No evidence for disassortative mating based on HLA genotype in a small-scale, endogamous population

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Abstract

Despite important early work suggesting humans prefer mates with dissimilar HLA genotypes, overall evidence for disassortative mating based on the human-specific MHC remains inconclusive. Recent studies have instead demonstrated assortative mating associated with cosmopolitan population structure, suggesting that social stratification may play a much stronger role in mate preferences. However, additional work in small-scale endogamous populations has been largely overlooked until now, even though this type of social structure characterized most of human evolution. Here, we evaluate HLA dissimilarity in one such group, Himba pastoralists from Namibia who have multiple relationship categories. Importantly, we compare partners who were in arranged marriages with those in chosen partnerships, i.e., "love matches" (n=249 observed partnerships). We find no difference in HLA dissimilarity (at the genotype and protein divergence level) between partnership types, nor in their fitness benefits to potential offspring as assessed via computationally predicted pathogen binding affinities. The effects of the partnership types likewise do not differ from a random, background distribution of 18,487 possible unrelated pairings. We do find extensive haplotype sharing at the HLA region, suggesting that episodes of fluctuating positive selection may be a more important driver of HLA polymorphism than disassortative mating.

1. Introduction
MHC dissimilarity is predicted to play a critical role in mate choice strategies, as choosing a dissimilar mate enhances offspring immune function by increasing their heterozygosity and therefore broadening pathogen resistance. However, empirical evidence of this preference is lacking, with multiple meta-analyses failing to show significant effects across a range of vertebrate species(1, 2). The literature on humans has followed a similar path, with early evidence showing promising results in both experimental work(3) and in two ethnically and genetically homogeneous populations(4, 5), but inconclusive results in the decades since(1, 6). In fact, rather than disassortative preferences, a recent meta-analysis of the human data showed evidence of the opposite trend, assortative mating(1).

These recent studies have largely focused on Western, heterogeneous study populations with cosmopolitan population structure, exhibiting more admixture and genetic diversity than small-scale societies. Studies in such populations can obscure patterns of disassortative mating due to cultural and behavioral preferences for ethnic similarity within a heterogeneous population. This can produce patterns of relatedness genome-wide, including at the HLA, obscuring more subtle mate preference effects–an obvious limitation affecting the majority of observational mate choice studies(1, 6). Further, of the few studies that have been conducted in non-Western, smaller scale societies(5, 7), sociocultural factors such as preferences for consanguineous marriages or arranged marriage systems have obscured the assessment of innate mate preferences.

Stronger tests for disassortative mating would benefit from sampling in smaller-scale endogamous populations, which are more reflective of human populations for the majority of human history and are less likely to show assortative mating resulting from cosmopolitan population structure. Such studies would also benefit from incorporating critical features of the local mating and marriage system. The current study aims to address both these gaps.

We aim to directly test the HLA dissimilarity hypothesis with a group of Himba agropastoralists living in northern Namibia, combining genetics with ethnographic and demographic data that help to disentangle these factors. Several features of the Himba marital system make them a particularly useful population for testing this hypothesis. First, marriages can be either arranged (via parental choice) or chosen ("love matches")(8). If there is an innate preference for HLA dissimilarity when selecting mates, we expect stronger effects among love matches than among couples in arranged marriages. Additionally, concurrent marital and nonmarital partnerships are common among Himba, providing another avenue for studying active mate choice, even when individuals are in an arranged marriage(9–11). We use paired genetic data (HLA and genome-wide) and partnership status to investigate whether there is greater HLA dissimilarity, at the genotype and protein sequence level, in unions resulting from partner choice than in those that are arranged. We also examine the pathogen binding ability of hypothetical offspring of arranged and chosen partnerships to test for any direct fitness benefits of parental HLA allele combinations.

2. Methods

(a) Study Population

The Himba are an agro-pastoralist group residing in Northwestern Namibia and Southern Angola, who have a total population size of around 30,000-40,000 individuals(12). The groups remain semi-nomadic but increasingly have access to formal education and the market economy. The present study was conducted in the community of Omuhonga as part of the Kunene Rural Health and Demography Project, which has been working in the community since 2010. There are about 1000 residents of Omuhonga, living in extended family compounds. Marriages are arranged by parents or other kin, although "love match" marriages are common, particularly after the first marriage(8). Extra-marital relationships are also common for both men and women(9).

