**The anti-cancer drug trametinib suppresses angiotensin-induced cardiac remodelling in mice but is detrimental to function**

Short title:  Trametinib inhibits cardiac remodelling causing dysfunction

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Early identification of cardiotoxicity from cancer drugs is crucial for patient welfare! This study shows novel, early changes on echocardiograms linked to toxic effects of melanoma drugs in a model with high blood pressure. □ #Echocardiogram #CardioOncology #HeartHealth

**Abstract**

**Background.**  The anti-cancer MEK inhibitor trametinib (alone or with the RAF inhibitor dabrafenib) causes cardiac dysfunction or heart failure in some patients.  Our hypothesis is that cardiotoxicity is exacerbated by an underlying co-morbidity such as hypertension causing early cardiac dysfunction detectable on echocardiograms.  **Objectives.**  The objectives were to assess the effects of trametinib on cardiac function in a mouse model of hypertension-induced cardiac hypertrophy and determine if this was moderated by dabrafenib.  **Methods.**  Male mice were treated with vehicle, trametinib or dabrafenib/trametinib in the absence/presence of angiotensin II (AngII; 0.8 mg/kg/d to increase blood pressure over 7 d).  Hearts were imaged using echocardiography. **Results.**  Inhibitors alone had a limited effect on mouse hearts over 28 d.  Trametinib or trametinib/dabrafenib inhibited cardiac hypertrophy induced by AngII over 7 d, reducing left ventricular (LV) wall thickness and mass.  AngII did not significantly affect cardiac function, but the inhibitors caused significant functional deterioration.  Segmental analysis revealed variation of contraction around the LV, with selective effects of AngII and trametinib or dabrafenib/trametinib in basal/mid-regional segments.  Frame-by-frame analysis of radial (not longitudinal) displacement of the LV endocardial wall demonstrated variation between consecutive cardiac cycles that enabled a high degree of classification according to treatment.  **Conclusions.**  Trametinib inhibits AngII-induced cardiac hypertrophy in mice but is detrimental to cardiac function, responses that are not moderated by dabrafenib.  AngII and MEK/RAF inhibition have regional effects around the LV with greater effects on radial displacement in basal/mid-regional segments.  Assessment of such changes may facilitate early identification of developing cardiotoxicity.

**Key words:**  Hypertension, MEK, RAF, extracellular signal-regulated kinases, echocardiography, cancer drugs

**Clinical perspectives.**

**Competencies in Medical knowledge.** The anti-cancer drugs trametinib and dabrafenib are cardiotoxic in some patients. These drugs had limited effects themselves on mouse hearts, but suppressed cardiac hypertrophy in a mouse model of hypertension whilst proving detrimental to cardiac function. Segments around the left ventricle were differentially affected by hypertension and inhibitors,with variation between consecutive cardiac cycles.

**Translational outlook.** Further research is needed to establish the relationship between hypertension and cardiotoxicity of trametinib in patients, and determine if regional effects in the ventricle wall can predict cardiac dysfunction. If so, a diagnostic algorithm may be a useful tool in identifying developing cardiotoxicity.

**Summary Figure. Effects of dabrafenib/trametinib on the cardiac response to angiotensin II in mice.** Inhibiting ERK1/2 suppresses cardiac fibrosis and cardiomyocyte hypertrophy induced by angiotensin II. This results in a decline in cardiac function, with particular effects on radial wall movement and in different segments of the left ventricle.

Extracellular signal-regulated kinases 1/2 (ERK1/2) drive cell proliferation and promote cancer.[1] The cascade [RAF family kinases (ARAF/BRAF/RAF1)→MEK1/2→ERK1/2] integrates signals from cell surface receptors, whilst downstream signals stimulate cell division and are cytoprotective. Mutations in BRAF are found in ~4.5% of all tumours,[2] and small molecule inhibitors (e.g. dabrafenib, encorafenib) are used clinically to treat cancers with a BRAF mutation.[3] These inhibitors do not differentiate between mutated BRAF and wild-type RAF, so may affect non-cancerous tissues. RAF inhibitors can, paradoxically, activate the pathway under certain circumstances and tumours can become resistant to the drugs. To avoid these issues, patients are often treated with a combination of RAF and MEK inhibitors (e.g. dabrafenib/trametinib).[4] Adverse effects of the drugs include cardiotoxicity. For example, trametinib (alone or with dabrafenib) promotes hypertension in up to 29% of patients, 12% develop left ventricular (LV) dysfunction and 1-5% develop heart failure.[5] Cardiotoxicity of RAF/MEK inhibitors appears reversible in most patients following withdrawal of treatment, but long-term effects on the heart, particularly if treatment is resumed, are unclear.

