMA. The apnoea rate is
expected to decrease with increased PMA across a broad age range[29]. It is also
plausible that the strength of cortico-respiratory coupling (i.e., PAC magnitude) may
change with PMA, with younger (i.e., preterm) infants potentially having weaker
coupling. However, in our dataset, we observe statistically significant PAC in preterm
infants (33^{+0}-36^{+6} weeks^{+days}; Figure 4B), and very preterm infants (28^{+0}-32^{+6}
weeks^{+days}; Figure 4C). Moreover, PAC and apnoea rate were not significantly
associated with PMA (Figure S6A-B, linear mixed-effects analysis). Similarly, PAC and
apnoea rate were not significantly related to postnatal age (PNA; defined as the age
in weeks from birth; Figure S6C-D). Whilst future studies should investigate the
relationship between cortico-respiratory coupling and age across an even broader age
range, the lack of significant associations here suggests that the relationship we
observed between apnoea rate and cortico-respiratory coupling is not determined by
age.

It is known that most apnoeas in infants occur during active sleep[6,30] and delta-
and theta-band frequencies in EEG are strongly related to sleep state[31]. We speculate
that cortico-respiratory coupling may change depending on sleep state, giving rise to
more episodes of apnoea during certain sleep stages. Indeed, adult studies have
suggested that cortical respiratory drive may occur predominantly during
wakefulness[6,32]. Future work should study the effects of sleep stages on cortico-
respiratory coupling to shed further light on the role of cortical motor drive for
respiration in infants.

In summary, we demonstrate the existence of cortico-respiratory coupling in
newborn infants. We believe that this coupling is related to central motor drive due to
its spatial location (predominantly over frontocentral EEG channels), timing (during
inspiration) and directionality (with the cortex driving respiration). Consistent with our
hypothesis, we found a significant relationship between apnoea rate and coupling
strength, with infants with stronger cortico-respiratory coupling less likely to experience
apnoea. This has far-reaching implications for our mechanistic understanding of
apnoea of prematurity as it demonstrates that cortical motor areas play a pivotal role
in apnoea in infants.

Data and code availability

- Due to ethical restrictions, the raw data that support the findings of this study are available on reasonable request. Requests can be made directly to the corresponding author (caroline.hartley@paediatrics.ox.ac.uk) or directed to the Paediatric Neuroimaging Group (University of Oxford) through the institutional email: neonatal.research@ouh.nhs.uk. Researchers will be required to agree to destroy the data within the time limits stipulated in the ethically approved study protocols. Upon agreement, data will be provided within 1 month.
- Summary-level data (before regression model adjustments of Figure 4), and channel- and recording-specific PAC spectra are provided at https://gitlab.com/paediatric_neuroimaging/cortico_respiratory_coupling.
- The codes for all analyses are available at: https://gitlab.com/paediatric_neuroimaging/cortico_respiratory_coupling. The algorithm for inter-breath interval detection is online at: https://gitlab.com/paediatric_neuroimaging/identify_ibi_from_ip.
- Any additional information required to reanalyse the data reported in this paper is available from the lead contact upon request.

Acknowledgements

This work is funded by the Wellcome Trust and Royal Society through a Sir Henry Dale Fellowship awarded to Caroline Hartley (grant number: 213486/Z/18/Z). Fatima Usman is funded by the Commonwealth Scholarship Commission. Odunayo Fatunla is funded by the Clarendon Fund in partnership with the Lincoln College Kingsgate Graduate Scholarship. We would like to thank members of the Paediatric Neuroimaging Group (University of Oxford) for help with data collection, and the babies and their parents who took part in this research.

Author contributions

Conceptualization: CSZ and CH; **Data curation:** FU, SR, OF, and CH; **Formal analysis:** CSZ; **Funding acquisition:** CH; **Investigation:** FU, SR, and OF; **Methodology:** CSZ; **Project administration:** CH; **Software:** CSZ; **Resources:** EA,

332   KTSP and SFF; **Supervision:** CH and EA; **Visualization:** CSZ; **Writing – original**

333   **draft:** CSZ; **Writing – review & editing:** all authors

335   ***Declaration of interests***

336   The authors declare no competing interests.

Material & Methods

Participants and study design

This study was conducted at the Newborn Care Unit of the John Radcliffe Hospital, Oxford University Hospitals NHS Foundation Trust, Oxford, UK. The National Research Ethics Service approved the study (references: 19/LO/1085; 12/SC/0447). Eligible families were approached and given verbal and written details about the study. Parents provided written consent to take part in the study. The study conformed to the standards set by the Declaration of Helsinki and Good Clinical Practice.

