Disruption of The Spinal Cord-Gut Axis Alters Gut Microbial Dynamics and Carbohydrate Cross-feeding

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Abstract

The spinal cord, a nexus for brain-body crosstalk, controls gut physiology and microbial homeostasis, but the underlying mechanisms remain unclear. Using genome-resolved longitudinal metagenomics in male and female C57BL/6 mice before and up to 6 months after disrupting the spinal cord-gut axis, we reconstructed over 6,500 microbial draft genomes. This "Mouse B6 Gut Catalog" improved or doubled species- and strain-level representation in other published catalogs. Impaired spinal cord-gut crosstalk induced persistent, sex-, time- and lesion-specific alterations in community composition, marked by a consistent loss of *Lactobacillus johnsonii*. Feeding this key bacterium to mice with a clinically relevant spinal cord injury improved host health. Genome-resolved, community-contextualized metabolic profiling revealed that shifts in carbohydrate-mediated microbe-microbe interactions explain the reduction of *L. johnsonii*. These findings identify carbohydrate metabolism as a keystone mechanism shaping gut microbiota and emphasize

that mammalian health and gut ecosystem function depend on a functional spinal cord-gut axis. Additionally, these data improve murine microbiome catalogs and demonstrate that metagenome-informed microbial interventions can improve host health and likely mitigate long-term dysbiosis.

INTRODUCTION

Mammals harbor a "second genome" composed of a complex microbial community residing in, on, and around their bodies. This microbial ecosystem, particularly prominent in the gut, assists the host in disease protection and influences the efficacy of drug therapies against illnesses like cancer[1]. These microbial beneficial roles depend on host control systems that monitor and regulate the microbiome[2]. One such system is the bi-directional signaling pathway connecting gut and brain functions[3,4]. Within this microbiota-gut-brain axis, the vagus (parasympathetic) nerve is often implicated as a solitary controller of host-microbiota mutualism[5,6]. However, the spinal sympathetic (fight-or-flight) nervous system also controls the gut, although its effects on the gut microbiota are rarely considered.

The spinal cord is the "hub" integrating nerve impulses before they influence the brain and body. Everyday stimuli, such as eating or bowel and bladder filling, activate spinal sympathetic nerves, affecting gut motility, mucus secretion, intestinal blood flow, and immune function[7]. Each of these gut physiological features can influence gut microbiota[8,9]. Even organ-specific signals directly transmitted to the brain via the vagus nerve, will directly[10,11] and indirectly[12,13] elicit spinal sympathetic reflexes that influence peripheral organs, including the gut[14]. Thus, to accurately assess gut microbiota stability in health and disease, a more holistic understanding of the "microbiota-gut-brain-spinal cord axis" is crucial.

Gut dysbiosis—imbalances in microbiome composition—is associated with the onset and severity of several health conditions, including central nervous system (CNS) diseases[15]. Accordingly, modifying gut microbiota is a focal point for therapeutic intervention. Despite wide use, gene-marker-based approaches like 16S rRNA gene sequencing[16–18] face limitations, including primer bias, reliance on reference genomes, and restricted functional representation of niche-defining traits, which evolve faster than core traits and drive ecological interactions[19–21]. Genome-resolved metagenomics overcomes these limitations, enhancing taxonomic resolution and constructing genomic potential of ecosystem-specific taxa[22,23]. Genome-resolved metagenomics has been applied to the spinal cord-gut axis only once, in our prior proof-of-concept murine study limited by single timepoint and sex sampling[24]. Here, we longitudinally sample both sexes to create and curate a large, high-resolution murine microbiome reference genome catalog, then use it to evaluate the spinal cord control over the gut ecosystem, establishing a foundational understanding of how microbiota influence mammalian health. These advances deconstruct conventional thinking about gut-brain communication, showing that disruption to the spinal cord-gut axis causes sex- and time-dependent microbiota changes, altering microbial carbohydrate metabolism with implications for host health.

RESULTS AND DISCUSSION

Expanding murine microbial reference genomes

We first sought to build a more comprehensive murine microbial reference genomes catalog using an expanded experimental design, which is summarized as follows (details under "*Animals and spinal cord injury"* in **Methods**). First, microbiomes were compared from three groups of mice: (i) control mice receiving a sham-lesion to the spinal cord (Laminectomy or Lam; surgical removal of vertebral body but no spinal lesion), (ii) mice that received a complete lesion in low-thoracic (T10) spinal cord, and (iii) mice that received a complete lesion in high-thoracic (T4) spinal cord. Complete lesions at these spinal levels partially (T10) or completely (T4) abolish supraspinal control over spinal sympathetic neurons that control the gut[25,26], effectively titrating the magnitude of "spinal cord-gut axis" dysfunction. Second, to enhance the longitudinal resolution of spinal cord-dependent changes (i.e., acute and chronic) on gut microbiota dynamics, samples were collected across six time points for each mouse. Third, to improve translational relevance and statistical power, we included female and male mice, and for each sex and spinal lesion level, we doubled the number of mice sampled at each time point.

Overall, these efforts resulted in 333 bulk metagenomes (294 fecal and 39 cecal) collected from 29 male and 30 female C57BL/6 mice at six time points spanning six months (**Fig. 1A**) – a considerable increase on our prior sampling[24] of five female samples per experimental group (Lam, T4 and T10 spinal lesions) at a single time point. The overall physiological state of each mouse in the three groups was assessed after six months using a standardized scale of neurological recovery for mice (i.e., the Basso Mouse Scale (BMS) for Locomotion[27]; hereafter referred to as "BMS score" (**Fig. S1**). This expanded experimental design was further augmented by enhancing metagenomic workflow at several key steps, including cross-assembly per mouse, ensemble binning, and bin refinement (see **Methods** and **Fig. S1A**).

The increased sampling and updated analytics drastically expanded microbiota representation. In total, recovered genomic representation increased with 6,635 bacterial metagenome-assembled genomes or MAGs (>50% completeness and <10% contamination), that when combined with 112 previously generated MAGs[24] led to 263 de-replicated MAGs (at >95% average nucleotide identity, or ANI) spanning 7 phyla and 31 families (**Fig. 1B**), which were used for all ecological and metabolic analyses. Of the 263 de-replicated MAGs, 238 were either above-medium (n = 76; >70% complete, <10% contamination) or high (n = 162; >90% complete, <5% contamination) quality (see **Methods**) – more than doubling MAG-based genomic representation from our prior pilot study[24] (**Fig. 1C**). Taxonomically, this larger MAG collection captured additional representative taxa above the species level (**Fig. 1B**). Furthermore, 20.5% (n=54/263) of newly identified MAGs are likely novel species-level taxa as they lacked 95% Average

106 Nucleotide Identity (ANI)-level representation, even against the ~62k bacterial species clusters
107 from the many environments represented in the Genome Taxonomy Database (GTDB[28]; Release
108 207). Ecologically, this updated MAG collection significantly improved the representation of
109 murine gut microbiota as the percentage of sequencing reads that align to de-replicated MAGs
110 increased from a mean of 55%[26] to ~80% (**Fig. 1D**). Further, this improved MAG-based read-
111 mapping finding was consistent with assembly-independent, read-based profiling of phylogenetic
112 marker genes (see **Methods**) where ~81% of the microbial diversity was recovered in the updated
113 MAGs (**Fig. S1B**).

114 To assess the taxonomic and ecological value of this new genomic resource, which we
115 named the "Mouse B6 Gut Catalog" (MB6GC), we compared MB6GC against other murine-
116 relevant microbiome catalogs. First, at a finer strain-level taxonomic resolution, MB6GC includes
117 480 medium- and high-quality MAGs (>50% completeness and <10% contamination) after de-
118 replication at 99% ANI. Comparing these MAGs against a similar strain-resolved MAG database
119 for CBA/J mice[29] showed minimal overlap, with over 400 MAGs specific to C57BL/6 mice (**Fig.**
120 **1E**; see **Methods**). Second, we compared species-level (95%ANI) MAGs in MB6GC to several
121 large murine gut metagenomic catalogs[30–32]. Despite these other catalogs integrating 3-8x more
122 metagenomic data from 36 to 74 projects across multiple mouse strains, MB6GC added ~6-10%
123 new MAGs, or improved the quality (i.e., completeness and contamination) of MAGs found in the
124 other catalogs (**Fig. 1F**; see **Methods**). Collectively, across all catalogs, 53 MAGs from MB6GC
125 were either novel (n=15) or were the best representative genomes of their species (n=38) (**Fig.**
126 **1G**). Overall, MB6GC is a new resource to support microbiome studies in the C57BL/6 mouse
127 strain, which is the most widely used in medical research[33]. Also, where strain-level (>99% ANI)
128 taxonomic resolution is needed to understand the (patho)physiological significance of
129 microbiome-host interactions[21,34], mouse strain-specific MAG resources should be used.

130 Disrupting the spinal cord-gut axis causes persistent gut dysbiosis

131 We next asked – '*Does the spinal cord regulate gut microbiome composition?*'. After data
132 curation (details under "*Data curation*" in **Methods** and **Fig. 1A**, **Fig. S1C-D**), abundance-based
133 beta-diversity comparisons (principal-coordinate analysis [PCoA] of Bray-Curtis dissimilarities)
134 were made using the MAG data. These comparisons revealed sex-specific microbial communities
135 in mice before surgery (**Fig. 2A**, **Fig. S2A**). Since sex-based differences persisted after surgery,
136 regardless of the experimental group (**Fig. 2B**, **Fig. S2B**), we analyzed the microbiome of male
137 and female cohorts separately. Notably, in both male and female mice, partial (T10 lesion) or
138 complete (T4 lesion) disruption of the spinal cord-gut axis significantly changed gut microbiome
139 composition (**Fig. 2C-D)** relative to mice with intact spinal cords (**Fig. 2E-F)**. Importantly, these
140 changes appear to be permanent, with marked shifts in community composition (beta-diversity)
141 evident for at least 6 months in all mice with spinal cord-gut axis dysfunction (**Fig. 3A**). Peak

142 changes in gut microbial community composition varied by sex, occurring either three or five
143 weeks after a partial or complete break of the spinal cord-gut axis in male or female mice,
144 respectively (**Fig. 3A**). Like beta-diversity, disrupting spinal cord control over the gut increased
145 alpha-diversity (estimated by Shannon's index) in both sexes (**Fig. 3B**, **Fig. S2C**) relative to mice
146 with intact spinal cords. An increase in alpha diversity is associated with increased gut transit
147 time[8], a common comorbidity of spinal cord dysfunction[35]. We confirmed this association in an
148 independent mouse cohort using a red carmine dye assay[36]. The data show that compared to mice
149 with intact spinal cords, total intestinal transit time increased 17-38% for at least 3 weeks in mice
150 with a spinal lesion (**Fig. 3C**).

151 Overall, instead of static, snapshot comparisons of microbiomes from different mice after
152 surgery, which is the norm in most microbiome studies, here, within-mouse longitudinal
153 comparisons (i.e., before and periodically after injury) (**Fig. 3**) reveal that spinal cord dysfunction
154 causes persistent and dynamic changes in gut microbiome composition. Moreover, our study
155 confirms that sex is a critical biological variable when analyzing the microbiome (see also[37,38]).
156 Thus, an intact spinal cord is required for regulating gut microbiome composition, regardless of
157 sex. Notably, prior animal studies have mostly interrogated the effects of spinal cord dysfunction
158 on gut microbiome composition in female mice only (reviewed in[16]). Sex-specific differences in
159 the microbiome are linked to development of, or susceptibility to, autoimmune disease[39–41],
160 gastrointestinal disease[42,43], metabolic disease[44,45], neuropsychiatric and neurodegenerative
161 disease[46,47]. Collectively, our analyses extend existing microbiome datasets to include longitudinal
162 spinal cord-dependent control over the gut ecosystem in both sexes, providing both a valuable new
163 experimental resource and a more comprehensive understanding of CNS regulation of the
164 microbiome.

165 Microbes that persist after disrupting the spinal cord-gut axis include taxa 166 previously linked to the host physiological state

167 Next, to determine which taxa respond to spinal cord-gut axis perturbation, microbial
168 relative abundances were compared in mice with or without an intact spinal cord (see **Methods**
169 for details on statistical analysis). Significant spinal cord-dependent effects are shown at an
170 aggregated phylum-level (**Fig. 4A**, **Fig. S3A**), and an individual MAG/species-level (**Fig. 4B-C**,
171 **Fig. S3B**). At the phylum level, a break in the spinal cord-gut axis decreased Bacteroidota and
172 Proteobacteria, with a concurrent increase in Firmicutes_A in males and females (**Fig. 4A, Fig.**
173 **S3A**). Whereas the ratio between the phyla Firmicutes and Bacteroidetes was also reported to
174 increase in our earlier work in mice with spinal cord injury[24,48], others have reported that this same
175 ratio decreases[49,50]. This discrepancy may be attributed to improvements in or differences between
176 databases, as the GTDB uses genomic data that now distinguish the Firmicutes phyla into three

separate phyla: Firmicutes, Firmicutes_A, and Firmicutes_B (see "*GTDB vs NCBI Taxonomy*" in **Methods**). In addition, other lower-resolution techniques (i.e., 16S rRNA gene amplicon sequencing) may not be able to distinguish between these three classically lumped lineages. Here, using genome-resolved metagenomics, Firmicutes_A was the only phylum that changed among the three.

At the species level, many differentially abundant MAGs were found in male (n=68; **Fig. 4B, Fig. S3B**) and female (n=14; **Fig. 4C**) mice. The magnitude of change in relative abundance of several discrete microbial species (**Fig. 4B-C**, **Fig S3B**) was differentially affected by spinal lesion level, an observation that was also shown for species alpha diversity (**Fig. 3B**). Temporal changes in MAG abundances also varied by spinal lesion level and sex (**Fig. S4**). However, time-resolved analysis of rare taxa shows persistent spinal lesion-dependent changes in *Lactobacillus johnsonii* (**Fig. 5A**) in both sexes. Notably, changes in *Weissella cibaria* (a significantly changing taxon in[24]) were time-dependent (**Fig. S4G**), highlighting the importance of well-controlled temporal analyses to discern changes in the gut microbiome that may only be sporadically evident. Indeed, static, time-delimited sampling at any single time point would likely under- or overestimate change(s) for a given microbe or the effect that sex or the spinal cord have on these changes.

Although abundant MAGs that decrease post-lesion in males and females were mostly Muribaculaceae family members (3 out of 4), most increasing MAGs (14 of 21) belong to the Lachnospiraceae family (**Table S1)**. A decrease in Muribaculaceae abundance with a concomitant increase in Lachnospiraceae has been linked to metabolic diseases, such as obesity, type 2 diabetes, insulin resistance, and non-alcoholic steatohepatitis[51,52]. Other rare taxa that are reduced in mice without normal spinal cord control of the gut, including *L. johnsonii* and Turicibacter sp002311155, may also play critical roles in influencing mammalian physiology and health[53–55]. Because *L. johnsonii*'s MAG was essentially depleted after lesioning the spinal cord, regardless of sex or spinal lesion level (**Fig. 5A**), and because this microbe was previously identified as a potential beneficial microbe capable of regulating various murine metabolic pathways[24], we directly tested the effects of reintroducing *L. johnsonii* into the gut of mice with spinal cord lesions.