Heart failure affects >64 million people worldwide, and results from various underlying causes.[6] Because the contractile cells of the adult mammalian heart (cardiomyocytes) have limited regenerative potential,[7] pathophysiological stresses that alter cardiomyocyte contractility and/or result in cardiomyocyte death can severely compromise cardiac function. Cardiac pathology associated with, for example, hypertension, is well studied. Cardiomyocytes initially hypertrophy (i.e. increase in size and contractile apparatus) to accommodate the increase in workload, but this adaptive hypertrophy may degenerate: cardiomyocytes become dysfunctional and die, whilst fibrosis increases.[8] The pathological changes associated with cardiotoxicity of anti-cancer drugs remain largely unknown.

Cardiomyocyte ERK1/2 signalling is tightly regulated during cardiac development. Germline mutations with moderate ERK1/2 activation cause severe congenital cardiac defects ("RASopathies"), and trametinib reduces cardiac hypertrophy and hypercontractility

in at least some of these patients.[9]  ERK1/2 promote hypertrophy of postnatal terminally-differentiated mammalian cardiomyocytes, but also stimulate cardiac fibrosis.[3]  RAF/MEK inhibitors could, therefore, be beneficial or detrimental for the heart.  Of the RAF inhibitors, dabrafenib inhibits ERK1/2 and reduces cardiac fibrosis in a mouse model of hypertension induced by angiotensin II (AngII), whereas encorafenib activates ERK1/2 and promotes adaptive hypertrophy in mouse hearts *in vivo.*[10,11]  In contrast, trametinib inhibits ERK1/2 phosphorylation in perfused hearts[10] and can be highly cardiotoxic in mice, causing heart failure over 2-4 months.[12]  Here, we show that trametinib is not necessarily cardiotoxic over the short term when administered alone or with dabrafenib, but inhibits cardiac hypertrophy induced by AngII and is detrimental to cardiac function.

## METHODS

**ETHICS STATEMENT.**  Male C57Bl/6J mice were housed at the BioResource Unit/Facility at University of Reading (studies of inhibitors only) or St. George's University of London (studies of inhibitors with AngII).  Females were not used because young female mice are resistant to AngII-induced hypertension.[13]  Procedures were performed in accordance with UK regulations, the European Parliament Directive 2010/63/EU for animal experiments, local institutional animal care committee procedures and the U.K. Animals (Scientific Procedures) Act 1986.  Studies were conducted under Project Licence P8BAB0744.

**MOUSE STUDIES, ECHOCARDIOGRAPHY, DATA ANALYSIS.**  C57Bl/6J male mice were treated with 1 mg/kg/d trametinib or trametinib with 3 mg/kg/d dabrafenib dissolved in vehicle or vehicle alone, using osmotic minipumps.  For studies with AngII, mice received a second minipump containing acidified PBS or AngII (0.8 mg/kg/d) in acidified PBS.  Echocardiograms were collected and analysed using VevoLab/VevoStrain software (Visualsonics).  Echocardiograms from previous studies[14,15] were also used.  Further

analysis used a statistical approach with machine-learning.  Full experimental details are in the Supplemental File.

**STATISTICAL ANALYSIS.**  GraphPad Prism 10.4.1 was used for analysis of VevoLab/VevoStrain data.  Outliers were excluded using a Grubb's outlier test.  Statistical testing used two-tailed two-way paired ANOVA with Holm-Sidak's multiple comparison post-test or Kruskal-Wallis test with Dunn's multiple comparisons as indicated.  Specific p values are provided with significance levels of $p<0.05$ in bold type.  Python was used for analysis of endocardial wall displacement (see **Supplemental Methods**).

**RESULTS**

**TRAMETINIB OR DABRAFENIB/TRAMETINIB INHIBITED CARDIAC HYPERTROPHY INDUCED BY ANGII IN MICE BUT WERE DETRIMENTAL TO CARDIAC FUNCTION.**  We first compared the effects of trametinib (1 mg/kg/d) alone or with dabrafenib (3 mg/kg/d) on the hearts of healthy male C57Bl/6J mice, monitoring cardiac function and dimensions by echocardiography (7 and 28 d) (**Figure 1**).  There were no significant differences between the treatments in body weights at 7 or 28 d, or in heart weight:body weight ratios at 28 d (**Supplemental Table S1**).  At 7 d, there were no differences in cardiac function/dimensions between the conditions, and at 28 d, the only difference was a small but significant increase in the thickness of the interventricular septum with trametinib or dabrafenib/trametinib (**Figure 1B-D; Supplemental Tables S2-S3**).  There was a small, significant increase in end diastolic LV mass between 7 and 28 d in vehicle-treated mice, with decreased global longitudinal strain (GLS), stroke volume and cardiac output, possibly as heart size grew to match body size.  This was not detected in mice treated with trametinib or dabrafenib/trametinib, and the inhibitors may have affected normal cardiac growth. Histologically, hearts from mice treated with trametinib or dabrafenib/trametinib were

indistinguishable from vehicle-treated mice (**Supplemental Figure S1**). Overall, trametinib or dabrafenib/trametinib did not have a substantial effect on mouse hearts over 28 d.