Clinically stable infants aged between 28- and 42-weeks postmenstrual age (PMA) at the time of the test occasion were eligible to be included in the study. Infants were excluded if they had intraventricular haemorrhage grades III or IV, hypoxic ischaemic encephalopathy, congenital malformations, there was a known history of maternal substance misuse during pregnancy, or at the time of the study, they were receiving mechanical ventilation, opioid analgesics, or antibiotics (as a surrogate marker of infection). These conditions are known to alter respiratory and brain activity. We enrolled 74 infants who were recorded on 123 separate occasions (range: 1-8 recordings per infant). Fifty-one infants were included in one or two test occasions; 23 infants took part in the ongoing *Breathing and Brain Development* study where they are studied approximately once a week whilst in the Newborn Care Unit (included on 3±2 [median ± interquartile range] occasions). Demographic details of all infants can be found in Table 1. Parts of this dataset have been reported earlier in Zandvoort et al. [33] to address a different research question.

Factors	
Age	
Postmenstrual age at recording (weeks)	34.5 ± 2.6
Gestational age at birth (weeks)	31.1 ± 3.4
Postnatal age at recording (weeks)	3.4 ± 2.2
Birthweight (g)	1,726 ± 816

Sex	
Females	27
Males	47
Mode of delivery	
Normal vaginal delivery	16
Vaginal assisted (ventouse/forceps/kiwi)	4
Elective Caesarean section	7
Emergency Caesarean section	47
Apgar scores	
Apgar at 1 minute	7.2 ± 2.2
Apgar at 5 minutes	9.1 ± 1.4
Apgar at 10 minutes	9.7 ± 0.8
Respiratory distress syndrome	
Ongoing condition	39
Past treated	48
No	36
Persistent pulmonary hypertension	
Past treated	5
No	118
Infection	
None	48
Treated	75
Respiratory support	
High flow therapy	25
Low flow therapy	16
Spontaneous ventilation	82
Caffeine received on the day of the EEG recording	
Yes	59
No	64

Table 1. Infant demographics. *Reported values are mean and standard deviation or number of babies*
or recordings. Values are reported over recordings for ages (postmenstrual, gestational, and postnatal),
respiratory distress syndrome, persistent pulmonary hypertension, infection and respiratory support.
Birthweight, sex, mode of delivery, and Apgar scores are provided over infants.

Data recordings

During each test occasion, approximately one to two hours of simultaneous
electroencephalography (EEG) and vital signs (respiration, heart rate and oxygen
saturation) were recorded. We continuously recorded the vital signs throughout their
hospital stay of the infants who took part in the *Breathing and Brain Development*
study.

Vital signs were recorded for each infant using Phillips IntelliVue MX800 or
MX750 monitors. These data were continuously downloaded from the vital signs
monitor using an electronic data capture software (iXtrend, ixitos, Germany). Both
electrocardiograph (ECG; sampling frequency: 250 Hz) and impedance
pneumography (IP; sampling frequency: 62.5 Hz) were recorded with three electrodes
placed on the infant's chest. Peripheral photoplethysmography was recorded at 125
Hz from a probe placed on the infant's foot or hand.

EEG was recorded from DC to 400 Hz using a SynAmps RT 64-channel
headbox and amplifiers and CURRYscan7 neuroimaging suite (Compumedics
Neuroscan) at a sampling rate of 2 kHz. Eight electrodes were placed at Cz, CPz, C3,
C4, Oz, FCz, T3, and T4. The ground electrode was placed at FPz and the reference
electrode at Fz. The infant's scalp was cleaned with NuPrep gel (D.O. Weaver and
Co., Aurora, USA) to achieve impedances of 5-10 kΩ. A conductive paste (Elefix EEG
paste, Nihon Kohden, Tokyo, Japan) was used to attach disposable Ag/AgCl cup EEG
electrodes (Ambu Neuroline, Ballerup, Denmark) to the infant's scalp. EEG was
recorded for an average of 1.7 ± 0.7 hours (mean ± SD). ECG was co-registered with
our EEG recordings using a single electrode (Ambu Neuroline, Ballerup, Denmark)
placed on the infant's chest (referenced to Fz).