Spinal cord injury-dependent weight loss is prevented by post-injury feeding with *Lactobacillus johnsonii*

We isolated *L. johnsonii* by culturing fecal samples from naive C57BL/6 mice on a selective growth medium for *Lactobacilli* (see **Methods**). Female C57BL/6 mice received complete mid-thoracic spinal lesions (injured) or were left uninjured (naive). Starting at 1 day post-lesion, naive and injured mice were fed 1 x 10^8 cfu of *L. johnsonii* or vehicle (water) daily via oral gavage. At 9 days post-lesion, fecal samples from all mice were plated then cfu/ml was

213 calculated. A significant decrease in *L. johnsonii* was noted in injured mice fed water only (**Fig.**
214 **5B**), confirming metagenomic data from this and our prior study (**Fig. 5A**; and[24]). Conversely,
215 feeding *L. johnsonii* significantly increased its abundance in lesioned mice, which was visually
216 evident in cultures, confirmed by quantifying cfu/ml and by 16S sequencing of fecal samples
217 obtained from mice in each cohort (**Fig. 5B-C**). Importantly, rapid weight loss, a consistent
218 phenomenon indicative of a changing metabolic state in mice with spinal injuries, was significantly
219 reduced when mice were fed *L. johnsonii* (**Fig. 5D**). Since feeding *L. johnsonii* did not affect
220 weight in naive mice (not shown), we conclude that these results reflect a gut-microbiome-
221 dependent "rescue" of metabolic perturbations caused by a break in the spinal cord-gut axis. For
222 example, spinal cord injury causes rapid muscle wasting[56], and *Lactobacillus* can prevent
223 sarcopenia by modulating inflammation, oxidative stress, and skeletal muscle metabolism[57].
224 Although further research is needed to determine the precise mechanisms underlying the rescue
225 effects of *L. johnsonii*, this bacterium can improve intestinal barrier function, modulate systemic
226 immunity, and prevent metabolic disease[53,58–61].

227 Altered carbohydrate metabolism in spinal cord-dependent dysbiotic gut 228 selects against rare beneficial taxa (e.g. *L. johnsonii*)

229 Data in **Fig. 5** demonstrate the physiologic importance of *L. johnsonii*, especially when the
230 spinal cord-gut axis is perturbed, as is common in most types of neurological disease. But, why
231 would a sustained reduction in *L. johnsonii* occur in these disease states? To answer this question,
232 we used the entire MB6GC to infer the complex metabolic interactions that occur across the gut
233 microbial community, focusing on the MAGs (n=13) that were most abundant and changing after
234 a break in the spinal cord-gut axis (**Fig. 4B-C**).

235 All MB6GC MAGs were annotated using DRAM (Distilled and Refined Annotation of
236 Metabolism[62]), a tool designed to functionally annotate microbial genes and categorize them across
237 various expert-curated metabolic pathways. Interestingly, the four most abundant MAGs in non-
238 lesioned mice (collectively ~28% and ~45% median relative abundance in female and male mice,
239 respectively) belong to the Bacteroidales order (**Table S1**), namely CAG-485 sp002362485, CAG-
240 873 sp910577315, Duncaniella sp910578515, and *B. thetaiotaomicron*. Their proportionally large
241 relative abundance can be attributed to these bacteria encoding a complete electron transport chain,
242 including the high-oxygen affinity cytochrome *bd* (**Fig. S5**), allowing them to survive and
243 potentially thrive in the low oxygen environment of the gut[63]. Additionally, under homeostatic
244 conditions, species from the Bacteroidales order can perform polysaccharide to oligo- and
245 monosaccharide conversions that dictate cross-feeding relationships broadly across a gut microbial
246 community[34,51,64]. The products of these carbohydrate conversions are primary energy sources for
247 many gut bacteria, and carbohydrate metabolism by gut microbes is essential for producing various

248 beneficial metabolites (e.g. short-chain fatty acids; SCFAs) that support host health[34]. However,
249 as mechanistic genome-resolved approaches are only just emerging in studies of mammalian gut
250 microbiome, community-contextualized microbial carbohydrate cross-feeding after dysbiosis has
251 not been well-resolved in any system, especially involving *L. johnsonii*. Here, we examine state-
252 dependent changes in systems used by bacteria to degrade and take up carbohydrates, namely
253 glycoside hydrolases (GHs), sugar transporters, and polysaccharide utilization loci (PUL) systems.

254 Based on the 13 most abundant changing MAGs, the capacity to degrade complex
255 polysaccharides remained unchanged after disrupting the spinal cord-gut axis. This conclusion was
256 drawn from analyzing the number and types of GHs, enzymes responsible for breaking down
257 complex carbohydrates into simple sugars for microbial energy. When GHs were grouped into 14
258 substrate-based categories (*sensu*[62]), 11 of 13 MAGs encoded diverse GHs that act on all 14
259 carbohydrate substrates (**Fig. S6A**). Of these 11 MAGs, three decreased and eight increased in
260 mice with spinal cord lesions (**Fig. S6B**). Unlike these abundant taxa, which can degrade these 14
261 carbohydrates, *L. johnsonii* scarcely encodes GHs (**Fig. S6A**; see also *Turicibacter* sp002311155,
262 another rare beneficial taxon that is decreased in mice with spinal lesions[53–55]). These data suggest
263 that rare beneficial taxa depend on abundant community members for polysaccharide degradation
264 and cross-feeding. Indeed, MAGs of these rare taxa encode many diverse types of sugar
265 transporters that would enable them to scavenge simple sugars (**Fig. S7**). This suggests that, under
266 homeostasis, the final products of polysaccharide degradation carried out by abundant community
267 members (e.g., Muribaculaceae) might be used by rare, beneficial taxa. After a significant
268 perturbation, as Muribaculaceae decrease, the increase in other abundant bacteria is unlikely to
269 support cross-feeding of rare taxa, a potential mechanism causing prolonged dysbiosis. For
270 example, Lachnospiraceae family members encode GHs with broad polysaccharide substrate
271 specificity (**Fig. S6**), but unlike Muribaculaceae, Lachnospiraceae also encode other numerous and
272 diverse simple sugar transporters (**Fig. S7**), so they are expected to consume the simple sugar
273 products of polysaccharide degradation. Conversely, the four most abundant MAGs changing post-
274 lesion (all Bacteroidales) scarcely encode these sugar transporters (**Fig. S7**), and instead
275 exclusively encode polysaccharide utilization loci (PULs), with multiple characteristic homologs
276 of the starch utilization system (**Fig. S8**). Bacteroidales use these systems to sense, bind, degrade,
277 and import polysaccharides and oligosaccharides[65]. Of these four Bacteroidales MAGs, one
278 increases after spinal lesioning (*B. thetaiotaomicron*) and three less-characterized Muribaculaceae
279 decrease (**Fig. 4 and Fig. S4**), with no evidence for such systems in any other taxa, consistent with
280 previous reports[65].

281 PUL systems exhibit selfish and altruistic behaviors in complex trophic networks like in
282 the gut. Some Bacteriodales species encode "selfish" PUL systems that rapidly import broken-
283 down oligosaccharides into the periplasm for further breakdown, conferring no direct benefit to

284 neighboring species, while other Bacteroidales encode "leaky" PULs that release partial
285 breakdown products (PBPs) for the benefit of the whole community[66]. We found evidence for a
286 selfish PUL system (utilizing GH99 and GH76; reviewed in[66]) encoded only by *B.*
287 *thetaiotaomicron*, potentially explaining the relative fitness of this taxon in the dysbiotic gut of
288 mice with spinal lesions, where transporter-rich Lachnospiraceae compete for and scavenge
289 monosaccharides. Other Bacteroidales species, notably those of the Muribaculaceae family,
290 exhibit an "altruistic" behavior by selectively packaging GHs into outer membrane vesicles that
291 are released into the extracellular environment, enabling the production of free saccharides for the
292 greater gut community[66–68]. More broadly, the Muribaculaceae family has been credited with
293 probiotic roles, including a protective oxygen-scavenging role[51] – consistent with the cytochrome
294 *bd* results above - see **Fig. S5**) and cross-feeding with other microbial species (reviewed in[51]).
295 Indeed, we find that some Muribaculaceae taxa (e.g., CAG-485 sp002362485) encode diverse
296 GHs, potentially boosting the fitness of other Bacteroidales (e.g, CAG-873 sp910577315) and rare
297 beneficial taxa (e.g., *L. jonhsonii*) that encode scarce GHs and instead depend on PBPs or
298 monosaccharide products, respectively.

299 A conceptual model that explains gut ecosystem function, both before and after a break in
300 the spinal cord-gut axis, can be developed from our data (**Fig. 6**). Fundamental to this model is our
301 data-driven hypothesis that reassembly of the microbial community post-injury, as orchestrated by
302 the most abundant and dynamic taxa, results in a unique carbohydrate metabolic fingerprint that
303 disfavors rare beneficial taxa and cooperative trophic interactions. First, under homeostatic
304 conditions, some abundant Bacteroidales members break down polysaccharides into oligo- and
305 mono-saccharides, "public goods" that other bacteroidales and beneficial microbes, such as *L.*
306 *johnsonii* and *Turicibacter* sp002311155, take up for energy using their sugar transporters. Second,
307 after a break in the spinal cord-gut axis, the niche of Bacteroidales (Muribaculaceae members and
308 *B. thetaiotaomicron*) is partially filled by other polysaccharide degraders, notably Lachnospiraceae
309 family members. While Muribaculaceae MAGs significantly decrease, *B. thetaiotaomicron* seem
310 to thrive, possibly because their PUL systems do not discard resources for other members and
311 instead can quickly import oligosaccharides back to the periplasm. Unlike Bacteroidales, other
312 polysaccharide degraders, mostly Lachnospiraceae family members, also encode large numbers of
313 diverse sugar transporters and thus can fully utilize carbohydrates and consume the
314 monosaccharides they produce, essentially starving rare beneficial taxa (e.g., *L. johnsonii*) and
315 reducing their abundance in the community (**Fig. 6**).

316 This model of gut ecosystem function provides a framework for predicting changes in gut
317 microbe-host interactions beyond disruptions in the spinal cord-gut axis. For example, a study
318 using paired metagenomics and metabolomics in insulin-resistant individuals (characterized by
319 elevated sugar levels in blood, urine, and feces) found that Lachnospiraceae taxa positively

correlate with monosaccharide concentrations in fecal matter, whereas Bacteroidales taxa show negative correlations with the same metabolites[69]. Our model suggests that high monosaccharide concentrations favor transporter-rich taxa like Lachnospiraceae while disfavoring transporter-scarce taxa like Bacteroidales.

Conclusions

This report establishes the spinal cord as an integral but largely overlooked part of the "CNS-gut-microbiota axis". By enhancing study design, sample diversity, and analytical methods, we introduce new resources (e.g., MB6GC) and microbial targets for better understanding disease mechanisms and potential interventions. For instance, feeding *L. johnsonii* helps restore host metabolic benefits during recovery from spinal cord injury (**Fig. 5**). A grand challenge in microbiome science is deciphering ecosystem-relevant changes in microbial functions without prior knowledge bias. Applying ecological theory and eco-systems biology concepts can organize significant taxa (~70 MAGs here) into key sub-communities controlled by a few dominant taxa, which represent priority targets for mechanistic studies. To achieve this and further reveal the scope and potential biological consequences of a break in the CNS-gut-microbiota-gut axis, a time-resolved, ecosystem-aware workflow is necessary to link host and gut microbiome metabolisms.

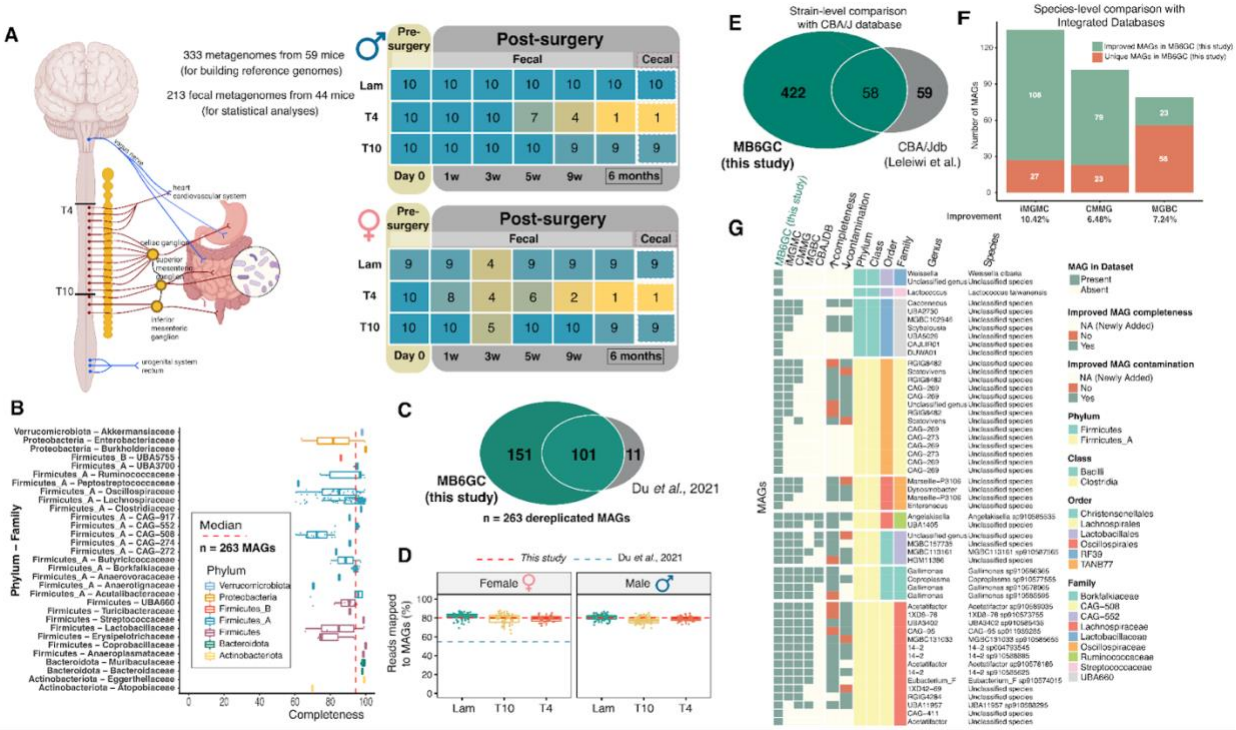


Fig. 1. Study design, characterization of recovered MAGs and comparative analysis of Mouse B6 Gut Catalog (MB6GC) against other published mouse gut microbiome catalogs.