(b) Data Generation

DNA was collected with informed consent from individuals via saliva using the prepIT-L2P kit and protocol. DNA was used to generate HLA genotype calls for 366 individuals. HLA class I and II loci were targeted for DNA sequencing using a well-established biotinylated DNA probe-based capture method(13). HLA alleles were determined from the sequence capture using the consensus calls obtained from two algorithms: NGSengine® 2.10.0 (GenDX, Utrecht, the Netherlands) and HLA Explore™ (Omixon Biocomputing Ltd. Budapest, Hungary). Alleles were then converted into their P-group designations from https://hla.alleles.org as of 10/7/2024 (2-field designation), which groups alleles whose nucleotide sequences encode identical protein sequence in the peptide binding domains (exon 2 and 3 for HLA class I and exon 2 for HLA class II alleles). Genotype array data using one of two arrays, Illumina

101 H3Africa or MEGAex, were previously generated for 360 of these individuals [dbGaP:
102 phs001995.v3.p1]. One individual was newly genotyped using H3Africa. Five individuals with HLA
103 capture data did not have corresponding SNP array data and were not included in the partnership analyses
104 requiring genome-wide relatedness metrics. These SNP array datasets previously underwent quality
105 control measures and were merged as previously described(10).
106
There were 219 adults (82 males, 137 females) having both genotype array and HLA genotype data who
were in at least one documented partnership: arranged marriage, love match marriage, self-reported
informal partnership, or a discovered informal partnership. Marriages and self-reported informal
partnerships were recorded during demographic interviews conducted between 2010 and 2017. Additional
informal partnerships were discovered due to shared biological offspring as determined by a list of parent
offspring trios generated via KING 2.1.3(14), using the identical by descent (IBD) segment flag. There
were 47 arranged marriages, 47 love match marriage pairs, and 155 informal pairs. Of the informal pairs,
48 were discovered only through shared biological offspring. These pairings may have been excluded
during the interviews because they occurred many years in the past and our interviews only asked about
current partnerships. The 47 love match marriages were combined with 203 informal partnerships and
designated as chosen partnerships to compare with the 47 arranged partnerships in our analyses.
118
(c) Genotype array data quality control
Individuals were genotyped using one of two arrays, H3Africa or MEGAex. These datasets previously
underwent quality control measures and were filtered using PLINK v1.90b3v(15) for missingness greater
than 5%, a MAF less than or equal to 1%, and a Hardy–Weinberg equilibrium exact test with a *p*-value
below 0.0001 as described previously(10) and documented in dbGaP accession phs001995.v3.p1. The
merged array dataset containing 376,657 SNPs was used for analyses.
125
(d) Creating a set unrelated sets of individuals
Traditional pedigree inference methods that use expected IBD sharing perform poorly in endogamous
populations with elevated background sharing and pedigree reticulations. We used PONDEROSA(16),
recently developed in our lab, to accurately infer close relationships in our sample. PONDEROSA uses a
machine learning approach to learn the population distribution of IBD summary statistics to more
accurately classify close relationships. Based on the inferred relations we excluded any pairs who were 2nd
degree or closer relatives from our background distributions in our linear models (n) Statistical Methods
eq 2-3). Third degree relative pairs were permitted in this background distribution to reflect the stated
cultural preference for cousin marriages in arranged unions(8). We also used this program to maximize a
set of unrelated individuals (n=102 individuals containing no 3rd degree or closer relatives) for allele and
haplotype frequency based analyses.
137
(e) Hardy-Weinberg Analysis
To test whether the HLA genotypes complied with Hardy-Weinberg equilibrium, we used the Asymptotic
Statistical Test with Ambiguity (ASTA) from the HWETests package(17). ASTA is designed to evaluate
deviations from Hardy-Weinberg equilibrium for highly polymorphic loci while accounting for ambiguity
in genotype calls. Using ASTA, we ran the full_algorithm() function with default parameters on the set of
102 unrelated individuals (maximum relation of 4th degree).
144
(f) Identity by Descent (IBD) Sharing
Genotype data was phased using SHAPEIT v2.r837(18) with --duohmm -W 5 using an ancestry-matched
recombination map(19), constructed based on linkage disequilibrium patterns in the 1000 Genomes
Project(20) Yorubans, matching the predominant West African ancestry present in the Himba. We called
IBD segments with phasedIBD(21) using default parameters with phase correction turned off. We
determined phasedIBD calls to be slightly more accurate than germline 1.5.3(22) and GERMLINE2(23)
by comparing the total cM of IBD shared between parent offspring pairs (Table S1). We expected parents