To synchronise the vital signs and EEG recordings, we registered
approximately 30 annotations simultaneously on both recordings using a bespoke
triggering device (the PiNe box)[34]. Annotations were used to mark resting state as well
as stimulus-evoked activity (these responses were used in other studies to investigate
infant brain maturity – see Zandvoort *et al.* [33] – but were used here only to synchronise
the EEG and vital signs recordings). The PiNe box enables synchronisation with an
average precision of 105 ± 14 ms (mean ± SD)[34]. Annotation timings were matched
between both vital signs and EEG recordings. Next, to further improve synchronisation,

we utilised the ECG signals which were recorded on both the vital signs and EEG

recording devices. To do so, ECG epochs were created from -10 to 10 seconds around

each annotation. We aligned the ECG signals around each of these annotation epochs

using cross correlations, providing a temporal delay between vital signs and EEG[34].

Temporal synchronisation between vital signs and EEG recordings was achieved by

taking the median of the time differences across all annotations.

From the total of 123 recordings, we excluded 19 recordings due to (a) missing

IP recordings (5 recordings), (b) missing annotations (9 recordings), and (c)

inaccuracies in EEG-IP syncing (5 recordings; standard deviations over annotations

exceeding 1 sec for annotation alignment or 0.1 sec during ECG alignment). Hence,

data comprising 104 recordings from 68 infants were used in the analysis.

Breath and apnoea identification

IP signals were processed to identify breaths (including apnoeas). To this end, we

used an algorithm validated to detect inter-breath intervals in infants[14]. Briefly, the

algorithm attenuates noise introduced by movements and cardiac activity by filtering

the signal. An adaptive amplitude threshold then identifies individual breaths. This

threshold was set at 0.4 times the standard deviation of the signal over the preceding

15 breaths. Finally, a support vector machine algorithm was applied to all long (here,

15 seconds or greater) inter-breath intervals to separate episodes of apnoea

(cessation of respiration) and periods of low amplitude erroneously identified as

apnoeas due to noise or shallow breathing. The inter-breath intervals marked as

noise/shallow breathing were discarded from further analysis. The model was trained

and validated on IP signals acquired during a clinical trial[35], which was a completely

independent data sample to the one used in the current study.

Phase-amplitude coupling

In line with Osipova *et al.* [12], we defined phase-amplitude coupling (PAC) as cross-

frequency coherence between a low-frequency phase and high-frequency amplitude.

Note that the term "phase-amplitude coupling" is used interchangeably in the literature

to mean the coupling principle of the statistical relationship between phases and

amplitudes as well as the actual measure to detect this. We reserve the abbreviation

for the latter. For our PAC measure, this meant that coherence was computed between

the filtered IP signal and instantaneous amplitude of the EEG signal at some higher

frequency (i.e., the second-order moment between the IP phase and EEG amplitude).

More formally, in our case PAC was defined as:

$$C(f_1, f_2) = \frac{|\sum_{n=1}^{N} X(f_1) * Y(f_1, f_2)^*|}{\sqrt{\sum_{n=1}^{N} |X(f_1)|^2 * \sum_{n=1}^{N} |Y(f_1, f_2)|^2}}$$ Eq. 1

with X being the IP signal at frequency f_1, Y the instantaneous EEG amplitude, and $*$

the complex conjugate. This instantaneous amplitude was obtained using the

magnitude of the Hilbert transform after narrowly filtering the data at frequency f_2. This

component is often filtered with a bandwidth that relates to the low-frequency

component f_1. Following earlier recommendations, the filter bandwidth of the high-

frequency component (at f_2) was equal to a single time the frequency of the low-

frequency component f_1[36] (e.g., when f_1 equals 5 Hz the filter bandwidth of f_2 is set to

5 Hz).

Data were segmented into 4-sec windows (50% overlap), which meant that the

frequency resolution was equal to 0.25 Hz (allowing to capture low delta-band

frequencies), which comprised around four breaths. To focus on regular respiration,

we discarded epochs which contained fewer than three detected breaths. An epoch

was also removed from further analysis when its standard deviation exceeded 3 times

the mean or median of the standard deviation of all epochs (which was done

separately for the EEG and IP). Finally, we rejected any epoch containing an absolute

value greater than 10 times the mean or median of the standard deviation of all epochs.