(**A**) Schematic illustrates the relative location of spinal lesions and depicts the spinal sympathetic and vagal parasympathetic innervation to the gut. Tables (middle) describe the number of samples in this study, categorized by sex, spinal lesion level (Lam, T10, T4), and time post-lesioning. Note that autonomic complications in mice with lesions at spinal level T4 significantly reduce lifespan, yielding fewer T4 SCI mice, regardless of sex, beyond 7 weeks. (**B**) The completeness of the de-replicated MAGs (n=263; de-replicated at 95% ANI from 6,635 MAGs) is shown together with the phyla and families they span. New representative taxa include 2 phyla (Verrucomicrobiota and Proteobacteria), 6 families (Akkermansiaceae, Burkholderiaceae, Atopobiaceae, CAG-552, Enterobacteriaceae, CAG-272), and 41 genera (see GTDB data under Zenodo DOI). The median completeness across all MAGs is shown with a vertical dotted red line at 94.35%. Contamination levels are <10% for all de-replicated MAGs. (**C**) Venn diagram comparing the number of de-replicated MAGs recovered in this study against those from our prior pilot study[24]. (**D**) The fraction of reads recruited to the de-replicated set of MAGs, where the mean (red dotted line) is ~80% across sexes and spinal injury levels (also see SingleM supporting data in **Fig. S1B**). The mean for[24] (blue dotted line) was only available for female mice. The mean number of reads mapped to MAGs, as evaluated by the Wilcoxon test, was significantly improved in the current study (*p*-value= 6.11e-11). Schematic created with BioRender. **(E)** Venn diagram showing that at the MAG strain level (99% ANI dereplication), MAGs from MB6GC (this study; C57BL/6 mice) are largely mouse-strain specific (see **Methods**), with minimal overlap with the CBA/J MAG catalog. (**F**) A stacked bar chart shows that at the MAG species level (95% ANI dereplication), MAGs from MB6GC improve upon each of the previously published catalogs of mouse-gut reference genomes, including the Comprehensive Mouse Microbiota Genome Catalog (CMMG), the Mouse Gastrointestinal Bacteria Catalogue (MGBC), and the Integrated Mouse Gut Metagenomic

363 Catalog (iMGMC). (**G**) A taxonomically resolved heatmap shows that 53 MAGs from MB6GC
364 were either novel (n=15) or were the best representative genomes of their species (n=38) compared
365 to other catalogs. MB6GC's improvements in genome completeness and contamination over all
366 catalogs are shown.
367

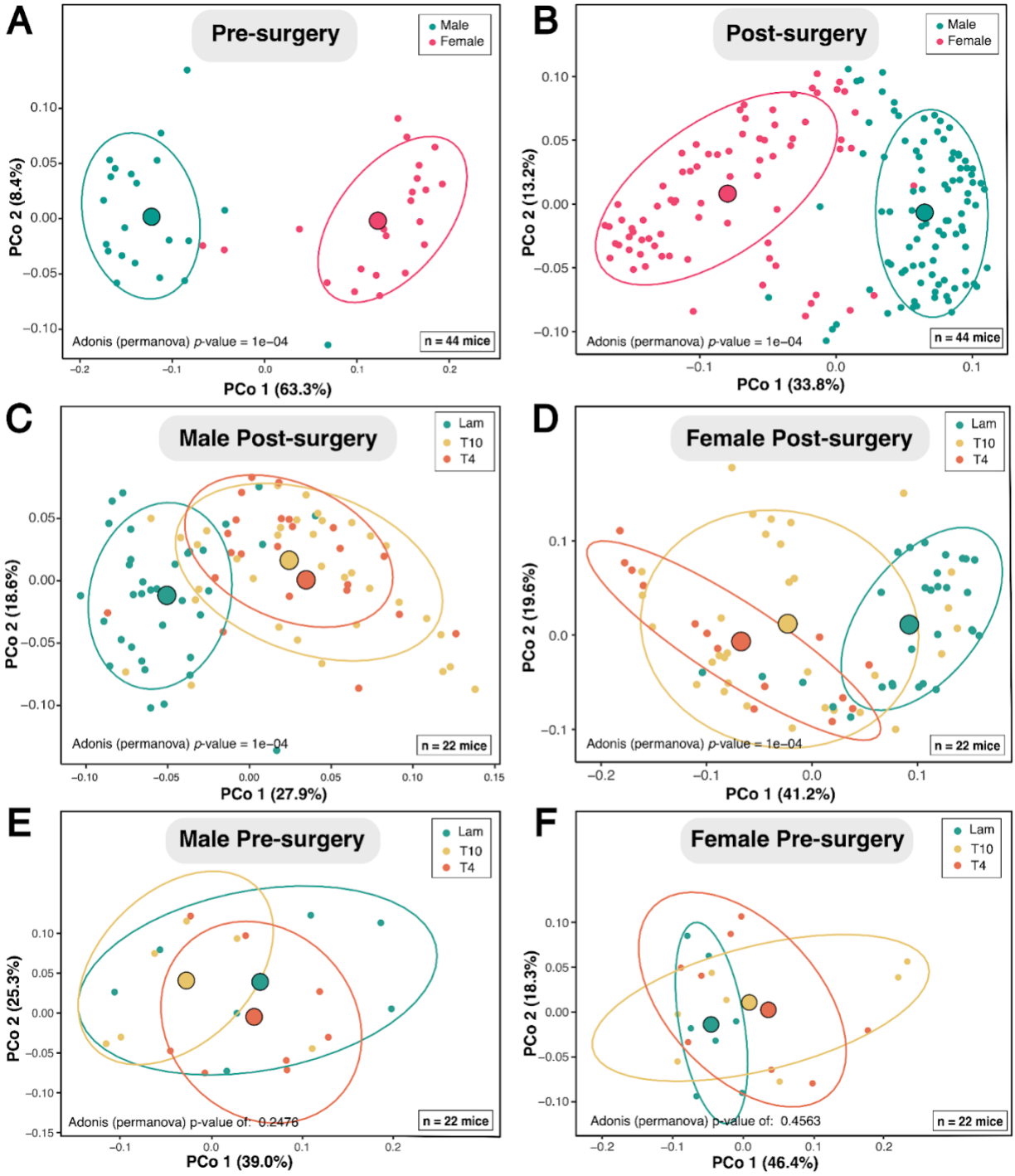


Fig. 2. Community-level microbiome changes are spinal cord-dependent and vary as a function of sex and spinal lesion level.

(**A-B**) MAG abundance-based principal coordinate analysis (PCoA) of Bray-Curtis dissimilarities from metagenomes of 44 mice (see "*Data Curation*" in **Methods** and **Fig. S1C**) show sex-based differences before (**A**) and after (**B**) spinal surgery. Consistent results arise from analyses before data curation (i.e., including outliers; see **Fig. S2A-B**). (**C-F**) PCoAs constructed using the same methods as in (**A-B**) show microbiome differences only after a break in the spinal cord-gut axis

(**C-D**). Before spinal lesioning (**E-F**) no group differences exist in male (n=22) or female (n=22) mice. See permanova *p*-values for assessment of significant differences in each panel. Small dots represent individual samples, while large dots represent the centroids per sex (**A-B**) or surgery (**C-F**).

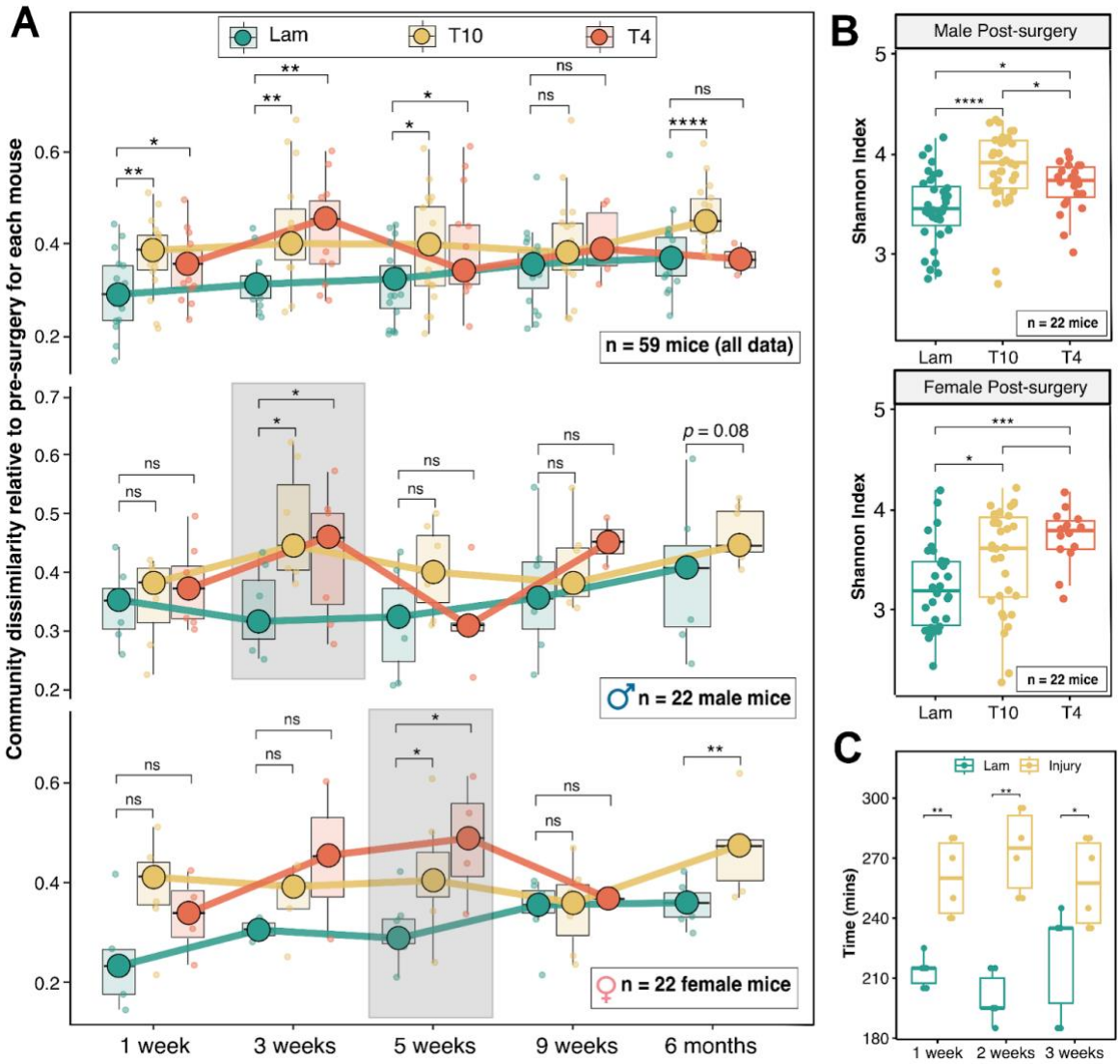
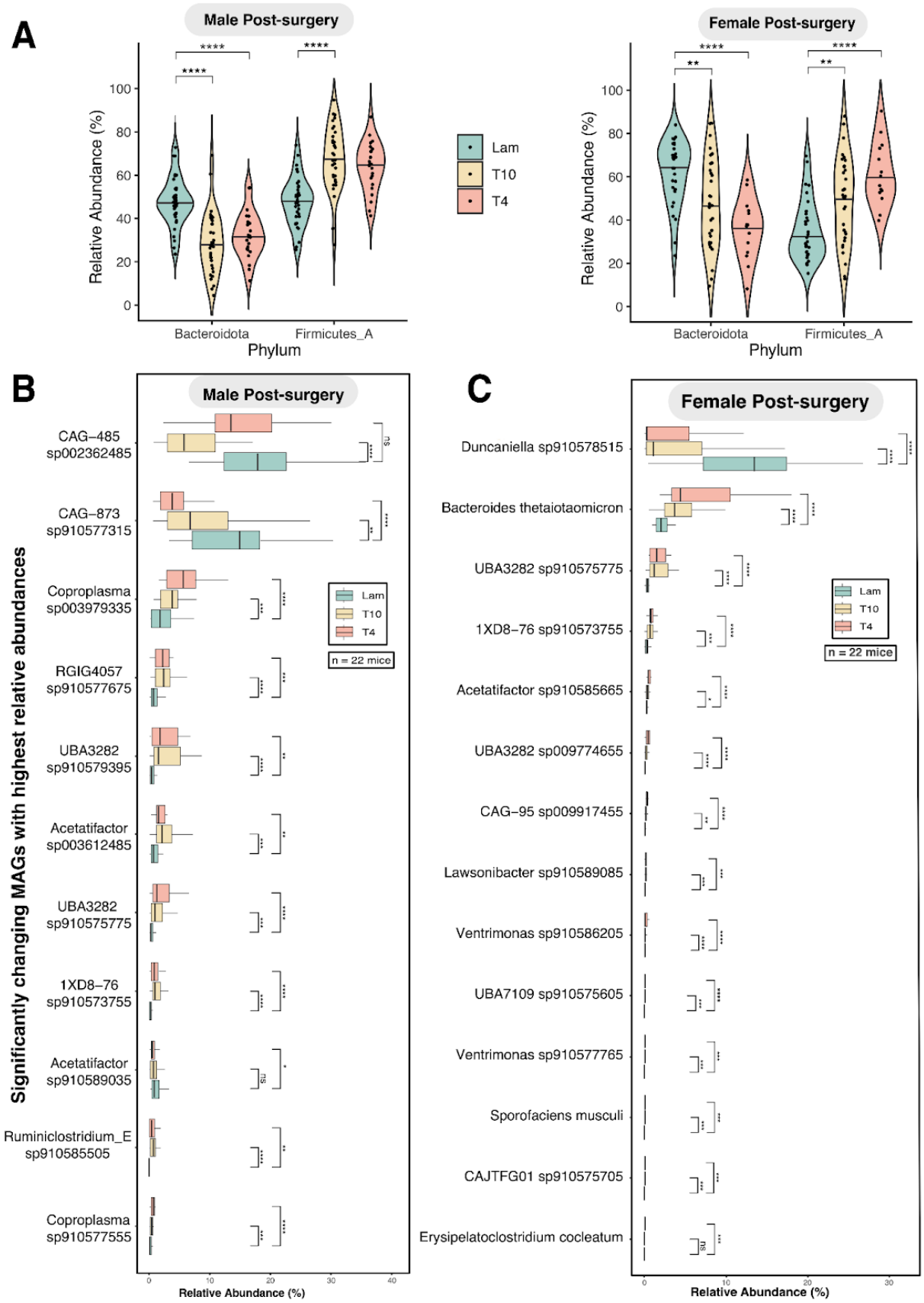


Fig. 3. Dynamic changes in microbiome beta- and alpha-diversity are spinal cord-dependent and vary as a function of sex and spinal lesion level.

(**A**) Boxplot injury-level- and time-resolved comparisons of Bray-Curtis dissimilarities (beta-diversity) relative to pre-surgery microbiome, shown for all data (n=59 mice) as well as curated male (n=22) and female (n=22) mouse data (see "*Data curation*" in **Methods** and **Fig. S1C**). Each small dot represents the microbiome change of an independent mouse relative to its baseline microbiome. Boxplot median values across mice per time-point are represented by large dots and are connected with thick lines. Peak changes are highlighted with gray boxes in male (3 weeks) and female (5 weeks) mice. (**B**) Boxplots showing spinal injury-level dependent microbiome alpha-diversity changes in curated male (n=22) and female (n=22) mice data. See baseline (Day 0) and sex- and time-resolved alpha diversity patterns in **Fig. S2C**. (**C**) Complete spinal injury slows intestinal transit time (Red carmine dye assay) relative to sham-lesion (Lam) controls. For all boxplots, whiskers represent the 25th and 75th percentile ranges while *p*-values represent those from Wilcoxon tests, adjusted for multiple comparisons by the fdr (false discovery rate) method (see **Methods**). ****: $p <= 0.0001$, ***: $p <= 0.001$, **: $p <= 0.01$, *: $p <= 0.05$, ns: $p > 0.05$.


A
Male Post-surgery
Female Post-surgery
Relative Abundance (%)
Bacteroidota
Firmicutes_A
Phylum
Lam
T10
T4
B
C
Significantly changing MAGs with highest relative abundances
CAG−485 sp002362485
CAG−873 sp910577315
Coproplasma sp003979335
RGIG4057 sp910577675
UBA3282 sp910579395
Acetatifactor sp003612485
UBA3282 sp910575775
1XD8−76 sp910573755
Acetatifactor sp910589035
Ruminiclostridium_E sp910585505
Coproplasma sp910577555
Duncaniella sp910578515
Bacteroides thetaiotaomicron
UBA3282 sp910575775
1XD8−76 sp910573755
Acetatifactor sp910585665
UBA3282 sp009774655
CAG−95 sp009917455
Lawsonibacter sp910589085
Ventrimonas sp910586205
UBA7109 sp910575605
Ventrimonas sp910577765
Sporofaciens musculi
CAJTFG01 sp910575705
Erysipelatoclostridium cocleatum
n = 22 mice
Relative Abundance (%)

398

399 **Fig. 4. Significant phylum- and MAG-level microbiome changes occur after disrupting the**
400 **spinal cord-gut axis.**

401 (**A**) Violin plots of the most abundant (median relative abundance >0.25%) phyla show that
402 Bacteriodota and Firmicutes_A significantly change only after disrupting the spinal cord-gut axis
403 in male (right) and female (left) mice. Pre-surgery control plots and plots of the least abundant
404 phyla (i.e., <0.25 median relative abundance) can be found in **Fig. S3A**. (**B**) Boxplots of the most
405 abundant (median relative abundance > 0.5%) significantly changing MAGs in male mice with
406 spinal lesions. All 68 changing MAGs can be found in **Fig. S3B**. (**C**) Boxplots of all significantly
407 changing MAGs (n=14) in female mice with spinal lesions. For violin plots and boxplots in all
408 panels (**A-C**), the 25th and 75th percentile ranges are shown. *p*-values represent those from
409 Wilcoxon tests, adjusted for multiple comparisons by the fdr (false discovery rate) method (see
410 "*Statistical methods for determining differentially abundant taxa*" in **Methods**). ****: $p <=$
411 0.0001, ***: $p <= 0.001$, **: $p <= 0.01$, *: $p <= 0.05$, ns: $p > 0.05$.