and offspring to share the full length of the recombination map in IBD1 and share an additional
background amount of IBD2 estimated to be the average cM of IBD shared between unrelated (> 3rd
degree relations) individuals in the population. The phasedIBD calls without phase correction most
closely matched this expectation. The phasedIBD calls with phase correction option turned on were the
least accurate. A minimum threshold of 3cM was used for all runs.
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(g) HLA similarity score
We assessed HLA similarity in terms of the number of alleles shared across the eight typed HLA genes
for each pair (minimum of 0 if no alleles are shared at any gene, maximum of 16 if both alleles at each
gene are shared). A shared allele in a homozygous state in one individual only counted as one shared
allele between the individuals. Importantly, due to the many alleles at each HLA gene, two individuals
with heterozygous genotypes at a locus do not necessarily imply shared alleles.
164
(h) Average log odds of homozygous offspring
Odds of homozygous offspring for each locus were determined according to standard mendelian
transmission probabilities, accounting for the multi-allelic states of each gene. Table S2 shows the
possible states for this variable considering only one gene. Importantly, for a given number of alleles
shared at a locus, there are multiple possibilities for the odds of homozygous offspring depending on the
genotype states of the individuals in the pair. We log transformed these odds to normalize the distribution
and then averaged across the 8 loci. We included a +0.1 boundary correction in the numerator and
denominator of the odds calculations to allow for numerators and denominators of 0 (eq 1).

$eq\ (1)\qquad Average\ log\ odds\ of\ homozygosity\ =\ \dfrac{\sum_{i=1}^{8} log(Odds_i)}{8}\quad$ where

$Odds_i\ is\ one\ of\ the\ values\ in\ set\ \{\ \dfrac{0+0.1}{4+0.1}\ ,\ \dfrac{1+0.1}{3+0.1}\ ,\ \dfrac{2+0.1}{2+0.1}\ ,\ \dfrac{4+0.1}{0+0.1}\ \}$

175
(i) Peptide Binding Groove Protein Divergence
We used a script available from a previous publication(24) to assess the mean Grantham amino acid
distance(25) of the peptide binding groove between HLA molecule allotypes, comparing the 4 pairwise
combinations of allotypes between two individuals. Only peptide binding groove sequences (exons 2 and
3 for HLA class I and exon 2 for HLA class II) were compared because all alleles were grouped
according to their P group (identical peptide binding grooves). We used the fasta files provided for the
HLA class I, DRB1, and DQB1 alleles and the Grantham distance matrix available in a previously
published package (https://granthamdist.sourceforge.io/). For the remaining HLA class II alleles we used
the IPD-MHC (https://www.ebi.ac.uk/ipd/mhc/alignment) multiple sequence alignment tool to align
alleles and extract exon 2 amino acid sequences.
186
(j) Pathogen Proteins
We modified a previously published list of representative human pathogen proteins(24) that was curated
from the Gideon database(26) based on global distributions, potential for high mortality, and historical
relevance. After personal correspondence with an infectious disease expert, Dr. Ashley Hazel, who has
extensive experience working with the Himba community we removed 6 pathogens unlikely to be
ecologically relevant in Northern Namibia: *Entamoeba histolytica* (amebiasis), *Trichinella spiralis*
(roundworm), *Schitosoma mansoni* (schistosomiasis), HIV, Hepatitis C, and *Yersinia pestis* (plague). We
added 4 sexually transmitted diseases known to be highly prevalent in the Himba (HSV1, HSV2,
Chlamydia trachomatis (chlamydia), and *Neisseria gonorrhoeae* (gonorrhea)) for a total of 25
pathogens(27). We used the NCBI accession numbers available in the previously published study(24) to
gather amino acid sequences for the antigenic proteins of the 21 pathogens overlapping that publication.
For the 4 additional pathogens we used IEDB https://www.iedb.org/ to find the pathogen proteins that are
MHC ligands and then identified these proteins' amino acid sequences using NCBI. The complete list of
298 proteins and their accession numbers are available in Appendix 1.

(k) Pathogen Binding Prediction

We used NetMHCpan (v4.1)(28) to predict peptide binding affinity for the allotypes of HLA class I (HLA-A, B, and C) and used NetMHCIIpan (v4.3)(29) for the allotypes of HLA class II (HLA-DP, DQ, and DR). These algorithms are trained using experimental binding affinity and eluted ligand data to predict binding affinity. Binding prediction was performed on all possible amino acid 9mers to match the 9-residue binding groove for HLA class I genes and all possible amino acid 15mers for HLA class II molecules. The HLA class II molecules DPα1 and DPβ1, DQα1 and DQβ1, and DRα1 and DRβ1 can often combine to form the DP, DQ, or DR epitopes, respectively(30, 31). DRα1 has limited polymorphism, and we did not include DRβ3/4/5 in this analysis. Thus, all possible combined DP and DQ molecules were cross-referenced for their binding affinities. The predicted binding affinity scores between every possible peptide and every MHC molecule were ranked as a percentage by the algorithms based on comparison with a large set of naturally occurring peptides. We used the default rank threshold for strong binding peptides (top 0.5% for HLA class I genes, and top 1% for HLA class II genes). One of the rare alleles (DPB1*61:01N) is a null allele, i.e., does not present peptides, and thus pairs including the individual with this allele were excluded for the DP binding analysis.

