NewroBus for the brain: humanized TfR1-targeting nanobodies with high BBB permeability and cargo transport capacity

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Abstract

Delivery of therapeutics to the brain is restricted by the blood-brain barrier (BBB). A promising strategy to overcome this involves exploiting receptor-mediated transcytosis pathways, such as those mediated by transferrin receptor 1 (TfR1), which is highly expressed on brain endothelial cells and naturally transports iron-bound transferrin across the BBB. To leverage this mechanism, we immunized camelids with human TfR1 and cloned 470 VHH nanobody sequences from their B cells. From this repertoire, 24 nanobodies (TfR1b-Nbs) were identified that bind cell membrane human TfR1. These nanobodies were produced in CHO-S cells, a mammalian expression system widely used in therapeutic protein production for its ability to support proper folding, disulfide bond formation, and human-compatible post-translational modifications. CHO-S cells also generate very low endotoxin levels, making them suitable for clinical-grade manufacturing. TfR1b-Nbs were screened for binding to human TfR1, lack of interference with transferrin binding and TfR1-mediated iron uptake, and the ability to cross the BBB via human TfR1-mediated transcytosis in newly generated humanized *Tfr1*^{h} knock-in rats.

To improve developability and reduce potential immunogenicity, selected TfR1b-Nbs were humanized and optimized using the AbNatiV algorithm, enhancing humanness, solubility, and VHH-nativeness. Eight optimized TfR1b-Nbs retained BBB permeability and were fused to humanized TNFα inhibitors (TNFI-α or TNFI-β), generating 16 heterodimers. Fusion to TNFI served as a functional readout, confirming that TfR1b-Nbs can shuttle biologically active, BBB-impermeable payloads into the CNS. All heterodimers demonstrated CNS delivery after intravenous administration, and selected constructs also reached the brain via subcutaneous injection, maintaining high serum and CSF levels for up to 72 hours. A pilot study with one heterodimer showed that chronic administration in rats humanized for both transferrin and TfR1 caused no hematological toxicity or signs of anemia—a key safety concern when targeting TfR1. These results establish humanized TfR1b-Nbs—designated *NewroBus*—as promising BBB shuttles for the safe and effective therapeutic delivery of biologics to the brain.

Introduction

Targeting transferrin receptor 1 (TfR1) to enhance drug delivery across the blood-brain barrier (BBB) has emerged as a rapidly advancing area of therapeutic development. This strategy leverages the fact that TfR1 is highly expressed on brain endothelial cells and its transcytosis activity can be exploited to deliver therapeutic molecules into the central nervous system (CNS) (Johnsen *et al*, 2019; Lee *et al*, 2000; Pardridge *et al*, 1991). Several companies are actively pursuing distinct TfR1-mediated strategies to enable CNS penetration of biologics. Notable examples include Denali Therapeutics, which developed the Transport Vehicle (TV) platform utilizing engineered Fc domains that bind TfR1 to drive receptor-mediated transcytosis (Ullman *et al*, 2020); Roche, with its Brain shuttle technology, based on a bispecific monoclonal antibody format (Weber *et al*, 2018); and Apertura Gene Therapy, which employs engineered AAV capsids specifically designed to target human TfR1 and improve CNS gene delivery (Huang *et al*, 2024). BioArctic has also introduced its Brain Transporter™ technology, although mechanistic and efficacy data remain unpublished.

Camelids naturally produce heavy-chain-only antibodies that lack light chains and consist of a single variable domain (VHH) followed by constant domains CH2 and CH3 (Hamers-Casterman *et al*, 1993). The isolated VHH domains, commonly referred to as nanobodies (Nbs), are compact, stable antigen-binding fragments with a typical molecular weight of 12–14 kDa. Compared to conventional monoclonal antibodies (mAbs), nanobodies offer several advantages: (1) their convex paratopes allow binding to recessed or cryptic epitopes, such as receptor binding pockets that are often inaccessible to the flat paratopes of mAbs; (2) they exhibit high stability across a wide pH range; (3) they have low immunogenicity, which can be further minimized through humanization or deimmunization; and (4) their small size and structural robustness make them more amenable to alternative delivery routes, including oral administration. Importantly, nanobodies targeting transferrin receptor 1 (TfR1) have been shown to mediate transcytosis across the BBB (Sehlin *et al*, 2020; Xiao & Gan, 2013), highlighting their potential as molecular buses. Additionally, nanobody-based TfR1 binders are monovalent and thus less likely to interfere with the physiological function of TfR1 in iron transport. Unlike bivalent antibodies, they are less likely to disrupt the transferrin (TF)–TfR1 interaction essential for cellular iron uptake or to promote TfR1 endocytosis, which typically requires engagement of both subunits of the TfR1 dimer. This minimizes the risk of iron deficiency-related toxicities, an important consideration in the development of TfR1-targeted therapeutics.

Based on these properties, we set out to identify and characterize novel anti-TfR1 nanobodies capable of efficiently crossing the BBB, with the long-term goal of using them as “Trojan horses” to deliver otherwise BBB-impermeable therapeutics into the brain.

Results

Identification of TfR1b-Nbs. 470 individual VHH domain clones isolated from PBMCs from 1 llama and 1 alpaca immunized with human TfR1 extracellular domain (Sino Biological, HPLC-11020-H07H) were tested for antigen binding. 106 unique α-TfR1-Nb VHH sequences were identified. The above experiments were performed at Abcore.

HEK293 human cells were transfected with a vector co-expressing human transferrin receptor 1 (hTfR1) and EGFP. Of the 106 α-TfR1 nanobodies (Nbs) produced in bacteria, 24 bound to cell-surface hTfR1 on transfected cells (**Supporting Figures S1**, **S2**, **S3** and **S4**). These nanobodies were designated TfR1b-Nbs (TfR1-binding nanobodies). None of the 24 TfR1b-Nbs cross-reacted with rodent (rat or mouse) Tfr1 (**Supporting Figures S5**, **S6**, **S7** and **S8**). Based on the complementarity-determining regions (CDRs) identities, these 24 TfR1b-Nbs were grouped into ten families (**Figure 1A**).

Assessing the binding of TfR1b-Nbs produced in mammalian cells to human TfR1. Chinese Hamster Ovary (CHO) cells are a preferred system to produce therapeutic proteins due to their ability to generate complex, properly folded proteins with significantly lower endotoxin levels compared to bacterial expression systems, enhancing both the safety and clinical suitability of the final product. CHO-derived biologics have a well-established track record of regulatory approval, making this platform highly reliable for biopharmaceutical manufacturing.

The data in **Table 1** and **Figure 1B and 1C** (showing representative results for 05F02R3 from Family A and 04G05R3 from Family D) demonstrate the high purity and yield of TfR1b-Nbs produced with transient transfection in CHO-S cells, supporting their suitability as a production platform for preclinical and clinical development. Protein production was performed at GenScript.

The biological activity of CHO-S–produced TfR1b-Nbs was assessed by testing their binding to human TfR1. HEK293 cells transfected with hTfR1 and EGFP showed strong staining with TfR1b-Nbs from Families A, B, D, G, and I, but not from Families E, F, J, or L (**Figure 2**). Similar results were obtained using CHEK-ATP089, a stable HEK293/hTfR1 cell line, where TfR1 expression was confirmed by transferrin-FITC staining (**Figure 3**).

Identifying TfR1-Nbs compatible with transferrin-mediated iron transport. TF is the major ferric iron transport protein, which binds ferric ($Fe3^{+}$) ions. The iron binding affinity of transferrin is pH dependent. In a neutral pH environment, TF (apotransferrin) binds iron with high affinity to form iron-bound TF (holotransferrin). In an acidic pH environment, the affinity of iron bound to transferrin decreases, dissociating iron from holotransferrin and releasing it into the environment. The importance of holotransferrin is to sequester Fe3+ ions in a relatively nonreactive and inert state to ensure normal free iron homeostasis in the body. Holotransferrin delivers iron to cells by binding to transferrin receptor (TfR1 or TfR2). Neutral pH at the cell surface promotes binding of holotransferrin to TfR1/TfR2. The receptor-ligand complex enters the cell through receptor-mediated endocytosis and is internalized into endosomes. Relatively lower endosomal pH results in the release of iron. The receptor-ligand complex is recycled to the cell surface, where apotransferrin dissociates from TfR1/TfR2 (Kawabata, 2019). Interference with these processes might lead to unintended toxic effects in patients treated with therapeutics based on TfR1b-Nbs.

To evaluate whether TfR1b-Nbs interfere with TF binding or uptake, we used CHEK-ATP089 cells. Unlabeled TF competes with TF-FITC for receptor binding in a dose-dependent manner—effective at TF/TF-FITC ratios of 3 and 1, but not 0.3—confirming assay sensitivity (**Figure 4**). Using the same setup, we tested whether TfR1b-Nbs (2.5 μM)

could reduce TF-FITC binding at a 1:1 molar ratio. All three Family I TfR1b-Nbs reduced TF-FITC binding, while none of the other 11 tested Nbs (from Families A, B, D, and G) showed interference (**Figure 4**).

We next assessed whether TfR1b-Nbs affect TF uptake, using pHrodo Red-TF and live-cell imaging on the Incucyte system. pHrodo Red fluorescence increases in acidic compartments, providing a readout of endocytic uptake. TF inhibited pHrodo Red-TF uptake in a dose-dependent manner, even at high pHrodo Red-TF/TF ratios (312.5 nM vs. 30 nM) (**Figure 5**). Despite potential imprecision in calculating pHrodo Red-TF molarity, these results confirm that competition can impair uptake. Consistent with binding data, Family I Nb 01B01R3 significantly reduced pHrodo Red-TF uptake (**Figure 5**), though less potently than 30 nM TF. TfR1b-Nbs from Families A, B, D, and G had no impact (**Figure 5**). A Family J Nb, which does not bind TfR1, also showed no effect (**Figure 5**).

Based on these results, TfR1b-Nbs from Families A, B, D, and G appear most suitable for further development, as they do not interfere with TF-TfR1 interactions and iron uptake.

Assessing in vivo BBB permeability of TfR1b-Nbs. Because TfR1b-Nbs do not bind rodent TfR1 (**Figures S5-S8**), rodents are unsuitable for in vivo BBB permeability studies. To overcome this problem, we generated knock-in (KI) rats in which the endogenous rat *Tfr1* gene was replaced with the human *TfR1* gene ($Tfr1^{h}$). The generation and characterization of these KI rats will be described in a separate publication. We used $Tfr1^{h}$ rats to assess whether TfR1b-Nbs from Families A, B, D, and G can cross the BBB. A diverse group of KI rats generated during the breeding and expansion of the $Tfr1^{h}$ line was used in these experiments, including animals of different genotypes—wild-type ($Tfr1^{w/w}$), heterozygous ($Tfr1^{h/w}$), and homozygous ($Tfr1^{h/h}$) for the humanized *Tfr1* allele—as well as both sexes and a range of ages. These animals were intravenously (IV) injected with 1 μL/g of a 40 μM PBS solution of each of the 11 TfR1b-Nbs. At 12–18 hours post-injection, blood was collected, and rats were perfused with PBS. Brain homogenates and serum samples were analyzed by a human TfR1-based ELISA to detect nanobody levels. As shown in **Figure 6A**, all tested TfR1b-Nbs were detected in brain homogenates of $Tfr1^{h}$ rats (even when only one allele was humanized), but not in rats expressing only rat *Tfr1* ($Tfr1^{w/w}$). Serum levels were also markedly higher in $Tfr1^{h}$ rats, likely reflecting increased nanobody stability upon target engagement. **Table 2** shows detailed information about age, sex, genotype of the rats utilized and the nanobodies injected.

In a follow-up experiment, nanobodies 04B05R3 (Family A) and 05D01R3 (Family G) were tested again in young $Tfr1^{h/w}$ or control $Tfr1^{w/w}$ rats. **Table 3** shows detailed information about age, sex, genotype of the rats utilized and nanobodies injected. Samples were collected 44–48 hours post-IV injection, including CSF. As before, serum levels were higher in $Tfr1^{h/w}$ rats, and only these rats showed TfR1b-Nbs in both the brain and CSF (**Figure 6B**). Both nanobodies exhibited remarkably high CSF/serum ratios, with 04B05R3 reaching ~1.5, suggesting preferential brain accumulation. These nanobodies remained detectable 2 days post-injection, reflecting prolonged stability—likely enhanced by target engagement—an uncommon feature for nanobodies, which typically have short half-lives.

Assessing CNS delivery of BBB-impermeable biologics via TfR1b-Nb-directed transcytosis. Figure 7A shows that TNFI-Nb was detected in the serum of both control ($Tfr1^{w/w}$) and humanized ($Tfr1^{h/w}$) rats following intravenous injection (see **Table 4** for experimental details). However, it was undetectable in brain homogenates of either genotype, indicating that TNFI-Nb does not cross the BBB. This absence confirms that the humanized BBB remains functionally restrictive to non-transported proteins. Supporting this, additional

time-course data (not shown) demonstrated that TNFI-Nb becomes undetectable in serum within 12 hours post-injection, consistent with its short systemic half-life.

To visually assess protein distribution between serum and CSF, we ran equal volumes of serum and CSF from *Tfr1*$^{h/h}$ rats on SDS-PAGE and stained the gels with Coomassie Blue. As shown in **Figure 7B**, strong protein bands were observed in serum, corresponding to abundant circulating proteins. In contrast, only a faint band at ~70 kDa was detectable in the CSF, reflecting the low protein content of CSF under physiological conditions and confirming the integrity of the BBB in *Tfr1*$^{h/h}$ rats. These findings further support that TNFI-Nb does not passively diffuse into the brain and reinforce the functionality of the humanized BBB model.

Next, we evaluated whether fusing TNFI-Nb1 to the TfR1b-Nb 04B05R3 (Family A) could enhance brain delivery of TNFI-Nb1. The heterodimer was produced in CHO-S cells by GenScript (**Figure 8A**). To assess CNS transcytosis, we employed a TNFα-specific ELISA. The same protocol used for monomeric TfR1b-Nbs was applied, including IV injection, perfusion, and sample collection. **Table 5** summarizes the age, sex, genotype of the rats used, and the nanobodies injected. The heterodimer was detected in both the brain and CSF of *Tfr1*$^{h/w}$ rats, but not in or control *Tfr1*$^{w/w}$ controls, confirming human TfR1-dependent BBB transport (**Figure 8B**). Like the monomeric 04B05R3, the heterodimer exhibited strong CNS localization, with a CSF/serum ratio of ~1—slightly lower than the monomer (~1.5) yet still indicating efficient brain accumulation.

To further evaluate BBB transcytosis of the heterodimer, we applied a fractionation protocol (Kariolis et al., 2020) used by Denali to assess CNS penetration of AVI-based therapeutics. One hemibrain from each of three *Tfr1*$^{h/w}$ rats was separated into vasculature and vascular-depleted parenchymal fractions. In this experiment, we tested the TNFI-Nb1–linker–04B05R3 heterodimer.

Western blot analysis confirmed effective fractionation: the endothelial marker Glut1 was enriched in the vasculature fraction and depleted in the parenchyma, while Gapdh—a general cellular marker—was enriched in the parenchyma. Cell-type–specific markers in the parenchymal fraction included: Vamp2 (neurons), NmdaR2b (neurons), Iba1(microglia), Eaat2 (astrocytes), and Mbp (oligodendrocytes) (**Figure 9A**). These results indicate successful enrichment of parenchymal CNS cell types, with particularly strong signal for microglial marker. Human TfR1 was detected in all brain fractions of *Tfr1*$^{h/w}$ rats, with highest expression in the vasculature, consistent with its known highest endothelial localization.

ELISA analysis (**Figure 9B**) confirmed the presence of TNFI-Nb1–04B05R3 in brain homogenates, vasculature, and parenchymal fractions of *Tfr1*$^{h/w}$ rats. After normalization to protein content, the parenchymal fraction showed the highest levels, supporting uptake of the heterodimer by CNS cells, likely via human TfR1.

These results support the concept that TfR1b-Nbs can serve as effective "NewroBuses," delivering otherwise BBB-impermeable biologics into the brain.

Assessing TNFI-Nb1-04B05R3 uptake by brain cells. To confirm human TfR1-dependent BBB penetration of TNFI-Nb1-04B05R3, we performed immunohistochemistry (IHC). One hemibrain from each rat used in the experiment shown in **Figure 9** was fixed in 4% paraformaldehyde following perfusion. Brain sections from *Tfr1*$^{h/w}$ and *Tfr1*$^{w/w}$ rats were stained using anti-His tag (for the nanobody) and anti-human TfR1 antibodies to examine nanobody localization and target engagement. TNFI-Nb1–04B05R3 (green) was detected in *Tfr1*$^{h/w}$ but not *Tfr1*$^{w/w}$ brains (**Figure 10A**). **Figure**

10B confirms human TfR1 expression (red) exclusively in *Tfr1*$^{h/w}$ brains. Colocalization of TNFI-Nb1–04B05R3 (green), human TfR1 (red), and the endothelial marker CD31 (purple) was observed in *Tfr1*$^{h/w}$ rats (**Figure 10C**). Merged and zoomed-in images show clear colocalization (white), confirming binding to the brain endothelium in a TfR1-dependent manner.

The presence of the heterodimer in the parenchymal fraction suggests cellular uptake within the CNS. To investigate whether TNFI-Nb1–04B05R3 localizes to specific brain cell types, we performed confocal imaging of cortex and hippocampus sections stained with DAPI (nuclei), NeuN (neurons), GFAP (astrocytes), IBA1 (microglia), and an anti-His tag antibody to detect TNFI-Nb1–04B05R3. Neuronal localization was negligible (data not shown). TNFI-Nb1–04B05R3 colocalizes with GFAP-positive astrocytes (yellow signal) in both cortex and hippocampus of *Tfr1*$^{h/w}$ rats, but not in *Tfr1*$^{w/w}$ controls (**Figure 11**). **Figure 12** shows similar colocalization with IBA1-positive microglia, again only in *Tfr1*$^{h/w}$ rats.

Together, these data demonstrate that TNFI-Nb1–04B05R3 crosses the BBB in a human TfR1-dependent manner, localizing to the CSF—an accessible surrogate for interstitial fluid within the brain parenchyma—with a portion taken up by CNS cells, predominantly astrocytes and microglia. The absence of signal in *Tfr1*$^{w/w}$ rats underscores the requirement for human TfR1. These findings support the potential of TfR1b-Nbs as targeted delivery vehicles for otherwise BBB-impermeable therapeutics.