412

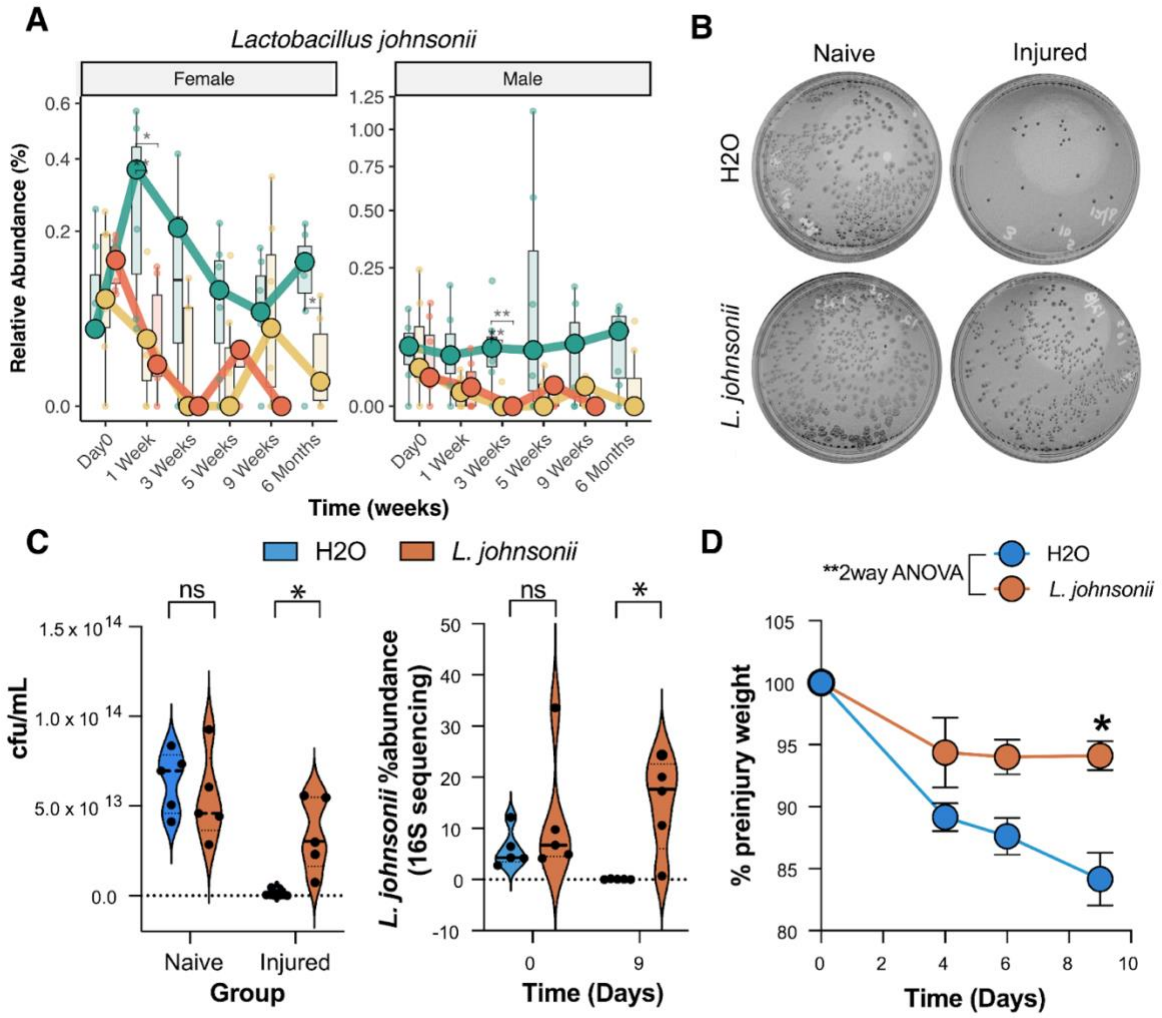


Fig. 5. The effects of feeding *L. johnsonii* to overcome the effects of a break in the spinal cord-gut axis.

(**A**) A rapid and protracted reduction in *L. johnsonii* MAG abundance occurs in the gut of male and female mice after a break in the spinal cord-gut axis. (**B-C**) Fecal cultures grown on MRS agar show reduced *L. johnsonii* colonies in mice subjected to a clinically-relevant model of spinal cord injury (SCI). SCI-dependent loss of *L. johnsonii* can be reversed by feeding mice 1 x 10^8cfu *L. johnsonii* daily after SCI (**C left**). 16S sequencing of fecal samples confirmed that this feeding regimen increased the abundance of *L. johnsonii* in the gut (**C right**). (**D**) Treatment with an *L. johnsonii* "probiotic" attenuates SCI-induced weight loss. *p*-values in (**A**) represent those from Wilcoxon tests, adjusted for multiple comparisons by the fdr (false discovery rate) method (see "*Statistical methods for determining differentially abundant taxa*" in **Methods**). ****: $p <= 0.0001$, ***: $p <= 0.001$, **: $p <= 0.01$, *: $p <= 0.05$, ns: $p > 0.05$. *p*-values for (**C-D**) are from 2-way ANOVA with Šídák's multiple comparisons test (**C**) or repeated measures 2-way ANOVA with Bonferroni's multiple comparisons test (**D**).

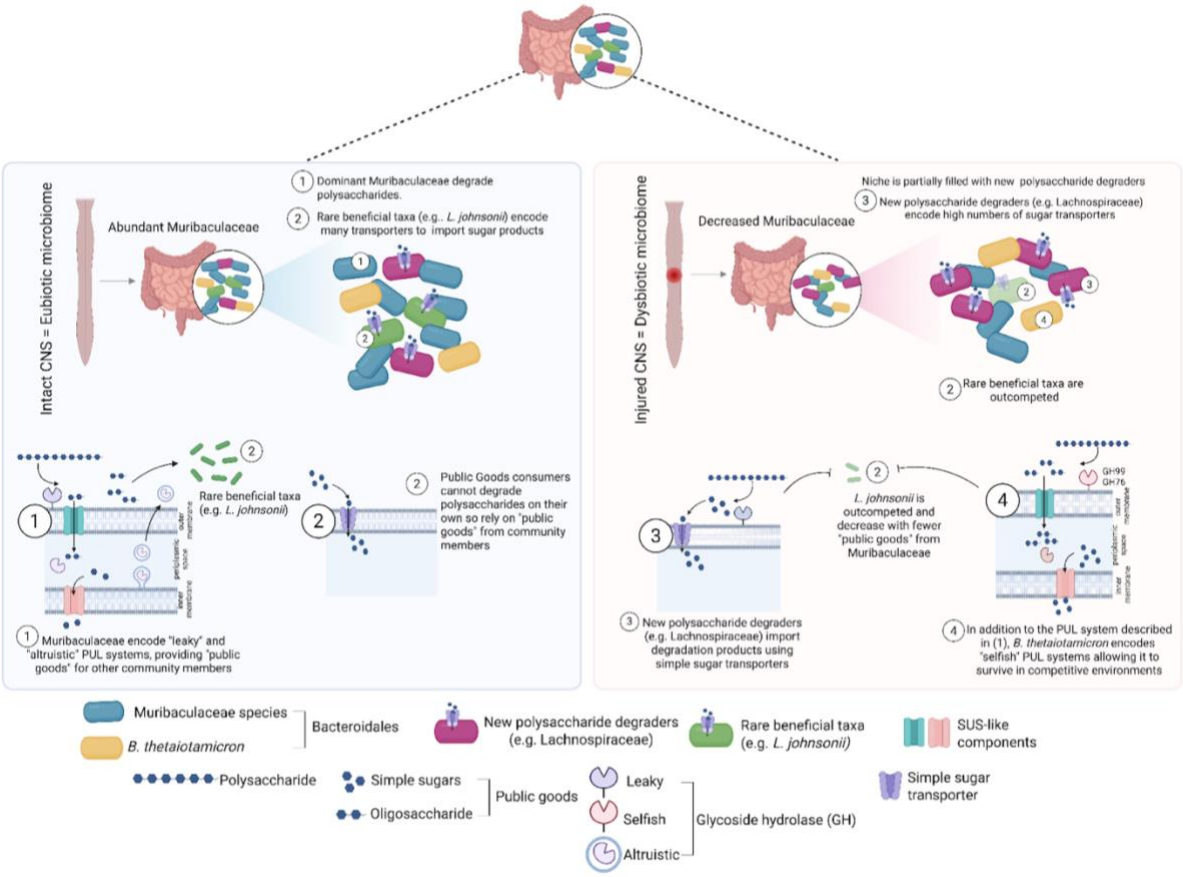


Fig. 6. Data-driven model of gut microbial community carbohydrate metabolism explains *L. johnsonii* depletion in mice with spinal lesions.

Metabolic cross-feeding model shows that a decrease in Muribaculaceae likely results in fewer public goods available to rare beneficial taxa like *L. johnsonii*. As Muribaculaceae decrease, their niche is partially replaced by other polysaccharide degraders that import most of the oligo- and mono-saccharides for their own use (e.g. Lachnospiraceae). This effectively starves rare beneficial taxa that cannot compete with taxa that encode "selfish" carbohydrate degradation and import systems.

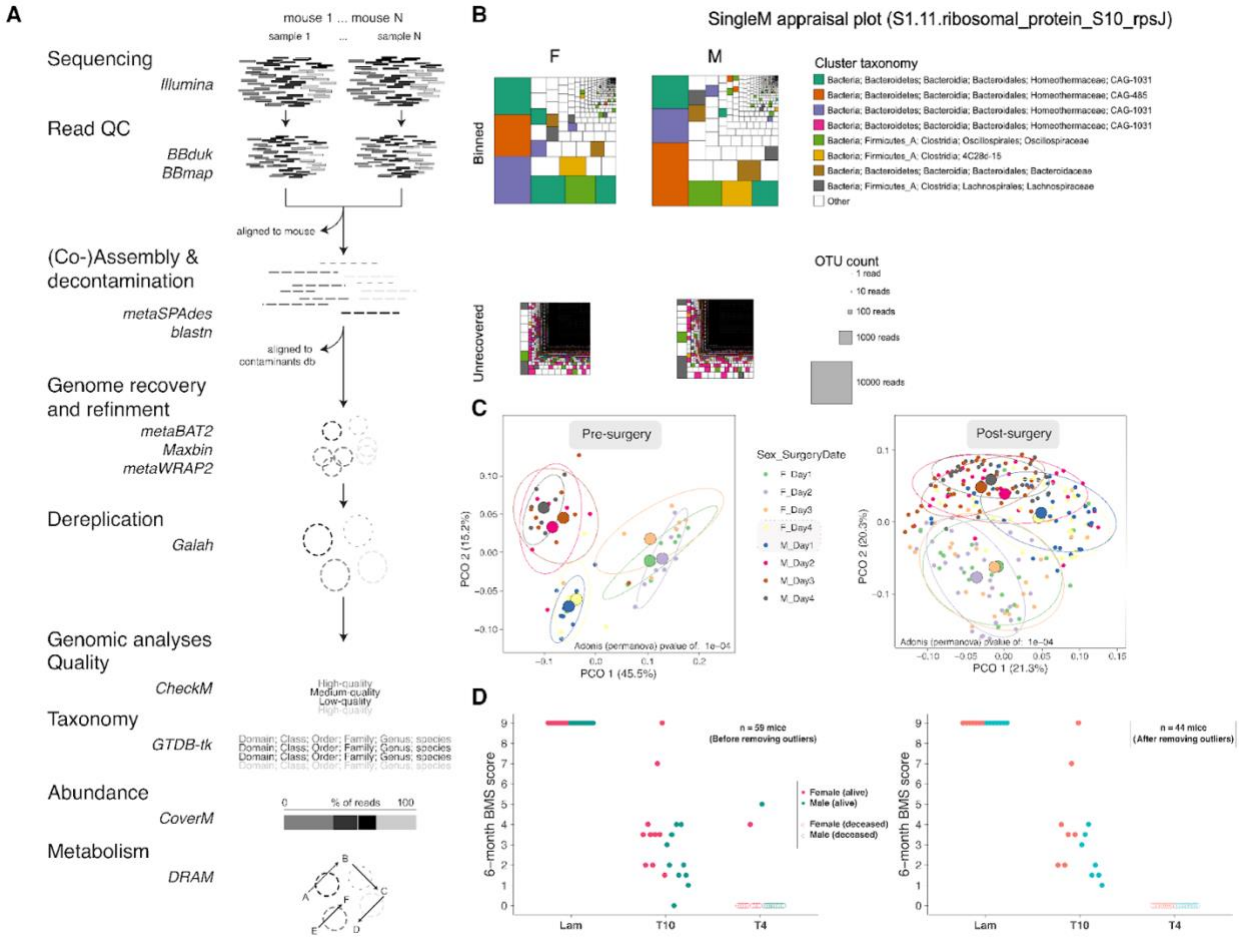
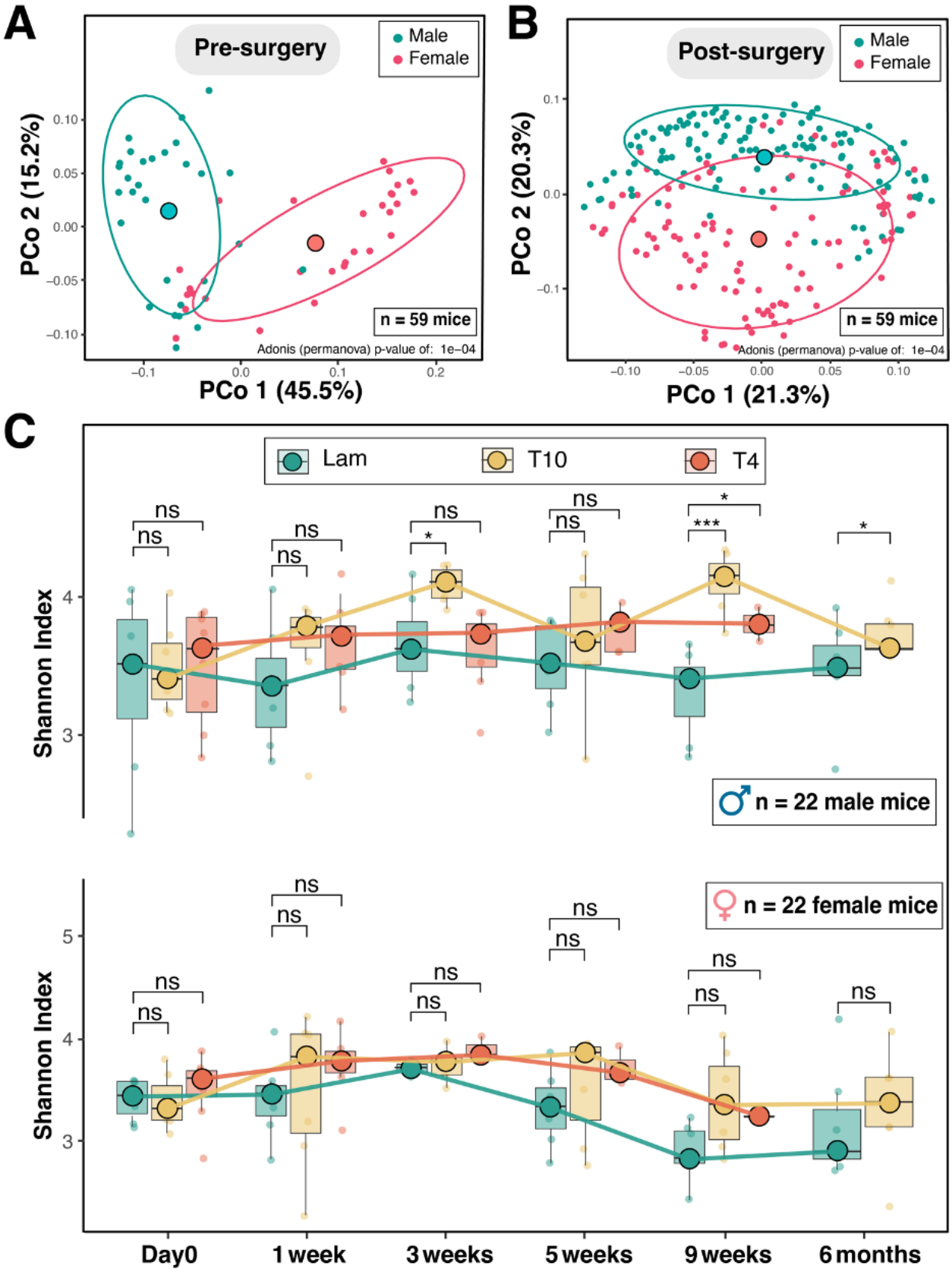


Fig. S1. Overview of the bioinformatic pipeline, read-profiling, and data curation.