The additional stress of trametinib or dabrafenib/trametinib on a hypertensive heart may exacerbate disease progression, so their effects on cardiac hypertrophy were assessed in a model of hypertension induced by 0.8 mg/kg/d AngII (**Figure 2A**), a dose that increases blood pressure over 7-14 days.[16] Analysis of M-mode images indicated that AngII (7 d) increased diastolic and systolic LV anterior and posterior wall thickness, whilst LV internal diameter was decreased (**Figure 2B-C; Supplemental Table S4**). Trametinib or dabrafenib/trametinib inhibited the increase in wall thickness by AngII, although diastolic wall thickness remained greater than in vehicle-treated mice. The decrease in LV internal diameter was not significantly inhibited. Speckle-tracking of long axis B-mode images indicated that AngII increased LV mass, an effect that was fully attenuated by trametinib or dabrafenib/trametinib (**Figure 3A-B; Supplemental Table S5**). This contrasts with the M-mode data and potentially reflects a change in geometry of the heart. In support of this, the diastolic length of the LV chamber was reduced in mice treated with AngII and trametinib or dabrafenib/trametinib (**Figure 3C**). Consistent with the effects on LV wall thickness and predicted mass, histological staining showed that dabrafenib/trametinib partially inhibited the increase in cardiomyocyte cross-sectional area and interstitial (though not perivascular) fibrosis induced by AngII.[17]

Although remodelling of the ventricular wall resulting from AngII treatment was suppressed by trametinib or dabrafenib/trametinib, the hearts were not normal. The decrease in LV internal diameter and end diastolic volume with AngII was not restored (**Figures 2C** and **3B**) suggesting the hearts may be hypercontracted. With respect to cardiac function, ejection fraction and fractional shortening were not reduced by AngII alone, but there was some decline in mice treated with trametinib and AngII (**Figure 3D**). Stroke volume was significantly decreased with AngII, with a greater decrease in the presence of trametinib or dabrafenib/trametinib, presumably contributing to the significant decrease in cardiac output. GLS and global circumferential strain (GCS) were unaffected by AngII alone,

but significantly reduced with AngII in the presence of inhibitors (**Figure 3E**). Thus, trametinib or dabrafenib/trametinib are detrimental to cardiac function in the presence of AngII. However, there were no significant differences between trametinib alone or dabrafenib with trametinib.

**SEGMENTAL ANALYSIS DEMONSTRATES DIFFERENTIAL EFFECTS OF ANGII AROUND THE LV WITH SELECTIVE EFFECTS OF THE INHIBITORS.** To determine if there are regional effects on the LV, we applied time-to-peak analysis to long axis B-mode images, dividing the LV walls into 6 segments each with 9 datapoints (**Figure 4A**). Averaging fewer datapoints increased the variation. To allow for this, the dataset was extended to include mice from previous studies treated with vehicle and AngII in the same way as this study.[14,15] Radial and longitudinal strain of the endocardium was measured perpendicular to and in line with, respectively, the endocardial wall. Variables assessed were time-to-peak and peak values for displacement, velocity, strain and strain rate.

In vehicle-treated mice, there was variation between adjacent and opposing segments, with more significant differences in peak than time-to-peak values (**Figure 4B-C**; **Supplemental Tables S6-S7**). The greatest variation was in longitudinal displacement and velocity, with the apex having little longitudinal movement. The pattern for radial movement differed, with greater movement of the posterior than anterior wall. AngII did not substantially affect variation between segments, but peak measurements in segment 1 were generally decreased (**Figures 4 and 5; Supplemental Figure S2**; **Supplemental Tables S8-S9**). Segregated analysis showed similar effects of trametinib and dabrafenib/trametinib on the AngII response with no significant differences (**Supplemental Figure S3**), so the data were combined (n=18). The inhibitors with AngII significantly reduced radial peak values relative to AngII alone, having a dominant effect on the anterior wall (segments 4-6), but did not normalise the decrease in peak values in segment 1 resulting from AngII treatment (**Figures 4 and 5; Supplemental Tables S10-S12**). Thus, trametinib without/with dabrafenib, has regional effects on the LV wall, reducing radial wall movement in an

asymmetric manner, effects that are potentially detrimental.

**HEARTBEAT VARIATION.** Since respiration and heartbeat are interconnected[18] and mice move whilst breathing (rendering speckle-tracking difficult), speckle-tracking was performed across 2 cardiac cycles measured immediately after a breath was taken, as contraction commenced. To identify variation between cycles, we conducted frame-by-frame analysis of the LV endocardial wall, focusing on peak displacement since this was affected most by AngII and the inhibitors (**Figure 4**). We first used a statistical approach (see **Supplemental Methods**) which showed no difference in average (for all segments) longitudinal displacement measurements between the cardiac cycles (**Figure 6A**). AngII reduced the maximum peak displacement in each cycle compared with vehicle only, an effect that was not modified by the inhibitors, and the two cycles remained synchronous. AngII did not affect the average peak radial displacement compared with vehicle-treated hearts, but the timing of the cardiac cycles diverged, starting during relaxation of the first cycle, with a phase shift becoming apparent during contraction in the second cycle (**Figure 6B**). There was substantial variation between segments, with the greatest phase shift induced by AngII in segments 5 and 4 (**Figure 6C**). Moreover, peak radial displacement was enhanced by AngII in segments 4/5, but reduced in segment 1. The inhibitors eliminated the phase shift induced by AngII, but also suppressed the peak radial displacement in all segments, consistent with the reduction in GLS and GCS (**Figure 3E**).