In Silico humanization and developability optimization of lead TfR1b nanobodies. Humanization and optimization are critical steps to enhance nanobody stability, solubility, and expression, while minimizing immunogenicity—key for safe and effective therapeutic use in humans. Accordingly, TfR1b-Nbs from Families A, B, D, and G underwent in silico humanization and developability optimization using a strategy similar to that applied to TNFI-Nbs (Yin *et al*, 2025). We employed AbNatiV, a machine learning algorithm that predicts both humanness and VHH-nativeness directly from sequence (Aubin Ramon, 2024). Humanness scores above 0.8 indicate high similarity to human variable domains and correlate with lower immunogenicity risk, while scores below 0.8 suggest greater likelihood of immune recognition due to non-human sequence features.

Based on the most favorable starting points for humanness, VHH-nativeness, and CamSol solubility, TFRI1b-Nbs 05F02, 04F01, and 04G05 were selected for TNF1b-Nbs Families A, B, and D, respectively. For Family G, 03E01 and 05D01 were chosen as representatives of the two subfamilies present in this group (**Table 6**).Two complementary mutational sampling strategies were applied: enhanced sampling, which rapidly converges on an optimized humanized sequence, and exhaustive sampling, which systematically evaluates all permissible residue substitutions based on position-specific scoring matrices (PSSMs) from human VH and VHH databases. Both methods identify designs on the Pareto front, balancing increased humanness with preserved VHH-nativeness—critical for maintaining nanobody stability and folding in the absence of a VL domain.

Humanization was restricted to solvent-exposed framework residues, while CDRs were preserved to avoid compromising antigen binding. As a further control for structural integrity post-humanization, the structures of wild-type (WT) and all humanized sequences were modelled with ImmuneBuilder (Abanades *et al*, 2023). The modelled structures were superimposed based on their framework regions, and the root-mean-square deviation (RMSD) calculated for the CDR regions was approximately 1 Å. This value is significantly smaller than the expected modelling accuracy for these regions suggesting minimal or no displacement of the CDR loops because of the humanizing framework mutations.

Following in silico humanization, we used the structural models of both Enhanced and Exhaustive humanized variants as inputs for the CamSol Combination pipeline (Rosace *et al*, 2023), by excluding all CDR regions from the design and by using an alignment of human VH sequences as input. CamSol Combination automatically identifies combinations of mutations predicted to improve solubility and conformational stability, or one of these properties without affecting the other. The apparent melting temperature of in silico mutants was predicted with NanoMelt (Rosace *et al.*, 2023). Of the mutations suggested by this approach, we retained only those that didn't reduce humanness according to AbNatiV scoring (**Table 7**). These variants were produced in CHO-S cells with production performed at GenScript**.**

We evaluated these humanized mutants for binding to TfR1 on the cell surface of transiently transfected HEK cells (**Figure 13**) and found that 13 optimized mutants retain binding to cell-membrane TfR1. The 8 mutant mutants highlighted in gray in **Table 7** (A2, A3, A4, B1, B2, D1, D2 and D3) exhibit the best features for drug development, including solubility and humanness. Consequently, we assessed their BBB permeability in vivo using our *Tfr1* humanized rats. All eight TfR1b-Nbs exhibited high CSF levels, indicating human TfR1-dependent BBB transcytosis (**Figure 14**). Among them, 05F02_enhanced WT-CDR3STEM-W demonstrated the highest CSF/serum ratio. **Table 8** provides details on the age, sex, genotype of the rats used, and the specific nanobodies administered.

The eight optimized TfR1b-Nbs were fused to either TNFI-α or TNFI-β—two humanized TNFα inhibitors derived from TNFI-Nb1 (Yin *et al.*, 2025)—to generate 16 unique heterodimers (**Figure 15A**). We first evaluated the ability of these heterodimers to bind cell-surface human TfR1, comparing their binding profiles to those of the corresponding parental humanized TfR1b-Nbs used in their construction. As shown in **Figure 15B**, several heterodimers, particularly those incorporating TfR1b-A4, -B1, and -B2, exhibited reduced binding to human TfR1. Whether this reduction can be mitigated by altering the heterodimer design—such as modifying the linker length or sequence—remains to be investigated in future studies.

For the purposes of this study, we prioritized heterodimers containing TfR1b-D1, -D2, -D3, and -A2 for further evaluation. Notably, all selected heterodimers retained TNFα inhibitory activity comparable to TNFI-α and TNFI-β (**Figure 15C**) (Yin *et al.*, 2025)—supporting their continued development as therapeutic candidates.

Evaluating the potential of subcutaneous administration. We selected two heterodimers—TNFI-β-TfR1b-A2 and TNFI-β-TfR1b-D1—for further evaluation of tissue distribution and subcutaneous (SQ) delivery. To test the feasibility of SQ administration—a more patient-friendly alternative to IV—we leveraged its known advantages: ease of use, steady absorption, reduced clinical resource needs, lower discomfort, and cost efficiency. Approved biologics such as Humira, Enbrel, and Simponi exemplify the success of this route in chronic conditions. In pilot experiments, rats expressing human TfR1 were injected SQ with 1 µL/g of a 40 µM of either TNFI-β-TfR1b-A2 or TNFI-β-TfR1b-D1 solution in PBS. Heterodimers levels were measured 72 hours post-injection. The results are summarized in **Figure 16**. In *Tfr1*$^{h/w}$ rats, CSF/serum ratios were 0.14 for TNFI-β-TfR1b-A2 and 0.37 for TNFI-β-TfR1b-D1, with average CSF concentrations of 18.8 pM (TNFI-β-TfR1b-A2) and 5.5 pM (TNFI-β-TfR1b-D1). TNFI-β-TfR1b-A2 achieved higher total systemic levels, while TNFI-β-TfR1b-D1 demonstrated superior BBB permeability. Only the heart (and possibly kidney) showed modest signs of tissue distribution dependent on human TfR1 expression. Importantly, heterodimers were detectable in both serum and CSF three days post-injection, indicating markedly extended in vivo stability compared to conventional nanobodies. Furthermore, the consistently higher serum levels in humanized *Tfr1* rats vs. wild-type rats suggest that target-mediated stabilization contributes to this prolonged half-life.

Assessing in vivo hematotoxicity. Interfering with TF-TfR1 interaction and/or uptake can have toxic effects, especially anemia. Homozygous null *Tfr1* mice display an embryonic lethal phenotype, while hypo-transferrinemic mice suffer severe anemia (Trenor *et al*, 2000). Although TfR1b-Nabs from Families D and A do not interfere with TF binding or uptake by CHEK-ATP089 cells (), we assessed the potential hematotoxicity of heterodimers in vivo. A complete blood count (CBC), conducted by the IRVS core at Rutgers, was performed on ~4 months-old rats humanized for both TF and TfR1, and administered either TNFI-β-TfR1b-A2 or PBS. This is because hematotoxicity, particularly anemia, is potentially influenced by TfR1 binding. The experimental design is detailed in **Table 9**. The initial CBC (Day -3) established baseline values. Subsequent CBCs (Day 1, Day 17 after three injections, and Day 24 after four injections) monitored for acute and long-term hematotoxic effects. All values remained within normal physiological ranges for rats, indicating that the humanization of *Tf* and *Tfr1* genes has preserved normal blood cell functions. TNFI-β-TfR1b-A2 did not cause significant changes in CBC parameters, including anemia indicators (RBC, HGB, HCT, **Figure 17**), compared to PBS controls. Given that these tests were performed in rats expressing human TF and TFf1, the findings are expected to closely reflect the INNs TNFI-β-TfR1b-A2' effects in humans, especially regarding holo-TF-TfR1 interactions and iron uptake. This supports the likelihood of low hematotoxicity in humans, particularly given that the therapeutic dosage of heterodimers may be significantly lower than those tested in these experiments.

In summary, these results demonstrate that heterodimers retain BBB permeability when administered subcutaneously and are viable for human therapeutic use, combining target specificity, extended stability, and ease of delivery.

Discussion

This study outlines a systematic approach to the selection, optimization, and in vivo validation of humanized TfR1b nanobodies (TfR1b-Nbs) as delivery vehicles for CNS-targeted biologics. The goal was to identify TfR1b-Nbs that not only bind human TfR1 with high specificity and affinity but also exhibit favorable developability profiles and the capacity to cross the BBB in a human TfR1-dependent manner—essential features for their therapeutic translation.

We began with immunization of camelids, cloning and identification of 24 TfR1b-Nbs spanning ten sequence-diverse families. These candidates had been previously validated for their ability to bind human TfR1 on the cell surface. For therapeutic use, production was transitioned from bacteria to CHO-S cells to ensure appropriate folding, post-translational modifications, and reduced immunogenicity—critical parameters for clinical-grade biologics.

Next, we tested each CHO-produced nanobody for its ability to bind cell-surface TfR1, while ensuring that this binding did not interfere with Tf binding or TfR1-mediated iron uptake—key physiological functions of TfR1 that must be preserved for therapeutic safety. Nanobodies that maintained Tf-TfR1 binding and iron transport were then evaluated for their ability to cross the BBB in vivo using newly generated *Tfr1*^{h} KI, in which the endogenous rodent TfR1 gene was replaced with the human ortholog. A subset of TfR1b-Nbs demonstrated robust CNS penetration in a human TfR1-dependent manner.

To enhance their clinical potential, the most promising TfR1b-Nbs were subjected to in silico humanization and developability optimization using *AbNatiV*, a machine learning tool that simultaneously evaluates sequence humanness and VHH-nativeness. Mutations were limited to framework regions to preserve antigen-binding activity, and final variants were selected based on high humanness scores, solubility, and retention of structural integrity. Eight optimized TfR1b-Nbs from three families passed this rigorous selection and retained their ability to bind cell-surface TfR1 and cross the BBB in vivo. These were advanced to generate heterodimers with either TNFI-α or TNFI-β—two humanized nanobodies targeting TNFα. All 16 resulting heterodimers were initially screened for BBB permeability, and several exhibited favorable pharmacokinetic profiles.

We then focused on two heterodimers—TNFI-β-TfR1b-A2 and TNFI-β-TfR1b-D1—for further characterization, including SQ delivery, a route preferred for chronic therapy due to its convenience, cost-effectiveness, and patient compliance. Remarkably, these heterodimers reached therapeutically relevant concentrations in the CSF and serum 72 hours post-injection, with CSF/serum ratios ranging from 0.14 to 0.37, and significantly extended systemic stability—properties not typical of conventional nanobodies.

Disruption of the TF–TfR1 interaction can result in hematological toxicity, particularly anemia, due to impaired iron uptake. The hematotoxicity study suggest that TfR1b-Nbs are well-tolerated and unlikely to impair iron homeostasis in humans. However, further studies will be required to confirm safety across a broader range of constructs and doses.

These findings validate the humanized TfR1b-Nbs as effective BBB shuttles and demonstrate that their fusion to therapeutic payloads like TNFI-Nbs preserves both BBB permeability and target engagement. Moreover, the ability to deliver these constructs via the SQ route highlights their translational potential for non-invasive, long-term CNS therapy. The pipeline—from immunization to in vivo validation and humanization—is summarized in **Figure 18**.

Conclusion

Together, our findings demonstrate that humanized TfR1b-Nbs enable efficient, TfR1-dependent transcytosis across the BBB, delivering otherwise impermeable therapeutic payloads into the CNS. These nanobodies exhibit favorable

pharmacokinetics, minimal hematotoxicity, and sustained CNS exposure following subcutaneous administration. Moreover, their ability to localize to astrocytes and microglia highlights their potential for cell-targeted intervention in neuroinflammatory and neurodegenerative diseases. These TfR1b-Nbs have been exclusively licensed to NanoNewron and are collectively referred to as **NewroBus** molecules. As such, NewroBus represents a promising platform for advancing the delivery of biologics to the brain.

Material sand Methods

Cell lines

Cell Line	Description	Vendor	Catalog No.
HEK293T	Human embryonic kidney cells with SV40 large T-antigen	ATCC	CRL-3216
WEHI-13VAR	Mouse fibrosarcoma TNF-sensitive reporter cell line	ATCC	CRL-2168
HEK-ATP089	HEK293 cells stably expressing human TfR1	Acro Biosystems	CHEK-ATP089

Expression constructs

Construct	Description	Vendor / Source	Catalog No. (VB)
Human TfR1 + EGFP	Mammalian expression vector for human TfR1 + EGFP	VectorBuilder	VB211221-1144bue
Mouse Tfr1 + EGFP	Mammalian expression vector for mouse Tfr1 + EGFP	VectorBuilder	VB220126-1168dfv
Rat Tfr1 + EGFP	Mammalian expression vector for rat Tfr1 + EGFP	VectorBuilder	VB220126-1169wgz

These constructs can be retrieved from VectorBuilder's plasmid lookup portal:

https://en.vectorbuilder.com/design/retrieve.html

Primary Antibodies

Target	Antibody Description	Vendor	Catalog No.
His tag	Anti-His tag (APC-conjugated)	R&D Systems	IC050A
His tag	Anti-His tag (Alexa Fluor 488-conjugated)	R&D Systems	IC050G
His tag	Rabbit mAb	Cell Signaling Technology (CST)	12698
His tag	Alexa Fluor 488 Rabbit mAb (D3I1O)	CST	14930
VHH domain	Goat Anti-Alpaca IgG, VHH domain	Jackson ImmunoResearch	128-005-230
VHH domain (biotinylated)	Biotinylated VHH domain	Jackson ImmunoResearch	128-065-232
Human TfR1	APC-conjugated monoclonal	Invitrogen	17071942
Mouse Tfr1	APC-conjugated monoclonal	Invitrogen	17071182
Rat Tfr1	APC-conjugated monoclonal	Invitrogen	17071082
Human TfR1	Mouse antibody	CST	54505S
Iba1 (microglia)	SPICA Dye™ 594-conjugated Rabbit antibody	Wako (FUJIFILM)	012-28401
GFAP (astrocytes)	Mouse monoclonal	BD Biosciences	556327
CD31 (endothelial cells)	Alexa Fluor® 647-conjugated recombinant antibody	Abcam	ab305210
Glut1	Rabbit monoclonal	CST	73015
GAPDH	Rabbit monoclonal	Sigma-Aldrich	G9545
MBP (oligodendrocytes)	Rabbit monoclonal	CST	78896
NMDA Receptor 2	Rabbit monoclonal	CST	4212
EAAT2 (astrocytes)	Rabbit polyclonal	Synaptic Systems	250203, 104202

Secondary antibodies

Target	Antibody Description	Vendor	Catalog No.
Goat IgG	Anti-Goat IgG Donkey SULFO-TAG	Meso Scale Discovery (MSD)	R32AG

Target	Antibody Description	Vendor	Catalog No.
Rabbit IgG	Anti-Rabbit IgG Donkey SULFO-TAG	MSD	R32AB
Mouse IgG	Goat anti-Mouse IgG (H+L), Alexa Fluor™ 594, HCA	Invitrogen	A-11032
Rabbit IgG	HRP-conjugated secondary antibody	CST	7074
Rabbit IgG	HRP-conjugated secondary antibody	Southern Biotech	OB405005

Recombinant proteins and detection reagents

Protein / Reagent	Description	Vendor	Catalog No.
Human TfR1	Biotinylated recombinant protein	Acro Biosystems	TFR-H82E5
Human TNFα	Recombinant active trimer	Acro Biosystems	TNA-H5228
Human Transferrin	Unlabeled	Jackson ImmunoResearch	009000050
Human Transferrin (FITC-conjugated)	Fluorescently labeled	Jackson ImmunoResearch	009090050
Human Transferrin (pHrodo™ Red-labeled)	pHrodo-conjugated transferrin	Thermo Fisher Scientific	P35376
Biotinylated TNFα	ELISA capture antigen	Acro Biosystems	TNA-H82E3-25ug
Biotinylated His Tag	ELISA control	R&D Systems	BAM050
Biotinylated VHH Domain	ELISA control	Jackson ImmunoResearch	128-065-232

General reagents and lab chemicals

Reagent	Description	Vendor	Catalog No.
Fugene® HD Transfection Reagent	Non-liposomal transfection reagent	Promega	E2311
Fetal Bovine Serum (FBS)	Heat-inactivated, cell culture supplement	Gibco	A3840102
EDTA	Chelating agent (disodium salt)	Sigma-Aldrich	N6507
DMEM Medium	High-glucose cell culture medium	Corning	10-017-CV
RPMI 1640 Medium	Cell culture medium for lymphoid cells	Corning	10-040-CV
Acridine Orange / Propidium Iodide (AO/PI)	Dual dye viability stain	DeNovix	CD-AO-PI-7.5
Propidium Iodide (PI)	DNA-binding dye for dead cell discrimination	Invitrogen	P3566
Actinomycin D	RNA synthesis inhibitor for cytotoxicity assays	Sigma-Aldrich	A9415
Caspase-3/7 Green Detection Reagent	Fluorescent apoptosis marker	Thermo Fisher Scientific	C10423
Dextran (MW 60,000)	Used in brain fractionation	Sigma-Aldrich	31397
Hygromycin B	selection marker	Sigma-Aldrich	H3272
S1 Buffer Components	Tris base, sucrose, EDTA, EGTA (lab-prepared)		

Consumables and tools for sample collection

Item	Description	Vendor
1 mL Tuberculin Syringe, 25G	For IV injection	Cardinal Health
VACUETTE Blood Collection Set, 25G	For tail vein blood collection	Greiner Bio-One
BD Microtainer SST Tubes	For tail vein serum isolation	Becton Dickinson
BD Vacutainer SST Tubes	For terminal serum isolation	Becton Dickinson
Air-Tite Syringe, 5 mL	For cardiac blood collection	Air-Tite Products Co., Inc.
Monoject Hypodermic Needle, 21G	For cardiac blood collection	Cardinal Health
EXEL Insulin Syringe, 28G	For CSF collection	EXEL

Item	Description	Vendor
Stereotaxic Frame	For CSF collection stabilization	Stoelting Co.
MiniCollect K2EDTA Tubes	For CBC analysis	Greiner Bio-One

Histology and microscopy reagents

Reagent	Description	Vendor	Catalog No.
Paraformaldehyde, 4%	Tissue fixative	Electron Microscopy Sciences	15714-S
30% Sucrose Solution	Cryoprotection	Lab-prepared	—
OCT Embedding Compound	Tissue embedding medium	Fisher	23-730-571
Superfrost Plus Glass Slides	Charged slides for cryosections	Fisher	22-037-246
Coverslips, 22x50 mm	For mounting stained sections	Corning	2980-225
Triton X-100	Detergent for permeabilization	Sigma-Aldrich	T9284
Super PAP Pen	Hydrophobic barrier pen for IHC	IHCWORLD	SPM0928
DAPI Mounting Medium	Antifade mounting with nuclear stain	Southern Biotech	0100-20

Instrumentation and software

Instrument / Software	Description	Vendor
MESO QuickPlex SQ 120	Electrochemiluminescence plate reader	Meso Scale Discovery
Incucyte Live-Cell Analysis System	Real-time imaging for cell-based assays	Sartorius
Trans-Blot Turbo System	Protein blotting system	Bio-Rad
ChemiDoc MP Imaging System	Gel and blot imaging	Bio-Rad
Image Lab Software	Western blot quantification	Bio-Rad
GraphPad Prism	Data analysis and statistics	GraphPad
Biacore 1K SPR System	Surface plasmon resonance (SPR) analysis	Cytiva
OpenSPR-XT	Benchtop SPR platform	Nicoya
Nikon A1R+ HD Confocal Microscope	Fluorescence imaging with Z-stack acquisition.	Nikon
Leica CM1950 Cryostat	Rotary cryostat for the sectioning of frozen tissue specimen	Leica
NIS Element Viewer	Confocal image processing	Nikon

Generation and screening of camelid anti-TfR1 nanobodies. To generate anti-TfR1 nanobodies (anti-TfR1-Nbs), one alpaca and one llama were immunized with the human TfR1 ectodomain recombinant protein (Acro Biosystems, TFR-H82E5). The immunization protocol began with an initial subcutaneous injection of 0.5 mg TfR1 mixed with Complete Freund's Adjuvant (CFA) at week 0, followed by booster injections of 0.5 mg TfR1 with Incomplete Freund's Adjuvant (IFA) every two weeks up to week 14. Serum samples were collected before immunization and at designated time points to assess antibody titers via ELISA, utilizing TfR1-coated plates with appropriate negative and positive controls to ensure specificity.