(**A**) Overview of the bioinformatic pipeline used in this study for metagenomic analysis of gut microbiomes. Following sequencing, read quality control (QC) was carried out including read trimming and removal of reads matching to the mouse genome. Co-assembly of QC'd reads per mouse was done, followed by decontamination of scaffolds to be used for binning. Genome recovery, refinement, and dereplication at 95% ANI resulted in 263 MAGs that were used for further analysis. (**B**) read-based profiling of phylogenetic marker genes (see "*Read-based profiling*" in **Methods**) from raw reads suggest that, in both male and female mice, ~81% of the microbial diversity in the ecosystem was recovered in our updated MAG database. (**C**) Principal coordinate analysis (PCoA) based on Bray-Curtis dissimilarities for pre-surgery (left = Day 0) and post-surgery (right) microbiome samples, colored based on the combination of sex and surgery date (see "*Data curation*" in **Methods**) to discern confounders. Male mice having their operation performed on the first surgery day and female mice having theirs at the fourth day clustered closer to each other and differently from the larger male and female clusters, as indicated by a significant separation pre- and post-surgery (adonis *p*-value = 1e-04). (**D**) Basso Mouse Scale (BMS) scores taken at 6 months post-injury after data curation (n = 44 mice) – see the caption of **Fig. 1**.


A
Pre-surgery
Male
Female
PCo 2 (15.2%)
PCo 1 (45.5%)
n = 59 mice
Adonis (permanova) p-value of: 1e−04
B
Post-surgery
Male
Female
PCo 2 (20.3%)
PCo 1 (21.3%)
n = 59 mice
Adonis (permanova) p-value of: 1e−04
C
Lam
T10
T4
Shannon Index
n = 22 male mice
n = 22 female mice
Day0
1 week
3 weeks
5 weeks
9 weeks
6 months

459

460 **Fig. S2. PCoAs for male and female and alpha diversity.**

461 (**A-B**) MAG abundance-based principal coordinate analysis (PCoA) of Bray-Curtis dissimilarities
462 from metagenomes of 59 mice (including outliers), showing sex-based differences before (**A**) and
463 after (**B**) surgery. See permanova *p*-values for assessment of significant differences in each panel.
464 Small dots represent individual samples while large dots represent the centroids per sex. (**C**)
465 Boxplot injury-level- and time-resolved comparisons of Shannon's index, shown for curated male
466 (n=22) and female (n=22) mice data. Each small dot represents the microbiome changes of an
467 independent mouse at that timepoint. Boxplot median values across mice per timepoint are
468 represented by large dots and are connected with thick lines. This panel provides a detailed
469 temporal overview of the collective changes shown in **Fig. 3**, highlighting changes at specific time
470 points and the apparent reduction in statistical power for per-timepoint analyses, especially in
471 female mice. For all boxplots, whiskers represent the 25th and 75th percentile ranges while *p*-
472 values represent those from Wilcoxon tests, adjusted for multiple comparisons by the fdr (false
473 discovery rate) method (see **Methods**). ****: $p <= 0.0001$, ***: $p <= 0.001$, **: $p <= 0.01$, *: p
474 $<= 0.05$, ns: $p > 0.05$.

475

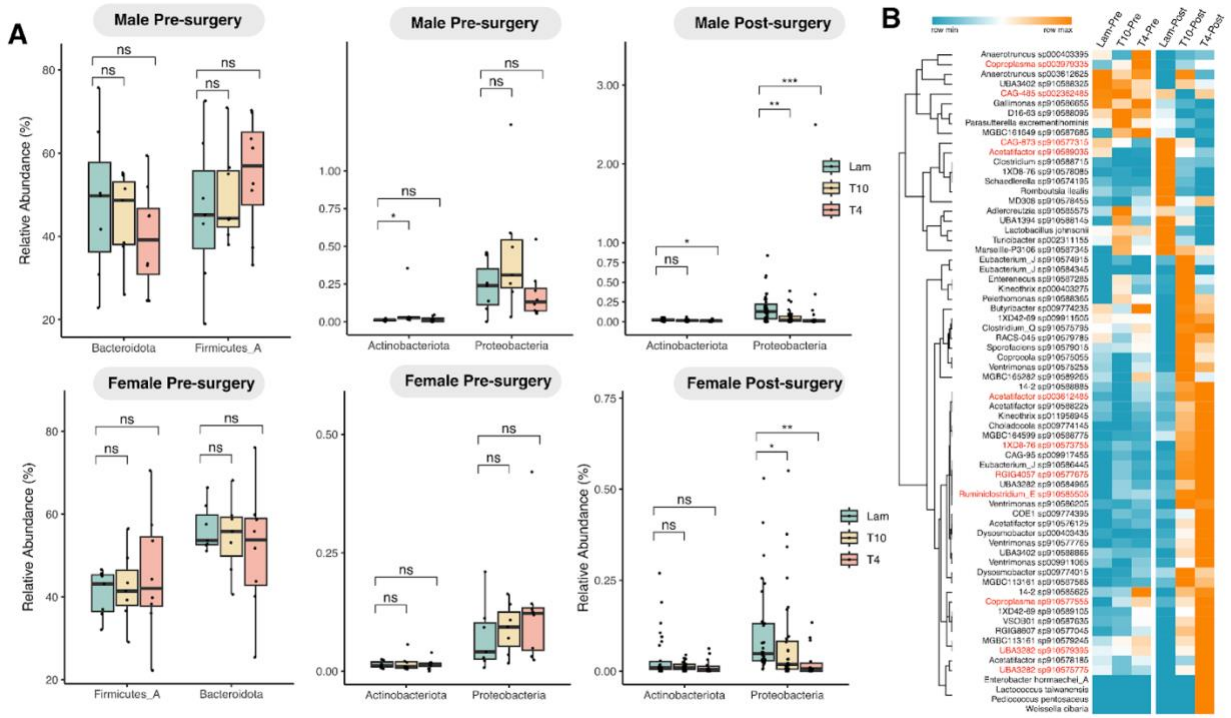
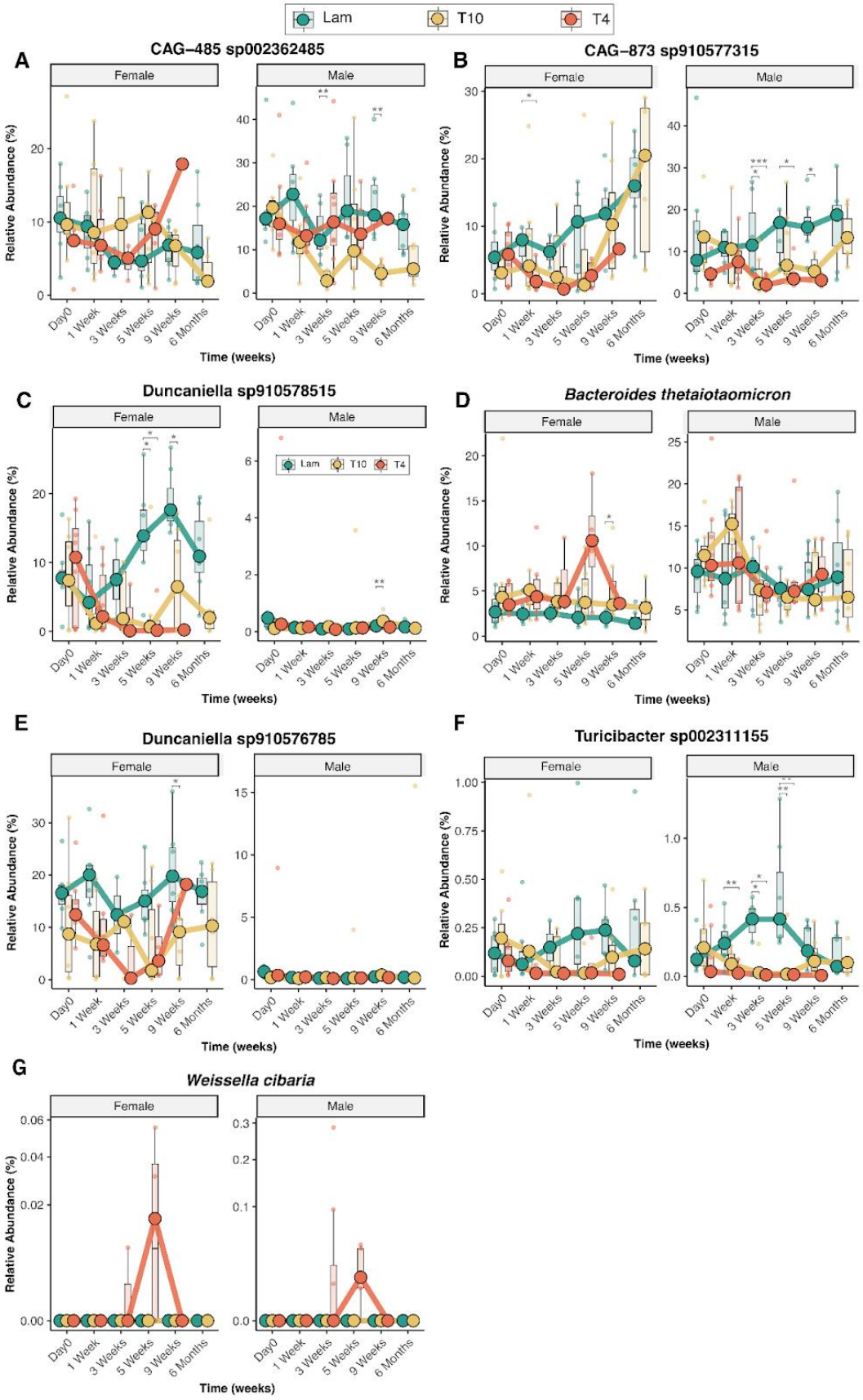


Fig. S3. Other significant phylum- and MAG-level microbiome changes after disrupting the spinal cord-gut axis (n=22).

(**A left**) Boxplots of the most abundant phyla, Bacteroidota and Firmicutes_A at baseline (Day0) for male (top) and female (bottom) mice; (**A middle**) Boxplots of the least abundant phyla, Actinobacteriota and Proteobacteria, at baseline in male (top) and female (bottom) mice; (**A right**) Boxplots of the least abundant phyla, Actinobacteriota and Proteobacteria, after surgery in male (top) and female (bottom) mice. (**B**) Heatmap of z-score-normalized MAG mean relative abundances before and after lesioning the spinal cord in male mice. Only significantly changing MAGs (n=68) are shown here. MAGs with the highest median relative abundances are highlighted in red and further shown in **Fig. 4B**. For boxplots in panel (A), the 25th and 75th percentile ranges are shown. *p*-values represent those from Wilcoxon tests, adjusted for multiple comparisons by the fdr (false discovery rate) method (see "*Statistical methods for determining differentially abundant taxa*" in **Methods**). ****: $p <= 0.0001$, ***: $p <= 0.001$, **: $p <= 0.01$, *: $p <= 0.05$, ns: $p > 0.05$.


Lam
T10
T4
A
CAG−485 sp002362485
B
CAG−873 sp910577315
C
Duncaniella sp910578515
D
Bacteroides thetaiotaomicron
E
Duncaniella sp910576785
F
Turicibacter sp002311155
G
Weissella cibaria
Female
Male
Relative Abundance (%)
Time (weeks)
Day0
1 Week
3 Weeks
5 Weeks
9 Weeks
6 Months

493

494

495 **Fig. S4. Temporal MAG abundance dynamics across the three experimental groups.**

496 Boxplots showing temporal dynamics of abundant bacteroidales MAGs (**A-E**), the MAG for the
497 rare beneficial Turicibacter taxon (**F**), and the sporadically changing *Weisella cibari* MAG (**G**) in
498 female (left insets) and male (right insets) mice resolved by experimental group and individual
499 samples. The most abundant differentially affected MAGs in male (i.e., CAG-485 sp002362485
500 and CAG-873 sp910577315 shown in **A-B**) and female (Duncaniella sp910578515 shown in **C**)
501 mice belong to the Muribaculaceae family, with persistent, spinal cord-dependent reductions in
502 Duncaniella sp910578515 noted in female, but not male mice, and mixed effects on CAG-485
503 sp002362485. Spinal-dependent changes in *Bacteroides thetaiotaomicron* (**D**) occurred in female
504 mice only. Some differentially abundant species identified post-lesion were reported in our
505 earlier pilot study[24], such as CAG-1031 bins – here CAG-1031 bins have been reclassified to
506 Duncaniella sp910578515 and Duncaniella sp910576785 (**D-E**) – and other rare taxa including
507 *Lactobacillus johnsonii* (**Fig. 5A**), *Weissella cibaria* (**F**), and the Turicibacter genus (its MAG
508 shown in **G**). Each small dot represents a MAG relative abundance at a specific time-point from
509 a single mouse over 6 months. Boxplot median values across mice per time-point are represented
510 by large dots and are connected with thick lines. For all boxplots, whiskers represent the 25th
511 and 75th percentile ranges while *p*-values represent those from Wilcoxon tests, adjusted for
512 multiple comparisons by the fdr (false discovery rate) method (see **Methods**). ****: $p <= 0.0001$, ***: $p <= 0.001$, **: $p <= 0.01$, *: $p <= 0.05$, ns: $p > 0.05$.