To validate the statistical analysis, we used machine-learning for further interpretation of the dynamics between the two cardiac cycles, using random forest classifiers (see **Supplemental Methods**). Results showed superior classification with the second cycle compared with the first (mean F1 = 0.687 vs 0.557). Segment ranking identified segment 5 as most informative (F1 = 0.680), then segments 1 and 4 (**Figure 6D**). F1 scores for individual and combinations of segments did not successfully classify all three conditions (**Figure 6E**). Pairwise comparisons demonstrated the second cardiac cycle (not the first) was largely successful in differentiating between AngII and vehicle, whereas both cycles

were useful in differentiating between the other groups. Furthermore, although the second cycle was dominant, data for both cycles together could increase performance for both segments. These data underscore the diagnostic value of spatial and temporal dissection in cardiac motion analysis.

**DISCUSSION.**

Many new, successful cancer therapies are small molecule inhibitors of protein kinases, targeting signalling pathways that promote cell growth and cytoprotection.[19] Since these pathways are essential for the heart to survive pathophysiological stresses, the emergence of inhibitor-associated cardiotoxic effects is unsurprising.[20] This study focused on dabrafenib and trametinib,[3] RAF and MEK inhibitors. Because of associated cardiotoxicity, it is now recommended that patients treated with dabrafenib/trametinib are monitored by echocardiography.[5] With greater numbers of cancer patients treated with RAF/MEK inhibitors, it is increasingly important to identify who may develop cardiac dysfunction, when that dysfunction develops and how it manifests. Our study showed that, whilst trametinib (alone or with dabrafenib) had relatively minor effects on healthy mouse hearts over the short term, it had much greater consequence in developing hypertension, suppressing the adaptive hypertrophic response and compromising cardiac function.

**CARDIOTOXICITY OF TRAMETINIB.** We identified little effect of trametinib on mouse hearts over 28 d (**Figure 1**), contrasting with previous work in which it induced severe cardiac dysfunction within 4-8 weeks, and high mortality over 2-4 months.[12] Beck et al.[12] used C57Bl/6 mice, starting at a similar age as our mice, theoretically with the same dose of trametinib (1 mg/kg/d). However, the substrain was not specified and, for example, C57Bl/6N mice can develop more severe cardiac phenotypes than C57Bl/6J mice.[21] A mixed sex protocol was also used which could confound interpretation. The study duration

was longer, but (given the severity of their response) we would expect significant cardiac dysfunction by 4 weeks. A key difference is the dosage and dosing regime. In Beck et al.,[12] trametinib was provided in food and, based on their information, the mice could have received an average of 1.5 mg/kg/d. With variation between individual mice there was, potentially, dosage variation. Since drug delivery from feeding is over a shorter time period, there is also likely to be higher peak dosage. Clearly, Beck et al. demonstrated the cardiotoxic potential of trametinib and provided substantial mechanistic information. The importance of our study is that we had limited effects of trametinib or dabrafenib/trametinib alone on cardiac function, but they compromised cardiac function in a model of hypertension.

**CARDIOTOXICITY OF TRAMETINIB IN HYPERTENSION.** Our hypothesis is that ERK1/2 signalling is not necessary for a healthy heart but is required for cardiac adaptation to pathophysiological stresses. Consequently, inhibiting the pathway is more damaging in patients (or mice) with an underlying co-morbidity such as hypertension, accounting for cardiotoxicity in only a subset of patients. The concept is embedded in recent guidelines supporting assessment of patients for cardiovascular risk factors prior to commencement of certain anti-cancer therapies.[5,22,23] Our data match this view and, whilst we detected little effect of the inhibitors alone, trametinib or dabrafenib/trametinib moderated the developing adaptive hypertrophic response to AngII in mice. The increase in LV wall thickness and mass were inhibited, consistent with a role for ERK1/2 signalling in promoting cardiomyocyte hypertrophy[3] (**Figures 2 and 3**). However, the inhibitors did not restore cardiac function. Whereas ejection fraction and fractional shortening were maintained in the presence of AngII at 7 d, inhibitor treatment caused some decline in these values, accompanied by significant decreases in stroke volume, cardiac output, GLS and GCS (**Figure 3D-E**). Dabrafenib/trametinib reduced interstitial fibrosis and cardiomyocyte hypertrophy resulting from AngII treatment.[17] Reduced fibrosis should improve cardiac function. However, cardiomyocyte hypertrophy is necessary to accommodate the increased workload, so the