At weeks 10 and 14, 500 mL of whole blood were drawn from each llama for peripheral blood mononuclear cell (PBMC) isolation. PBMCs were isolated within four hours of blood collection using density gradient centrifugation, ensuring cell viability above 99% as determined by trypan blue exclusion. Total RNA was extracted from the isolated PBMCs using the RNeasy Maxi Kit (Qiagen) and quantified by spectrophotometry, ensuring an A300/A280 ratio greater than 1.9. High-quality RNA was confirmed by agarose gel electrophoresis,

displaying distinct 18S and 28S rRNA bands without signs of degradation. Complementary DNA (cDNA) was synthesized from the purified RNA using the SuperScript IV First-Strand Synthesis System (Thermo Fisher Scientific). A nanobody-specific library was constructed by amplifying the variable regions of heavy-chain-only antibodies (VHH) through a two-step PCR process using camelid-specific degenerate primers. The amplified VHH fragments were cloned into the pADL-20c phagemid vector using SfiI restriction sites and transformed into E. coli TG1 cells, resulting in a phage display library containing approximately 2.57×10^9 individual clones. Library diversity was confirmed by sequencing 74 random clones, which revealed 88% contained VHH inserts with intact open reading frames and no duplicate sequences. Library panning was performed against immobilized TfR1 through three rounds of selection to enrich for specific binders. To reduce non-specific interactions, the phage pool was pre-absorbed on BSA-coated wells before each panning round. Enrichment of specific phage binders was monitored using dot assays, which demonstrated increased binding to TfR1 with each successive round.

Following panning, 470 individual clones were screened using an off-phage ELISA to identify those with specific binding to TfR1 and minimal binding to BSA. Positive clones were further validated through repeat screening and sequencing to ensure specificity and diversity. Selected nanobodies were expressed in a non-amber-suppressor strain of E. coli, and periplasmic fractions containing His-tagged nanobodies were purified using His-tag affinity chromatography. Purity was confirmed by SDS-PAGE, and nanobody concentrations were determined by absorbance at 280 nm. Purified nanobodies were dialyzed into PBS (pH 7.4) and filter-sterilized for downstream applications.

FACS analysis using transfected HEK293T cells and stable HEK293-hTfR1 cells. HEK293T cells (ATCC, CRL-3216) were cultured in DMEM (Corning, 10-017-CV) supplemented with 10% fetal bovine serum (Gibco, A3840102) at 37°C in a humidified 5% CO_2 incubator. For transient expression of transferrin receptor 1 (TfR1), cells were transfected with plasmids encoding human TfR1-EGFP, mouse Tfr1-EGFP, or rat Tfr1-EGFP (VectorBuilder; VB211221-1144bue, VB220126-1168dfv, and VB220126-1169wgz, respectively) using Fugene HD transfection reagent (Promega, E2311), according to the manufacturer's protocol. Cells were harvested for analysis 24–48 hours post-transfection. HEK293-hTfR1 cells (Acro Biosystems, CHEK-ATP089), a stable clone expressing human TfR1, were maintained under the same conditions and used as a positive control for TfR1 surface expression and nanobody binding. 100µg/mL of Hygromycin B (Sigma, H3272) was used for specific selection of TfR1 expressed clones.

Staining procedure: Cells were resuspended in ice-cold FACS buffer (PBS containing 1% BSA and 1 mM EDTA, pH 7.2) and incubated with nanobodies (final concentration 40–100 nM) at 4°C for 45 minutes with gentle shaking. After three washes in FACS buffer, cells were incubated for 30 minutes at 4°C with APC-conjugated anti-His tag antibody (R&D Systems, IC050A) at 1:100 dilution to detect bound His-tagged nanobodies. Following additional washes, cells were stained with propidium iodide (Invitrogen, P3566; 1:1000 dilution) to exclude dead cells.

Flow cytometry analysis: Samples were analyzed on a BD LSR Fortessa or similar flow cytometer. Data were gated on live, EGFP-positive cells (for transfection marker) and analyzed for APC signal using FlowJo software. For comparative binding across species-specific TfR1, anti-human, anti-mouse, and anti-rat APC-conjugated antibodies (Invitrogen; Cat. Nos. 17071942, 17071182, 17071082) were also used in parallel at 1:100 dilution.

TNFα inhibition and IC_{50} determination using incucyte caspase-3/7 assays. WEHI-13VAR cells (ATCC, CRL-2168), a TNFα-sensitive mouse fibrosarcoma line, were cultured in RPMI 1640 medium (Corning, 10-040-CV)

supplemented with 10% fetal bovine serum (Gibco, A3840102) at 37°C with 5% CO_2. Cells were seeded into 96-well plates at 30,000 cells per well in RPMI containing 10% FBS and 1 μg/mL Actinomycin D (Sigma-Aldrich, A9415) to sensitize cells to TNFα-induced apoptosis. Recombinant human TNFα (Acro Biosystems, TNA-H5228) was pre-incubated with varying concentrations of TNFI nanobodies for 30 minutes at room temperature, then added to the cells at a final TNFα concentration of 0.25 ng/mL. Caspase-3/7 activity was monitored using Caspase-3/7 Green Detection Reagent (Thermo Fisher Scientific, C10423) added at a final concentration of 0.5 μM. Plates were immediately placed in the Incucyte Live-Cell Imaging System (Sartorius), and fluorescent signals were monitored every 2–4 hours for up to 24 hours. Fluorescent caspase-3/7 signal intensity was quantified using Incucyte analysis software. TNFα-induced apoptosis was set to 100%, and inhibition was expressed as a percentage of this maximum signal. IC_{50} values for each TNFI-Nb were calculated using GraphPad Prism with nonlinear regression (log[inhibitor] vs. response – variable slope).

Transferrin–TfR1 binding competition assay. HEK293-hTfR1 stable cells were cultured in DMEM (Corning, 10-017-CV) supplemented with 10% fetal bovine serum (Gibco, A3840102) and 100μg/mL of Hygromycin B under standard conditions (37°C, 5% CO_2). Cells were harvested at ~80% confluence, washed with ice-cold PBS, and resuspended in FACS buffer (PBS + 1% BSA + 1 mM EDTA).

Competition assay: Cells were incubated for 60 minutes at 4°C in 96-well plates with the following conditions (in 200 μL final volume per well):

TF-FITC alone: 2.5 μM FITC-conjugated human transferrin (Jackson ImmunoResearch, 009090050)

Nanobody alone: 2.5 μM of individual TfR1b-Nbs

TF-FITC + Nanobody: 2.5 μM of both reagents co-incubated

TF-FITC + Unlabeled TF: Co-incubation with increasing concentrations of unlabeled human TF (30 nM, 750 nM, 2.5 μM, and 7.5 μM; Jackson ImmunoResearch, 009000050)

Following incubation, cells were washed 3× with FACS buffer and resuspended in buffer containing 1:1000 dilution of propidium iodide (Invitrogen, P3566) to exclude dead cells.

Flow cytometry analysis: Fluorescence was measured using a flow cytometer. FITC signal was gated on live cells, and the mean fluorescence intensity (MFI) of the FITC channel was used to quantify TF-FITC binding. Competitive inhibition was determined by comparing MFI between conditions. Decreased MFI in the presence of excess unlabeled TF or TfR1b-Nbs indicated interference with TF-TfR1 binding.

Transferrin uptake interference assay using incucyte live-cell imaging. HEK293-hTfR1 stable cells were plated in black-walled 96-well tissue culture plates at a density of 30,000 cells per well in DMEM (Corning, 10-017-CV) with 10% fetal bovine serum (Gibco, A3840102) and 100μg/mL of Hygromycin B. After 24 hours, cells reached approximately 80% confluence and were ready for assay. To assess TfR1-mediated uptake, pHrodo™ Red-conjugated human transferrin (Thermo Fisher Scientific, P35376) was used at a final concentration of 312.5 nM. Uptake competition was evaluated by co-incubating with either:

Unlabeled human transferrin at increasing concentrations (30 nM, 150 nM, 750 nM, and 3750 nM), or, selected TfR1b-Nbs at 40 nM.

All reagents were diluted in serum-free DMEM. Cells were incubated at 37°C immediately following reagent addition.

Live imaging and quantification: Plates were placed into the Incucyte® Live-Cell Analysis System (Sartorius), and images were acquired every 1–2 hours over a 24-hour period using the red fluorescence channel (excitation/emission ~560/585 nm). The pHrodo dye emits fluorescence only in acidic intracellular compartments, allowing real-time tracking of transferrin uptake via receptor-mediated endocytosis.

Data analysis: Total integrated fluorescence intensity per well was quantified using Incucyte software. Uptake inhibition by TfR1b-Nbs or unlabeled transferrin was calculated as the percent decrease in red signal compared to wells treated with pHrodo-TF alone. All conditions were performed in triplicate and repeated in independent experiments.

In vivo drug administration, perfusion, and sample collection. ***Intravenous (IV) injection****:* Rats were briefly warmed under a heat lamp to promote vasodilation. Intravenous drug administration was performed via the lateral tail vein using 1 mL Tuberculin Syringes fitted with 25G needles (Cardinal Health). Nanobodies or fusion constructs were injected slowly at a volume of 1 µL per gram of body weight in sterile PBS to minimize stress and ensure accurate dosing.

SQ injection: For SQ administration, animals were gently restrained using a towel, and the compound was injected into the interscapular area using a 1mL Tuberculin syringes fitted with 25G needles (Cardinal Health). Dosing volume was the same as IV: 1 µL of a 40 µM solution per gram of body weight in PBS. Animals were monitored for signs of discomfort or inflammation at the injection site.

Perfusion and brain tissue collection: At the designated time points post-injection, animals were deeply anesthetized using isoflurane. For terminal experiments involving brain collection, transcardiac perfusion was performed with cold PBS (without calcium or magnesium) at a flow rate of 10 mL/min for 10 minutes to remove blood from the vasculature. Brains were immediately extracted and processed for either biochemical assays or immunohistochemistry.

Serum collection:

Non-terminal: Blood samples (~300 µL) were collected via tail vein using a 25G VACUETTE Safety Blood Collection Set (Greiner Bio-One) into BD Microtainer Serum Separator Tubes (Becton Dickinson). Samples were allowed to clot for 30 minutes at room temperature, followed by centrifugation at 9000 × g for 30 seconds.

Terminal**:** Blood was collected by cardiac puncture using a 5 mL syringe (Air-Tite Products Co., Inc) fitted with a 21G needle (Cardinal Health), transferred into BD Vacutainer SST tubes, allowed to clot for 30 minutes at room temperature, and centrifuged at 2000 × g for 10 minutes. All serum samples were aliquoted and stored at –80°C.

CSF collection: Following transcardiac perfusion and prior to brain tissue collection, CSF was collected via cisterna magna puncture. Animals were placed in a stereotaxic frame (Stoelting Co.). The head was flexed forward, and a midline incision was made at the base of the skull to expose the translucent dura mater over the cisterna magna, using a 28G insulin syringe (EXEL), CSF was slowly withdrawn, carefully avoiding blood contamination. Typically, 50–80 µL of CSF was collected per animal. Samples were snap-frozen in liquid nitrogen and stored at –80°C.

ELISA for TfR1- and TNFα-based detection of nanobodies. ***TfR1-based ELISA****:* Streptavidin-coated 96-well plates (Meso Scale Discovery, L15SA or L45SA) were blocked overnight at 4°C with 3% BSA in PBST (PBS + 0.05% Tween-20). Plates were then coated with 0.25 µg/mL biotinylated human transferrin receptor 1 (TfR1) protein (Acro Biosystems, TFR-H82E5) in PBS for 1 hour at room temperature with shaking.

After washing 4× with PBST, samples containing nanobodies were added and incubated overnight at 4°C. Following three PBST washes, wells were incubated with 1 µg/mL Goat Anti-Alpaca IgG, VHH domain (Jackson ImmunoResearch, 128-005-230) for 1 hour at room temperature. After additional washes, SULFO-TAG-labeled anti-goat IgG secondary antibody (Meso Scale Discovery, R32AG; 0.5–1 µg/mL) was added and incubated for 1 hour.

TNFα-based ELISA: The same procedure was used with the following modifications: plates were coated with 0.2 µg/mL biotinylated human TNFα (Acro Biosystems, TNA-H82E3-25ug) instead of TfR1. Detection of nanobody binding was performed using either anti-His tag rabbit monoclonal antibody (Cell Signaling Technology, 12698; 1:500 dilution) or Goat Anti-Alpaca IgG VHH domain antibody, depending on the tag configuration. Corresponding SULFO-TAG-labeled anti-rabbit (R32AB) or anti-goat (R32AG) secondary antibodies were used.

Plate reading and analysis: After final washes, wells were developed using 2× MSD Read Buffer (Meso Scale Discovery, R92TC) and read on a MESO QuickPlex SQ 120 instrument. Background-subtracted signals were normalized to control wells. TNFI-Nb1 (specific for TNFα) and irrelevant control nanobodies were used to confirm specificity.

Hematotoxicity Assessment via Complete Blood Count (CBC). To evaluate hematotoxicity following intravenous administration of TfR1-targeting nanobody constructs, complete blood counts (CBCs) were performed in treated and control rats at multiple time points. Animals received IV injections of either PBS (Group 1, n=7; 3 males, 4 females) or TNFI-β–TfR1b–A2 (Group 2, n=8; 3 males, 5 females) at a dose of 1 µL of a 40 µM solution per gram of body weight. Prior the blood collection, rats were briefly warmed under a heat lamp. Blood samples were collected from the tail lateral vein using a VACUTTE Safety Blood Collection Set (Greiner Bio-One) and transferred into MiniCollect K2EDTA tubes (Greiner Bio-One). Samples were immediately placed on an icepack and kept at +4°C until analysis and transported to the In Vivo Research Services (IRVS) Core Facility at Rutgers University. CBCs were performed using the Heska Element HT5 CBC Analyzer.

CBCs were collected at four time points:

- **Day –3 (D–3):** Baseline, prior to treatment
- **Day 1 (D1):** 24 hours after the first injection
- **Day 17 (D17):** After the third injection
- **Day 24 (D24):** After the fourth injection

The following parameters were measured:

White blood cell (WBC) profile:

- Total WBC ($\times 10^3$/µL)
- Neutrophils (NEU), Lymphocytes (LYM), Monocytes (MONO), Eosinophils (EOS), Basophils (BAS)
- Percent distribution: NEU%, LYM%, MONO%, EOS%, BAS%

Red blood cell (RBC) profile:

- RBC count ($\times 10^6$/µL), Hemoglobin (HGB, g/dL), Hematocrit (HCT, %)
- Mean corpuscular volume (MCV, fL), Mean corpuscular hemoglobin (MCH, pg)
- Mean corpuscular hemoglobin concentration (MCHC, g/dL)
- Red cell distribution width (RDW, %)

Platelet profile:

- Platelet count (PLT, $\times 10^3/\mu L$), Mean platelet volume (MPV, fL)

These data were used to monitor for treatment-related hematologic changes, particularly indicators of anemia, leukocyte shifts, and thrombocytopenia.

Fractionation of brain tissue. To evaluate the distribution of nanobodies between the brain vasculature and parenchyma, rats were deeply anesthetized and perfused transcardially with cold PBS. The right hemisphere was dissected, and choroid plexuses were removed. Tissue was homogenized in 5 mL of ice-cold S1 buffer (250 mM sucrose, 20 mM Tris-base pH 7.4, 1 mM EDTA, 1 mM EGTA) using a glass-Teflon homogenizer (10 strokes). Homogenates were centrifuged at 1,000 × g for 10 minutes at 4°C. Pellets were resuspended in 2 mL of 17% dextran (MW 60,000; Sigma, 31397) and centrifuged at 4,200 × g for 15 minutes. The pellet was collected as the capillary-enriched vasculature fraction. The supernatant was diluted in S1 buffer and centrifuged again at 4,200 × g for 15 minutes. The resulting pellet was collected as the vascular-depleted parenchymal fraction.

Both fractions were lysed in ice-cold S1 buffer containing protease and phosphatase inhibitors and sonicated (50% amplitude, 3 × 10 s bursts with 30 s interval rests). Protein concentrations were determined by Bradford assay. Aliquots were used for ELISA and Western blot.