Before cross-spectrum computation, EEG and IP epochs were convolved with a

wavelet constructed from a Hanning window.

Changes in PAC throughout the respiratory cycle

To investigate how cortico-respiratory coupling changes throughout the respiratory

cycle, we estimated PAC at 50 equally spaced timepoints within the respiratory cycle.

To this end, we time-locked both EEG and IP to each inter-breath interval as identified

by the algorithm of Adjei *et al.* [14]. At each respiratory cycle phase, cross- and auto-

spectra were obtained to estimate PAC over all breaths within a recording. We limited

the analysis to EEG channels FCz and Cz, because they exhibited the highest PAC

magnitude (Figure 2), as well as statistically significant frequency-frequency samples

relative to the surrogate PAC (see Figure 2D for the *p*-values of this statistical

comparison). PAC was computed within the different frequency bands (delta: 0.5-4

Hz; and theta: 4-8 Hz) to see whether its modulation was specific for each frequency

band. The same preprocessing steps apply as for PAC computation described earlier

apart from the fact that data were segmented into 2/3 sec windows (so that temporal

changes would appear within the respiratory cycle).

Directionality of phase-amplitude coupling

To assess whether the cortical activity was preceding or trailing relative to the

respiratory activity, we employed a cross-frequency version of the phase-slope index.

Briefly, this metric estimates the phase lags over narrow frequency ranges and

quantifies whether they increase or decrease over this range by estimating the phase-

frequency slopes. Increasing or decreasing slopes signify changing time lags

depending on the frequency of the oscillation and can therefore be indicative of

whether a signal leads or trails another signal. We used the phase-slope index (Ψ) as

proposed by Jiang *et al.* [13], which was defined as:

$$\Psi(f_1, f_2) = imag\left(\sum_{f_2 - \frac{\beta}{2}}^{f_2 + \frac{\beta}{2}} \tilde{C}^*(f_1, f_2)\, \tilde{C}(f_1, f_2 + \Delta f)\right) \qquad \text{Eq. 2}$$

481 with $\tilde{C}$ being the coherency (i.e., the normalised cross-spectrum) of C defined in Eq. 1.

482 Δf is the frequency resolution and β the bandwidth used to estimate the phase slope.

483 In line with previous recommendations by Jiang *et al.* [13], we set β to 1 Hz (i.e., four

484 times the frequency resolution).

Surrogate analysis

To assess the statistical significance of PAC (i.e., coherence) and phase-slope index at each of the EEG channels, we performed a surrogate analysis under the null hypothesis that the means of the true and surrogate PAC (and phase-slope index) were equal. The surrogate analysis consisted of randomly shuffling epochs of the EEG amplitude after which PAC was computed with the IP signal. The random shuffling of the EEG amplitude epochs destroys any temporal relationships whilst retaining the signals' characteristics. We obtained a surrogate spectrum for each recording. Next, the true and surrogate spectra were statistically compared on a sample-by-sample basis using dependent *t*-tests[37] (using the FieldTrip toolbox – version 20210325). This comparison was performed over all recordings. Statistical significance was defined using the false discovery rate correction (with an alpha level set at 0.001).

Statistical analysis of relationships between cortico-respiratory coupling and apnoea

We tested our main hypothesis that PAC is negatively associated with apnoea rate. Vital signs were continuously monitored for the infants that took part in the *Breathing and Brain Development* study, which allowed us to analyse their vital signs in longer time windows than the EEG. We here selected up to 24 hours of vital signs from the day of the EEG recording (mean ± standard deviation: 9.5 ± 9.9 hours) and expressed this as a rate per hour. We then constructed linear mixed-effects models to study the relationships between PAC and apnoea rate. We included the coherence magnitude as a fixed factor, which was averaged over the statistically significant samples from channels FCz and Cz (see Figure S4 for the channel-specific relationships). As fixed effects, we included (1) data length of the respiratory signal, (2) mode of ventilation ('Self-ventilating', 'Low flow', 'High flow'; Figure S7), and (3) PMA (Figure S6). Infant ID was included as a random effect. Data length, mode of ventilation and PMA were included as confounding factors. P-values (for the regression coefficient of PAC) and partial correlation coefficients were reported from these models. The normality assumption was verified with Q-Q plots. We also tested the linearity assumption (see Supplementary text). For visualisation (Figure 4), we used the plotAdjustedResponse function in MATLAB to adjust for the fixed factors (calculating a separate model without

517   the random effect of an infant). Regression lines are visualised by calculating the line-

518   of-best-fit on the adjusted data.