reducing cardiomyocyte hypertrophy will most likely compromise cardiac function. ERK1/2 directly modulate ion fluxes in cardiomyocytes and these will also be affected. For example, ERK1/2 regulates the sodium proton exchanger (NHE1) which impacts on intracellular pH and, therefore, contraction.[24] ERK1/2 also influence intercellular communication via connexin 43 phosphorylation in gap junctions.[25] Further research is needed to understand the implications, but effects of trametinib on contractility via reversible phosphorylation is compatible with the observation that most patients with cardiac dysfunction from trametinib improve following cessation of the therapy.[5]

Dabrafenib inhibits cardiac ERK1/2 signalling and the hypertrophic response to AngII but, in contrast to trametinib, restores cardiac function.[10] The reason for the difference is unclear since both reduce fibrosis and cardiomyocyte hypertrophy. However, RAF kinases have substrates other than MEK1/2 (e.g. MST2[26]) and dabrafenib inhibits other kinases (e.g. RIPK3[27]), so it could have ancillary beneficial effects on cardiac function. There is probably also greater inhibition of ERK1/2 with MEK inhibitors, with more severe consequences, because of additional inputs at the level of MEK1/2.[28] Whatever the reason, there were no significant differences between trametinib and dabrafenib/trametinib on the effects of AngII on the heart, and the detrimental effects of trametinib appeared to dominate.

**ECHOCARDIOGRAPHY FOR DIAGNOSIS OF CARDIOTOXICITY.** Echocardiography is the most widely-used imaging modality for assessment of cardiac function in patients treated with cardiotoxic anti-cancer drugs.[29] Cardiotoxicity is generally based on ejection fraction (a decrease of >10% to below 50%) and/or GLS (a decline of >15%).[22] We wanted to determine if other changes in the LV may be additional indicators of developing dysfunction. Our B-mode images of mouse hearts were suitable for segmental analysis of different regions of the LV but, because of the limited number of points tracked for each segment, there was substantially more variation. By including echocardiograms for vehicle and AngII-treated mice from previous studies, we generated a powerful dataset suitable to assess variation around the LV wall, and to compare different cardiac cycles. The data

demonstrated clear variation in contractile response between segments in vehicle-treated mice, selective effects of AngII particularly on segment 1 at the base of the posterior wall and selective effects of trametinib or dabrafenib/trametinib particularly on peak radial displacement of the anterior wall (**Figures 4** and **5**). Frame-by-frame analysis of radial displacement produced further insight showing that AngII promoted a phase shift in the second cardiac cycle after a breath that was normalised by the inhibitors, even though the inhibitors curtailed peak radial displacement, potentially accounting for the reduction in cardiac function (**Figure 6**).

In this study, we used machine learning only to support the statistical analysis. However, it illustrates the power of combinatorial multi-segmented analysis for the effects of the inhibitors. A similar multi-faceted approach could be used for patient echocardiograms, and extending the analysis using feature engineering may enable the development of diagnostic algorithms to identify abnormalities associated with early stages of cardiotoxicity. Notably, the inhibitors seemed to have much greater effects on LV basal segments, and on radial rather than longitudinal displacement. This may be because apical segments are more anatomically constrained, and basal segments have greater movement in normal circumstances. Additionally, radial displacements are generally greater than longitudinal displacements. Small differences in function or movement will be more readily detected in these areas that move more freely. This may be an important consideration for early detection of LV dysfunction in the clinical setting. Rather than using a global measure of longitudinal movement (i.e. GLS), perhaps measures of radial displacement in specific segments may be more sensitive in detecting LV dysfunction.

**LIMITATIONS.** A major limitation of this study is the lack of blood pressure data. Since trametinib causes hypertension in some patients,[5] it could have increased blood pressure in the mice. Trametinib or dabrafenib/trametinib did not affect cardiomyocyte size or cardiac fibrosis (**Supplemental Figure S1**), suggesting that blood pressure was not increased, but blood pressure assessments should form part of any future studies. We only used young

male mice, whilst cancers treated with trametinib affect men and women. However, hypertension affects many more men at younger ages than women.[30] Moreover, the focus of the study was cardiotoxicity of trametinib in hypertension and, since young female mice are resistant to AngII-induced hypertension,[13] a separate study is needed. Hypertension generally develops later in life,[30] and further studies with older mice and with stable hypertension would be useful. Finally, hypertension is one factor that may influence the risk of cardiotoxicity. Other cardiovascular risk factors (e.g. obesity) or genetic variation (e.g. titin mutations[31]) are likely to impact cardiotoxicity risk and further research in these areas would be beneficial.