(l) HLA haplotype resolution

The HLA allele calls from target sequence capture are unphased. We resolved HLA haplotypes by imputing closely matched HLA allele calls from the paired, phased SNP data across the 5Mb region encompassing the typed HLA genes. Although HLA imputation based on sparse genotype data is not accurate at detailed resolutions, we were still able to resolve the haplotypes by comparing imputed alleles to the known alleles from the HLA targeted sequencing. We trained HIBAG HLA imputation classifier models(32) on the Himba SNP data across the whole 5Mb region with paired HLA allele calls from the target capture (n=361 individuals). The models had "out of bag" accuracies of 97.96%, 96.97%, 98.60%, 98.75%, 98.73%, 98.75%, 97.66% across A, B, C, DPB1, DQA1, DQB1, and DRB1, respectively. The program does not impute DPA1 genotypes. We then extracted each phased SNP-based haplotype at the HLA region and created two pseudo-homozygous genotypes for each individual (i.e., ID1_hap1|ID1_hap1 and ID1_hap2|ID1_hap2). We ran these pseudo-homozygous genotypes through our curated HIBAG model to predict the HLA allele associated with each pseudo-homozygous genotype. We ensured that the predicted alleles across the two pseudo-homozygous genotypes matched one of the known target sequenced HLA allele calls for at least the first field. In this way, we filled in the allele calls across the two phased haplotypes. We were able to resolve 698/722 haplotypes with this method. The linkage disequilibrium patterns determined via asymmetric conditional linkage disequilibrium closely correspond to previously identified patterns(33) with high LD between HLA-B and C and also among HLA-DRB1, DQA1, and DQB1, demonstrating the accuracy of our haplotype resolution (Figure S1).

(m) Calculating IBD sharing rates

To calculate IBD sharing rates at each base pair genome-wide, we used a line sweep algorithm (Figure S2). In this method, the start of an IBD segment is marked +1, and the base pair directly following the last base pair in the IBD segment, is marked -1. A 'line' sweeps across the chromosome-length array summing the tally at the current base pair with the tally carried over from the previous base pair to generate IBD coverage scores for each base pair. In this way, a base pair located within a larger set of IBD fragment markings receives a larger coverage value during the line sweep step. The IBD sharing rate for each base pair is then computed by dividing the IBD coverage score by the total number of possible haplotypes in the dataset, the number of pairs multiplied by 2. This procedure reduces the complexity of the IBD sharing calculation problem, allowing for efficient computation of the IBD sharing rate at each base pair, genome-wide. IBD sharing rates were binned and averaged in 1000 base pair chunks.

(n) Statistical Models

We constructed a dyadic data model(34) to assess the effect of partnership type on IBD sharing (eq 2) and
on our HLA metrics (eq 3): HLA similarity score, mean log odds of homozygous offspring, peptide
binding groove divergence, and mean number of predicted pathogen peptides bound by hypothetical
offspring genotypes. Each partnership type (arranged, chosen) was encoded as a binary variable, and
random effects were included for each female and each male to account for individual-level variation.
IBD sharing was included as a covariate in equation 3 to determine whether differences in HLA metrics
between partnership types remained significant after conditioning on overall genetic relatedness.

The intercepts of both models represent a baseline derived from all 18,487 possible opposite sex pairings
that are not in known partnerships, are no closer than 3rd degree relatives, and also constrained to those
that have a permissible age differential based on the range observed in the known partnership data. This
baseline provides a background distribution for the expected similarity of partners in the population if
there is random mating, but with avoidance of close kin. We allowed 3rd degree relatives to exist in our
background distribution of partners as Himba individuals report a cultural preference for cousin marriages
in arranged unions(8). However, surprisingly, no individuals in actual arranged partnerships were related
at 3rd degree or closer, but 1 "love" marriage and 2 informal partnerships discovered through pedigree
analysis were found to be 3rd degree relatives.

This background distribution allows us to test whether each pair type deviates from the baseline
expectation of similarity while providing additional information to estimate the random effects. The
random effects of each female and male capture the unique contributions of each individual to their
dyadic HLA metrics/IBD sharing across all pairings (both real and random)(34). This approach allows us
to identify whether individuals' real partnerships (arranged and chosen) differ systematically in their
similarity scores when compared to their average similarity with potential random partners in the
population.