Western Blotting of Brain Fractions. Equal amounts of protein from homogenate, vasculature, and parenchymal fractions were mixed with LDS sample buffer (Invitrogen, NP0007) containing 10% β-mercaptoethanol, heated at 95°C for 5 minutes, and loaded onto 4–12% Bis-Tris polyacrylamide gels (Bio-Rad, 3450125). Electrophoresis was followed by transfer to nitrocellulose membranes using the Trans-Blot Turbo System (Bio-Rad) at 25 V for 7 minutes. Membranes were blocked with 5% non-fat milk (Bio-Rad, 1706404) in PBST (PBS + 0.05% Tween-20) for 45 minutes, then incubated overnight at 4°C with primary antibodies (1:1000 dilution in blocking buffer):

- Anti-human TfR1 (CST, 13113)
- Anti-Glut1 (CST, 73015) – endothelial cell marker
- Anti-GAPDH (Sigma, G9545) – loading control
- Anti-VAMP2, NMDAR2B (CST, 4212) – neuronal markers
- Anti-IBA1 (Wako, 01620001) – microglial marker
- Anti-EAAT2 (Synaptic Systems, 250203 or 104202) – astrocytic marker
- Anti-MBP (CST, 78896) – oligodendrocyte marker

After washing, membranes were incubated for 45 minutes at room temperature with HRP-conjugated secondary antibodies (anti-rabbit: CST 7074 or Southern Biotech OB405005, 1:1000 in 5% milk). Detection was performed using Clarity ECL substrate (Bio-Rad, 1705061), and bands were visualized with the ChemiDoc MP Imaging System (Bio-Rad). Densitometry analysis was performed using Image Lab software.

These blots validated the separation of vascular and parenchymal compartments and confirmed human TfR1 expression and nanobody localization across fractions.

Immunohistochemistry (IHC). Rats were deeply anesthetized and perfused transcardially with PBS (without calcium and magnesium), followed by fixation with 4% paraformaldehyde (PFA; Electron Microscopy Sciences, 15714-S) in PBS at a flow rate of 10 mL/min for 10 minutes. Brains were extracted and post-fixed in 4% PFA at 4°C

for 24 hours on a shaker. After fixation, brains were washed twice in PBS and incubated in 30% sucrose in PBS at 4°C for 48 hours for cryoprotection. Brains were embedded in OCT compound (Fisher, 23-730-571), frozen, and stored at –80°C. Coronal sections (20 µm thick) were cut on a cryostat (Leica CM1950) and mounted onto charged glass slides (Fisher, 22-037-246), then stored at –80°C until use.

Immunostaining procedure: Slides were brought to room temperature, and a hydrophobic barrier was drawn around each section using a Super PAP Pen. Sections were rehydrated in PBS for 10 minutes and blocked in 10% normal goat serum containing 0.3% Triton X-100 (Sigma, T9284) for 1 hour at room temperature.

Sections were incubated overnight at 4°C with the following primary antibodies diluted in 5% serum with 0.3% Triton X-100:

Primary Antibody	Target	Dilution	Vendor / Cat. No.
His-tag (Alexa Fluor 488)	Nanobody localization	1:250	CST, 14930
Anti-IBA1 (SPICA Dye 594)	Microglia	1:250	Wako, 012-28401
Anti-GFAP	Astrocytes	1:250	BD Biosciences, 556327
Anti-human TfR1	Human TfR1	1:250	CST, 54505S
Anti-CD31 (Alexa Fluor 647)	Endothelial cells	1:250	Abcam, ab305210

After three washes in PBS containing 0.3% Triton X-100, sections were incubated for 2 hours at room temperature with appropriate fluorescent secondary antibodies diluted 1:1000 in PBS with 5% serum and 0.3% Triton X-100. The following secondary antibody was used:

Secondary Antibody	Target	Dilution	Vendor / Cat. No.
Goat anti-Mouse IgG (Alexa Fluor 594)	GFAP, TfR1	1:1000	Invitrogen, A-11032

Sections were washed in PBS and mounted using DAPI-containing aqueous mounting medium (Southern Biotech, 0100-20) and coverslipped (Corning, 2980-225).

Imaging and analysis: Confocal imaging was performed using a Nikon A1R inverted laser-scanning confocal microscope. Z-stacks and tile scans were acquired using consistent settings across experimental conditions. Colocalization of nanobody signal with cell-type markers was assessed in the cortex and hippocampus. Image processing was performed using NIS element view.

Statistical Analysis. All quantitative data are presented as mean ± standard error of the mean (SEM), unless otherwise indicated. Statistical analyses were performed using GraphPad Prism software. Comparisons between two groups were made using unpaired two-tailed Student's t-tests. For multiple group comparisons, one-way or two-way ANOVA followed by appropriate post hoc tests (e.g., Tukey's or Sidak's) were used. A p-value < 0.05 was considered statistically significant. The number of biological replicates (n) is indicated in the figure legends or methods.

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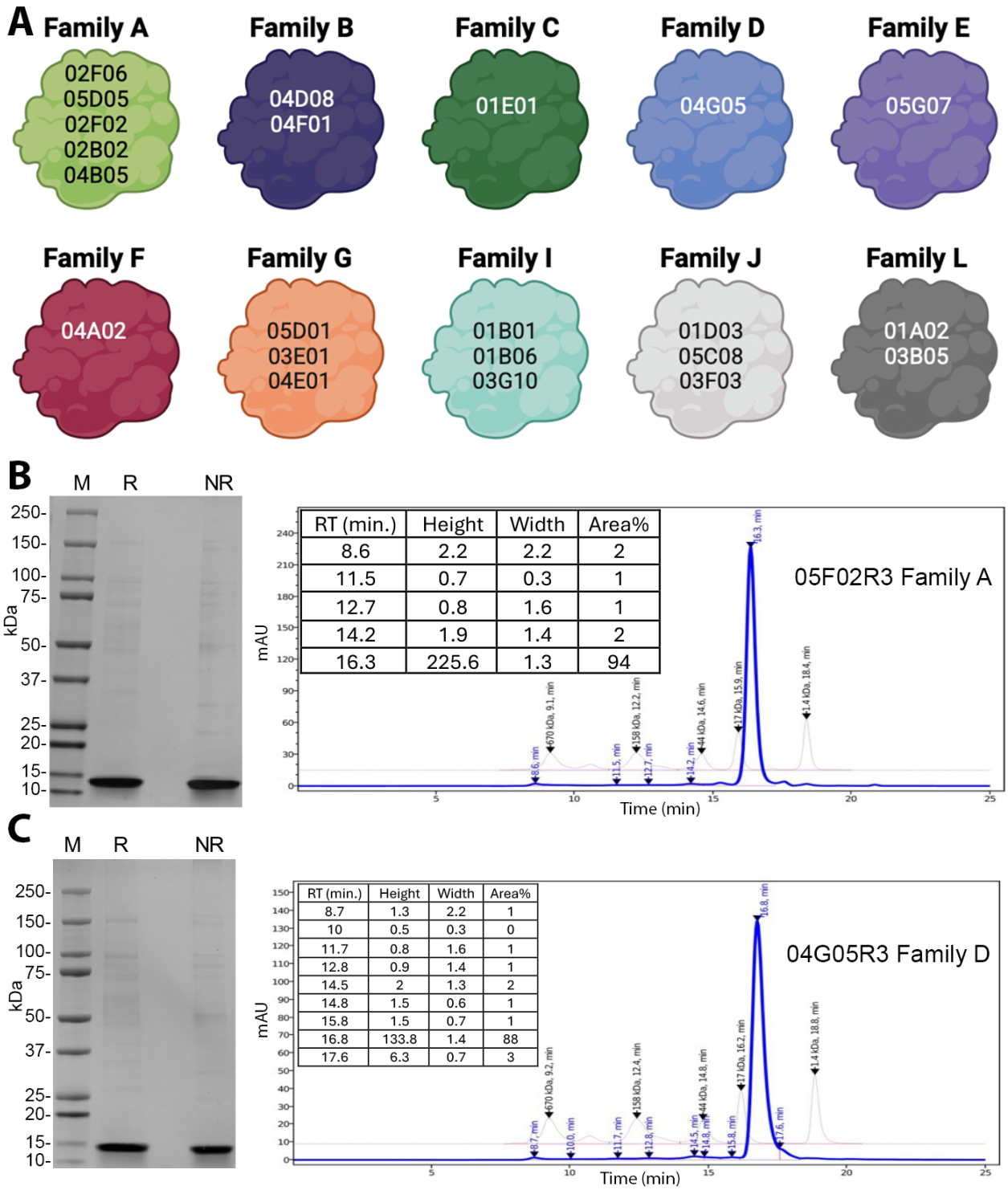
Yin T, Ramon A, Greenig M, Sormanni P, D'Adamio L (2025) Development of potent humanized TNFα inhibitory nanobodies for therapeutic applications in TNFα-mediated diseases. *BioRxiv*

Figures and Figures legends.



RT (min.)	Height	Width	Area%
8.6	2.2	2.2	2
11.5	0.7	0.3	1
12.7	0.8	1.6	1
14.2	1.9	1.4	2
16.3	225.6	1.3	94

RT (min.)	Height	Width	Area%
8.7	1.3	2.2	1
10	0.5	0.3	0
11.7	0.8	1.6	1
12.8	0.9	1.4	1
14.5	2	1.3	2
14.8	1.5	0.6	1
15.8	1.5	0.7	1
16.8	133.8	1.4	88
17.6	6.3	0.7	3

Figure 1. Protein purification and analysis of selected TfR1b-Nbs. A) Schematic representation of the 24 TfR1b-Nbs produced in mammalian cells. **B)** SDS-PAGE and SEC-HPLC analysis of TfR1b-Nb 02B02R3 (Family A). Coomassie-stained SDS-PAGE shows M = molecular weight marker; Lane 1 = purified 02B02R3 under reducing conditions; Lane 2 = under non-reducing conditions. Left panel: SEC-HPLC profile of purified 02B02R3. **C)** SDS-PAGE and SEC-HPLC analysis of TfR1b-Nb 04G05R3 (Family D), shown as in (B).

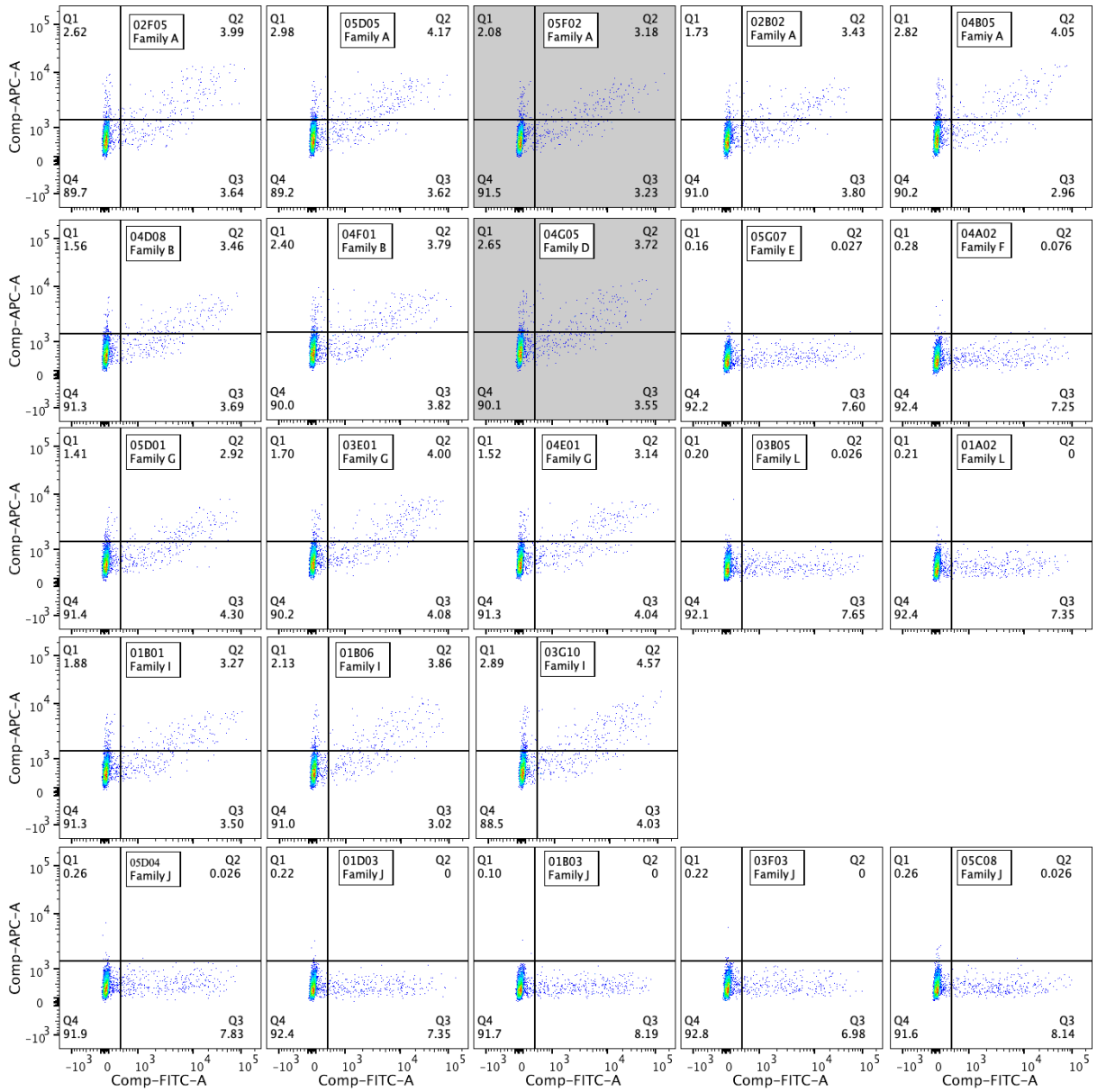


Figure 2. TfR1b-Nbs produced by mammalian cells bind to human TfR1. HEK293 cells were transfected with a vector expressing human TfR1 (alongside EGFP). Subsequently, cells were treated with each TfR1b-Nb at a concentration of 400 nanomolar, followed by incubation with an anti-His-APC antibody. The two nanobodies highlighted in grey are the parental sequences from which the final selected NewroBus molecules were derived.

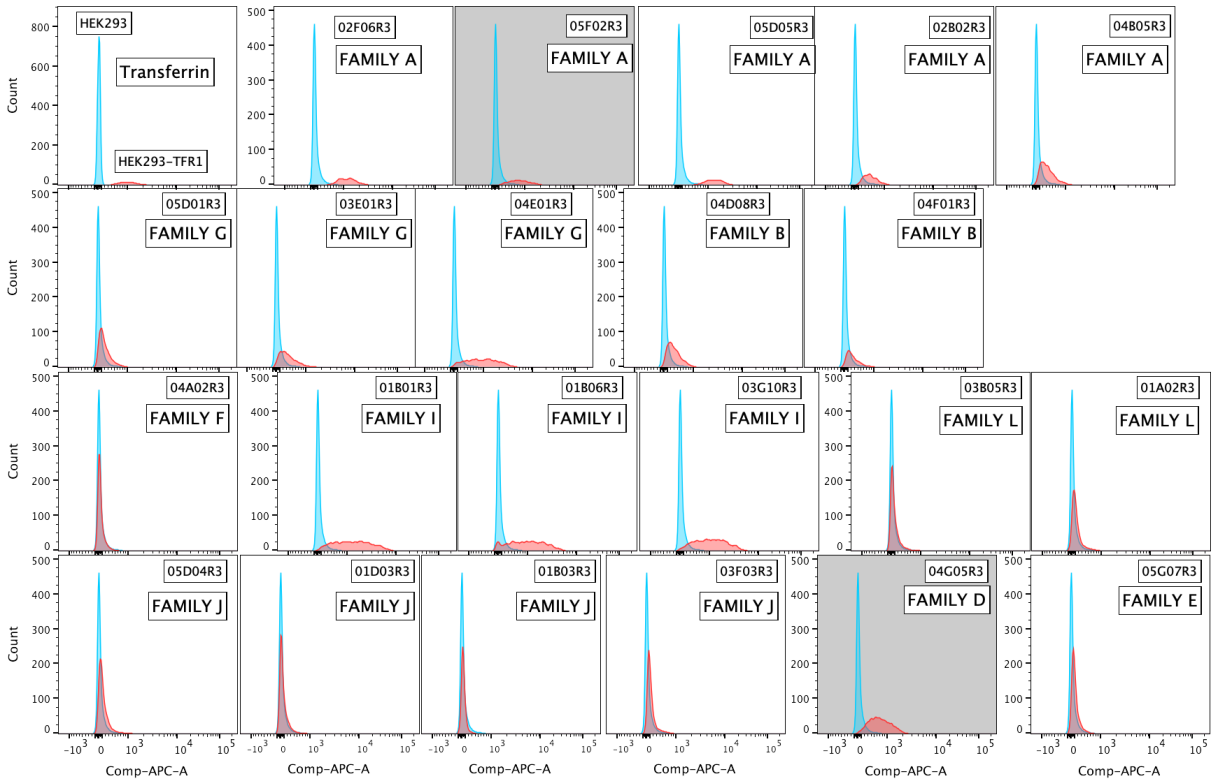


Figure 3. TfR1b-Nbs produced by mammalian cells bind to human TfR1. The upper-left panel shows the specific binding of human Transferrin-FITC (at a concentration of 2.5 nanomolar) to CHEK-ATP089 cells, with no binding observed on HEK293 cells. The remaining panels depict the binding of each TfR1b-Nb at a concentration of 40 nanomolar to CHEK-ATP089 cells. No binding was observed on HEK293 cells. The two nanobodies highlighted in grey are the parental sequences from which the final selected NewroBus molecules were derived.