514

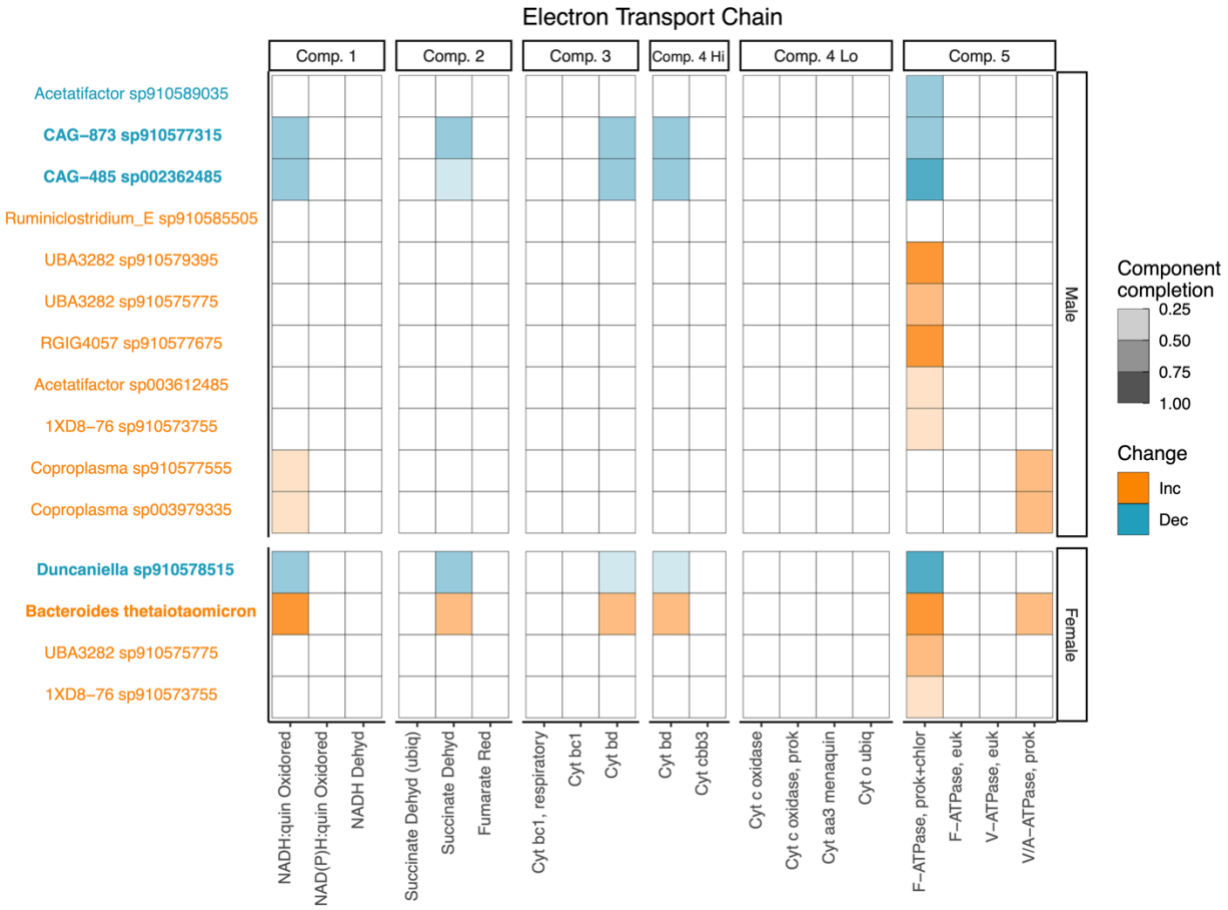
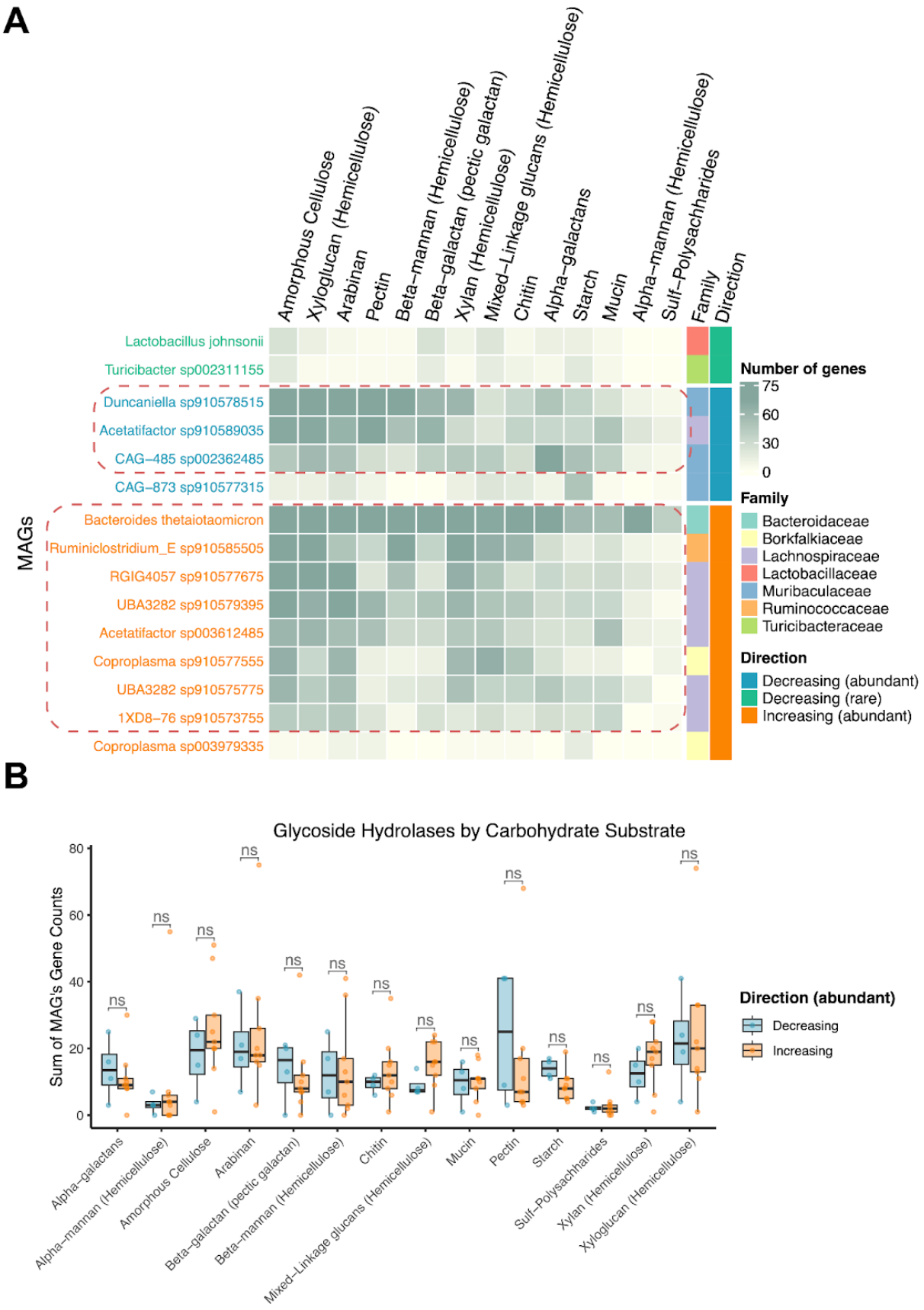


Fig. S5. Electron transport chain (ETC) components encoded by the most abundant changing MAGs in male and female mice.

A heatmap showing the completeness of each component of the ETC encoded by the most abundant changing MAGs according to annotations by DRAM. The color indicates the direction of change (orange for increasing and blue for decreasing) after a break in the spinal cord-gut axis while color intensity indicates component completion. MAGs in bold encode the minimum number of components required for a complete ETC. For example, a MAG encoding cytochrome *bd* or cytochrome *cbb3* is capable of respiring oxygen via their high-affinity cytochrome (Comp. 4 Hi). None of the MAGs encoded the low oxygen-affinity cytochromes (Comp. 4 Lo) while bacteroidales encoded the high oxygen-affinity cytochrome *bd*.


A
Amorphous Cellulose
Xyloglucan (Hemicellulose)
Arabinan
Pectin
Beta−mannan (Hemicellulose)
Beta−galactan (pectic galactan)
Xylan (Hemicellulose)
Mixed−Linkage glucans (Hemicellulose)
Chitin
Alpha−galactans
Starch
Mucin
Alpha−mannan (Hemicellulose)
Sulf−Polysachharides
Family
Direction
Lactobacillus johnsonii
Turicibacter sp002311155
Duncaniella sp910578515
Acetatifactor sp910589035
CAG−485 sp002362485
CAG−873 sp910577315
MAGs
Bacteroides thetaiotaomicron
Ruminiclostridium_E sp910585505
RGIG4057 sp910577675
UBA3282 sp910579395
Acetatifactor sp003612485
Coproplasma sp910577555
UBA3282 sp910575775
1XD8−76 sp910573755
Coproplasma sp003979335
Number of genes
75
60
30
0
Family
Bacteroidaceae
Borkfalkiaceae
Lachnospiraceae
Lactobacillaceae
Muribaculaceae
Ruminococcaceae
Turicibacteraceae
Direction
Decreasing (abundant)
Decreasing (rare)
Increasing (abundant)
B
Glycoside Hydrolases by Carbohydrate Substrate
Sum of MAG's Gene Counts
80
60
40
20
0
ns
Alpha−galactans
Alpha−mannan (Hemicellulose)
Amorphous Cellulose
Arabinan
Beta−galactan (pectic galactan)
Beta−mannan (Hemicellulose)
Chitin
Mixed−Linkage glucans (Hemicellulose)
Mucin
Pectin
Starch
Sulf−Polysachharides
Xylan (Hemicellulose)
Xyloglucan (Hemicellulose)
Direction (abundant)
Decreasing
Increasing

526

527 **Fig. S6. Carbohydrate substrate-resolved glycoside hydrolases (GHs) encoded by the most**
528 **abundant changing MAGs and rare beneficial taxa.**

529 (**A**) A heatmap showing the number of GHs encoded by each MAG organized by carbohydrate
530 substrate cleavage specificity according to DRAM's annotations (from the Carbohydrate-Active
531 enZymes Database or CAZy[70]). GHs that act on multiple substrates were accounted for in each
532 substrate category separately. Increasing abundant taxa after spinal cord lesions are shown in
533 orange, decreasing abundant taxa are shown in blue, and decreasing rare taxa are shown in green.
534 Dotted boxes highlight MAGs enriched in GHs. (**B**) Boxplot comparisons of the number of GHs
535 encoded by abundant increasing and decreasing MAGs organized by carbohydrate substrate
536 cleavage specificity as defined in (A). For all boxplots, whiskers represent the 25th and 75th
537 percentile ranges while *p*-values represent those from Wilcoxon tests, adjusted for multiple
538 comparisons by the fdr (false discovery rate) method (see **Methods**). ns: $p > 0.05$.

539

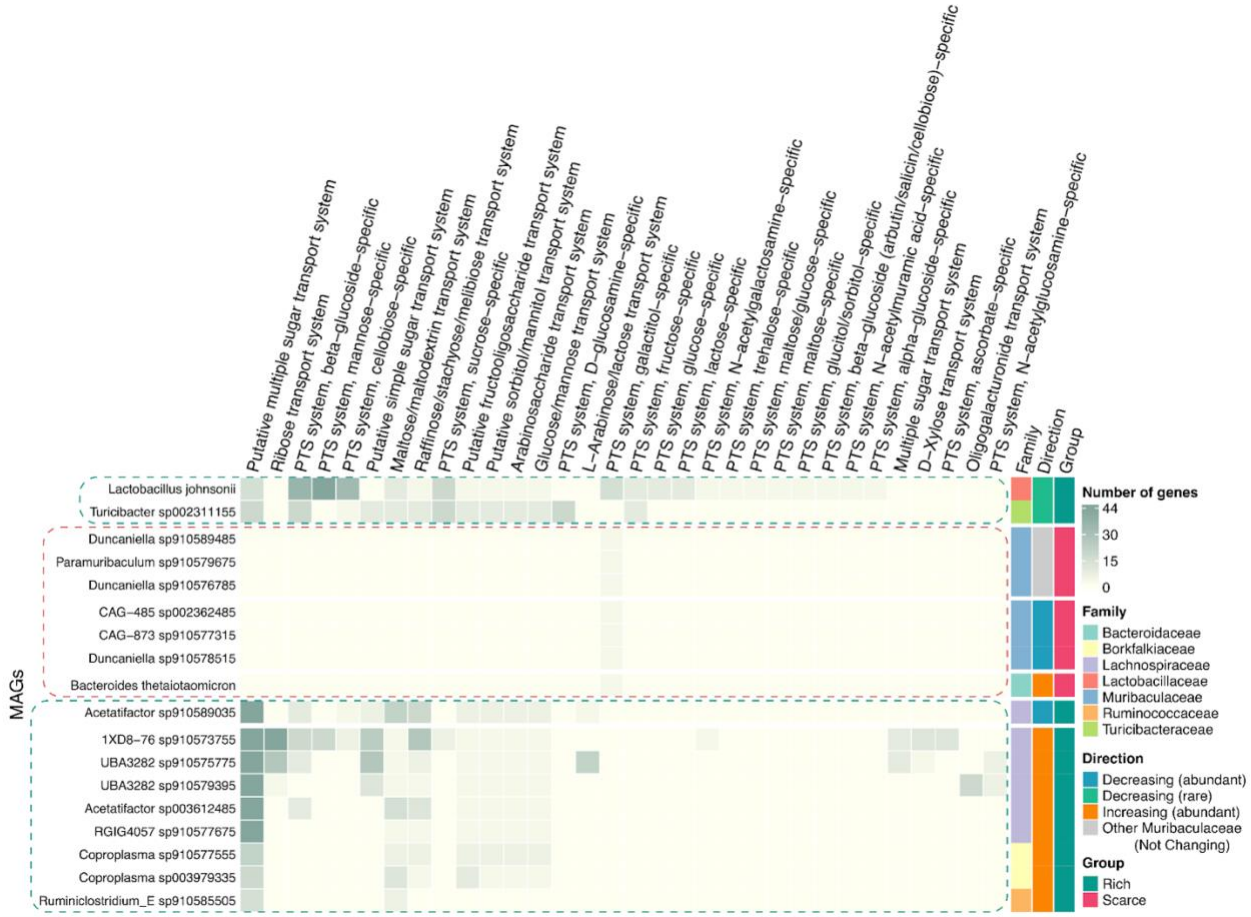


Fig. S7. Sugar-resolved transporters and phosphotransferase systems encoded by the most abundant changing MAGs, other Muribaculaceae MAGs, and rare beneficial taxa.

A heatmap showing the number of transporters and phosphotransferase systems (PTS) encoded by each MAG organized by sugar specificity according to DRAM's annotations. Component II of PTS was used as a marker for the sugar-specific system by DRAM. Direction of change is indicated by orange for increasing abundant taxa after spinal cord lesions, blue for decreasing abundant taxa, grey for other non-changing Muribaculaceae MAGs, and green for decreasing rare taxa. All bacteriodales (highlighted with a red dotted box) scarcely encode sugar transporters and PTS, while rare beneficial taxa and abundant increasing taxa (both highlighted with green dotted boxes) are relatively rich in these sugar import systems.