**CONCLUSIONS.** Trametinib alone or with dabrafenib, had limited effects on unstressed hearts but significantly affected AngII hypertensive hearts, reducing cardiac remodelling but proving detrimental to cardiac function (**Summary Figure**). Segmental analysis of echocardiograms showed that AngII and trametinib have differential effects around the LV. Furthermore, analysis of sequential cardiac cycles (following a breath) identified more abnormalities in the second cycle. These data identify options for monitoring cardiotoxicity apart from standard measures (ejection fraction, GLS). In the future, an algorithm based on such changes may be developed for early diagnosis of cardiotoxicities of RAF/MEK inhibitors and other anti-cancer drugs.

**REFERENCES**

1.  Martin-Vega A, Cobb MH. Navigating the ERK1/2 MAPK cascade. Biomolecules 2023;13:1555.doi:10.3390/biom13101555

2.  Hanrahan AJ, Chen Z, Rosen N, Solit DB. BRAF - a tumour-agnostic drug target with lineage-specific dependencies. Nat Rev Clin Oncol 2024;21:224-247.10.1038/s41571-023-00852-0

3.  Clerk A, Amadi, S.U., Smith, S.J., Sugden, P.H. Inhibiting the extracellular signal-regulated kinase 1/2 (ERK1/2) cascade in cancer and the heart: for better or worse, in sickness and health? IJDDP 2024;3:100006.doi.org/10.53941/ijddp.2024.100006

4.  Imani S, Roozitalab G, Emadi M, Moradi A, Behzadi P, Jabbarzadeh Kaboli P. The evolution of BRAF-targeted therapies in melanoma: overcoming hurdles and unleashing novel strategies. Front Oncol 2024;14:1504142.10.3389/fonc.2024.1504142

5.  Senechal I, Andres MS, Tong J et al. Risk stratification, screening and treatment of BRAF/MEK inhibitors-associated cardiotoxicity. Curr Oncol Rep 2024;26:1431-1441.10.1007/s11912-024-01599-2

6.  Savarese G, Becher PM, Lund LH, Seferovic P, Rosano GMC, Coats A. Global burden of heart failure: A comprehensive and updated review of epidemiology. Cardiovasc Res 2023;118:3272-3287.10.1093/cvr/cvac013

7.  Soonpaa MH, Zebrowski DC, Platt C, Rosenzweig A, Engel FB, Field LJ. Cardiomyocyte cell-cycle activity during preadolescence. Cell 2015;163:781-2.10.1016/j.cell.2015.10.037

8.  Bazgir F, Nau J, Nakhaei-Rad S et al. The microenvironment of the pathogenesis of cardiac hypertrophy. Cells 2023;12.10.3390/cells12131780

9.  De Brouchoven I, Lorand J, Bofferding L, Sorlin A, Van Damme A, Danhaive O. Trametinib as a targeted treatment in cardiac and lymphatic presentations of Noonan syndrome. Front Pediatr 2025;13:1475143.10.3389/fped.2025.1475143

10. Meijles DN, Cull JJ, Cooper STE et al. The anti-cancer drug dabrafenib is not cardiotoxic and inhibits cardiac remodelling and fibrosis in a murine model of hypertension. Clin Sci (Lond) 2021;135:1631-1647.10.1042/CS20210192

11. Clerk A, Meijles DN, Hardyman MA et al. Cardiomyocyte BRAF and type 1 RAF inhibitors promote cardiomyocyte and cardiac hypertrophy in mice in vivo. Biochem J 2022;479:401-424.10.1042/BCJ20210615

12. Beck TC, Arhontoulis DC, Morningstar JE et al. Cellular and molecular mechanisms

of MEK1 inhibitor-induced cardiotoxicity. JACC CardioOncol 2022;4:535-548.10.1016/j.jaccao.2022.07.009

13. Brooks HL, Pollow DP, Hoyer PB. The VCD mouse model of menopause and perimenopause for the study of sex differences in cardiovascular disease and the metabolic syndrome. Physiology (Bethesda) 2016;31:250-7.10.1152/physiol.00057.2014

14. Marshall JJ, Cull JJ, Alharbi HO et al. PKN2 deficiency leads both to prenatal congenital cardiomyopathy and defective angiotensin II stress responses. Biochem J 2022;479:1467-1486.10.1042/BCJ20220281

15. Cull JJ, Cooper STE, Alharbi HO et al. Striatin plays a major role in angiotensin II-induced cardiomyocyte and cardiac hypertrophy in mice in vivo. Clin Sci (Lond) 2024;138:573-597.10.1042/CS20240496

16. Zimmerman MC, Lazartigues E, Sharma RV, Davisson RL. Hypertension caused by angiotensin II infusion involves increased superoxide production in the central nervous system. Circ Res 2004;95:210-6.10.1161/01.RES.0000135483.12297.e4

17. Cooper STE. RAF kinases and cardiac endothelial cells: characterising RAF 'paradox breaker' inhibitor PLX8394 in hypertension. PhD Thesis Chapter 5, 2022:103-118.