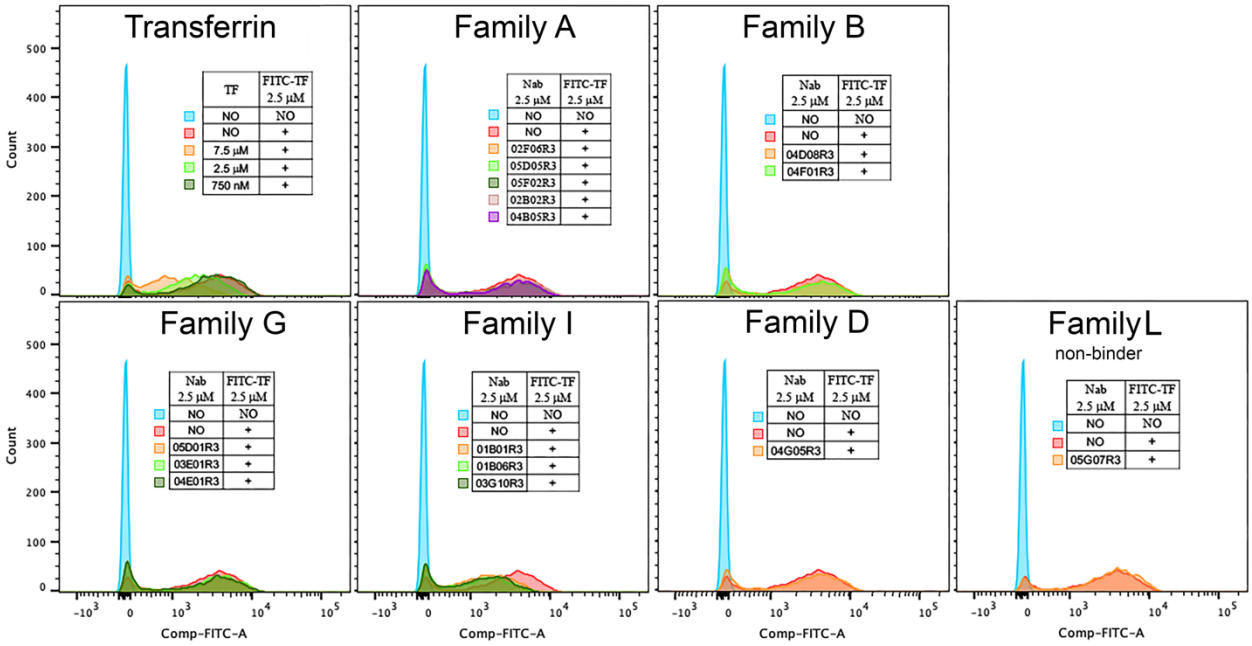


Figure 4. Selection of TfR1b-Nbs that do not interfere with TF-TfR1 interaction. The upper-left panel demonstrates the dose-dependent competition of unlabeled Transferrin for binding to TfR1 on the cell surface of CHEK-ATP089 cells in the presence of FITC-Transferrin. Subsequent panels assess the inhibitory activity of 14 TfR1b-Nbs from Families A, B, D, G, and I. A nanobody from Family L (which lacks binding to TfR1 on CHEK-ATP089 cells) serves as a negative control. In these experiments, CHEK-ATP089 cells were incubated with the specified proteins for 1 hour on ice before FACS analysis. Human Transferrin-FITC and TfR1b-Nbs were utilized at a concentration of 2.5 micromolar.

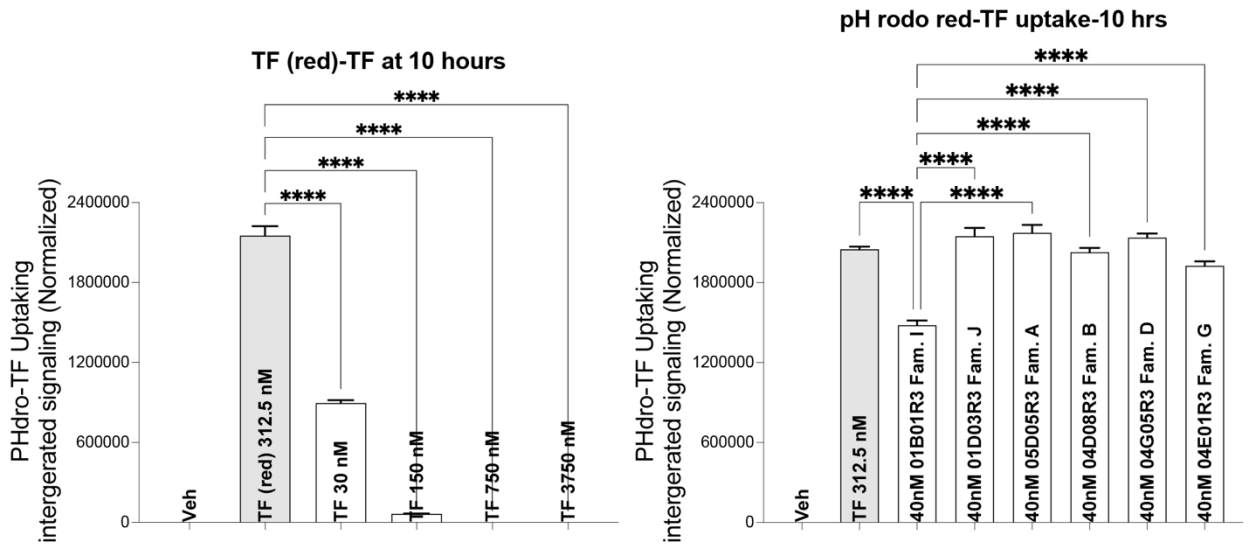


Figure 5. Selection of TfR1b-Nbs that do not interfere with TF uptake. The panel demonstrates the dose-dependent competition of unlabeled Transferrin pHrodo Red-TF uptake by CHEK-ATP089 cells. The right panel assess the inhibitory activity of representative TfR1b-Nbs from Families A, B, D, G, and I. A nanobody from Family J (which lacks binding to TfR1 on CHEK-ATP089 cells) serves as a negative control.

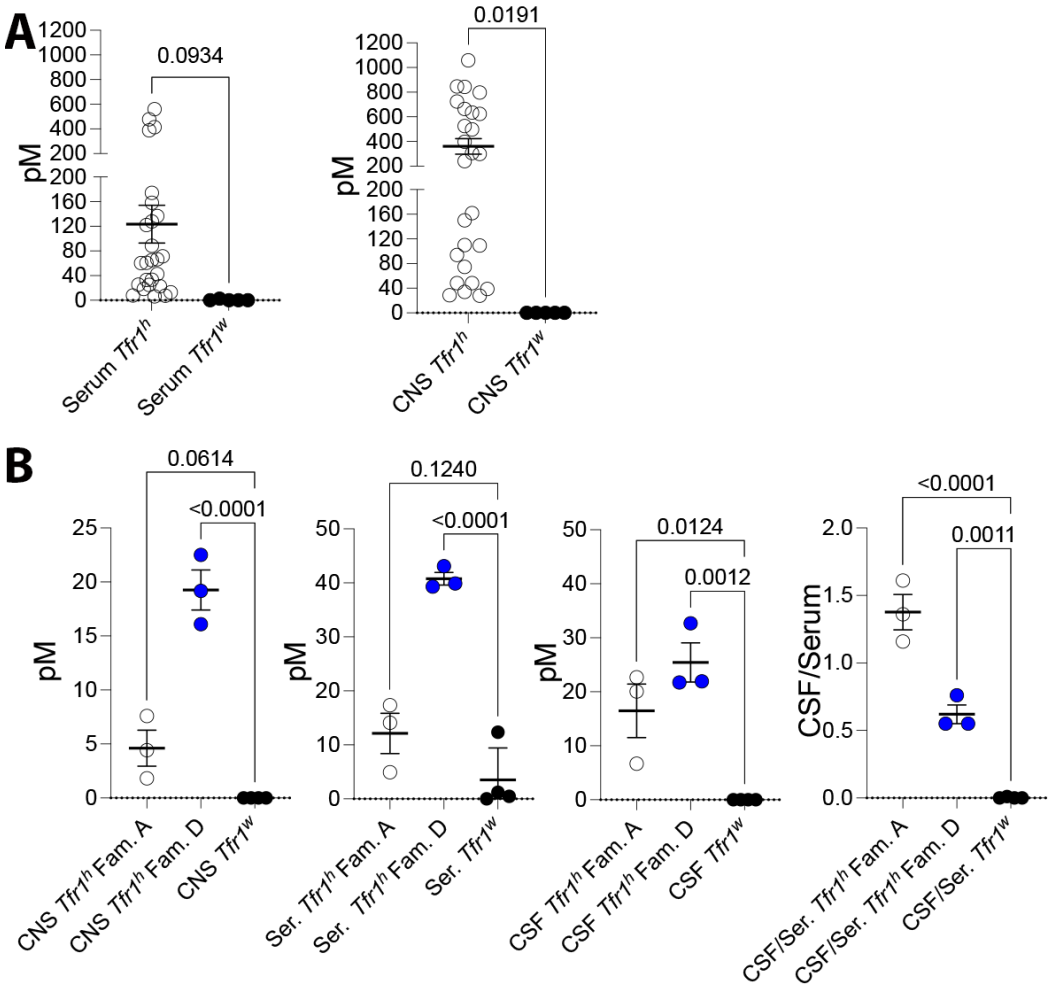


Figure 6. In vivo BBB permeability of TfR1b-Nbs in humanized Tfr1 rats. A) ELISA detection of TfR1b-Nbs in brain homogenates of rats expressing human TfR1 (*Tfr1*$^{h/w}$ or *Tfr1*$^{h/h}$), but not in wild-type rats (*Tfr1*$^{w/w}$), 16-18 hrs. following IV injection. **B)** Detection of TfR1b-Nbs 04B05R3 (Family A) and 05D01R3 (Family D) in serum, CSF, and brain homogenates of *Tfr1*$^{h/w}$ and *Tfr1*$^{w/w}$ rats 44–48 hours post-injection. Both nanobodies showed high CSF/serum ratios in *Tfr1*$^{h/w}$ rats, indicating efficient BBB penetration and sustained stability.

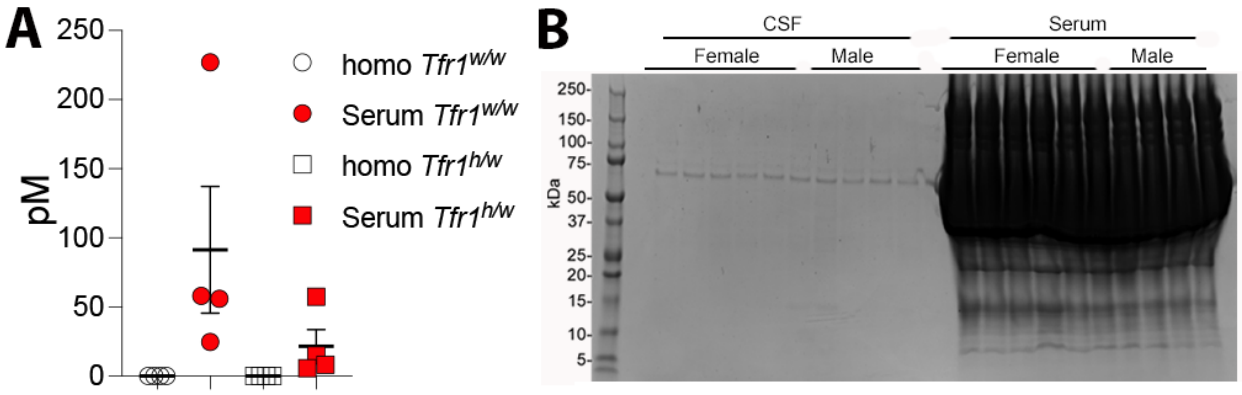


Figure 7. TNFI-Nb1 does not cross the BBB and is excluded from the brain and CSF. A) ELISA quantification of TNFI-Nb1 in serum and brain homogenates from *Tfr1*$^{w/w}$ and *Tfr1*$^{h/w}$ rats 6 hours after intravenous injection. TNFI-Nb1 is detected in serum but not in brain tissue, indicating that it does not cross the BBB. **B)** SDS-PAGE and Coomassie Blue staining of equal volumes of serum and CSF from *Tfr1*$^{h/h}$ rats. While serum contains abundant proteins, only a faint ~70 kDa band is detected in the CSF, confirming the low protein content of CSF and intact BBB exclusion of serum proteins.

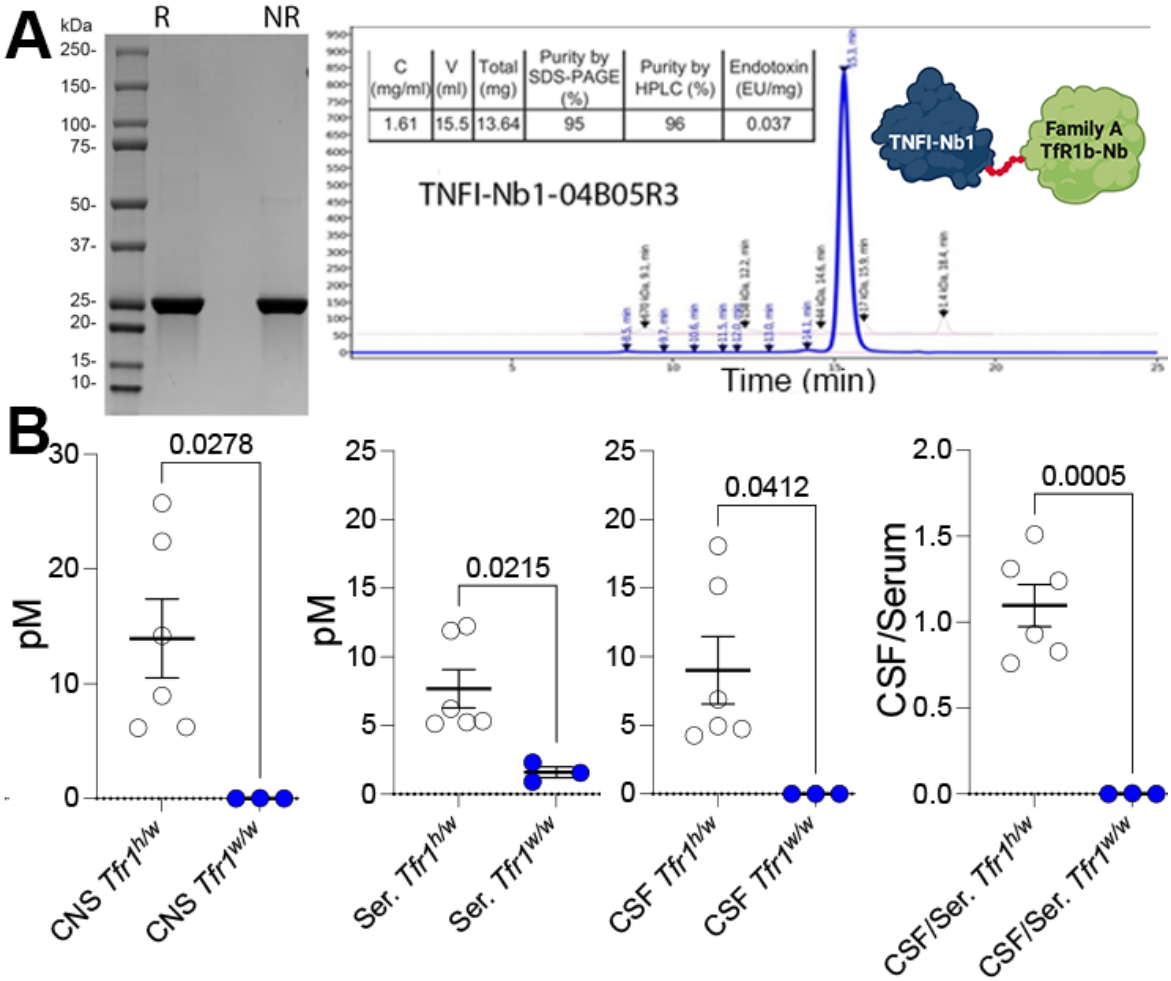


Figure 8. CNS delivery of a BBB-impermeable TNFα inhibitor via TfR1b-Nb-directed transcytosis. A) Schematic of the heterodimer composed of TNFI-Nb1 fused to TfR1b-Nb 04B05R3 (Family A), produced in CHO-S cells. **B)** ELISA-based detection of the heterodimer in serum, CSF, and brain homogenates of *Tfr1*$^{h/w}$ rats, but not in *Tfr1*$^{w/w}$ controls, following IV injection. The heterodimer exhibited efficient BBB penetration with a CSF/serum ratio of ~1, confirming human TfR1-dependent CNS delivery. Samples were analyzed 48 hours post-IV injection.

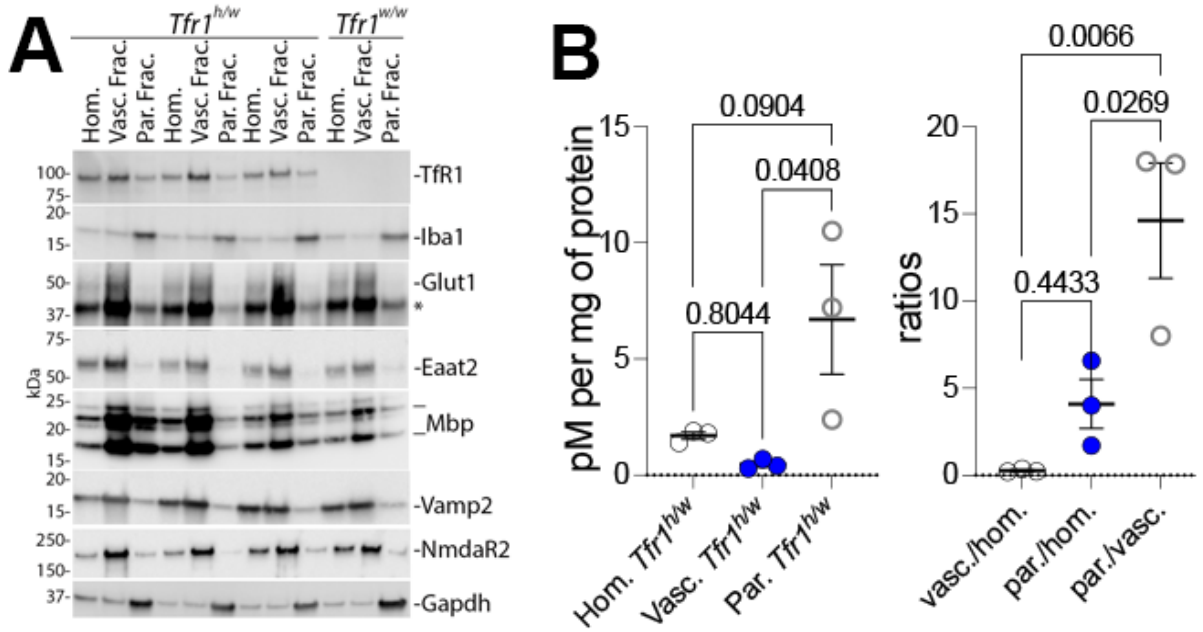


Figure 9. Brain fractionation analysis of TNFI-Nb1–linker–04B05R3 distribution in Tfr1$^{h/w}$ rats. **A)** Western blot validation of brain fractionation into vasculature and parenchyma. Glut1 (endothelial marker) was enriched in the vasculature fraction, while Gapdh and cell-type–specific markers—Vamp2 and NmdaR2b (neurons), Iba1 (microglia), Eaat2 (astrocytes), and Mbp (oligodendrocytes)—were enriched in the parenchymal fraction. Human TfR1 was detected in all fractions, with strongest signal in the vasculature. **B)** ELISA quantification of TNFI-Nb1–04B05R3 in homogenate, vasculature, and parenchymal fractions. After normalization to protein content, the parenchymal fraction showed the highest nanobody levels, supporting uptake by CNS cells via human TfR1-mediated transcytosis.