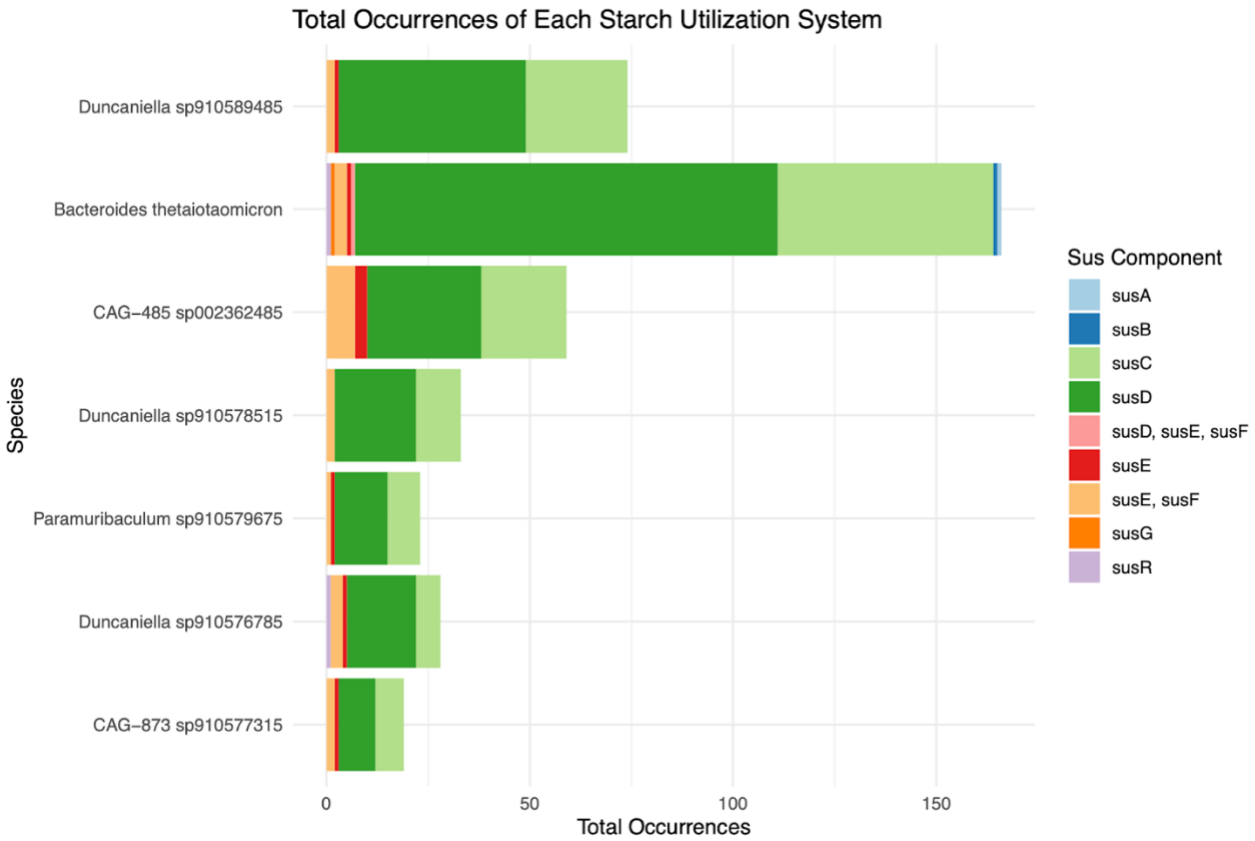


Fig. S8. Prevalence of starch utilization system (SUS) in members of the bacteroidales order.

Stacked bar chart showing the different SUS components detected in members of the bacteroidales order as defined in[65] and annotated by DRAM. *B. thetaiotamicron*, the only increasing bacteroidales MAG after injury as seen in female mice data, encodes a markedly larger number of SUS homologs (using *susC* and *susD* as marker genes).

Table S1. Taxonomic classification of the most abundant and rare beneficial changing MAGs, as well as non-changing Bacteroidales MAGs. Abundant decreasing and increasing MAGs are shown in blue and orange, respectively. Rare beneficial MAGs are shown in green, while non-changing MAGs are not colored. The Bacteroidales order is colored in green, with different shades showing different families (Muribaculaceae and Bacteroidaceae). Lachnospirales are colored in grey at the order and family (Lachnospiraceae) levels.

Direction after injury	Phylum	Class	Order	Family	Genus	Species
Decreasing	Bacteroidota	Bacteroidia	Bacteroidales	Muribaculaceae	CAG-873	CAG-873 sp910577315
Decreasing	Bacteroidota	Bacteroidia	Bacteroidales	Muribaculaceae	CAG-485	CAG-485 sp002362485
Decreasing	Bacteroidota	Bacteroidia	Bacteroidales	Muribaculaceae	Duncaniella	Duncaniella sp910578515
Increasing	Bacteroidota	Bacteroidia	Bacteroidales	Bacteroidaceae	Bacteroides	*Bacteroides thetaiotaomicron*
Decreasing	Firmicutes_A	Clostridia	Lachnospirales	Lachnospiraceae	Acetatifactor	Acetatifactor sp910589035
Increasing	Firmicutes_A	Clostridia	Lachnospirales	Lachnospiraceae	Acetatifactor	Acetatifactor sp003612485
Increasing	Firmicutes_A	Clostridia	Lachnospirales	Lachnospiraceae	1XD8-76	1XD8-76 sp910573755
Increasing	Firmicutes_A	Clostridia	Lachnospirales	Lachnospiraceae	UBA3282	UBA3282 sp910575775
Increasing	Firmicutes_A	Clostridia	Lachnospirales	Lachnospiraceae	UBA3282	UBA3282 sp910579395
Increasing	Firmicutes_A	Clostridia	Lachnospirales	Lachnospiraceae	RGIG4057	RGIG4057 sp910577675
Increasing	Firmicutes_A	Clostridia	Oscillospirales	Ruminococcaceae	Ruminiclostridium_E	Ruminiclostridium_E sp910585505
Increasing	Firmicutes_A	Clostridia	Christensenellales	Borkfalkiaceae	Coproplasma	Coproplasma sp003979335

Direction after injury	Phylum	Class	Order	Family	Genus	Species
Increasing	Firmicutes_A	Clostridia	Christensenellales	Borkfalkiaceae	Coproplasma	Coproplasma sp910577555
Decreasing (rare beneficial)	Firmicutes	Bacilli	Lactobacillales	Lactobacillaceae	Lactobacillus	*Lactobacillus johnsonii*
Decreasing (rare beneficial)	Firmicutes	Bacilli	Haloplasmatales_A	Turicibacteraceae	Turicibacter	Turicibacter sp002311155
No change	Bacteroidota	Bacteroidia	Bacteroidales	Muribaculaceae	Paramuribaculum	Paramuribaculum sp910579675
No change	Bacteroidota	Bacteroidia	Bacteroidales	Muribaculaceae	Duncaniella	Duncaniella sp910576785
No change	Bacteroidota	Bacteroidia	Bacteroidales	Muribaculaceae	Duncaniella	Duncaniella sp910589485

Methods

Animals and creating spinal cord lesions

All surgical and post-operative care procedures were performed in accordance with the Ohio State University Institutional Animal Care and Use Committee. Food (Teklad Global 14% Protein Rodent Maintenance Diet) and water were available at libitum. For metagenomic sequencing, N= 29 female C57BL/6 mice and N=30 male C57BL/6 mice from Jackson Laboratories (Bar Harbor, Maine) were used in this study. To prevent gut microbial cross-contamination due to co-habitation, all mice were single-housed upon arrival at our animal facility and for the duration of the study. Mice were anesthetized with an intraperitoneal cocktail of ketamine (80 mg/kg)/xylazine (10 mg/kg) after which a partial laminectomy was performed at the fourth thoracic spine (T4) or the tenth thoracic spine (T10). In mice modeling spinal cord injury (SCI), modified #5 Dumont forceps were inserted laterally and held closed for 3 seconds to produce a complete crush spinal cord injury. In total, 20 mice received a complete crush at T4, 20 mice received a complete crush at T10, and 19 mice received laminectomy (Lam) only (**Fig. 1A**).

Post-operatively, animals were hydrated with 2 mL Ringer's solution (subcutaneous) for 5 days. Bladders were voided manually at least twice daily for the duration of the study. No prophylactic antibiotics were used during or after surgery. Fecal samples for metagenomic sequencing were collected pre-lesion, 1-week, 2-weeks, 3-weeks, 5-weeks, 9-weeks, and 6-months post-lesion. Mice were removed from their home cage and placed into a ventilated, aseptic polystyrene compartment and fresh fecal samples were collected from each mouse into sterile tubes and immediately frozen in liquid nitrogen. Mice were returned to their home cage after sample collection. At 6-months post-lesion, mice were anesthetized with ketamine/xylazine for tissue collection. Cecal contents were collected by removal of the cecum and extrusion of cecal contents into a sterile Eppendorf tube. All samples were immediately frozen in liquid nitrogen and stored at -80°C until further use.

Analysis of neurological function

Spinally-mediated neurological function was assessed using the Basso Mouse Scale for Locomotion (BMS score) at 6 months post lesion. Two experimenters assessed BMS scores for mice during a 4-minute interval in the open-field. Scores were tallied for each hind paw and then averaged to obtain a single score per animal. Lam mice have a score of 9 indicating normal locomotor function. Mice with spinal lesions have scores below 9, with scores varying as function of lesion level, with higher lesion levels creating greater neurological impairment across the body and correspondingly lower BMS scores. Deceased mice were assigned a score of 0 in this study.

Lactobacillus johnsonii isolation:

603 Fresh fecal samples were collected from naïve C57BL/6 mice (Jackson Laboratories) into
604 sterile tubes and homogenized in 500ul sterile 0.1M PBS. 100ul of this homogenate was plated
605 on agar plates made with De Man, Rogosa and Sharpe agar (Lactobacilli MRS agar; Neogen
606 culture media NCM0035A), a selective culture medium designed to encourage growth of
607 Lactobacillus species. These plates were cultured under anaerobic conditions using Mitsubishi™
608 AnaeroPack- Anaero Anaerobic Gas Generator (Thermo Scientific™ R681001) for 48hrs and
609 numerous colonies were visible on the plates. To determine whether the resulting colonies were
610 indeed L. johnsonii we selected 10 isolated colonies for further characterization. These colonies
611 were expanded by culturing in MRS broth (MRS Broth; Millipore 69966) under static anaerobic
612 conditions for 48hrs at 37C. After the incubation, the cultures were centrifuged to pellet the
613 bacteria and cells were frozen in 40% glycerol-MRS broth and DNA was isolated from each
614 culture using the Qiagen DNeasy PowerSoil Pro kit (Qiagen 47014). After DNA isolation, 16S
615 PCR was run using universal 16S primers (GM3: AGA GTT TGA TCM TGG C; GM4: TAC CTT
616 GTT ACG ACT T) following protocol (Natalie Solonenko 2016. 16S PCR Universal. protocols.io
617 https://dx.doi.org/10.17504/protocols.io.c5my45). These primers amplify the 16S gene region for
618 all bacteria (they are not genus or species specific). PCR products were then run on an agarose gel
619 and gel-purified using Zymoclean Gel DNA Recovery Kit (Zymo Research; D4001). This gel-
620 purified DNA was sent to the Genomics Core Lab at The Ohio State University for Sanger
621 sequencing. The resulting sequences were then compared to all known DNA sequences using
622 nucleotideBLAST (https://blast.ncbi.nlm.nih.gov/). The results of this query for isolate #10 are
623 shown in Figure 1D. All results returned were specific for strains of *Lactobacillus johnsonii*,
624 indicating that we have been successful in isolating and culturing this bacterium from the mouse
625 gut microbiome. The same was true for all 10 isolates.

626 <u>Preparation of L. johnsonii glycerol stocks:</u>

627 After identifying that bacterial isolates were L. johnsonii, cultures were prepared to
628 generate glycerol stocks for storing and future use in experimental studies. Frozen glycerol stocks
629 (see above) were thawed and 1mL was transferred to a 50ml conical tube with 5mL sterile 0.1M
630 PBS to remove glycerol. Cells were centrifuged at 4000xg for 10 minutes to pellet bacteria and
631 supernatant was removed. Pellet was resuspended in 10mL of sterile MRS broth (MRS Broth;
632 Millipore 69966) and cultured for 24hrs at 37C in anaerobic conditions (Mitsubishi™ AnaeroPack-
633 Anaero Anaerobic Gas Generator; Thermo Scientific™ R681001). This 10mL culture was then
634 divided evenly (3.3mL) into 3 new sterile 50mL conical tubes containing a final volume of 25mL.
635 These cultures are incubated or 6-9hrs at 37oC in anaerobic chambers until absorbance/OD is
636 between 0.35 and 0.55 when read at 600nm. We compared the OD values to a growth curve chart
637 to determine approximate cfu/mL values. Bacteria are pelleted as above and resuspended in 40%
638 glycerol MRS broth (MRS Broth; Millipore 69966) at a final concentration of 1 x 109 cfu/ml in

639 cryovials and stored at -80C. To ensure accurate concentrations prior to use in animal studies, an
640 aliquot if then thawed, washed and plated on MRS agar (Lactobacilli MRS agar; Neogen culture
641 media NCM0035A) plates using serial dilutions to determine exact cfu/mL.

642 Oral delivery of L. johnsonii:

643 C57BL/6 female mice (Jackson Laboratories) received a mid-thoracic spinal cord lesion as
644 described above. Mice received 1 x 108 cfu/mL L. johnsonii from stored glycerol stocks, washed
645 in sterile 0.1M PBS, in 100ul sterile saline by oral gavage each day starting on day 1 post-injury
646 through day 9 post-injury. Fecal samples were collected before and again 9 days post-lesioning.
647 Fecal samples were homogenized in 500uL sterile 0.1M PBS and 100ul was plated on MRS agar
648 (Lactobacilli MRS agar; Neogen culture media NCM0035A) to enumerate Lactobacillus colonies.
649 From the same fecal samples, DNA was isolated using Qiagen DNeasy PowerSoil Pro kit (Qiagen
650 47014) for 16S sequencing (Novogene). Body weights were measured before surgery and
651 throughout the 9-day survival period.

652 Intestinal transit time assay:

653 Total intestinal transit time was measured using a 6% Red Carmine dye assay following
654 the protocol published in[36]. C57BL/6 female mice (Jackson laboratories) were given either a T9
655 complete crush spinal cord injury or a laminectomy (sham) control surgery (see surgical details
656 above). Starting at 7 days post-injury, and weekly thereafter, mice were given 150uL of 6% red
657 carmine (Sigma-Aldrich C1022) dissolved in a 0.5% methylcellulose (Sigma-Aldrich M0512)
658 solution. After oral gavage of the 6% carmine solution mice were fasted for the duration of the
659 assay. After 180 minutes, each mouse was placed in a clean cage and monitored at 5 minutes
660 intervals to assess fecal pellets for the passage of the red dye (fecal pellets are visibly red color).
661 The time of first red fecal pellet is recorded and whole gut transit time is calculated as the time
662 interval between the gavage and the first observed red fecal pellet.