References

1. Martin, R.J., and Abu-Shaweesh, J.M. (2005). Control of breathing and neonatal apnea. Neonatology *87*, 288-295.
2. Finer, N.N., Higgins, R., Kattwinkel, J., and Martin, R.J. (2006). Summary proceedings from the apnea-of-prematurity group. Pediatrics *117*, S47-S51.
3. Zandvoort, C.S., Dereymaeker, A., Baxter, L., Jansen, K., Naulaers, G., de Vos, M., and Hartley, C. (2024). Apnoea suppresses brain activity in infants. Imaging Neuroscience *2*, 1-14.
4. Williamson, M., Poorun, R., and Hartley, C. (2021). Apnoea of prematurity and neurodevelopmental outcomes: Current understanding and future prospects for research. Frontiers in Pediatrics *9*, 755677.
5. Martin, R.J., Abu-Shaweesh, J.M., and Baird, T.M. (2004). Apnoea of prematurity. Paediatric Respiratory Reviews *5*, S377-S382.
6. Tremoureux, L., Raux, M., Hudson, A.L., Ranohavimparany, A., Straus, C., and Similowski, T. (2014). Does the supplementary motor area keep patients with Ondine's curse syndrome breathing while awake? PLoS One *9*, e84534.
7. Launois, C., Attali, V., Georges, M., Raux, M., Morawiec, E., Rivals, I., Arnulf, I., and Similowski, T. (2015). Cortical drive to breathe during wakefulness in patients with obstructive sleep apnea syndrome. Sleep *38*, 1743-1749.
8. Kluger, D.S., Balestrieri, E., Busch, N.A., and Gross, J. (2021). Respiration aligns perception with neural excitability. Elife *10*, e70907.
9. Kluger, D.S., and Gross, J. (2021). Respiration modulates oscillatory neural network activity at rest. PLoS biology *19*, e3001457.
10. Tort, A.B., Brankačk, J., and Draguhn, A. (2018). Respiration-entrained brain rhythms are global but often overlooked. Trends in neurosciences *41*, 186-197.
11. Brændholt, M., Kluger, D.S., Varga, S., Heck, D.H., Gross, J., and Allen, M.G. (2023). Breathing in waves: Understanding respiratory-brain coupling as a gradient of predictive oscillations. Neuroscience & Biobehavioral Reviews *152*, 105262.
12. Osipova, D., Hermes, D., and Jensen, O. (2008). Gamma power is phase-locked to posterior alpha activity. PloS one *3*, e3990.
13. Jiang, H., Bahramisharif, A., van Gerven, M.A., and Jensen, O. (2015). Measuring directionality between neuronal oscillations of different frequencies. Neuroimage *118*, 359-367.
14. Adjei, T., Purdy, R., Jorge, J., Adams, E., Buckle, M., Fry, R.E., Green, G., Patel, C., Rogers, R., and Slater, R. (2021). New method to measure interbreath intervals in infants for the assessment of apnoea and respiration. BMJ Open Respiratory Research *8*, e001042.
15. Perl, O., Ravia, A., Rubinson, M., Eisen, A., Soroka, T., Mor, N., Secundo, L., and Sobel, N. (2019). Human non-olfactory cognition phase-locked with inhalation. Nature human behaviour *3*, 501-512.
16. Kluger, D.S., and Gross, J. (2020). Depth and phase of respiration modulate cortico-muscular communication. Neuroimage *222*, 117272.
17. Park, H.-D., Barnoud, C., Trang, H., Kannape, O.A., Schaller, K., and Blanke, O. (2020). Breathing is coupled with voluntary action and the cortical readiness potential. Nature communications *11*, 289.

18. Urfy, M.Z., and Suarez, J.I. (2014). Breathing and the nervous system. Handbook of clinical neurology *119*, 241-250.

19. Evans, K.C., Shea, S.A., and Saykin, A.J. (1999). Functional MRI localisation of central nervous system regions associated with volitional inspiration in humans. The Journal of Physiology *520*, 383-392.