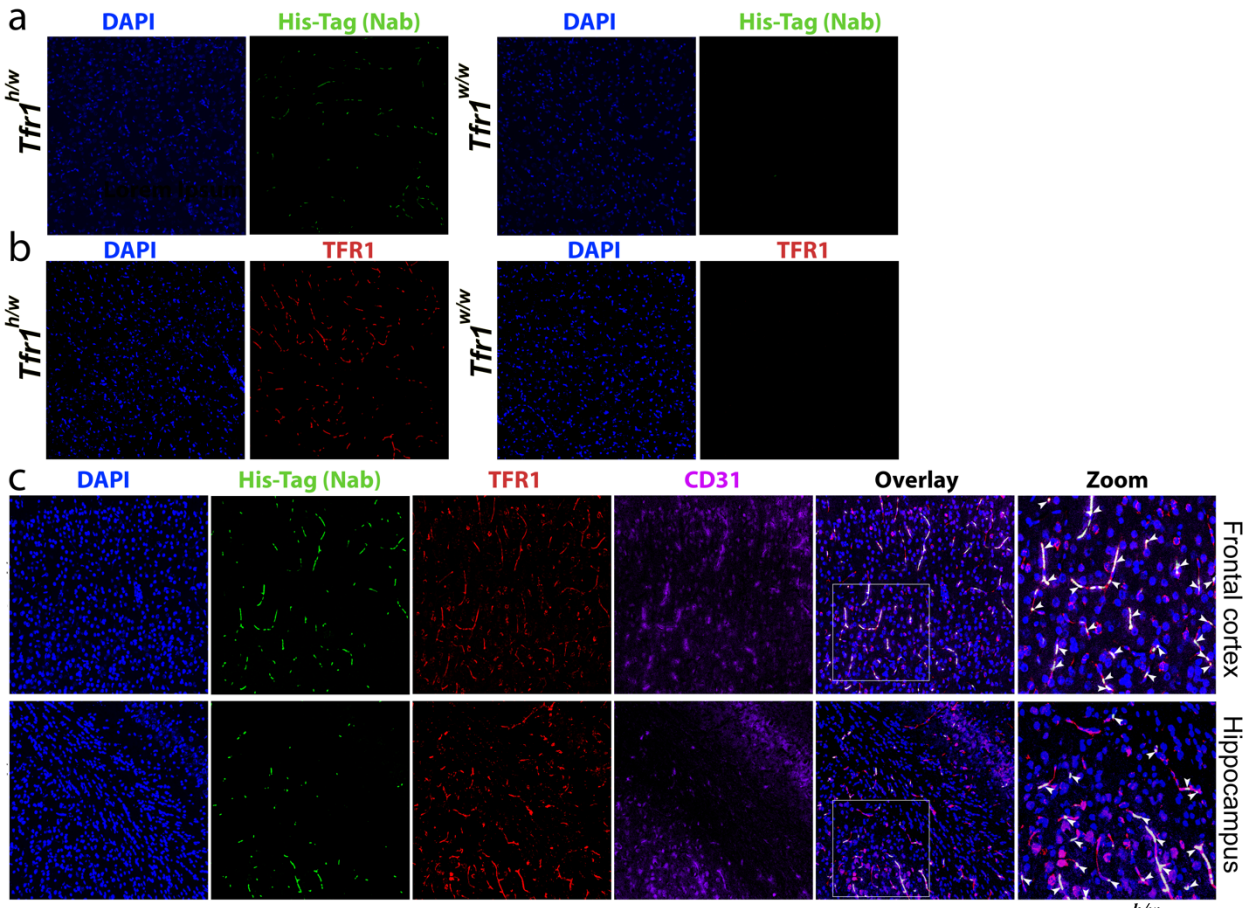


Figure 10. TNFI-Nb1-linker-04B05R3 colocalize with human TfR1in brain vessels of *Tfr1$^{h/w}$* rats. A) Anti-His tag antibody (green) and (**B**) anti-human TfR1 antibody (red) stain TNFI-Nb1-linker-04B05R3 and human TfR1, respectively, in the brains of *Tfr1$^{h/w}$* but not *Tfr1$^{w/w}$* rats. **C**) Human TfR1 (red), CD31 (purple)—an endothelial cell marker—and TNFI-Nb1-linker-04B05R3 (green) colocalize in vessels in the brains of *Tfr1$^{h/w}$* rats. White arrows in the zoomed-in panel indicate the colocalization spots within the region highlighted by the white square in the overlay panel.

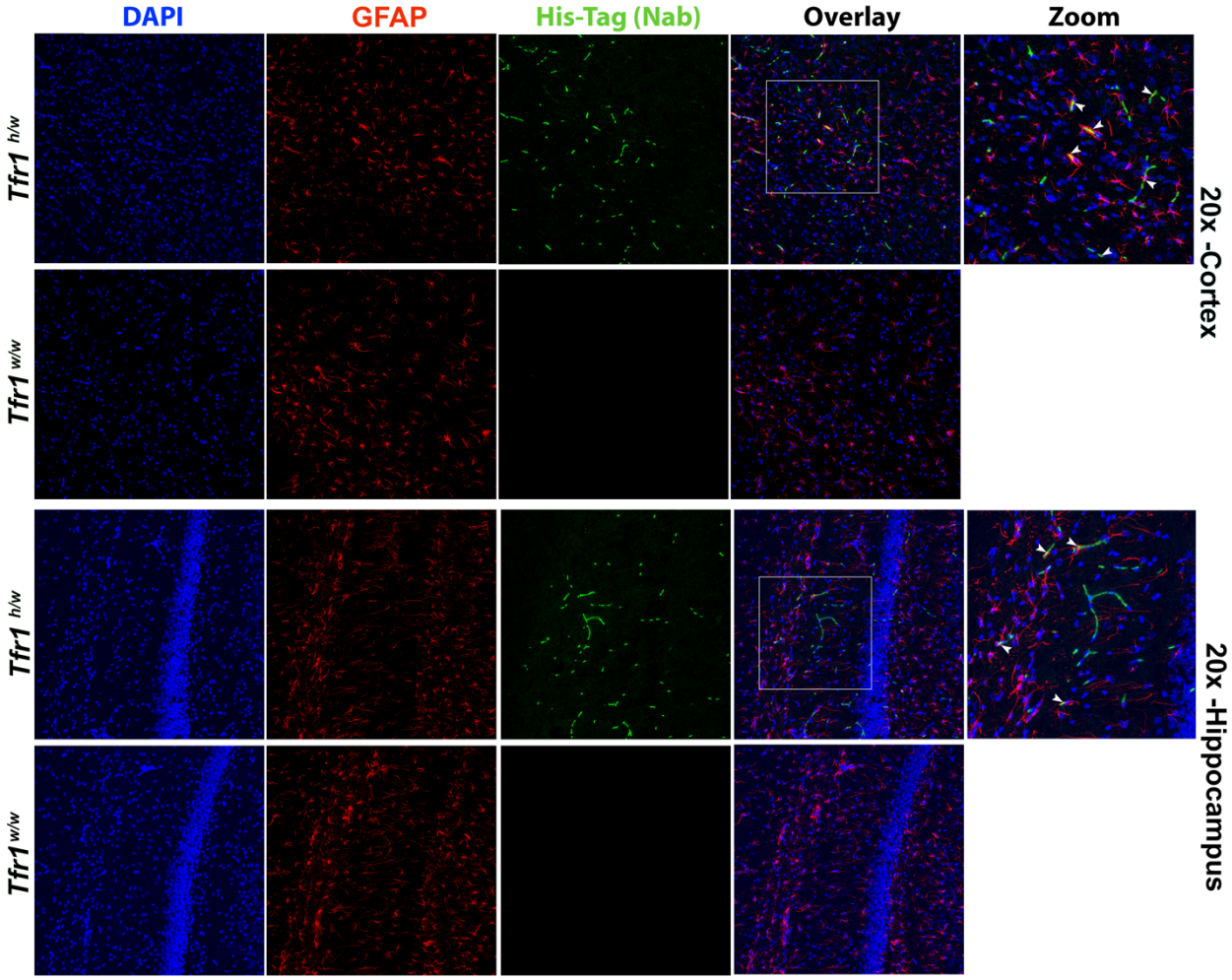


Figure 11. Confocal images showing the localization of TNFI-Nb1-linker-04B05R3 in astrocytes of the cortex and hippocampus in *Tfr1*$^{h/w}$ and *Tfr1*$^{w/w}$ rats. Sections were stained with an anti-His tag antibody (green) to detect TNFI-Nb1-linker-04B05R3 and an anti-GFAP antibody (red) to label astrocytes. TNFI-Nb1-linker-04B05R3 is present in the brains of *Tfr1*$^{h/w}$ rats but not in *Tfr1*$^{w/w}$ rats. In *Tfr1*$^{h/w}$ rats, TNFI-Nb1-linker-04B05R3 colocalizes with some astrocytes, indicated by yellow overlays in the merged images. White arrows in the zoomed-in panel point to colocalization spots within the region highlighted by the white square in the overlay panel.

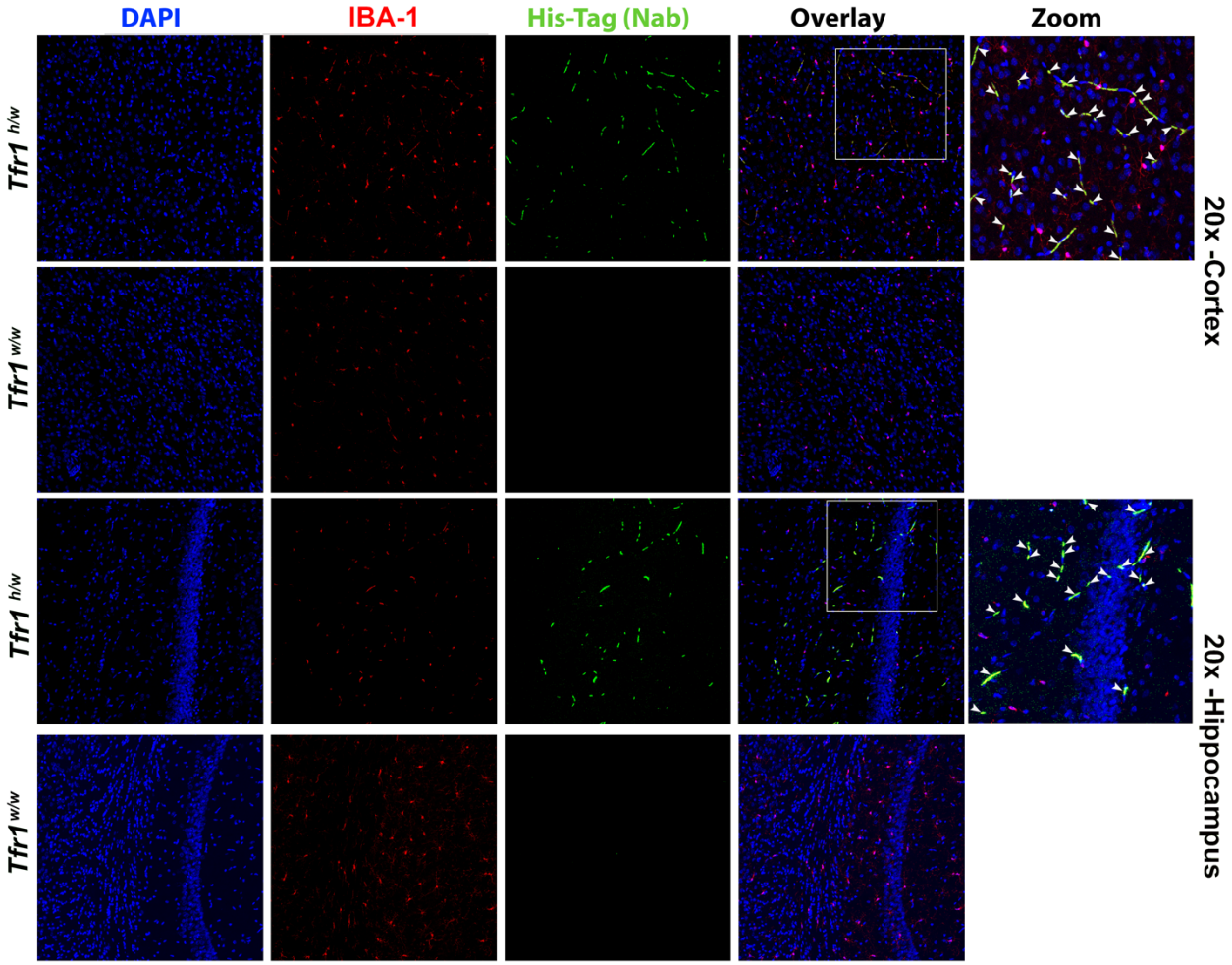


Figure 12. Confocal images showing the localization of TNFI-Nb1-linker-04B05R3 in microglia of the cortex and hippocampus in *Tfr1*$^{h/w}$ and *Tfr1*$^{w/w}$ rats. Sections were stained with an anti-His tag antibody (green) to detect TNFI-Nb1-linker-04B05R3 and an anti-IBA1 antibody (red) to label microglia. TNFI-Nb1-linker-04B05R3 is present in the brains of *Tfr1*$^{h/w}$ rats but not in *Tfr1*$^{w/w}$ rats. In *Tfr1*$^{h/w}$ rats, TNFI-Nb1-linker-04B05R3 colocalizes with most microglia, as indicated by yellow overlays in the merged images. White arrows in the zoomed-in panel point to colocalization spots within the region highlighted by the white square in the overlay panel.

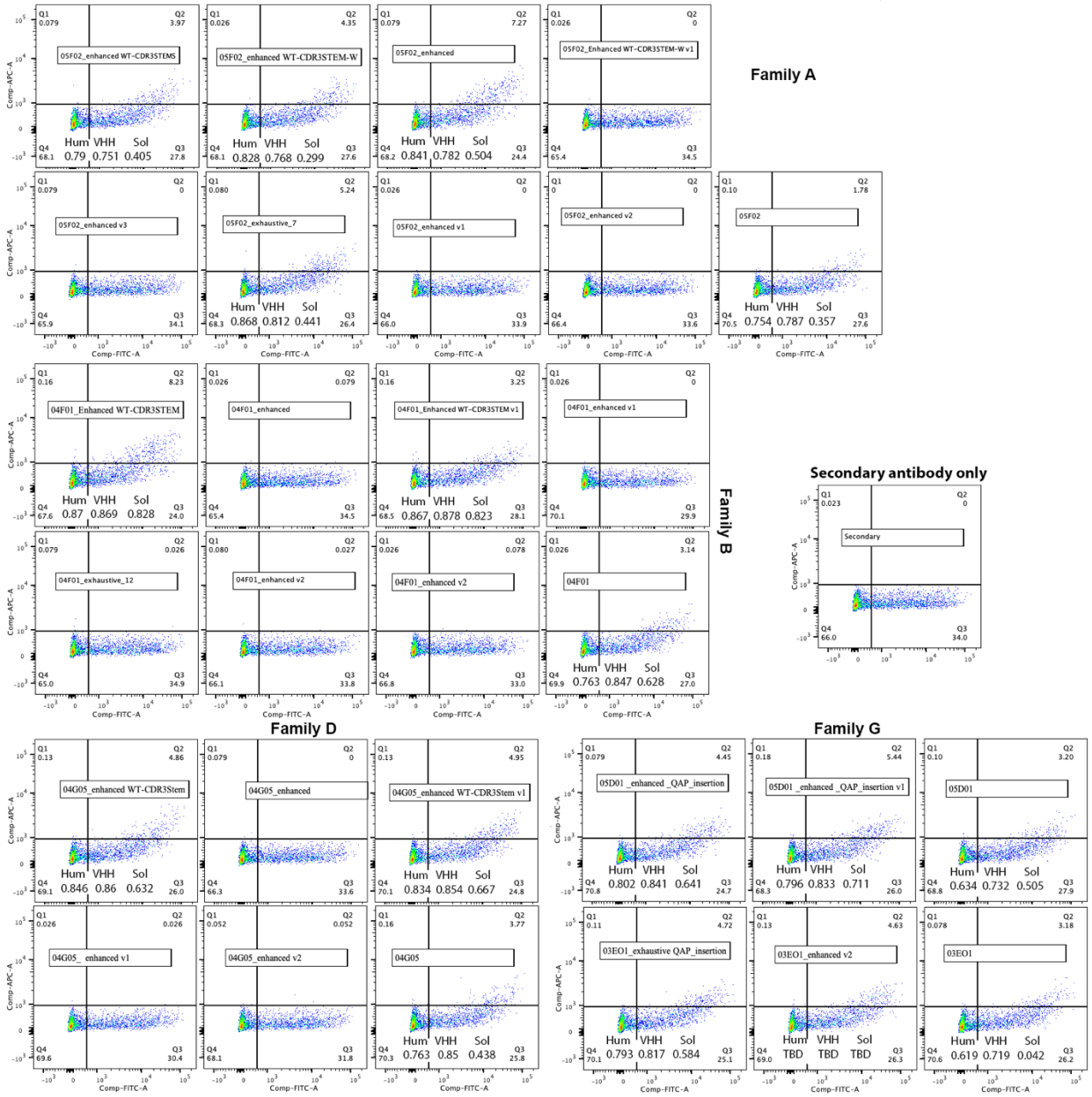


Figure 13. Binding of humanized mutant TfR1b-Nbs to human TfR1. HEK293 cells were transfected with a vector expressing human TfR1 alongside EGFP. The cells were then treated with each TfR1b-Nb at a concentration of 400 nM, followed by incubation with an anti-His-APC antibody. Secondary antibody staining alone is shown as a negative control. Binding of the wild-type TfR1-Nb from which the mutants originated is also shown for comparison.