18. Fisher JP, Zera T, Paton JFR. Respiratory-cardiovascular interactions. Handb Clin Neurol 2022;188:279-308.10.1016/B978-0-323-91534-2.00006-0

19. Roskoski R, Jr. Properties of FDA-approved small molecule protein kinase inhibitors: A 2024 update. Pharmacol Res 2024;200:107059.10.1016/j.phrs.2024.107059

20. Shyam Sunder S, Sharma UC, Pokharel S. Adverse effects of tyrosine kinase inhibitors in cancer therapy: pathophysiology, mechanisms and clinical management. Signal Transduct Target Ther 2023;8:262.10.1038/s41392-023-01469-6

21. Garcia-Menendez L, Karamanlidis G, Kolwicz S, Tian R. Substrain specific response to cardiac pressure overload in C57BL/6 mice. Am J Physiol Heart Circ Physiol 2013;305:H397-402.10.1152/ajpheart.00088.2013

22. Lyon AR, Lopez-Fernandez T, Couch LS et al. 2022 ESC Guidelines on cardio-oncology developed in collaboration with the European Hematology Association (EHA), the European Society for Therapeutic Radiology and Oncology (ESTRO) and the International Cardio-Oncology Society (IC-OS). Eur Heart J 2022;43:4229-4361.10.1093/eurheartj/ehac244

23. Tong J, Senechal I, Ramalingam S, Lyon AR. Risk assessment prior to cardiotoxic anticancer therapies in 7 steps. Br J Hosp Med (Lond) 2025;86:1-21.10.12968/hmed.2024.0632

24. Fliegel L. Structural and Functional Changes in the Na(+)/H(+) Exchanger Isoform 1, Induced by Erk1/2 Phosphorylation. Int J Mol Sci 2019;20:2378.10.3390/ijms20102378

25. Zhang M, Wang ZZ, Chen NH. Connexin 43 phosphorylation: implications in multiple diseases. Molecules 2023;28.10.3390/molecules28134914

26. O'Neill E, Rushworth L, Baccarini M, Kolch W. Role of the kinase MST2 in suppression of apoptosis by the proto-oncogene product Raf-1. Science 2004;306:2267-2270.10.1126/science.1103233

27. Li JX, Feng JM, Wang Y et al. The B-Raf(V600E) inhibitor dabrafenib selectively inhibits RIP3 and alleviates acetaminophen-induced liver injury. Cell DeathDis 2014;5:e1278.doi:10.1038/cddis.2014.241

28. Clerk A, Kemp TJ, Harrison JG, Pham FH, Sugden PH. Integration of protein kinase signaling pathways in cardiac myocytes: signaling to and from the extracellular signal- regulated kinases. AdvEnzyme Regul 2004;44:233-248

29. Tamaki N, Manabe O, Hirata K. Cardiovascular imaging in cardio-oncology. Jpn J Radiol 2024;42:1372-1380.10.1007/s11604-024-01636-x

30. Connelly PJ, Currie G, Delles C. Sex differences in the prevalence, outcomes and management of hypertension. Curr Hypertens Rep 2022;24:185-192.10.1007/s11906-022-01183-8

31. Garcia-Pavia P, Kim Y, Restrepo-Cordoba MA et al. Genetic variants associated with

cancer therapy-induced cardiomyopathy. Circulation 2019;140:31-41

**Figure legends**

**Figure 1. Trametinib or dabrafenib/trametinib have limited effects on the hearts of mice over 28 d. A,** Schematic: male C57Bl/6J mice (9 weeks) received baseline (BL) echocardiograms (Echo) with additional echocardiogams after osmotic minipumps (MP) were implanted to deliver vehicle (Veh), 1 mg/kg/d trametinib (T), or 3 mg/kg/d dabrafenib with trametinib (DT). **B**, Representative M-mode traces (28 d) from the short axis (SAX) and B-mode images from the SAX or long axis (LAX). **C**, Analysis of B-mode SAX or M-mode images for interventricular septum (IVS) and diastolic dimensions of left ventricle (LV) anterior wall (AW), posterior wall (PW) or internal diameter (ID). **D**, Analysis of B-mode LAX images for end diastolic LV mass (EDLVM), end diastolic volume (EDV), global longitudinal and circumferential strain (GLS, GCS) and measures of cardiac function. Data are individual values with means ± SEM. Statistical analysis used two-way repeated measures ANOVA with Holm-Sidak's post-test. Significant differences are in bold.

**Figure 2. Trametinib or dabrafenib/trametinib inhibit the increase in left ventricular wall thickness induced by AngII in mice. A,** Schematic: male C57Bl/6J mice (9 weeks) received a baseline (BL) echocardiogram (Echo) with a second taken 7 d after osmotic minipumps (MP) were implanted to deliver vehicle (Veh), 0.8 mg/kg/d AngII (A) or AngII with 1 mg/kg/d trametinib (Tram, T/A), or 3 mg/kg/d dabrafenib with trametinib (DabTram, DT/A). **B**, Representative short axis (SAX) images (7 d). **C**, Analysis of B-mode SAX or M-mode images for the interventricular septum (IVS) and left ventricle (LV) anterior wall (AW), posterior wall (PW) or internal diameter (ID). Data are individual values with means ± SEM. Statistical analysis used one-way ANOVA with Holm-Sidak's post-test. Significant differences are in bold.