663 DNA extraction and metagenomic sequencing

664 From 59 mice, 333 bulk metagenomic samples were prepared for sequencing (batch #1).
665 Both fecal and cecum samples were extracted using a DNeasy PowerSoil Pro Kit (# 47014,
666 QIAGEN) following the manufacturer's instructions. The samples were then normalized to 12
667 ng/uL and sent to the JP Sulzberger Columbia Genome Center. At the center, using a Riptide kit
668 from iGenomX for library preparation, sequencing was performed using Illumina NovaSeq 6000
669 targeting 26M 2x150bp reads per sample. Reads from bulk metagenomes ranged from 3.5 million
670 paired-end reads to 64.5, with a median value of 25.1 million paired-end reads. For 11 samples
671 with the lowest coverage, from 3.5 to 13.6 million reads, sequencing was redone at the same center,
672 at 25M 2x100bp reads per sample. Library prep was performed using Nextera XT kit from Illumina
673 (batch #2) at the Sullivan Lab before sending samples to the sequencing center. The prefix

674 "Repeat_" was added to their sample labels in all metadata spreadsheets and codes. Reads ranged
675 from 5.7 million paired-end reads to 26.8 for these 11 samples, with a median value of 24.2 million
676 paired-end reads.
677 These samples starting with "Repeat_" were used in addition to the original ones for
678 binning bacterial genomes but were discarded after calculations of abundances and further
679 analyses due to having different read lengths (batch #2 having 2x100bp).
680
681 Read quality control
682 Adapter trimming: Reads were trimmed with bbduk[71] (https://jgi.doe.gov/data-and-tools/bbtools/)
683 from the right end (ktrim=r) to keep ones longer than 30bp (minlength=30) by using a kmer of 23
684 (k=23) and allowing for a mismatch of 1 (hdist=1 hdist2=1) and a lower kmer at the ends of the
685 read (mink=11). In addition, reads mapping to adapters, artifacts, phix and overrepresented
686 sequences (i.e., low-complexity sequences) using "ref= adapters,artifacts,phix,<path-to-
687 overrepresented_sequences>" to remove all reads having a 23-mer match to these commonly
688 found sequences in Illumina runs.
689 Mouse reads removal: Reads mapping with a minimum identity of 95% to our mouse strain
690 C57BL/6NJ (genome downloaded from the NCBI assembly database using the accession number
691 "GCA_001632555.1") were removed using bbmap[71] (https://sourceforge.net/projects/bbmap).
692 Additional settings were used in bbmap (maxindel=3 bwr=0.16 bw=12 quickmatch fast minhits=2
693 overwrite=t usemodulo).
694 Quality trimming: Next, using bbduk, reads with average quality of 10 or ones where base quality
695 drops to 10 are trimmed from both sides keeping reads with a minimum length of 30 basepairs
696 (qtrim=rl maq=10 maxns=0 minlength=30 trimq=10).
697
698 Read-based profiling
699 Read-based profiling was performed using SingleM v 0.13.2
700 (https://github.com/wwood/singlem) using raw reads. First, the "pipe" command was run on raw
701 reads and on the 263 set of MAGs. Operational taxonomic unit (OTU) tables resulting from each
702 run (for paired reads of one sample) were then combined by sex. Finally, the "appraisal" method
703 was used to assess how well these MAGs represented the whole community, resulting in **Fig. S1B**.
704
705 Co-assembly and decontamination of scaffolds for binning (see Fig. S1A)
706 After quality control, all trimmed paired-end reads per mouse were combined and
707 assembled using SPAdes (v3.14.1) in the meta mode (i.e. metaSPAdes[72];
708 https://github.com/ablab/spades). All scaffolds below 500bp were discarded. The scaffolds longer
709 than 500bp were blasted (-task megablast) against an in-house contaminants database (see DOI:
710 10.5281/zenodo.13871704 with details in the ReadMe file). This database included the mouse

711 genome (strain C57BL/6NJ) and genomes of non-gut microbes and viruses that were sequenced
712 together with samples from this study (batches #1 and #2). Alignments that have 95% nucleotide
713 query-subject identity covering 80% of the scaffold were considered contaminants and were
714 therefore discarded.
715 After removing contaminants, trimmed reads from all bulk metagenomes were mapped to
716 the set of co-assembled decontaminated scaffolds using CoverM v0.6.1
717 (https://github.com/wwood/CoverM) in the "contig" mode with the following parameters "--min-
718 read-percent-identity .95 --min-read-aligned-percent .75 --discard-unmapped". Resulting bam
719 files were used as inputs for binning tools.
720
721 Construction of MAGs, estimation of abundance, and taxonomic classification (see Fig. S1A)
722 Metabat2[73] and Maxbin2[74] (with the 107 marker gene set) were run with co-assemblies and
723 per-sample bam files through UniteM v0.0.18 (https://github.com/dparks1134/UniteM) with a
724 minimum contig length to be binned of 1500bp. The resulting bins from each tool were refined per
725 mouse with metaWRAP v1.2 (bin_refinement module with the --keep-ambiguous flag, keeping
726 bins with more than 50% completion). The quality of all bins was assessed with CheckM[75] v
727 1.0.18. All 6635 bins from all 59 mice were then pooled with the 112 bins from[24]. The 6149 bins
728 that were estimated to be more than 60% complete and less than 10% contaminated were de-
729 replicated via Galah v0.3.1 (https://github.com/wwood/galah), resulting in 263 bins that captured
730 redundant sequence variation resulting from mouse-constrained co-assembly. These 263 bins were
731 used for further analysis.
732 Bins were taxonomically classified by the Genome Database Taxonomy Toolkit[76] (GTDB-
733 TK) v2.1.1 through the "classify_wf" module, which uses "release 207" of GTDB. The abundance
734 of de-replicated MAGs was estimated using CoverM v0.6.1 (https://github.com/wwood/CoverM)
735 in the "genome" mode with the trimmed mean method through the parameters "-m trimmed_mean
736 --min-read-percent-identity .95 --min-read-aligned-percent .75 --discard-unmapped".
737
738 GTDB vs NCBI Taxonomy:
739 Differences between NCBI and GTDB (release 207) include the following: (a) NCBI's
740 Firmicutes is divided into Firmicutes (includes order Bacilli), Firmicutes_A (includes order
741 Clostridia), and Firmicutes_B (includes order Dehalobacteriia); (b) Many members of the families
742 Bacteroidaceae, Prevotellaceae, Muribaculaceae are reclassified as Muribaculaceae, belonging to
743 the phylum Bacteroidota (i.e, Bacteroidetes in NCBI); (c) Some members of the Clostridiaceae
744 family (p__Firmicutes; c__Clostridia; o__Eubacteriales) are classified as CAG-508
745 (p__Firmicutes_A; c__Clostridia; o__TANB77) and include mostly unclassified species;
746 members classified as family CAG-552 (p__Firmicutes_A; c__Clostridia; o__Christensenellales)
747 were largely unclassified firmicutes belonging to order Eubacteriales or novel orders; there have

748 been also many events of reclassification in the class Clostridia from order Eubacteriales to
749 Lachnospirales, Oscillospirales, Peptostreptococcales, and TANB77.
750
751 Comparisons of genomic catalogs
752 First, genomes from different catalogs were collected. These include the Comprehensive
753 Mouse Microbiota Genome Catalog (CMMG)'s 1573 species-level representative genomes (95%
754 ANI) (https://ezmeta.unige.ch/CMMG/), the Integrated Mouse Gut Metagenomic Catalog
755 (iMGMC)'s 1296 representative genomes (https://github.com/tillrobin/iMGMC), the Mouse
756 Gastrointestinal Bacteria Catalogue (MGBC)'s 26,640 high-quality, non-redundant genomes
757 (MGBC-hqnr_26640; https://github.com/BenBeresfordJones/MGBC), and CBAJDB's 504 strain-
758 level (99% ANI) medium- and high-quality genomes (CBAJDB_MQ-HQ;
759 https://zenodo.org/records/8395759). Second, CheckM[75] v1.0.18 was run on MAG catalogs to
760 provide completeness and contamination information for each MAG – except for MGBC, for
761 which MAG completeness and contamination data were extracted from genomes metadata
762 (MGBC_md_26640). Third, for MGBC and CBAJDB (catalogs with no publicly available
763 species-level representative genomes), Galah v0.3.1 (https://github.com/wwood/galah) was run (-
764 -ani 95 --precluster-ani 90) to get species-level representative genomes for MGBC (n=1091) and
765 CBAJDB (n=113). Finally, at the strain level, MB6GC MAGs (n=499) were clustered with
766 CBAJDB (n=504) using Galah (--ani 99 --precluster-ani 90) - **Fig. 1E**. At the species level,
767 MB6GC MAGs (n=263) were clustered with CMMG, iMGMC, MGBC individually (**Fig. 1F**)
768 using Galah (--ani 95 --precluster-ani 90). Additionally, MB6GC MAGs (n=263) were also
769 clustered with MAGs from all representative genomes of the 4 catalogs together (**Fig. 1G**) using
770 Galah (--ani 95 --precluster-ani 90). Stacked bar charts were plotted using ggplot2
771 (https://ggplot2.tidyverse.org) while heatmap (as in all heatmaps hereinafter unless otherwise
772 stated) were made using the ComplexHeatmap package[77]
773 (https://github.com/jokergoo/ComplexHeatmap). All tables hereinafter were processed using the
774 tidyverse package in R[78].
775
776 Alpha- and beta-diversity estimation
777 Beta-diversity was assessed using principal coordinate analyses (PCoAs) with bray-curtis
778 dissimilarity matrices. Bray-curtis distances were calculated using the "vegdist" function from the
779 R package "vegan" (https://github.com/vegandevs/vegan). Vegan's "adonis2" function was used
780 to perform permanova tests on bray-curtis distances with 9999 permutations. Longitudinal
781 analyses were done by plotting bray-curtis distances to the mouse's Day 0 sample to account for
782 individual variations using ggplot2 (https://ggplot2.tidyverse.org). Statistical testing was
783 performed via the wilcox_test function from the "rstatix" package (https://CRAN.R-
784 project.org/package=rstatix), with the following parameters: "paired=FALSE, ref.group = 'Lam',

785 detailed=TRUE, alternative='less', p.adjust.method ='fdr'. Significance was shown using the
786 "ggpubr" package's stat_pvalue_manual function (https://rpkgs.datanovia.com/ggpubr).
787 Alpha diversity was assessed by Shannon's index using the "diversity" function from the
788 vegan package. Shannon's indices for samples were plotted by timepoint (i.e., samples at one time
789 point) and as an aggregate (e.g., all samples post-injury). Longitudinal statistical comparisons
790 shown in boxplots were made in a fashion similar to beta-diversity. Aggregate statistical
791 comparisons shown in main figures were performed using ggpubr's stat_compare_means, with the
792 following parameters: method = "wilcox.test", paired=FALSE, method.args = list(exact=FALSE),
793 label = "p.signif", hide.ns =TRUE, ref.group = ".all".
794
795 Data curation
796 Upon observing beta-diveristy in all samples after sham or spinal cord injury, samples were
797 shown to cluster into 3 main groups. Since mice had their operations in 4 batches, at 4 different
798 dates, we hypothesized that this might be one of the confounders. Colored by the day (i.e., date)
799 on which the surgery was performed on mice, male mice having their operation performed at the
800 first surgery day and female mice having theirs at the fourth day clustered closer to each other and
801 differently from their respective larger male and female clusters (see **Fig. S1C**). These mice were
802 excluded from further analyses (e.g., alpha and beta diversity, etc.). In addition, repeated samples
803 (batch #2), having a different read length from batch #1, were also excluded from these analyses.
804 Main male and female clusters were treated separately and as different cohorts.
805
806 Statistical methods for determining differentially abundant taxa (Fig. 4, Fig. S3)
807 The trimmed-mean coverage values from CoverM were converted to relative abundance
808 per sample after excluding repeated samples. Relative abundances were summed per each taxon
809 and used for further tests. Considering the complex experimental design and the large number of
810 taxa to be investigated (**Fig. 1**), it was challenging to test significance per time point due to losing
811 statistical power, especially for female mice. Therefore, two statistical approaches were used to
812 examine the data. First, samples collected from every two consecutive timepoints (e.g., "Day0 and
813 1week", "1week and 3weeks", and so on) were combined and tested for omnibus changes between
814 the three experimental groups; the relative abundance values were fed into a Kruskal-Wallis test
815 (from the kruskal.test function in R) followed by a Benjamini-Hochberg correction for multiple
816 testing to check if there is a difference between this taxon's abundance in Lam, T10, and T4 mice
817 collectively. To examine the specific changes (e.g., differences between the taxon's abundances
818 in Lam vs T10 or Lam vs T4), statistically significant taxa from Kruskal-Wallis tests (adjusted *p*-
819 values of less than 0.05) were then tested by a Wilcoxon signed-rank test (from the
820 pairwise.wilcox.test function in R) followed by a Benjamini-Hochberg correction. No taxa turned
821 out to be differentially abundant between Lam and T10 or T4 from the first timepoint pair (i.e.,

822 "Day0 and 1week"). Taxa deemed differentially abundant between injured and control (i.e., Lam)
823 mice by these tests were then collected and plotted in **Fig. 4** and **Fig. S3**. Taxa were plotted
824 depending on their abundance as follows: if the median abundance of the taxon (in any of the three
825 treatments) was above 20% for phyla or 0.5% for species, it was plotted in the high-relative
826 abundance plots. Second, these taxa were compared post-surgery (> Day0) with a Wilcoxon
827 signed-rank test followed by fdr correction. Hence, consensus taxa plotted in **Fig. 4** and **Fig. S3**
828 were both statistically significant upon testing the samples aggregated at the post-injury level (>
829 Day 0) *and* at the two consecutive timepoints level.
830

831 MAG functional annotation

832 DRAM v1.3.6 (https://github.com/WrightonLabCSU/DRAM) was used to annotate MAGs
833 against multiple databases, including the Carbohydrate-Active enZymes Database or CAZy[70].
834 DRAM's distillate outputs, *i.e.*, the "product.tsv" and the Carbon Utilization and Transporters tabs
835 of the "metabolism_summary.xlsx," were used to profile each MAG's electron transport chain
836 components (**Fig. S5**), GHs (**Fig. S6**), and sugar transporters (including PTS components; **Fig.**
837 **S7**), respectively, whereas each MAG's "Annotations" table was used to profile its PUL systems
838 (**Fig. S8**). MAG-resolved heatmaps were generated for (i) the electron transport chain, (ii) GHs by
839 counting the number of genes belonging to each "subheader" (representing a different
840 carbohydrate substrate), including oligo- and backbone-cleavage enzymes, (iii) sugar transporters
841 by counting the number of genes belonging to each "module" (representing a different transport
842 system), and (iv) PUL systems by counting the number of genes whose annotations contain the
843 strings "susA", "susB", "susC","susD","susE", "susF","susG", and "susR".

844 Temporal dynamics of select taxa

845 Relative abundances of select taxa were plotted longitudinally using ggplot2
846 (https://ggplot2.tidyverse.org). Statistical testing was performed via the wilcox_test function from
847 the "rstatix" package (https://CRAN.R-project.org/package=rstatix), with the following
848 parameters: "paired=FALSE, ref.group = 'Lam', detailed=TRUE, alternative='two.sided',
849 p.adjust.method='fdr'. Significance was shown using the "ggpubr" package's stat_pvalue_manual
850 function (https://rpkgs.datanovia.com/ggpubr).
851

852 Data availability

853
854 All data produced in this study are publicly available. Curated data and all used scripts and
855 databases can be found in the following Zenodo DOI: 10.5281/zenodo.13871704 (unpublished

until review is complete). Raw reads are uploaded to NCBI SRA under the BioProject accession number: PRJNA1168861 (to download, see relevant tables in the Zenodo directory).

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Author Contributions

P.G.P., M.B.S., K.A.K., A.A.Z. conceived the project and experimental design. P.G.P., M.B.S., K.A.K., A.A.Z., and M.M. developed the data analysis plan. P.G.P., M.B.S., K.A.K., A.A.Z., M.M., G.J.S., and J.M.S. helped with data interpretation and writing of the manuscript. M.M. performed the statistical, profiling, functional, and bioinformatic analyses, and managed and coordinated responsibilities for the manuscript preparation. A.A.Z. guided and supervised statistical, profiling, functional, and bioinformatic analyses, performed the decontamination analyses, and drafted the manuscript with M.M. M.M., K.A.K., A.A.Z., and G.J.S. contributed to figure generation and functional analyses and, with P.G.P., contributed to improving illustrations. G.J.S. uploaded raw reads to SRA. K.A.K. performed the intestinal transit time assay, isolation and culture of *L. johnsonii, L. jonhsonii* supplementation experiments, murine spinal cord injuries, as well as daily animal care and sample collection. J.D. performed the DNA extraction for metagenomic sequencing. All authors read and reviewed the manuscript and approved it in its final form.

Declaration of interests

All authors declare no competing interest.

Declaration of generative AI and AI-assisted technologies

During preparation of this work, Microsoft CoPilot and ChatGPT were used periodically to improve writing or shorten passages. After using it, the authors reviewed and edited the content as needed and take full responsibility for the content of the publication.

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