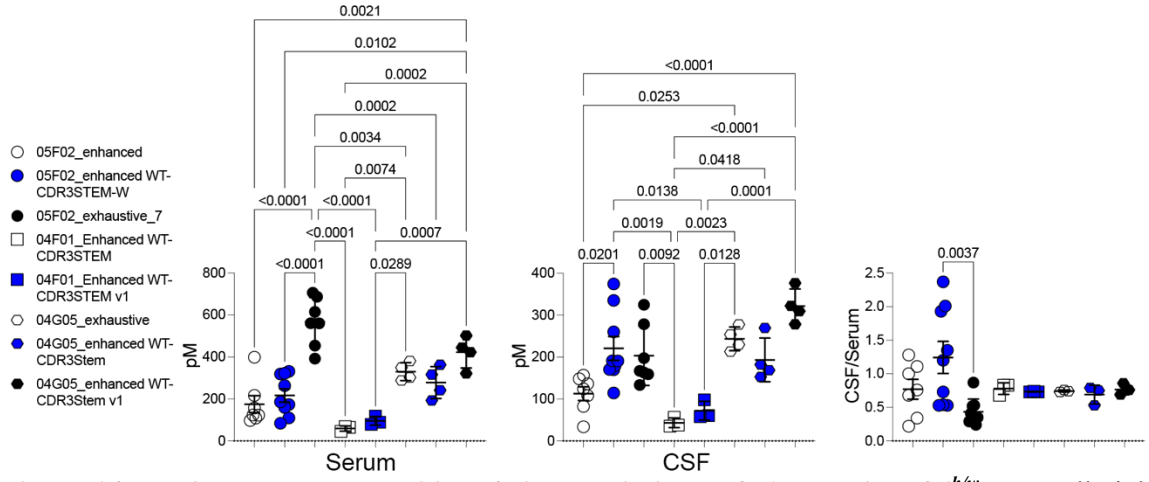
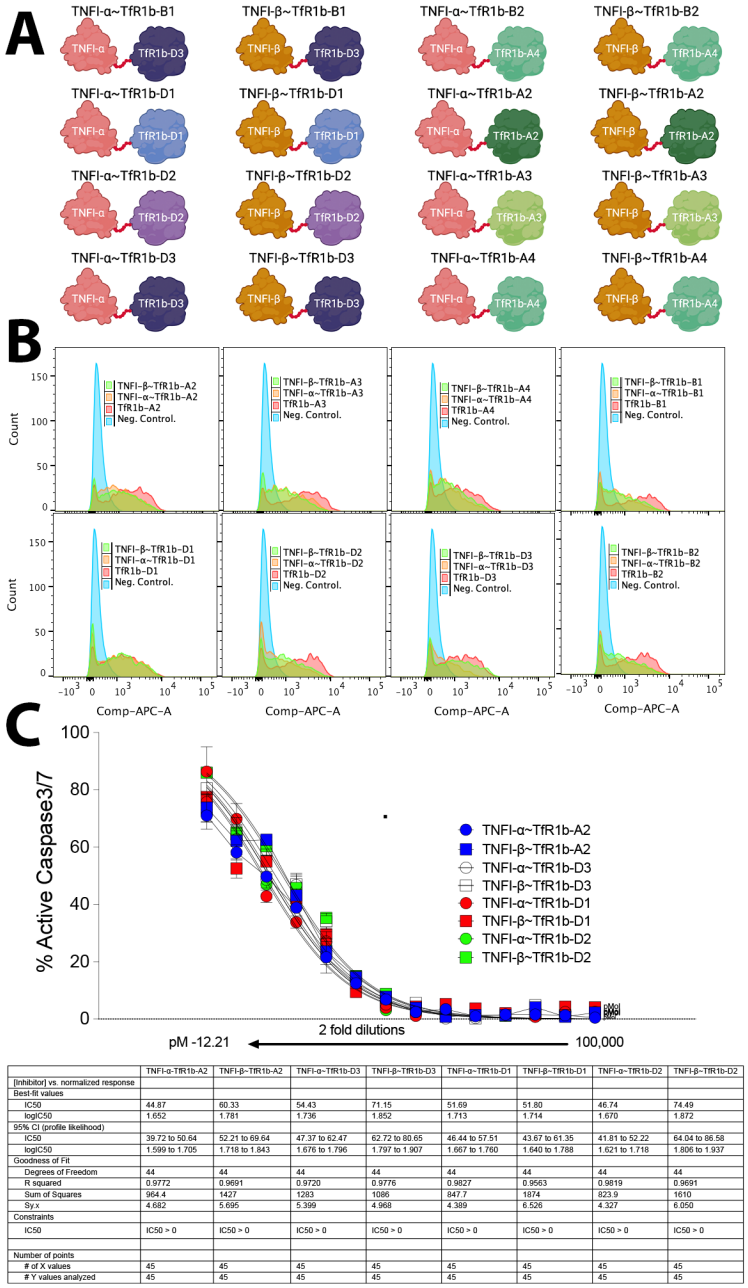


Figure 14. In vivo BBB permeability of eight optimized TfR1b-Nbs in *Tfr1*$^{h/w}$ rats. All eight TfR1b-Nbs exhibited elevated CSF levels following IV injection, confirming human TfR1-dependent BBB transcytosis. Among them, **05F02_enhanced WT-CDR3STEM-W** showed the highest CSF/serum ratio. Details of the animals and nanobody administration are provided in Table 9.



	TNFI-α-TfR1b-A2	TNFI-β~TfR1b-A2	TNFI-α~TfR1b-D3	TNFI-β~TfR1b-D3	TNFI-α~TfR1b-D1	TNFI-β~TfR1b-D1	TNFI-α~TfR1b-D2	TNFI-β~TfR1b-D2
[Inhibitor] vs. normalized response								
Best-fit values								
IC50	44.87	60.33	54.43	71.15	51.69	51.80	46.74	74.49
logIC50	1.652	1.781	1.736	1.852	1.713	1.714	1.670	1.872
95% CI (profile likelihood)								
IC50	39.72 to 50.64	52.21 to 69.64	47.37 to 62.47	62.72 to 80.65	46.44 to 57.51	43.67 to 61.35	41.81 to 52.22	64.04 to 86.58
logIC50	1.599 to 1.705	1.718 to 1.843	1.676 to 1.796	1.797 to 1.907	1.667 to 1.760	1.640 to 1.788	1.621 to 1.718	1.806 to 1.937
Goodness of Fit								
Degrees of Freedom	44	44	44	44	44	44	44	44
R squared	0.9772	0.9691	0.9720	0.9776	0.9827	0.9563	0.9819	0.9691
Sum of Squares	964.4	1427	1283	1086	847.7	1874	823.9	1610
Sy.x	4.682	5.695	5.399	4.968	4.389	6.526	4.327	6.050
Constraints								
IC50	IC50 > 0	IC50 > 0	IC50 > 0	IC50 > 0	IC50 > 0	IC50 > 0	IC50 > 0	IC50 > 0
Number of points								
# of X values	45	45	45	45	45	45	45	45
# Y values analyzed	45	45	45	45	45	45	45	45

Figure 15. TfR1 binding and TNFα inhibitory activity of optimized and humanized heterodimers. A) Schematic representation of the 16 humanized and optimized TNFI–TfR1b heterodimers. **B**) Flow cytometry analysis in HEK293 cells stably expressing human TfR1, comparing the binding of heterodimers to that of their parental humanized TfR1b-Nbs. **C**) Dose-response apoptosis inhibition curves in WEHI-13VAR cells treated with human TNFα and 2-fold serial dilutions of TNFI–TfR1b heterodimers (100,000 to 12.21 pM). Apoptosis was measured using a fluorogenic caspase-3/7 activation assay. The IC_{50} values of the heterodimers were comparable to those of the parental humanized nanobodies TNFI-α (IC_{50} = 48.84 pM, range: 42.11–56.56) and TNFI-β (IC_{50} = 64.31 pM, range: 57.17–72.31).

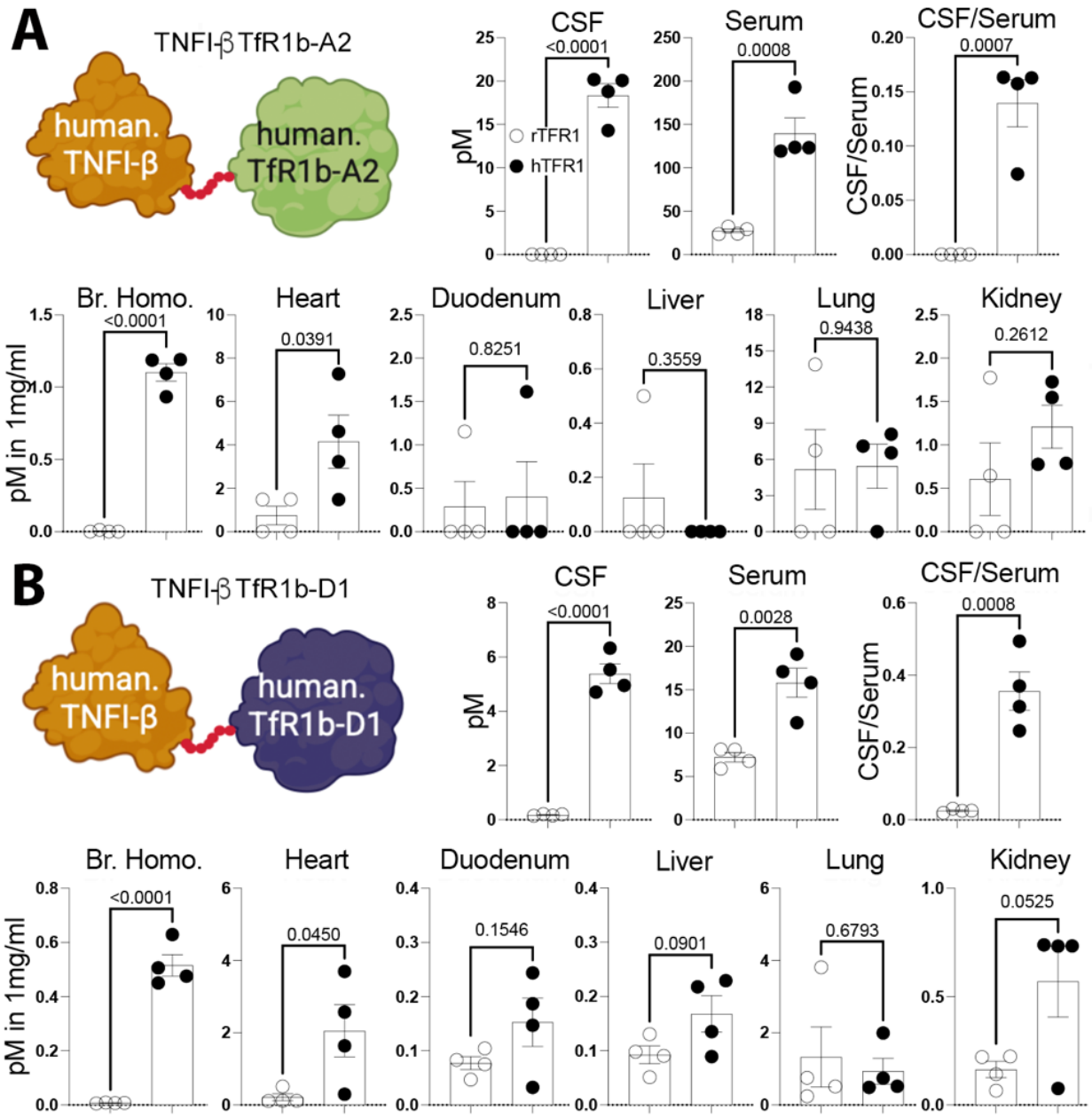


Figure 16. Subcutaneous delivery of humanized heterodimers demonstrates effective BBB transcytosis and extended in vivo stability. *Tfr1*$^{h/w}$ and *Tfr1*$^{w/w}$ rats were injected subcutaneously with TNFI-β–TfR1b-A2 (**A**) and TNFI-β–TfR1b-D1 (**B**). Heterodimers levels were measured 72 hours post-injection. TNFI-β–TfR1b-A2 achieved higher serum and CSF levels (CSF/serum ratio: 0.14), while TNFI-β–TfR1b-D1 showed superior BBB permeability (CSF/serum ratio: 0.37). Heterodimers remained detectable in both CSF and serum three days post-injection, confirming prolonged in vivo stability. Tissue distribution was primarily brain-specific and human TfR1-dependent, with additional enrichment observed in the heart and, potentially, the kidney. with some enrichment in the heart and, potentially, the kidney.

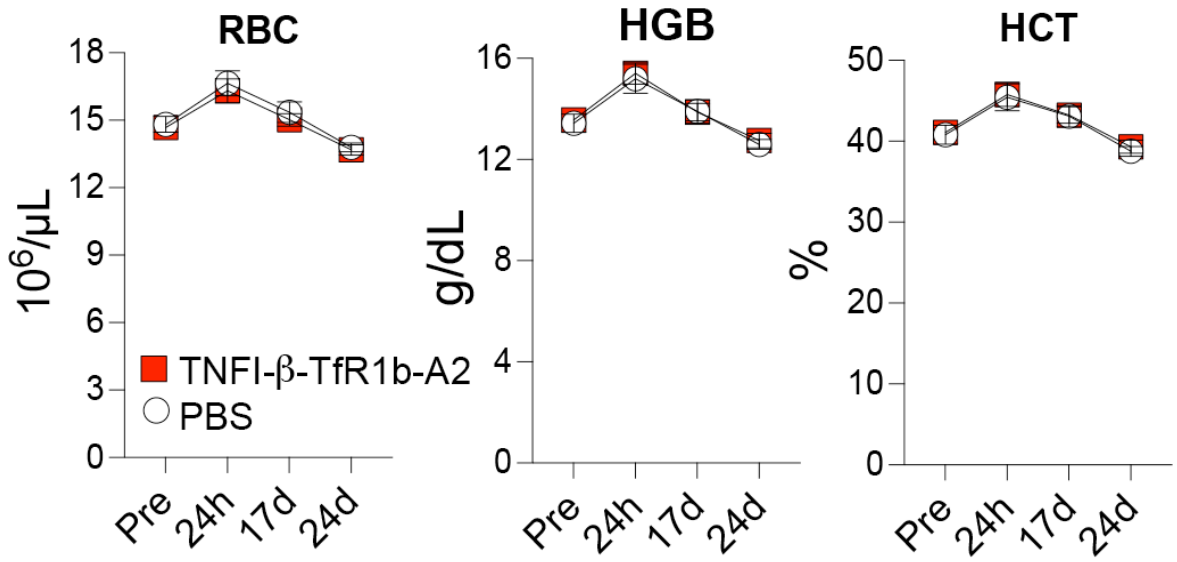


Figure 17. In vivo assessment of hematotoxicity following TNFI-β–TfR1b-A2 administration in humanized rats. CBC analysis on ~4-month-old rats humanized for both TF and TfR1 (*Tf*$^{h/w}$:*Tfr1*$^{h/h}$), following administration of TNFI-β–TfR1b-A2 or PBS. No significant changes were observed in key hematological parameters—including RBC, hemoglobin (HGB), and hematocrit (HCT)—compared to PBS-treated controls. All values remained within normal physiological ranges, indicating no detectable hematotoxicity. These results suggest that TNFI-β–TfR1b-A2 is well tolerated in vivo and unlikely to induce anemia through interference with human TF–TfR1–mediated iron transport.

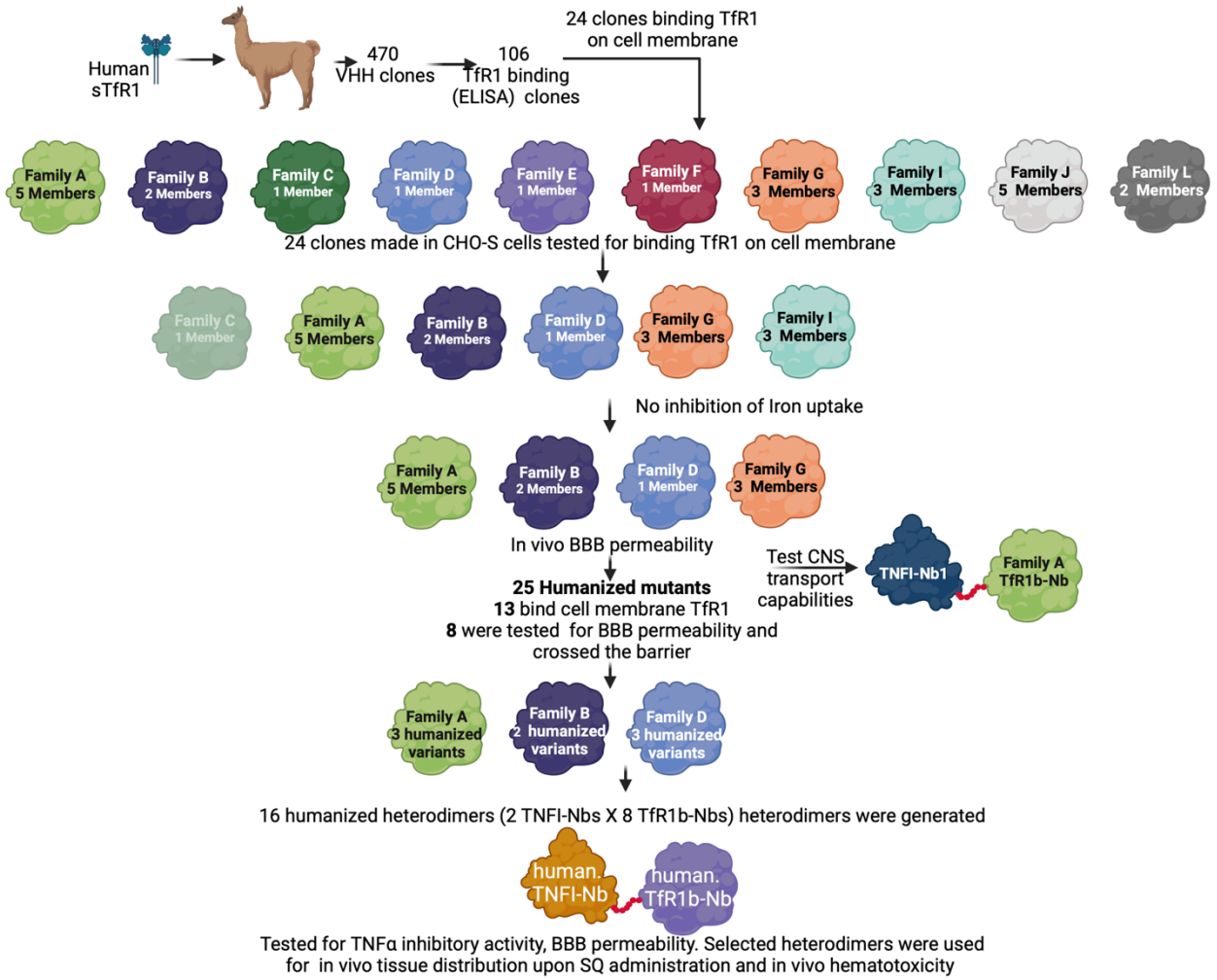


Figure 18. Schematic representation of the workflow for the identification of NewroBus molecules. A step-by-step overview illustrating the process used to discover and optimize NewroBus nanobodies for efficient BBB transcytosis and CNS delivery. The Family C nanobody was not tested.