$(eq\ 2) \quad IBD = \mu + \beta_{arranged\ partnership} + \beta_{chosen\ partnership} + \Upsilon_{(1|female\ ID)} + Z_{(1|male\ ID)} + \varepsilon$

$(eq\ 3) \quad HLA\ metric = \mu + \beta_{IBD} * IBD + \beta_{arranged\ partnership} + \beta_{chosen\ partnership} +$
$\Upsilon_{(1|female\ ID)} + Z_{(1|male\ ID)} + \varepsilon$

Pairwise contrasts in the effect size estimates of the two different partnership types (arranged vs chosen)
were assessed by calculating the Z-statistic, defined as the contrast estimate divided by its standard error.
P-values for these Z-statistics were derived from a two-tailed normal distribution. Statistical significance
for the main effects and pairwise contrasts was determined using a p-value threshold of 0.05.

3. Results
To help contextualize our findings, we first compared HLA diversity in the Himba, which has not been
previously characterized, with other publicly available populations, curated to represent a diverse
world-wide sample(35) and all available sub-Saharan African groups(36). With HLA allele calls from 366
individuals, we assessed the diversity of HLA alleles based on their P group designation, that is, alleles
were grouped according to identical peptide binding domains to indicate differences in pathogen binding
function between alleles, across three HLA class I (A, B, C) and five HLA class II genes (DPA1, DPB1,
DQA1, DQB1, DRB1) (Figure S3). In order to make unbiased comparisons of allelic diversity based on
sample size, we subset our sample to the maximum set of unrelated Himba individuals (n=102, maximum
4th degree relations) (Figure S4). Despite endogamy and a recent population bottleneck(37), the allelic
diversity at each gene falls along the linear prediction based on sample size constructed from the publicly
available data, with expected or somewhat greater diversity than expected given the Himba sample size
compared to other African populations (Figure 1). We ensured genotype frequencies at each HLA gene

did not deviate from Hardy Weinberg equilibrium (Table S3) as this could indicate genotyping error or
population substructure(38).
302

303 **Figure 1:** *Allelic diversity in the Himba follows or somewhat exceeds linear predictions based on sample size from publicly*
304 *available cohorts.* Allelic diversity for the 8 genotyped HLA genes was measured in the set of 102 unrelated Himba. African
305 cohorts are labeled in orange, non-African cohorts in blue, and the Himba with a green diamond. Linear predictions of the
306 number of unique alleles based on sample size were constructed from the available datasets, excluding the Himba. Ribbons
307 indicate the 95% confidence interval for the linear prediction of the number of unique alleles based on sample size.
308

Next, we assessed whether there was an effect of partnership type (chosen or arranged) on overall genetic
relatedness. If mate choice is, in part, driven by minimizing overall genetic relatedness to reduce fitness
consequences associated with increased homozygosity in offspring(37, 39, 40), chosen partnerships could
be more genetically dissimilar overall than arranged partnerships. This could produce an associated, but
spurious pattern of HLA dissimilarity in chosen partnerships as compared to arranged partners and the
background distribution. Additionally, preferences for cross-cousin marriage and strong ties between
allied families could lead to greater sharing of IBD segments among arranged marriage partners than a
background distribution would predict, leading to associated, but spurious patterns of greater HLA
similarity in arranged partners as compared to chosen partners and the background distribution.
318

We assessed overall relatedness via the total number of centimorgans shared in genome-wide IBD
segments between the individuals in the partnership (see eq 2 in *Methods*). There was no effect of
arranged partnership or chosen partnership on genome-wide IBD sharing, relative to a background
population distribution created from random, minimally related (maximum 3rd degree relations),
heterosexual pairings with realistic age gaps (Figure 2A) ($p = 0.84$, 0.82, for arranged and chosen
partnership effects, respectively) nor a significant difference between the effects of arranged and chosen
partnerships in a pairwise contrast of effect size estimates ($p = 0.93$). 3rd degree relatives were allowed in
the background distribution to reflect the stated cultural preference for cousin marriages in arranged
unions(8).
328

Figure 2: *No effect of partnership types on genome-wide IBD sharing.* A) We consider a linear model where the intercept is the expected genome-wide IBD sharing based on a background distribution of unrelated female - male pairs from the population. We plot the effect size estimates of arranged partnerships and chosen partnerships on IBD sharing and find that the 95% confidence intervals of the effects contain zero, indicating there is no significant effect of arranged or chosen partnerships on IBD sharing. B) The distributions of the IBD sharing (cM) for arranged and chosen partnerships clearly overlap. Means of each distribution are noted with a dashed line.