**Figure 3. Trametinib or trametinib/dabrafenib inhibit the increase in left ventricular mass induced by AngII in mice, but are detrimental to cardiac function.**

Male C57Bl/6J mice were treated as in **Figure 2A**. **A**, Representative long axis (LAX) images (7 d). **B, D-E**, Speckle-tracking of B-mode LAX and SAX images for end diastolic LV mass (EDLVM) or volume (EDV) (**B**), measures of cardiac function (**D**), and endocardial global longitudinal and circumferential strain (GLS, GCS) (**E**). **C**, LV chamber internal length from the mitral valve to the apex. Data are individual values with means ± SEM. Statistical analysis used one-way ANOVA with Holm-Sidak's post-test. Significant differences are in bold type.

**Figure 4. Segmental analysis demonstrates variation in function around the left ventricle with selective/regional effects of AngII and trametinib.** Long axis B-mode images were used for segmental analysis using speckle-tracking, measuring longitudinal (Long) and radial (Rad) endocardial movement: displacement (D), strain (S), strain rate (R), velocity (V). Mice were treated with vehicle (n=38), AngII (n=41) or AngII with trametinib or dabrafenib/trametinib (n=18). **A**, Positions of the segments. **B**, Heatmap for statistical analysis of segmental data showing p values comparing adjacent and opposing segments. Analysis used Kruskal-Wallis tests with Dunn's multiple comparison post-test. **C**, Heatmaps for mean values for each parameter studied. Data for each segment were normalised to the mean value for all the segments for each parameter. **D**, Heatmap shows p values for comparisons between mice treated with vehicle and AngII (left), vehicle and Inhibitors/AngII (centre) plus AngII and Inhibitors/AngII (right). Analysis used the data from C, applying Kruskal-Wallis tests with Dunn's multiple comparison post-test.

**Figure 5. Segmental analysis of cardiac function in mice treated with AngII without/with trametinib: peak values.** Long axis B-mode images were used for segmental analysis, measuring longitudinal (Long) and radial endocardial movement. Data were generated for each of 6 segments from mice treated with vehicle (n=38), 0.8 mg/kg/d AngII (n=41), AngII with 1 mg/kg/d trametinib or trametinib and 3 mg/kg/d dabrafenib (DabTram/AngII, n=18). Profiles for peak measurements for longitudinal (**A**) and radial

movement (**B**) are shown.  Individual values are provided with means ± 95% CI.

**Figure 6.  Frame-by-frame analysis of radial displacement across consecutive cardiac cycles identifies greater differentiating potential between conditions in the second cycle.  A-C,** Data for longitudinal and radial displacement of the LV endocardial wall were collated for each frame across consecutive cardiac cycles (C1 and C2; frames 1-54) for the average of all segments (**A-B**) and individual segments **(C)**.  Data are mean displacement (solid lines) with 95% CI (shaded areas).  Vehicle (n=38): blue line/shade; AngII (n=41): yellow line/grey shade; AngII with trametinib or dabrafenib/trametinib (Inhibs, n=18): red line/shade. Statistically significant differences in each frame (p<0.05) are marked with vertical dashed lines (Kruskal-Wallis tests with Dunn's post hoc correction).  **D,** Machine learning used random forest models trained across all segments.  Evaluation used stratified 5-fold cross-validation (random seed = 42), and F1 scores were averaged per configuration. Segment ranking identified segment 5 as the most informative (F1 = 0.680).  **E**, Heatmaps for F1 scores for individual and combinations of segments three-way (all conditions) or pairwise analysis for each or both cardiac cycle(s).

**Summary Figure.  Effects of dabrafenib/trametinib on the cardiac response to angiotensin II in mice.**  Inhibiting ERK1/2 suppresses cardiac fibrosis and cardiomyocyte hypertrophy induced by angiotensin II.  This results in a decline in cardiac function, with particular effects on radial wall movement and in different segments of the left ventricle.

Cooper_Duvenage Summary Figure

**Angiotensin II**
(hypertension)

Dabrafenib —| BRAF RAF1

**Trametinib** —| MKK1/2

ERK1/2

Cardiac
remodelling
(Fibrosis,
Cardiomyocyte
hypertrophy)

**Cardiac
function**

LV Radial displacement

Heartbeat 1    Heartbeat 2

Vehicle
AngII
AngII/Inhibitors

**Longitudinal (Long)**

Anterior wall

5    4

6                **Radial (Rad)**

Apex                Mitral valve
region

2    1

Posterior wall

Segment 5

Segment 4

Segment 2

Segment 1