Tables

Name	Family	Conc. (mg/ml)	Purity SDS-PAGE (%)	Purity SEC-HPLC (%)	Endotoxin Level (EU/mg)	Total (mg)
02F06R3	A	0.45	75	92	0.111	~6.525
05D05R3	A	0.44	85	94	0.114	~6.38
05F02R3	A	0.46	90	94	0.109	~6.67
02B02R3	A	0.52	90	95	0.096	~7.54
04B05R3	A	0.53	50	96	0.094	~7.685
04D08R3	B	0.46	95	91	0.109	~6.67
04F01R3	B	0.42	80	89	0.119	~6.09
04G05R3	D	0.42	90	88	0.119	~6.09
05G07R3	E	0.29	70	83	0.172	~2.75
04A02R3	F	0.67	70	88	0.075	~9.045
05D01R3	G	0.6	75	89	0.083	~8.4
03E01R3	G	0.81	80	90	0.062	~11.34
04E01R3	G	0.88	70	92	0.057	~12.58
05D04R3	J	0.36	85	95	0.139	~5.04
01D03R3	J	0.84	75	94	0.064	~12.18
05C08R3	J	0.68	70	91	0.074	~9.18
01B03R3	J	0.73	80	94	0.068	~10.22
03F03R3	J	0.79	90	93	0.063	~11.06
01B01R3	I	0.92	95	96	0.054	~12.88
01B06R3	I	0.58	75	86	0.086	~8.7
03G10R3	I	0.79	85	88	0.063	~11.85
03B05R3	L	0.85	85	84	0.059	~12.75
01A02R3	L	0.72	70	83	0.069	~10.8

Table 1. Production of TfR1b-Nbs in mammalian cells was performed as for TNFI-Nbs. The table 1 shows quantity and purity of TfR1b-Nb purified from 100 ml culture supernatants.

Nanobody	Family	*Tfr1*	sex	Age in days	Time (hrs)	Conc. in Serum [pM]	Conc. in Brain homog. [pM]
02B02R3	A	*h/h*	f	70	16-18	33.2	28.6
05F02R3	A	*h/h*	f	70	16-18	22.7	28.0
04F01R3	B	*h/h*	m	39	16-18	128.1	241.2
04G05R3	D	*h/h*	m	39	16-18	414.8	624.4
03E01R3	G	*h/h*	m	31	16-18	559.7	843.8
04E01R3	G	*h/h*	f	34	16-18	388.2	796.9
01B01R3	I	*h/h*	m	122	16-18	60.6	109.9
01B01R3	I	*h/h*	f	34	16-18	60.4	299.0
03G10R3	I	*h/h*	m	121	16-18	13.0	48.4
02B02R3	A	*h/w*	m	114	16-18	42.4	94.0
02F06R3	A	*h/w*	m	61	16-18	71.2	161.8
04B05R3	A	*h/w*	f	107	12	478.5	1058.6
04B05R3	A	*h/w*	f	237	16-18	174.1	525.2
05D05R3	A	*h/w*	m	237	16-18	25.0	39.0
05F02R3	A	*h/w*	f	147	16-18	33.4	109.3
04D08R3	B	*h/w*	m	147	16-18	66.7	498.6
04D08R3	B	*h/w*	m	34	16-18	25.4	48.2
04F01R3	B	*h/w*	m	114	16-18	88.5	635.1
04G05R3	D	*h/w*	m	147	16-18	121.9	847.2
03E01R3	G	*h/w*	f	64	16-18	158.3	663.6
04E01R3	G	*h/w*	f	212	16-18	136.6	724.3
05D01R3	G	*h/w*	f	94	12	65.3	398.1
05D01R3	G	*h/w*	m	61	16-18	18.6	306.8
01B01R3	I	*h/w*	m	149	16-18	7.6	74.5
01B06R3	I	*h/w*	m	65	16-18	6.3	34.1
03G10R3	I	*h/w*	m	51	16-18	7.8	150.1
02F06R3	A	*w/w*	m	49	16-18	0.0	0.0
04B05R3	A	*w/w*	f	217	16-18	0.0	0.0
05D05R3	A	*w/w*	m	217	16-18	2.8	0.0
05D01R3	G	*w/w*	m	51	16-18	0.0	0.0
04G05R3	D	*w/w*	f	139	12	0.0	0.0

Table 2. Detection of TfR1b-Nbs in brain samples of rats expressing human TfR1, compared with rat Tfr1-expressing animals. Columns 1–6 list the nanobody name, family, *Tfr1* genotype, sex, age, and time of tissue collection post-injection. Columns 7 and 8 report the concentrations of TfR1b-Nbs in serum and brain homogenates, respectively. Data for this experiment are shown in Figure 6A.

Nanobody	Family	*Tfr1*	sex	Age in days	Time (hrs)	Serum [pM]	CSF[pM]	CSF/Serum	CNS [pM]
04B05R3	A	*h/w*	m	43	44-48	14.101	22.706	1.61	4.431
04B05R3	A	*h/w*	m	43	44-48	17.371	20.066	1.16	7.593
04B05R3	A	*h/w*	f	42	44-48	4.927	6.6769	1.36	1.83
05D01R3	G	*h/w*	m	44	44-48	43.093	32.691	0.76	22.517
05D01R3	G	*h/w*	m	44	44-48	39.896	21.941	0.55	19.183
05D01R3	G	*h/w*	f	44	44-48	39.326	21.714	0.55	16.104
04B05R3	A	*w/w*	m	42	44-48	1.226	0	0	0
04B05R3	A	*w/w*	m	44	44-48	0.498	0	0	0
05D01R3	G	*w/w*	m	42	44-48	0	0	0	0
05D01R3	G	*w/w*	m	43	44-48	12.371	0	0	0

Table 3. Detection of TfR1b-Nbs in brain samples of rats expressing human TfR1, compared with rat Tfr1-expressing animals. A human TfR1-specific ELISA detected TfR1b-Nbs in brain tissues and CSF of *Tfr1*$^{h/w}$ rats, but not in *Tfr1*$^{w/w}$ controls. Columns 1–6 list the nanobody name, family, *Tfr1* genotype, sex, age, and time of tissue collection post-injection. Columns 7–10 report the concentrations of TfR1b-Nbs in serum, CSF, the CSF/serum ratio, and brain homogenates, respectively. Data for this experiment are shown in Figure 6B.

Nanobody	*Tfr1*	sex	Age in days	Time (hrs)	Serum [pM]	CNS [pM]
TNFI-Nb1	*h/w*	f	48	6	14.81634	0
TNFI-Nb1	*h/w*	f	48	6	5.659766	0
TNFI-Nb1	*h/w*	m	40	6	8.185005	0
TNFI-Nb1	*h/w*	f	108	6	57.38742	0
TNFI-Nb1	*w/w*	f	139	6	226.9179	0
TNFI-Nb1	*w/w*	f	39	6	55.92783	0
TNFI-Nb1	*w/w*	m	39	6	57.97242	0
TNFI-Nb1	*w/w*	m	39	6	24.78752	0

Table 4. Details of the animals used in the experiment presented in Figure 7, including genotype, sex, age, and nanobody administered.

Nanobody	Family	*Tfr1*	sex	Age in days	Time (hrs)	Serum [pM]	CSF[pM]	CSF/Serum	CNS [pM]
TNFI-Nb1-04B05R3	A	*h/w*	m	44	44-48	5.14	4.25	0.83	6.23
TNFI-Nb1-04B05R3	A	*h/w*	f	44	44-48	5.32	4.95	0.93	6.17
TNFI-Nb1-04B05R3	A	*h/w*	m	44	44-48	6.21	4.73	0.76	8.96
TNFI-Nb1-04B05R3	A	*h/w*	f	51	44-48	5.24	6.88	1.31	14.18
TNFI-Nb1-04B05R3	A	*h/w*	m	50	44-48	12.23	15.16	1.24	25.73
TNFI-Nb1-04B05R3	A	*h/w*	m	51	44-48	11.91	18.08	1.51	22.40
TNFI-Nb1-04B05R3	A	*w/w*	m	51	44-48	0.91	0.004	0.004	0.00
TNFI-Nb1-04B05R3	A	*w/w*	f	44	44-48	2.30	0.004	0.001	0.03
TNFI-Nb1-04B05R3	A	*w/w*	f	44	44-48	1.56	0.002	0.001	0.00

Table 5. Shuttle Activity of TfR1b-Nbs for TNFI-Nb1 in brain samples of rats expressing human TfR1. A human TNFα-specific ELISA detected TNFI-Nb1-04B05R3 in brain tissues and CSF of *Tfr1*$^{h/w}$ rats, but not in *Tfr1*$^{w/w}$ controls. The linker used to link these two nanobodies was: GGGGSGGG. Columns 1–6 list the nanobody name, family, *Tfr1* genotype, sex, age, and time of tissue collection post-injection. Columns 7–10 report the concentrations of TfR1b-Nbs in serum, CSF, the CSF/serum ratio, and brain homogenates, respectively. Data for this experiment are shown in Figure 8.

ID	F	H	V	S
02F06	A	0.721	0.744	0.382
05D05		0.697	0.783	0.405
05F02		0.754	0.787	0.357
02B02		0.745	0.785	0.291
04B05		0.734	0.772	0.304
04D08	B	0.774	0.831	0.620
04F01		0.763	0.847	0.628
04G05	D	0.763	0.850	0.438
05D01	G	0.634	0.732	0.505
03E01		0.619	0.719	0.042
04E01		0.601	0.735	0.026

Table 6. AbNatiV Assessment of TfR1b-Nbs. Key metrics include Humaness (H), VHH-ness (VH), and CamSol Intrinsic (S). The Nbs underlined, in bold and italic have been selected for humanization.

ID	H	V	S	Code	Fam.
05F02	0.754	0.787	0.357		
Enhanced WT-CDR3STEMS	0.790	0.751	0.405		
enhancedWT-CDR3STEM-W	0.828	0.768	0.299	**A2**	
enhanced	0.841	0.782	0.504	**A3**	
Enhanced WT-CDR3STEM-W v1	0.821	0.769	0.598		**A**
enhanced v3	0.840	0.753	0.602		
exhaustive_7	0.868	0.812	0.441	**A4**	
enhanced v1	0.812	0.749	0.800		
enhanced v2	0.827	0.769	0.802		
04F01	0.763	0.847	0.628		
Enhanced WT-CDR3STEM	0.870	0.869	0.828	**B1**	
enhanced	0.882	0.866	0.960		
Enhanced WT-CDR3STEM v1	0.867	0.878	0.823	**B2**	
enhanced v1	0.884	0.850	1.029		**B**
exhaustive_12	0.899	0.876	0.978		
enhanced v2	0.882	0.869	0.991		
enhanced v3	0.884	0.858	1.023		
04G05	0.763	0.850	0.438		
Enhanced WT-CDR3Stem	0.846	0.860	0.632	**D1**	
enhanced	0.854	0.863	0.847		
Enhanced WT-CDR3Stem v1	0.834	0.854	0.667	**D2**	**D**
exhaustive	0.843	0.841	0.572	**D3**	
enhanced v1	0.814	0.838	0.939		
enhanced v2	0.831	0.854	0.888		
05D01	0.763	0.850	0.438		
enhanced_QAP_insertion	0.802	0.841	0.641		
enhanced_QAP_insertion v1	0.796	0.833	0.711		**G**
03E01	0.619	0.719	0.042		
exhaustive QAP_insertion	0.793	0.817	0.584		
enhanced	0.784	0.820	0.500		

Table 7. TfR1b-Nb mutants with enhanced therapeutic potential determined by AbNatiV analysis. Changes in Humanness (H), VHH-ness (VH), and CamSol Intrinsic (S) are presented in the last three columns.

Nanobody	Family	*Tfr1*	sex	Age in days	Time (hrs)	Conc. in Serum [pM]	Conc. in CSF [pM]	CSF/ Ser.
05F02_enhanced	A	*h/h*	m	131	16-24	122.75	156.99	1.28
05F02_enhanced	A	*h/h*	m	131	16-24	216.82	148.09	0.68
05F02_enhanced	A	*h/h*	m	100	16-24	123.96	124.15	1.00
05F02_enhanced	A	*h/h*	m	100	16-24	97.37	108.01	1.11
05F02_enhanced	A	*h/w*	m	212	16-24	399.12	136.37	0.34
05F02_enhanced	A	*h/w*	f	212	16-24	156.72	33.77	0.22
05F02_enhanced	A	*h/w*	f	212	16-24	108.90	82.48	0.76
05F02_enhanced WT-CDR3STEM-W	A	*h/h*	m	131	16-24	319.32	169.85	0.53
05F02_enhanced WT-CDR3STEM-W	A	*h/h*	m	131	16-24	331.44	179.84	0.54
05F02_enhanced WT-CDR3STEM-W	A	*h/h*	m	131	16-24	322.09	170.56	0.53
05F02_enhanced WT-CDR3STEM-W	A	*h/h*	m	100	16-24	263.54	191.09	0.73
05F02_enhanced WT-CDR3STEM-W	A	*h/w*	m	82	16-24	109.81	260.22	2.37
05F02_enhanced WT-CDR3STEM-W	A	*h/w*	f	94	16-24	173.53	335.17	1.93
05F02_enhanced WT-CDR3STEM-W	A	*h/w*	f	96	16-24	159.62	190.97	1.20
05F02_enhanced WT-CDR3STEM-W	A	*h/w*	f	94	16-24	186.12	374.53	2.01
05F02_enhanced WT-CDR3STEM-W	A	*h/w*	m	82	16-24	84.60	114.36	1.35
05F02_exhaustive_7	A	*h/h*	m	131	16-24	615.23	324.83	0.53
05F02_exhaustive_7	A	*h/h*	m	100	16-24	560.37	197.48	0.35
05F02_exhaustive_7	A	*h/h*	m	131	16-24	687.31	278.54	0.41
05F02_exhaustive_7	A	*h/h*	m	100	16-24	392.66	133.47	0.34
05F02_exhaustive_7	A	*h/w*	m	87	16-24	561.22	163.28	0.29
05F02_exhaustive_7	A	*h/w*	f	212	16-24	454.19	159.10	0.35
05F02_exhaustive_7	A	*h/w*	f	225	16-24	706.45	167.02	0.24
04F01_Enhanced WT-CDR3STEM	B	*h/w*	m	92	16-24	64.86	56.40	0.87
04F01_Enhanced WT-CDR3STEM	B	*h/w*	f	92	16-24	68.06	35.65	0.52
04F01_Enhanced WT-CDR3STEM	B	*h/w*	f	90	16-24	45.49	37.75	0.83
04F01_Enhanced WT-CDR3STEM v1	B	*h/w*	m	92	16-24	118.29	98.00	0.83
04F01_Enhanced WT-CDR3STEM v1	B	*h/w*	f	92	16-24	89.13	60.64	0.68
04F01_Enhanced WT-CDR3STEM v1	B	*h/w*	f	90	16-24	79.08	57.87	0.73
04G05_exhaustive	D	*h/w*	m	84	16-24	381.19	277.88	0.73
04G05_exhaustive	D	*h/w*	f	89	16-24	288.57	213.07	0.74
04G05_exhaustive	D	*h/w*	m	89	16-24	302.11	229.89	0.76
04G05_exhaustive	D	*h/w*	f	91	16-24	349.42	253.78	0.73
04G05_enhanced WT-CDR3Stem	D	*h/w*	m	82	16-24	362.82	269.43	0.74
04G05_enhanced WT-CDR3Stem	D	*h/w*	f	91	16-24	316.27	168.69	0.53
04G05_enhanced WT-CDR3Stem	D	*h/w*	m	89	16-24	241.76	182.20	0.75
04G05_enhanced WT-CDR3Stem	D	*h/w*	f	89	16-24	192.70	152.89	0.79
04G05_enhanced WT-CDR3Stem v1	D	*h/w*	m	84	16-24	321.78	278.02	0.86
04G05_enhanced WT-CDR3Stem v1	D	*h/w*	f	94	16-24	445.31	309.46	0.69
04G05_enhanced WT-CDR3Stem v1	D	*h/w*	f	94	16-24	423.02	322.14	0.76
04G05_enhanced WT-CDR3Stem v1	D	*h/w*	f	96	16-24	502.33	376.44	0.75

Table 8. Assessment of BBB permeability of optimized TfR1b-Nbs.

D -3	D0	D1	D7	D14	D17	D21	D24
CBC	Inj.	CBC	Inj.	Inj.	CBC	Inj.	CBC

Table 9. Hematotoxicity assessment. Rats were injected IV on the indicated days (inj.) with either PBS (G1, 3♂/4♀), or TNFI-β-TfR1b-A2 (G2, 3♂/5♀) (1 µL of a 40 µM INN solution in PBS per gram of rat body weight). Complete blood counts (CBC) were performed at several time points: 3 days before the first injection (D -3), 24 hours after the first injection (D1), on Day 17 following three injections, and on Day 24 after four injections. The CBC measurements included white blood cells (WBC), neutrophils (NEU), lymphocytes (LYM), monocytes (MONO), eosinophils (EOS), basophils (BAS), as well as their percentages (NEU %, LYM %, MONO %, EOS %, BAS %), red blood cells (RBC), hemoglobin concentration (HGB), hematocrit (HCT), mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), red cell distribution width (RDW %), platelet count (PLT), and mean platelet volume (MPV).