(a) HLA similarity does not differ among partnership types

We next addressed our main question: do couples who have chosen to be together have lower HLA similarity than arranged partnerships? That is, is mate choice driven by minimizing relatedness at the HLA region rather than genome-wide relatedness? We assessed HLA genotype similarity in two ways, first by constructing a HLA similarity score, the number of shared alleles between a pair of individuals across all 8 genes (i.e. minimum score of 0, maximum score of 16) and also by computing the average log odds of homozygous offspring across all 8 genes (see *Methods*). These two metrics are highly correlated (Pearson r = 0.96), but average log odds of homozygous offspring provides a finer measurement (Figure S5). Using linear mixed models for dyadic data that correct for overall IBD sharing and account for the unique influence of each female and male to their similarity scores across all pairings(34) (see eq 3 in *Methods*), we found no effect of arranged or chosen partnership on overall HLA similarity (p = 0.7, 0.61, respectively) (Figure 3A) or on average log odds of HLA homozygosity in potential offspring (p = 0.79, 0.80, respectively) (Figure 3C). In a pairwise contrast, there were no differences between the effects of chosen and arranged partnerships on HLA similarity score (p = 0.56) or on average log odds of HLA homozygosity in potential offspring (p = 0.72). We also assessed whether there was a difference in HLA peptide binding groove protein divergence (exon 2 and 3 for HLA class I genes, exon 2 for HLA class 2 genes) by calculating the grantham distance between the amino acid sequences(24, 25) coded by partners' alleles. Again, we found no significant effects of arranged or chosen partnerships on mean amino acid sequence divergence between the molecules coded by partners' HLA genotypes ($p = 0.74, 0.70$, for arranged and chosen partnership effects, respectively, Figure S6) and no significant difference in their effects from a pairwise contrast ($p = 0.64$)

Figure 3: *No effect of partnership types on HLA similarity or average log odds of offspring HLA homozygosity.* We again consider linear models where the intercept is the expected outcome based on a background distribution of random, unrelated, opposite sex pairs from the population and control for genome-wide IBD sharing. We apply the model to the HLA similarity score (A) and to the average log odds of HLA homozygous offspring (C) across all 8 genes. We plot the effect size estimates of arranged partnerships and chosen partnerships on HLA similarity score (A) and average log odds of offspring homozygosity (C) and find the 95% confidence intervals contain zero, indicating no significant effect of arranged or chosen partnerships on HLA similarity or average log odds of offspring homozygosity. The distributions of HLA similarity scores (C) and average log odds of offspring HLA homozygosity (D) clearly overlap between arranged and chosen partnerships. Means of each distribution are noted with a dashed line.

(b) Pathogen binding repertoires of potential offspring do not differ among partnership types
Increased pathogen binding ability of offspring is theorized to be the alleged evolutionary driver of disassortative mating at the MHC region(41, 42)(42). Therefore, even if similarity scores, as assessed by allelic or protein similarity do not differ among pair types, individuals might seek out complementary HLA genotypes in mates that increase the pathogen binding repertoire of potential offspring. Using the same linear mixed model (*Methods* eq 3) as above, we looked for effects of arranged and chosen partnerships on the average computationally-predicted pathogen binding repertoires in hypothetical offspring (i.e., the average number of unique pathogen peptides predicted to be bound by each offspring HLA genotype). We used a curated list of 25 pathogens, taking a subset of those listed in a previous study of evolutionarily significant human pathogens(24) that have plausible transmission in the Himba, and included additional sexually transmitted pathogens known to be prevalent in the Himba(43–45) (personal correspondence with Dr. Ashley Hazel) (see Appendix Table 1). At each locus, the hypothetical offspring genotypes, in expected proportions, were cross-referenced to respective binding affinities to our list of antigenic peptides producing a mean number of unique pathogen peptides that strongly bind per HLA genotype. Many of the HLA class II DPA/B or DQA/B gene products can form both cis and trans heterodimers, comprising DP and DQ epitopes, respectively(30, 31). Thus, an individual heterozygous for both DPA1 and DPB1 may form 4 unique DPα1/DPβ1 molecules. We assessed the mean number of bound antigenic peptides for all possible offspring DP and DQ epitope combinations, again based on expected genotype proportions.