20. Aminoff, M., and Sears, T. (1971). Spinal integration of segmental, cortical and breathing inputs to thoracic respiratory motoneurones. The Journal of Physiology *215*, 557-575.

21. Gandevia, S., and Rothwell, J. (1987). Activation of the human diaphragm from the motor cortex. The Journal of physiology *384*, 109-118.

22. Welch, J.F., Argento, P.J., Mitchell, G.S., and Fox, E.J. (2020). Reliability of diaphragmatic motor-evoked potentials induced by transcranial magnetic stimulation. Journal of Applied Physiology *129*, 1393-1404.

23. Essouri, S., Baudin, F., Mortamet, G., Beck, J., Jouvet, P., and Emeriaud, G. (2019). Relationship between diaphragmatic electrical activity and esophageal pressure monitoring in children. Pediatric Critical Care Medicine *20*, e319-e325.

24. Fang, S.-J., Chen, C.-C., Liao, D.-L., and Chung, M.-Y. (2023). Neurally adjusted ventilatory assist in infants: a review article. Pediatrics & Neonatology *64*, 5-11.

25. Rice, A., Fuglevand, A.J., Laine, C.M., and Fregosi, R.F. (2011). Synchronization of presynaptic input to motor units of tongue, inspiratory intercostal, and diaphragm muscles. Journal of neurophysiology *105*, 2330-2336.

26. Hammer, M., Schwale, C., Brankačk, J., Draguhn, A., and Tort, A.B. (2021). Theta-gamma coupling during REM sleep depends on breathing rate. Sleep *44*, zsab189.

27. Mckay, L.C., Evans, K.C., Frackowiak, R.S., and Corfield, D.R. (2003). Neural correlates of voluntary breathing in humans. Journal of applied physiology *95*, 1170-1178.

28. Del Negro, C.A., Funk, G.D., and Feldman, J.L. (2018). Breathing matters. Nature Reviews Neuroscience *19*, 351-367.

29. Henderson-Smart, D. (1981). The effect of gestational age on the incidence and duration of recurrent apnoea in newborn babies. Journal of Paediatrics and Child Health *17*, 273-276.

30. Lehtonen, L., and Martin, R.J. (2004). Ontogeny of sleep and awake states in relation to breathing in preterm infants. In 3. (Elsevier), pp. 229-238.

31. Jenni, O.G., Borbély, A.A., and Achermann, P. (2004). Development of the nocturnal sleep electroencephalogram in human infants. American journal of physiology-regulatory, integrative and comparative physiology *286*, R528-R538.

32. Dubois, M., Chenivesse, C., Raux, M., Morales-Robles, A., Nierat, M.-C., Garcia, G., Navarro-Sune, X., Chavez, M., Martinerie, J., and Similowski, T. (2016). Neurophysiological evidence for a cortical contribution to the wakefulness-related drive to breathe explaining hypocapnia-resistant ventilation in humans. Journal of Neuroscience *36*, 10673-10682.

33. Zandvoort, C.S., van der Vaart, M., Robinson, S., Usman, F., Mellado, G.S., Fry, R.E., Worley, A., Adams, E., Slater, R., and Baxter, L. (2024). Sensory event-related potential morphology predicts age in premature infants. Clinical Neurophysiology *157*, 61-72.

34.   Worley, A., Fabrizi, L., Boyd, S., and Slater, R. (2012). Multi-modal pain
measurements in infants. J Neurosci Methods *205*, 252-257.
35.   Hartley, C., Moultrie, F., Hoskin, A., Green, G., Monk, V., Bell, J.L., King, A.R.,
Buckle, M., van der Vaart, M., and Gursul, D. (2018). Analgesic efficacy and
safety of morphine in the Procedural Pain in Premature Infants (Poppi) study:
randomised placebo-controlled trial. The Lancet *392*, 2595-2605.
36.   Zandvoort, C.S., and Nolte, G. (2021). Defining the filter parameters for phase-
amplitude coupling from a bispectral point of view. Journal of Neuroscience
Methods *350*, 109032.
37.   Maris, E., and Oostenveld, R. (2007). Nonparametric statistical testing of EEG-
and MEG-data. J Neurosci Methods *164*, 177-190.
626
627