Across all 8 HLA genes tested, there was no effect of arranged or chosen partnerships on the mean number of pathogen peptides bound by offspring allotypes that differed from the background distribution of random partners (Figure S7, Table S4). Likewise, we found no differences in the effect estimates of arranged and chosen partnerships in pairwise contrasts (Table S5). HLA class I molecules generally bind peptides derived from intracellular pathogens (e.g. viruses) whereas HLA class II molecules generally bind those derived from extracellular pathogens (e.g. bacteria and parasites)(46). Thus, we also performed this analysis including only intracellular pathogens for HLA-A, B, and C genotypes and only extracellular pathogens for HLA-DP, DQ, and DR genotypes. We still found no significant effects of arranged or chosen partnerships in mean pathogen peptides bound and no significant differences of effects in pairwise contrasts (Figure S8, Tables S6-7).

(c) Extensive haplotype sharing could indicate recent episodes of fluctuating positive selection
We find no evidence that disassortative mating is driving maintenance of polymorphism at the HLA. Other forms of selection, besides sexually mediated balancing selection, could therefore play a larger role in maintaining polymorphism at this locus. We assessed haplotype sharing by visualizing the SNP-based haplotypes (from genotyping array) across the entire 5Mb region surrounding the 8 genotyped HLA genes. The SNP-based haplotypes along this much larger 5Mb region were grouped by their HLA allele haplotypes (typed via short-read sequencing) (Figure S9). Although the 7 HLA genes used to create allele-based haplotypes comprise only ~1% of the base pairs in this 5Mb region, we see virtually identical SNP haplotypes across unrelated individuals (maximum relatedness of 4th degree) who share frequent HLA allele-based haplotypes (n=92 haplotypes with >4 occurrences in the 102 unrelated individuals) (Figure S9-10). This finding indicates that the entire 5Mb HLA region was inherited without recombination in these unrelated individuals, consistent with recent positive selection on these haplotypes(47, 48).

Unlike our previous study of sub-Saharan African populations, where in 6 of the 7 populations the majority of haplotypes were observed only once(36), 70% of haplotypes were observed at least twice in our set of unrelated individuals, with the most frequent haplotype (*A*30:02 - C*03:04 - B*15:10 - DRB1*03:01 - DQA1*05:01 - DQB1*02:01 - DPB1*01:01*) observed 23 times in the unrelated set. Nineteen of the 29 haplotypes occurring more than once in the unrelated set of individuals were observed in at least one of the 7 sub-Saharan African populations from our previous study (Figure S11). We assessed whether there was a relationship between haplotype frequency and number of unique peptides computationally predicted to be bound by each haplotype(49), but found no significant relationship ($p = 0.91$, Figure S12) and likewise found no significant relationship with number of peptides bound when we analyzed each pathogen independently (Figure S13).

Consistent with the SNP-based haplotype visualization, we found increased IBD sharing at the HLA region relative to genome-wide levels. This pattern implies recent (within 200 generations) positive selection as opposed to overdominance (i.e., heterozygote advantage)(47, 48). The mean rate of IBD sharing across the HLA region between random pairs of unrelated individuals from the background distribution used in our pair analyses (n=352 individuals, n=18,487 pairs, related at a maximum of 3rd degree) is 5.3%, 1.97 standard deviations above the genome-wide median of 3.1%. The rate of IBD sharing across the HLA region in the subset of unrelated individuals from our allele frequency analyses (related at a maximum of 4th degree) who have common HLA-based haplotypes (n=71 individuals, haplotypes shown in figure S9) is even higher at 10.4%, 6.4 standard deviations above the genome-wide median of 2.8%.

Figure 4: *Elevated IBD sharing rates at the HLA region is consistent with recent, fluctuating positive selection.* Panels A and B depict IBD sharing rates across chromosome 6 (rates were averaged within 1000 base pair bins). The dashed line depicts the genome-wide median IBD sharing rate, excluding chromosome 6. Panel A depicts IBD sharing rates for the pairs that make up the background distribution for the partnership analyses (random heterosexual pairings, with relevant age gaps, related at no closer than 3rd degree; n=352 individuals). Panel B depicts the IBD sharing rates for the unrelated individuals included in the allele frequency estimations (related at no closer than 4th degree) with the common haplotypes shown in Figure S9 (n=71 individuals). The HLA region is designated in grey. The pink shaded region depicts +2 and -2 standard deviations from the genome-wide (excluding chromosome 6) median IBD sharing rate, 3.1% and 2.8% for panels A and B, respectively.

4. Discussion

This study adds an important new data point to the growing literature on HLA and mate choice. Our study is free from the limitations of cosmopolitan datasets, which may exhibit spurious patterns of HLA similarity in realized partners due to preferences for ethnically, and therefore, relatively genetically similar partners. Cultural norms like arranged marriage and cross-cousin marriage preferences are predicted to promote similarity across the HLA, while tolerance of extramarital partnerships and "love match" marriages could enhance the possibility of finding an HLA-dissimilar partner. Therefore, the unique socioecology of Himba marriage patterns sets up a strong test of the dissimilarity hypothesis.

Surprisingly, we find that chosen partnerships are neither less similar at their HLA than random pairs constructed from the population background distribution, nor do they differ in their HLA similarity from arranged partnerships. We also find no evidence that chosen partners are selected to directly maximize an offspring pathogen binding repertoire. Unlike studies of Hutterites(4) and Mormons(5) that do show evidence for greater HLA dissimilarity in married pairs, background genetic diversity in the Himba is likely high enough that any unrelated individual is sufficiently dissimilar at the HLA and there may be no fitness benefit to maximizing dissimilarity. Furthermore, it has been suggested that offspring may not benefit from maximal diversity at the HLA; T cells that bind self peptide-MHC complexes too strongly undergo pruning in the thymus. Possessing many different MHC alleles could deplete the T cell repertoire required for appropriate immune response(50, 51). This process of negative selection for extremely autoreactive T-cells has been hypothesized to inhibit the expansion of the MHC genomic region to include additional copy numbers.(52) Supporting this optimality hypothesis, decreased parasite load in intermediate versus maximally diverse MHC individuals has been observed in voles(51) and stickleback fish(53) and our data suggest that this may also be relevant for humans.

We find extensive IBD sharing at the HLA region consistent with recent, fluctuating positive selection which may contribute to polymorphism maintenance at this region, rather than heterozygote advantage(47, 48). Despite association with many diseases(54) and consistently showing up as a target of natural selection(49, 52), the selective forces shaping HLA diversity are not well understood(52). We suggest that innate drive for disassortative mating based on HLA genotype is unlikely to be a major evolutionary force in maintaining diversity among humans. Future work should focus on assessing the contribution of other selective regimes (e.g., negative frequency dependent, fluctuating selection, heterozygote advantage, divergent allele advantage, etc.) to the maintenance of polymorphism at the HLA region.

We conclude that, given the limited benefits of HLA dissimilarity, assortative mating for sociocultural traits may be a more important driver of mate choice for groups like the Himba than maximizing genetic diversity. Our previous work has shown that resource scarcity affects women's partner preferences,(55, 56) and that relationship success is correlated with similarity in mate value(57). Concurrent partnerships, rather than increasing the chance of finding a partner with a dissimilar HLA genotype, may be more important for garnering resource security for women, and distributing reproductive opportunities for men who are otherwise limited by polygyny and a later age at marriage.(9, 58)

Ethics

Ethical approval for this study was granted by the University of California, Los Angelse (IRB-10-000238), the State University of New York, Stony Brook (IRB-636415-12), and was approved by the Namibian Ministry of Home Affairs and the University of Namibia Office of Academic Affairs and Research. Chief Basekama Ngombe provided permission to work in the community and local approval of the study. Community leaders were actively involved in discussions regarding what genetic data could be used for, who would have access to it, and whether there was a for-profit element involved (there was not). Individual informed consent, and for minors' parental assent, was obtained orally from all participants, given low rates of literacy in the community. Care was taken to protect participants' privacy, for example via a double-blind procedure for DNA collection(59). These data were collected as part of the longitudinal Kunene Rural Health and Demography Project, which has been working in the community since 2010.

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Author Contributions

BS, PN, and BMH conceived of the study. GLM conducted analyses with help from KH and CB. GLM and BS wrote the manuscript with input from PN, SP, and BMH. KK, GH, NFP, TDJF, and NP generated HLA sequence data under the supervision of PN. Sample and ethnographic data collection were conducted by BS and SP. BMH and BS supervised the research.

Use of AI-assistance

528 AI was not used in the preparation of this article.
529
530 **Data and code availability:** Genotype and HLA allele call data for the individuals included in this study
531 have been deposited at dbGap as phs001995.v3.p1. All original code and data used to conduct analyses
532 are deposited in the following github repository https://github.com/gillianmeeks/Himba_HLA_mating and
533 on dryad https://doi.org/10.5061/dryad.02v6wwqg2. Partnership type assignments are not available due to
534 the sensitive nature of the data.
535
536 **Competing Interests**
537 The authors declare no competing interests.
538
539 **Supplemental information**
540 SI. Supplementary Figures S1–S13 and Supplementary Tables S1–S7
541 Appendix Table 1. Spreadsheet listing pathogen peptide NCBI accessions, related to methods.
542

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